ACS central science

Cite This: ACS Cent. Sci. 2018, 4, 805-819

A Friend or Foe: Calcineurin across the Gamut of Neurological Disorders

Jackson Saraf,[†] Pallab Bhattacharya,^{*,†}[©] Kiran Kalia,[†] Anupom Borah,[‡] Deepaneeta Sarmah,[†] Harpreet Kaur,[†] Kunjan R Dave,[§] and Dileep R Yavagal[§]

[†]Department of Pharmacology and Toxicology, National Institute of Pharmaceutical Education and Research (NIPER), Ahmedabad, Gandhinagar, Gujarat 382355, India

[‡]Cellular and Molecular Neurobiology Laboratory, Department of Life Science and Bioinformatics, Assam University, Silchar, Assam 788011, India

[§]Department of Neurology, University of Miami Miller School of Medicine, Miami, Florida 33136, United States

ABSTRACT: The serine/threonine phosphatase calcineurin (CaN) is a unique but confounding calcium/calmodulin-mediated enzyme. CaN has shown to play essential roles from regulating calcium homeostasis to being an intricate part of learning and memory formation. Neurological disorders, despite differing in their etiology, share similar pathological outcomes, such as mitochondrial dysfunction and apoptotic signaling brought about by excitotoxic elements. CaN, being deeply integrated in vital neuronal functions, may be implicated in various neurological disorders. Understanding the enzyme and its physiological niche in the nervous system is vital in uncovering its roles in the spectrum of brain disorders. By reviewing the crosstalk in different neurological pathologies, a possible grasp of CaN's complex signaling may lead to forming better neurotherapy. This Outlook attempts to explore the various neuronal functions of CaN and investigate its pervasive role through the gamut of neurological disorders.



1. CALCINEURIN: A PROLOGUE

Even 40 years after its initial discovery by Wang and Desai as a column fraction, calcineurin (CaN) continues to intrigue the scientific community with its vast biological role.¹ In its initial findings, CaN was often referred to as an "affinity-purified phosphodiesterase".² Klee and Krinks were the first to purify CaN and presumed it to be a regulatory subunit of a phosphodiesterase based on its ability to inhibit the same.³ Wallace and Lynch further purified CaN from bovine brain extracts using a two-column approach and found its activity to hinder the adenylate cyclase and nucleotide phosphodiesterases.^{4,5} Due to similarity in biological activity traced by various research groups, it was therefore speculated that CaN may possibly be a regulatory enzyme. Its abundant allocation in the nervous system prompted Klee et al. to coin the term "calcineurin".⁶ Klee's version of CaN was corroborated by the work of Philip Cohen in which he found a fraction in cellular extracts that dephosphorylated the α -/ β -subunits of phosphorylase kinase; the fraction revealed to be nearly identical to the CaN discovered by Klee.7,8

As per consensus, CaN is deemed as a calcium/calmodulindependent (Ca²⁺/CaM) serine-threonine phosphatase, also known as protein phosphatase 2B (PP2B), ubiquitous in most mammalian tissues but predominating in the brain.⁹ It is the only reported phosphatase thus far known to be completely dependent on CaM. From the initial presumption as being a calmodulin-dependent cyclic nucleotide phosphodiesterase inhibitor to the radical revelation of being the target of immunosuppressant drugs such as cyclosporin A in organ transplants, CaN seems to manifest itself in key biological pathways throughout various tissues in the body and necessitates a deeper investigation in its intricate physiological function.¹⁰ Its prevalence in the nervous tissue has recently made it a target of investigation for many neurological disorders, particularly those constituted by calcium excitotoxicity and neurotransmitter imbalance.^{11,12} It is believed that CaN plays a major role in bridging Ca²⁺ homeostasis to brain function; studies even point to CaN being the link between aging and brain dysregulation.¹³

> CaN is deemed as a calcium/ calmodulin-dependent (Ca²⁺/CaM) serine-threonine phosphatase, also known as protein phosphatase 2B (PP2B), ubiquitous in most mammalian tissues but predominating in the brain.

 Received:
 April 17, 2018

 Published:
 June 27, 2018

Though discussing the vast role of CaN in all the tissues is far beyond the realm of this Outlook, an attempt shall be made to delve into the CaN's role in the brain and across the spectrum of neurological disorders such as stroke, Alzheimer's disease, Parkinson's disease, and other neurodegenerative disorders involving CaN. This crosstalk may clarify the extent of CaN's involvement in these perplexing disorders.

2. CALCINEURIN: CLASSIFICATION AND STRUCTURE

The classification of serine/threonine phosphatases was established by Cohen in which he proposed two categories: type 1 and type 2 protein phosphatases (PPs).¹⁴ Type 1 PPs dephosphorylate the β -subunit of phosphorylase kinase while type 2 act on the α -subunit. The two PPs can be distinguished by their inhibition study in which phosphopeptide inhibitors act on type 1 but are unable to inhibit type 2 PPs. Cohen also later subclassified type 2 PPs according to the dependence on divalent ions for activity.¹⁵ He designated PP2A, PP2B, and PP2C as having no requirement for divalent ions, requiring Ca²⁺ (CaN), and requiring Mg²⁺ for activity, respectively. Evolutionarily speaking, PP1, PP2A, and PP2B (CaN) are some of the most highly conserved PP families throughout nature.¹⁶

Human CaN is a heterodimeric serine/threonine phosphatase, comprising a unique structure composed of two subunits connected jointly: a 61 kDa catalytic A subunit (though variations between species ranges from 58 to 64 kDa) and a 19 kDa regulatory B subunit.¹⁷ The B subunit, although smaller in size, consists of a helical B-binding segment, a calmodulin-binding (CaM-binding) segment, and an autoinhibitory domain.¹⁸ The B-binding segment tightly adheres to the A subunit. Half of CaN appears to be cytosolic while the amino terminal glycine of the B subunit is believed to be myristoylated for greater association with the plasma membrane.¹⁹ Furthermore, the B segment consists of four Ca²⁺-binding sites (EF-motifs).^{20,21} Two Ca²⁺ sites reside on the N-terminal and have lower affinity (dissociation constant in the micromolar range). These seemingly serve as sensors for the fluctuations in intracellular Ca^{2+} levels and thereby serve a regulatory role, especially for the binding of CaM to the CaM-binding segment of subunit B. Another two Ca²⁺ sites reside on the C-terminal of the B subunit. These have much higher affinity (have dissociation constants in the nanomolar range) and act as stabilizers to keep both subunits intact and confer structural stability to the entirety of CaN.²² The CaM-binding segment relies on Ca²⁺ to help associate with subunit A, the catalytic site. This structure therefore stands to reason how CaN is able to couple Ca²⁺ signals to cellular responses. However, the enzyme undergoes complicated conformations to reach the active state. In previous studies, the conformational shift from inactive to active enzymatic states raised several questions. After all, the autoinhibitory site of subunit B (residues 467-486) lies at a relative distance from the B-binding site (341-372) and the CaMbinding segment (391-414). These questions were answered through studies done by Shen et al. in which the group discovered there are a separate sequence of conformational shifts that eventually lead to the fully activated enzyme.²³ They found that, during elevated levels, Ca2+ initially binds to the lower-affinity Ca²⁺ sites, thereby causing a conformation which exposes the CaM-binding site. This fulfills subunit B's main role as a regulator. When CaM binds to its respective site, another conformational change stabilizes subunit B by releasing the autoinhibitory peptide and renders the enzyme fully active.

In summary, when the enzyme is inactive (resting Ca^{2+} levels), influx (micromolar) of Ca^{2+} binding to subunit B causes a conformational change to reveal the CaM-binding site on subunit A (partial activation), which upon binding to CaM displaces the autoinhibitory peptide and stabilizes and fully activates the enzyme (see Figure 1).

3. CALCINEURIN: VITAL HOMEOSTATIC MECHANISMS

3.1. Ca²⁺ Homeostasis. As CaN is ubiquitous throughout the body (several isoforms exist among different tissues), its roles are accordingly highly diversified. However, this Outlook will focus on CaN's roles solely in the brain, where the highest levels can be found.²⁴ CaN is known to constitute slightly more than 1% of total protein in the brain,²⁵ and it is most densely populated in the caudate putamen and hippocampus but distributed throughout other regions of the brain.²⁶ CaN is also found at the level of dendrites, axons, neuronal spines, presynaptic terminals, and postsynaptic densities. Magnifying the grandeur of CaN's role in neuronal homeostasis is its very existence as the sole phosphatase heavily dependent on $Ca^{2+}/$ CaM signaling. It is well-established that calcium and its various calcium channels play a major and intricate role in neuronal function and activity. Calcium is arguably one of the most important secondary messengers capable of initiating pivotal cellular events.²⁷ Calcium plays an essential role in many biological pathways that may be operating simultaneously within the neuron; drastic changes in its levels are associated with numerous neurological disorders.^{28,29} As Ca²⁺ relays synaptic signals to the nucleus then to synapse again, it is immensely responsible for turning on and off the various genetic expressions necessary for normal neuronal activity.³⁰ Therefore, dysregulation (excess and deficient Ca^{2+} stimulation) in the Ca²⁺ synapse-to-nucleus signaling can cause breakdown in essential gene expressions that underlie most neurological pathologies, both degenerative and psychiatric in nature.

CaN is an integral member of the Ca^{2+} signaling pathway on both sides of the synapse.³¹ It behaves as a sensor and is vigilant of the intracellular levels of Ca^{2+} and maintains the proper equilibria necessary for cell maintenance and operation. There are several mechanisms CaN deploys to regulate intracellular Ca^{2+} levels and the downstream activities that maintain an optimum neuronal microenvironment.

CaN is an integral member of the Ca²⁺ signaling pathway on both sides of the synapse. It behaves as a sensor and is vigilant of the intracellular levels of Ca²⁺ and maintains the proper equilibria necessary for cell maintenance and operation.

3.2. Synaptic Exocytosis. Presynaptically, CaN and cyclindependent kinase 5 (CDK5) oversee vesicular release of neurotransmitters and prevent excessive exocytosis to maintain an adequate amount of neurotransmitters reaching the post-synaptic sites.³² These exocytotic processes require an ensemble of proteins that coordinate action potential and Ca²⁺ signaling to neurotransmitter release. CDK5 is able to phosphorylate



Figure 1. Activation of calcineurin (CaN). The phosphatase is activated through the sequence of two subsequent conformations. Initially, the influx of calcium (Ca²⁺) raises the intracellular calcium load. At the nanomolar range, Ca²⁺ first binds to the two higher-affinity EF motifs (H_x) of subunit B, tightly adhering subunit B to subunit A and stabilizing the enzyme. As Ca²⁺ levels rise to the micromolecular threshold, it binds to the lower-affinity EF motifs (L_x), spurring a conformational change that unveils the calmodulin (CaM) site to which CaM binds and initiates a second conformational change responsible for displacing the autoinhibitory peptide. The final conformation stabilizes and fully renders CaN active.

proteins, like synapsin 1 which attaches to synaptic vesicles and sequesters them to the release-reluctant resting pool.³³ Studies have shown that CDK5 knockout accelerates synaptic cycling and exocytotic kinetics; similarly, CaN knockout has shown to slow down exocytosis rates.³⁴ CaN counteracts this effect, dephosphorylates synapsin 1, and is able to reverse the suppression of exocytosis and synaptic cycling induced by CDK5.³⁵ Therefore, during greater Ca²⁺ influx, CaN is able to increase the amount of neurotransmitters released into the synaptic cleft³⁶ (see Figure 2a).

3.3. Synaptic Endocytosis. CaN also participates in endocytosis as it facilitates clathrin-dependent recycling by targeting different dephosphins, endocytic proteins, one in particular called dynamin 1.³⁷ The interaction with dynamin 1 is based on CaN's sensitivity toward Ca²⁺ and spurs the depolarization-induced vesicular recycling; disturbance to such an interaction blocks endocytosis.³⁸ However, such a role has been under controversy as CaN has shown to be implicated at different endocytic sites catalyzing different and opposing events.³⁹ Regardless, studies continue to confirm that CaN is a key role player in endocytotic processes at both nerve terminals as well as in non-neuronal cells⁴⁰ (see Figure 2a).

3.4. Ca²⁺ Influx and Regulation. Postsynaptically, CaN manages the Ca²⁺ homeostasis in the neuron by modulating its influx rates. The entry of Ca²⁺ activates the cell and triggers a cascade of a wide variety of biomolecular events. This intracytoplasmic Ca²⁺ influx is either via voltage-gated calcium channels (VGCCs) or ligand-gated channels such as the

N-methyl-D-aspartate receptors (NMDARs).^{41,42} For NMDARs, CaN is located in the postsynaptic densities, near the cytoplasm and colocalized with NMDA receptors.⁴³ The high affinity of CaN for the incoming Ca²⁺ activates it to dephosphorylate the C-terminal of the NR2A subunits of NMDARs, leading to glycine-independent desensitization and weakening of Ca²⁺ currents into the cell by decreasing the average duration of opened Ca²⁺ channels as well as their frequency.⁴⁴

It has also been documented that VGCCs are directly involved in synaptic plasticity and dendritic excitability.^{45,46} VGCCs, specifically the L-type CCs and to some extent the N-type CCs, are phosphorylated.⁴⁷ CaN is targeted toward these channels through A-kinase anchoring protein 79/150 (AKAP) when calcium-dependent inactivation (CDI) is stimulated by the Ca²⁺ influx. CDI then further triggers microtubule-associated protein 2B (MAP2B) to guide protein kinase A (PKA) toward the VGCCs for priming and reinitiating Ca²⁺ current.^{48,49} CaN is able to reduce the Ca²⁺ influx through this negative feedback effect and utilizes it to keep a check on the amount of Ca²⁺ entering the cell.

CaN also regulates the Ca²⁺-induced Ca²⁺ release (CICR), a universal mechanism by which cells are able to couple the inositol triphosphate receptors (IP3Rs) and ryanodine receptors (RyRs) to Ca²⁺ channels to trigger the release of Ca²⁺ from intracellular stores.^{50,51} Intracellular stores of Ca²⁺ reside in the sarcoplasmic reticulum and are mediated by IP3Rs and RyRs.³⁰ It is believed this is another way CDI can be initiated alternatively to VGCC dephosphorylation due to the intimate association of VGCCs and the CCs opened through RyRs



Figure 2. (a) Presynaptic homeostatic functions of calcineurin (CaN). (A) CaN plays an integral role in vesicle exocytosis by regulating the release in coordination with neurotransmission signaling. The influx of calcium (Ca^{2+}) activates CaN and causes it to dephosphorylate synapsin 1, removing its suppression of vesicles and allowing them to move from the release-reluctant resting pool toward the axonal terminal for release. The process is reversed by the phosphorylation by cyclin-dependent kinase 5 (CDK5). (B) CaN also plays a role in endocytosis of vesicles after activation by Ca^{2+} and then dephosphorylation of dynamin 1. CaN and dynamin 1 form a complex and move toward the endocytotic packaging assembly where involved proteins allow for the packaging of the vesicle at the axonal terminal, its uptake, and delivery of the vesicle toward the vesicle pool for recycling. This series of synaptic cycling and recycling by CaN is tightly coordinated and coupled to the Ca²⁺-dependent signaling along with the crosstalk of involved proteins. Interruptions in the processes can disturb the equilibria of neurotransmission and often lead to neuronal impair. (b) Postsynaptic homeostatic functions of calcineurin (CaN). Postsynaptically, CaN tightly regulates several functions. It controls intracellular Ca^{2+} levels through modulation of voltage-gated calcium channels (VGCCs), NMDA (N-methly-D-aspartate), and AMPA (α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid) receptors. Influx of Ca^{2+} through VGCCs tends to trigger the release of more Ca^{2+} from the endoplasmic reticulum (calcium-induced calcium release) via ryanodine and inositol triphosphate receptors (RyRs and IP3Rs). The increased Ca²⁺ load activates CaN, and through a negative feedback control, it dephosphorylates VGCCs, RyRs, and IP3Rs, decreasing the duration of opening and frequency as well as weakening the incoming cationic currents. Ca²⁺ influx via NMDA receptors (NMDARs) and the Ca²⁺-permeable AMPA receptors (AMPARs) follows a similar pathway of Ca²⁺ regulation and decreases long-term potentiation (LTP) through internalization and reduced NMDAR and AMPAR expression and increasing long-term depotentiation (LTD). This Ca²⁺-dependent inactivation (CDI) reinitiates the opening of these channels by triggering kinases, like protein kinase A (PKA) which complexes with A-kinase anchoring protein 79/150 (AKAP79/150) after guided by microtubule-associated protein 2B (MAP2B), that cause rephosphorylation. In the case of voltage-gated A-type potassium 4.2 (Kv4.2) channels, however, phosphorylation of the channels results in internalization and increased long-term potentiation (LTP) while the process is reversed after dephosphorylation by CaN.

located on the sarcoplasmic reticulum (SR).⁵² CaN is able to dephosphorylate a phosphokinase C (PKC) phosphosite on both IP3Rs and RyRs and reduce their sensitivity toward alterations in intracellular Ca²⁺ concentrations and the subsequent affinity for their ligands (IP3 and Ry, respectively), thereby preventing the mechanism of CICR⁵³ (see Figure 2b).

4. SYNAPTIC PLASTICITY

In conjunction to maintaining homeostatic Ca^{2+} levels throughout the neuronal network, CaN also facilitates homeostasis to synaptic plasticity, a term that denotes the flexibility and versatility of the neuronal network to change the synaptic strength in a direction that opposes the elevation or reduction in activity.⁵⁴ This synaptic scaling often tends to be a negative feedback process and responds to altered synaptic strength to stabilize the neuronal network.^{55,56} CaN has been shown to be an important mediator of synaptic plasticity as its activation or inhibition can block or initiate synaptic scaling which further tends to enhance synaptic strength. CaN is able to regulate this through Ca²⁺ signaling, but at times even Ca²⁺ signaling can be disrupted and in turn can disrupt CaN activity. For the neuronal network to function properly synaptic plasticity must be sustained, and in such cases restoring Ca²⁺ signals becomes imperative. Studies suggest that chronic Ca²⁺-dependent inhibition (CDI) can lead to an attenuation of neuronal firing and

decrease neuronal activity. As Ca²⁺ influx decreases, CaN activity also decreases and sets off a compensatory mechanism involving the phosphorylation of serine 845 of GluA1 by PKA bound to AKAP79/150 and the elevated expression of α -amino-3hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AM-PARs) which are more sensitive and permeable to Ca²⁺, thereby restoring Ca²⁺ signaling⁵⁷ (see Figure 2b). Upon restoration of Ca²⁺ signals, CaN is reactivated and reverts to facilitating proper neuronal signaling.

> In conjunction to maintaining homeostatic Ca²⁺ levels throughout the neuronal network, CaN also facilitates homeostasis to synaptic plasticity, a term that denotes the flexibility and versatility of the neuronal network to change the synaptic strength in a direction that opposes the elevation or reduction in activity.

Although mainly associated with LTD, CaN has shown its ability to control neuronal excitability via regulation of voltage-gated A-type potassium (K^+) Kv4.2 channels.^{71,72}

These channels are highly populated in postsynaptic terminals as well as dendrites.^{73,74} Their role is repolarization after the passage of action potentials and preventing their reverse propagation into dendritic regions.^{75,76} Overexpressed Kv4.2 channels have shown to shorten action potentials and mitigate LTP; this negative regulation of LTP resonates with that of CaN's. Increase in NR2B/NR2A subunit ratio has been shown to facilitate synaptic plasticity, and LTP ensues. Overexpressed Kv4.2 channels reduce this ratio and trigger LTD.⁷⁷ It has been shown that CaN modulates the activity of these channels in arms with the PKA/AKAP79/150 complex. The dephosphorvlation by CaN stabilizes Kv4.2 channels at the postsynaptic terminal, dampening of neurotransmission (LTD). PKA counters this activity through phosphorylation of the channels by binding to AKAP79/150 to induce their internalization into the cytoplasm.⁷⁸ This presents fewer Kv4.2 channels at the postsynaptic terminal and leads to LTP. Deficits in Kv4.2 channels have shown to produce spatial learning deficits and memory impairment⁷⁹ (see Figure 2b).

5. RECEPTOR SIGNALING

AMPARs are the most abundant excitatory receptors throughout the nervous system when it comes to fast synaptic transmission.⁸⁰ Their expansive distribution and association with postsynaptic protein complexes grant them a high-level priority in regards to synaptic transmission and the electrophysiology of neurons.⁸¹ Their dynamic nature is conducive for changes in synaptic plasticity and therefore generates acute responsiveness within cells toward extracellular signals. Similar to metabotropic glutamate receptors and NMDARs, these receptors are also governed by phosphorylation (by PKA bound to AKAP79/150) and dephosphorylation (CaN), both targeting the highly sensitive s845 residue of the receptor.⁸² Phosphorvlation increases greater expressions of the GluA1 on the postsynaptic membrane and the amount of current flowing through the AMPARs, thereby strengthening synaptic transmission.^{83,} Dephosphorylation of s845 sequesters the receptor and induces LTD.85 PKA and CaN regulate synaptic plasticity at AMPARs and serve a vital role in maintaining the optimum signaling throughout the neural circuitry.

CaN also serves another regulatory role by dephosphorylating the metabotropic glutamate receptor 5 (mGluR5) which is responsible for augmenting the responsiveness of neurons to different signals from their microenvironment.86 In other words, mGluR5 elevates LTP. Belonging to group I of mGluRs and located postsynaptically, mGluR5 works via the G_q pathway: it stimulates phospholipase C to hydrolyze phosphatidylinositol biphosphate (PIP2) into IP3 and diacylglycerol (DAG) and further leads to the opening of Ca²⁺ stores via IP3Rs and RyRs and alteration in activities of various voltagegated channels.⁸⁷ In the process, it triggers a Ca²⁺ influx through NMDARs and becomes desensitized. CaN, after being activated by the NMDAR-mediated influx of Ca²⁺, resensitizes mGluR5 by dephosphorylation of its C-terminus and prolongs its activity; this is an important aspect of synaptic plasticity.88

6. TRANSCRIPTION REGULATION

One of the major associations of CaN involves the nuclear factor for activated T-cell (NFAT) transcription factor. This family consists of five NFAT downstream targets residing in the cytoplasm in a hyperphosphorylated idle state (NFAT_c1-4,

CaN has proven to modulate synaptic activity through Ca²⁺ influx through VGCCs and NMDARs.58 Furthermore, CaN has proven to confer bidirectional plasticity toward memory and learning while its deficiency has shown to impair working memory.⁵⁹ This faculty of CaN to control bidirectional synaptic plasticity has great implications in memory and learning.⁶⁰ Both phosphorylation and dephosphorylation play vital roles in memory formation and, respectively, trigger long-term potentiation (LTP) and long-term depression (LTD).⁶¹ Phosphokinases such as CaMKII, PKA, PKC, and MAPK have been involved in producing LTP, a phenomenon marked by both presynaptic and postsynaptic stimulations occurring at higher than normal frequencies or basically an enhanced synaptic transmission (the presynaptic stimulation often precedes the postsynaptic by varying milliseconds).⁶² Phosphatases, such as CaN, tend to depress this high-frequency stimulation and work in an opposing manner. Overexpression of CaN has shown to impair LTP while its inhibition can disrupt depotentiation.⁶³ Whereas LTP will produce greater NMDARs/AMPARs expression, LTD will tend to decrease the same.⁶⁴ CaN can indirectly also enhance LTP by reducing the LTD through its extensive manipulation of inhibitory synapses. Its affinity for GABA-A receptors and their subsequent dephosphorylation renders them for disinhibition as less GABA firing leads to less inhibition of the excitatory neurotransmission, thereby raising the LTP and overall synaptic efficacy.^{65–67} This is exemplary of the sophistication of CaN and its unique command over its own actions and the ability to adjust them as per subcellular requirements via downstream dephosphorylation. However, even though LTP may offer a larger collection of studies supporting its role in memory formation (particularly in the CA1 region of the hippocampus), LTD may be just as integral.^{88,69} Therefore, regardless of which direction the debate on LTP and LTD swings, CaN has forged its own niche in the continuous quest to elucidate memory formation and learning.70

NFAT5/TonEBP).^{89,90} The CaN-NFAT signaling contributes to a wide array of biological developments such as cardiac and lung morphogenesis, differentiation and the growth of skeletal muscle, and immune responses like T-cell activation.⁹¹ As CaN is enriched throughout the brain regions, the tightly regulated signaling makes NFAT as proportionally and deeply integrated in many neuronal responses and activities such as axonal growth, memory formation, and neuronal apoptosis.⁹² Furthermore, the pathway is heavily involved in the development of the nervous system (synaptogenesis, corticogenesis), maintenance (myelination, neuronal survival), and nervous system operation (synaptic connectivity, plasticity, and neurotransmission).⁹³ CaN-NFAT pathway begins when Ca²⁺ influx activates CaN to dephosphorylate the CaN-phosphosites from the N-terminal of NFAT_c subunits, exposing their nuclear sites.⁹⁴ Once within the nucleus, these NFAT_c subunits partner with their NFAT_n counterparts on their specific response elements and promote various transcription sequences necessary for the homeostatic functioning of the neuron. However, the Ca²⁺ that activates CaN may not be sufficient to trigger this pathway. Rather, a greater influx of Ca^{2+} is provided by unique Ca²⁺ release activated Ca²⁺ (CRAC) channels.⁹⁵ Some studies also suggest that the NFAT complexes induce a positive feedback loop to increase the intracellular Ca²⁺ levels via IP3R-RyRs Ca²⁺ release.⁹⁶ Their job is to sustain the Ca²⁺ threshold necessary to maintain the NFAT_c-NFAT_n complexes within the nucleus for extended transcription to fulfill cellular needs at specified time points. A drop-off in intracellular Ca²⁺ levels signals the rephosphorylation of the NFAT complexes by initial priming by dual-specificity tyrosine-phosphorylation regulated kinases (DYRKs) particularly DYRK1A and DYRK2, recently discovered regulators of NFAT.97 Once primed, they are further subjected to rephosphorylation for export to the cytoplasm by glycogen synthase kinase 3 β (GSK-3), which acts to fine-tune the NFAT signaling⁹⁸ (see Figure 3).

