



# Graphene as an Imaging Platform of Charged Molecules

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Supporting Information

**ABSTRACT:** Graphene, a single atom layer of carbon atoms, provides a two-dimensional platform with an extremely high sensitivity to charges due to its unique band structure and high surface-to-volume ratio. Graphene field-effect transistor (G-FET) biosensors have, indeed, demonstrated a detection limit of subnanomolar or even subpicomolar. However, in G-FET, signal is averaged throughout the whole channel, so there remains a need to visualize the spatial distribution of target molecules on a single G-FET, to provide further insight into target molecules and/or biological functions. Here, we made use of graphene as an imaging platform of charged molecules via Raman microscopy. Positively (or negatively) charged



microbeads with a diameter of 1  $\mu$ m were dispersed in a buffer solution and were attached on graphene. We found out that Raman peaks of graphene, where positively (or negatively) charged beads contacted, were up-shifted (or down-shifted) significantly, indicating that the carrier density in the graphene was locally modulated by the charged beads and the charge state of the beads was represented by the peak-shift direction. From the peak shift, the change in the carrier density was calculated to be +1.4  $\times$  10<sup>12</sup> cm<sup>-2</sup> (or -1.0  $\times$  10<sup>12</sup> cm<sup>-2</sup>). By taking Raman peak-shift images, we visualized distribution of charged molecules on graphene with a spatial resolution below 1  $\mu$ m. The technique described here overcomes the limitation of spatial resolution of G-FET and provides a new route to graphene-based chemical and biosensors.

# INTRODUCTION

Graphene, a two-dimensional (2D) sheet of hexagonally arranged carbon atoms, offers an ideal sensing platform owing to its 2D nature and its unique band structure. The 2D nature creates a uniform and large sensing field with ultrahigh sensitivity to charges because every atom in a graphene sheet makes contact with its environment. Therefore, even a few chemical dopants can lead to significant modulation of its carrier (electron or hole) density. In graphene field-effect transistor (G-FET) sensors, where graphene is used as a channel, a change in the carrier density is monitored as a drain current change. There have been reports on G-FET sensors showing successful detections of various gases,<sup>1</sup> ions,<sup>2</sup> biomolecules,<sup>3</sup> and so on. However, in G-FET, the signal is averaged throughout the whole channel, so that spatial information of target molecules on G-FET cannot be obtained, and the signal is canceled out if both positively and negatively charged molecules are adsorbed in a single FET sensor. There are several methods to see local charge puddles in graphene with a high spatial resolution, including local potential Kelvin probe microscopy<sup>4</sup> and scanning single-electron transistors.<sup>5</sup> Here, we made use of graphene as an imaging platform of charged molecules via Raman microscopy. Raman microscopy is a noncontact, nondestructive, and label-free imaging technique with a spatial resolution below 1  $\mu$ m and can be

applied to samples in liquid environment, used to analyze biological materials,<sup>6</sup> nanocarbon materials,<sup>7</sup> polymers,<sup>8</sup> and so on. Furthermore, it has been known that a change in the carrier density in graphene appears as distinct Raman peak shifts.<sup>9</sup> Thus, by monitoring the peak shifts, local chemical dopants on graphene can be visualized via Raman microscopy. This method also enables one to investigate the charge state of biomolecules in liquid. Indeed, Paulus et al. have demonstrated a biological cell monitoring on graphene via Raman microscopy.<sup>10</sup> We conducted our study with polystyrene (PS) beads of known  $\zeta$ potential and demonstrated that a graphene imaging platform can visualize both positively and negatively charged molecules.

# RESULTS AND DISCUSSION

Analysis about Raman Images of Graphene Films under a Buffer Solution. To ensure that graphene provides a uniform imaging platform in a liquid environment, we took Raman images of graphene films using Raman microscopy. The schematic experimental setup is depicted in Figure 1a. A phthalate buffer solution at pH = 4 (50 mM, Horiba) was dropped on graphene films. Figure 1b shows a typical Raman

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**Figure 1.** (a) Schematic of setup for Raman measurement of graphene films immersed in a buffer solution. (b) A typical Raman spectrum of graphene showing G-band (1591 cm<sup>-1</sup>), 2D-band (2686 cm<sup>-1</sup>), and D-band (1341 cm<sup>-1</sup>) as well as N<sub>2</sub> in air at 2329 cm<sup>-1</sup>. (c, d) Raman image of graphene constructed by  $I_G$  (c) and  $I_{2D}$  (d). The image is 10 × 184 pixels, where the pixel sizes are 205.2 nm in x and 214.2 nm in y directions. The scale bar is 5  $\mu$ m. (e, f) Histograms of  $G_{pos}$  (e) and  $2D_{pos}$  (f). Solid lines are Gaussian fitting curves for a guide to the eye.

spectrum of graphene. The peak parameters are determined using the Lorentz function. The Raman spectrum shows distinctive peaks at 1591, 2686, and 1341 cm<sup>-1</sup> assigned to Gband, 2D-band, and D-band of graphene, respectively. The peak at 2329 cm<sup>-1</sup> is assigned to N<sub>2</sub> in air, which does not disturb Raman analysis of graphene. The intensity ratio of the G-band to 2D-band  $(I_G/I_{2D})$  and the full width at half-maximum of 2Dband ( $\Gamma_{2D}$ ) are calculated to be 0.64 and 29 cm<sup>-1</sup>, respectively. The symmetric Lorentzian line shape of the 2D-band indicates that the graphene films are a single layer.<sup>11</sup> Figure 1c,d shows Raman images constructed by  $I_{G}$  and  $I_{2D}$ , respectively. The Raman images clearly show that the obtained graphene films were uniform throughout the imaged area. To study more in detail, the Raman spectrum variance is investigated by making histograms of G-band peak position  $(G_{pos})$  and 2D-band peak position (2D<sub>pos</sub>; Figure 1e,f). The deviation in the peak positions was sufficiently small ( $< 2 \text{ cm}^{-1}$ ), which supports that the graphene provides a uniform imaging platform. The deviation is likely attributed to the inhomogeneous charged impurities in the SiO<sub>2</sub> substrate.<sup>12</sup>

To prove that Raman spectrum of graphene is sensitive to the carrier density change, we changed the carrier density by changing the pH of the buffer solution on graphene and investigated Raman spectrum of graphene. It is known that a pH change leads to a modulation of the carrier density of graphene.<sup>2</sup> Raman images of the graphene films were taken at pH = 4 (phthalate buffer, 50 mM, Horiba), 7 (phosphate buffer, 25 mM, Horiba), and 9 (tetraborate buffer, 10 mM, Horiba). The averaged Raman spectra throughout the Raman images (see Figure S1 in the Supporting Information) are shown in Figure 2a–c. The results clearly show that both  $G_{pos}$  and  $2D_{pos}$ are down-shifted with increasing pH. In contrast, no shift was observed at D-band and at the 2329 cm<sup>-1</sup> peak assigned to N<sub>2</sub>, as expected. In addition to the peak position, the  $I_G/I_{2D}$  is also sensitive to the carrier density change. The  $I_G/I_{2D}$  as a function of pH is plotted in Figure 2d, showing that  $I_G/I_{2D}$  decreases with pH increase. The reliability was confirmed using Student's *t* test with probability limits of p < 0.01. On the other hand,  $\Gamma_{2D}$ did not show pH dependence, as shown in Figure 2e, because  $\Gamma_{2D}$  is not sensitive to the carrier density change.<sup>13</sup> Both the down shift of  $G_{pos}$  and  $2D_{pos}$  and the decrease in  $I_G/I_{2D}$  are attributed to decrease in the carrier density in graphene.<sup>13</sup> At high pH, deprotonated ions are adsorbed on graphene,<sup>2,14</sup> which should lead to either increase in holes or decrease in electrons in graphene. As the Raman data indicate carrier density decrease at high pH, we concluded that the graphene films were n-doped in our system, where the graphene films were electrically floated under the buffer solution.

To provide further insights, we proved the reversible nature of the carrier density modulation by pH change. A buffer solution on the graphene films was replaced from pH = 4, 7, 9, 7, to 4, sequentially, and Raman images of the graphene films were taken at each condition (Figure S2). Figure 3 presents the transient response of the averaged  $G_{pos}$  against pH change. Averaged  $G_{pos}$  was calculated from the Raman images in Figure S2. The result shows that  $G_{\text{pos}}$  is monotonically down-shifted (up-shifted) with increasing (decreasing) pH, although there is a small offset shift probably due to insufficient rinsing such that some buffer solution residue remained. The validity was confirmed using Student's t test with probability limit of p <0.01. The tests show that the  $G_{pos}$  at different pHs is statistically different from each other. It is also found that  $2D_{pos}$  is monotonically down-shifted with increasing pH (Figure S3). The sensitivity of  $2D_{pos}$  against pH change is smaller than that of  $G_{pos}$ , in agreement with the previous report<sup>13</sup> showing that 2D<sub>pos</sub> against carrier density modulation is less sensitive compared to  $G_{pos}$ . Figure 3 also indicates that the  $G_{pos}$  instantly follows the pH change, which may lead to real-time monitoring of chemical dopants on graphene.



**Figure 2.** (a) Raman spectra of graphene taken at pH = 4 (red), 7 (green), and 9 (blue). (b, c) The enlarged spectra of G-band region (b) and 2D-band region (c) in (a). (d, e)  $I_G/I_{2D}$  (d) and  $\Gamma_{2D}$  (e) as a function of pH. The asterisk in (d) indicates the values are statistically different (p < 0.01). Each point represents the average value throughout the Raman images (Figure S1). The error bar represents the standard deviation.



**Figure 3.** Transient response of  $G_{\text{pos}}$  against pH change. Each dot represents the average value from the Raman images in Figure S2. The error bar represents the standard deviation.

Visualization of Charged Molecular Distribution via Raman Imaging of Graphene Films. Finally, we demonstrated a detection of locally modulated carrier density via Raman microscopy using charged microbeads. Negatively charged polystyrene (PS) beads (1  $\mu$ m diameter, surface density of COOH ~2 × 10<sup>13</sup> cm<sup>-2</sup>, micromod GmbH) were dispersed in a buffer solution at pH = 7. At pH = 7, the  $\zeta$ potential of the beads was about -30 mV and thus were negatively charged in the solution. Figure 4a shows a Raman image constructed by  $I_{G}$ . The PS beads were clearly observed as bright spots in Figure 4a, proving that our method has a high spatial resolution (down to 1  $\mu$ m). The Raman image constructed by  $G_{pos}$  is also shown in Figure 4b. The image shows that  $G_{pos}$  where the PS beads are located is lower than that without PS beads. The down shift indicates that the negatively charged beads moved electrons away in the graphene, where beads were in contact. Typical Raman spectra taken from the area with and without casted PS beads are shown in Figure 4c. A peak at 3160 cm<sup>-1</sup> is assigned to aromatic CH vibration in PS beads,<sup>15</sup> confirming that Raman signal from the beads was detectable even on graphene. It is interesting to note that the intensity ratio of the G-band to Dband  $(I_{\rm C}/I_{\rm D})$  with PS beads (~1.64) was smaller than that without PS beads ( $\sim$ 2.67). This result also supports that the PS beads locally modulate carrier density in graphene, resulting in the local scatters. The averaged  $G_{pos}$  and  $2D_{pos}$  with beads area are calculated to be 1594.7 and 2689.4, respectively, and the averaged  $G_{pos}$  and  $2D_{pos}$  without beads area are calculated to be 1595.8 and 2690.3, respectively.  $G_{pos}$  is correlated to its electron density  $n_e$  through eq  $1^{13,16}$ 

$$n_{\rm e} = \left(\frac{21\Delta G_{\rm pos} + 75}{\hbar |v_{\rm F}| \sqrt{\pi}}\right)^2 \tag{1}$$

