

Oligochitosan as a potential anti-acne vulgaris agent: combined antibacterial effects against *Propionibacterium acnes*

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Abstract To develop an antibacterial treatment for acne vulgaris using natural substance with few side effects, we investigated the antibacterial activities of oligochitosan against acne-related bacteria, particularly *Propionibacterium acnes*. Oligochitosan showed potent antibacterial effect on *P. acnes*. Especially, 10 kDa oligochitosan presented the highest antimicrobial effect with minimum inhibitory concentration values of 32–64 µg/mL on *P. acnes*. In addition, oligochitosan clearly reversed the antibacterial effect of tetracycline and erythromycin on *P. acnes* in the combination mode. The combination of tetracycline- or erythromycin-10 kDa oligochitosan resulted in a median ΣFIC range of 0.02–0.56, suggesting that the antibiotics–oligochitosan combination resulted in an antibacterial synergy against *P. acnes*. Thus, the results obtained in this research strongly supported that erythromycin and tetracycline will restore the antibacterial activity against *P. acnes* in the combination mode with 10 kDa oligochitosan.

Keywords Acne-related bacteria · Antibacterial activity · Erythromycin · Oligochitosan · Synergistic effect · Tetracycline

Introduction

Acne vulgaris is one of the most typical skin diseases, affecting approximately 80% of adults aged from 11 to 30 years [1]. There are several significant factors causing acne in people such as skin pathogenic bacteria, keratinization of the follicle, and increase in sebum secretion [2]. Although it is not life threatening, it can cause a permanent scar and also negatively affects psychological developments, resulting in a severe emotional scar, which may lead to clinical depression and social phobias [3]. It has been known that *Staphylococcus epidermidis*, *Propionibacterium acnes*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Candida albicans* are skin pathogens associated with the development of abnormal follicular keratinization and inflammation [4]. Among a variety of factors contributing to the development of acne vulgaris, bacterium *P. acnes* is the main factor in the pathogenesis of acne vulgaris [5]. In general, topical therapies for acne vulgaris, including retinoids, benzoyl peroxide, and antibiotics, have been used to improve the control of moderate acne. As described, an antibiotic application for treating acne vulgaris is usually used to kill acne-related bacteria. Among the various antibiotics, tetracycline, erythromycin, and lincomycin are commonly taken for the antibiotic therapy [6]. However, treatment by antibiotics and benzoyl peroxide, used to treat acne vulgaris, normally causes several side effects such as the emergence of resistant bacteria, immune hypersensitivity, and organ damage in the case of using those medicines for a long

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period; moreover, they are toxic to human cells [7, 8]. Therefore, recent researches have focused on the development of alternative therapeutic substance with less adverse and strong antibacterial effects [9].

Chitosan is a linear polysaccharide comprising randomly distributed β -(1 \rightarrow 4)-linked *N*-acetyl-D-glucosamine and D-glucosamine and possess several bioactivities, including antioxidant, antitumor, and enzymatic inhibitory effect. Recently, considerable attention has been focused on its applications in the food and pharmaceutical industry [8, 10]. However, high-molecular weight chitosan has a limitation on practical applications due to its insolubility at above pH 6.3 [11]. To overcome these disadvantages, recent researches have been conducted focusing on the transformation of chitosan into oligochitosan with a molecular weight of less than 10 kDa [12]. Oligochitosan, obtained by hydrolysis or degradation of chitosan, is a water-soluble chitosan with a low molecular as well as is known to be non-toxic, biodegradable and biocompatible due to their shorter chain lengths [13]. Oligochitosan is well absorbed in the body and can be added to any food as the main ingredient because it is easily soluble in water [14]. It has also been reported to exhibit high medicinal and functional aspects involved in the hypocholesterolemic effect [12], antitumor effects [14], accelerating calcium and ferrum absorption [15], immunoenhancement effect, antibacterial effect, and cholesterol regulation [16]. However, limited information is available on the antibacterial activity of low-molecular weight oligochitosan against acne-related bacteria. Therefore, the object of this research was to evaluate the antibacterial effect of oligochitosan against acne-related bacteria.

Materials and methods

Materials

Chitosan (MW 250 kDa) (80% degree of deacetylation) prepared from crab shell chitin was provided by Kitto Life Co. (Seoul, Korea) and oligochitosan (MW 1 kDa, MW 3 kDa, MW 5 kDa, and MW 10 kDa) (80% degree of deacetylation) were manufactured as previously reported by Park et al. [17] and Jung et al. [18]. Antibiotics were purchased from Sigma-Aldrich (St. Louis, USA). All other reagents were of the highest grade available commercially.

Evaluation of antimicrobial activity

Strains and culture conditions

The following microbial strains were obtained from the Korean Collection for Type Cultures (KCTC; Daejeon, Korea) and Korean Culture Center of Microorganisms

(KCCM, Seoul, Korea): *S. aureus* KCTC1927, *S. epidermidis* KCCM 40,416, *P. aeruginosa* KCTC 1637, and *P. acnes* KCTC 3314. Five *P. acnes* clinical isolates were provided by the Gyeongsang National University Hospital (Jinju, Korea), a member of the National Biobank of Korea. *P. acnes* strains were anaerobically cultivated in brain heart infusion broth (Difco Inc., Detroit, USA) supplemented with 1.0% glucose and incubated at 37 °C for 24 h in a CO₂ incubator (NAPCO 5400; General Laboratory Supply, Pasadena, USA) in a 10% CO₂ humidified atmosphere.

Measurement of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC)

MIC is the method of quantitatively evaluating the antimicrobial activity. It is defined as the lowest concentration of antimicrobial agents that will inhibit the visible growth of a microorganism after 20–24 h of incubation [19]. The experiment procedures by the guideline of Clinical and Laboratory Standards Institute were followed [20]. MIC assay was performed using the serial two-fold dilution method with Mueller–Hinton broth (MHB; Difco Inc.) and 96-well microtiter plates (with clear flat bottoms). After incubation, MIC values were determined by reading the plates visually. This test was repeated three times.

MBC is the lowest concentration of an antibacterial agent required to kill a particular bacterium [21]. It can be determined from broth dilution MIC tests by sub-culturing to agar plates that do not contain the test agent. MBC is identified by determining the lowest concentration of antibacterial agent that reduces the viability of the initial bacterial inoculum by $\geq 99.9\%$.

Morphology assessment by scanning electron microscopy (SEM)

The morphological change of *P. acnes* cells by the treatment of 10 kDa oligochitosan was monitored using a SEM VEGA II LSU microscope (Tescan, Brno, Czech Republic). The specimens were primary fixed in 2.5% buffered glutaraldehyde (Sigma-Aldrich) for 2 h, rinsed in distilled water, and post-fixed in 1% buffered osmium tetroxide (Sigma-Aldrich) for 2 h. The fixed samples were then dehydrated stepwise with ethanol (25, 50, 70, 94, and 100%) and subsequently samples were freeze dried and observed by SEM.

Antibiotic susceptibility test (AST)

AST is used to determine whether an organism is susceptible or resistant to an antimicrobial agent [22]. The antibiotic resistance of the test strains was confirmed

against commercial antibiotics by MIC assay. An antibiotic was serially diluted and then the bacterial growth was visually checked.

Determination of fractional inhibitory concentration (FIC)

FIC assay is widely used for evaluation of in vitro synergy for multiple agents [23]. This test method assesses interaction between antimicrobial agents by exposing bacteria to varying concentrations of the antimicrobial drugs [24]. In this study, the synergy effect between oligochitosan and antibiotics (tetracycline, erythromycin, and lincomycin) against pathogens of human skin was evaluated. FIC index was calculated using the following formula:

$$FIC_A = MIC_A \text{ in combination} / MIC_A$$

$$FIC_B = MIC_B \text{ in combination} / MIC_B$$

$$FIC \text{ index} = FIC_A + FIC_B$$

The interaction was defined as synergistic if the FIC index was <1; additive if the FIC index was 1.0; sub-additive if the FIC index was between 1.0 and 2.0; indifferent if the FIC index was 2, and antagonistic if the FIC index >2. Synergy was further sub-classified as marked (FIC index, ≤0.50) and weak (FIC index, between 0.50 and 1.0) [4].

Statistical analysis

In all cases, analyses were performed in triplicate and data were averaged over the three measurements. The standard deviation was also calculated. Significance of differences between the average MICs for each individual microorganism was determined by Student's t test at the 95% significance level using SPSS version 12.0 (SPSS Inc., Chicago, USA).

Results and discussion

Determination of MIC and MBC of chitosan and oligochitosan

Acne vulgaris is one of the most typical skin diseases causing disfiguration and permanent scarring. According to Lee et al. [4], *P. acnes* appears to be the most probable organism to cause acne vulgaris and is therefore the target of topical antibiotic treatments. However, antibiotic therapies normally have several adverse effects such as organ damage, emergence of resistant bacteria, and immune hypersensitivity. To discover an alternative therapeutic

agent with few side effects from marine resources, the antibacterial effects of chitosan and oligochitosan were quantitatively evaluated by the MIC assay. The MIC values of the chitosan and oligochitosan were 32–4096 µg/mL against Gram positive acne-related bacteria tested in this study. No antibacterial activity was observed against Gram negative acne-related bacterium, *P. aeruginosa* (data not shown). Interestingly, oligochitosan showed strong antibacterial activity against *P. acnes* strains in the MICs range of 16–512 µg/mL. Moreover, oligochitosan exhibits equal or lower MIC values than those of the chitosan against *P. acnes*. However, oligochitosan showed less antimicrobial effects against other acne-related bacteria, including *P. aeruginosa*, *S. epidermidis* and *S. aureus*. Oligochitosan (MW 1, 3, 5, and 10 kDa) showed stronger antibacterial effect on *P. acnes* than that of chitosan. Among oligochitosan, the highest antibacterial effect was observed in the treatment of 10 kDa oligochitosan. The MIC value of oligochitosan was 16–64 µg/mL against *P. acnes* strains.

In order to evaluate the bactericidal activity of chitosan and oligochitosan, MBC assay was performed. MBC values of chitosan and oligochitosan were 32–8192 µg/mL against acne-inducing bacteria associated with acne vulgaris. Similar to the MIC results obtained in Table 1, oligochitosan showed strong antibacterial activity against *P. acnes* strains in MBC range of 32–2048 µg/mL.

The antibacterial activity of oligochitosan was superior to that of the chitosan against *P. acnes*. Among oligochitosan, 10 kDa oligochitosan showed the highest antibacterial activity in a range of 32–128 µg/mL against *P. acnes*. However, the MBC values of 10 kDa oligochitosan were two-folds increased compared to those of MIC values. Similar patterns between MIC and MBC values were reported by several studies [9, 25]. Considering both MIC and MBC results, 10 kDa oligochitosan exhibited the highest antibacterial activity against *P. acnes* strains. As reported by Rabea et al. [26] and Champer et al. [27], chitosan with high-molecular weight exhibited strong antimicrobial activity against cutaneous pathogens, excluding *P. acnes*. Interestingly, however, 10 kDa oligochitosan showed superior antibacterial activity against *P. acnes* strains in the ranges of MIC values from 16 to 128 µg/mL.

The mechanism of antimicrobial activity of oligochitosan is still yet to be elucidated. However, chitosan molecules are reported to be stacked over the microbial cell surface, blocking the nutrients, or bind to DNA, as such inhibiting transcription or permeability of the microbial cell wall [28, 29]. Chitosan samples with a Mw <5 kDa induce filamentation in *B. megaterium* by blocking the synthesis of DNA. Thus, the molecular weight of chitosan can influence the permeability of chitosan through the cell membrane [30]. Therefore, further studies are necessary to

Table 1 Minimum inhibitory concentration (MIC) of chitosan and oligochitosan against acne-related bacteria

Strains	Oligochitosan (MW 1 kDa)		Oligochitosan (MW 3 kDa)		Oligochitosan (MW 5 kDa)		Oligochitosan (MW 10 kDa)		Chitosan (MW 250 kDa)	
	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC
<i>S. aureus</i> KCTC1927	4096	8192	4096	8192	2048	4096	4096	8192	128	512
<i>S. epidermidis</i> KCCM 40416	4096	8192	4096	8192	2048	4096	4096	8192	512	1024
<i>P. acnes</i> KCTC 3314	512	1024	32	64	32	64	16	32	512	2048
<i>P. acnes</i> isolate 2874	512	1024	64	256	32	64	32	64	512	2048
<i>P. acnes</i> isolate 2875	512	1024	128	512	64	128	64	128	512	2048
<i>P. acnes</i> isolate 2876	512	1024	64	128	64	128	32	64	512	2048
<i>P. acnes</i> isolate 2877	512	1024	64	256	32	128	32	64	512	2048
<i>P. acnes</i> isolate 2878	512	1024	128	256	64	128	64	64	512	2048

address the detail antimicrobial mechanism of 10 kDa oligochitosan particularly against *P. acnes*.

According to the previous reports, chitosan and chitosan phytochemical conjugates showed MIC value of 128–512 µg/mL for *P. acnes* and isolated *P. acnes* [9], and *n*-hexane extract of *Sargassum serratifolium* C. Agardh showed MICs of 128 µg/mL [25]. Lee et al. [4] also reported that phlorotannins from *Eisenia bicyclis* showed strong antibacterial activity in the ranges of 64–1024 µg/mL against *P. acnes* strains. Considering these results, oligochitosan exhibiting MICs of 16–64 µg/mL possessed superior antibacterial activity than that of phlorotannins. In addition, these results strongly indicate that oligochitosan will be a potential candidate to develop an alternative therapeutic agent for the treatment of *P. acnes* infection.

Effect of 10 kDa oligochitosan on *P. acnes* cells morphology

As described above, 10 kDa oligochitosan exhibited the highest antibacterial activity. To investigate the antibacterial effect of oligochitosan on *P. acnes* cells, changes in the cells morphology were monitored using SEM. Considering the MIC values of oligochitosan, *P. acnes* cells were treated by various concentrations of oligochitosan (0, 16, 32, and 64 µg/mL) and cells were cultured at 37 °C under anaerobic for a day. As shown in Fig. 1. SEM micrographs showed distorted and damaged cells by the treatment of 10 kDa oligochitosan above 32 µg/mL. In more detail, untreated *P. acnes* cells show normal cell surface architecture, which appears smooth and sharply layered [Fig. 1(A)]. In contrast, the bacterial cells treated by 10 kDa oligochitosan showed loosen integrity on their surface structure [Fig. 1(C), (D)]. These results coincide with the previous report of Champer et al. [27] that

chitosan can bind to the surface of bacteria causing shrinkage of *P. acnes* cell surface structure. The antibacterial mechanism of chitosan has not been fully elucidated yet, but the most accepted antibacterial mechanism of chitosan is directly dependent on its deacetylation and amino group [31]. Negatively charged bacterial cell surface interacts with positively charged chitosan, resulting in weakening of the cell wall either accompanied by cell lysis or cell wall damage [32]. Moreover, a higher molecular weight of chitosan is reported to have stronger antimicrobial activity than low-molecular chitosan. In this study, however, oligochitosan with low-molecular weight is more effective than chitosan with high-molecular weight for controlling *P. acnes*, suggesting that the different antibacterial mechanism against *P. acne* compared to chitosan with high-molecular weight. In order to address this issue in more detail, further study will be needed.

Antibiotics and benzoyl peroxide resistance of *P. acnes* strains

The antibiotic resistance profile of *P. acnes* and isolated *P. acnes* was determined based on the analysis of MIC breakpoint [33]. MIC breakpoint values of tetracycline, lincomycin, and erythromycin were 1–4 µg/mL, 2–8 µg/mL, and 4–8 µg/mL, respectively. The MIC values of lincomycin against *P. acnes* strains were in the range of the Soussy's MIC breakpoint values, indicating susceptibility to the test antibiotic (Table 2). The MIC of *P. acnes* standard strain against tetracycline is 16 µg/mL, which was over the range of MIC breakpoint. Moreover, some *P. acnes* isolates were found to be resistant to erythromycin and tetracycline. However, *P. acnes* strains tested in this research were judged to be sensitive to benzoyl peroxide based on the analysis of the MIC values.

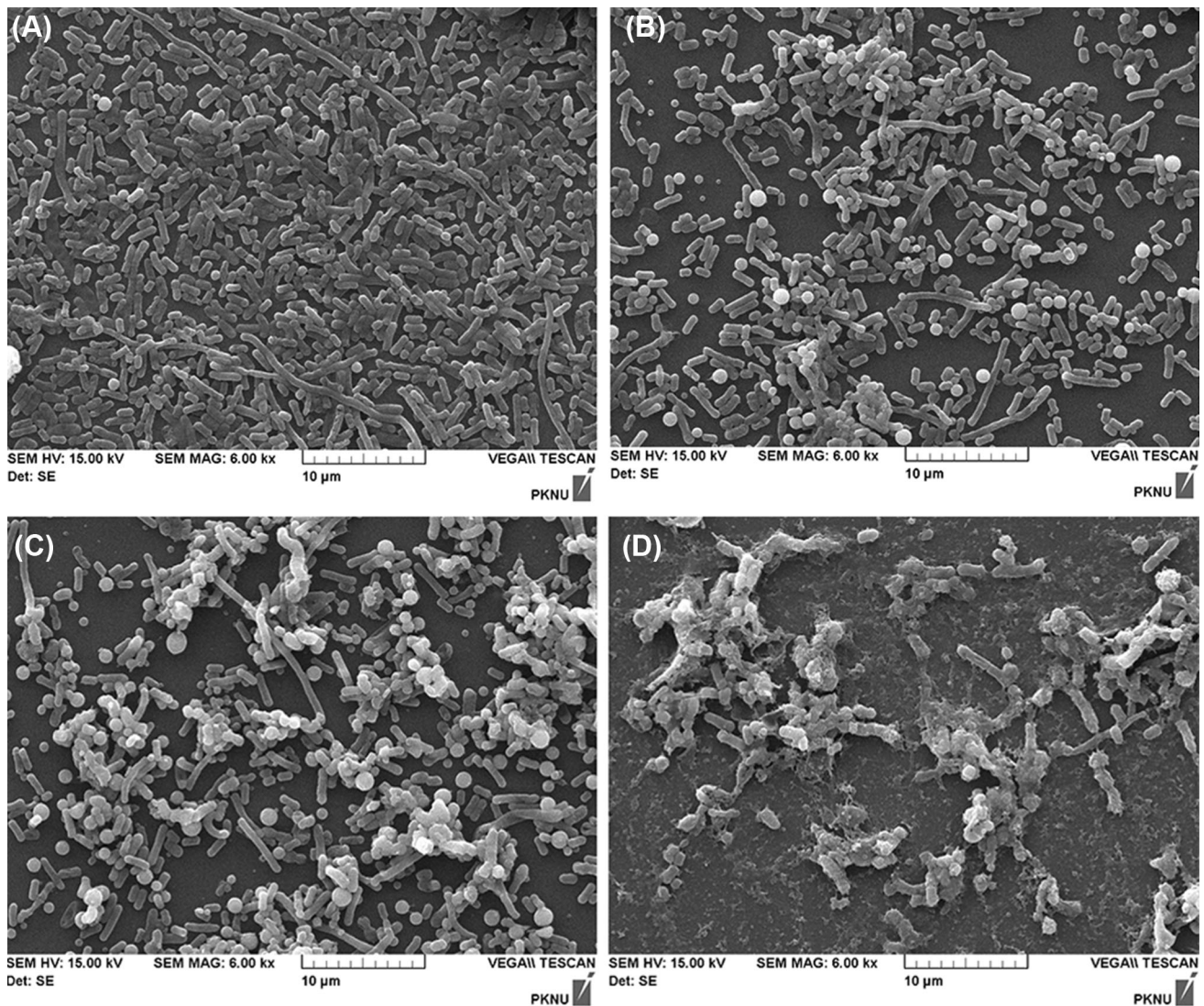


Fig. 1 Scanning electron microscopy profiles on the antibacterial effect of 10 kDa oligochitosan against *Propionibacterium acnes* ($\times 6000$ K magnification) (A) control (B) treated with oligochitosan of 16 $\mu\text{g/mL}$ (C) treated with oligochitosan of 32 $\mu\text{g/mL}$ (D) treated with oligochitosan of 64 $\mu\text{g/mL}$

Table 2 Minimum inhibitory concentration (MIC) of antibiotics against *Propionibacterium acnes* strains

Strains	MIC ($\mu\text{g/mL}$)			
	Tetracycline	Lincomycin	Erythromycin	Benzoyl peroxide
Break point	4–8	2–8	1–4	250
<i>P. acnes</i> KCTC 3314	16	4	<1	64
<i>P. acnes</i> isolate 2874	32	8	<1	64
<i>P. acnes</i> isolate 2875	2	<1	<1	64
<i>P. acnes</i> isolate 2876	<1	<1	64	64
<i>P. acnes</i> isolate 2877	32	8	<1	64
<i>P. acnes</i> isolate 2878	32	8	<1	64

These antibiotic-resistant profiles against *P. acnes* strains are almost similar with that reported by Lee et al. [4], Kim et al. [9], and Kim et al. [25]. Likewise, the profiles of MICs to antibiotics were similar with the previous results [4, 9, 25]. The results

obtained in this study indicated that some commercial antibiotics are no longer effective for treating bacterial infection related with *P. acnes*. Thus, alternative therapeutic agents for no longer effective antibiotics are required.

