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# **IL-17 in Neonatal Health and Disease**

**Shelley M. Lawrence, MD, MS<sup>1,2</sup>, J. Lauren Ruoss, MD<sup>3</sup>, and James L. Wynn, MD<sup>†,3,4</sup>** <sup>1</sup>University of California, San Diego, College of Medicine, Department of Pediatrics, Division of Neonatal-Perinatal Medicine

<sup>2</sup>University of California, San Diego, Department of Pediatrics, Division of Host-Microbe Systems and Therapeutics

3University of Florida, College of Medicine, Department of Pediatrics, Division of Neonatal-Perinatal Medicine

<sup>4</sup>University of Florida, Department of Pathology, Immunology, and Laboratory Medicine

# **Abstract**

Over the last few years, scientific interest in the cytokine IL-17A has intensified as its role in human health and disease has been elucidated. Discovered almost a quarter century ago, IL-17A is known to have poor biologic activity when acting alone, but attains robust actions when working synergistically with potent mediators of proinflammatory immune responses, such as IL-6 and IL-8. IL-17A is produced by specialized innate immune cells that protect host barriers from the outside world. Like sentries, these innate immune cells can "sound the alarm" through increased production of IL-17A, causing activation and recruitment of primed neutrophils and monocytes when pathogens escape initial host defenses. In this way, IL-17A promulgates mechanisms responsible for pathogen death and clearance. However, when IL-17A pathways are triggered during fetal development, due to chorioamnionitis or *in utero* inflammatory conditions, IL-17A can instigate and/or exacerbate fetal inflammatory responses that increase neonatal morbidities and mortality associated with common neonatal conditions such as sepsis, bronchopulmonary dysplasia (BPD), patent ductus arteriosus (PDA), and necrotizing enterocolitis (NEC). This review details the ontogeny of IL-17A in the fetus and newborn, discusses how derangements in its production can lead to pathology, and describes known and evolving therapies that may attenuate IL-17A-mediated human conditions.

## **Keywords**

IL-17; neonate; sepsis; patent ductus arteriosus; necrotizing enterocolitis; bronchopulmonary dysplasia; retinopathy of prematurity

<sup>†</sup>**Corresponding Author:** James L. Wynn, MD. Division of Perinatal-Neonatal Medicine, Department of Pediatrics, The University of Florida; 1600 SW Archer Rd, P.O. Box 100296, Gainesville, Florida 32610-0296; james.wynn@peds.ufl.edu. Phone: (352) 273-8985. Shelley M. Lawrence MD, MS. Division of Perinatal-Neonatal Medicine, Department of Pediatrics, The University of California, San Diego; 9500 Gilman Drive, MC0760, La Jolla, California 92093-0760; slawrence@ucsd.edu. Phone: (303) 704-8908; Fax: (858) 822-3593.

#### **Introduction**

First discovered in 1993<sup>1</sup>, interleukin (IL)-17A (aka CTLA8) belongs to a family of six IL-17 cytokines (IL17A-F) and demonstrates diverse biologic functions<sup>2-4</sup>. In general, IL-17A is produced by a wide spectrum of innate immune cells that are strategically located in barrier tissues that protect the human body from the outside environment. If breached by pathogenic microorganisms, these specialized immune cells become key instigators of early innate immune responses that may negatively impact the future health of the host<sup>5</sup>. IL-17Amediated fetal inflammatory responses can manifest in childhood with debilitating disease, including increased severity of respiratory syncytial virus (RSV) infection<sup>6,7</sup>, asthma exacerbations<sup>8</sup>, and Crohn's and inflammatory bowel disease<sup>9-11</sup>. In adults, IL-17A is strongly associated with disease severity and progression of psoriatic responses in the skin<sup>12</sup>, depression<sup>13,14</sup>, and post-ischemic phases of stroke<sup>2,15–19</sup>. This review will detail our current understanding of IL-17A and its association with common, debilitating neonatal conditions. In addition, the ontogeny of IL-17A-producing immune cells will be discussed, including changes in lymphocyte composition with fetal and neonatal maturation. Lastly, novel pharmacologic interventions, which target IL-17A pathways to alleviate disease will be addressed.

# **Cells that produce IL-17**

Differentiation of naïve T lymphocytes into distinct effector T helper (Th) cells is essential for proper adaptive immune responses<sup>20</sup>. Naïve T helper cells are currently known to differentiate into: (i) Th1 cells, which are vital for cell-mediated immunity and have a key role initiating early resistance to pathogens, (ii) Th2 cells, which stimulate antibodymediated responses and promote immune tolerance rather than defend against microbial infection, (iii) regulatory T (Treg) cells, which suppress immune activity, and (iv) Th17 cells, which are important in the clearance of extracellular bacteria through IL-17A production $20-23$ .

Th17 cells generate proinflammatory cytokines including IL-17A, IL-22, IL-26, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), chemokine (C–C motif) ligand 20 (CCL20)<sup>24</sup>, and granulocyte macrophage colony stimulating factor  $(GM-CSF)^{11,25}$ . Differentiation of Th17 cells from naïve T cells is stimulated by proinflammatory cytokines, including IL-1β, IL-6, IL-21, and IL-23, in coordination with transforming growth factor-β (TGF-β) in a RAR-related orphan receptor-γ t (RORγt) – dependent manner<sup>11,26–29</sup>. Whereas RORγt triggers the development of Th17 cells and IL-23 induces their maturation and expansion, TGF-β promotes Th17 cell differentiation in a dose-dependent manner<sup>11</sup>. At low concentrations TGF-β activates RORγt expression and Th17 differentiation, while high TGF-β concentrations stimulate forkhead box P3 (FOXP3) and Treg formation $11,30$ . Because of their opposing immune responses, the Th17/Treg balance is crucial for maintaining immune homeostasis. However, in extreme inflammatory or infectious conditions, the formation of Treg lymphocytes can become defective, thereby permitting these cells to phenotypically transform into Th17-like cells<sup>3,31,32</sup> through a process referred to as *plasticity* or transdifferentiation<sup>28</sup>.

The clinical importance of Th17 cells is apparent in preterm neonates with chorioamnionitis exposure. Chorioamnionitis is a pathologic state of inflammation of the fetal membranes and is associated with increased cord blood quantities of IL-17A-producing Th17 lymphocytes<sup>33</sup> and Tregs that exhibit Th17-like phenotypes<sup>4</sup>. Because a common lineage is shared by Tregs and Th17 cells via developmental regulation of the transcription factor  $ROR\gamma\delta^{34}$ , extremely high levels of RORγδ expression during inflammation contributes to loss of Treg antiinflammatory function and facilitates the conversion of Tregs to Th17 subtypes<sup>35,36</sup>. Likewise, cord blood T lymphocytes with effector memory (TEM) cells can also express Th17-related markers and produce IL-17A under inflammatory conditions<sup>37</sup>. Increased concentrations of IL-17A during early inflammatory conditions enables Th17 cells to assume Th1-like effector functions, thereby expediting the recruitment of stimulated neutrophils to the site of infection and activating of regional macrophages to promote pathogen clearance<sup>20, 38, 39</sup>. In the most extreme preterm neonates, however, clinically relevant deficiencies of Th17 and Th1 cellular responses, due to impaired induction of the p40 subunit common to IL-12 and IL-23, leads to decreased concentrations of these interleukins<sup>21,40,41</sup>. Reductions in IL-12 and IL-23 in this vulnerable patient population can heighten risks of early onset neonatal sepsis despite the presence of higher concentrations of other pro-inflammatory cytokines<sup>21,41</sup>. Alternatively, permutations in the normal fetal immune development towards a proinflammatory Th17 response, instead of a tolerant Th2 response, may initiate fetal inflammatory pathways that result in debilitating chronic diseases in the neonatal period or throughout a person's lifespan<sup>33</sup>.

IL-17A is also produced by  $\gamma$  δT cells, lymphoid-tissue inducer (LTi) cells<sup>42</sup>,  $\gamma$  δT cells<sup>43</sup>, invariant natural killer T (iNKT) cells<sup>44</sup>, natural killer (NK) cells<sup>45</sup>, neutrophils<sup>46</sup>, Paneth cells<sup>47,48</sup>, microglia<sup>49</sup>, and innate lymphoid cells group 3 (ILC3)<sup>2,5</sup>. These innate immune cells are strategically located in barrier tissues that protect the human body from the outside environment and provide vital host protections against foreign threats<sup>5</sup>. Serving primarily as sentinels, these innate immune cells can rapidly (within 4 to 8 hours) increase IL-17A production after a pathogenic challenge, luring primed neutrophils to inflamed sites to expedite the death and clearance of offending microorganisms<sup>5</sup>.

Many of these innate immune cells constitutively express the cell surface receptors IL-1R1 and IL-23R<sup>5,50–53</sup> and, therefore, achieve maximal IL-17A production following exposure to both IL-1 $\beta$  and IL-23 cytokines<sup>5,54</sup>. Given the close proximity of these immune cells with the outside environment, commensal microorganisms that comprise the host's microbiome may induce IL-1R1 expression on their cell surface to enhance the biological effects of IL-17A $5,50$ . As a result, this relationship may enable this group of innate immune cells to 're-shape' the gut microbiota as necessary to maintain a healthy physiologic state through increased production of IL-17, IL-22, and antimicrobial peptides that target specific groups of bacteria<sup>5</sup>. Dysregulation of these cells, however, may also ignite or aggravate chronic inflammatory and/or autoimmune conditions, becoming clinically relevant at any age<sup>5,54,55</sup>.

# **IL-17 function**

IL-17A has diverse proinflammatory effects but may be poorly active when operating alone3,56. IL-17A must therefore synergize with other proinflammatory cytokines, such as

TNF-α, IL-1β, IL-22, IFN-γ, and GM-CSF, to generate high levels of IL-6 and IL-8, which are potent mediators of host immune responses $4,57$ . By bolstering granulopoiesis, IL-17A promotes the recruitment of large numbers of activated neutrophils to inflamed sites<sup>58</sup>, where neutrophil cytotoxic<sup>59</sup> and phagocytic activities<sup>60</sup> are enhanced, thereby providing an amplification loop for neutrophil proinflammatory responses<sup>61</sup>. IL-17A also enhances the recruitment and survival of macrophages $4.62$ , stimulates the production of antimicrobial proteins and peptides from a variety of immune and non-immune cells<sup>4,34,58,63,64</sup>, and promotes the secretion of IL-2 from T helper cells, which accelerates the proliferation of Treg and conventional T lymphocytes $4,65$ .