where  $\Delta G_{\text{pos}}$  is the frequency shift from  $G_{\text{pos}}^{0} = 1581 \text{ cm}^{-113}$  of graphene where  $n_{\text{e}}$  is equal to hole density  $(n_{\text{h}})$  and  $\nu_{\text{F}} = 1.1 \times 10^{6} \text{ ms}^{-1}$  is the Fermi velocity.<sup>17,18</sup> By substituting  $\Delta G_{\text{pos}}$  in eq 1, the  $n_{\text{e}}$  can be calculated to be  $8.0 \times 10^{12} \text{ cm}^{-2}$  for graphene with beads and  $9.0 \times 10^{12} \text{ cm}^{-2}$  without beads, yielding a decrease in  $n_{\text{e}}$  of  $1.0 \times 10^{12} \text{ cm}^{-2}$  by external potential of the beads. In addition to a decrease in  $n_{e'}$  mechanical strain may also attribute to the peak shifts of  $G_{\text{pos}}$  and  $2D_{\text{pos}}$ . According to the previous reports,  $^{13,19-21}$  the peaks were up-shifted along a slope of  $\Delta 2D_{\text{pos}}/\Delta G_{\text{pos}} = 2.2$  under tensile strain and up-shifted along a slope of  $\Delta 2D_{\text{pos}}/\Delta G_{\text{pos}}$  = 0.2 under increase in  $n_{e}$ . Therefore, the peak shifts ( $\Delta G_{\text{pos}}, \Delta 2D_{\text{pos}}$ ) can be decomposed into the contribution of  $n_{\text{e}}$  modulation and mechanical strain. It was found that the contribution of  $n_{e}$  decrease to the peak shifts of  $(-1.1, -0.9 \text{ cm}^{-1})$  was 69% and the rest was induced by compressive strain.

To ensure that the peak shifts were truly caused by the charge of the beads, we carried out a control experiment using positively charged beads. Positively charged PS beads (1  $\mu$ m diameter, surface density of  $NR_3^+ \sim 2 \times 10^{13} \text{ cm}^{-2}$ , micromod GmbH) were dispersed in a buffer solution at pH = 4, where the  $\zeta$  potential of the beads was about +32 mV. Raman images constructed by  $I_G$  and by  $G_{pos}$  are shown in Figure 5a,b. The image in Figure 5b shows that  $G_{pos}$  where the PS beads are located is higher than that without PS beads, as opposed to the data in Figure 4. The scatter plot in Figure 5c also clearly shows that both  $G_{\text{pos}}$  and  $2D_{\text{pos}}$  are shifted toward high frequency with beads. From these results, we confirmed that the peak shifts were truly related to local increase/decrease in  $n_{\rm e}$  due to the charged beads. In addition, we can conclude that Raman microscopy can visualize the locally modulated  $n_{\rm e}$  of graphene. The averaged  $G_{pos}$  and  $2D_{pos}$  with beads are calculated to be 1596.5 and 2691.6, respectively. Also, the averaged  $G_{\rm pos}$  and  $2D_{pos}$  without beads are calculated to be 1595.1 and 2689.1, respectively. Therefore, using eq 1, we found that the  $n_e$  with beads was higher by  $1.4 \times 10^{12}$  cm<sup>-2</sup> than that without beads. This value is close to that obtained using negatively charged beads, although the sign is opposite. This result is reasonable as



**Figure 4.** (a, b) Raman images of the graphene films and negatively charged polystyrene (PS) beads constructed by  $I_G$  (a) and  $G_{pos}$  (b). The scale bar is 2  $\mu$ m. (c–e) Typical Raman spectra with beads (blue) and without beads (red) (c) and the enlarged spectra in G-band region (d) and 2D-band region (e).