Table 3 Fractional inhibitory concentration (FIC) indices of 10 kDa oligochitosan in combination with antibiotics against *Propionibacterium acnes*

Strains	Test compound	MIC ($\mu\text{g/mL}$)	$\sum\text{FIC}_{\text{max}}^{\text{a}}$	$\sum\text{FIC}_{\text{min}}^{\text{b}}$	Median $\sum\text{FIC}$	Minimum concentration for observing synergy
<i>P. acnes</i> KCTC 3314	Oligochitosan	16	1.03	0.28	0.28	0.5
	Tetracycline	16				4
	Oligochitosan	16	1.50	1.00	1.03	0.5
	Benzoyl peroxide	64				64
<i>P. acnes</i> isolate 2874	Oligochitosan	32	1.06	0.50	0.56	1
	Tetracycline	32				16
	Oligochitosan	32	2.00	1.00	1.02	0.5
	Benzoyl peroxide	64				64
<i>P. acnes</i> isolate 2875	Oligochitosan	64	1.25	0.52	1.00	1
	Benzoyl peroxide	64				64
<i>P. acnes</i> isolate 2876	Oligochitosan	32	0.02	0.02	0.02	0.02
	Erythromycin	64				1
	Oligochitosan	32	1.50	1.00	1.03	1
	Benzoyl peroxide	64				64
<i>P. acnes</i> isolate 2877	Oligochitosan	32	1.25	0.25	0.34	1
	Tetracycline	32				8
	Oligochitosan	32	1.50	1.00	1.03	1
	Benzoyl peroxide	64				64
<i>P. acnes</i> isolate 2878	Oligochitosan	64	1.03	0.50	0.52	2
	Tetracycline	32				16
	Oligochitosan	64	2	2	2	64
	Benzoyl peroxide	64				64

The FIC index indicated synergistic effect: <0.5 , marked synergy; 0.5 to <1.0 , weak synergy; 1.0 , additive; >1.0 to <2.0 , subadditive; 2.0 , indifferent; >2.0 , antagonistic

^a $\sum\text{FIC}_{\text{max}}$, maximum FIC

^b $\sum\text{FIC}_{\text{min}}$, minimum FIC

Synergistic effect between oligochitosan (MW 10 kDa) and antibiotics against *P. acnes* strains

With the emergence of multidrug resistant bacteria, the need for new antibiotics or therapeutic agents has increased [34]. It has been proved that one of the more effective approaches in developing alternative therapies or new drugs is the restoration of antibiotic effect in combination with antibacterial substances derived from natural products and traditional drug against drug-resistant bacteria [34]. Based on these reports, an interaction between 10 kDa oligochitosan and commercial antibiotics (or benzole peroxide) against *P. acnes* strains, antibiotic-resistant strains as reported previously using AST, was tested by the checkerboard method using FIC assay, as stated in “Materials and methods” section [24]. As shown in Table 1, 10 kDa oligochitosan was chosen for further study among the chitosan and its oligo since oligochitosan showed the highest antibacterial effect on *P. acnes*.

MICs of tetracycline against *P. acnes* strains combining with 10 kDa oligochitosan fairly decreased from 16 to 4 $\mu\text{g/mL}$. On comparing the MIC of the tetracycline alone, it can be observed that the MIC has decreased 2 to fourfolds in the combination of tetracycline and 10 kDa oligochitosan. In all test strains, the median $\sum\text{FIC}$ indices were from 0.28 to 0.56, indicating a synergistic antibacterial effect between tetracycline and oligochitosan (Table 3).

The highest synergistic antibacterial activity was observed in the combination with erythromycin and 10 kDa oligochitosan against *P. acnes* isolate 2876 strain. The MIC of erythromycin against the isolates combining with 32 $\mu\text{g/mL}$ of 10 kDa oligochitosan dramatically decreased from 64 to 1 $\mu\text{g/mL}$. The median FIC index was 0.02, indicating very strong synergistic effect against *P. acnes* isolate. However, no synergistic antibacterial effect against *P. acnes* stains was observed in the combination of benzoyl peroxide and oligochitosan (Table 3).

It has been previously reported that phlorotannins of *E. bicyclis* showed the median $\sum\text{FIC}$ indices ranging from

0.502 to 0.750 in combination with β -lactam antibiotics against *P. acnes* [4]. Kim et al. [25] also reported that a synergy effect between chitosan phytochemical conjugate and tetracycline with the median Σ FIC indices from 0.502 to 0.533 against *P. acnes* strains. Compared with these results, the synergistic effect of 10 kDa oligochitosan with antibiotics is remarkably high. Thus, the 10 kDa oligochitosan has a potential to restore the antibacterial activity of old-fashioned antibiotics such as tetracycline and erythromycin against *P. acnes*. It was anticipated that the 10 kDa oligochitosan would be a good candidate in the remedy of antibiotic-resistant *P. acnes* infection.

In conclusion, the oligochitosan possessed combined antibacterial effects on *P. acnes*. The finding obtained in this research strongly suggested that 10 kDa oligochitosan will be a potential alternative therapeutic substance for acne vulgaris treatment since it has strong antibacterial effect on acne-related bacteria.

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Compliance with ethical standards

Conflict of interest The authors declare no conflict of interest.

References

- Uhlenhake E, Yentzer BA, Feldman SR. Acne vulgaris and depression: a retrospective examination. *J. Cosmet. Dermatol.* 9: 59–63 (2010)
- Farrar MD, Ingham, E. Acne: inflammation. *Clin. Dermatol.* 22: 380–384 (2004)
- Ash C, Harrison A, Drew S, Whittall R. A randomized controlled study for the treatment of acne vulgaris using high-intensity 414 nm solid state diode arrays. *J. Cosmet. Laser Ther.* 17: 170–176 (2015)
- Lee JH, Eom SH, Lee EH, Jung YJ, Kim HJ, Jo MR, Son KT, Lee HJ, Kim JH, Lee MS, Kim YM. *In vitro* antibacterial and synergistic effect of phlorotannins isolated from edible brown seaweed *Eisenia bicyclis* against acne-related bacteria. *Algae* 29: 47–55 (2014)
- Liu PF, Nakatsuji T, Zhu W, Gallo RL, Huang CM. Passive immunoprotection targeting a secreted CAMP factor of *Propionibacterium acnes* as a novel immunotherapeutic for acne vulgaris. *Vaccine* 29: 3230–3238 (2011)
- Han S, Lee K, Yeo J, Baek H, Park K. Antibacterial and anti-inflammatory effects of honeybee (*Apis mellifera*) venom against acne-inducing bacteria. *J. Med. Plants Res.* 4: 459–464 (2010)
- Kim JY, Oh TH, Kim BJ, Kim SS, Lee NH, Hyun CG. Chemical composition and anti-inflammatory effects of essential oil from *Farfugium japonicum* flower. *J. Oleo Sci.* 57: 623–628 (2008)
- Lee SH, Ryu BM, Je JY, Kim SK. Diethylaminoethyl chitosan induces apoptosis in HeLa cells via activation caspase-3 and p53 expression. *Carbohydr. Polym.* 84: 571–578 (2011)
- Kim JH, Je JY, Kim YM. Anti-inflammatory effects of chitosan-phytochemical conjugates against *Propionibacterium acnes*-induced inflammation. *Korean J. Fish. Aquat. Sci.* 49: 589–593 (2016)
- Cho YS, Kim SK, Ahn CB, Je JY. Preparation, characterization, and antioxidant properties of gallic acid-grafted-chitosans. *Carbohydr. Polym.* 83: 1617–1622 (2011)
- Seo S, King JM, Prinyawiwatkul W. Simultaneous depolymerization and decolorization of chitosan by ozone treatment. *J. Food Sci.* 72: 522–526 (2007)
- Kim SK, Rajapakse N. Enzymatic production and biological activities of chitosan oligosaccharides (COS): a review. *Carbohydr. Polym.* 62: 357–368 (2005)
- Lodhi G, Kim YS, Hwang JW, Kim SK, Jeon YJ, Je JY, Ahn CB, Moon SH, Jeon BT, Park PJ. Chitoooligosaccharide and its derivatives: preparation and biological applications. *BioMed Res. Int.* Article ID 654913 (2014)
- Xia WS. Physiological activities of chitosan and its application in functional foods. *J. Chinese Inst. Food Sci. Technol.* 3: 77–81 (2003)
- Bravo-Osuna I, Millotti G, Vauthier C, Ponchel G. In vitro evaluation of calcium binding capacity of chitosan and thiolated chitosan poly (isobutyl cyanoacrylate) core-shell nanoparticles. *Int. J. Pharm.* 338: 284–290 (2007)
- Xia W, Liu P, Zhang J, Chen J. Biological activities of chitosan and chitoooligosaccharides. *Food Hydrocolloid.* 25: 170–179 (2011)
- Park PJ, Je JY, Jung WK, Byun HG, Kim SK. Free radical scavenging activities of chitoooligosaccharides hydrolyzed from chitosan with high degree of deacetylation on 1,1-diphenyl-2-picrylhydrazyl radical. *J. Chitin Chitosan* 9: 108–113 (2004)
- Jung WK, Moon SH, Kim SK. Effect of chitoooligosaccharides on calcium bioavailability and bone strength in ovariectomized rats. *Life Sci.* 78: 970–976 (2006)
- Grierson DS, Afolayan AJ. Antibacterial activity of some indigenous plants used for the treatment of wounds in the Eastern Cape, South Africa. *J. Ethnopharmacol.* 66: 103–106 (1999)
- Clinical and Laboratory Standards Institute (CLSI). Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically: Approved standard, 8th ed. CLSI document M07-A8. CLSI, Wayne, PA, pp 68 (2006)
- Amyes S, Miles RS, Thomson CJ, Tillotson G. Antimicrobial Chemotherapy: Pocketbook. CRC Press, Florida, pp 25 (1996)
- Jenkins SG, Schuetz AN. Current concepts in laboratory testing to guide antimicrobial therapy. *Mayo Clin. Proc.* 87: 290–308 (2012)
- Odds FC. Synergy, antagonism, and what the checkerboard puts between them. *J. Antimicrob. Chemother.* 52: 1 (2003)
- Hsieh MH, Yu CM, Yu VL, Chow JW. Synergy assessed by checkerboard. A critical analysis. *Diagn. Microbiol. Infect. Dis.* 16: 343–349 (1993)
- Kim YH, Kim JH, Kim DH, Kim SH, Kim HR, Kim YM. Synergistic Antimicrobial Effect of *Sargassum serratifolium* (C. Agardh) C. Agardh extract against human skin pathogens. *Korea J. Food Sci. Technol.* 48: 241–246 (2016)
- Rabea EI, Badawy ME, Stevens CV, Smaghe G, Steurbaut W. Chitosan as antimicrobial agent: applications and mode of action. *Biomacromolecules* 4: 1457–1465 (2003)
- Champer J, Patel J, Fernando N, Salehi E, Wong V, Kim J. Chitosan against cutaneous pathogens. *AMB Express*, 3: 37 (2013)
- Shon DH. Chitosan oligosaccharides for functional foods and microbial enrichment of chitosan oligosaccharides in soy-paste.

- In: Proceedings of the International workshop on Bioactive Natural Products. The Committee on Science and Technology in Developing Countries (COSTED) and the Science Council of Japan, Tokyo, Japan: 56–66. (2001)
29. Tharanathan RN, Kittur FS. Chitin—The undisputed biomolecule of great potential. *Crit. Rev. Food Sci. Nutr.* 43: 61–87 (2003)
 30. Park SC, Nah JW, Park Y. pH-dependent mode of antibacterial actions of low molecular weight water-soluble chitosan (LMWSC) against various pathogens. *Macromol. Res.* 19: 853–860 (2011)
 31. Goy RC, Britto DD, Assis OB. A review of the antimicrobial activity of chitosan. *Polimeros* 19: 241–247 (2009)
 32. Eaton P, Fernandes JC, Pereira E, Pintado ME, Malcata FX. Atomic force microscopy study of the antibacterial effects of chitosans on *Escherichia coli* and *Staphylococcus aureus*. *Ultramicroscopy* 108: 1128–1134 (2008)
 33. Soussy CJ, Cluzel R, Courvalin P. Definition and Determination of in vitro antibiotic susceptibility breakpoints for bacteria in France. *Eur. J. Clin. Microbiol. Infect. Dis.* 13: 238–246 (1994)
 34. Eom SH, Park JH, Yu DU, Choi JI, Choi JD, Lee MS, Kim YM. Antimicrobial activity of brown alga *Eisenia bicyclis* against methicillin-resistant *Staphylococcus aureus*. *Fish. Aquat. Sci.* 14: 251–256 (2011)