

Original Article

Add-On therapy with Chinese herb medicine Bo-Er-Ning capsule (BENC) improves outcomes of gastric cancer patients: a randomized clinical trial followed with bioinformatics-assisted mechanism study

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Abstract: As a Chinese herb medicine (CHM), Bo-Er-Ning capsule (BENC) has been approved in China for adjuvant treatment of cancer, but the particular therapeutic effect of BENC on gastric cancer (GC) has yet to be evaluated. In this study, we implemented an efficacy-driven approach by directly starting the study with a randomized clinical trial to assess the add-on therapeutic effect of BENC on advanced GC patients. Our results showed that the addition of BENC to chemotherapy resulted in higher Karnofsky performance scores and better 3-year overall survival, compared to those treated with the conventional chemotherapy regimen. Subsequently, we explored the mechanism of BENC action on GC cells in the assistance of BATMAN-TCM, the first online bioinformatics analysis tool designed especially for the mechanism study of CHM, by which we identified 263 candidate protein targets of BENC involved in GC treatment. The further enrichment analysis suggested that BENC treatment affected a diversity of biological processes of GC cells, such as cell proliferation, cell cycle and apoptosis, which were further validated in the following *in vitro* and *in vivo* assays, indicating such a bioinformatics-assisted approach was feasible and powerful to CHM mechanism study. Thus, as exemplified by BENC, we provided an efficacy-driven and bioinformatics-assisted strategy for CHM research, which may help promote the discovery and application of novel CHM drugs on patients with refractory diseases.

Keywords: Bo-Er-Ning capsule, gastric cancer, Chinese herb medicine, efficacy-driven approach

Introduction

Gastric cancer (GC) is the fifth most common cancer and the second most frequent cancer-related death worldwide. More than 900,000 new cases of GC are diagnosed every year and over 70% of new cases occur in developing countries, accounting for half of the caseloads of the world [1, 2]. Unfortunately, approximately two thirds of GC patients are diagnosed at advanced stages, resulting in disappointing outcomes. The median survival time of GC patients is only about one year, with an 5-year overall survival rate ranging 5-20%, primarily due to a high recurrence or metastasis rate [3]. Current standard interventions for GC have not

changed for the last decades. Thus, it is an urgent need to develop novel pharmaceuticals to improve the outcomes of GC patients.

So far, over two thirds of chemo-drugs are derived from natural products since launching chemotherapy for cancer treatment over half a century ago [4]. Chinese herb medicine (CHM) is characterized by the application of a blend of several natural sourced ingredients to treat illnesses based on patients' symptoms. As a key component of traditional Chinese medicine, CHM has a long history and distinguished clinical effect on various diseases, including cancer [5]. Nowadays, CHM research mainly focuses on identifying the active substances and inves-

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Table 1. General data of two groups (cases)

Category	Characteristics	Control group	Trial group
Gender	Male	24	36
	Female	30	22
Histological Type (WHO)	Moderate or poorly differentiated and undifferentiated adenocarcinoma	26	30
	Mucinous adenocarcinoma	6	4
	Tubular adenocarcinoma	6	4
	Signet-ring cell carcinoma	4	6
	Well differentiated adenocarcinoma	12	14
Median Age/Year	~	58.5	56.6

tigating mechanisms of action, under the influence of western biomedical research mode. However, though tremendous efforts have been made, this mechanism-centered approach has led to little progress in developing new CHM formula/drugs in the last decades. Thus, to facilitate the development and clinical application of novel CHM formula/drugs, an efficacy-driven approach for CHM research has recently been proposed [6, 7]. Efficacy-driven approach starts the research with a randomized clinical trial to directly evaluate the CHM efficacy in humans, and conducts the mechanism studies *in vitro* or in animals after the clinical efficacy has been validated. Ethically, efficacy-driven approach is acceptable as some folk CHM recipes have been used in China as well as in other countries for many years, and new CHM drugs will continue to be invented and directly applied to patients in places where traditional Chinese medicine is officially recognized [6, 8].

Chinese medicine characterizes cancer as blood stasis, heat toxins, knotted accumulation and formation of swellings and lumps [9]. Bo-Er-Ning capsule (BENC), a patent CHM formula approved by the State Food and Drug Administration (SFDA) of China (Z20054459), is used for the adjuvant treatment of cancer due to its essences of heat-clearing and detoxifying as well as its functions in activating blood flow to dissipate blood stasis. BENC is composed of a series of key pharmaceutical ingredients, including *Radix Astragali Seu Hedysari Praeparata*, *Ligustrum Lucidum*, *Tulipa Edulis*, *Portulaca Oleracea*, *Rhizoma Paridis*, *Solanum Nigrum*, *Fructus Perillae*, *Gallus Gallus Domesticus*, *Rheum Officinale*, *Dryobalanops Aromatica* and *Bombyx Batryticatus* (See [Supplementary Table 1](#) for details). Previous

study has revealed that BENC exhibited desirable pharmacological effects on preventing recurrence of bladder cancer [10]. Moreover, when applied with HEP regimen (hydroxycarbonythecine, etoposide, cisplatin), BENC showed a synergistic anti-tumor effect on advanced non-small cell lung cancer [11]. However, the therapeutic effect of BENC on GC has not been evaluated.

In this study, we started with a randomized clinical trial and confirmed the add-on therapeutic efficacy of BENC on advanced GC patients. Next, in the assistance of bioinformatic analysis tools, we conducted a virtual study to explore the mechanism of BENC action on GC cells. Finally and importantly, we further validated the predicted cellular functional and related signaling impacts of BENC on GC cells using *in vitro* and *in vivo* assays. Thus, taking advantage of the efficacy-driven approach and recent advent in bioinformatics, we provided an efficacy-driven and bioinformatics-assisted strategy for CHM research to facilitate the discovery and application of novel CHM formula/drugs on cancer patients.

Material and methods

Patients and clinical data

From June 2008 to June 2012, 112 patients with advanced GC treated by FP regimen (Tegafur + Cisplatin) in Zhangqiu People's Hospital of Shandong were enrolled for this clinical study by the 3th author and randomly divided into two groups by the 4th author. Among those, 58 patients received BENC accompanying FP regimen (trial group), while 54 patients received FP regimen alone (control group). The general clinical data of two groups were in

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Table 1. The eligibility criteria were: (1) age from 25 to 75 years; (2) all patients were pathologically confirmed for stage IV or postoperative recurrence and metastasis GC; (3) Karnofsky performance score (KPS) \geq 80; (4) no previous anticancer therapy; (5) no dysfunction of heart, liver, lung and kidney. All study procedures were approved by the Ethical Committee of Zhangqiu People's Hospital of Shandong. All patients signed the informed consents, and the study conformed to the ethical principles set forth by the Declaration of Helsinki.

Treatment regimen for control group: Tegafur 40~60 mg, oral administration, two times a day, from day 1 to day 14 and Cisplatin 40 mg, intravenous infusion administration, from day 1 to day 3. Treatment regimen for trial group: BENC 4 capsules, take orally three times a day (eating with warm water half an hour after a meal) for 14 days (from day 1 to day 14), accompanying Tegafur and Cisplatin treatment (the same drug dosage and usage as the control group). Both groups were evaluated after 3 cycles (both groups take 21 days as one cycle) of chemotherapy (14 days of treatment and 7 days of rest). After 3 cycles of chemotherapy, all patients were observed for appetite, body weight, and Karnofsky performance score (KPS).

Patients were followed up by clinic visit, phone, or mail at least once 6 months starting from day 1 after the first hospitalization. Study endpoints took the overall survival (OS) as the standard. The survival status was obtained from medical records or by direct follow up via telephone or mail. Due to the serious loss of the follow-up visit, the number of control group was 20, and the trial group was 23. Death was defined as GC related death.

Bioinformatics analysis

The potential protein targets for BENC were predicted using the online BATMAN-TCM server [12] and the known GC related targets were obtained from Gene Expression Omnibus (GEO) database (See [Supplementary Table 2](#) for details). We chose four gene expression chips (GSE33429; GSE63089; GSE13195; GSE33335) derived from human GC and adjacent normal tissue.

We first constructed an interaction network for predicted putative drug targets of BENC and

the known GC related targets based on the data obtained from the Cytoscape plugin Bisogenet. The network was visualized using Cytoscape (Version 3.2.1) [13]. Next, the topology parameters of each node in the overlap network were calculated using a Cytoscape plugin CytoNCA. The node with a score twice bigger than the median of "Degree centrality" (DC) is considered important and appeared in the network. We finally performed enrichment analysis of the potentially significant targets using ClueGO, a Cytoscape plugin that visualizes non-redundant biological terms for large clusters of genes in a functionally grouped interaction network [14]. These candidates were divided into two types, GO biological process and KEGG signaling pathways.

Cell culture and reagents

Human GC SGC-7901 and BGC-823 cells were obtained from the China Infrastructure of Cell Line Resources (School of Basic Medicine Peking Union Medical College, China) and were maintained in RPMI-1640 medium supplemented with 10% (v/v) FBS and 100 U/ml streptomycin/penicillin in the humidified atmosphere of 5% CO₂ at 37°C. RPMI-1640 medium and fetal bovine serum (FBS) were obtained from GIBCO (USA). Annexin-V FITC apoptosis detection kit (#556547) and FITC BrdU Flow kit-PartA (#559619) were purchased from BD Biosciences Pharmingen (USA). Bcl-2 rabbit polyclonal antibody, Bax rabbit polyclonal antibody, Caspase 9/p35/p10 rabbit polyclonal antibody, E-cadherin rabbit polyclonal antibody, Vimentin rabbit polyclonal antibody and Beta-actin mouse monoclonal antibody were bought from Proteintech. BENC were obtained from Shenzhen Jian An Pharmaceutical Limited.

Cellular functional assays

The cell proliferation was assessed by the CCK-8 assay, in which 10 μ l of CCK-8 was added to each well, and the cells were incubated for an additional two hours before the absorbance value was measured at a wavelength of 450 nm using an automated microplate reader. Cell invasion and migration assays were performed in Transwell chambers, with or without matrigel on the upper chamber respectively. Cell colony formation assay was carried out and cell colonies were counted after stained with 0.1% crystal violet. Apoptosis detection, cell

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Table 2. Changes in appetite and body weight of patients' in two groups (cases)

Group	Appetite				Body weight			
	Increase	Decrease	Unchange	Effective rate/%	Increase	Decrease	Unchange	Effective rate/%
Trial Group	45	3	10	77.6	35	10	13	60.3
Control Group	6	30	18	11.1	5	25	24	9.2

Table 3. Karnofsky score in two groups (cases)

Group	Cases	Scores Decrease	Scores Unchange	Add 10 Points	Add 20 Points	Effective rate/%
Trial Group	58	8	12	26	12	65.5
Control Group	54	23	19	11	1	22.2

cycle and Western blot analysis were performed according to the manufacturer's instructions.

Xenograft mouse work

SGC-7901 cells (2×10^5 /ml) treated with BENC ($200 \mu\text{g mL}^{-1}$) were inoculated subcutaneously into the left and right flank of 8 NOD/SCID mice using 1-ml needles. Two weeks later, the tumor volumes were measured once every two or three days using the following formula: long diameter \times (short diameter)²/2. On day 16, mice were sacrificed and the tumor tissues were weighed. None of mice died during the experiments.

Immunohistochemistry

The slides of tumor tissue sections were disposed of deparaffinization and antigen unmasking, and were then incubated with the antibody against Ki-67 (Abcam, UK) at 4°C overnight. After washing with PBS, the slides were incubated with Polymer Helper and Poly peroxidase-anti-mouse/rabbit IgG (PV-9000, ORIGENE, China), followed by further incubation with diaminobenzidine (DAB). The tissue sections were mounted after counterstained with hematoxylin.

Statistical analysis

The results were analyzed using SPSS17.0 software (USA). Data were represented as the mean \pm standard deviation. The differences expressed were using the Student's t-test.

Results

BENC add-on therapy improved the outcomes of advanced GC patients

We firstly compared the add-on effect of BENC on the quality of life (QOL) between the trial and

control groups, as manifested by appetite, body weight as well as Karnofsky Performance Scores (KPS). As showed in **Table 2**, the appetite was greatly improved in the trial group (77.6%) compared to the control group (11.1%), which was in line with the body weight gaining rate in the trial group (60.3%) verse the control group (9.2%). The differences between two groups were statistically significant ($P < 0.01$).

Since the KPS has been widely recognized as a reliable predictor of survival in patients with various cancers [15, 16], we next assessed the KPS for both groups. Our result showed a significant increase of KPS in the trial group versus the control group (65.5% vs. 22.2%, **Table 3**). Finally, we used Kaplan-Meier analysis to evaluate the add-on therapy effect of BENC on advanced GC patients, and the result showed that the 3-year overall survival of the trial group (23.57 months) was greatly improved compared to the control group (18.75 months, **Figure 1**). The difference between two groups was statistically significant ($P < 0.05$). Together, these results indicated that the adjuvant therapy of BENC significantly improved the QOL and 3-year overall survival of the GC patients in the trial group.

BENC inhibited GC cells growth and decreased GC cells viability

It is well-known that CHM exhibits its maximum strength in treating various diseases in a manner of multi-mode of action [15]. Accordingly, the add-on therapeutic efficacy of BENC on GC patients could be attributed to multiple factors, such as modulation on patient's immune system, synergistic effect with the chemotherapy or direct inhibitory effect on GC cell growth and invasion. Thus, we next determined the role of BENC in GC cell growth and viability.

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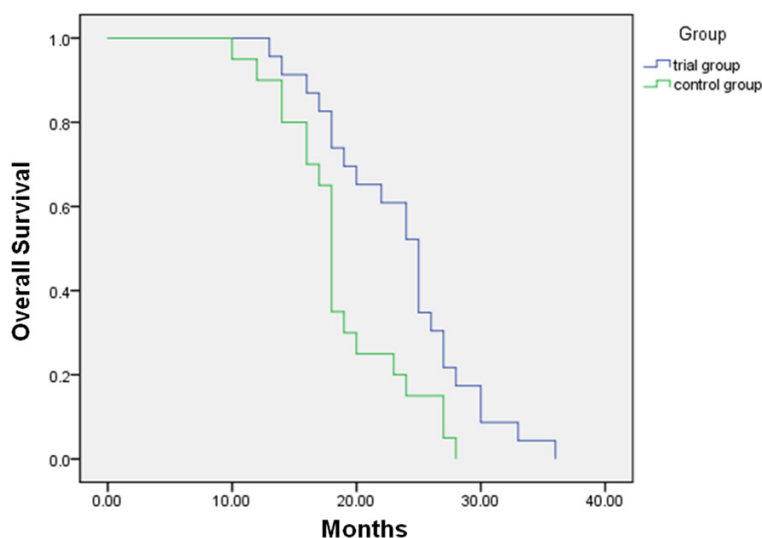


Figure 1. Kaplan-Meier analysis showed significantly improved overall survival of GC patients by add-on therapy with BENC ($P=0.01$).

