

Review

Current and Emerging Therapies in the Management of Hypoxic Ischemic Encephalopathy in Neonates

Jayasree Nair  and Vasantha H.S. Kumar * 

Division of Neonatology, Jacobs School of Medicine and Biomedical Sciences, University at Buffalo, Buffalo, NY 14203, USA; jnair2@buffalo.edu

* Correspondence: vkumar3@buffalo.edu; Tel.: +1-716-3230260

Received: 27 May 2018; Accepted: 16 July 2018; Published: 19 July 2018



Abstract: Neonatal hypoxic ischemic encephalopathy (HIE) presents a significant clinical burden with its high mortality and morbidity rates globally. Therapeutic hypothermia (TH) is now standard of care for infants with moderate to severe HIE, but has not definitively changed outcomes in severe HIE. In this review, we discuss newer promising markers that may help the clinician identify severity of HIE. Therapies that are beneficial and agents that hold promise for neuroprotection are described, both for use either alone or as adjuncts to TH. These include endogenous pathway modifiers such as erythropoietin and analogues, melatonin, and remote ischemic post conditioning. Stem cells have therapeutic potential in this condition, as in many other neonatal conditions. Of the agents listed, only erythropoietin and analogues are currently being evaluated in large randomized controlled trials (RCTs). Exogenous therapies such as argon and xenon, allopurinol, monosialogangliosides, and magnesium sulfate continue to be investigated. The recognition of tertiary mechanisms of brain damage has opened up new research into therapies not only to attenuate brain damage but also to promote cell repair and regeneration in a developmentally disorganized brain long after the perinatal insult. These alternative modalities may be especially important in mild HIE and in areas of the world where there is limited access to expensive hypothermia equipment and services.

Keywords: birth asphyxia; hypoxic ischemic encephalopathy; neonatal encephalopathy

1. Introduction

Significant progress has been made globally in reducing mortality in children under five years of age following the United Nations Millennium Declaration, signed in 2000. Millennium developmental goals (MDG) 4 and 5 specifically focus on reducing child mortality and improving maternal health [1]. Despite the decline in under five mortality by 49% to 46/1000 live births, there is a much slower decline in neonatal mortality, which constitutes about 45% of all childhood deaths. The devised action plan of sustainable development goals aims to reduce neonatal mortality to 12/1000 and under five mortality to 25/1000 live births by 2030 (<https://sustainabledevelopment.un.org/sdg3>).

The most common contributor to early neonatal mortality is birth asphyxia with prematurity, infections, and low birth weight being other major contributors. Four million newborn infants experience birth asphyxia each year, accounting for an estimated one million deaths and 42 million disability-adjusted life years [2]. Many of these infants sustain significant brain injury and develop long-term sequelae, most commonly cerebral palsy, epilepsy, and sensory deficits [2]. Advances in managing infants with birth asphyxia, leading to hypoxic ischemic encephalopathy (HIE) on a global scale will contribute significantly to achieving the 2030 sustainable developmental goals.

With the advent of therapeutic hypothermia (TH), improved outcomes are being reported in moderate HIE. TH, however, has not demonstrated improvement in outcomes related to severe HIE. As hypothermia, both whole body and head cooling, is being evaluated and used across the globe,

several limitations for its use, related to accessibility, provision of adequate facilities for initiation and monitoring hypothermia and financial limitations—especially in developing countries—are being recognized. This has led clinicians and researchers to continue evaluating complementary and/or alternative therapies for infants with HIE. In this review, we will discuss current and emerging therapies in the management of HIE, other than hypothermia.

2. Current Standard of Care

Any pregnancy that is identified as being at high risk for neonatal complications should ideally be delivered at a tertiary care center with trained and experienced resuscitators. Management of an infant who is depressed at birth involves following accepted guidelines such as those published by ILCOR and Neonatal Resuscitation Program [3]. The infant is evaluated for hypothermia, which should ideally commence within 6 h of birth for infants with moderate to severe HIE [4]. Improved motor outcomes have been noted with earlier cooling within 3 h after birth [5]. Supportive management of seizures, fluid balance, and hematological and cardiovascular abnormalities is essential in ensuring optimal outcomes [6]. Presence of a multidisciplinary team including pediatric neurologists, cardiologists, and other subspecialties as well as institutional capabilities for long term EEG, MRI, and physical and occupational therapies are a requisite for establishment of a cooling protocol at tertiary institutes [6]. Follow up with a developmental pediatrician and enrollment in Early interventional programs are also essential to optimize outcomes for infants with HIE [7].

3. Pathophysiology of HIE and Implications for Intervention

The primary pathophysiologic event that results in HIE is impaired oxygen delivery to the brain tissue. This could be caused by reduced oxygen carrying capacity primarily from the hypoxic component or from reduced cerebral blood flow due to volume loss or impaired circulation. Often, a combination of these events is seen. While in some infants the exact causative event cannot be identified, the pathophysiology of brain injury in HIE is well described in term and near term infants.

Primary energy failure occurring from the initial hypoxic ischemic (HI) insult (Figure 1) is characterized by decreased ATP production and systemic acidosis from increased lactate. Energy failure at the cellular level leads to loss of integrity of the neuronal cell membrane, with calcium entry into the cell facilitated by activation of NMDA receptor and other excitotoxic neurotransmitters. There are very few intervention strategies at this stage to limit damage from the primary energy failure that occurs within minutes to hours of the initial insult. However, a period of latency lasting several hours follows primary energy failure, which opens up the window of opportunity for interventions to limit further neuronal damage. In the absence of any intervention, secondary energy failure ensues in moderate to severe HIE. However, the degree of resuscitation following HI injury and the severity of primary energy failure mechanisms may impact not only the latency period, but also the subsequent changes in secondary energy failure in the brain. Secondary energy failure typically occurs 6 to 48 h following HI insult and appears to be related to oxidative stress, inflammation, excitotoxicity, and ultimately to cell death. In a subset of infants, persistent active mechanisms may prevent regeneration of neurons or exacerbate brain damage resulting in tertiary brain injury [8]. Tertiary damage to the neurons—such as myelin deficits, reduced plasticity, and altered cell number—could persist for months to years after the initial insult. Potential mechanisms that oversee these effects include persistent inflammation and epigenetic changes resulting from blockade of oligodendrocyte maturation, impaired neurogenesis, impaired axonal growth, or altered synaptogenesis [9,10].

Timely clinical assessment is essential to facilitate intervention in the latent period. Anticipation from prenatal history and risk factors prepares the team for potential HIE. A detailed neurological exam helps the clinician determine the severity, generally classified as mild, moderate or severe HIE. TH is typically started within 6 h of birth for it to be effective, before secondary energy failure ensues. However, the concept of tertiary brain injury and alternate therapies that could continue to affect prognosis has expanded the treatment window of HIE. Despite advances in management options, it is

difficult to predict HIE in infants, as some of them develop HIE in spite of a normal neurological exam. Recently, specific biomarkers of neuronal injury are being explored for earlier identification of HIE in these infants.

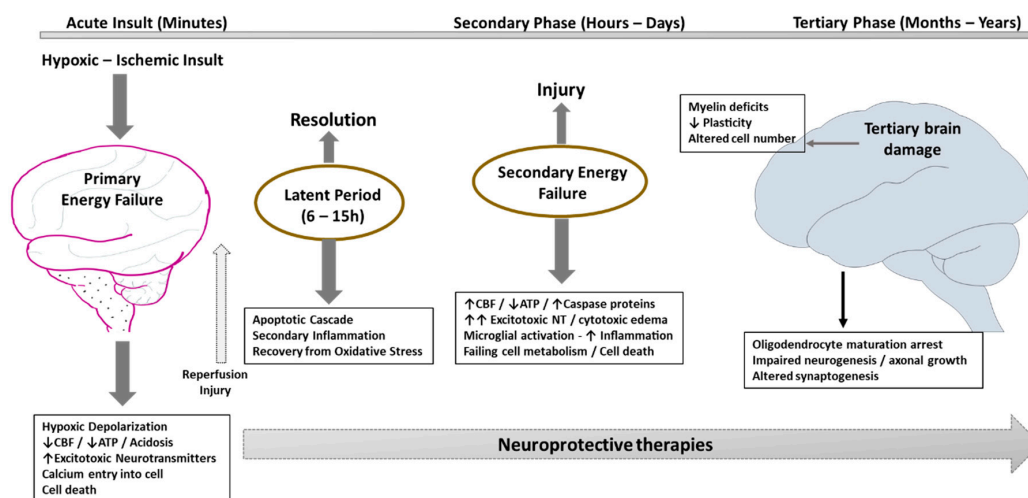


Figure 1. Schematic illustration of pathophysiology of HIE in relation to hypoxic ischemic (HI) insult resulting in primary (acute phase) and secondary energy failure (secondary phase) in the brain. Brain damage (tertiary phase) continues to occur months to years after the injury resulting in decreased plasticity and reduced number of neurons. Latent period following resuscitation is ideal for interventions to decrease the impact of secondary energy failure. However, strategies are developed to attenuate tertiary brain damage which will expand the therapeutic window, substantially increasing the beneficial effects of neuroprotection in these infants and hence its impact on long-term outcomes. CBF—cerebral blood flow; ATP—Adenosine tri phosphate; NT—neurotransmitters.

4. Evaluation of HIE: Emerging Modalities

4.1. Biomarkers

Due to the varied clinical presentation of HIE, it is difficult to predict outcomes early during the course, when intervention with targeted therapies would have maximal effect. Hence, biomarkers that could help detect presence and severity of HIE are being evaluated. These are chemicals that are measurable in blood, urine, cerebrospinal fluid (CSF), or from multiple locations reflective of brain injury or regeneration. Currently, there are no easily available and specific biomarkers for clinical use. In this section, we will describe emerging biomarkers in HIE.

4.1.1. Specific Biomarkers of Brain Injury

S100B is a calcium binding protein released by brain glial cells in response to injury [11]. Elevated levels of S100B have been evaluated in cord blood [12], urine [13], CSF, as well as amniotic fluid for newborns with HIE or encephalopathy. A recent clinical study elevated umbilical cord blood levels of this protein in neonates suffering from HIE stages II-III, suggesting that this biomarker may correlate with the severity of disease and the risk of adverse neurodevelopmental outcomes and/or death [12]. Elevated S100B protein in plasma within 24 h after birth is associated with increased brain injury as evaluated by MRI in a cohort of 50 newborns with HIE [14].

Neuron specific enolase (NSE) is a glycolytic isoenzyme that is released after neuronal death. Celtik et al. identified NSE cut off values that could help differentiate mild moderate and severe encephalopathy [15]. Another recent study correlated NSE levels in the early postnatal period with poor outcomes at two years of age [16]. However, other studies evaluating plasma biomarkers in HIE

have found no correlation between NSE levels and brain injury outcomes [14,17]. Additionally, NSE is affected by any hemolytic process which is a drawback to its utility as an effective biomarker [18].

Glial fibrillary acidic protein (GFAP) is released with damage to astroglial cells and levels in blood and CSF have been seen to correlate with severity of HIE, MRI changes, and developmental outcomes in small studies [19,20]. However, there are no norms for GFAP values in various body fluids and a recent study in newborns with HIE found very low GFAP levels, below the assay lower limits for detection [14].

Several other biomarkers of brain injury including ubiquitin carboxyl-terminal hydrolase L1 (UCH-L1) and total tau protein also hold promise as early biomarkers of brain injury in HIE that are being studied further [14,21].

4.1.2. Inflammatory Markers

Several cytokines are released in acute HIE and their potential as early and accurate biomarkers is being extensively researched. Interleukin (IL)-6 and IL-16 in cord blood were found to be significantly associated with electrographic and clinical HIE severity in a study of full term infants [22]. In another study, plasma levels of IL-6, IL-8, and vascular endothelial growth factor was associated not only with severity of HIE but also predicted abnormal neurological outcomes [20]. However, inflammatory cytokines continue to be limited in their lack of specificity, with other causes of inflammation such as infections and hemolysis also producing a cytokine response.

4.2. Metabolomic Analysis and Metabolites

Metabolomic analyses are being used in various conditions to identify its presence in time for appropriate therapy. Distinct patterns of changes in body metabolites have been evaluated in a recent metabolomic analysis in a non-human primate model of HIE. Eight metabolites (arachidonic acid, butanoic acid, citric acid, fumaric acid, lactate, malate, propanoic acid, and succinic acid) were noted to correlate with adverse neurodevelopmental outcomes [23] in this model. While promising, larger studies evaluating metabolomics are required before this can be used for clinical prognostication.

4.3. Imaging

Magnetic resonance spectroscopy (MRS) has significantly improved our understanding of the changes in brain metabolism and evolution of brain injury after a hypoxic ischemic insult. It could help define the severity of injury if done during the treatment window (within 6 h) by estimating phosphocreatine (PCr) and cerebral lactate or inorganic phosphate. High PCr has been associated with a favorable outcome [24] while high lactate and Pi may indicate a poor outcome [25]. MRS after 6 h continues to provide useful clues to help the clinician prognosticate HIE insult. Low cerebral PCr/Pi, high cerebral lactate, and decreased nucleoside triphosphate are associated with poor outcomes. Persisting brain lactic acidosis on MRS is seen in infants with poor neurodevelopmental outcomes through the first year after birth [26].

5. Emerging Therapies

Most of the neuroprotective strategies being evaluated for use in management of HIE (Table 1, Figure 2), are primarily to mitigate the devastating effects from secondary energy failure on the brain. Therapies that are applied in experimental and animal models of HIE generally work on slowing the pathophysiology that include decreasing oxidative stress, antagonizing excitatory neurotransmitter release or receptor blockade, anti-inflammatory effects, immunomodulation or by decreasing apoptosis among others. While some are undergoing randomized controlled trials such as erythropoietin and its analogues, others are being researched in experimental animal models. Several others have been evaluated anecdotally.

Table 1. Promising therapies in management of HIE by mechanism.

| Mechanism | Clinical Trials | Pre-Clinical Studies |
|------------|--|--|
| Endogenous | Erythropoietin, darbepoietin Stem cells Melatonin | Remote ischemic postconditioning Endocannabinoids |
| Exogenous | Monosialogangliosides Xenon Allopurinol Topiramate Magnesium sulfate | Argon Azithromycin |

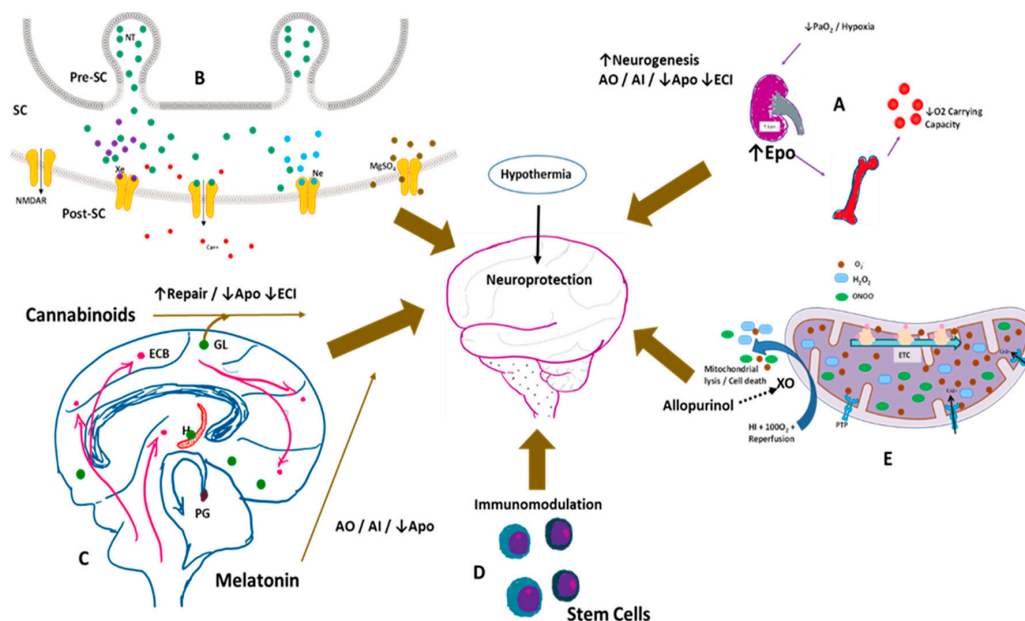


Figure 2. Potential neuroprotective therapies in the management of hypoxic-ischemic encephalopathy. Hypothermia is currently the standard of care in the management of moderate to severe HIE in infants. (A) Hypoxia stimulates erythropoietin (Epo) production by the kidneys, increasing RBC production in the bone marrow, thereby increasing O₂ carryign capacity. Epo favors neurogenesis; is an antioxidant (AO), anti-inflammatory (AI), decreases apoptosis (Apo) and excitotoxic cell injury (ECI); (B) Xenon (Xe—purple dots), neon (Ne—teal dots), and magnesium sulphate (Mgso₄—brown dots) antagonize the N-methyl-D-aspartate (NMDA) mediated excitotoxicity via the NMDA receptor (NMDAR), decreasing calcium entry into the cell. SC—synaptic cleft, Pre-SC—presynaptic cleft, Post-SC—postsynaptic cleft; (C) Endogenous cannabinoids (ECBs) and melatonin are neuroprotective. Spingolipids such as gangliosides (GL) also protect against apoptotic injury; (D) Umbilical cord blood derived stem cells and mesenchymal stem cells modulate the immune system and affect long-term outcomes; (E) Allopurinol by inhibiting the enzyme xanthine oxidase (XO), decreases reactive oxygen species, mitochondrial lysis, and cell death. O₂—superoxide, H₂O₂—hydrogen peroxide, ONOO—peroxynitrite, ETC—electron transfer chain; PTP—permeability transition pore of inner mitochondrial membrane.

5.1. Resuscitation

5.1.1. Optimizing Placental Transfusion

Either by delayed cord clamping or by the process or cord milking/stripping may impart a neuroprotective effect, besides improving hemodynamics [27]. Full-term infants who underwent DCC had a 45% higher mean ferritin concentration at four months of age [28], demonstrating a beneficial effect in preventing iron deficiency anemia. Iron plays an important role in brain myelination [29], suggesting a neuroprotective effect of placental transfusion. Additionally, umbilical cord derived stem

cells are being evaluated for treatment of HIE, and ensuring an optimal placental transfusion ensures that the compromised infant receives the entire complement of stem cells.

5.1.2. Vasopressin

Use of vasopressin as an alternative to epinephrine in neonatal resuscitation is being evaluated in pre-clinical studies. This medication has a neuroprotective potential as studies in rat and guinea pig have demonstrated that vasopressin activates hippocampal interneurons, silencing synchronous neuronal activity [30]. This may reduce neuronal energy demand, which could be a neuroprotective mechanism.

5.2. Erythropoietin/Analogues (Endogenous)

Erythropoietin (Epo) is an endogenous protein, synthesized in the fetal liver that has an impact on multiple critical pathways. Besides stimulating erythropoiesis, Epo is a cytokine that influences the body's immune response [31]. Additionally, its neuroprotective role has been recognized and evaluated in pre-clinical and clinical studies. Epo receptors (EpoR) are widely expressed throughout the central nervous system in several cell types including progenitor cells, astrocytes, oligodendrocytes, and microglia [32,33], to name a few. Epo and EpoR are upregulated following hypoxic ischemic injury and Epo has an anti-oxidant [34] as well as anti-inflammatory [35] effect. It reduced apoptotic [36,37] and excitotoxic cell injury [38]. Clinical trials evaluating Epo alone [39,40] in infants with HIE have shown promising results. Since hypothermia has become standard of care therapy for HIE, larger trials are currently on going evaluating Epo as a complement to cooling therapy. A Phase I trial evaluating effective dose and safety [41] demonstrated that a moderately high dose of 1000 U/kg achieved levels (based on animal studies) that would protective maximal neuroprotection and minimize risks of excessive Epo. In a Phase II double-blinded, placebo-controlled trial in infants undergoing TH for HIE, multiple doses of Epo (1000 U/kg) resulted in less MRI brain injury and potential for improved short-term motor outcomes [42]. A Phase III trial evaluating the effect of Epo with TH on the combined outcome of death or neurodevelopmental disability is currently underway [43].

Darbepoietin (darbe) is a long acting erythropoietin analogue that offers the additional benefit of once weekly administration. A small randomized trial evaluating the use of darbe as adjunctive therapy to hypothermia in the first 12 h of life and repeated at one week of life demonstrated a good safety profile of this medication [44]. A large multicenter randomized controlled trial evaluating use of darbe in mild HIE is currently underway (MEND study ClinicalTrials.gov Identifier: NCT03071861).

5.3. Stem Cells

Recent experimental studies in animal models have indicated that various mechanisms of action are involved in the process by which umbilical cord blood cells (UCBCs) protect the brain from hypoxic ischemic injury. These stem cells are predominantly derived from two sources—bone marrow derived mesenchymal stem cells (BM-MSC) and umbilical cord blood derived mesenchymal stem cells (UCB-MSC) [45]. Preclinical studies have been done evaluating stem cells in neonatal brain injury. Cord blood stem cell therapy has could have protective effects mainly on inflammation, apoptosis, oxidative stress, and may enhance regeneration [46]. Unlike hypothermia, cell based therapy may provide a longer therapeutic window as repair and regeneration take place over longer periods of time. Donor MSCs do not last for a long time in the brain, but do have paracrine effects by secreting trophic and immunomodulatory factors that help in repair from brain injury. A meta-analysis of MSCs demonstrated a significant positive effect on neurobehavioral outcome following HIE injury [47]. Specifically in a preterm model of global hypoxic injury, IV mesenchymal stem cells reduced cerebral inflammation by induction of T-cell tolerance [48]. A Phase I clinical trial evaluating UCB cells to neonates with HIE showed that collection, preparation, and infusion of autologous, volume- and RBC-reduced, non-cryopreserved cord blood cells within the first few postnatal days was feasible and relatively safe in their cohort [49]. Five newborns (four with moderate and one with severe HIE)

have undergone autologous UCBCs therapy without any significant adverse effects of treatment with survival up to one year [46]. While promising in its potential, larger well designed multi center trials are required to prove safety and efficacy prior to using this modality in standard practice.

5.4. Remote Ischemic Postconditioning (Endogenous)

The concept of remote ischemic conditioning (RIPC) involves delivery of sub-lethal small ischemic insults, remote from the area of injury, that activate endogenous repair pathways which potentially help in reducing the extent of original ischemic injury. This has previously been studied in adult cardiac injury [50], however is now being evaluated both in adult and neonatal brain ischemia and stroke [51]. RIPC, as it applies in neonatal HIE, would involve conditioned ischemic insults to the limb soon after the initial hypoxic ischemic insult. The ischemic skeletal muscle releases several endogenous compounds that activate not only the neuronal and humoral pathways, but also the systemic inflammatory response [52]. Delayed RIPC has significantly improved long term sensory motor deficits in a neonatal HI rat model [53]. In a piglet model of neonatal asphyxia, RIPC demonstrated a beneficial effect, mediated by modulation of nitrosative stress, despite glial activation [54]. Due to ease of application facilitating use even in low resource setting, this modality holds great promise both as an adjunct to therapeutic hypothermia, and also as a primary therapy in infants who may only have mild HIE and do not undergo cooling. Safety and efficacy of RIPC needs further evaluation in preclinical studies, especially in translational large animal models of neonatal HIE.

5.5. Endocannabinoids (Endogenous)

The endocannabinoid system has been recognized as an important neuroregulatory mechanism that could help in protection from brain injury. Activation of this system has been shown to decrease glutamate excitotoxicity and activation of microglia and cell death pathways. Use of a cannabinoid (CBD) receptor 1 and 2 (CBR1 and CBR2) agonist WIN 55212-2 in a rodent model of neonatal HIE demonstrated protective effects by prevention of glutamate release, TNF alpha accumulation, and iNOS induction, resulting in decreased cell death [55]. In another rodent model, WIN promoted neural stem cell proliferation in the sub ventricular zone and the short-term generation and migration of neuroblasts to the adjacent injured caudate [56]. Additionally, use of CBR agonists promotes oligodendrocyte generation, survival, and differentiation, thus contributing to repair from white matter injury [56]. In asphyxiated newborn piglets, intravenous administration of CBD improved brain oxygenation and induced a partial EEG recovery after HI [57]. As with other modalities describe above, while use of this therapy is promising, its safety and efficacy in preclinical studies continues to be evaluated.

5.6. Melatonin (Endogenous)

Melatonin is an endogenous neuroendocrine moiety secreted by the pineal gland and well known for its role in modulating the circadian rhythm. Besides this, melatonin has several other mechanisms that suggest an important role in recovery and repair from brain injury. Melatonin plays an important role in normal glial development [58] and has anti-apoptotic [59], anti-inflammatory, and anti-oxidant [60] effects. In a piglet model of neonatal HI injury, melatonin used along with hypothermia greatly decreased the HI-induced injury measured by magnetic resonance spectroscopy [61]. In a randomized controlled pilot trial evaluating melatonin with cooling in term infants with HIE, compared to controls, the HIE groups had increased serum melatonin and nitric oxide and plasma superoxide dismutase. The melatonin/hypothermia group also had fewer seizures, less evidence of white matter injury on MRI, and a lower rate of mortality without developmental or neurological abnormalities [62]. Melatonin has a good safety profile with low risk for toxicity [63,64] and thus holds tremendous promise in management of infants with HIE [65]. However, it has yet to be validated in larger randomized controlled trials.

5.7. Monosialoganglioside

Gangliosides are sphingolipids that serve an important function in maintaining cell membrane integrity. In a rat model of neonatal hypoxic ischemic injury, reduced ganglioside, phospholipid, and cholesterol contents in the hippocampus were noted [66]. Monosialoganglioside therapy has been shown to protect against apoptotic injury [67] and attenuate brain injury [68]. This led to consideration of monosialogangliosides as an adjuvant therapy in HIE. In a meta-analysis of all published clinical studies, Sheng et al. evaluated 10 trials consisting of 787 neonates. They concluded that adjuvant treatment with monosialoganglioside potentially offers additional benefits in terms of improving short-term clinical effects and reducing long-term neurodevelopmental disabilities [69]. However, they did acknowledge methodologic flaws in the included studies as well as limited information on optimal dosing, safety, as well as long term outcomes.

5.8. Xenon

Acute hypoxic ischemic insult leads to NMDA receptor activation through neuronal depolarization. Xenon inhibits NMDA signaling and thus may play a role in reducing the acute cell injury. While studies in the piglet model of birth asphyxia suggest a benefit to the combined modality of treatment with hypothermia and xenon [70,71], a proof of concept clinical trial (TOBY-Xe) [72] in 92 neonates concluded that xenon is unlikely to enhance the neuroprotective effects of cooling after birth asphyxia. Additionally, xenon is a noble gas that is expensive and requires a specialized delivery system [73]. Due to these factors, alternative therapies are currently being evaluated.

5.9. Argon

This is another significantly less expensive noble gas, that has demonstrated significant neuroprotection in animal models of HIE. In an extensive piglet model of ischemic injury, Broad et al. showed augmentation of hypothermic neuroprotection with argon use. They demonstrated reduced brain cell death, MRS, and aEEG improvements with combined argon hypothermia treatment [74]. Additionally, both xenon and argon have good blood brain barrier penetration [75].

5.10. Allopurinol

Oxidant injury by free radicals and superoxides formed through activation of the xanthine oxidase pathway contribute to the damage caused by a hypoxic ischemic insult. Allopurinol is a xanthine oxidase inhibitor that is being investigated as a potential agent for use in treatment of HIE. Preclinical studies in various rodent [76] and mammalian models [77] of HIE have shown neuroprotective effects with use alone and recently, as a complement to TH [78]. Only a few small trials in human neonates have been published so far and a Cochrane review in 2012 did not reveal any statistically significant difference in the risk of death or a composite of death or severe neurodevelopmental disability [79]. However a follow-up of two earlier performed randomized controlled trials at 4–8 years suggested a neuroprotective effect of neonatal allopurinol treatment in the subset of moderately asphyxiated infants [80]. Additional use of allopurinol antenatally in pregnancies at risk of hypoxic ischemic injury is being evaluated. A randomized blinded study of 53 pregnancies with evidence of fetal hypoxia demonstrated lower cord blood levels of S-100B, a short term marker of brain injury [81]; however, long term beneficial outcomes are yet to be proven conclusively.

5.11. Magnesium Sulfate

Magnesium sulfate is an NMDA receptor antagonist believed to reduce excitotoxic damage after a hypoxic ischemic insult. It is now being widely used antenatally for neuroprotection in preterm deliveries. Initial interest in use of this medication was generated due to low magnesium levels being noted in infants with HIE [82]. A prospective, longitudinal, placebo-controlled trial of MgSO₄ use in infants with severe asphyxia, without hypothermia therapy, revealed encouraging short term

outcomes compared to standard supportive treatment [83]. A systematic review of preclinical evidence for MgSO₄ use in HIE noted inconsistent evidence of benefit as well as little consensus in dose and timing of administration [84]. Though generally considered a safe medication, there is some concern regarding risk of hypotension and bradycardia, especially with high doses of this medication which could limit its use [85]. A meta-analysis of five small randomized controlled trials evaluating MgSO₄ in HIE concluded that there was improvement in short-term outcomes without significant increase in side effects [86]. However, there is a need for large well designed studies to determine if there are long-term benefits of magnesium and to confirm its safety, as a trend towards increased mortality was noted in the magnesium group [86]. Due to its low cost and ease of delivery, MgSO₄ is thought to be a potentially viable therapy in low resource settings, in spite of its side effects [87].

5.12. Topiramate

Topiramate blocks the voltage-dependent sodium and calcium channels and also inhibits the excitatory glutamate pathway while enhancing the inhibitory effects of gamma-aminobutyric acid (GABA). All these effects would work favorably in the pathophysiology of HIE. In newborns, it has been extensively studied in the management of HIE in combination with hypothermia. The short-term and safety data support its use in combination with hypothermia in exploring the possible neuroprotective effects [88]. Although administration of topiramate in newborns with HIE is safe, it did not reduce the combined frequency of mortality and severe neurological disability [89]. The role of topiramate co-treatment in preventing subsequent epilepsy deserve further studies.

5.13. Azithromycin

Preclinical studies in models of ischemic stroke have revealed that azithromycin has a neuroprotective effect [90]. Recent abstracts have investigated the possibility of using azithromycin in neonatal HIE alone and as an adjunct to hypothermia [91]. This is a medication that has an established safety profile and is already being used in neonates, hence promising preclinical studies could quickly lead to clinical trials evaluating azithromycin as an adjunct to hypothermia therapy.

5.14. Combination Therapies

With the discovery of more therapeutic targets for management of HIE, there is potential for combination and adjunctive therapies with agents that may affect the pathophysiological process at different phases. Additionally, TH is being explored for mild HIE, as neurological deficits have been noted in these infants as well [92]. Melatonin, epo, darbe, xenon, and topiramate are all being studied as adjuncts to TH (Clinicaltrials.gov NCT02071160, NCT01913340, NCT01471015, NCT00934700, NCT01241019). Stem cells and Epo have actions that extend into the tertiary phase and thus may prove to be complementary to TH [93]. Creative ways of combining pharmacological and cell based therapies in secondary and tertiary phases of injury would be exciting and would especially benefit infants with HIE that evolved over time and missed the window for TH.

6. Management of Hypoxic Ischemic Injury beyond the Neonatal Period

Changes that occur in the brain after perinatal ischemic injury over time include loss of myelin, decreased neurogenesis, and reduced plasticity of the brain leading to tertiary brain damage [8]. These mechanisms contributing to tertiary brain damage may provide us with opportunities for long-term interventions in improving neurological function. Modulating persistent inflammation to promote neurogenesis is being experimented with IL-6 [94]. Imaging for gliosis, a function of chronic inflammation from activation of microglia [8], could be used as a diagnostic aid in the development of long-term treatments. Enhancing myelination by oligodendrocyte (ODC) stimulation could overcome ODC maturational arrest; with its ability to reduce perinatal inflammation and promoting white matter development [95,96].

Trichostatin-A, a histone deacetylase (HDAC) inhibitor has been shown to abolish behavioral and neuroendocrine deficits from early life stresses by promoting demethylation and possibly inducing epigenetic modifications [97]. HDAC inhibitors may restore loss of acetylation associated with inflammation and HI and improve and behavior outcomes in injury models in animals [98]. HDAC inhibitors are shown to modulate innate and humoral immune responses to enhance Th2 and TLR4 activation [99]. Modulation of innate of adaptive immune responses by lymphocyte activation is shown to reduce microglial activation and disease progression in animal models [100]. Administration of microRNAs (miRNA-124) to modulate microglial function and to improve neuronal function long after injury is also being explored as therapeutic targets [101].

Stem cell therapy offers enormous potential for stimulation of endogenous neural stem cells; to redirect new cell production from astroglia to oligodendrocytes and neurons following acute insult. Repeated treatments and delayed treatment in animal models has expanded the therapeutic window for treatment of tertiary brain damage by inducing repair and regeneration [102,103]. Activated microglia and macrophage phenotypes support neurogenesis and progenitor cell proliferation similar to stem cells, and hence are exciting candidates for repair and regeneration, as they migrate to site of injury and generate trophic factors [104]. Growth factors can prevent tertiary brain damage by attenuating inflammatory processes and promoting plasticity of the brain [105]. Ampakines, small and diffusible growth factors that easily diffuse across the blood–brain barrier and induce brain derived neurotrophic factor (BDNF) production in the brain when administered systemically are being explored [106].

Most of the therapies discussed above are studied in animal models with few current clinical studies, particularly in neonates with hypoxic ischemic encephalopathy. However, current research suggesting targets for therapy in infants with HIE in the tertiary phase, months to years after the acute insult offers hope to children with this devastating disease.

7. Limitations of Management in Resource Poor Settings

Therapeutic hypothermia, while standard of care in developed countries, remains an expensive and sometimes unaffordable modality of treatment in resource limited settings. Practical difficulties are often encountered in transferring patients, often in rural settings to tertiary care centers within the 6-h window for therapeutic hypothermia. An additional concern initially identified in a randomized controlled trial of TH by Robertson et al. was the high mortality rate that was noted in the enrolled infants [107]. A systematic review and meta-analysis of trials evaluating TH for neonatal encephalopathy in low-and middle-income countries (LMIC) also showed that TH was not associated with a statistically significant reduction in neonatal mortality [108]. Major contributing factors, besides lack of tertiary care facilities and adequate neonatal transport, were thought to be the high rate of neonatal sepsis, inflammation and growth retardation [109] suggesting a chronic perinatal insult in these infants in LMIC. Rodent models primed with infection prior to a hypoxic ischemic insult have demonstrated reduced protection with hypothermia [110]. Thus, with current limitations in LMIC, alternative low cost cooling methods as well as adjuncts and affordable treatment options such as magnesium are being evaluated [87].

8. Conclusions

Birth asphyxia contributes to a significantly higher burden of neonatal mortality and morbidity globally, more so in developing countries. Neonatal HIE, apart from increased mortality leads to devastating neurological consequences such as cerebral palsy, epilepsy, and mental retardation. With the advent and widespread clinical use of TH over the last decade, the prognosis of moderate HIE has significantly improved. However, TH is expensive, needs a multidisciplinary team and is further limited by narrow therapeutic window for initiation. With issues of access to health care and the burden of birth asphyxia shifting to developing and least developed nations, there is a need for alternative and supplementary neuroprotective agents. Low cost and easy availability along with ease of use would assist in ensuring that these therapies have global applicability. Several of the promising

modalities we have described in this review are still in the preclinical stage. It is uncertain how many of these therapies, alone or in combination, will achieve widespread use in the management of neonatal HIE. Large well designed randomized control trials in developed countries as well as in low resource settings, are required to prove the safety and efficacy of these therapies, prior to widespread clinical use. Neonatal HIE remains a condition of global importance with significant disparities in available treatment options among developed and developing countries. Hence, a concerted effort to address this problem on a global scale is essential.

Author Contributions: J.N. and V.H.S.K. conceived and wrote the paper.

Funding: This research received no external funding.

Acknowledgments: Supported by research funding grant 1 R03 HD086531-01 NIH-NICHD (J.N.) and AAP NRP research grant 2018 (J.N.).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Gaffey, M.F.; Das, J.K.; Bhutta, Z.A. Millennium development goals 4 and 5: Past and future progress. *Semin. Fetal Neonatal. Med.* **2015**, *20*, 285–292. [[CrossRef](#)] [[PubMed](#)]
- Saugstad, O.D. Reducing global neonatal mortality is possible. *Neonatology* **2011**, *99*, 250–257. [[CrossRef](#)] [[PubMed](#)]
- Wyckoff, M.H.; Aziz, K.; Escobedo, M.B.; Kapadia, V.S.; Kattwinkel, J.; Perlman, J.M.; Simon, W.M.; Weiner, G.M.; Zaichkin, J.G. Part 13: Neonatal resuscitation: 2015 american heart association guidelines update for cardiopulmonary resuscitation and emergency cardiovascular care. *Circulation* **2015**, *132* (Suppl. 2), S543–S560. [[CrossRef](#)] [[PubMed](#)]
- Shankaran, S.; Laptook, A.R.; Ehrenkranz, R.A.; Tyson, J.E.; McDonald, S.A.; Donovan, E.F.; Fanaroff, A.A.; Poole, W.K.; Wright, L.L.; Higgins, R.D.; et al. Whole-body hypothermia for neonates with hypoxic-ischemic encephalopathy. *N. Engl. J. Med.* **2005**, *353*, 1574–1584. [[CrossRef](#)] [[PubMed](#)]
- Thoresen, M.; Tooley, J.; Liu, X.; Jary, S.; Fleming, P.; Luyt, K.; Jain, A.; Cairns, P.; Harding, D.; Sabir, H. Time is brain: Starting therapeutic hypothermia within three hours after birth improves motor outcome in asphyxiated newborns. *Neonatology* **2013**, *104*, 228–233. [[CrossRef](#)] [[PubMed](#)]
- Committee on Fetus and Newborn; Papile, L.A.; Baley, J.E.; Benitz, W.; Cummings, J.; Carlo, W.A.; Eichenwald, E.; Kumar, P.; Polin, R.A.; Tan, R.C.; et al. Hypothermia and neonatal encephalopathy. *Pediatrics* **2014**, *133*, 1146–1150. [[PubMed](#)]
- Robertson, C.M.; Perlman, M. Follow-up of the term infant after hypoxic-ischemic encephalopathy. *Paediatr. Child Health* **2006**, *11*, 278–282. [[PubMed](#)]
- Fleiss, B.; Gressens, P. Tertiary mechanisms of brain damage: A new hope for treatment of cerebral palsy? *Lancet Neurol.* **2012**, *11*, 556–566. [[CrossRef](#)]
- Favrais, G.; van de Looij, Y.; Fleiss, B.; Ramanantsoa, N.; Bonnin, P.; Stoltenburg-Didinger, G.; Lacaud, A.; Saliba, E.; Dammann, O.; Gallego, J.; et al. Systemic inflammation disrupts the developmental program of white matter. *Ann. Neurol.* **2011**, *70*, 550–565. [[CrossRef](#)] [[PubMed](#)]
- Leviton, A.; Gressens, P. Neuronal damage accompanies perinatal white-matter damage. *Trends Neurosci.* **2007**, *30*, 473–478. [[CrossRef](#)] [[PubMed](#)]
- Michetti, F.; Gazzolo, D. S100b protein in biological fluids: A tool for perinatal medicine. *Clin. Chem.* **2002**, *48*, 2097–2104. [[PubMed](#)]
- Zaigham, M.; Lundberg, F.; Olofsson, P. Protein s100b in umbilical cord blood as a potential biomarker of hypoxic-ischemic encephalopathy in asphyxiated newborns. *Early Hum. Dev.* **2017**, *112*, 48–53. [[CrossRef](#)] [[PubMed](#)]
- Gazzolo, D.; Marinoni, E.; Di Iorio, R.; Bruschetini, M.; Kornacka, M.; Lituania, M.; Majewska, U.; Serra, G.; Michetti, F. Measurement of urinary s100b protein concentrations for the early identification of brain damage in asphyxiated full-term infants. *Arch. Pediatr. Adolesc. Med.* **2003**, *157*, 1163–1168. [[CrossRef](#)] [[PubMed](#)]
- Massaro, A.N.; Wu, Y.W.; Bammler, T.K.; Comstock, B.; Mathur, A.; McKinstry, R.C.; Chang, T.; Mayock, D.E.; Mulkey, S.B.; Van Meurs, K.; et al. Plasma biomarkers of brain injury in neonatal hypoxic-ischemic encephalopathy. *J. Pediatr.* **2018**, *194*, 67–75. [[CrossRef](#)] [[PubMed](#)]

15. Celtik, C.; Acunas, B.; Oner, N.; Pala, O. Neuron-specific enolase as a marker of the severity and outcome of hypoxic ischemic encephalopathy. *Brain Dev.* **2004**, *26*, 398–402. [[CrossRef](#)] [[PubMed](#)]
16. Kelen, D.; Andorka, C.; Szabo, M.; Alafuzoff, A.; Kaila, K.; Summanen, M. Serum copeptin and neuron specific enolase are markers of neonatal distress and long-term neurodevelopmental outcome. *PLoS ONE* **2017**, *12*, e0184593. [[CrossRef](#)] [[PubMed](#)]
17. Nagdyman, N.; Komen, W.; Ko, H.K.; Muller, C.; Obladen, M. Early biochemical indicators of hypoxic-ischemic encephalopathy after birth asphyxia. *Pediatr. Res.* **2001**, *49*, 502–506. [[CrossRef](#)] [[PubMed](#)]
18. Gazzolo, D.; Abella, R.; Marinoni, E.; Di Iorio, R.; Li Volti, G.; Galvano, F.; Pongiglione, G.; Frigiola, A.; Bertino, E.; Florio, P. Circulating biochemical markers of brain damage in infants complicated by ischemia reperfusion injury. *Cardiovasc. Hematol. Agents Med. Chem.* **2009**, *7*, 108–126. [[CrossRef](#)] [[PubMed](#)]
19. Blennow, M.; Hagberg, H.; Rosengren, L. Glial fibrillary acidic protein in the cerebrospinal fluid: A possible indicator of prognosis in full-term asphyxiated newborn infants? *Pediatr. Res.* **1995**, *37*, 260–264. [[CrossRef](#)] [[PubMed](#)]
20. Chalak, L.F.; Sanchez, P.J.; Adams-Huet, B.; Laptook, A.R.; Heyne, R.J.; Rosenfeld, C.R. Biomarkers for severity of neonatal hypoxic-ischemic encephalopathy and outcomes in newborns receiving hypothermia therapy. *J. Pediatr.* **2014**, *164*, 468–474. [[CrossRef](#)] [[PubMed](#)]
21. Takahashi, K.; Hasegawa, S.; Maeba, S.; Fukunaga, S.; Motoyama, M.; Hamano, H.; Ichiyama, T. Serum tau protein level serves as a predictive factor for neurological prognosis in neonatal asphyxia. *Brain Dev.* **2014**, *36*, 670–675. [[CrossRef](#)] [[PubMed](#)]
22. Walsh, B.H.; Boylan, G.B.; Livingstone, V.; Kenny, L.C.; Dempsey, E.M.; Murray, D.M. Cord blood proteins and multichannel-electroencephalography in hypoxic-ischemic encephalopathy. *Pediatr. Crit. Care Med.* **2013**, *14*, 621–630. [[CrossRef](#)] [[PubMed](#)]
23. Chun, P.T.; McPherson, R.J.; Marney, L.C.; Zangeneh, S.Z.; Parsons, B.A.; Shojaie, A.; Synovec, R.E.; Juul, S.E. Serial plasma metabolites following hypoxic-ischemic encephalopathy in a nonhuman primate model. *Dev. Neurosci.* **2015**, *37*, 161–171. [[CrossRef](#)] [[PubMed](#)]
24. Iwata, O.; Iwata, S.; Bainbridge, A.; De Vita, E.; Matsuishi, T.; Cady, E.B.; Robertson, N.J. Supra- and sub-baseline phosphocreatine recovery in developing brain after transient hypoxia-ischaemia: Relation to baseline energetics, insult severity and outcome. *Brain* **2008**, *131*, 2220–2226. [[CrossRef](#)] [[PubMed](#)]
25. Cady, E.B.; Iwata, O.; Bainbridge, A.; Wyatt, J.S.; Robertson, N.J. Phosphorus magnetic resonance spectroscopy 2 h after perinatal cerebral hypoxia-ischemia prognosticates outcome in the newborn piglet. *J. Neurochem.* **2008**, *107*, 1027–1035. [[CrossRef](#)] [[PubMed](#)]
26. Robertson, N.J.; Cox, I.J.; Cowan, F.M.; Counsell, S.J.; Azzopardi, D.; Edwards, A.D. Cerebral intracellular lactic alkalosis persisting months after neonatal encephalopathy measured by magnetic resonance spectroscopy. *Pediatr. Res.* **1999**, *46*, 287–296. [[CrossRef](#)] [[PubMed](#)]
27. Katheria, A.C.; Lakshminrusimha, S.; Rabe, H.; McAdams, R.; Mercer, J.S. Placental transfusion: A review. *J. Perinatol.* **2017**, *37*, 105–111. [[CrossRef](#)] [[PubMed](#)]
28. Andersson, O.; Hellstrom-Westas, L.; Andersson, D.; Domellof, M. Effect of delayed versus early umbilical cord clamping on neonatal outcomes and iron status at 4 months: A randomised controlled trial. *BMJ* **2011**, *343*, d7157. [[CrossRef](#)] [[PubMed](#)]
29. Georgieff, M.K. Nutrition and the developing brain: Nutrient priorities and measurement. *Am. J. Clin. Nutr.* **2007**, *85*, 614S–620S. [[PubMed](#)]
30. Spoljaric, A.; Seja, P.; Spoljaric, I.; Virtanen, M.A.; Lindfors, J.; Uvarov, P.; Summanen, M.; Crow, A.K.; Hsueh, B.; Puskarjov, M.; et al. Vasopressin excites interneurons to suppress hippocampal network activity across a broad span of brain maturity at birth. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E10819–E10828. [[CrossRef](#)] [[PubMed](#)]
31. Wu, Y.W.; Gonzalez, F.F. Erythropoietin: A novel therapy for hypoxic-ischaemic encephalopathy? *Dev. Med. Child. Neurol.* **2015**, *57* (Suppl. 3), 34–39. [[CrossRef](#)] [[PubMed](#)]
32. Sugawa, M.; Sakurai, Y.; Ishikawa-Ieda, Y.; Suzuki, H.; Asou, H. Effects of erythropoietin on glial cell development; oligodendrocyte maturation and astrocyte proliferation. *Neurosci. Res.* **2002**, *44*, 391–403. [[CrossRef](#)]
33. Nagai, A.; Nakagawa, E.; Choi, H.B.; Hatori, K.; Kobayashi, S.; Kim, S.U. Erythropoietin and erythropoietin receptors in human CNS neurons, astrocytes, microglia, and oligodendrocytes grown in culture. *J. Neuropathol. Exp. Neurol.* **2001**, *60*, 386–392. [[CrossRef](#)] [[PubMed](#)]

34. Maiese, K.; Chong, Z.Z.; Hou, J.; Shang, Y.C. Erythropoietin and oxidative stress. *Curr. Neurovasc. Res.* **2008**, *5*, 125–142. [[CrossRef](#)] [[PubMed](#)]
35. Sun, Y.; Calvert, J.W.; Zhang, J.H. Neonatal hypoxia/ischemia is associated with decreased inflammatory mediators after erythropoietin administration. *Stroke* **2005**, *36*, 1672–1678. [[CrossRef](#)] [[PubMed](#)]
36. Juul, S.E.; Beyer, R.P.; Bammler, T.K.; McPherson, R.J.; Wilkerson, J.; Farin, F.M. Microarray analysis of high-dose recombinant erythropoietin treatment of unilateral brain injury in neonatal mouse hippocampus. *Pediatr. Res.* **2009**, *65*, 485–492. [[CrossRef](#)] [[PubMed](#)]
37. Yis, U.; Kurul, S.H.; Kumral, A.; Tugyan, K.; Cilaker, S.; Yilmaz, O.; Genc, S.; Genc, K. Effect of erythropoietin on oxygen-induced brain injury in the newborn rat. *Neurosci. Lett.* **2008**, *448*, 245–249. [[CrossRef](#)] [[PubMed](#)]
38. Dziejko, M.; Felderhoff-Mueser, U.; Sifringer, M.; Krutz, B.; Bittigau, P.; Thor, F.; Heumann, R.; Buhner, C.; Ikonomidou, C.; Hansen, H.H. Erythropoietin protects the developing brain against n-methyl-d-aspartate receptor antagonist neurotoxicity. *Neurobiol. Dis.* **2004**, *15*, 177–187. [[CrossRef](#)] [[PubMed](#)]
39. Zhu, C.; Kang, W.; Xu, F.; Cheng, X.; Zhang, Z.; Jia, L.; Ji, L.; Guo, X.; Xiong, H.; Simbruner, G.; et al. Erythropoietin improved neurologic outcomes in newborns with hypoxic-ischemic encephalopathy. *Pediatrics* **2009**, *124*, e218–e226. [[CrossRef](#)] [[PubMed](#)]
40. Elmahdy, H.; El-Mashad, A.R.; El-Bahrawy, H.; El-Gohary, T.; El-Barbary, A.; Aly, H. Human recombinant erythropoietin in asphyxia neonatorum: Pilot trial. *Pediatrics* **2010**, *125*, e1135–e1142. [[CrossRef](#)] [[PubMed](#)]
41. Wu, Y.W.; Bauer, L.A.; Ballard, R.A.; Ferriero, D.M.; Glidden, D.V.; Mayock, D.E.; Chang, T.; Durand, D.J.; Song, D.; Bonifacio, S.L.; et al. Erythropoietin for neuroprotection in neonatal encephalopathy: Safety and pharmacokinetics. *Pediatrics* **2012**, *130*, 683–691. [[CrossRef](#)] [[PubMed](#)]
42. Wu, Y.W.; Mathur, A.M.; Chang, T.; McKinstry, R.C.; Mulkey, S.B.; Mayock, D.E.; Van Meurs, K.P.; Rogers, E.E.; Gonzalez, F.F.; Comstock, B.A.; et al. High-dose erythropoietin and hypothermia for hypoxic-ischemic encephalopathy: A phase ii trial. *Pediatrics* **2016**, *137*, e20160190. [[CrossRef](#)] [[PubMed](#)]
43. Juul, S.E.; Comstock, B.A.; Heagerty, P.J.; Mayock, D.E.; Goodman, A.M.; Hauge, S.; Gonzalez, F.; Wu, Y.W. High-dose erythropoietin for asphyxia and encephalopathy (heal): A randomized controlled trial—Background, aims, and study protocol. *Neonatology* **2018**, *113*, 331–338. [[CrossRef](#)] [[PubMed](#)]
44. Baserga, M.C.; Beachy, J.C.; Roberts, J.K.; Ward, R.M.; DiGeronimo, R.J.; Walsh, W.F.; Ohls, R.K.; Anderson, J.; Mayock, D.E.; Juul, S.E.; et al. Darbepoetin administration to neonates undergoing cooling for encephalopathy: A safety and pharmacokinetic trial. *Pediatr. Res.* **2015**, *78*, 315–322. [[CrossRef](#)] [[PubMed](#)]
45. Mitsialis, S.A.; Kourembanas, S. Stem cell-based therapies for the newborn lung and brain: Possibilities and challenges. *Semin. Perinatol.* **2016**, *40*, 138–151. [[CrossRef](#)] [[PubMed](#)]
46. Nabetani, M.; Shintaku, H.; Hamazaki, T. Future perspectives of cell therapy for neonatal hypoxic-ischemic encephalopathy. *Pediatr. Res.* **2018**, *83*, 356–363. [[CrossRef](#)] [[PubMed](#)]
47. Archambault, J.; Moreira, A.; McDaniel, D.; Winter, L.; Sun, L.; Hornsby, P. Therapeutic potential of mesenchymal stromal cells for hypoxic ischemic encephalopathy: A systematic review and meta-analysis of preclinical studies. *PLoS ONE* **2017**, *12*, e0189895. [[CrossRef](#)] [[PubMed](#)]
48. Jellema, R.K.; Wolfs, T.G.; Lima Passos, V.; Zwanenburg, A.; Ophelders, D.R.; Kuypers, E.; Hopman, A.H.; Dudink, J.; Steinbusch, H.W.; Andriessen, P.; et al. Mesenchymal stem cells induce t-cell tolerance and protect the preterm brain after global hypoxia-ischemia. *PLoS ONE* **2013**, *8*, e73031. [[CrossRef](#)] [[PubMed](#)]
49. Cotten, C.M.; Murtha, A.P.; Goldberg, R.N.; Grotegut, C.A.; Smith, P.B.; Goldstein, R.F.; Fisher, K.A.; Gustafson, K.E.; Waters-Pick, B.; Swamy, G.K.; et al. Feasibility of autologous cord blood cells for infants with hypoxic-ischemic encephalopathy. *J. Pediatr.* **2014**, *164*, 973–979. [[CrossRef](#)] [[PubMed](#)]
50. Heusch, G.; Botker, H.E.; Przyklenk, K.; Redington, A.; Yellon, D. Remote ischemic conditioning. *J. Am. Coll. Cardiol.* **2015**, *65*, 177–195. [[CrossRef](#)] [[PubMed](#)]
51. Baillieux, S.; Chacaroun, S.; Doutreleau, S.; Detante, O.; Pepin, J.L.; Verges, S. Hypoxic conditioning and the central nervous system: A new therapeutic opportunity for brain and spinal cord injuries? *Exp. Biol. Med.* **2017**, *242*, 1198–1206. [[CrossRef](#)] [[PubMed](#)]
52. Lim, S.Y.; Hausenloy, D.J. Remote ischemic conditioning: From bench to bedside. *Front. Physiol.* **2012**, *3*, 27. [[CrossRef](#)] [[PubMed](#)]
53. Drunalini Perera, P.N.; Hu, Q.; Tang, J.; Li, L.; Barnhart, M.; Doycheva, D.M.; Zhang, J.H.; Tang, J. Delayed remote ischemic postconditioning improves long term sensory motor deficits in a neonatal hypoxic ischemic rat model. *PLoS ONE* **2014**, *9*, e90258. [[CrossRef](#)] [[PubMed](#)]

54. Rocha-Ferreira, E.; Rudge, B.; Hughes, M.P.; Rahim, A.A.; Hristova, M.; Robertson, N.J. Immediate remote ischemic postconditioning reduces brain nitrotyrosine formation in a piglet asphyxia model. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 5763743. [[CrossRef](#)] [[PubMed](#)]
55. Fernandez-Lopez, D.; Martinez-Orgado, J.; Nunez, E.; Romero, J.; Lorenzo, P.; Moro, M.A.; Lizasoain, I. Characterization of the neuroprotective effect of the cannabinoid agonist win-55212 in an in vitro model of hypoxic-ischemic brain damage in newborn rats. *Pediatr. Res.* **2006**, *60*, 169–173. [[CrossRef](#)] [[PubMed](#)]
56. Fernandez-Lopez, D.; Pradillo, J.M.; Garcia-Yebenes, I.; Martinez-Orgado, J.A.; Moro, M.A.; Lizasoain, I. The cannabinoid win55212-2 promotes neural repair after neonatal hypoxia-ischemia. *Stroke* **2010**, *41*, 2956–2964. [[CrossRef](#)] [[PubMed](#)]
57. Alvarez, F.J.; Lafuente, H.; Rey-Santano, M.C.; Mielgo, V.E.; Gastiasoro, E.; Rueda, M.; Pertwee, R.G.; Castillo, A.I.; Romero, J.; Martinez-Orgado, J. Neuroprotective effects of the nonpsychoactive cannabinoid cannabidiol in hypoxic-ischemic newborn piglets. *Pediatr. Res.* **2008**, *64*, 653–658. [[CrossRef](#)] [[PubMed](#)]
58. Villapol, S.; Fau, S.; Renolleau, S.; Biran, V.; Charriaut-Marlangue, C.; Baud, O. Melatonin promotes myelination by decreasing white matter inflammation after neonatal stroke. *Pediatr. Res.* **2011**, *69*, 51–55. [[CrossRef](#)] [[PubMed](#)]
59. Jou, M.J.; Peng, T.I.; Yu, P.Z.; Jou, S.B.; Reiter, R.J.; Chen, J.Y.; Wu, H.Y.; Chen, C.C.; Hsu, L.F. Melatonin protects against common deletion of mitochondrial DNA-augmented mitochondrial oxidative stress and apoptosis. *J. Pineal Res.* **2007**, *43*, 389–403. [[CrossRef](#)] [[PubMed](#)]
60. Pandi-Perumal, S.R.; BaHamman, A.S.; Brown, G.M.; Spence, D.W.; Bharti, V.K.; Kaur, C.; Hardeland, R.; Cardinali, D.P. Melatonin antioxidative defense: Therapeutical implications for aging and neurodegenerative processes. *Neurotox. Res.* **2013**, *23*, 267–300. [[CrossRef](#)] [[PubMed](#)]
61. Robertson, N.J.; Faulkner, S.; Fleiss, B.; Bainbridge, A.; Andorka, C.; Price, D.; Powell, E.; Lecky-Thompson, L.; Thei, L.; Chandrasekaran, M.; et al. Melatonin augments hypothermic neuroprotection in a perinatal asphyxia model. *Brain* **2013**, *136*, 90–105. [[CrossRef](#)] [[PubMed](#)]
62. Aly, H.; Elmahdy, H.; El-Dib, M.; Rowisha, M.; Awany, M.; El-Gohary, T.; Elbatch, M.; Hamisa, M.; El-Mashad, A.R. Melatonin use for neuroprotection in perinatal asphyxia: A randomized controlled pilot study. *J. Perinatol.* **2015**, *35*, 186–191. [[CrossRef](#)] [[PubMed](#)]
63. Seabra, M.L.; Bignotto, M.; Pinto, L.R., Jr.; Tufik, S. Randomized, double-blind clinical trial, controlled with placebo, of the toxicology of chronic melatonin treatment. *J. Pineal Res.* **2000**, *29*, 193–200. [[CrossRef](#)] [[PubMed](#)]
64. Fulia, F.; Gitto, E.; Cuzzocrea, S.; Reiter, R.J.; Dugo, L.; Gitto, P.; Barberi, S.; Cordaro, S.; Barberi, I. Increased levels of malondialdehyde and nitrite/nitrate in the blood of asphyxiated newborns: Reduction by melatonin. *J. Pineal Res.* **2001**, *31*, 343–349. [[CrossRef](#)] [[PubMed](#)]
65. Hendaus, M.A.; Jomha, F.A.; Alhammadi, A.H. Melatonin in the management of perinatal hypoxic-ischemic encephalopathy: Light at the end of the tunnel? *Neuropsychiatr. Dis. Treat.* **2016**, *12*, 2473–2479. [[CrossRef](#)] [[PubMed](#)]
66. Ramirez, M.R.; Muraro, F.; Zylbersztejn, D.S.; Abel, C.R.; Arteni, N.S.; Lavinsky, D.; Netto, C.A.; Trindade, V.M. Neonatal hypoxia-ischemia reduces ganglioside, phospholipid and cholesterol contents in the rat hippocampus. *Neurosci. Res.* **2003**, *46*, 339–347. [[CrossRef](#)]
67. Ferrari, G.; Anderson, B.L.; Stephens, R.M.; Kaplan, D.R.; Greene, L.A. Prevention of apoptotic neuronal death by gm1 ganglioside. Involvement of trk neurotrophin receptors. *J. Biol. Chem.* **1995**, *270*, 3074–3080. [[CrossRef](#)] [[PubMed](#)]
68. Ballough, G.P.; Cann, F.J.; Smith, C.D.; Forster, J.S.; Kling, C.E.; Filbert, M.G. GM1 monosialoganglioside pretreatment protects against soman-induced seizure-related brain damage. *Mol. Chem. Neuropathol.* **1998**, *34*, 1–23. [[CrossRef](#)] [[PubMed](#)]
69. Sheng, L.; Li, Z. Adjuvant treatment with monosialoganglioside may improve neurological outcomes in neonatal hypoxic-ischemic encephalopathy: A meta-analysis of randomized controlled trials. *PLoS ONE* **2017**, *12*, e0183490. [[CrossRef](#)] [[PubMed](#)]
70. Faulkner, S.; Bainbridge, A.; Kato, T.; Chandrasekaran, M.; Kapetanakis, A.B.; Hristova, M.; Liu, M.; Evans, S.; De Vita, E.; Kelen, D.; et al. Xenon augmented hypothermia reduces early lactate/*N*-acetylaspartate and cell death in perinatal asphyxia. *Ann. Neurol.* **2011**, *70*, 133–150. [[CrossRef](#)] [[PubMed](#)]

71. Chakkarapani, E.; Dingley, J.; Liu, X.; Hoque, N.; Aquilina, K.; Porter, H.; Thoresen, M. Xenon enhances hypothermic neuroprotection in asphyxiated newborn pigs. *Ann. Neurol.* **2010**, *68*, 330–341. [[CrossRef](#)] [[PubMed](#)]
72. Azzopardi, D.; Robertson, N.J.; Bainbridge, A.; Cady, E.; Charles-Edwards, G.; Deierl, A.; Fagiolo, G.; Franks, N.P.; Griffiths, J.; Hajnal, J.; et al. Moderate hypothermia within 6 h of birth plus inhaled xenon versus moderate hypothermia alone after birth asphyxia (TOBY-Xe): A proof-of-concept, open-label, randomised controlled trial. *Lancet Neurol.* **2016**, *15*, 145–153. [[CrossRef](#)]
73. Faulkner, S.D.; Downie, N.A.; Mercer, C.J.; Kerr, S.A.; Sanders, R.D.; Robertson, N.J. A xenon recirculating ventilator for the newborn piglet: Developing clinical applications of xenon for neonates. *Eur. J. Anaesthesiol.* **2012**, *29*, 577–585. [[CrossRef](#)] [[PubMed](#)]
74. Broad, K.D.; Fierens, I.; Fleiss, B.; Rocha-Ferreira, E.; Ezzati, M.; Hassell, J.; Alonso-Alconada, D.; Bainbridge, A.; Kawano, G.; Ma, D.; et al. Inhaled 45–50% argon augments hypothermic brain protection in a piglet model of perinatal asphyxia. *Neurobiol. Dis.* **2016**, *87*, 29–38. [[CrossRef](#)] [[PubMed](#)]
75. Zhuang, L.; Yang, T.; Zhao, H.; Fidalgo, A.R.; Vizcaychipi, M.P.; Sanders, R.D.; Yu, B.; Takata, M.; Johnson, M.R.; Ma, D. The protective profile of argon, helium, and xenon in a model of neonatal asphyxia in rats. *Crit. Care Med.* **2012**, *40*, 1724–1730. [[CrossRef](#)] [[PubMed](#)]
76. Yamaguchi, M.; Okamoto, K.; Kusano, T.; Matsuda, Y.; Suzuki, G.; Fuse, A.; Yokota, H. The effects of xanthine oxidoreductase inhibitors on oxidative stress markers following global brain ischemia reperfusion injury in C57BL/6 mice. *PLoS ONE* **2015**, *10*, e0133980. [[CrossRef](#)] [[PubMed](#)]
77. Marro, P.J.; Mishra, O.P.; Delivoria-Papadopoulos, M. Effect of allopurinol on brain adenosine levels during hypoxia in newborn piglets. *Brain Res.* **2006**, *1073–1074*, 444–450. [[CrossRef](#)] [[PubMed](#)]
78. Rodriguez-Fanjul, J.; Duran Fernandez-Feijoo, C.; Lopez-Abad, M.; Lopez Ramos, M.G.; Balada Caballe, R.; Alcantara-Horillo, S.; Camprubi Camprubi, M. Neuroprotection with hypothermia and allopurinol in an animal model of hypoxic-ischemic injury: Is it a gender question? *PLoS ONE* **2017**, *12*, e0184643. [[CrossRef](#)] [[PubMed](#)]
79. Chaudhari, T.; McGuire, W. Allopurinol for preventing mortality and morbidity in newborn infants with hypoxic-ischaemic encephalopathy. *Cochrane Database Syst. Rev.* **2012**, CD006817. [[CrossRef](#)] [[PubMed](#)]
80. Kaandorp, J.J.; van Bel, F.; Veen, S.; Derks, J.B.; Groenendaal, F.; Rijken, M.; Roze, E.; Venema, M.M.; Rademaker, C.M.; Bos, A.F.; et al. Long-term neuroprotective effects of allopurinol after moderate perinatal asphyxia: Follow-up of two randomised controlled trials. *Arch. Dis. Child. Fetal Neonatal. Ed.* **2012**, *97*, F162–F166. [[CrossRef](#)] [[PubMed](#)]
81. Torrance, H.L.; Benders, M.J.; Derks, J.B.; Rademaker, C.M.; Bos, A.F.; Van Den Berg, P.; Longini, M.; Buonocore, G.; Venegas, M.; Baquero, H.; et al. Maternal allopurinol during fetal hypoxia lowers cord blood levels of the brain injury marker s-100b. *Pediatrics* **2009**, *124*, 350–357. [[CrossRef](#)] [[PubMed](#)]
82. Ilves, P.; