CROSSTALK

CrossTalk opposing view: the hypoxic ventilatory response does not include a central, excitatory hypoxia sensing component

Luc J. Teppema

Department of Anaesthesiology, Leiden University Medical Centre, Leiden, The Netherlands

Email: luc.teppema@kpnmail.nl

Edited by: Francisco Sepúlveda & Frank Powell

Combining state-of-the-art (opto)genetic, molecular, viral transfection, imaging and staining techniques, Gourine and coworkers have constructed a framework with astrocytes as brain interoceptors particularly focusing on a putative role as central respiratory $CO₂$ and $O₂$ sensors (Gourine *et al.* 2005; Gourine & Kasparov, 2011; Angelova *et al.* 2015; Gourine & Funk, 2017; Rajani *et al.* 2018). In the opinion of this author, this work does not provide conclusive evidence for their hypothesis of an involvement of astrocytes as central $O₂$ sensors in the ventilatory response to hypoxia (HVR) especially in awake animals and humans.

Several animal species show immediate hypoventilation, hypercapnia and substantial reduction or entire absence of their HVR after bilateral carotid body denervation (CBD). Partial or complete restoration of the HVR is observed in rat, cat, piglet, goat and pony, all with different rates. This results from up-regulation of aortic bodies, release from cortical inhibition, recruitment of accessory glomus tissue in the trunk, neuroplastic changes consisting of axon regeneration, building alternative circuitries and recruiting central (including astrocytic) O_2 sensors that may be silent in carotid body-intact conditions (Tenny & Ou, 1977; Martin-Body et al. 1986; Olson *et al.* 1988; Roux *et al.* 2000; Teppema & Dahan, 2010; Mouradian *et al.* 2012). Surprisingly, Angelova *et al.* (2015) did not challenge CBD animals with acute hypoxia or NaCN infusion shortly (a few days) after surgery; after 10 weeks, hypoxic responses in these animals may thus be due to neuroplastic adaptations. In the conscious goat, isolated central hypoxia does not lead to a progressive increase in ventilation as occurs during chronic carotid body (CB) hypoxia (Weizhen *et al.* 1992). The central hypoxia-induced rise in ventilation in the awake dog depends on peripheral chemoreceptor integrity (Curran *et al.* 2000).

Up to two decades after CB removal or carotid endarterectomy, humans show no HVR (Wade *et al.* 1970; Swanson *et al.* 1978; Honda *et al.* 1979; Dahan *et al.* 2007). Exceptionally, on a hypercapnic background, a small response develops that can be ascribed to an involvement of the aortic bodies (Swanson *et al.* 1978; Timmers *et al.* 2003) but not to central mechanisms (Honda *et al.* 1979). Gourine and Funk (2017) propose an important stimulatory role of mitochondrial ROS in the HVR. To support their hypothesis involving a role of electron flow in the mitochondria in the HVR, they refer to patients with a mutation in the gene encoding succinate dehydrogenase (*SDHD*). These patients, however, do indeed show increased mitochondrial ROS (Cerecer-Gil*et al.* 2010) but *do not* have an abnormally large HVR but rather one at the lower end of normal (Dahan *et al.* 2007). Carotid body type I cells from SDHD^{+/−} mice show an unaltered response to hypoxia, and mitochondrial complex II is not involved in oxygen sensing in these mice (note that total *SDHD* gene knockout is lethal in these mice; Piruat*et al.* 2004).

Utilizing barometric plethysmography, Gourine and coworkers tested their

hypothesis *in vivo*. Even if air humidity and temperature in the plethysmograph are controlled, body temperature measured and the inspired $CO₂$ concentration maintained constant, the pressure signal may be influenced by frequency and airway resistance (Enhorning *et al.* 1998). Percentages of oxygen in the inspired air as the independent variable (i.e. the chamber O_2 %) is not the same as the inspired P_{O_2} (Fig. 5 in Angelova *et al.* 2015), and has little predictive value as to the actual stimulus, in this case, the P_{aO_2} . Using arbitrary or relative units is not meaningful and can even be misleading. In other words, what would be of interest is the following: (1) to show blood gases or at least oxygen saturation (e.g. by using a tail probe); (2) to employ a useful index of the HVR such as \dot{V}_{A}/\dot{V}_{CO} , or \dot{V}_{A}/\dot{V}_{O} , (normalized to body weight; see also Olson *et al.* 1988; Morgan *et al.* 2014); and (3) precise control of the P_{aCO_2} Useful quantitative comparisons between groups require exposure to equal stimulus levels that reach their final values at equal rates. The hypoxic challenges in Angelova *et al.* (2015) were poikilocapnic, which is a confounding factor, given the CO2 sensitivity of astrocytes (Gourine *et al.* 2005) and the known O_2 – CO_2 interaction in the rat (Wilson & Teppema, 2016). Finally, Angelova *et al.* (2015) claim lower respiratory activity in PINK1-deficient mice, but failure of astrocytes to sense low oxygen in these mice does not lead to a reduced response to hypoxia (Fig. 7E in Angelova *et al.* 2015). In conclusion, the data of Gourine and coworkers convincingly show O_2 sensitivity of brain astrocytes and provide details of the resulting stimulus-tranduction cascade, involving ROS, the spread of Ca^{2+} waves and a role of ATP. Concerning the role of Ca^{2+} , could gap-junctional Ca^{2+} exchange between glial cells and retrotrapezoid (RTN) neurons excite the latter? And what is

Luc J. Teppema received his MSci (1978) and PhD (1984) degrees from the Catholic University of Nijmegen, The Netherlands (currently Radboud University). As a staff member at the Department of Anaesthesiology, Leiden University Medical Centre, his research focuses on the effects of hypercapnia and hypoxia on the control of breathing and the effects of anaesthetics, analgesics and carbonic anhydrase inhibitors.

the effect of glial depolarization (previously qualified as 'glial impairment' by Holleran *et al.* 2000) and ATP on the release of H^+ by glia? Whether the reported increase in respiratory activity in CBD animals (10 weeks after surgery) is due to $O₂$ sensing by astrocytes remains to be determined, as is the case with an astrocytic role in the normal poikilocapnic (and consequently hypocapnic) and isocapnic HVR.

The human *poikilocapnic* response to mild acute hypoxemia (saturation ~80%) is biphasic, consisting of an initial increase in ventilation, followed by a secondary fall and a rise in cerebral blood flow (CBF; Steinback & Poulin, 2007, 2016). The modest rise in CB activity will induce little central depression other than that caused by a local fall in P_{CO_2} and may account entirely for the small sustained rise in ventilation. Consequently, astrocytes are unlikely to be involved in maintaining an appropriate minute ventilation. If a modest decrease in brainstem P_{O_2} (which is smaller than in the arterial blood) would result in astrocytic release of ATP, then, apart from directly impacting RTN neurons, the lower P_{CO_2} would tend to reduce it. If, as claimed by Gourine *et al.* (2005) and Rajani *et al.* (2018), a prolonged astrocytic ATP release in the anaesthetized rat maintains phrenic activity in the depressing phase, why then is a similar release after CBD not able to augment respiratory activity (Gourine *et al.* 2005)? The data from the poikilocapnic studies in Angelova *et al.* (2015) were collected in the last 5 min of a 10 min lasting exposure, but the time courses of both stimuli and responses are not shown, so it is unclear to what extent these data include those from the depressing phase.

In the laboratory setting the HVR is often measured *isocapnically* to quantitatively estimate hypoxic sensitivity and $O₂$ –CO₂ interaction effects. The isocapnic HVR is also biphasic, and the secondary fall (HVD) is related to a rise in cerebral blood flow combined with central depression (Teppema & Dahan, 2010). During HVD ventilation reaches a level \sim 30% above control, maintained by the carotid bodies that operate at lower gain but do not show a biphasic response (Teppema & Dahan, 2010). In anaesthesia, however, HVD is uncoupled from carotid body activity (see Teppema & Dahan, 2010) and may involve a different role of ATP, as suggested in the rat (Rajani *et al.* 2018).

If the $O₂$ sensitivity of astrocytes is of less relevance to the awake ventilatory response to mild hypoxia, what then is its physiological significance? In the event of local brain hypoxia or ischaemia, an increase in ventilation would be counterproductive: the resulting fall in P_{aCO} would cause vasoconstriction and thus impede blood flow to the affected areas and other brain regions. A crucial role of astrocytes is sensing (and adapting) neural activity and metabolism, and translating this also into an adaptation of local vasomotor activity to defend the supply of oxygen and nutrients (Gordon *et al.* 2016; Mukandala *et al.* 2016). That O_2 sensitivity is a general property of astrocytes, rather than a distinctive feature of those located in the ventral medullary surface may suggest that although the latter are located in close association with the RTN, their primary role is to guard the supply of oxygen and nutrients by other means than by stimulating ventilation.

Call for comments

Readers are invited to give their views on this and the accompanying CrossTalk articles in this issue by submitting a brief (250 word) comment. Comments may be submitted up to 6 weeks after publication of the article, at which point the discussion will close and the CrossTalk authors will be invited to submit a 'Last Word'. Please email your comment, including a title and a declaration of interest, to jphysiol@physoc.org. Comments will be moderated and accepted comments will be published online only as 'supporting information' to the original debate articles once discussion has closed.

References

- Angelova PR, Kasymov V, Christie I, Sheikbaei S, Turovsky E, Marina N, Korsak A, Zwicker J, Teschemacher AG, Ackland GL, Funk GD, Kasparov S, Abramov AI & Gourine A (2015). Functional oxygen sensitivity of astrocytes. *J Neurosci* **35**, 10640–10473.
- Cerecer-Gil NY, Figuera LE, Llamas FJ, Lara M, Escamilla JG, Ramos R, Estrada G, Hussain AK, Gaal J, Korpershoek E, de Krijger RR, Dinjens WN, Devilee P & Bayley JP (2010). Mutation of SDHB is a cause of hypoxia-related high-altitude paraganglioma. *Clin Cancer Res* **16**, 4148–4154.
- Curran AK, Rodman JR, Eastwood PR, Henderson KS, Dempsey JA & Smith CA (2000). Ventilatory responses to specific CNS hypoxia in sleeping dogs. *J Appl Physiol* **88**, 1840–1852.
- Dahan A, Nieuwenhuijs D & Teppema L (2007). Plasticity of central chemoreceptors: effect of bilateral carotid body resection on central CO2 sensitivity. *PLoS Medicine* **4**, e239.
- Enhorning G, van Schaik S, Lundgren C & Vargas I (1998). Whole-body plethysmography, does it measure tidal volume of small animals? *Can J Physiol Pharmacol* **76**, 945–951.
- Gordon GRJ, Howarth C & MacVicar BA (2016). Bidirectional control of blood flow by astrocytes: a role for tissue oxygen and other metabolic factors. *Adv Exp Med Biol* **903**, 209–219
- Gourine AV & Funk GD (2017). On the existence of a central respiratory oxygen sensor. *J Appl Physiol* **123**, 1344–1349. Gourine AV & Kasparov S (2011). Astrocytes as
- brain interoceptors. *Exp Physiol* **96**, 411–416. Gourine AV, Llaudet E, Dale N & Spyer KM
- (2005). Release of ATP in the ventral medulla during hypoxia in rats: role in hypoxic ventilatory response. *J Neurosci* **25**, 1211–1218.
- Holleran J, Babbie M & Erlichman JS (2000). Ventilatory effects of impaired glial function in a brain stem chemoreceptor region in the conscious rat. *J Appl Physiol* **90**, 1539– 1547.
- Honda Y, Watanabe S, Hashizume I, Satomura Y, Hata N, Sakakibara Y & Severinghaus JW (1979). Hypoxic chemosensitivity in asthmatic patients two decades after carotid body resection. *J Appl Physiol* **46**, 632–638.
- Martin-Body RL, Robson GJ & Sinclair JD (1986). Restoration of hypoxic respiratory responses in the awake rat after carotid body sinus denervation by sinus nerve section *J Physiol* **380**, 61–73.
- Morgan BJ, Adrian R, Bates ML, Dopp JM & Dempsey JA (2014). Quantifying hypoxia-induced chemoreceptor sensitivity in the awake rodent *J Appl Physiol* **117**, 816–824.
- Mouradian GC, Forster HV & Hodges MR (2012) Acute and chronic effects of carotid body denervation on ventilation and chemoreflexes in three rat stains. *J Physiol* **590**, 3335–3347.
- Mukandala G, Tynan R, Lanigan S & O'Connor JJ (2016). The effects of hypoxia and inflammation on synaptic signaling in the CNS. *Brain Sci* **6**, 6.
- Olson EB Jr, Vidruk EH & Dempsey JA (1988). Carotid body excision significantly changes ventilator control in awake rats. *J Appl Physiol* **64**, 666–671.
- Piruat JI, Pintado CO, Ortega-Saenz P, Roche M & Lopez-Barneo J (2004). The mitochondrial *SDHD* gene is required for early embryogenesis and its partial deficiency results in persistent carotid body glomus cell activation with full responsiveness to hypoxia. *Mol Cell Biol* **24**, 10933–10940.

Rajani V, Zhang Y, Jalabula V, Rancic V, SheikhBahaei S, Zwicker JD, Pagliardini S, Disckson CT, Ballanyi, K, Kasparov S, Gourine AV & Funk GD (2018). Release of ATP by pre-Bötzinger complex astrocytes contributes to the hypoxic ventilatory response via a Ca^{2+} -dependent P2Y₁ receptor mechanism. *J Physiol* **596**, 3245–3269.

Roux JC, Peyronnet J, Pascual O, Dalmaz Y & Pequignot JM (2000). Ventilatory and central neurochemical reorganisation of O2 chemoreflex after carotid sinus nerve transection in rat. *J Physiol* **522**, 493–501.

Steinback CD & Poulin MJ (2007). Ventilatory responses to isocapnic and poikilocapnic responses in humans. *Respir Physiol Neurobiol* **155**, 104–113.

- Steinback CD & Poulin MJ (2016). Influence of hypoxia on cerebral blood flow regulation in humans. *Adv Exp Biol Med* **903**, 131–142.
- Swanson GD, Whipp BJ, Kaufman RD, Aqleh KA, Winter B & Bellville JW (1978). Effect of hypercapnia on hypoxic ventilatory drive in carotid body-resected man *J Appl Physiol* **45**, 871–877.
- Tenny SM & Ou LC (1977). Ventilatory response of decorticate and decerebrate cats to hypoxia and CO2. *Respir Physiol* **29**, 81–92.
- Teppema LJ & Dahan A (2010). The ventilatory response to hypoxia: in mammals: mechanisms, measurements and analysis. *Physiol Rev* **90**, 675–754.
- Timmers HJ, Karemaker JM, Wieling W, Marres HA, Folgering HT & Lenders JW (2003). Baroreflex and chemoreflex function after bilateral carotid body tumor resection. *J Hypertens* **21**, 591–599.
- Wade JG, Larson CP Jr, Hickey RF, Ehrenfeld WK & Severinghaus JW (1970). Effect of carotid endarterectomy on chemoreceptor and baroreceptor function in man. *N Engl J Med* **282**, 823–29.
- Weizhen N, Engwall MJA, Daristotle L, Pizarro J & Bisgard GE (1992). Ventilatory effects of prolonged (CNS) hypoxia in awake goats. *Respir Physiol* **87**, 37–48.
- Wilson RJA & Teppema LJ (2016). Integration of central and peripheral respiratory chemoreflexes. *Compr Physiol* **6**, 1005–1041.

Additional information

Competing interests

None declared.