The CCK-8 assay results showed that the growth of GC cells (SGC-7901 and BGC-823 cell lines) were significantly inhibited following BENC treatment in a time and dose-dependent manner (**Figure 2A**). Meanwhile, the viability of these cells were determined by the cell colony formation assay, showing the cell clonality of these cells were also decreased in a dose-dependent manner following BENC treatment for 24 hours (**Figure 2B**). These results demonstrated a direct suppressive role of BENC in GC cell growth and viability.

Identification and enrichment analysis of candidate targets for BENC against GC

Having confirmed the clinical therapeutic efficacy of BENC on GC patients was attributed to its direct inhibitory effects on GC cells, we next conducted an virtual study to explore the underlying mechanism involved in the anti-tumor activity of BENC on GC cells by using the online BATMAN-TCM server [12] and Gene Expression Omnibus (GEO) database.

Since the protein-protein interaction (PPI) networks are relevant to visualize the role of various key proteins in cancer, we constructed a putative target network for BENC, containing 2297 nodes and 52226 edges, and a known GC related target network, containing 3645 nodes and 92722 edges, using the Bisogenet, a plugin for Cytoscape (3.2.1). Furthermore, to clarify the pharmacological mechanisms of

BENC against GC, we intersected the two networks and thus obtained 1154 nodes and 31722 edges. Then we calculated the topology parameters of each node in the overlap network by using a Cytoscape plugin CytoNCA. Referring to a previous method [16], we identified nodes with degrees that were bigger than twice the median degree of whole nodes as the significant targets. As thus, we constructed a significant target network that contained 263 nodes and 9845 edges (**Figure 3A**).

Finally, we performed an enrichment analysis of the identified 263 candidate targets for BENC against

GC cells using ClueGO, a Cytoscape plugin [14], and divided these candidates into GO biological process and KEGG signaling pathway, respectively. In this analysis, a wide range of biological processes were disturbed in response to BENC treatment, such as cell growth, cell cycle, cell death and apoptosis (**Figure 3B**). Meanwhile, the impaired signaling pathways included viral carcinogenesis, transcriptional misregulation in cancer and so on (**Figure 3C**). Thus, we reasonably speculated that the potential mechanism of BENC acting against GC cells was related to cell proliferation, cell cycle and apoptosis.

BENC promoted apoptosis and arrested GC cells in G2/M phase

To validate the above *in silico* analysis result, we carried out a series of cellular functional assays, such as apoptosis assay and cell cycle analysis, by using different GC cell lines. Flow cytometry analysis showed that the apoptotic population that positively stained with annexin V-FITC was significantly increased upon BENC treatment in a dose-dependent manner (**Figure 4A** and **4B**). Consistently, immunoblotting results showed that BENC promoted accumulation of the pro-apoptosis proteins Bax and cleaved Caspase-9, whereas the level of anti-apoptosis protein Bcl-2 was down-regulated by BENC in a dose-dependent manner (**Figure 4C**). Furthermore, to determine whether BENC

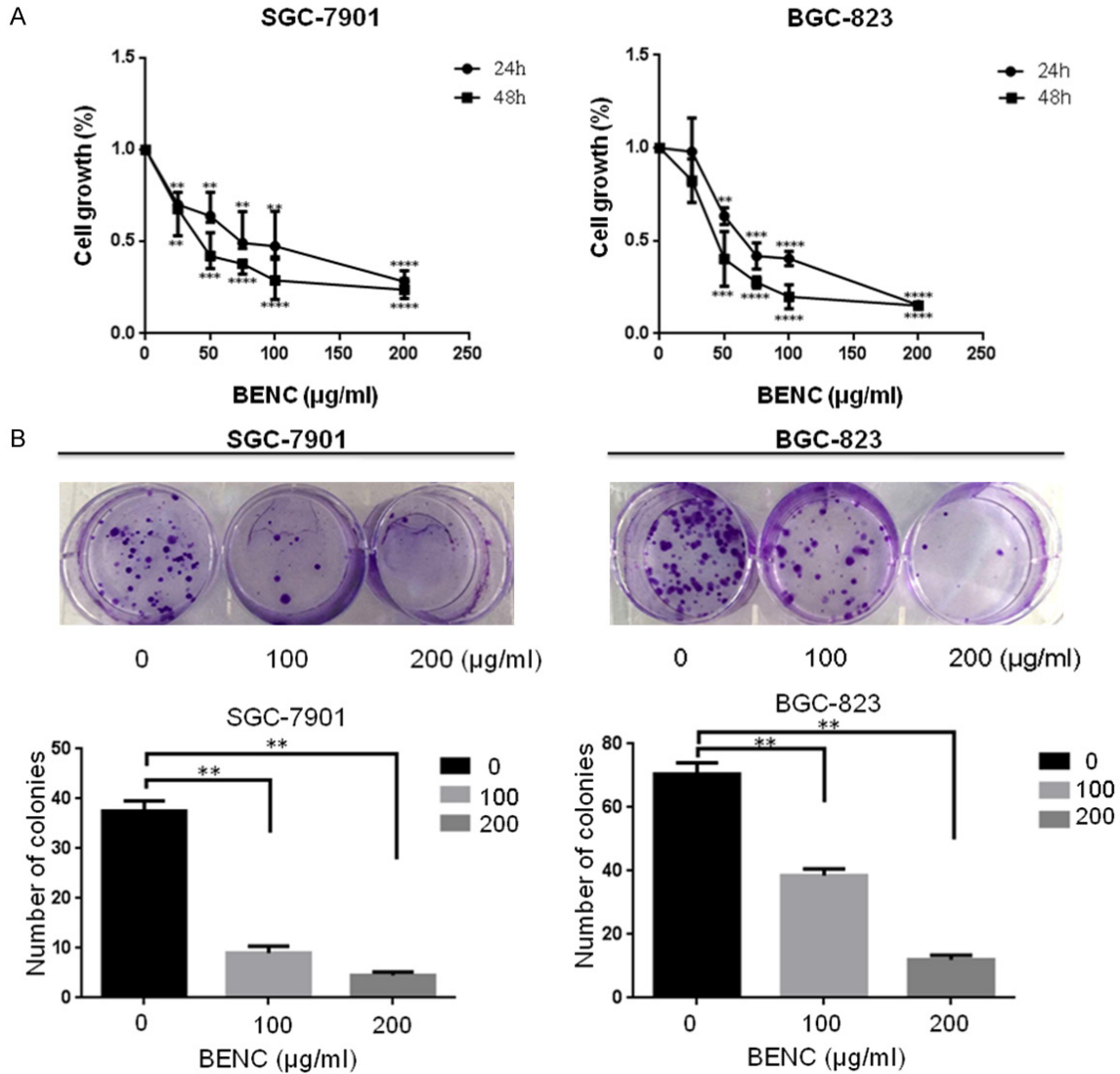


Figure 2. BENC inhibited GC cells growth and decreased GC cells viability. A. CCK-8 assay was performed to measure cell growth rate at 24 h and 48 h after BENC treatment at different dosages. B. Cell colony formation assay was performed to measure cell clonality after BENC treatment for 24 h. ** $P < 0.01$ based on the Student t-test.

induces the disturbance of cell cycle progression, the BrdU-incorporated cell cycle analysis was performed in GC cells. The results showed that BENC treatment could dramatically decrease the proliferation rate of these cells, evidenced by reduced proportion of G0/G1 and S phase, while arresting the cells at G2/M phase (Figure 4D and 4E). Consistently, immunoblotting results showed that BENC promoted accumulation of the G2/M phase-specific marker Cyclin B1, while down-regulating the levels of Cyclin D1 and Cyclin E in both SGC-7901 and BGC-823 cells (Figure 4F). Together, these data indicated that BENC treatment

induced apoptosis, inhibition of proliferation and G2/M arrest in GC cells.

BENC suppressed migration and invasion of GC cells

Since the epithelial-mesenchymal transition (EMT) plays an important role in the course of tumor progression, we next determine the effect of BENC on EMT in GC cells. The transwell assay results revealed that both cell migration and cell invasion were inhibited by BENC in a dose-dependent manner (Figure 5A and 5B). This cellular phenotype might be associated

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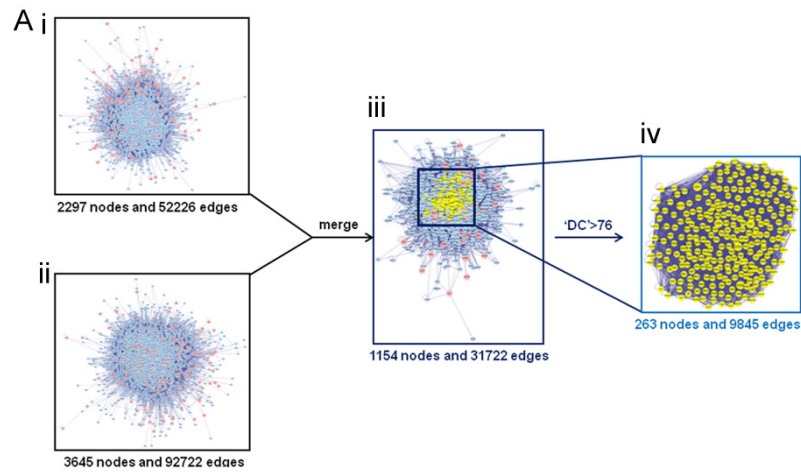
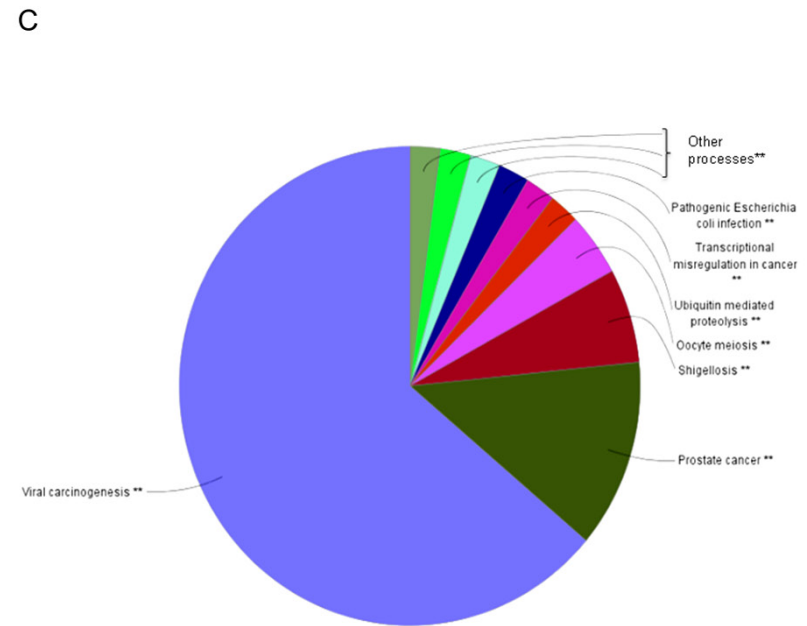
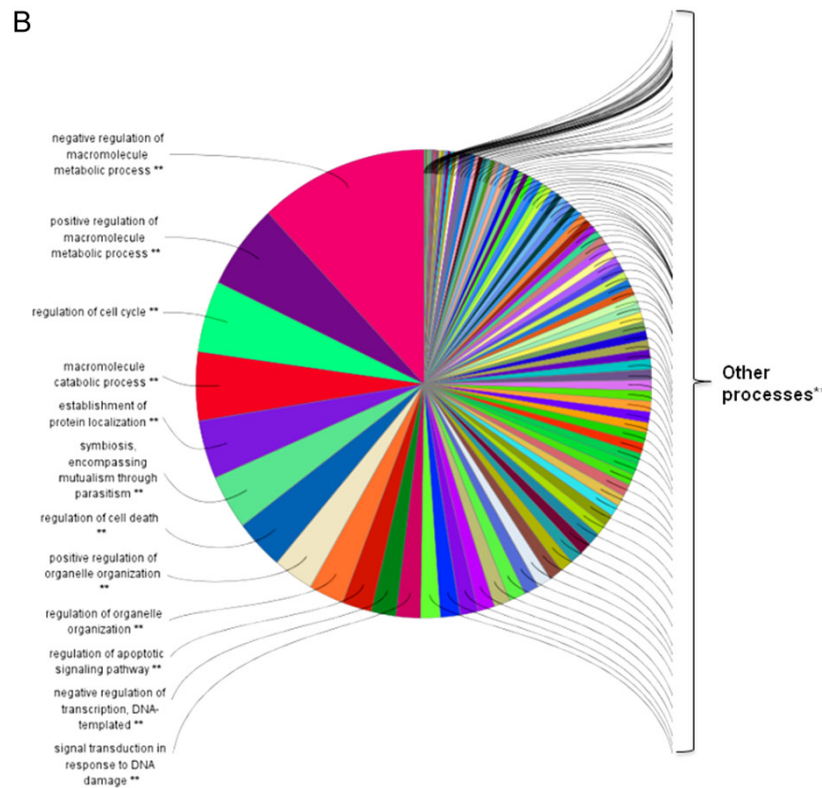
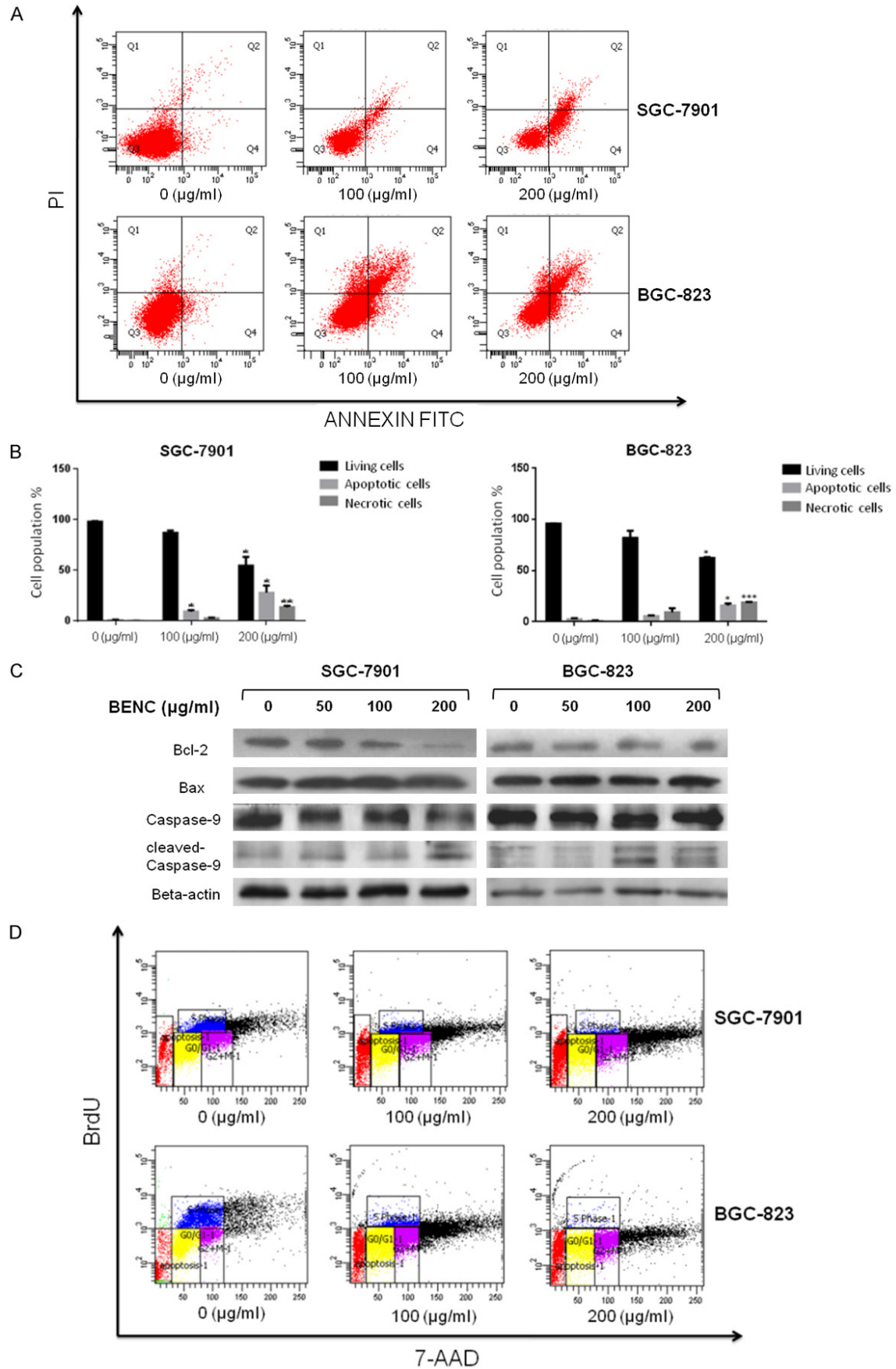