7. CaN-MITOCHONDRIA CONNECTION

The intrinsic and paramount role of mitochondria in orchestrating cell survival and apoptosis has been well-documented.99-101 Studies over time have delineated several mechanistic pathways of mitochondria and their tightly regulated control over cell viability.¹⁰² Members of the Bcl-2 family, BAX and BAD, remain in an inactive state after phosphorylation by phosphokinases such as PKA, Akt, and Raf-1.¹⁰³⁻¹⁰⁵ Therefore, phosphorylation promotes cell survival and is employed by various growth factors. CaN has been postulated to be a key element in several of these pathways, particularly those favoring apoptosis.¹⁰⁶ Its ability to dephosphorylate BAD activates the pro-apoptotic factor to bind and complex with the mitochondrial membrane-bound anti-apoptotic factors, Bcl-2 and Bcl-xL.¹⁰⁷ The dephosphorylation occurs at primarily two serine residues of human BAD, s75 and s99.108 This dephosphorylation causes BAD to dissociate from the protein 14-3-3 and relocate to the outer mitochondrial membrane (OMM). The dimerization of BAD with Bcl-2 and Bcl-xL (suppression of pro-survival signals) prompt the translocation of BAX to the OMM to initiate the formation of mitochondrial permeability transition pores (mPTP) which exacerbate the apoptotic cascade through the release of cytochrome c from the mitochondria and result in execution of cell death by caspases 3, 6, and 7.¹⁰⁹

As it has been elucidated that CaN plays a major role in programmed cell death, would the changes in CaN expression



Figure 3. Regulation of NFAT by CaN. CaN drives the transcriptional role of the nuclear factor of activated T-cells (NFAT) following activation by the calcium (Ca2+) influx via voltage-gated calcium channels (VGCCs). NFAT is hyperphosphorylated and therefore idle in the cytoplasm (hence, NFAT_c). CaN dephosphorylates its N-terminal phosphosites and reveals its element binding domain (EB). The Ca^{2+} influx triggers further release of Ca^{2+} from the endoplasmic reticulum, driving the translocation of NFAT_c into the nucleus to partner with its nuclear counterpart, NFAT_n. The complex binds to its DNA site and transcribes neuronal requisites essential for growth, maintenance, and survival. The transcription remains as long as intracellular Ca²⁺ levels remain high (further maintained by calcium release activated Ca²⁺ channels or CRAC). The fall of Ca²⁺ levels activates dual-specificity tyrosine-phosphorylation regulated kinases (DYRKs) and triggers the cessation of transcription by the NFAT_c-NFAT_n complex. DYRKs dissociate the complex and prime NFAT_c for rephosphorylation by glycogen synthase kinase 3 β (GSK-3^{β}) and relocation to the cytoplasm, where the next cycle begins contingent to the next calcium drive.

affect this role? Such a query arises from the relatively higher concentrations of the PP2B distributed in the brain. What role would a denser population of constitutive CaN play in influencing neuronal survival? Asai et al. went on to address this in a study where they investigated how deeply integrated CaN was in the intrinsic cell death pathway.¹¹⁰ What the group discovered was that CaN predisposes neurons to the cyt c-caspase 3 dependent pathway more frequently in comparison to other cells in the body. The elevated CaN levels (due to Ca²⁺ overload) render neurons to undergo apoptosis under even normal cell conditions. Thus, Ca²⁺-induced excitotoxicity through NMDARs increased the probability of inducing cell death due to the greater expression and sensitivity of CaN in neurons.

Despite establishing CaN's role in apoptosis, its inhibition alone is not enough to prevent the same, and therefore other mechanisms independent of CaN's dephosphorylation of BAD must exist.¹⁰⁷ This curiosity fueled the search for alternative CaN-linked pathways related to apoptosis. Falling in line with CaN's role in the intrinsic pathway of apoptosis is another recently discovered component of mitochondrial dysfunction also facilitated by CaN during cellular duress: dynamin-related peptide 1 (DRP1). A cytosolic dynamin GTPase, DRP1 was one of the initial fission proteins to be discovered, but an understanding of its function still remains unclear.¹¹¹ Mitochondrial fission is often necessary during biogenesis of new mitochondria, but it may also indicate dysfunction in the organelle and trigger apoptosis.¹¹² DRP1 is proposed to trigger



Figure 4. Calcineurin–mitochondrial (CaN–mitochondrial) connection. (A) At abnormally high calcium (Ca^{2+}) influx rates, such as during elevated glutamate signaling, calcineurin (CaN) becomes highly active and triggers the pathway of apoptosis by dephosphorylating the proapoptotic factor Bcl-2-associated death promoter (BAD) which translocates to the outer mitochondrial membrane (OMM) and dimerizes with the anti-apoptotic factors B-cell lymphphoma-2 (Bcl-2) and B-cell lymphoma-extra large (Bcl-xL). The event spurs another pro-apoptotic factor, Bcl-2-associated-X (BAX), to transition to the OMM and catalyze the formation of the mitochondrial permeability transition pore (mPTP). The sustained high Ca^{2+} load continues to maintain the opening of the mPTP, leading to the release of cytochrome c and formation of the apoptosome, culminating in the caspase cascade leading to apoptosis. (B) CaN has also shown to be involved in triggering mitochondrial fission by dephosphorylating the dynamin-related protein 1 (DRP1). DRP1 translocates to the OMM, forms puncta, and constricts the mitochondria with an oligomeric chain and splices the organelle similar to vesicular cleavage by dynamin. The fission of mitochondria is thought to come before the release of cytochrome c, leading to dysfunction and triggering apoptosis or necrosis based on the rate and severity of the event. Evidence has shown that mitochondrial fission is indicative of mitochondrial dysfunction, and the spherical mitochondrial debris void of cytochrome c implicates the occurrence of cell death pathways.

mitochondrial fission by transferring to the OMM surface and forming puncta of DRP1 to form an oligomeric chain that constricts mitochondria to facilitate fission.¹¹³ This event resembles to the way dynamin cleaves endocytic vessels.¹¹⁴ Being a GTPase in nature, DRP1 uses GTP-hydrolysis to cause scission of mitochondria through the twisting movement of the chain.¹¹⁵ CaN mediates the dephosphorylation at s637 required to activate DRP1 and translocate it to the OMM to initiate fission.¹¹⁶ However, controversy surrounds whether it is BAD-Bcl-xL dimerization or mitochondrial fission that transpires prior to eliciting cell death.¹¹⁷ Some studies suggest that BAX activation and cytochrome c release follow fission in mammalian cells.¹¹⁸ Other studies point to the overexpression of Bik, triggering CICR that fission occurs following Ca²⁺.¹¹⁹ The lack of evidence of elongated mitochondrial void of cytochrome c resulting from mitochondrial fusion suggests that fission, not fusion, is more involved in apoptosis.¹²⁰ This makes sense as healthy cells require the fusion process to create a dynamic interconnected network and therefore take on an elongated morphology in contrast to the spherical shapes encountered in mitochondrial fission¹²¹ (see Figure 4).

This pivotal role of CaN in the intrinsic apoptotic signaling pathway further emphasizes the importance of the balance between phosphorylating and dephosphorylating events that dictate the fate of a cell.

8. CALCINEURIN: ROLE IN NEUROLOGICAL DISORDERS

The complexity of neurological disorders makes it difficult to dissect and diagnose the underlying etiology. However, certain etiologies ranging from dysfunctional VGCCs to disturbed mitochondrial dynamics have been common hallmarks of several neurological disorders.^{122,123} Keeping in perspective that Ca^{2+} is the engine the drives nearly all neuronal processes it is therefore worth investigating mediators that are intimately and heavily dependent on Ca^{2+} signaling, such as CaN.⁴¹ CaN, being an integral sensor and regulator of Ca^{2+} flux, may be deeply involved in many neurodegenerative disorders. The following sections explore the involvement of CaN through the spectrum of neurological disorders.

8.1. Ischemic Stroke. The global burden of stroke is a major medical concern that continues to rise in the oncoming years.¹²⁴ It currently ranks as the second highest cause of mortality worldwide for the elderly population aged 60 years or older.¹²⁵ Ischemic stroke accounts for nearly 85% of strokes (the rest being hemorrhagic and transient in nature).¹²⁶ Needless to say, more efficient therapy is in dire need to treat patients facing a narrow therapeutic window. A glance at the etiology shows that stroke, particularly ischemic stroke, is encompassed by excitotoxicity, oxidative and nitrosative stress, inflammation, and edema.^{127–129} Cerebral ischemia expresses a cumbersome

pathology that threatens neuronal viability not only during ischemia but after reperfusion as well; the underlying mecha-nisms have been well-delineated over the years.^{127,130–133} The ischemic phase sets the stage for cell damage as acidic conditions trigger various membrane pumps to dysfunction and thereby elicit heightened depolarization (Na⁺-H⁺ exchanger pumps more Na^+ , the Na^+/K^+ ATPase slows down, while Na^+/K^+ Ca^{2+} exchanger reverses and raises intracellular Ca^{2+} levels). Due to the elevated intracellular Ca²⁺ load, CaN has been reported to be elevated during cerebral ischemia.¹¹⁰ Reperfusion also entails Ca²⁺ overload (by reduction of extracellular $\mathrm{H}^{\scriptscriptstyle +}$ levels and therefore accelerating $\mathrm{Na}^{\scriptscriptstyle +}/\mathrm{H}^{\scriptscriptstyle +}$ exchange and in turn more Ca2+ loading via Na+/Ca2+ exchange) and consequently further elevates CaN activation. There is also greater $\hat{C}a^{2+}$ uptake by the mitochondria beyond the capacity via the mitochondrial Ca²⁺ uniporter (MCU) due to restoration of the electrochemical potential.¹³³ Often referred to as the "oxygen paradox", this oxidative burst has been believed to be the culprit behind mPTP formation, and reperfusion often sends the cell mainly through necrotic/necroptotic signaling along with apoptotic signaling contingent on the extent of Ca^{2+} loading along with duration and severity of the ischemic milieu.¹³⁴

During the reperfusion phase, studies have discerned CaN's ability to dephosphorylate BAD and induce apoptosis.¹⁰⁷ Furthermore, CaN also dephosphorylates DRP-1 to induce mitochondrial fission and trigger apoptosis (see Table 1).¹¹⁶ Many of the mechanistic pathways still remain cryptic, and an understanding of CaN's behavior throughout not only cerebral ischemia but also other neurodegenerative pathologies is difficult to pinpoint. However, a crosstalk of CaN with other biological molecules during cerebral ischemia may be proposed. In summary, CaN seems to play an integral role in ischemic stroke through the elevated Ca²⁺ influx, by virtue of activating several markers involved in mitochondrial and subsequent apoptotic signaling.

CaN seems to play an integral role in ischemic stroke through the elevated Ca²⁺ influx, by virtue of activating several markers involved in mitochondrial and subsequent apoptotic signaling.

8.2. Alzheimer's Disease (AD). The global burden of Alzheimer's disease (AD) is a major concern as 47 million AD patients have been diagnosed worldwide.¹³⁵ The most prevalent neurodegenerative disease (and predominant cause of dementia) is projected to grow to 76 million cases by 2030. Often reported to impair brain histology, cognitive functions, and behavior as well as deeply impacting the physical, emotional, and financial lifestyle of patients and their close ones, AD is the most lethal and complicated cause of dementia.¹³⁶ The ever-controversial pathology of AD has been hallmarked by accumulative lesions of two degenerate proteins, amyloid- β $(A\beta)$ and τ , and have been extensively studied.^{137,138} As AD is a disease where memories become lost, and CaN is a crucial protein deeply associated with learning and memory, this link therefore warrants a deeper investigation. Several studies honing on amyloidosis have detected abnormally high levels of CaN localized around the A β lesions and furthermore have detected the same in astrocytic processes enveloping amyloid

elevated NFAT exacerbates the inflammatory milieu by signaling for cytokine release and astrocyte and microglial activation⁹

astrocytic and microglial activation, cytokine release and T-cell

stress

activation, oxidative and nitrative

neuroinflammation

ed

pathology involved	potential role of calcineurin (CaN)
ischemia/reperfusion injury, excitotoxicity, oxidative stress, inflammation, mitochondrial dysfunction	calcium-driven BAD and DRP1 dephosphorylation leading to mitochondrial dysfunction and cell death pathways; elevated neurotransmitter exocytotic kinetics ^{107,116}
proteinopathy: neurofibrillary tangles, β -amyloid (A β) plaques, neurotransmitter imbalance, inflammation	A β lesions trigger greater glutamate signaling and calcium excitotoxicity and drives CaN toward mitochondrial dysfunction and neuronal death; CaN accelerates dendritic spine loss and neuritic dystrophy; CaN increases CaMKII dephosphorylation, decreasing AMPAR-mediated LTP ^{139,141}
proteinopathy: <i>a</i> -synuclein, neurotransmitter imbalance	elevated nuclear translocation of NFAT by CaN and increased inflammatory cascade ^{98,145}
CAG expansion repeats leading to mutant huntingtin accumulation	mitochondrial dysfunction due to excessive NMDAR activation by huntingtin; greater dephosphorylation at s421 of huntingtin shows disrupted axonal disruption and BDNF transport ^{188,150}
proteinopathy: TDP-43, C9orf72, and mutant superoxide dismutase (SOD 1 ^{G93A}), excitotoxicity	mitochondrial dysfunction driven by CaN via calcium overload; CaN fails to dephosphorylate TDP-43 after poorly binding to mutant SOD 1 ¹⁵²⁻¹⁵⁷
autoimmune inflammatory demyelination,	CaN is involved in a the cycle of autoimmune T-cell-induced glutamate excitotoxicity that exacerbates neuroinflammation through CaN-activated NFAT transcription of cytokines, mitochondrial dysfunction and apoptosis; CaN may also drive exocytosis of glutamate at axonal terminals ^{36,107,139}
neurotransmitter imbalance (dopamine, glutamate, and GABA), mitochondrial dysfunction	altered synaptic cycling and exocytotic kinetics of neurotransmitters like dopamine, glutamate, and GABA by CaNy subunit absence $^{3/2}$
neurotransmitter imbalance, cerebrovascular injury and impairment	CaN may sequester GABA _A receptors and increase neuronal excitability; mitochondrial dysfunction through greater glutamate signaling and calcium excitotoxicity ^{160,162}
focal/global physical injury, axonal injury, cerebrovascular injury and impairment, consequential neurotransmitter imbalance	CaN initiates astrocyte activation by NFAT transcription and worsens the inflammation; it may also be involved in the subsequent excessive neurotransmitter due to contributions to exo-/endocytosis processes ¹⁶¹

Huntington's disease

amyotrophic lateral

sclerosis

multiple sclerosis

chizophrenia

Parkinson's disease

Alzheimer's disease

ys;

51

Table 1. Summary of Calcineurin'S Role in Neurodegenerative Disorders

neurodegenerative disorder

stroke

epilepsy

traumatic brain

injury

deposits.¹³⁹ Certain studies report CaN to become aberrant due to the calcium overload triggered by $A\beta$ oligomer manipulation of glutamate-dependent cascades, though the mechanisms remain unclear.¹⁴⁰ CaN seems to become dysregulated, upregulating the NFAT translocation and transcription, causing dendritic spine loss and neuritic dystrophy, changes that lie in complete contrast to homeostatic CaN-dependent NFAT that underlies axonal growth, elevated dendritic branching, and overall neuronal survival. Deregulated CaN also increases CaMKII dephosphorylation, which downregulates AMPARdependent LTP. This imbalance between the two enzymes results in greater LTD, synapse weakening, and memory loss (see Table 1).¹⁴¹ Thus, deregulated CaN may spur dendritic spine loss, dysregulate AMPAR-mediated LTP, and trigger apoptosis through mitochondrial-mediated pathways.

8.3. Parkinson's Disease (PD). PD follows AD as the second most burdensome neurodegenerative disease worldwide affecting a wide range of age groups: from 41 per 100 000 in the fourth decade to over 1900 per 100 000 for elderly above 80 years of age.¹⁴²

PD is the most notorious movement disorder plagued by inclusions of aberrant α -synuclein in both the perikaryon and the processes of dopaminergic distributed throughout the substantia nigra pars compacta region.¹⁴³ Studies have reported altered and abnormal Ca²⁺ signaling induced by the synucleinopathy en route to weakening of LTP and memory impairment.¹⁴⁴ CaN, deeply involved with LTP and memory formation, compellingly points to a link in PD. A study elicited CaN's confounding switch between protective Ca²⁺ regulation to a cascade of downstream sequences that exacerbate the toxicity perpetuated by α -synuclein.¹⁴⁵ The study also witnessed enhanced nuclear localization of NFAT, 4 in the brains of PD patients. This association of NFAT to PD further bolsters elevated cytosolic Ca²⁺ load and the failure of CaN. This may be due to several mechanisms unexplored at the time. However, it may be proposed that elevated Ca^{2+} due to α -synuclein may be responsible for overactivating CaN which drives it toward LTP weakening.44 Meanwhile, sustained Ca2+ load prevents the rephosphorylation of NFAT and may explain its retention in the nucleus.⁹⁸ The consequent inflammatory signaling interferes with synaptic connectivity and neurotransmission, exacerbating neuronal survival (see Table 1). Thus, CaN seems to entail LTP weakening, prolonged NFATmediated inflammation, and the consequential breakdown of neurotransmission.

8.4. Huntington's Disease (HD). Hallmarked by the death of medium spiny neurons of the striatum and the consequential aggressive progression of motor, cognitive, and behavioral decline, HD is now the most common monogenic brain disorder in developed nations (Europe and North America) with a prevalence of nearly 10 times more than that of Asian nations.¹⁴⁶ The CAG expansion (beyond 35 repeats) in the gene responsible for the production of huntingtin (HTT) protein has been discovered in nearly 90% of HD patients. Pathology manifests through the aberrant splicing pattern of HTT fragments, particularly those accumulating in HTT exon 1 protein.¹⁴⁷ However, the pathology comprises proteins involved in a vast array of neuronal functions, from regulating the cell cycle to cell signaling, emphasizing the complexity of HD and the quest for more promising therapeutic targets. A link between CaN and HD, associating elevated levels of the phosphatase in the disease after NMDAR activation triggering the intrinsic apoptotic pathway, has been reported.¹⁴⁸ Studies

have pointed to the phosphorylation of s421 evincing an inverse relationship with disease susceptibility.¹⁴⁹ Greater dephosphorylation of s421 seems to disrupt the axonal transport and diminished BDNF transport, promoting neurodegeneration; the inhibition of CaN increased phosphorylation of s421 thereby restoring antero- and retrograde axonal transport in mutant HTT.¹⁵⁰ Mutant HTT has also shown to downregulate CaN and trigger hyperphosphorylation of τ protein in another study without any noticeable changes in τ kinases (see Table 1).¹⁵¹ It can therefore be proposed that elevated CaN, due to hyperactivation of NMDARs (induced by mutant HTT), seems to disrupt axonal and BDNF transport and triggers mitochondrial-mediated apoptotic pathways.

8.5. Amyotrophic Lateral Sclerosis (ALS). ALS is an age-dependent insidious and fatal motor neuron (MN) disease of the cortex, brain stem, and spinal cord, encompassing damage to both upper and lower motoneurons. It has a current global prevalence of 1.9 per 100 000 that threatens to amass by 69% by 2040.¹⁵² Along with the recent breakthrough discovery of a genetic culprit of ALS (an intronic hexanucleotide expansion in the C9orf72 gene), the predominance of pathological studies revolve around proteinopathies and Ca2+ overload excitotoxicity brought about hyperactivated glutamatergic signaling, mainly a ramification of the lack of GluR2 subunits and heightened permeability in AMPARs.^{153,154} The altered intracellular Ca²⁺ milieu sets off a Ca²⁺-dependent cascade, still not clearly understood, which culminates in cell death. The affected neurons eventually lose their Ca²⁺ buffering capacities, and therefore, mitochondrial dysfunction and apoptosis via the intrinsic pathway have been reported.¹⁵⁵ CaN has shown to be highly involved in triggering this pathway during Ca²⁺ overload. Plaques of trans-activating response region DNA binding protein (TDP-43) have been depicted in nearly 90% of ALS cases, and Liachko et al. have found that dephosphorylation of TDP-43 at sites s409/410 by CaN helps to limit toxic protein accumulation.¹⁵⁶ Kim et al. shed further light on the interaction between TDP-43 and CaN when the group observed weakened dephosphorylation of TDP-43 by CaN after its poor binding with mutated superoxide dismutase 1 (SOD 1^{G93A}).¹⁵⁷ Furthermore, TDP-43 pathology is now believed to drive ALS progression along with being a factor in other proteinopathies like AD and PD (see Table 1). CaN seems to therefore exacerbate neuronal damage through its role in mitochondrial dysfunction as well through its failure of limiting the intracellular toxic TDP-43 levels due to interfering binding by SOD 1^{G93A}

8.6. Multiple Sclerosis (MS). MS is a chronic, autoimmune inflammatory neurodegenerative disease that plagues about 2.5 million people worldwide.¹⁵⁸ Glutamate-induced excitotoxicity has been implicated to be a driving force in neuronal death. Studies report that inflammatory cells in foci ensuing autoimmune inflammatory insult are a major source of extracellular glutamate, releasing it via cysteine/glutamate transporters Xc^{-.159} The surplus glutamate not only further activates more T-cells via their upregulated expression of NMDARs, but it also triggers the rapid influx of Ca²⁺ in neuronal cells that accelerates pro-apoptotic signaling. CaN is speculated to play a role in MS by virtue of the large intracellular Ca²⁺ load driving it to facilitate the intrinsic pathway and mitochondrial dysfunction.¹⁰⁷ It also can be postulated that excitotoxicity drives the exocytotic kinetics through the activated CaN and synapsin 1 complex to release greater glutamate, which fuels the vicious T-cell-mediated autoimmunity and exacerbates excitotoxic milieu

in neighboring neurons (see Table 1).^{36,159} CaN, by virtue of Ca²⁺ overload, seems to accelerate pro-apoptotic signaling as well drive the excess exocytosis of glutamate into the synapse, exacerbating inflammation-induced neurodegeneration. Such a cascade may deserve deeper investigation in future studies concerning MS and the role of CaN.

9. OTHER NEURODEGENERATIVE DISORDERS

CaN is involved in other intricate neurological pathologies (Table 1). Knockdown of the CaN γ subunit has shown to alter synaptic vesicle cycling and has shown to cause psychiatric disorders like schizophrenia.³⁷ In an epileptic study, CaN has shown to augment neuronal excitability by greater endocytosis and internalization of GABA_A receptors, disturbing the excitatory/inhibitory equilibrium to prevent inhibitory actions of GABA that mitigate the synchronous neuronal discharge responsible for seizure activity.¹⁶⁰ The same study also showed the prevention of mitochondrial dysfunction by inhibition of CaN with cyclosporin A (CsA), a cyclophilin D binding drug that complexes with CaN to render it inactive and thereby preventing the opening of mPTP. A study based on traumatic brain injury (TBI) showed how the CaN/NFAT pathway following TBI induced astrocyte activation that further compounded the inflammatory environment and escalated neuronal damage.¹⁶¹ Neuroinflammation is a major component in CNS disorders and acute and chronic brain injury. Inhibition of CaN mitigated the detrimental activation of NFAT that transcribes inflammatory markers and recruits astrocytes and glial cells to the site of injury. In neuroinflammation studies, CaN plays both anti- and pro-inflammatory roles based on its interactions with transcription factors and displays a complex but unique dichotomy in mediating inflammatory milieu. For instance, it can associate with NFAT and activator protein 1 (AP1) to activate T-cells while inducing their anergy and tolerance by associating NFAT with forkhead box P3 (FOXP3).9 In conjunction, activation of Nuclear Factor KB (NFKB) and NFAT triggers the transcription of cytokines, activating astrocytes and microglia while promoting T-cell expansion in the CNS tissue, ultimately leading to inflammation-mediated neuronal damage and cell death.