In general, IL-17A is important in combating extracellular pathogens including Grampositive, Gram-negative, and fungal microbes $40,66,67$ . Neonates demonstrate reduced baseline production of IL-17A compared to adults, which may diminish neonatal immune responses and contribute to their increased susceptibility to infection by group B Streptococcus, Escherichia coli<sup>40</sup>, and Klebsiella pneumoniae<sup>20,68</sup>. Because IL-17A levels are similar between preterm and term neonates, its production is not dependent upon immune maturation or advancing gestational age<sup>20</sup>. In mice, IL-17A-producing  $\gamma \delta T$  cells originate only within the embryonic thymus beginning at embryonic day 1569. This period corresponds to approximately 24-28 weeks' gestation in humans, which is associated with the highest risks of IL-17A-associated neonatal pathology including sepsis, necrotizing enterocolitis, and brain injury<sup>70</sup>. Although other factors contribute to the heightened susceptibility to pathology at this developmental stage, there is burgeoning evidence that IL-17 plays a prominent role in neonatal pathology.

# **The role of IL-17 in neonatal sepsis**

The capacity for the host to respond to sepsis is strongly influenced by the patient's developmental age<sup>71–76</sup>. It is commonly accepted that IL-17A is important for neonatal host defense and protective immunity. Physiologic levels of IL-17A produced by type 3 innate lymphoid cells (ILCs) improved resistance to infectious challenge in healthy neonatal mice via microbiota-driven granulocyte-colony stimulating factor (G-CSF) production and neutrophil recruitment<sup>77</sup>. Similarly, reduced whole blood IL-17A in human preterm neonates was associated with an increased risk of bacteremia<sup>20</sup>. IL-17A produced by intestinal  $\gamma \delta T$ cells plays an important role in maintenance and protection of epithelial barriers in the intestinal mucosa78. In contrast, excessive production of IL-17A during disease can be detrimental to the host and lead to organ injury and death. Emerging information implicates pathologic IL-17A signaling during experimental inflammation<sup>79</sup>, sepsis<sup>79–81</sup>, NEC<sup>82</sup>, acute lung injury<sup>83,84</sup>, and brain injury<sup>15,17,85</sup>. The deleterious role of excessive IL-17A, produced by many different cellular sources including γδT cells that can rapidly produce IL-17A via an IL-1R1-dependent pathway<sup>50,86</sup>, is well established in the pathogenesis of severe inflammatory and ischemic-injury models including sepsis<sup>15,80,87</sup>. IL-17A mRNA was increased in the intestine following necrotizing enterocolitis in baboons<sup>88</sup>, and excessive IL-17A impaired enterocyte tight junctions, increased enterocyte apoptosis, and reduced enterocyte proliferation in murine NEC<sup>82</sup>. Thus, excessive IL-17A leads to decreased intestinal integrity that, in turn, increases the likelihood of developing bacteremia, organ injury, and sepsis.

Mortality to murine neonatal sepsis was dependent upon excessive and deleterious IL-17A production by  $\gamma$  ST cells in the lung and gut. Sepsis mortality was significantly attenuated with antibody-mediated transient IL-17 receptor (IL-17R) blockade. Notably, pathologic IL-17A production was dependent upon IL-1R1-mediated signaling and was heavily augmented by IL-18<sup>19</sup>. The importance of discovering the contribution of IL-18 to pathologic IL-17A production in neonates is underscored by studies that showed serum IL-18 varies inversely with age among children $89$ , and preterm neonates exhibited elevated blood IL-18 levels when uninfected, that rose further with infection<sup>90</sup>. Taken together, these data support the paradigm of a developmental age-related propensity to pathologic IL-17A production with stimulation<sup>19,82,85</sup>.

#### **The role of IL-17 in bronchopulmonary dysplasia**

BPD develops in very low birthweight (VLBW; < 1500 grams at birth) infants secondary to abnormal growth of distal lung structures and tissue damage caused by the combination of the underlying disease pathology and injury caused by standard ventilation techniques that are lifesaving in this patient population $91-95$ . Neonates destined to develop BPD demonstrate increased numbers of circulating neutrophils and macrophages, in addition to higher concentrations of leukotrienes, endothelin- $1^{96}$ , IL- $6^{97}$ , and IL-17A $^{91,98}$ . New studies confirm an increased number of TEMs, progenitor Th17, mature Th17, and IL-17+ Treg lymphocytes in placental cord blood samples of extremely preterm human neonates who developed BPD compared to controls<sup>98</sup>. These same investigators previously employed a murine model of *in utero* infection to demonstrate enhanced lung expression of Th17 cells exhibiting intense proinflammatory properties resistant to Treg-mediated suppression<sup>99,100</sup>. The rise in Th17 cells resulted from heightened RORγt expression and transformation of Tregs into Th17-like cells via  $ROR\gamma t$ -mediated mechanisms. This phenotypic alteration of Tregs therefore contributes to both compromised Treg suppressor functions and perpetuation of Th17-mediated proinflammatory responses. The authors concluded that chorioamnionitismediated Th17-biased proinflammatory responses in the preterm infant attenuates immune programing during a critical developmental window resulting in "dysfunctional immune priming". This immune modification ultimately leads to uncontrolled inflammatory processes and tissue damage that increases the risks of morbidity and mortality associated with BPD and chronic lung disease<sup>33</sup>.

IL-17A induces airway epithelial cells to secrete chemokines and cytokines that are also important in protecting the lung against extracellular infections including: (i) CXCL1, CXCL2, IL-6, IL-8, and GM-CSF, which promote neutrophil differentiation in the bone marrow and recruitment of activated cells to the lung epithelium<sup>101–103</sup>, (ii) CCR6, which is associated with trafficking of T, B, and dendritic cells to epithelial sites<sup>104</sup>, (iii) IL-19, which can influence the differentiation of T helper cells into Th2 cells<sup>105</sup>, (iv) CCL28, which enables the migration of IgE-secreting B cells<sup>106</sup>, and (v) Th1 cell-like chemoattractants, which facilitate more effective pathogen clearance when necessary<sup>107,108</sup>. Furthermore, IL-17A contributes to the maintenance of a healthy respiratory epithelial border by promoting the production of adhesion and cell junction molecules through induction of ICAM-1 by airway epithelial cells<sup>11,106,109</sup>. Therefore, tight regulation of IL-17A appears vital for proper maintenance of the respiratory epithelial border, whereas altered production

can injure developing lung tissue resulting in detrimental pulmonary pathologies in our youngest patients.

# **The role of IL-17 in necrotizing enterocolitis**

Necrotizing enterocolitis is a life-threatening gastrointestinal ailment that affects up to 5% of all VLBW infants admitted to the neonatal intensive care unit<sup>110</sup>. Devastatingly, 10% to 50% of neonates diagnosed with NEC will succumb to their illness, while one-third will suffer debilitating long-term complications, such as short-gut syndrome<sup>111</sup>. NEC onset is epidemiologically linked to a "window of susceptibility" with symptoms typically occurring in VLBW infants around 30-32 weeks' corrected gestational age (GA). Therefore, the youngest gestational-aged neonates will develop disease later in life than those who are older at birth<sup>112,113</sup>. Because the incidence of NEC is also inversely proportional to GA at birth<sup>114</sup>, immature host defenses are thought to contribute to its pathogenesis<sup>115</sup>. Heightened baseline expression of Toll-like receptor (TLR)-4 and STAT3 in the intestinal mucosa of premature compared to full-term neonates<sup>116,117</sup>, in addition to changes in microbial colonization and maturation of immune responses have been implicated in this association<sup>115,118</sup>.

Immune homeostasis in the gut is normally maintained by the production of low-levels of IL-17A by resident Th17 lymphocytes and  $ILCs<sup>119</sup>$ . Secretion of IL-17A is influenced by the release of IL-1β and TGF-β from intestinal epithelial cells and functions to inhibit excessive bacterial growth in the gastrointestinal tract<sup>6,120</sup>. Th17 cells are also directly regulated by local Treg lymphocytes to ensure IL-17A is not disproportionally produced under basal conditions<sup>28</sup>. Treg control of Th17 cells is particularly important during the formation of the intestinal microbiome following birth to prevent excessive proinflammatory immune responses as the infant is newly exposed to trillions of microorganisms<sup>28</sup>. Disruption of the normal establishment of the neonatal microbiome through exposure to intrapartum or postpartum antibiotics, however, can alter the proper functioning of ILCs and Th17 lymphocytes. This modification can attenuate neutrophil homeostasis through altered production of G-CSF and IL-17A, which increases the neonate's susceptibility to NEC and/or sepsis by enabling the translocation of virulent microorganisms<sup>118,119</sup>.

During a pathogenic challenge or inflammatory trigger, expression of TLR-4 by the intestinal mucosa is upregulated and accompanied by a notable rise in serum concentrations of IL-6 and prostaglandin E2 (PGE2)<sup>6,121,122</sup>. Together, these adaptions result in preferential differentiation of Th17 lymphocytes with a coinciding drop in Treg numbers $82,111$ . Excessive IL-17A concentrations promote mucosal injury that is commonly associated with NEC, including impaired enterocyte tight junctions, increased enterocyte apoptosis, and reduced proliferation of enterocytes  $82$ . Hence, blocking IL-17A or inducing Treg lymphocyte differentiation can decrease or reverse NEC severity $82$ .

In summary, specific T helper subsets are controlled by interactions between commensal microbes and mucosal immunity<sup>123</sup>. Tonic TLR-4 activation by pathogenic bacteria leads to exaggerated recruitment and activation of Th17 cells but suppression of Tregs, which can alter the normal balance of gut immunity and initiate pathologic responses commonly

associated with necrotizing enterocolitis. Although specific mechanisms involved in IL-17Amediated NEC remains elusive, continued investigation of this pathway remains intriguing. A better understanding of IL-17A-mediated actions may lead to the development of novel pharmacologic therapies or change standard clinical management to improve patient outcomes resulting from necrotizing enterocolitis.

#### **The role of IL-17 in retinopathy of prematurity**

Retinopathy of prematurity is a common, biphasic vasoproliferative disorder of the developing retina that occurs in VLBW, critically ill, and severely growth restricted infants124. ROP consists of initial blunting of retinal vascular growth followed by retinal vessel proliferation around 4-6 weeks of  $age<sup>124</sup>$ . The incidence of ROP is inversely related to gestational age, with increased risk and severity of disease in the youngest, sickest preterm infants. Recently, pre- and post- natal inflammatory conditions, including chorioamnionitis and neonatal sepsis, were shown to be positively correlated to ROP development $^{125,126}$ . Alarmingly, ROP remains a major cause of blindness in infancy despite improved screening and treatment options<sup>90</sup>.

Fetal inflammatory responses have recently been identified as an important instigator of ROP and results from greatly increased levels of IL- $6^{127}$ , but reduced concentrations of IL-17A and IL-18, shortly after birth<sup>90</sup>. Diminished IL-17A levels in preterm infants during critical time points may be responsible for the arrest of retinal angiogenesis, while also predisposing the infant to late onset sepsis<sup>90</sup>.

During retinal neovascularization, however, resident immunocompetent cells of the retina, or microglial cells, secrete proinflammatory cytokines, like IL-17A, when chronically activated. These inflammatory signals stimulate vascular endothelial growth factor (VEGF) production from neuroglial cells, such as Müller and ganglion cells, to promote aberrant growth of retinal vessels 49,124,128–131. This abnormal growth is the hallmark of ROP and can be attenuated in murine models by IL-17A neutralizing antibodies through modulation of the VEGF pathway132. In knockout models of IL-17A, inflammation was also alleviated and tissue repair facilitated by biasing macrophage polarization towards a M2 phenotype. This macrophage transformation leads to the production of high levels of IL-10, which dampens pro-inflammatory responses to enable wound healing<sup>132–135</sup>. In contrast, IL-17 drives macrophage differentiation towards a M1 subtype, facilitating secretion of TNF-α, IL-1β, and IL-6 to enhance pro-angiogenic VEGF pathways and ROP<sup>132</sup>.

Currently, anti-VEGF agents are routinely used to treat vasoproliferative diseases, including ROP<sup>136</sup>. However, a substantial number of eyes do not respond adequately to these specific therapies124,137–139. Development of innovative therapies based on IL-17A neutralization may be an effective treatment for ROP by attenuating microglial cell density, Müller cell gliosis, and ganglion cell loss<sup>124</sup>.

#### **The role of IL-17 in the patent ductus arteriosus**

The ductus arteriosus (ductus; PDA) is vital to the fetus allowing blood to be shunted away from the fetal lungs and into the systemic circulation. In term infants, the ductus constricts

after birth due to an increase in tissue oxygenation and decreased levels of prostaglandin<sup>140,141</sup>. The internal layer of the ductus becomes hypoxic with ductal constriction leading to neointimal cushion formation and extensive vascular remodeling<sup>140,141</sup>. This mechanism of vascular remodeling is similar to the development of atherosclerosis seen after vascular wall injury<sup>140,142</sup>. Cytokines, such as IL-17, play a role in vascular remodeling in atherosclerosis and may similarly mediate remodeling of the ductus<sup>143,144</sup>. IL-17 has also been shown to affect the aorta (large vessel) and the coronaries (small vessels) in different ways revealing that IL-17 has variable expression in different tissues145. Although IL-17 plays a role in vascular remodeling and in expression of prostaglandin, its role in ductal closure is presently unknown.

In contrast to term infants, the ductus may remain patent for a prolonged period in preterm infants, which can result in a shunt that leads to pulmonary over-circulation and systemic hypoperfusion. The ductus in a premature infant is thinner which can decrease vasoconstriction requiring other means of closure including platelet aggregation and thrombus formation<sup>140,142</sup>. IL-17 increases platelet activation and platelet adhesion which may be needed for ductal closure in the premature infant<sup>142,146</sup>. In addition, premature infants are more likely to have a symptomatic PDA if there is a history of chorioamnionitis<sup>147</sup> or history of postnatal inflammation<sup>148</sup> including early onset and late onset sepsis<sup>149,150</sup>. Of note, IL-17 is an important cytokine in inflammation and it in some cells it has been shown to stimulate the increase of prostaglandins, a potent vasodilator of the ductus<sup>146,149,151</sup>. The mechanistic interplay between prostaglandin and IL-17 may affect ductal closure in the setting of sepsis or other significant inflammatory states. IL-17 concentration at physiologic levels may promote vascular remodeling and platelet aggregation assisting ductal closure. However, the concentration at higher levels may induce pathologic ductal dilation due to its effect on prostaglandin production which could be the etiology of inflammatory associated symptomatic PDA. Thus, therapeutic approaches that target attenuation of pathologic IL-17 production require further study prior to implementation in the preterm population.

#### **Pharmacologic interventions targeting IL-17 pathways**

Presently, 2 monoclonal antibodies directed against IL-17A and one against the IL-17A receptor have completed phase III human trials. These agents include: (i) Ixekizumab (LY2439821), a humanized IL-17A specific antibody, (ii) Secukinumab (AIN457), a fully human IL-17A specific antibody<sup>152</sup>, and (iii) Brodalumab (AMG 827), an IL-17A specific antibody152. Because the inhibition of IL-17A may increase the risks of extracellular  $bacterial<sup>153</sup>$  and/or fungal infections, careful attention must be paid when this cytokine is specifically targeted<sup>152</sup>. Adult and animal studies provide important information as we ponder the use of these therapies in neonatal patients. However, our understanding of IL-17A pathways in term and preterm infants remains incomplete and is a barrier to clinical trials that target IL-17 signaling in this unique population.

# **Conclusion**

IL-17 is important for immune surveillance and protection of the neonate. Conversely, pathologically-elevated IL-17 plays a deleterious role in many devastating inflammatory diseases that occur in neonates including sepsis, NEC, and ROP. Targeting excessive IL-17 signaling in the setting of these diseases represents a novel therapeutic strategy that brings hope for improved outcomes in this vulnerable population.

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#### **Abbreviations**





#### **References**

- 1. Rouvier E, Luciani MF, Mattéi MG, Denizot F, Golstein P. CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences, and homologous to a herpesvirus saimiri gene. J Immunol. 1993 Jun 15; 150(12):5445–56. [PubMed: 8390535]
- 2. Waisman A, Hauptmann J, Regen T. The role of IL-17 in CNS diseases. Acta Neuropathol. 2015 May; 129(5):625–37. DOI: 10.1007/s00401-015-1402-7 [PubMed: 25716179]
- 3. Beringer A, Noack M, Miossec P. IL-17 in Chronic Inflammation: From Discovery to Targeting. Trends Mol Med. 2016 Mar; 22(3):230–41. DOI: 10.1016/j.molmed.2016.01.001 [PubMed: 26837266]
- 4. Crome SQ, Wang AY, Levings MK. Translational mini-review series on Th17 cells: function and regulation of human T helper 17 cells in health and disease. Clin Exp Immunol. 2010 Feb; 159(2): 109–19. DOI: 10.1111/j.1365-2249.2009.04037.x [PubMed: 19912252]
- 5. Cua DJ, Tato CM. Innate IL-17-producing cells: the sentinels of the immune system. Nat Rev Immunol. 2010 Jul; 10(7):479–89. DOI: 10.1038/nri2800 [PubMed: 20559326]
- 6. Bystrom J, Al-Adhoubi N, Al-Bogami M, Jawad AS, Mageed RA. Th17 lymphocytes in respiratory syncytial virus infection. Viruses. 2013 Mar 5; 5(3):777–91. DOI: 10.3390/v5030777 [PubMed: 23462708]
- 7. Huang H, Saravia J, You D, Shaw AJ, Cormier SA. Impaired gamma delta T cell-derived IL-17A and inflammasome activation during early respiratory syncytial virus infection in infants. Immunol Cell Biol. 2015 Feb; 93(2):126–35. DOI: 10.1038/icb.2014.79 [PubMed: 25267484]
- 8. Molet S, Hamid Q, Davoine F, Nutku E, Taha R, Pagé N, Olivenstein R, Elias J, Chakir J. IL-17 is increased in asthmatic airways and induces human bronchial fibroblasts to produce cytokines. J Allergy Clin Immunol. 2001 Sep; 108(3):430–8. DOI: 10.1067/mai.2001.117929 [PubMed: 11544464]
- 9. Kleinschek MA, Boniface K, Sadekova S, Grein J, Murphy EE, Turner SP, Raskin L, Desai B, Faubion WA, de Waal Malefyt R, Pierce RH, McClanahan T, Kastelein RA. Circulating and gutresident human Th17 cells express CD161 and promote intestinal inflammation. J Exp Med. 2009 Mar 16; 206(3):525–34. DOI: 10.1084/jem.20081712 [PubMed: 19273624]
- 10. Kobayashi T, Okamoto S, Hisamatsu T, Kamada N, Chinen H, Saito R, Kitazume MT, Nakazawa A, Sugita A, Koganei K, Isobe K, Hibi T. IL23 differentially regulates the Th1/Th17 balance in ulcerative colitis and Crohn's disease. Gut. 2008 Dec; 57(12):1682–9. DOI: 10.1136/gut. 2007.135053 [PubMed: 18653729]
- 11. Tsai HC, Velichko S, Hung LY, Wu R. IL-17A and Th17 cells in lung inflammation: an update on the role of Th17 cell differentiation and IL-17R signaling in host defense against infection. Clin Dev Immunol. 2013; 2013:267971.doi: 10.1155/2013/267971 [PubMed: 23956759]
- 12. Croxford AL, Karbach S, Kurschus FC, Wörtge S, Nikolaev A, Yogev N, Klebow S, Schüler R, Reissig S, Piotrowski C, Brylla E, Bechmann I, Scheller J, Rose-John S, Wunderlich FT, Münzel T, von Stebut E, Waisman A. IL-6 regulates neutrophil microabscess formation in IL-17A-driven psoriasiform lesions. J Invest Dermatol. 2014 Mar; 134(3):728–35. DOI: 10.1038/jid.2013.404 [PubMed: 24067382]
- 13. Beurel E, Harrington LE, Jope RS. Inflammatory T helper 17 cells promote depression-like behavior in mice. Biol Psychiatry. 2013 Apr 1; 73(7):622–30. DOI: 10.1016/j.biopsych. 2012.09.021 [PubMed: 23174342]
- 14. Liu Y, Ho RC, Mak A. The role of interleukin (IL)-17 in anxiety and depression of patients with rheumatoid arthritis. Int J Rheum Dis. 2012 Apr; 15(2):183–7. DOI: 10.1111/j.1756-185X. 2011.01673.x [PubMed: 22462422]