Figure 5. (a–c) Raman images of the graphene films and positively charged polystyrene (PS) beads constructed by  $I_{\rm G}$  (a) and  $G_{\rm pos}$  (b). The scale bar is 2  $\mu$ m. (c) Scatter plot of  $G_{\rm pos}$  vs 2D<sub>pos</sub>. The blue dots and the red dots represent the data from spectra with beads and without beads, respectively. The averaged  $G_{\rm pos}$  and 2D<sub>pos</sub> are also plotted as diamond marks in (c). The error bar represents standard deviation.

the absolute values of the  $\zeta$  potential of the beads are close with each other (-30 and +32 mV). The vector decomposition into

mechanical strain and  $n_e$  modulation was also applied for the peak shifts (1.4, 2.5 cm<sup>-1</sup>). It was found that the contribution of  $n_e$  increase to the peak shifts was 20% and the rest was induced by tensile strain. The reason why the mechanical strain estimated from ( $\Delta G_{\text{pos}}$ ,  $\Delta 2D_{\text{pos}}$ ) was in opposite direction between positively and negatively charged beads may be due to the limited accuracy of  $\Delta 2D_{\text{pos}}$ , and this issue will be addressed in the future study.

## CONCLUSIONS

In conclusion, we have measured the  $n_e$  change of graphene via Raman microscopy, using the 2D nature and high sensitivity of graphene. Both  $G_{pos}$  and  $2D_{pos}$  shifted responding to changes in pH. We have also found the locally modulated  $n_e$  by charged PS microbeads. The charged molecular distribution on graphene was imaged with a spatial resolution of submicrometer. The sensitivity relies on the Debye length in an analogous fashion of G-FET biosensors.<sup>22</sup> We used buffer solutions of concentrations of 50 mM (pH = 4) and 25 mM (pH = 7), and so the Debye length is estimated to be 1 and 2 nm, respectively. This means that only charged molecules within the Debye length (<2 nm) will affect the local carrier density modulation, even though the diameter of the charged beads is 1  $\mu$ m. Even in this condition, the results show that the Raman peaks were shifted by  $\sim 1$  cm<sup>-1</sup>, which represents that our method exhibits very high sensitivity. To further improve the sensitivity, some superresolution techniques, including structured illumination Raman microscopy<sup>23</sup> or tip-enhanced Raman microscopy,<sup>24</sup> may be applied to our method. In addition, hexagonal boron nitride may be applied to suppress inhomogeneous charged impurities underneath graphene films,<sup>12</sup> which may also lead to further enhancement in the sensitivity of our method. Because our method is a noncontact, nondestructive, and label-free measurement and is applicable to a liquid environment, it is possible to combine the system with G-FET biosensors, which will expand the application range of G-FET. Furthermore, although Raman spectroscopy generally provides no information regarding charge state of target molecules, we can find out qualitative information by putting them on a graphene imaging platform. Our technique described here, thus, opens the door to a more versatile sensing platform for biomolecules and biological functions.

# MATERIALS AND METHODS

Graphene films were grown on Cu foils by chemical vapor deposition. To transfer the graphene films to  $Si/SiO_2$  substrates, a poly(methyl methacrylate) (PMMA) solution was coated and subsequently the Cu foils were etched away using an ammonium persulfate solution. PMMA/graphene films were transferred onto Si/SiO<sub>2</sub> substrates and finally PMMA was dissolved using acetone. The obtained graphene films were studied.

Raman analysis of graphene films was carried out using Raman microscopy (Raman-11, Nanophoton Corp.). A laser beam emitting at 532 nm was used for the excitation. The laser beam was focused by an objective lens ( $100\times$ , NA 0.9, Nikon) onto the graphene films. The resolution of the spectrometer was about 4 cm<sup>-1</sup> with the grating of 600 line/mm. It should be noted that the inaccuracy of determination of the peak position is much smaller than the resolution when the spectral shape is known. Laser intensity and exposure time were optimized to take Raman spectra with sufficient signal to noise ratio.

# ASSOCIATED CONTENT

#### **S** Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.7b02008.

Details about Raman images of graphene films and pHdependent Raman spectra and images of graphene (PDF)

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#### **Author Contributions**

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

## Notes

The authors declare no competing financial interest.

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