10. IMMUNOSUPPRESSION OF CaN AND ITS EFFECTS ON THE BRAIN

CaN inhibitors (CNIs), namely, cyclosporine A and tacrolimus, have been immensely utilized in the field of organ transplantation, becoming the cornerstone therapy for preventing immune rejections.¹⁶³ However, their effects on the brain remain a concern. CNI therapy used in kidney transplant patients has elicited neurological symptoms such as recurrent headaches, tremors, seizures, extrapyramidal syndrome, encephalopathy, and posterior encephalopathy syndrome.¹⁶⁴ Other clinical studies have shown neurotoxicity with the initiation of CNI therapy.^{165,166} A recent study focusing on the neural effects of CNIs in orthotopic liver transplant patients by Pflugrad et al. showcased cognitive decline and altered brain structure.¹⁶⁵ The neurological adverse reactions observed in CNI therapy further emphasize the complex role of CaN in the brain. Inhibiting CaN with CNIs alone to treat neurological disorders is still up for debate and requires extensive investigation to overcome the associated neurotoxicity. Furthermore, safer drugs void of neurotoxic side effects are desperately required in organ transplantation.

There are certain risk factors that can be assessed prior to organ transplantation that may help avoid possible neurologic complications. Encephalopathy, although often indicated in solid organ transplant patients, can be assessed by varying characteristic abnormalities observed in magnetic resonance imaging (MRI). For instance, altered signal changes can be observed in the insular and cingulated cortices with relatively lesser changes in occipital and perirolandic cortex may indicate possible neurotoxicity.¹⁶⁶ Other diagnostic suggestions involve the requirement for more sensitive Doppler devices to monitor intracranial pressure noninvasively, monitoring of EEG levels post-transplant, and impaired verbal and motor responsiveness without electrolyte imbalance, tinnitus, or hearing loss (improved after discontinuation of tacrolimus).¹⁶⁷Alternative therapies to CNI involve monoclonal antibodies (belatacept),¹⁶⁸ combinations of mycophenolate mofetil and sirolimus,¹⁶⁹ along with newer CNI-free therapies in the pipeline.¹⁷⁰ CNI-free immunosuppressant therapy may prove to be beneficial in organ transplant patients in the future.¹⁷

11. CONCLUSION

Neurodegenerative disorders present a major conundrum in medical science. The complex neuropathologies, despite sharing common pathological manifestations, arise from dysregulated crosstalk and genetic expressions that are difficult to trace. Some biological molecules involved in regulating much of the neuronal functions may be honed in on as potential therapeutic targets. CaN is one such target that is inveterate to running essential neuronal components such as regulating Ca²⁺ homeostasis and Ca²⁺-dependent downstream processes, synaptic plasticity, neurotransmission release, and synaptic cycling. Furthermore, it is involved in regulating transcription processes and the overall neuronal survival, operation, and maintenance. Due to such a diversified and crucial biological role, CaN may lie at the crux of many neurodegenerative disorders. From stroke to proteinopathies to inflammatory pathologies, CaN has been reported in underlying mechanistic pathways. However, it is rather perplexing to pinpoint whether CaN serves to protect and salvage neurons during neurological disorders or whether it perpetuates neuronal damage. The convoluted pathways of CaN exhibit its rescue of one aspect of the cell while deteriorating another during neuropathic conditions. Furthermore, some pathways are not truly delineated due to their erratic and volatile dispositions. Retentive in mind should also be that simple inhibition of an endogenous molecule as vital as CaN is to neuronal function alone may bring more harm than good in neurotherapy. Although many of the signaling cascades remain to be elucidated, and manipulating CaN may seem daunting, CaN warrants a deeper investigation. By bringing a greater focus to understanding its roles, it may be possible to strategize methods to modulate CaN as a potential therapeutic target to alleviate neurological disorders.

AUTHOR INFORMATION

Corresponding Author

*Phone: +91-79 66745555. Fax: +91-79 66745560. E-mail: pallab.bhu@gmail.com; pallab.bhattacharya@niperahm.ac.in. ORCID ©

Pallab Bhattacharya: 0000-0003-2867-1650

Author Contributions

J.S., P.B., K.R.D., and D.R.Y. conceived and designed the study. J.S., D.S., H.K., K.R.D., and P.B. outlined the performed

rigorous literature search. J.S., D.S., H.K., K.K., and P.B. conceived and designed the figures and images. J.S., P.B., K.R.D., K.K., and D.R.Y. wrote the manuscript.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors acknowledge Department of Science and Technology (DST), Government of India, for their financial support through Grant (SB/YS/LS-196/2014), International Society for Neurochemistry (ISN) Return Home Grant, Department of Pharmaceuticals, Ministry of Chemical and Fertilizers, Government of India, and National Institute of Pharmaceutical Education and Research (NIPER) Ahmedabad, Gandhinagar, India. The authors also want to express their thanks to Prof. Larry Benowitz, Boston Children's Hospital, Harvard Medical School, Boston, MA, United States, and the Director, NIPER Ahmedabad, for providing necessary support.

REFERENCES

(1) Wang, J. H.; Desai, R. A brain protein and its effect on the Ca²⁺and protein modulator-activated cyclic nucleotide phosphodiesterase. *Biochem. Biophys. Res. Commun.* **1976**, 72 (3), 926.

(2) Watterson, D. M.; Vanaman, T. C. Affinity chromatography purification of a cyclic nucleotide phosphodiesterase using immobilized modulator protein, a troponin C-like protein from brain. *Biochem. Biophys. Res. Commun.* **1976**, 73 (1), 40.

(3) Klee, C.; Krinks, M. Purification of cyclic 3', 5'-nucleotide phosphodiesterase inhibitory protein by affinity chromatography on activator protein coupled to Sepharose. *Biochemistry* **1978**, *17* (1), 120.

(4) Wallace, R. W.; Lynch, T. J.; Tallant, E. A.; Cheung, W. Y. An endogenous inhibitor protein of brain adenylate cyclase and cyclic nucleotide phosphodiesterase. *Arch. Biochem. Biophys.* **1978**, *187* (2), 328.

(5) Wallace, R. W.; Lynch, T. J.; Tallant, E. A.; Cheung, W. Y. Purification and characterization of an inhibitor protein of brain adenylate cyclase and cyclic nucleotide phosphodiesterase. *J. Biol. Chem.* **1979**, 254 (2), 377.

(6) Klee, C.; Crouch, T.; Krinks, M. Calcineurin: a calcium-and calmodulin-binding protein of the nervous system. *Proc. Natl. Acad. Sci. U. S. A.* **1979**, *76* (12), 6270.

(7) Stewart, A. A.; Ingebritsen, T. S.; Manalan, A.; Klee, C. B.; Cohen, P. Discovery of A Ca2+-and calmodulin-dependent protein phosphatase. *FEBS Lett.* **1982**, *137* (1), 80.

(8) INGEBRITSEN, T. S.; COHEN, P. The protein phosphatases involved in cellular regulation. *Eur. J. Biochem.* **1983**, *132* (2), 255.

(9) Furman, J. L.; Norris, C. M. Calcineurin and glial signaling: neuroinflammation and beyond. *J. Neuroinflammation* **2014**, *11* (1), 158.

(10) Liu, J.; Farmer, J. D., Jr; Lane, W. S.; Friedman, J.; Weissman, I.; Schreiber, S. L. Calcineurin is a common target of cyclophilincyclosporin A and FKBP-FK506 complexes. *Cell* **1991**, *66* (4), 807.

(11) Carriedo, S. G.; Yin, H. Z.; Sensi, S. L.; Weiss, J. H. Rapid Ca2+ entry through Ca2+-permeable AMPA/Kainate channels triggers marked intracellular Ca2+ rises and consequent oxygen radical production. *J. Neurosci.* **1998**, *18* (19), 7727.

(12) Cheng, C.; Fass, D. M.; Reynolds, I. J. Emergence of excitotoxicity in cultured forebrain neurons coincides with larger glutamate-stimulated [Ca2+] i increases and NMDA receptor mRNA levels. *Brain Res.* **1999**, *849* (1–2), *97*.

(13) Foster, T. C.; Sharrow, K. M.; Masse, J. R.; Norris, C. M.; Kumar, A. Calcineurin links Ca2+ dysregulation with brain aging. *J. Neurosci.* **2001**, *21* (11), 4066.

(14) Ingebritsen, T. S.; Cohen, P. Protein phosphatases: properties and role in cellular regulation. *Science* **1983**, *221* (4608), 331.

(15) Cohen, P. The structure and regulation of protein phosphatases. *Annu. Rev. Biochem.* **1989**, 58 (1), 453.

(16) Cohen, P.; Cohen, P. Protein phosphatases come of age. J. Biol. Chem. **1989**, 264 (36), 21435.

(17) Klee, C.; Draetta, G.; Hubbard, M. Calcineurin. Adv. Enzymol. Relat. Areas Mol. Biol. 2006, 61, 149.

(18) Griffith, J. P.; Kim, J. L.; Kim, E. E.; Sintchak, M. D.; Thomson, J. A.; Fitzgibbon, M. J.; Fleming, M. A.; Caron, P. R.; Hsiao, K.; Navia, M. A. X-ray structure of calcineurin inhibited by the immunophilinimmunosuppressant FKBP12-FK506 complex. *Cell* **1995**, *82* (3), 507.

(19) Coghlan, V. M.; Perrino, B. A.; Howard, M.; Langeberg, L. K.; Hicks, J. B.; Gallatin, W. M.; Scott, J. D. Association of protein kinase A and protein phosphatase 2B with a common anchoring protein. *Science* **1995**, *267* (5194), 108.

(20) Kissinger, C. R.; Parge, H. E.; Knighton, D. R.; Lewis, C. T.; Pelletier, L. A.; Tempczyk, A.; Kalish, V. J.; Tucker, K. D.; Showalter, R. E.; Moomaw, E. W. Crystal structures of human calcineurin and the human FKBP12–FK506–calcineurin complex. *Nature* **1995**, *378* (6557), 641.

(21) Feng, B.; Stemmer, P. M. Interactions of calcineurin A, calcineurin B, and Ca2+. *Biochemistry* **1999**, *38* (38), 12481.

(22) Gallagher, S. C.; Gao, Z.-H.; Li, S.; Dyer, R. B.; Trewhella, J.; Klee, C. B. There is communication between all four Ca2+-bindings sites of calcineurin B. *Biochemistry* **2001**, *40* (40), 12094.

(23) Shen, X.; Li, H.; Ou, Y.; Tao, W.; Dong, A.; Kong, J.; Ji, C.; Yu, S. The secondary structure of calcineurin regulatory region and conformational change induced by calcium/calmodulin binding. *J. Biol. Chem.* **2008**, *283* (17), 11407.

(24) Jiang, H.; Xiong, F.; Kong, S.; Ogawa, T.; Kobayashi, M.; Liu, J. O. Distinct tissue and cellular distribution of two major isoforms of calcineurin. *Mol. Immunol.* **1997**, *34* (8–9), 663.

(25) Solà, C.; Tusell, J. M.; Serratosa, J. Comparative study of the distribution of calmodulin kinase II and calcineurin in the mouse brain. *J. Neurosci. Res.* **1999**, *57* (5), *6*51.

(26) Yakel, J. L. Calcineurin regulation of synaptic function: from ion channels to transmitter release and gene transcription. *Trends Pharmacol. Sci.* **1997**, *18* (4), 124.

(27) Miller, R. J. Multiple calcium channels and neuronal function. *Science* **1987**, 235 (4784), 46.

(28) Kennedy, M. B. Regulation of neuronal function by calcium. *Trends Neurosci.* **1989**, *12* (11), 417.

(29) Berridge, M. J.; Lipp, P.; Bootman, M. D. The versatility and universality of calcium signalling. *Nat. Rev. Mol. Cell Biol.* 2000, 1 (1), 11.

(30) Bading, H. Nuclear calcium signalling in the regulation of brain function. *Nat. Rev. Neurosci.* **2013**, *14* (9), 593.

(31) Mansuy, I. M. Calcineurin in memory and bidirectional plasticity. *Biochem. Biophys. Res. Commun.* 2003, 311 (4), 1195.

(32) Verstegen, A. M.; Tagliatti, E.; Lignani, G.; Marte, A.; Stolero, T.; Atias, M.; Corradi, A.; Valtorta, F.; Gitler, D.; Onofri, F. Phosphorylation of synapsin I by cyclin-dependent kinase-5 sets the ratio between the resting and recycling pools of synaptic vesicles at hippocampal synapses. *J. Neurosci.* **2014**, *34* (21), 7266.

(33) Alabi, A. A.; Tsien, R. W. Synaptic vesicle pools and dynamics. Cold Spring Harbor Perspect. Biol. 2012, 4 (8), a013680.

(34) Cottrell, J. R.; Levenson, J. M.; Kim, S. H.; Gibson, H. E.; Richardson, K. A.; Sivula, M.; Li, B.; Ashford, C. J.; Heindl, K. A.; Babcock, R. J. Working memory impairment in calcineurin knock-out mice is associated with alterations in synaptic vesicle cycling and disruption of high-frequency synaptic and network activity in prefrontal cortex. J. Neurosci. 2013, 33 (27), 10938.

(35) Kim, S. H.; Ryan, T. A. CDK5 serves as a major control point in neurotransmitter release. *Neuron* **2010**, *67* (5), 797.

(36) Kim, S. H.; Ryan, T. A. Balance of calcineurin A α and CDK5 activities sets release probability at nerve terminals. *J. Neurosci.* **2013**, 33 (21), 8937.

(37) Cottrell, J. R.; Li, B.; Kyung, J. W.; Ashford, C. J.; Mann, J. J.; Horvath, T. L.; Ryan, T. A.; Kim, S. H.; Gerber, D. J. Calcineurin Aγ is a functional phosphatase that modulates synaptic vesicle endocytosis. J. Biol. Chem. 2016, 291 (4), 1948.

(38) Ferguson, S. M.; De Camilli, P. Dynamin, a membraneremodelling GTPase. *Nat. Rev. Mol. Cell Biol.* **2012**, *13* (2), 75.

(39) Clayton, E. L.; Anggono, V.; Smillie, K. J.; Chau, N.; Robinson, P. J.; Cousin, M. A. The phospho-dependent dynamin–syndapin interaction triggers activity-dependent bulk endocytosis of synaptic vesicles. *J. Neurosci.* **2009**, *29* (24), 7706.

(40) Wu, X.-S.; Zhang, Z.; Zhao, W.-D.; Wang, D.; Luo, F.; Wu, L.-G. Calcineurin is universally involved in vesicle endocytosis at neuronal and nonneuronal secretory cells. *Cell Rep.* **2014**, *7* (4), 982.

(41) Berridge, M. J. Neuronal calcium signaling. *Neuron* 1998, 21 (1), 13.

(42) Ross, W. N. Understanding calcium waves and sparks in central neurons. *Nat. Rev. Neurosci.* **2012**, *13* (3), 157.

(43) Wang, L.-Y.; Orser, B. A.; Brautigan, D. L.; MacDonald, J. F. Regulation of NMDA receptors in cultured hippocampal neurons by protein phosphatases 1 and 2A. *Nature* **1994**, *369* (6477), 230.

(44) Krupp, J. J.; Vissel, B.; Thomas, C. G.; Heinemann, S. F.; Westbrook, G. L. Calcineurin acts via the C-terminus of NR2A to modulate desensitization of NMDA receptors. *Neuropharmacology* **2002**, 42 (5), 593.

(45) Fino, E.; Paille, V.; Cui, Y.; Morera-Herreras, T.; Deniau, J. M.; Venance, L. Distinct coincidence detectors govern the corticostriatal spike timing-dependent plasticity. *J. Physiol.* **2010**, *588* (16), 3045.

(46) Plotkin, J. L.; Day, M.; Surmeier, D. J. Synaptically driven state transitions in distal dendrites of striatal spiny neurons. *Nat. Neurosci.* **2011**, *14* (7), 881.

(47) Oliveria, S. F.; Dell'Acqua, M. L.; Sather, W. A. AKAP79/150 anchoring of calcineurin controls neuronal L-type Ca2+ channel activity and nuclear signaling. *Neuron* **2007**, *55* (2), 261.

(48) Budde, T.; Meuth, S.; Pape, H.-C. Calcium-dependent inactivation of neuronal calcium channels. *Nat. Rev. Neurosci.* 2002, 3 (11), 873.

(49) Dittmer, P. J.; Dell'Acqua, M. L.; Sather, W. A. Ca2+/ calcineurin-dependent inactivation of neuronal L-type Ca2+ channels requires priming by AKAP-anchored protein kinase A. *Cell Rep.* **2014**, 7 (5), 1410.

(50) Cameron, A. M.; Steiner, J. P.; Roskams, A. J.; Ali, S. M.; Ronnettt, G. V.; Snyder, S. H. Calcineurin associated with the inositol 1, 4, 5-trisphosphate receptor-FKBP12 complex modulates Ca2+ flux. *Cell* **1995**, 83 (3), 463.

(51) Sun, H.; Leblanc, N.; Nattel, S. Mechanisms of inactivation of L-type calcium channels in human atrial myocytes. *American Journal of Physiology-Heart and Circulatory Physiology* **1997**, 272 (4), H1625.

(52) Sham, J.; Cleemann, L.; Morad, M. Functional coupling of Ca2+ channels and ryanodine receptors in cardiac myocytes. *Proc. Natl. Acad. Sci. U. S. A.* **1995**, *92* (1), 121.

(53) Fabiato, A. Calcium-induced release of calcium from the cardiac sarcoplasmic reticulum. *American Journal of Physiology-Cell Physiology* **1983**, 245 (1), C1.

(54) Arendt, K. L.; Zhang, Z.; Ganesan, S.; Hintze, M.; Shin, M. M.; Tang, Y.; Cho, A.; Graef, I. A.; Chen, L. Calcineurin mediates homeostatic synaptic plasticity by regulating retinoic acid synthesis. *Proc. Natl. Acad. Sci. U. S. A.* **2015**, *112* (42), E5744.

(55) Turrigiano, G. Homeostatic synaptic plasticity: local and global mechanisms for stabilizing neuronal function. *Cold Spring Harbor Perspect. Biol.* **2012**, *4* (1), a005736.

(56) Stellwagen, D.; Malenka, R. C. Synaptic scaling mediated by glial TNF- α . *Nature* **2006**, 440 (7087), 1054.

(57) Kim, S.; Ziff, E. B. Calcineurin mediates synaptic scaling via synaptic trafficking of Ca2+-permeable AMPA receptors. *PLoS Biol.* **2014**, *12* (7), e1001900.

(58) Graef, I. A.; Mermelstein, P. G.; Stankunas, K.; Neilson, J. R.; Deisseroth, K.; Tsien, R. W.; Crabtree, G. R. L-type calcium channels and GSK-3 regulate the activity of NF-ATc4 in hippocampal neurons. *Nature* **1999**, 401 (6754), 703.

(59) Zeng, H.; Chattarji, S.; Barbarosie, M.; Rondi-Reig, L.; Philpot, B. D.; Miyakawa, T.; Bear, M. F.; Tonegawa, S. Forebrain-specific

calcineurin knockout selectively impairs bidirectional synaptic plasticity and working/episodic-like memory. *Cell* **2001**, *107* (5), 617. (60) Malleret, G.; Haditsch, U.; Genoux, D.; Jones, M. W.; Bliss, T. V.; Vanhoose, A. M.; Weitlauf, C.; Kandel, E. R.; Winder, D. G.; Mansuy, I. M. Inducible and reversible enhancement of learning, memory, and long-term potentiation by genetic inhibition of calcineurin. *Cell* **2001**, *104* (5), 675.

(61) Giese, K. P.; Mizuno, K. The roles of protein kinases in learning and memory. *Learn. Mem.* **2013**, *20* (10), 540.

(62) Lisman, J.; Yasuda, R.; Raghavachari, S. Mechanisms of CaMKII action in long-term potentiation. *Nat. Rev. Neurosci.* **2012**, *13* (3), 169.

(63) Baumgärtel, K.; Mansuy, I. M. Neural functions of calcineurin in synaptic plasticity and memory. *Learn. Mem.* **2012**, *19* (9), 375.

(64) Beattie, E. C.; Carroll, R. C.; Yu, X.; Morishita, W.; Yasuda, H.; von Zastrow, M.; Malenka, R. C. Regulation of AMPA receptor endocytosis by a signaling mechanism shared with LTD. *Nat. Neurosci.* **2000**, 3 (12), 1291.

(65) Jones, M. V.; Westbrook, G. L. Shaping of IPSCs by endogenous calcineurin activity. J. Neurosci. **1997**, *17* (20), 7626.

(66) Lu, Y. M.; Mansuy, I. M.; Kandel, E. R.; Roder, J. Calcineurinmediated LTD of GABAergic inhibition underlies the increased excitability of CA1 neurons associated with LTP. *Neuron* **2000**, *26* (1), 197.

(67) Wang, J.; Liu, S.; Haditsch, U.; Tu, W.; Cochrane, K.; Ahmadian, G.; Tran, L.; Paw, J.; Wang, Y.; Mansuy, I. Interaction of calcineurin and type-A GABA receptor $\gamma 2$ subunits produces longterm depression at CA1 inhibitory synapses. *J. Neurosci.* **2003**, *23* (3), 826.

(68) Winder, D. G.; Mansuy, I. M.; Osman, M.; Moallem, T. M.; Kandel, E. R. Genetic and pharmacological evidence for a novel, intermediate phase of long-term potentiation suppressed by calcineurin. *Cell* **1998**, *92* (1), 25.

(69) Lüscher, C.; Malenka, R. C. NMDA receptor-dependent longterm potentiation and long-term depression (LTP/LTD). *Cold Spring Harbor Perspect. Biol.* **2012**, *4* (6), a005710.

(70) Malenka, R. C.; Bear, M. F. LTP and LTD: an embarrassment of riches. *Neuron* 2004, 44 (1), 5.

(71) Lin, L.; Sun, W.; Kung, F.; Dell'Acqua, M. L.; Hoffman, D. A. AKAP79/150 impacts intrinsic excitability of hippocampal neurons through phospho-regulation of A-type K+ channel trafficking. *J. Neurosci.* **2011**, *31* (4), 1323.

(72) Yao, J.-j.; Zhao, Q.-R.; Liu, D.-D.; Chow, C.-W.; Mei, Y.-A. Neuritin Up-regulates Kv4. 2 α -Subunit of Potassium Channel Expression and Affects Neuronal Excitability by Regulating the Calcium-Calcineurin-NFAT_c4 Signaling Pathway. *J. Biol. Chem.* **2016**, 291 (33), 17369.

(73) Jinno, S.; Jeromin, A.; Kosaka, T. Postsynaptic and extrasynaptic localization of Kv4. 2 channels in the mouse hippocampal region, with special reference to targeted clustering at gabaergic synapses. *Neuroscience* **2005**, *134* (2), 483.

(74) Alonso, G.; Widmer, H. Clustering of KV4. 2 potassium channels in postsynaptic membrane of rat supraoptic neurons: an ultrastructural study. *Neuroscience* **1997**, 77 (3), 617.

(75) Kim, J.; Wei, D. S.; Hoffman, D. A. Kv4 potassium channel subunits control action potential repolarization and frequency-dependent broadening in rat hippocampal CA1 pyramidal neurones. *J. Physiol.* **2005**, *569* (1), 41.

(76) Hoffman, D. A.; Magee, J. C.; Colbert, C. M.; Johnston, D. K+ channel regulation of signal propagation in dendrites of hippocampal pyramidal neurons. *Nature* **1997**, *387* (6636), 869.

(77) Yashiro, K.; Philpot, B. D. Regulation of NMDA receptor subunit expression and its implications for LTD, LTP, and metaplasticity. *Neuropharmacology* **2008**, 55 (7), 1081.

(78) Schrader, L. A.; Anderson, A. E.; Mayne, A.; Pfaffinger, P. J.; Sweatt, J. D. PKA modulation of Kv4. 2-encoded A-type potassium channels requires formation of a supramolecular complex. *J. Neurosci.* **2002**, 22 (23), 10123. (79) Lockridge, A.; Yuan, L. L. Spatial learning deficits in mice lacking A-type K+ channel subunits. *Hippocampus* **2011**, *21* (11), 1152.

(80) Traynelis, S. F.; Wollmuth, L. P.; McBain, C. J.; Menniti, F. S.; Vance, K. M.; Ogden, K. K.; Hansen, K. B.; Yuan, H.; Myers, S. J.; Dingledine, R. Glutamate receptor ion channels: structure, regulation, and function. *Pharmacol. Rev.* **2010**, *62* (3), 405.

(81) Shepherd, J. D.; Huganir, R. L. The cell biology of synaptic plasticity: AMPA receptor trafficking. *Annu. Rev. Cell Dev. Biol.* 2007, 23, 613.

(82) Lee, H.-K.; Takamiya, K.; Han, J.-S.; Man, H.; Kim, C.-H.; Rumbaugh, G.; Yu, S.; Ding, L.; He, C.; Petralia, R. S. Phosphorylation of the AMPA receptor GluR1 subunit is required for synaptic plasticity and retention of spatial memory. *Cell* **2003**, *112* (5), 631.

(83) Man, H.-Y.; Sekine-Aizawa, Y.; Huganir, R. L. Regulation of α amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor trafficking through PKA phosphorylation of the Glu receptor 1 subunit. *Proc. Natl. Acad. Sci. U. S. A.* **2007**, 104 (9), 3579.

(84) Roche, K. W.; O'Brien, R. J.; Mammen, A. L.; Bernhardt, J.; Huganir, R. L. Characterization of multiple phosphorylation sites on the AMPA receptor GluR1 subunit. *Neuron* **1996**, *16* (6), 1179.

(85) Lee, H.-K.; Kameyama, K.; Huganir, R. L.; Bear, M. F. NMDA induces long-term synaptic depression and dephosphorylation of the GluR1 subunit of AMPA receptors in hippocampus. *Neuron* **1998**, *21* (5), 1151.

(86) Jong, Y.-J. I.; Kumar, V.; O'Malley, K. L. Intracellular metabotropic glutamate receptor 5 (mGluR5) activates signaling cascades distinct from cell surface counterparts. *J. Biol. Chem.* **2009**, 284 (51), 35827.

(87) Fagni, L.; Chavis, P.; Ango, F.; Bockaert, J. Complex interactions between mGluRs, intracellular Ca2+ stores and ion channels in neurons. *Trends Neurosci.* **2000**, *23* (2), 80.

(88) Alagarsamy, S.; Saugstad, J.; Warren, L.; Mansuy, I. M.; Gereau IV, R. W.; Conn, P. J. NMDA-induced potentiation of mGluR5 is mediated by activation of protein phosphatase 2B/calcineurin. *Neuropharmacology* **2005**, *49*, 135.

(89) Crabtree, G. R.; Olson, E. N. NFAT signaling: choreographing the social lives of cells. *Cell* **2002**, *109* (2), S67.

(90) Macian, F. NFAT proteins: key regulators of T-cell development and function. *Nat. Rev. Immunol.* 2005, 5 (6), 472.

(91) Crabtree, G. R.; Schreiber, S. L. SnapShot: Ca2+-calcineurin-NFAT signaling. *Cell* **2009**, *138* (1), 210.

(92) Shin, S.-Y.; Yang, H. W.; Kim, J.-R.; Do Heo, W.; Cho, K.-H. A hidden incoherent switch regulates RCAN1 in the calcineurin–NFAT signaling network. *J. Cell Sci.* **2011**, *124* (1), 82.

(93) Kipanyula, M. J.; Kimaro, W. H.; Etet, P. F. The emerging roles of the calcineurin-nuclear factor of activated T-lymphocytes pathway in nervous system functions and diseases. *Journal of Aging Research* **2016**, 2016, 5081021.

(94) Shin, S.-Y.; Choo, S.-M.; Kim, D.; Baek, S. J.; Wolkenhauer, O.; Cho, K.-H. Switching feedback mechanisms realize the dual role of MCIP in the regulation of calcineurin activity. *FEBS Lett.* **2006**, *580* (25), 5965.

(95) Parekh, A. B.; Putney, J. W., Jr Store-operated calcium channels. *Physiol. Rev.* **2005**, *85* (2), 757.

(96) Nguyen, T.; Di Giovanni, S. NFAT signaling in neural development and axon growth. *Int. J. Dev. Neurosci.* **2008**, *26* (2), 141.

(97) Gwack, Y.; Sharma, S.; Nardone, J.; Tanasa, B.; Iuga, A.; Srikanth, S.; Okamura, H.; Bolton, D.; Feske, S.; Hogan, P. G. A genome-wide Drosophila RNAi screen identifies DYRK-family kinases as regulators of NFAT. *Nature* **2006**, *441* (7093), 646.

(98) Beals, C. R.; Sheridan, C. M.; Turck, C. W.; Gardner, P.; Crabtree, G. R. Nuclear export of NF-ATc enhanced by glycogen synthase kinase-3. *Science* **1997**, *275* (5308), 1930.

(99) Green, D. R.; Reed, J. C. Mitochondria and apoptosis. *Science* **1998**, *281* (5381), 1309–1312.

(100) Rossé, T.; Olivier, R.; Monney, L.; Rager, M.; Conus, S.; Fellay, I.; Jansen, B.; Borner, C. Bcl-2 prolongs cell survival after Baxinduced release of cytochrome c. *Nature* **1998**, *391* (6666), 496.

(101) Nicholls, D. G.; Budd, S. L. Mitochondria and neuronal survival. *Physiol. Rev.* 2000, 80 (1), 315.

(102) Green, D. R.; Kroemer, G. The pathophysiology of mitochondrial cell death. *Science* **2004**, 305 (5684), 626.

(103) Guyton, K. Z.; Liu, Y.; Gorospe, M.; Xu, Q.; Holbrook, N. J. Activation of mitogen-activated protein kinase by ho role in cell survival following oxidant injury. *J. Biol. Chem.* **1996**, *271* (8), 4138. (104) Philpott, K. L.; McCarthy, M. J.; Klippel, A.; Rubin, L. L.

Activated phosphatidylinositol 3-kinase and Akt kinase promote survival of superior cervical neurons. *J. Cell Biol.* **1997**, *139* (3), 809.

(105) Chen, J.; Fujii, K.; Zhang, L.; Roberts, T.; Fu, H. Raf-1 promotes cell survival by antagonizing apoptosis signal-regulating kinase 1 through a MEK–ERK independent mechanism. *Proc. Natl. Acad. Sci. U. S. A.* **2001**, *98* (14), 7783.

(106) Datta, S. R.; Ranger, A. M.; Lin, M. Z.; Sturgill, J. F.; Ma, Y.-C.; Cowan, C. W.; Dikkes, P.; Korsmeyer, S. J.; Greenberg, M. E. Survival factor-mediated BAD phosphorylation raises the mitochondrial threshold for apoptosis. *Dev. Cell* **2002**, *3* (5), 631.

(107) Wang, H.-G.; Pathan, N.; Ethell, I. M.; Krajewski, S.; Yamaguchi, Y.; Shibasaki, F.; McKeon, F.; Bobo, T.; Franke, T. F.; Reed, J. C. Ca2+-induced apoptosis through calcineurin dephosphorylation of BAD. *Science* **1999**, *284* (5412), 339.

(108) Niemi, N. M.; MacKeigan, J. P. Mitochondrial phosphorylation in apoptosis: flipping the death switch. *Antioxid. Redox Signaling* **2013**, *19* (6), 572.

(109) Harris, M.; Thompson, C. The role of the Bcl-2 family in the regulation of outer mitochondrial membrane permeability. *Cell Death Differ.* **2000**, 7 (12), 1182.

(110) Asai, A.; Qiu, J.-h.; Narita, Y.; Chi, S.; Saito, N.; Shinoura, N.; Hamada, H.; Kuchino, Y.; Kirino, T. High level calcineurin activity predisposes neuronal cells to apoptosis. *J. Biol. Chem.* **1999**, 274 (48), 34450.

(111) Slupe, A. M.; Merrill, R. A.; Flippo, K. H.; Lobas, M. A.; Houtman, J. C.; Strack, S. A calcineurin docking motif (LXVP) in dynamin-related protein 1 contributes to mitochondrial fragmentation and ischemic neuronal injury. *J. Biol. Chem.* **2013**, *288* (17), 12353.

(112) Manczak, M.; Reddy, P. H. Abnormal interaction between the mitochondrial fission protein Drp1 and hyperphosphorylated tau in Alzheimer's disease neurons: implications for mitochondrial dysfunction and neuronal damage. *Hum. Mol. Genet.* **2012**, *21* (11), 2538.

(113) Qi, X.; Qvit, N.; Su, Y.-C.; Mochly-Rosen, D. A novel Drp1 inhibitor diminishes aberrant mitochondrial fission and neurotoxicity. *J. Cell Sci.* **2013**, *126* (3), 789.

(114) Faelber, K.; Held, M.; Gao, S.; Posor, Y.; Haucke, V.; Noé, F.; Daumke, O. Structural insights into dynamin-mediated membrane fission. *Structure* **2012**, *20* (10), 1621.

(115) Fröhlich, C.; Grabiger, S.; Schwefel, D.; Faelber, K.; Rosenbaum, E.; Mears, J.; Rocks, O.; Daumke, O. Structural insights into oligomerization and mitochondrial remodelling of dynamin 1-like protein. *EMBO J.* **2013**, 32 (9), 1280.

(116) Archer, S. L. Mitochondrial dynamics—mitochondrial fission and fusion in human diseases. *N. Engl. J. Med.* **2013**, *369* (23), 2236. (117) Youle, R. J.; Karbowski, M. Mitochondrial fission in apoptosis. *Nat. Rev. Mol. Cell Biol.* **2005**, *6* (8), 657.

(118) Breckenridge, D. G.; Stojanovic, M.; Marcellus, R. C.; Shore, G. C. Caspase cleavage product of BAP31 induces mitochondrial fission through endoplasmic reticulum calcium signals, enhancing cytochrome c release to the cytosol. *J. Cell Biol.* **2003**, *160* (7), 1115. (119) Germain, M.; Mathai, J. P.; McBride, H. M.; Shore, G. C. Endoplasmic reticulum BIK initiates DRP1-regulated remodelling of mitochondrial cristae during apoptosis. *EMBO J.* **2005**, *24* (8), 1546. (120) Martinou, J.-C.; Youle, R. Which came first, the cytochrome c release or the mitochondrial fission? *Cell Death Differ.* **2006**, *13* (13), 1291–1295.

(121) Chen, H.; Detmer, S. A.; Ewald, A. J.; Griffin, E. E.; Fraser, S. E.; Chan, D. C. Mitofusins Mfn1 and Mfn2 coordinately regulate

mitochondrial fusion and are essential for embryonic development. J. Cell Biol. 2003, 160 (2), 189.

(122) Zamponi, G. W. Targeting voltage-gated calcium channels in neurological and psychiatric diseases. *Nat. Rev. Drug Discovery* **2016**, 15 (1), 19.

(123) Burté, F.; Carelli, V.; Chinnery, P. F.; Yu-Wai-Man, P. Disturbed mitochondrial dynamics and neurodegenerative disorders. *Nat. Rev. Neurol.* 2015, 11 (1), 11.

(124) Kaur, H.; Sarmah, D.; Saraf, J.; Vats, K.; Kalia, K.; Borah, A.; Yavagal, D. R.; Dave, K. R.; Ghosh, Z.; Bhattacharya, P. Noncoding RNAs in ischemic stroke: time to translate. *Ann. N. Y. Acad. Sci.* **2018**, *1421* (1), 19–36.

(125) Feigin, V. L.; Forouzanfar, M. H.; Krishnamurthi, R.; Mensah, G. A.; Connor, M.; Bennett, D. A.; Moran, A. E.; Sacco, R. L.; Anderson, L.; Truelsen, T. Global and regional burden of stroke during 1990–2010: findings from the Global Burden of Disease Study 2010. *Lancet* **2014**, *383* (9913), 245.

(126) Sarmah, D.; Agrawal, V.; Rane, P.; Bhute, S.; Watanabe, M.; Kalia, K.; Ghosh, Z.; Dave, K. R.; Yavagal, D. R.; Bhattacharya, P. Mesenchymal Stem Cell therapy in Ischemic stroke: A meta-analysis of preclinical studies. *Clin. Pharmacol. Ther.* **2018**, *103* (6), 990–998.

(127) Chamorro, Á.; Dirnagl, U.; Urra, X.; Planas, A. M. Neuroprotection in acute stroke: targeting excitotoxicity, oxidative and nitrosative stress, and inflammation. *Lancet Neurol.* **2016**, *15* (8), 869.

(128) Bhattacharya, P.; Pandey, A. K.; Paul, S.; Patnaik, R. Melatonin renders neuroprotection by protein kinase C mediated aquaporin-4 inhibition in animal model of focal cerebral ischemia. *Life Sci.* **2014**, *100* (2), 97.

(129) Sarmah, D.; Kaur, H.; Saraf, J.; Pravalika, K.; Goswami, A.; Kalia, K.; Borah, A.; Wang, X.; Dave, K. R.; Yavagal, D. R. Getting Closer to an Effective Intervention of Ischemic Stroke: The Big Promise of Stem Cell. *Transl. Stroke Res.* **2017**, DOI: 10.1007/s12975-017-0580-0.

(130) Bhosale, G.; Sharpe, J. A.; Sundier, S. Y.; Duchen, M. R. Calcium signaling as a mediator of cell energy demand and a trigger to cell death. *Ann. N. Y. Acad. Sci.* **2015**, *1350* (1), 107.

(131) Manzanero, S.; Santro, T.; Arumugam, T. V. Neuronal oxidative stress in acute ischemic stroke: sources and contribution to cell injury. *Neurochem. Int.* **2013**, *62* (5), 712.

(132) Niatsetskaya, Z. V.; Sosunov, S. A.; Matsiukevich, D.; Utkina-Sosunova, I. V.; Ratner, V. I.; Starkov, A. A.; Ten, V. S. The oxygen free radicals originating from mitochondrial complex I contribute to oxidative brain injury following hypoxia-ischemia in neonatal mice. *J. Neurosci.* **2012**, *32* (9), 3235.

(133) Qiu, J.; Tan, Y.-W.; Hagenston, A. M.; Martel, M.-A.; Kneisel, N.; Skehel, P. A.; Wyllie, D. J.; Bading, H.; Hardingham, G. E. Mitochondrial calcium uniporter Mcu controls excitotoxicity and is transcriptionally repressed by neuroprotective nuclear calcium signals. *Nat. Commun.* **2013**, *4*, 2034.

(134) Kalogeris, T.; Baines, C. P.; Krenz, M.; Korthuis, R. J. Cell Biology of Ischemia/Reperfusion Injury. *Int. Rev. Cell Mol. Biol.* 2012, 298, 229–317.

(135) dos Santos Picanc, L.; Ozela, P.; Pinheiro, A.; Padilha, E.; Braga, F.; Dos, C. S.; Rosa, J. Alzheimer's disease: A review from the pathophysiology to diagnosis, new perspectives for pharmacological treatment. *Curr. Med. Chem.* **2016**, *23*, 1.

(136) Alzheimer's Association. *Alzheimer's Dementia* **2017**, *13* (4), 325..

(137) Ittner, L. M.; Götz, J. Amyloid- β and tau—a toxic pas de deux in Alzheimer's disease. *Nat. Rev. Neurosci.* **2011**, *12* (2), 67.

(138) Bloom, G. S. Amyloid- β and tau: the trigger and bullet in Alzheimer disease pathogenesis. *JAMA neurology* **2014**, 71 (4), 505.

(139) Abdul, H. M.; Sama, M. A.; Furman, J. L.; Mathis, D. M.; Beckett, T. L.; Weidner, A. M.; Patel, E. S.; Baig, I.; Murphy, M. P.; LeVine, H. Cognitive decline in Alzheimer's disease is associated with selective changes in calcineurin/NFAT signaling. *J. Neurosci.* **2009**, 29 (41), 12957. (140) Benilova, I.; Karran, E.; De Strooper, B. The toxic $A\beta$ oligomer and Alzheimer's disease: an emperor in need of clothes. *Nat.*

Neurosci. 2012, 15 (3), 349. (141) Popugaeva, E.; Pchitskaya, E.; Bezprozvanny, I. Dysregulation of neuronal calcium homeostasis in Alzheimer's disease-A therapeutic opportunity? *Biochem. Biophys. Res. Commun.* 2017, 483 (4), 998.

(142) Cacabelos, R. Parkinson's disease: from pathogenesis to pharmacogenomics. *Int. J. Mol. Sci.* **2017**, *18* (3), 551.

(143) Irwin, D. J.; Lee, V. M.-Y.; Trojanowski, J. Q. Parkinson's disease dementia: convergence of α -synuclein, tau and amyloid- β pathologies. *Nat. Rev. Neurosci.* **2013**, 14 (9), 626.

(144) Martin, Z. S.; Neugebauer, V.; Dineley, K. T.; Kayed, R.; Zhang, W.; Reese, L. C.; Taglialatela, G. α -Synuclein oligomers oppose long-term potentiation and impair memory through a calcineurin-dependent mechanism: relevance to human synucleopathic diseases. J. Neurochem. **2012**, 120 (3), 440.

(145) Caraveo, G.; Auluck, P. K.; Whitesell, L.; Chung, C. Y.; Baru, V.; Mosharov, E. V.; Yan, X.; Ben-Johny, M.; Soste, M.; Picotti, P. Calcineurin determines toxic versus beneficial responses to α -synuclein. *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111* (34), E3544.

(146) Pringsheim, T.; Wiltshire, K.; Day, L.; Dykeman, J.; Steeves, T.; Jette, N. The incidence and prevalence of Huntington's disease: A systematic review and meta-analysis. *Mov. Disord.* **2012**, *27* (9), 1083.

(147) Bates, G. P.; Dorsey, R.; Gusella, J. F.; Hayden, M. R.; Kay, C.; Leavitt, B. R.; Nance, M.; Ross, C. A.; Scahill, R. I.; Wetzel, R. Huntington disease. *Nature reviews Disease primers* **2015**, *1*, 15005.

(148) Xifró, X.; García-Martínez, J. M.; Del Toro, D.; Alberch, J.; Pérez-Navarro, E. Calcineurin is involved in the early activation of NMDA-mediated cell death in mutant huntingtin knock-in striatal cells. J. Neurochem. 2008, 105 (5), 1596.

(149) Kratter, I. H.; Zahed, H.; Lau, A.; Tsvetkov, A. S.; Daub, A. C.; Weiberth, K. F.; Gu, X.; Saudou, F.; Humbert, S.; Yang, X. W. Serine 421 regulates mutant huntingtin toxicity and clearance in mice. *J. Clin. Invest.* **2016**, *126* (9), 3585.

(150) Pineda, J. R.; Pardo, R.; Zala, D.; Yu, H.; Humbert, S.; Saudou, F. Genetic and pharmacological inhibition of calcineurin corrects the BDNF transport defect in Huntington's disease. *Mol. Brain* **2009**, *2* (1), 33.

(151) Gratuze, M.; Noël, A.; Julien, C.; Cisbani, G.; Milot-Rousseau, P.; Morin, F.; Dickler, M.; Goupil, C.; Bezeau, F.; Poitras, I. Tau hyperphosphorylation and deregulation of calcineurin in mouse models of Huntington's disease. *Hum. Mol. Genet.* **2015**, *24* (1), 86. (152) Arthur, K. C.; Calvo, A.; Price, T. R.; Geiger, J. T.; Chio, A.; Traynor, B. J. Projected increase in amyotrophic lateral sclerosis from 2015 to 2040. *Nat. Commun.* **2016**, *7*, 12408.

(153) Renton, A. E.; Majounie, E.; Waite, A.; Simón-Sánchez, J.; Rollinson, S.; Gibbs, J. R.; Schymick, J. C.; Laaksovirta, H.; Van Swieten, J. C.; Myllykangas, L. A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* 2011, 72 (2), 257.

(154) Robberecht, W.; Philips, T. The changing scene of amyotrophic lateral sclerosis. *Nat. Rev. Neurosci.* 2013, 14 (4), 248.

(155) Turner, M. R.; Hardiman, O.; Benatar, M.; Brooks, B. R.; Chio, A.; De Carvalho, M.; Ince, P. G.; Lin, C.; Miller, R. G.; Mitsumoto, H. Controversies and priorities in amyotrophic lateral sclerosis. *Lancet Neurol.* **2013**, *12* (3), 310.

(156) Liachko, N. F.; Saxton, A. D.; McMillan, P. J.; Strovas, T. J.; Currey, H. N.; Taylor, L. M.; Wheeler, J. M.; Oblak, A. L.; Ghetti, B.; Montine, T. J. The phosphatase calcineurin regulates pathological TDP-43 phosphorylation. *Acta Neuropathol.* **2016**, *132* (4), 545.

(157) Kim, J. M.; Billington, E.; Reyes, A.; Notarianni, T.; Sage, J.; Agbas, E.; Taylor, M.; Monast, I.; Stanford, J. A.; Agbas, A. Impaired Cu–Zn Superoxide Dismutase (SOD1) and Calcineurin (Cn) Interaction in ALS: A Presumed Consequence for TDP-43 and Zinc Aggregation in Tg SOD1 G93A Rodent Spinal Cord Tissue. *Neurochem. Res.* **2018**. DOI: 10.1007/s11064-017-2461-z.

(158) Macrez, R.; Stys, P. K.; Vivien, D.; Lipton, S. A.; Docagne, F. Mechanisms of glutamate toxicity in multiple sclerosis: biomarker and therapeutic opportunities. *Lancet Neurol.* **2016**, *15* (10), 1089.

(159) Evonuk, K. S.; Baker, B. J.; Doyle, R. E.; Moseley, C. E.; Sestero, C. M.; Johnston, B. P.; De Sarno, P.; Tang, A.; Gembitsky, I.; Hewett, S. J. Inhibition of system xc- transporter attenuates autoimmune inflammatory demyelination. J. Immunol. 2015, 195 (2), 450.

(160) Jung, S.; Yang, H.; Kim, B. S.; Chu, K.; Lee, S. K.; Jeon, D. The immunosuppressant cyclosporin A inhibits recurrent seizures in an experimental model of temporal lobe epilepsy. *Neurosci. Lett.* **2012**, 529 (2), 133.

(161) Furman, J. L.; Sompol, P.; Kraner, S. D.; Pleiss, M. M.; Putman, E. J.; Dunkerson, J.; Abdul, H. M.; Roberts, K. N.; Scheff, S. W.; Norris, C. M. Blockade of astrocytic calcineurin/NFAT signaling helps to normalize hippocampal synaptic function and plasticity in a rat model of traumatic brain injury. *J. Neurosci.* **2016**, *36* (5), 1502.

(162) Gross, C.; Yao, X.; Engel, T.; Tiwari, D.; Xing, L.; Rowley, S.; Danielson, S. W.; Thomas, K. T.; Jimenez-Mateos, E. M.; Schroeder, L. M. MicroRNA-mediated downregulation of the potassium channel Kv4. 2 contributes to seizure onset. *Cell Rep.* **2016**, *17* (1), 37.

(163) Azzi, J. R.; Sayegh, M. H.; Mallat, S. G. Calcineurin inhibitors: 40 years later, can't live without.... J. Immunol. 2013, 191 (12), 5785.

(164) Ishikura, K.; Ikeda, M.; Hamasaki, Y.; Hataya, H.; Shishido, S.; Asanuma, H.; Nishimura, G.; Hiramoto, R.; Honda, M. Posterior reversible encephalopathy syndrome in children: its high prevalence and more extensive imaging findings. *Am. J. Kidney Dis.* **2006**, *48* (2), 231.

(165) Pflugrad, H.; Schrader, A. K.; Tryc, A. B.; Ding, X.; Lanfermann, H.; Jäckel, E.; Schrem, H.; Beneke, J.; Barg-Hock, H.; Klempnauer, J. Longterm calcineurin inhibitor therapy and brain function in patients after liver transplantation. *Liver Transplantation* **2018**, 24 (1), 56.

(166) Bindu, P. S.; Sinha, S.; Taly, A. B.; Christopher, R.; Kovoor, J. M. Cranial MRI in acute hyperammonemic encephalopathy. *Pediatric neurology* **2009**, *41* (2), 139.

(167) Pruitt, A. A.; Graus, F.; Rosenfeld, M. R. Neurological complications of solid organ transplantation. *Neurohospitalist* **2013**, *3* (3), 152.

(168) Kumar, D.; LeCorchick, S.; Gupta, G. Belatacept as an alternative to calcineurin inhibitors in patients with solid organ transplants. *Front. Med.* **2017**, *4*, 60.

(169) Teperman, L.; Moonka, D.; Sebastian, A.; Sher, L.; Marotta, P.; Marsh, C.; Koneru, B.; Goss, J.; Preston, D.; Roberts, J. P. Calcineurin Inhibitor-Free Mycophenolate Mofetil/Sirolimus Maintenance in Liver Transplantation: The Randomized Spare-the-Nephron Trial. *Liver transplantation* **2013**, *19* (7), 675.

(170) Webber, A. B.; Vincenti, F. An Update on Calcineurin Inhibitor–Free Regimens: The Need Persists, but the Landscape has Changed. *Transplantation* **2016**, *100* (4), 836.

(171) Goede, L.; Pflugrad, H.; Schmitz, B.; Tryc, A. B.; Barg-Hock, H.; Klempnauer, J.; Weissenborn, K.; Lanfermann, H.; Ding, X.-Q. Neurotoxic Side Effects of Calcineurin Inhibitors in Patients After Liver Transplantation: Preliminary Results of a Quantitative MRI Study of the Brain. J. Clin. Exp. Hepatol. 2017, 7, S31.