- 15. Gelderblom M, Weymar A, Bernreuther C, Velden J, Arunachalam P, Steinbach K, Orthey E, Arumugam TV, Leypoldt F, Simova O, Thom V, Friese MA, Prinz I, Hölscher C, Glatzel M, Korn T, Gerloff C, Tolosa E, Magnus T. Neutralization of the IL-17 axis diminishes neutrophil invasion and protects from ischemic stroke. Blood. 2012 Nov 1; 120(18):3793–802. DOI: 10.1182/ blood-2012-02-412726 [PubMed: 22976954]
- 16. Li GZ, Zhong D, Yang LM, Sun B, Zhong ZH, Yin YH, Cheng J, Yan BB, Li HL. Expression of interleukin-17 in ischemic brain tissue. Scand J Immunol. 2005 Nov; 62(5):481–6. DOI: 10.1111/j. 1365-3083.2005.01683.x [PubMed: 16305645]
- 17. Shichita T, Sugiyama Y, Ooboshi H, Sugimori H, Nakagawa R, Takada I, Iwaki T, Okada Y, Iida M, Cua DJ, Iwakura Y, Yoshimura A. Pivotal role of cerebral interleukin-17-producing gammadeltaT cells in the delayed phase of ischemic brain injury. Nat Med. 2009 Aug; 15(8):946– 50. DOI: 10.1038/nm.1999 [PubMed: 19648929]
- 18. Wang DD, Zhao YF, Wang GY, Sun B, Kong QF, Zhao K, Zhang Y, Wang JH, Liu YM, Mu LL, Wang DS, Li HL. IL-17 potentiates neuronal injury induced by oxygen-glucose deprivation and affects neuronal IL-17 receptor expression. J Neuroimmunol. 2009 Jul 25; 212(1–2):17–25. DOI: 10.1016/j.jneuroim.2009.04.007 [PubMed: 19457561]
- 19. Zhang J, Mao X, Zhou T, Cheng X, Lin Y. IL-17A contributes to brain ischemia reperfusion injury through calpain-TRPC6 pathway in mice. Neuroscience. 2014 Aug 22.274:419–28. DOI: 10.1016/ j.neuroscience.2014.06.001 [PubMed: 24928352]
- 20. Schelonka RL, Maheshwari A, Carlo WA, Taylor S, Hansen NI, Schendel DE, Thorsen P, Skogstrand K, Hougaard DM, Higgins RD, NICHD Neonatal Research Network. T cell cytokines and the risk of blood stream infection in extremely low birth weight infants. Cytokine. 2011 Feb; 53(2):249–55. DOI: 10.1016/j.cyto.2010.11.003 [PubMed: 21145756]
- 21. Maddux AB, Douglas IS. Is the developmentally immature immune response in paediatric sepsis a recapitulation of immune tolerance? Immunology. 2015 May; 145(1):1–10. DOI: 10.1111/imm. 12454 [PubMed: 25691226]
- 22. Adkins B, Leclerc C, Marshall-Clarke S. Neonatal adaptive immunity comes of age. Nat Rev Immunol. 2004 Jul; 4(7):553–64. DOI: 10.1038/nri1394 [PubMed: 15229474]
- 23. Schaub B, Liu J, Schleich I, Höppler S, Sattler C, von Mutius E. Impairment of T helper and T regulatory cell responses at birth. Allergy. 2008 Nov; 63(11):1438–47. DOI: 10.1111/j. 1398-9995.2008.01685.x [PubMed: 18925880]
- 24. Hirota K, Yoshitomi H, Hashimoto M, Maeda S, Teradaira S, Sugimoto N, Yamaguchi T, Nomura T, Ito H, Nakamura T, Sakaguchi N, Sakaguchi S. Preferential recruitment of CCR6-expressing Th17 cells to inflamed joints via CCL20 in rheumatoid arthritis and its animal model. J Exp Med. 2007 Nov 26; 204(12):2803–12. DOI: 10.1084/jem.20071397 [PubMed: 18025126]
- 25. El-Behi M, Ciric B, Dai H, Yan Y, Cullimore M, Safavi F, Zhang GX, Dittel BN, Rostami A. The encephalitogenicity of T(H)17 cells is dependent on IL-1- and IL-23-induced production of the cytokine GM-CSF. Nat Immunol. 2011 Jun; 12(6):568–75. DOI: 10.1038/ni.2031 [PubMed: 21516111]
- 26. Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B. TGFbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. Immunity. 2006 Feb; 24(2):179–89. DOI: 10.1016/j.immuni.2006.01.001 [PubMed: 16473830]
- 27. de Jong E, Suddason T, Lord GM. Translational mini-review series on Th17 cells: development of mouse and human T helper 17 cells. Clin Exp Immunol. 2010 Feb; 159(2):148–58. DOI: 10.1111/j.1365-2249.2009.04041.x [PubMed: 19912248]
- 28. Sehrawat S, Rouse BT. Interplay of Regulatory T Cell and Th17 Cells during Infectious Diseases in Humans and Animals. Front Immunol. 2017 Apr 3.8:341.doi: 10.3389/fimmu.2017.00341 [PubMed: 28421070]
- 29. Ziegler SF, Buckner JH. FOXP3 and the regulation of Treg/Th17 differentiation. Microbes Infect. 2009 Apr; 11(5):594–8. DOI: 10.1016/j.micinf.2009.04.002 [PubMed: 19371792]
- 30. Zhou L, Lopes JE, Chong MM, Ivanov II, Min R, Victora GD, Shen Y, Du J, Rubtsov YP, Rudensky AY, Ziegler SF, Littman DR. TGF-beta-induced Foxp3 inhibits T(H)17 cell differentiation by antagonizing RORgammat function. Nature. 2008 May 8; 453(7192):236–40. DOI: 10.1038/nature06878 [PubMed: 18368049]

- 31. Noack M, Miossec P. Th17 and regulatory T cell balance in autoimmune and inflammatory diseases. Autoimmun Rev. 2014 Jun; 13(6):668–77. DOI: 10.1016/j.autrev.2013.12.004 [PubMed: 24418308]
- 32. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, Weiner HL, Kuchroo VK. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. Nature. 2006 May 11; 441(7090):235–8. [PubMed: 16648838]
- 33. Rito DC, Viehl LT, Buchanan PM, Haridas S, Koenig JM. Augmented Th17-type immune responses in preterm neonates exposed to histologic chorioamnionitis. Pediatr Res. 2017 Apr; 81(4):639–645. DOI: 10.1038/pr.2016.254 [PubMed: 27870827]
- 34. Weaver CT, Hatton RD. Interplay between the TH17 and TReg cell lineages: a (co-) evolutionary perspective. Nat Rev Immunol. 2009 Dec; 9(12):883–9. DOI: 10.1038/nri2660 [PubMed: 19935807]
- 35. Blatner NR, Mulcahy MF, Dennis KL, Scholtens D, Bentrem DJ, Phillips JD, Ham S, Sandall BP, Khan MW, Mahvi DM, Halverson AL, Stryker SJ, Boller AM, Singal A, Sneed RK, Sarraj B, Ansari MJ, Oft M, Iwakura Y, Zhou L, Bonertz A, Beckhove P, Gounari F, Khazaie K. Expression of RORγt marks a pathogenic regulatory T cell subset in human colon cancer. Sci Transl Med. 2012 Dec 12.4(164):164ra159.doi: 10.1126/scitranslmed.3004566
- 36. Yang XO, Nurieva R, Martinez GJ, Kang HS, Chung Y, Pappu BP, Shah B, Chang SH, Schluns KS, Watowich SS, Feng XH, Jetten AM, Dong C. Molecular antagonism and plasticity of regulatory and inflammatory T cell programs. Immunity. 2008 Jul 18; 29(1):44–56. DOI: 10.1016/ j.immuni.2008.05.007 [PubMed: 18585065]
- 37. Zhang X, Mozeleski B, Lemoine S, Dériaud E, Lim A, Zhivaki D, Azria E, Le Ray C, Roguet G, Launay O, Vanet A, Leclerc C, Lo-Man R. CD4 T cells with effector memory phenotype and function develop in the sterile environment of the fetus. Sci Transl Med. 2014 May 28.6(238): 238ra72.doi: 10.1126/scitranslmed.3008748
- 38. Mills KH. Induction, function and regulation of IL-17-producing T cells. Eur J Immunol. 2008 Oct; 38(10):2636–49. DOI: 10.1002/eji.200838535 [PubMed: 18958872]
- 39. Weaver CT, Hatton RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. Annu Rev Immunol. 2007; 25:821–52. DOI: 10.1146/ annurev.immunol.25.022106.141557 [PubMed: 17201677]
- 40. Caron JE, La Pine TR, Augustine NH, Martins TB, Kumánovics A, Hill HR. Severely depressed interleukin-17 production by human neonatal mononuclear cells. Pediatr Res. 2014 Dec; 76(6): 522–7. DOI: 10.1038/pr.2014.133 [PubMed: 25192396]
- 41. Lavoie PM, Huang Q, Jolette E, Whalen M, Nuyt AM, Audibert F, Speert DP, Lacaze-Masmonteil T, Soudeyns H, Kollmann TR. Profound lack of interleukin (IL)-12/IL-23p40 in neonates born early in gestation is associated with an increased risk of sepsis. J Infect Dis. 2010 Dec 1; 202(11): 1754–63. DOI: 10.1086/657143 [PubMed: 20977341]
- 42. Chen L, He Z, Slinger E, Bongers G, Lapenda TLS, Pacer ME, Jiao J, Beltrao MF, Soto AJ, Harpaz N, Gordon RE, Ochando JC, Oukka M, Iuga AC, Chensue SW, Blander JM, Furtado GC, Lira SA. IL-23 activates innate lymphoid cells to promote neonatal intestinal pathology. Mucosal Immunol. 2015 Mar; 8(2):390–402. DOI: 10.1038/mi.2014.77 [PubMed: 25160819]
- 43. Li M, Wang B, Sun X, Tang Y, Wei X, Ge B, Tang Y, Deng Y, He C, Yuan J, Li X. Upregulation of Intestinal Barrier Function in Mice with DSS-Induced Colitis by a Defined Bacterial Consortium Is Associated with Expansion of IL-17A Producing Gamma Delta T Cells. Front Immunol. 2017 Jul 12.8:824.doi: 10.3389/fimmu.2017.00824 [PubMed: 28747917]
- 44. Thapa P, Manso B, Chung JY, Romera Arocha S, Xue HH, Angelo DBS, Shapiro VS. The differentiation of ROR-γt expressing iNKT17 cells is orchestrated by Runx1. Sci Rep. 2017 Aug 1.7(1):7018.doi: 10.1038/s41598-017-07365-8 [PubMed: 28765611]
- 45. Milosavljevic N, Gazdic M, Simovic Markovic B, Arsenijevic A, Nurkovic J, Dolicanin Z, Djonov V, Lukic ML, Volarevic V. Mesenchymal stem cells attenuate acute liver injury by altering ratio between interleukin 17 producing and regulatory natural killer T cells. Liver Transpl. 2017 Aug; 23(8):1040–1050. DOI: 10.1002/lt.24784 [PubMed: 28481005]
- 46. Hu S, He W, Du X, Yang J, Wen Q, Zhong XP, Ma L. IL-17 Production of Neutrophils Enhances Antibacteria Ability but Promotes Arthritis Development During Mycobacterium tuberculosis

Infection. EBioMedicine. 2017 Aug 9. pii: S2352-3964(17)30303-1. doi: 10.1016/j.ebiom. 2017.08.001

- 47. Park SW, Kim M, Brown KM, D'Agati VD, Lee HT. Paneth cell-derived interleukin-17A causes multiorgan dysfunction after hepatic ischemia and reperfusion injury. Hepatology. 2011 May; 53(5):1662–75. DOI: 10.1002/hep.24253 [PubMed: 21360570]
- 48. Song X, Qian Y. IL-17 family cytokines mediated signaling in the pathogenesis of inflammatory diseases. Cell Signal. 2013 Dec; 25(12):2335–47. DOI: 10.1016/j.cellsig.2013.07.021 [PubMed: 23917206]
- 49. Grigsby JG, Cardona SM, Pouw CE, Muniz A, Mendiola AS, Tsin AT, Allen DM, Cardona AE. The role of microglia in diabetic retinopathy. J Ophthalmol. 2014; 2014:705783.doi: 10.1155/2014/705783 [PubMed: 25258680]
- 50. Duan J, Chung H, Troy E, Kasper DL. Microbial colonization drives expansion of IL-1 receptor 1 expressing and IL-17-producing gamma/delta T cells. Cell Host Microbe. 2010 Feb 18; 7(2):140– 50. DOI: 10.1016/j.chom.2010.01.005 [PubMed: 20159619]
- 51. Yoshiga Y, Goto D, Segawa S, Ohnishi Y, Matsumoto I, Ito S, Tsutsumi A, Taniguchi M, Sumida T. Invariant NKT cells produce IL-17 through IL-23-dependent and -independent pathways with potential modulation of Th17 response in collagen-induced arthritis. Int J Mol Med. 2008 Sep; 22(3):369–74. [PubMed: 18698497]
- 52. Riol-Blanco L, Lazarevic V, Awasthi A, Mitsdoerffer M, Wilson BS, Croxford A, Waisman A, Kuchroo VK, Glimcher LH, Oukka M. IL-23 receptor regulates unconventional IL-17-producing T cells that control bacterial infections. J Immunol. 2010 Feb 15; 184(4):1710–20. DOI: 10.4049/ jimmunol.0902796 [PubMed: 20083652]
- 53. Rachitskaya AV, Hansen AM, Horai R, Li Z, Villasmil R, Luger D, Nussenblatt RB, Caspi RR. Cutting edge: NKT cells constitutively express IL-23 receptor and RORgammat and rapidly produce IL-17 upon receptor ligation in an IL-6-independent fashion. J Immunol. 2008 Apr 15; 180(8):5167–71. [PubMed: 18390697]
- 54. Sutton CE, Lalor SJ, Sweeney CM, Brereton CF, Lavelle EC, Mills KH. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. Immunity. 2009 Aug 21; 31(2):331–41. DOI: 10.1016/j.immuni.2009.08.001 [PubMed: 19682929]
- 55. Veldhoen M, Hirota K, Westendorf AM, Buer J, Dumoutier L, Renauld JC, Stockinger B. The aryl hydrocarbon receptor links TH17-cell-mediated autoimmunity to environmental toxins. Nature. 2008 May 1; 453(7191):106–9. DOI: 10.1038/nature06881 [PubMed: 18362914]
- 56. Onishi RM, Gaffen SL. Interleukin-17 and its target genes: mechanisms of interleukin-17 function in disease. Immunology. 2010 Mar; 129(3):311–21. DOI: 10.1111/j.1365-2567.2009.03240.x [PubMed: 20409152]
- 57. Fossiez F, Djossou O, Chomarat P, Flores-Romo L, Ait-Yahia S, Maat C, Pin JJ, Garrone P, Garcia E, Saeland S, Blanchard D, Gaillard C, Das Mahapatra B, Rouvier E, Golstein P, Banchereau J, Lebecque S. T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines. J Exp Med. 1996 Jun 1; 183(6):2593–603. [PubMed: 8676080]
- 58. Schwarzenberger P, La Russa V, Miller A, Ye P, Huang W, Zieske A, Nelson S, Bagby GJ, Stoltz D, Mynatt RL, Spriggs M, Kolls JK. IL-17 stimulates granulopoiesis in mice: use of an alternate, novel gene therapy-derived method for in vivo evaluation of cytokines. J Immunol. 1998 Dec 1; 161(11):6383–9. [PubMed: 9834129]
- 59. Hoshino H, Laan M, Sjöstrand M, Lötvall J, Skoogh BE, Linden A. Increased elastase and myeloperoxidase activity associated with neutrophil recruitment by IL-17 in airways in vivo. J Allergy Clin Immunol. 2000 Jan; 105(1 Pt 1):143–9. [PubMed: 10629464]
- 60. Lu YJ, Gross J, Bogaert D, Finn A, Bagrade L, Zhang Q, Kolls JK, Srivastava A, Lundgren A, Forte S, Thompson CM, Harney KF, Anderson PW, Lipsitch M, Malley R. Interleukin-17A mediates acquired immunity to pneumococcal colonization. PLoS Pathog. 2008 Sep 19.4(9):e1000159.doi: 10.1371/journal.ppat.1000159 [PubMed: 18802458]
- 61. Abi Abdallah DS, Egan CE, Butcher BA, Denkers EY. Mouse neutrophils are professional antigenpresenting cells programmed to instruct Th1 and Th17 T-cell differentiation. Int Immunol. 2011 May; 23(5):317–26. DOI: 10.1093/intimm/dxr007 [PubMed: 21422151]

- 62. Sergejeva S, Ivanov S, Lötvall J, Lindén A. Interleukin-17 as a recruitment and survival factor for airway macrophages in allergic airway inflammation. Am J Respir Cell Mol Biol. 2005 Sep; 33(3): 248–53. DOI: 10.1165/rcmb.2004-0213OC [PubMed: 15901616]
- 63. Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, Fouser LA. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. J Exp Med. 2006 Oct 2; 203(10):2271–9. DOI: 10.1084/jem.20061308 [PubMed: 16982811]
- 64. Ouyang W, Kolls JK, Zheng Y. The biological functions of T helper 17 cell effector cytokines in inflammation. Immunity. 2008 Apr; 28(4):454–67. DOI: 10.1016/j.immuni.2008.03.004 [PubMed: 18400188]
- 65. Crome SQ, Wang AY, Kang CY, Levings MK. The role of retinoic acid-related orphan receptor variant 2 and IL-17 in the development and function of human CD4+ T cells. Eur J Immunol. 2009 Jun; 39(6):1480–93. DOI: 10.1002/eji.200838908 [PubMed: 19449310]
- 66. Ishigame H, Kakuta S, Nagai T, Kadoki M, Nambu A, Komiyama Y, Fujikado N, Tanahashi Y, Akitsu A, Kotaki H, Sudo K, Nakae S, Sasakawa C, Iwakura Y. Differential roles of interleukin-17A and -17F in host defense against mucoepithelial bacterial infection and allergic responses. Immunity. 2009 Jan 16; 30(1):108–19. DOI: 10.1016/j.immuni.2008.11.009 [PubMed: 19144317]
- 67. Huang W, Na L, Fidel PL, Schwarzenberger P. Requirement of interleukin-17A for systemic anti-Candida albicans host defense in mice. J Infect Dis. 2004 Aug 1; 190(3):624–31. DOI: 10.1086/422329 [PubMed: 15243941]
- 68. Happel KI, Zheng M, Young E, Quinton LJ, Lockhart E, Ramsay AJ, Shellito JE, Schurr JR, Bagby GJ, Nelson S, Kolls JK. Cutting edge: roles of Toll-like receptor 4 and IL-23 in IL-17 expression in response to Klebsiella pneumoniae infection. J Immunol. 2003 May 1; 170(9):4432– 6. [PubMed: 12707317]
- 69. Haas JD, Ravens S, Düber S, Sandrock I, Oberdörfer L, Kashani E, Chennupati V, Föhse L, Naumann R, Weiss S, Krueger A, Förster R, Prinz I. Development of interleukin-17-producing γδ T cells is restricted to a functional embryonic wave. Immunity. 2012 Jul 27; 37(1):48–59. DOI: 10.1016/j.immuni.2012.06.003 [PubMed: 22770884]
- 70. Stoll BJ, Hansen NI, Bell EF, Shankaran S, Laptook AR, Walsh MC, Hale EC, Newman NS, Schibler K, Carlo WA, Kennedy KA, Poindexter BB, Finer NN, Ehrenkranz RA, Duara S, Sánchez PJ, O'Shea TM, Goldberg RN, Van Meurs KP, Faix RG, Phelps DL, Frantz ID 3rd, Watterberg KL, Saha S, Das A, Higgins RD, Eunice Kennedy Shriver National Institute of Child Health and Human Development Neonatal Research Network. Neonatal outcomes of extremely preterm infants from the NICHD Neonatal Research Network. Pediatrics. 2010 Sep; 126(3):443–56. DOI: 10.1542/peds.2009-2959 [PubMed: 20732945]
- 71. Wynn JL, Guthrie SO, Wong HR, Lahni P, Ungaro R, Lopez MC, Baker HV, Moldawer LL. Postnatal Age Is a Critical Determinant of the Neonatal Host Response to Sepsis. Mol Med. 2015 Jun 2.21:496–504. DOI: 10.2119/molmed.2015.00064 [PubMed: 26052715]
- 72. Wynn JL, Cvijanovich NZ, Allen GL, Thomas NJ, Freishtat RJ, Anas N, Meyer K, Checchia PA, Lin R, Shanley TP, Bigham MT, Banschbach S, Beckman E, Wong HR. The influence of developmental age on the early transcriptomic response of children with septic shock. Mol Med. 2011; 17(11–12):1146–56. DOI: 10.2119/molmed.2011.00169 [PubMed: 21738952]
- 73. Wynn J, Cornell TT, Wong HR, Shanley TP, Wheeler DS. The host response to sepsis and developmental impact. Pediatrics. 2010 May; 125(5):1031–41. DOI: 10.1542/peds.2009-3301 [PubMed: 20421258]
- 74. Wynn JL, Scumpia PO, Winfield RD, Delano MJ, Kelly-Scumpia K, Barker T, Ungaro R, Levy O, Moldawer LL. Defective innate immunity predisposes murine neonates to poor sepsis outcome but is reversed by TLR agonists. Blood. 2008 Sep 1; 112(5):1750–8. DOI: 10.1182/ blood-2008-01-130500 [PubMed: 18591384]
- 75. Wynn JL, Scumpia PO, Delano MJ, O'Malley KA, Ungaro R, Abouhamze A, Moldawer LL. Increased mortality and altered immunity in neonatal sepsis produced by generalized peritonitis. Shock. 2007 Dec; 28(6):675–683. DOI: 10.1097/SHK.0b013e3180556d09 [PubMed: 17621256]
- 76. Raymond SL, López MC, Baker HV, Larson SD, Efron PA, Sweeney TE, Khatri P, Moldawer LL, Wynn JL. Unique transcriptomic response to sepsis is observed among patients of different age

groups. PLoS One. 2017 Sep 8.12(9):e0184159.doi: 10.1371/journal.pone.0184159 [PubMed: 28886074]

- 77. Deshmukh HS, Liu Y, Menkiti OR, Mei J, Dai N, O'Leary CE, Oliver PM, Kolls JK, Weiser JN, Worthen GS. The microbiota regulates neutrophil homeostasis and host resistance to Escherichia coli K1 sepsis in neonatal mice. Nat Med. 2014 May; 20(5):524–30. DOI: 10.1038/nm.3542 [PubMed: 24747744]
- 78. Lee JS, Tato CM, Joyce-Shaikh B, Gulen MF, Cayatte C, Chen Y, Blumenschein WM, Judo M, Ayanoglu G, McClanahan TK, Li X, Cua DJ. Interleukin-23-Independent IL-17 Production Regulates Intestinal Epithelial Permeability. Immunity. 2015 Oct 20; 43(4):727–38. DOI: 10.1016/ j.immuni.2015.09.003 [PubMed: 26431948]
- 79. Wynn JL, Wilson CS, Hawiger J, Scumpia PO, Marshall AF, Liu JH, Zharkikh I, Wong HR, Lahni P, Benjamin JT, Plosa EJ, Weitkamp JH, Sherwood ER, Moldawer LL, Ungaro R, Baker HV, Lopez MC, McElroy SJ, Colliou N, Mohamadzadeh M, Moore DJ. Targeting IL-17A attenuates neonatal sepsis mortality induced by IL-18. Proc Natl Acad Sci U S A. 2016 May 10; 113(19):E2627–35. DOI: 10.1073/pnas.1515793113 [PubMed: 27114524]
- 80. Flierl MA, Rittirsch D, Gao H, Hoesel LM, Nadeau BA, Day DE, Zetoune FS, Sarma JV, Huber-Lang MS, Ferrara JL, Ward PA. Adverse functions of IL-17A in experimental sepsis. FASEB J. 2008 Jul; 22(7):2198–205. DOI: 10.1096/fj.07-105221 [PubMed: 18299333]
- 81. Li J, Zhang Y, Lou J, Zhu J, He M, Deng X, Cai Z. Neutralisation of peritoneal IL-17A markedly improves the prognosis of severe septic mice by decreasing neutrophil infiltration and proinflammatory cytokines. PLoS One. 2012; 7(10):e46506.doi: 10.1371/journal.pone.0046506 [PubMed: 23056325]
- 82. Egan CE, Sodhi CP, Good M, Lin J, Jia H, Yamaguchi Y, Lu P, Ma C, Branca MF, Weyandt S, Fulton WB, Niño DF, Prindle T Jr, Ozolek JA, Hackam DJ. Toll-like receptor 4-mediated lymphocyte influx induces neonatal necrotizing enterocolitis. J Clin Invest. 2016 Feb; 126(2):495– 508. DOI: 10.1172/JCI83356 [PubMed: 26690704]
- 83. Mikacenic C, Hansen EE, Radella F, Gharib SA, Stapleton RD, Wurfel MM. Interleukin-17A Is Associated With Alveolar Inflammation and Poor Outcomes in Acute Respiratory Distress Syndrome. Crit Care Med. 2016 Mar; 44(3):496–502. DOI: 10.1097/CCM.0000000000001409 [PubMed: 26540401]
- 84. Li Q, Gu Y, Tu Q, Wang K, Gu X, Ren T. Blockade of Interleukin-17 Restrains the Development of Acute Lung Injury. Scand J Immunol. 2016 Mar; 83(3):203–11. DOI: 10.1111/sji.12408 [PubMed: 26709006]
- 85. Choi GB, Yim YS, Wong H, Kim S, Kim H, Kim SV, Hoeffer CA, Littman DR, Huh JR. The maternal interleukin-17a pathway in mice promotes autism-like phenotypes in offspring. Science. 2016 Feb 26; 351(6276):933–9. DOI: 10.1126/science.aad0314 [PubMed: 26822608]
- 86. Paget C, Chow MT, Gherardin NA, Beavis PA, Uldrich AP, Duret H, Hassane M, Souza-Fonseca-Guimaraes F, Mogilenko DA, Staumont-Sallé D, Escalante NK, Hill GR, Neeson P, Ritchie DS, Dombrowicz D, Mallevaey T, Trottein F, Belz GT, Godfrey DI, Smyth MJ. CD3bright signals on γδ T cells identify IL-17A-producing Vγ6Vδ1+ T cells. Immunol Cell Biol. 2015 Feb; 93(2): 198–212. DOI: 10.1038/icb.2014.94 [PubMed: 25385067]
- 87. Li L, Huang L, Vergis AL, Ye H, Bajwa A, Narayan V, Strieter RM, Rosin DL, Okusa MD. IL-17 produced by neutrophils regulates IFN-gamma-mediated neutrophil migration in mouse kidney ischemia-reperfusion injury. J Clin Invest. 2010 Jan; 120(1):331–42. DOI: 10.1172/JCI38702 [PubMed: 20038794]
- 88. Namachivayam K, Blanco CL, MohanKumar K, Jagadeeswaran R, Vasquez M, McGill-Vargas L, Garzon SA, Jain SK, Gill RK, Freitag NE, Weitkamp JH, Seidner SR, Maheshwari A. Smad7 inhibits autocrine expression of TGF-β2 in intestinal epithelial cells in baboon necrotizing enterocolitis. Am J Physiol Gastrointest Liver Physiol. 2013 Jan 15; 304(2):G167–80. DOI: 10.1152/ajpgi.00141.2012 [PubMed: 23154975]
- 89. Kleiner G, Marcuzzi A, Zanin V, Monasta L, Zauli G. Cytokine levels in the serum of healthy subjects. Mediators Inflamm. 2013; 2013:434010.doi: 10.1155/2013/434010 [PubMed: 23533306]
- 90. Sood BG, Shankaran S, Schelonka RL, Saha S, Benjamin DK Jr, Sánchez PJ, Adams-Chapman I, Stoll BJ, Thorsen P, Skogstrand K, Ehrenkranz RA, Hougaard DM, Goldberg RN, Tyson JE, Das A, Higgins RD, Carlo WA, Eunice Kennedy Shriver National Institute of Child Health and Human

Development Neonatal Research Network. Cytokine profiles of preterm neonates with fungal and bacterial sepsis. Pediatr Res. 2012 Aug; 72(2):212–20. DOI: 10.1038/pr.2012.56 [PubMed: 22562288]