Figure 3. *In silico* identification and enrichment analysis of candidate targets for BENC against GC. A. (i) The interactive PPI network of BENC putative targets was made of 2297 nodes and 52226 edges. (ii) The interactive PPI network of GC related protein targets was composed of 3645 nodes and 92722 edges. (iii) The interactive PPI network of BENC putative targets and known GC related targets made of 1154 nodes and 31722 edges was shown. (iv) PPI network of significant proteins extracted from iii, in which 263 nodes and 9845 edges were included. B. Candidate significant targets were enriched in the representative biological processes ($P < 0.05$). C. Candidate significant targets were enriched in the representative signalling pathways ($P < 0.05$).



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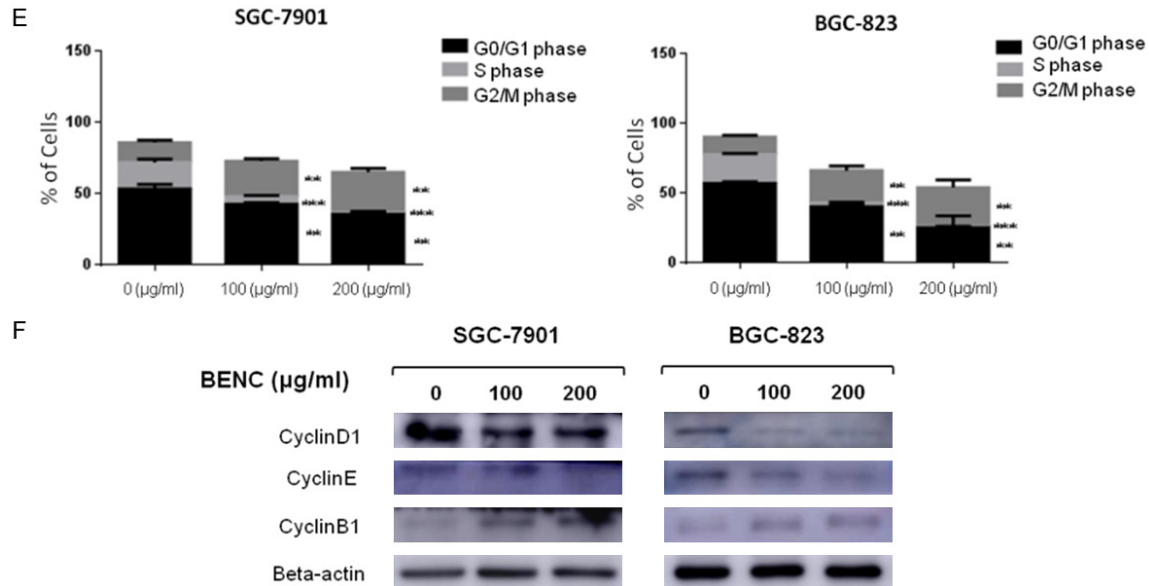


Figure 4. BENC treatment resulted in apoptosis and disturbance of cell cycle progression in GC Cells. A. Induction of apoptosis of SGC-7901 and BGC-823 cells after BENC treatment. Both cell lines were treated with BENC at different concentrations (0, 100, 200 µg/ml) for 24 h when apoptotic events was assessed by flow cytometry. B. Statistical analysis of the percentages of the apoptotic cells. C. SGC-7901 and BGC-823 cells were treated with BENC at different concentrations (0, 50, 100, 200 µg/ml) for 24 h. After proteins were extracted, the levels of Bcl-2, Bax, pro-caspase-9, cleaved-caspase-9 were analyzed by Western Blot. D. Cell cycle analysis of GC cells following BENC treatment (0, 100, 200 µg/ml) for 36 h by flow cytometry. E. Statistical analysis of the proportions of the cells at different phases. $**P < 0.01$ based on the Student t-test. F. SGC-7901 and BGC-823 cells were treated with BENC at different concentrations (0, 100, 200 µg/ml) for 36 h. After proteins were extracted, the levels of Cyclin D1, Cyclin E, Cyclin B1 were analyzed by Western Blot.

with the inhibitory effect of BENC on EMT process, evidenced by the expression patterns of EMT-related proteins in these cells in response to BENC treatment at different dosages (Figure 5C).

Anti-tumor efficacy of BENC *in vivo*

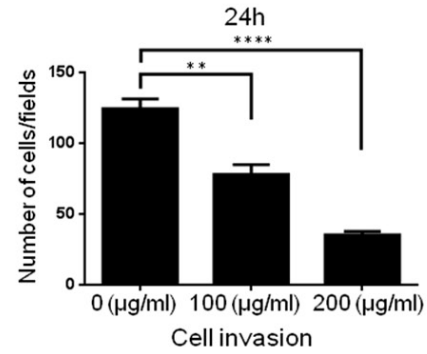
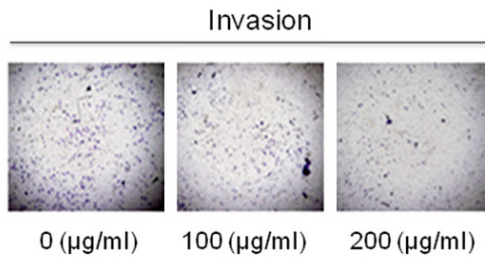
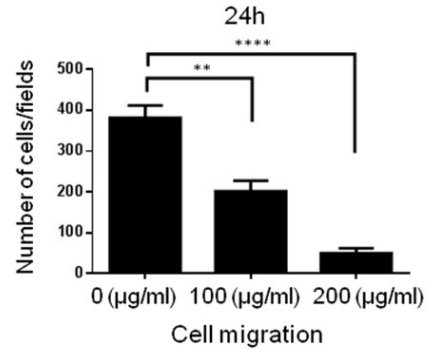
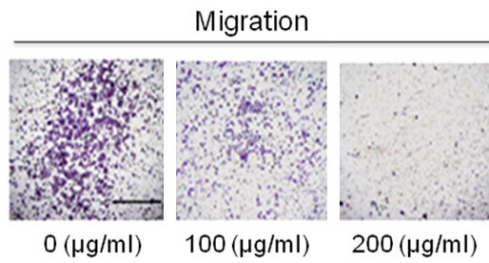
To investigate the direct tumor suppressive role of BENC *in vivo*, we further determined the effect of BENC on tumors xenografted onto immunodeficient mice. As shown in Figure 6A, the development of tumors were greatly suppressed by BENC treatment on these mice. By day 16, the tumor volume of BENC-treated group were approximately 7-fold smaller than the control group ($P < 0.05$), in line with the tumor weights, which showed a striking difference between two groups (Figure 6B-D). The subsequent immunohistochemistry analysis revealed that the number of Ki-67 positive cells was significantly decreased in BENC-treated tumors, compared to those in control tumors, indicating an anti-proliferative effect of BENC

on these tumors (Figure 6E and 6F). Thus, these data provided convincing evidence showing that BENC possesses direct anti-tumor activity *in vivo*.

Discussion

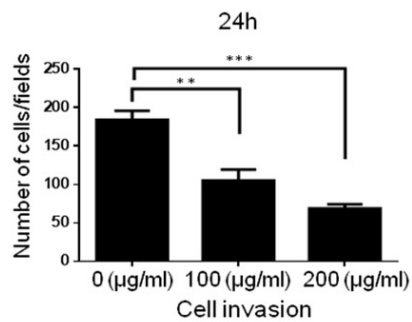
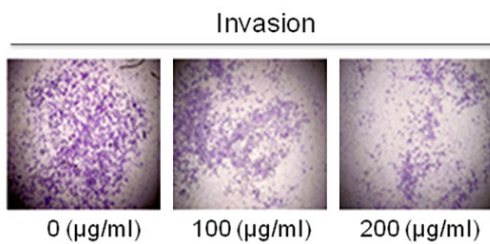
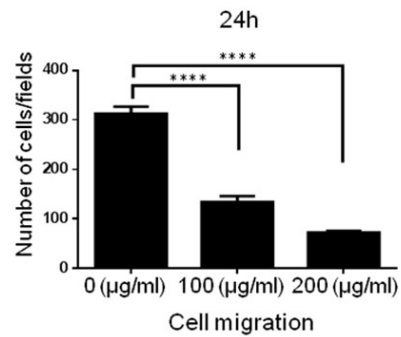
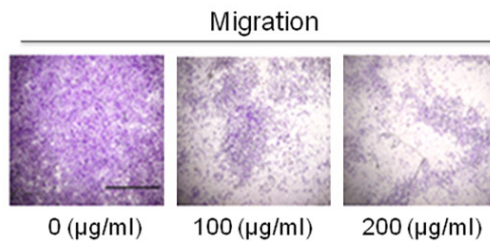
As an important resource for innovative drug discovery, CHM research has attracted great attention for many years [16, 17]. More and more new compounds with various therapeutic activities have been identified from CHM in recent years [18]. In the view of exploring novel drugs, an efficacy study is always beneficial no matter whether the result is positive or not, because if a drug is proved effective, its immediate application in clinical will benefit patients, whereas if the drug is not effective, stopping using it will save resource by avoiding unnecessary basic research. But from a patient's point of view, it is apparently not the best choice to try a new drug before its efficacy has been fully determined, even if the drug is a CHM that has been applied in clinical practice for a long time.

A



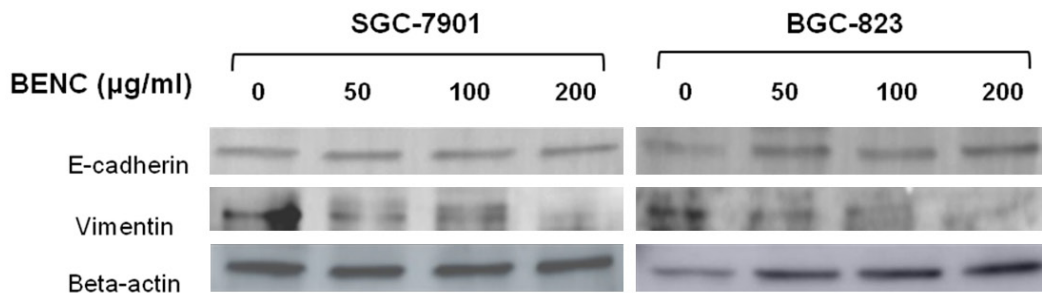
SGC-7901

B



BGC-823

C



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Figure 5. BENC suppressed migration and invasion of SGC-7901 and BGC-823 cells. A, B. Transwell assay was performed to measure cell migration and invasion at 24 h after BENC treatment. C. SGC-7901 and BGC-823 cells were treated with BENC (0, 50, 100, 200 $\mu\text{g}/\text{ml}$) for 24 h. Proteins were extracted, then E-cadherin and Vimentin levels were analyzed by Western Blot. $**P < 0.01$ based on the Student t-test.

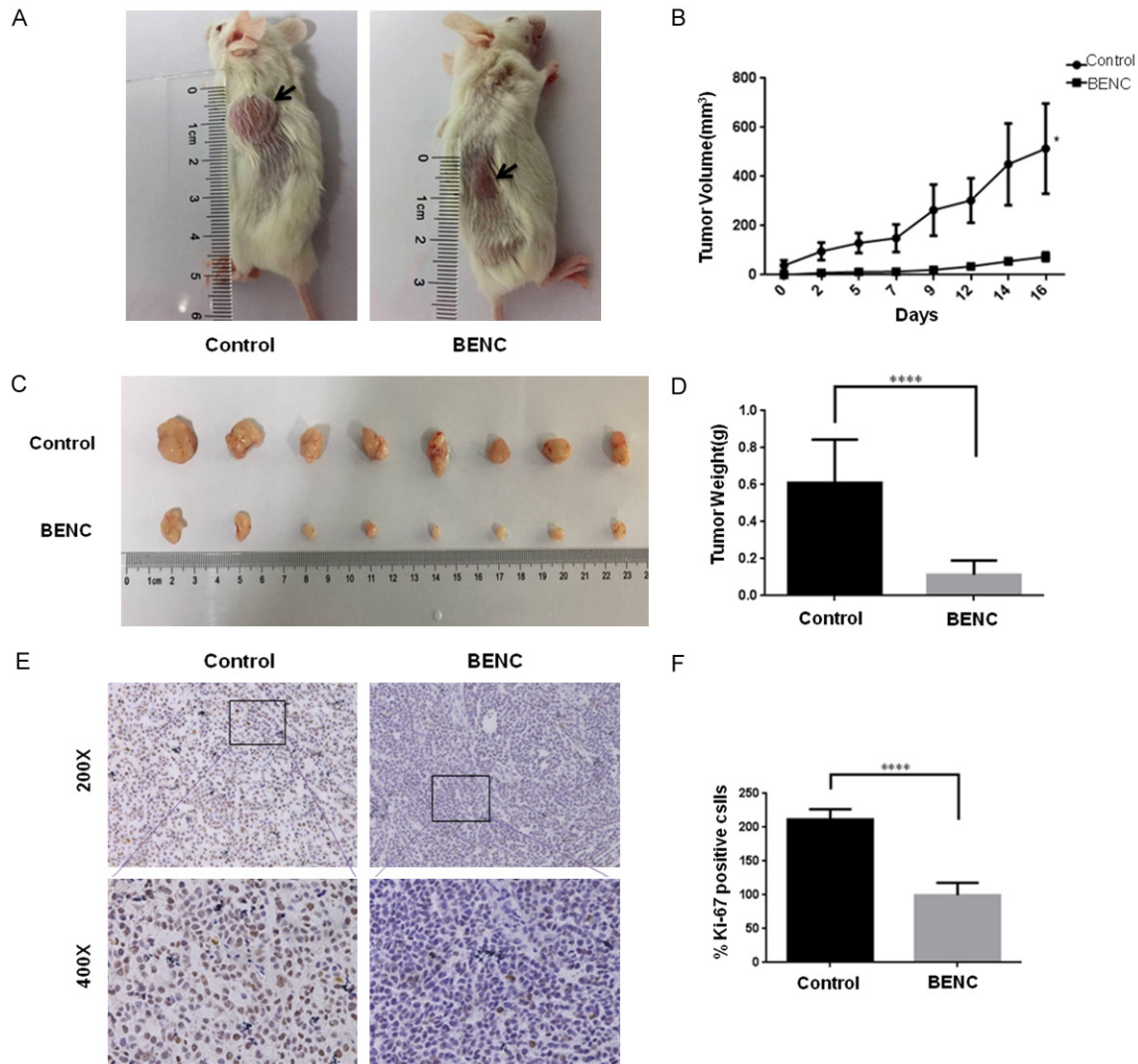


Figure 6. BENC suppressed development of the tumors on xenografted mice. A. The tumor masses inoculated on NOD/SCID mice were carefully checked every another day with naked eyes. B. The tumor volumes were measured and calculated once every two or three days. C, D. The tumors were resected and weighted on day 16. E. Immunohistochemistry staining for Ki-67 was performed by using the tumor slides from control and BENC-treated groups. F. Statistical analysis of the positive ratio of Ki-67 staining. $*P < 0.05$ based on the Student t-test.

Especially, for the patients who are suffering from serious diseases such as cancer, it is unethical to put them on a new drug to take the risk of failure in efficacy trial, as thus they may miss the optimal treatment window, resulting in serious consequences. Actually, surveys have found that most cancer patients were inclined to take the CHM on the basis of conventional

therapy [19]. Therefore, to evaluate the efficacy of BENC on GC, we conducted a clinical trial in advanced GC patients using BENC combined with FP regimen rather than in lieu of FP regimen. Application of this add-on therapy in the efficacy-driven approach took advantage of the efficacy study to promote the clinical use of BENC, but also precluding the ethical problems

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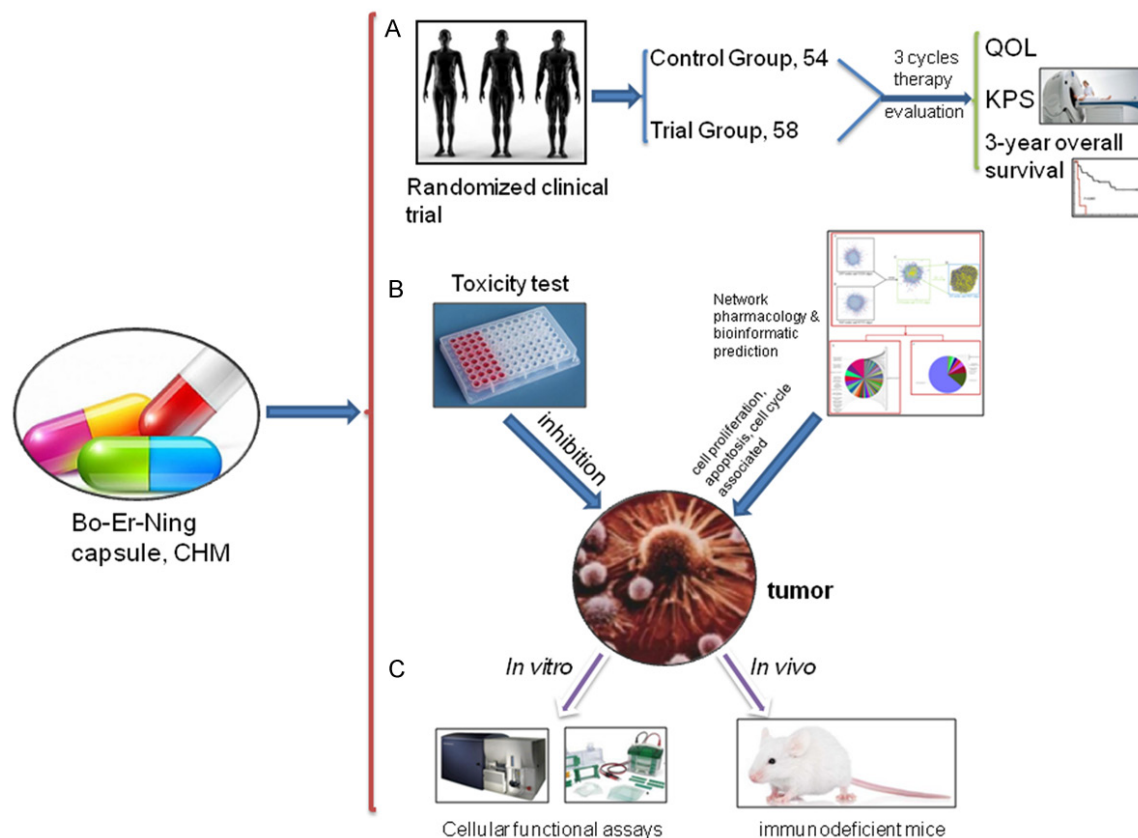


Figure 7. The overall schematic design of the study. A. Starting the study with a randomized clinical trial to assess the add-on therapeutic effect of BENC on GC patients. B. An virtual study was conducted to explore the mechanism of BENC action on GC cells in the assistance of bioinformatic analysis tools. C. Further validation of the predicted cellular functional and related signaling impacts of BENC on GC cells using *in vitro* and *in vivo* assays.

caused by the failure of the clinical trial. In this study, BENC add-on therapy significantly improved the outcomes of advanced GC patients, evidenced by higher Karnofsky scores and better 3-year overall survival. Importantly, the following cellular functional assays demonstrated that BENC exhibited its anti-tumor effect by directly suppressing the growth and decreasing the viability of GC cells. Thus, having validated the add-on therapeutic efficacy of BENC on GC cells, the clinical trial designed for GC patients treated with BENC combined with lower-dosed chemotherapy or the trial for patients treated with BENC alone warrants further investigation.

In an efficacy-driven approach, mechanism study is still important, though it is undertaken after the efficacy has been confirmed. Identification of the potential targets is a critical step to reveal the underlying mechanisms of CHM actions at the molecular level. However,

due to the complexity of the interaction between diverse ingredients of CHM and the networks of various diseases, it remains to be challenging to find the CHM targets by conventional methods [20]. Nowadays, with the advance in computational methods, many informatics methods such as bioinformatics and cheminformatics have been proposed to apply on the target prediction for CHM [21-23]. Several online databases for herbal ingredients and a number of compound-target interaction prediction methods have been recently developed [24-30]. Among these, BATMAN-TCM is the first online bioinformatics analysis tool designed especially for the mechanism study of TCM, which supports the comprehensive target prediction for all components of CHM and subsequent network pharmacology analyses [12].

Using BATMAN-TCM server and GEO database, we constructed and analyzed BENC and GC-related protein targets network, and finally

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identified 263 candidate protein targets for BENC involved in GC treatment. The further enrichment analysis using ClueGO plugin predicted that BENC inhibited GC cells growth by affecting the biological processes such as cell cycle and apoptosis. Indeed, the subsequent flow cytometry analysis of BrdU incorporation and Annexin-V staining assays showed a G2/M arrest and significant increased apoptotic events upon BENC treatment. Furthermore, the trans-well assays revealed that BENC could suppress cell migration and invasion, associated with its inhibitory effect on EMT process. The anti-tumor activity of BENC was further determined in immunodeficient mice, in which the growth of xenografted tumors was significantly suppressed by BENC treatment. Thus, using *in vitro* and *in vivo* assays, we validated the anti-tumor effects of BENC on GC cells according to the target prediction results by BATMAN-TCM, indicating such a bioinformatics-guided approach was feasible and powerful to CHM mechanism study.

The signaling pathway enrichment was conducted using KEGG PATHWAY. Notably, the viral carcinogenesis related pathways accounted for 2/3 of the total hits. It has been well documented that many signaling pathways are involved in viral carcinogenesis. For instance, the NF- κ B signaling pathway plays important roles in HBV-associated hepatocellular carcinoma (HCC) and HPV-associated cervical carcinoma [31, 32]. As a key marker for NF- κ B signaling pathway, VCAM1 has been identified as a candidate protein target for BENC in this study. Mutant VCAM1 could confer growth and infiltration capacity to HCC cells and down-regulation of VCAM1 inhibits cancer cell adhesion to endothelial cells and EMT process [31, 33]. Suppression of VCAM1 activity has been reported as a potential therapeutic strategy for GC [34]. Moreover, a recent study showed that silencing of IKBKG, another candidate protein target for BENC, promoted tumor growth and invasion of breast cancer through the dys-regulation of NF- κ B/STAT3 signaling pathways [35]. Thus, the therapeutic effects of BENC on GC cells are attributed to the disturbance of the viral carcinogenesis related pathways, such as NF- κ B/STAT3 signaling pathway.

CHM has been recognized for its holistic therapeutic effect on various diseases by combined

usage of multiple herbs containing a diversity of active compounds. It shows distinguished medical advantages and efficacy in the treatment of complex diseases such as cancer, owing to its special nature characteristics of multi-components, multi-targets and multi-modes of action [9]. In the present study, our results demonstrated the add-on benefits of TCM in improving outcomes of GC patients, and revealed that BENC suppressed cell growth, invasion and decreased the viability of GC cells through inductions of apoptosis, G2/M arrest and MET process, respectively, displaying a promising potential for its multi-levels, multi-targets and coordinated intervention effects on GC cells. BENC contains a diversity of key active compounds from multiple herbs, such as emodin and rhein from *Rheum Officinale*, polyphyllin I (PPI) from *Rhizoma Paridis*, colchicine from *Tulipa Edulis* and astragaloside IV (AS-IV) from *Radix Astragali Seu Hedysari Praeparata*. Recent studies have shown that emodin and rhein could induce apoptosis in nasopharyngeal carcinoma cells and glioma cells via targeting the chloride channels and ERK signaling pathway, respectively [36, 37], whereas PPI could suppress human osteosarcoma cell growth via inactivation of Wnt/ β -catenin pathway to arrest the cells at G2/M phase [38]. In human colon cancer cells, colchicine induced apoptosis via activation of p38 signaling pathway [39], while AS-IV inhibited EMT process by suppressing CREB1 signaling pathway [40]. Therefore, maintaining the diversity of active compounds plays an essential role to exhibit the strength of CHM in treating various cancers, and may help overcome the major challenges encountered in conventional chemotherapy, such as the drug-resistant phenotype.

On the other hand, to bring the BENC into line with modern medicine and better benefit GC patients, it is necessary to further carry out virtual and experimental assays for identification of the bioactive lead components, thereby trimming or withdrawing the redundant and deleterious components/herbs from BENC. At present, many chemoinformatics methods based on molecular docking, pharmacophore and quantitative structure activity relationships have been used for the bioactive leads identification against the drug targets [41, 42]. Thus, our next step is to perform a virtual screening of bioactive components from BENC, based on

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the 263 candidate targets we have identified in this study, and followed by experimental validation.

Conclusion

To facilitate the application of BENC on GC patients, we conducted a randomized clinical trial, in which we demonstrated that the add-on therapy with BENC resulted in better QOL, higher KPS and improved 3-year overall survival in advanced GC patients. The subsequent bioinformatics-assisted mechanism study revealed that BENC disturbed GC cell growth through affecting cell proliferation, cell cycle, apoptosis and EMT process. In perspective, the integration with cell-based biotechnologies and computational technologies will initiate the next revolution of CHM research. Here, we provided a proof-of-concept evidence showing how to use the efficacy-driven and bioinformatics-assisted strategy to benefit the advancement of CHM (Figure 7). In this connection, we speculate that the future development of CHM will be emphasized on accelerating its clinical application so as to benefit the mankind at larger extents.

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Disclosure of conflict of interest

None.

Abbreviations

CHM, Chinese herb medicine; BENC, Bo-Er-Ning capsule; GC, Gastric cancer; OS, Overall survival; KPS, Karnofsky performance scores; QOL, Quality of life; DC, Degree centrality; PPI, Protein-protein interaction; CCK-8, Cell Counting Kit-8; EMT, Epithelial-Mesenchymal transition; HCC, Hepatocellular carcinoma.

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Supplementary Table 1. The pharmaceutical ingredients of Bo-Er-Ning capsules

Latin name	English name	Chinese name
<i>Radix Astragali Seu Hedysari Praeparata</i>	Prepared Manyinflorescenced Sweetvetch Root	Zhi Huang Qi
<i>Ligustrum Lucidum</i>	Glossy Privet Fruit	Nv Zhen Zi
<i>Tulipa Edulis</i>	Edible Tulip	Guang Ci Gu
<i>Portulaca Oleracea</i>	Purslane	Ma Chi Xian
<i>Rhizoma Paridis</i>	Paris Root	Chong Lou
<i>Solanum Nigrum</i>	Black Nightshade	Long Kui
<i>Fructus Perillae</i>	Perilia Fruit	Zi Su Zi
<i>Gallus Gallus Domesticus</i>	Chicken's Gizzard Endothelium	Ji Nei Jin
<i>Rheum Officinale</i>	Rheum Palmatum	Da Huang
<i>Dryobalanops Aromatica</i>	Borneol	Bing Pian
<i>Bombyx Batryticatus</i>	Stiff Silkworm	Jiang Can