- 91. Groneck P, Götze-Speer B, Oppermann M, Eiffert H, Speer CP. Association of pulmonary inflammation and increased microvascular permeability during the development of bronchopulmonary dysplasia: a sequential analysis of inflammatory mediators in respiratory fluids of high-risk preterm neonates. Pediatrics. 1994 May; 93(5):712–8. [PubMed: 8165067]
- 92. Merritt TA, Cochrane CG, Holcomb K, Bohl B, Hallman M, Strayer D, Edwards DK 3rd, Gluck L. Elastase and alpha 1-proteinase inhibitor activity in tracheal aspirates during respiratory distress syndrome. Role of inflammation in the pathogenesis of bronchopulmonary dysplasia. J Clin Invest. 1983 Aug; 72(2):656–66. DOI: 10.1172/JCI111015 [PubMed: 6603478]
- 93. Bry K, Hallman M, Teramo K, Waffarn F, Lappalainen U. Granulocyte-macrophage colonystimulating factor in amniotic fluid and in airway specimens of newborn infants. Pediatr Res. 1997 Jan; 41(1):105–9. DOI: 10.1203/00006450-199704001-00638 [PubMed: 8979297]
- 94. Paananen R, Husa AK, Vuolteenaho R, Herva R, Kaukola T, Hallman M. Blood cytokines during the perinatal period in very preterm infants: relationship of inflammatory response and bronchopulmonary dysplasia. J Pediatr. 2009 Jan; 154(1):39–43.e3. DOI: 10.1016/j.jpeds. 2008.07.012 [PubMed: 18760808]
- 95. Gotsch F, Romero R, Kusanovic JP, Mazaki-Tovi S, Pineles BL, Erez O, Espinoza J, Hassan SS. The fetal inflammatory response syndrome. Clin Obstet Gynecol. 2007 Sep; 50(3):652–83. DOI: 10.1097/GRF.0b013e31811ebef6 [PubMed: 17762416]
- 96. Niu JO, Munshi UK, Siddiq MM, Parton LA. Early increase in endothelin-1 in tracheal aspirates of preterm infants: correlation with bronchopulmonary dysplasia. J Pediatr. 1998 Jun; 132(6):965–70. [PubMed: 9627587]
- 97. Mittendorf R, Covert R, Montag AG, el Masri W, Muraskas J, Lee KS, Pryde PG. Special relationships between fetal inflammatory response syndrome and bronchopulmonary dysplasia in neonates. J Perinat Med. 2005; 33(5):428–34. DOI: 10.1515/JPM.2005.076 [PubMed: 16238538]
- 98. Gleditsch DD, Shornick LP, Van Steenwinckel J, Gressens P, Weisert RP, Koenig JM. Maternal inflammation modulates infant immune response patterns to viral lung challenge in a murine model. Pediatr Res. 2014 Jul; 76(1):33–40. DOI: 10.1038/pr.2014.57 [PubMed: 24727945]
- 99. Miao J, Zhang K, Lv M, Li Q, Zheng Z, Han Q, Guo N, Fan C, Zhu P. Circulating Th17 and Th1 cells expressing CD161 are associated with disease activity in rheumatoid arthritis. Scand J Rheumatol. 2014; 43(3):194–201. DOI: 10.3109/03009742.2013.846407 [PubMed: 24392804]
- 100. Basdeo SA, Moran B, Cluxton D, Canavan M, McCormick J, Connolly M, Orr C, Mills KH, Veale DJ, Fearon U, Fletcher JM. Polyfunctional, Pathogenic CD161+ Th17 Lineage Cells Are Resistant to Regulatory T Cell-Mediated Suppression in the Context of Autoimmunity. J Immunol. 2015 Jul 15; 195(2):528–40. DOI: 10.4049/jimmunol.1402990 [PubMed: 26062995]
- 101. Laan M, Prause O, Miyamoto M, Sjöstrand M, Hytönen AM, Kaneko T, Lötvall J, Lindén A. A role of GM-CSF in the accumulation of neutrophils in the airways caused by IL-17 and TNFalpha. Eur Respir J. 2003 Mar; 21(3):387–93. [PubMed: 12661990]
- 102. Ye P, Rodriguez FH, Kanaly S, Stocking KL, Schurr J, Schwarzenberger P, Oliver P, Huang W, Zhang P, Zhang J, Shellito JE, Bagby GJ, Nelson S, Charrier K, Peschon JJ, Kolls JK. Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. J Exp Med. 2001 Aug 20; 194(4):519–27. [PubMed: 11514607]
- 103. Lindén A, Laan M, Anderson GP. Neutrophils, interleukin-17A and lung disease Neutrophils, interleukin-17A and lung disease. Eur Respir J. 2005 Jan; 25(1):159–72. DOI: 10.1183/09031936.04.00032904 [PubMed: 15640338]
- 104. Singh SP, Zhang HH, Foley JF, Hedrick MN, Farber JM. Human T cells that are able to produce IL-17 express the chemokine receptor CCR6. J Immunol. 2008 Jan 1; 180(1):214–21. [PubMed: 18097022]
- 105. Liao SC, Cheng YC, Wang YC, Wang CW, Yang SM, Yu CK, Shieh CC, Cheng KC, Lee MF, Chiang SR, Shieh JM, Chang MS. IL-19 induced Th2 cytokines and was up-regulated in asthma patients. J Immunol. 2004 Dec 1; 173(11):6712–8. [PubMed: 15557163]

- 106. Kawaguchi M, Kokubu F, Kuga H, Matsukura S, Hoshino H, Ieki K, Imai T, Adachi M, Huang SK. Modulation of bronchial epithelial cells by IL-17. J Allergy Clin Immunol. 2001 Nov; 108(5):804–9. [PubMed: 11692108]
- 107. Umemura M, Yahagi A, Hamada S, Begum MD, Watanabe H, Kawakami K, Suda T, Sudo K, Nakae S, Iwakura Y, Matsuzaki G. IL-17-mediated regulation of innate and acquired immune response against pulmonary Mycobacterium bovis bacille Calmette-Guerin infection. J Immunol. 2007 Mar 15; 178(6):3786–96. [PubMed: 17339477]
- 108. Khader SA, Bell GK, Pearl JE, Fountain JJ, Rangel-Moreno J, Cilley GE, Shen F, Eaton SM, Gaffen SL, Swain SL, Locksley RM, Haynes L, Randall TD, Cooper AM. IL-23 and IL-17 in the establishment of protective pulmonary CD4+ T cell responses after vaccination and during Mycobacterium tuberculosis challenge. Nat Immunol. 2007 Apr; 8(4):369–77. [PubMed: 17351619]
- 109. Kinugasa T, Sakaguchi T, Gu X, Reinecker HC. Claudins regulate the intestinal barrier in response to immune mediators. Gastroenterology. 2000 Jun; 118(6):1001–11. [PubMed: 10833473]
- 110. Dasgupta S, Jain SK. Protective effects of amniotic fluid in the setting of necrotizing enterocolitis. Pediatr Res. 2017 Jul 5.doi: 10.1038/pr.2017.144
- 111. Hui L, Dai Y, Guo Z, Zhang J, Zheng F, Bian X, Wu Z, Jiang Q, Guo M, Ma K, Zhang J. Immunoregulation effects of different γδT cells and toll-like receptor signaling pathways in neonatal necrotizing enterocolitis. Medicine (Baltimore). 2017 Feb.96(8):e6077.doi: 10.1097/MD.0000000000006077 [PubMed: 28225489]
- 112. González-Rivera R, Culverhouse RC, Hamvas A, Tarr PI, Warner BB. The age of necrotizing enterocolitis onset: an application of Sartwell's incubation period model. J Perinatol. 2011 Aug; 31(8):519–23. DOI: 10.1038/jp.2010.193 [PubMed: 21273988]
- 113. Gordon PV, Clark R, Swanson JR, Spitzer A. Can a national dataset generate a nomogram for necrotizing enterocolitis onset? J Perinatol. 2014 Oct; 34(10):732–5. DOI: 10.1038/jp.2014.137 [PubMed: 25078862]
- 114. Guthrie SO, Gordon PV, Thomas V, Thorp JA, Peabody J, Clark RH. Necrotizing enterocolitis among neonates in the United States. J Perinatol. 2003 Jun; 23(4):278–85. DOI: 10.1038/sj.jp. 7210892 [PubMed: 12774133]
- 115. Denning TW, Bhatia AM, Kane AF, Patel RM, Denning PW. Pathogenesis of NEC: Role of the innate and adaptive immune response. Semin Perinatol. 2017 Feb; 41(1):15–28. DOI: 10.1053/ j.semperi.2016.09.014 [PubMed: 27940091]
- 116. Sodhi CP, Shi XH, Richardson WM, Grant ZS, Shapiro RA, Prindle T Jr, Branca M, Russo A, Gribar SC, Ma C, Hackam DJ. Toll-like receptor-4 inhibits enterocyte proliferation via impaired beta-catenin signaling in necrotizing enterocolitis. Gastroenterology. 2010 Jan; 138(1):185–96. DOI: 10.1053/j.gastro.2009.09.045 [PubMed: 19786028]
- 117. Wolfs TG, Derikx JP, Hodin CM, Vanderlocht J, Driessen A, de Bruïne AP, Bevins CL, Lasitschka F, Gassler N, van Gemert WG, Buurman WA. Localization of the lipopolysaccharide recognition complex in the human healthy and inflamed premature and adult gut. Inflamm Bowel Dis. 2010 Jan; 16(1):68–75. DOI: 10.1002/ibd.20995 [PubMed: 20014022]
- 118. Neu J. Neonatal necrotizing enterocolitis: an update. Acta Paediatr Suppl. 2005 Oct; 94(449): 100–5. DOI: 10.1080/08035320510043637 [PubMed: 16214774]
- 119. Zhang X, Zhivaki D, Lo-Man R. Unique aspects of the perinatal immune system. Nat Rev Immunol. 2017 Aug; 17(8):495–507. DOI: 10.1038/nri.2017.54 [PubMed: 28627520]
- 120. Shaw MH, Kamada N, Kim YG, Núñez G. Microbiota-induced IL-1β, but not IL-6, is critical for the development of steady-state TH17 cells in the intestine. J Exp Med. 2012 Feb 13; 209(2): 251–8. DOI: 10.1084/jem.20111703 [PubMed: 22291094]
- 121. Zhou L, Ivanov II, Spolski R, Min R, Shenderov K, Egawa T, Levy DE, Leonard WJ, Littman DR. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat Immunol. 2007 Sep; 8(9):967–74. DOI: 10.1038/ni1488 [PubMed: 17581537]
- 122. Boniface K, Bak-Jensen KS, Li Y, Blumenschein WM, McGeachy MJ, McClanahan TK, McKenzie BS, Kastelein RA, Cua DJ, de Waal Malefyt R. Prostaglandin E2 regulates Th17 cell

differentiation and function through cyclic AMP and EP2/EP4 receptor signaling. J Exp Med. 2009 Mar 16; 206(3):535–48. DOI: 10.1084/jem.20082293 [PubMed: 19273625]

- 123. Dimmitt RA, Staley EM, Chuang G, Tanner SM, Soltau TD, Lorenz RG. Role of postnatal acquisition of the intestinal microbiome in the early development of immune function. J Pediatr Gastroenterol Nutr. 2010 Sep; 51(3):262–73. DOI: 10.1097/MPG.0b013e3181e1a114 [PubMed: 20639773]
- 124. Talia DM, Deliyanti D, Agrotis A, Wilkinson-Berka JL. Inhibition of the Nuclear Receptor RORγ and Interleukin-17A Suppresses Neovascular Retinopathy: Involvement of Immunocompetent Microglia. Arterioscler Thromb Vasc Biol. 2016 Jun; 36(6):1186–96. DOI: 10.1161/ATVBAHA. 115.307080 [PubMed: 27055905]
- 125. Dammann O, Brinkhaus MJ, Bartels DB, Dördelmann M, Dressler F, Kerk J, Dörk T, Dammann CE. Immaturity, perinatal inflammation, and retinopathy of prematurity: a multi-hit hypothesis. Early Hum Dev. 2009 May; 85(5):325–9. DOI: 10.1016/j.earlhumdev.2008.12.010 [PubMed: 19217727]
- 126. Gantert M, Been JV, Gavilanes AW, Garnier Y, Zimmermann LJ, Kramer BW. Chorioamnionitis: a multiorgan disease of the fetus? J Perinatol. 2010 Oct; 30(Suppl):S21–30. DOI: 10.1038/jp. 2010.96 [PubMed: 20877404]
- 127. Deliyanti D, Wilkinson-Berka JL. Inhibition of NOX1/4 with GKT137831: a potential novel treatment to attenuate neuroglial cell inflammation in the retina. J Neuroinflammation. 2015; 12:136.doi: 10.1186/s12974-015-0363-z [PubMed: 26219952]
- 128. Vecino E, Rodriguez FD, Ruzafa N, Pereiro X, Sharma SC. Glia-neuron interactions in the mammalian retina. Prog Retin Eye Res. 2016 Mar.51:1–40. DOI: 10.1016/j.preteyeres. 2015.06.003 [PubMed: 26113209]
- 129. Sato T, Kusaka S, Shimojo H, Fujikado T. Simultaneous analyses of vitreous levels of 27 cytokines in eyes with retinopathy of prematurity. Ophthalmology. 2009 Nov; 116(11):2165–9. DOI: 10.1016/j.ophtha.2009.04.026. [PubMed: 19700197]
- 130. Joyal JS, Sitaras N, Binet F, Rivera JC, Stahl A, Zaniolo K, Shao Z, Polosa A, Zhu T, Hamel D, Djavari M, Kunik D, Honoré JC, Picard E, Zabeida A, Varma DR, Hickson G, Mancini J, Klagsbrun M, Costantino S, Beauséjour C, Lachapelle P, Smith LE, Chemtob S, Sapieha P. Ischemic neurons prevent vascular regeneration of neural tissue by secreting semaphorin 3A. Blood. 2011 Jun 2; 117(22):6024–35. DOI: 10.1182/blood-2010-10-311589 [PubMed: 21355092]
- 131. Amadi-Obi A, Yu CR, Liu X, Mahdi RM, Clarke GL, Nussenblatt RB, Gery I, Lee YS, Egwuagu CE. TH17 cells contribute to uveitis and scleritis and are expanded by IL-2 and inhibited by IL-27/STAT1. Nat Med. 2007 Jun; 13(6):711–8. DOI: 10.1038/nm1585 [PubMed: 17496900]
- 132. Zhu Y, Tan W, Demetriades AM, Cai Y, Gao Y, Sui A, Lu Q, Shen X, Jiang C, Xie B, Sun X. Interleukin-17A neutralization alleviated ocular neovascularization by promoting M2 and mitigating M1 macrophage polarization. Immunology. 2016 Apr; 147(4):414–28. DOI: 10.1111/ imm.12571. [PubMed: 26694999]
- 133. Mokarram N, Bellamkonda RV. A perspective on immunomodulation and tissue repair. Ann Biomed Eng. 2014 Feb; 42(2):338–51. DOI: 10.1007/s10439-013-0941-0 [PubMed: 24297492]
- 134. Lamagna C, Aurrand-Lions M, Imhof BA. Dual role of macrophages in tumor growth and angiogenesis. J Leukoc Biol. 2006 Oct; 80(4):705–13. DOI: 10.1189/jlb.1105656 [PubMed: 16864600]
- 135. Ligresti G, Aplin AC, Zorzi P, Morishita A, Nicosia RF. Macrophage-derived tumor necrosis factor-alpha is an early component of the molecular cascade leading to angiogenesis in response to aortic injury. Arterioscler Thromb Vasc Biol. 2011 May; 31(5):1151–9. DOI: 10.1161/ ATVBAHA.111.223917 [PubMed: 21372301]
- 136. Wallace DK, Kraker RT, Freedman SF, Crouch ER, Hutchinson AK, Bhatt AR, Rogers DL, Yang MB, Haider KM, VanderVeen DK, Siatkowski RM, Dean TW, Beck RW, Repka MX, Smith LE, Good WV, Hartnett ME, Kong L, Holmes JM, Pediatric Eye Disease Investigator Group (PEDIG). Assessment of Lower Doses of Intravitreous Bevacizumab for Retinopathy of Prematurity: A Phase 1 Dosing Study. JAMA Ophthalmol. 2017 Jun 1; 135(6):654–656. DOI: 10.1001/jamaophthalmol.2017.1055 [PubMed: 28448664]

- 137. Yonekawa Y, Wu WC, Nitulescu CE, Chan RVP, Thanos A, Thomas BJ, Todorich B, Drenser KA, Trese MT, Capone A Jr. Progressive retinal detachment in infants with retinopathy of prematurity treated with intravitreal bevacizumab or ranibizumab. Retina. 2017 May 3.doi: 10.1097/IAE. 0000000000001685
- 138. VanderVeen DK, Melia M, Yang MB, Hutchinson AK, Wilson LB, Lambert SR. Anti-Vascular Endothelial Growth Factor Therapy for Primary Treatment of Type 1 Retinopathy of Prematurity: A Report by the American Academy of Ophthalmology. Ophthalmology. 2017 May; 124(5):619– 633. DOI: 10.1016/j.ophtha.2016.12.025 [PubMed: 28341474]
- 139. Fassnacht-Riederle H, Becker M, Graf N, Michels S. Effect of aflibercept in insufficient responders to prior anti-VEGF therapy in neovascular AMD. Graefes Arch Clin Exp Ophthalmol. 2014 Nov; 252(11):1705–9. DOI: 10.1007/s00417-014-2589-3 [PubMed: 24614949]
- 140. Clyman RI. Mechanisms regulating the ductus arteriosus. Biol Neonate. 2006; 89(4):330–5. DOI: 10.1159/000092870 [PubMed: 16770073]
- 141. Kajino H, Chen YQ, Chemtob S, Waleh N, Koch CJ, Clyman RI. Tissue hypoxia inhibits prostaglandin and nitric oxide production and prevents ductus arteriosus reopening. Am J Physiol Regul Integr Comp Physiol. 2000 Jul; 279(1):R278–86. [PubMed: 10896892]
- 142. Demir N, Peker E, Ece, A engin K, Bulan KA, Tuncer O. Is platelet mass a more significant indicator than platelet count of closure of patent ductus arteriosus? J Matern Fetal Neonatal Med. 2016; 29(12):1915–8. DOI: 10.3109/14767058.2015.1067296 [PubMed: 26169703]
- 143. Vucovich MM, Cotton RB, Shelton EL, Goettel JA, Ehinger NJ, Poole SD, Brown N, Wynn JL, Paria BC, Slaughter JC, Clark RH, Rojas MA, Reese J. Aminoglycoside-mediated relaxation of the ductus arteriosus in sepsis-associated PDA. Am J Physiol Heart Circ Physiol. 2014 Sep 1; 307(5):H732–40. DOI: 10.1152/ajpheart.00838.2013 [PubMed: 24993047]
- 144. Iwasaki S, Minamisawa S, Yokoyama U, Akaike T, Quan H, Nagashima Y, Nishimaki S, Ishikawa Y, Yokota S. Interleukin-15 inhibits smooth muscle cell proliferation and hyaluronan production in rat ductus arteriosus. Pediatr Res. 2007 Oct; 62(4):392–8. DOI: 10.1203/PDR. 0b013e31813c9339 [PubMed: 17667861]
- 145. Robert M, Miossec P. Effects of Interleukin 17 on the cardiovascular system. Autoimmun Rev. 2017 Sep; 16(9):984–991. DOI: 10.1016/j.autrev.2017.07.009 [PubMed: 28705781]
- 146. Maione F, Cicala C, Liverani E, Mascolo N, Perretti M, D'Acquisto F. IL-17A increases ADPinduced platelet aggregation. Biochem Biophys Res Commun. 2011 May 20; 408(4):658–62. DOI: 10.1016/j.bbrc.2011.04.080 [PubMed: 21530487]
- 147. Kim ES, Kim EK, Choi CW, Kim HS, Kim BI, Choi JH, Park JS, Moon KC. Intrauterine inflammation as a risk factor for persistent ductus arteriosus patency after cyclooxygenase inhibition in extremely low birth weight infants. J Pediatr. 2010 Nov; 157(5):745–50.e1. DOI: 10.1016/j.jpeds.2010.05.020 [PubMed: 20598319]
- 148. Gonzalez A, Sosenko IR, Chandar J, Hummler H, Claure N, Bancalari E. Influence of infection on patent ductus arteriosus and chronic lung disease in premature infants weighing 1000 grams or less. J Pediatr. 1996 Apr; 128(4):470–8. [PubMed: 8618179]
- 149. Veldhoen M. Interleukin 17 is a chief orchestrator of immunity. Nat Immunol. 2017 May 18; 18(6):612–621. DOI: 10.1038/ni.3742 [PubMed: 28518156]
- 150. Stoll BJ, Gordon T, Korones SB, Shankaran S, Tyson JE, Bauer CR, Fanaroff AA, Lemons JA, Donovan EF, Oh W, Stevenson DK, Ehrenkranz RA, Papile LA, Verter J, Wright LL. Late-onset sepsis in very low birth weight neonates: a report from the National Institute of Child Health and Human Development Neonatal Research Network. J Pediatr. 1996 Jul; 129(1):63–71. [PubMed: 8757564]
- 151. Valdez PA, Vithayathil PJ, Janelsins BM, Shaffer AL, Williamson PR, Datta SK. Prostaglandin E2 suppresses antifungal immunity by inhibiting interferon regulatory factor 4 function and interleukin-17 expression in T cells. Immunity. 2012 Apr 20; 36(4):668–79. DOI: 10.1016/ j.immuni.2012.02.013. [PubMed: 22464170]
- 152. Miossec P, Kolls JK. Targeting IL-17 and TH17 cells in chronic inflammation. Nat Rev Drug Discov. 2012 Oct; 11(10):763–76. DOI: 10.1038/nrd3794 [PubMed: 23023676]

153. Kolls JK, Kanaly ST, Ramsay AJ. Interleukin-17: an emerging role in lung inflammation. Am J Respir Cell Mol Biol. 2003 Jan; 28(1):9–11. DOI: 10.1165/rcmb.2002-0255PS [PubMed: 12495927]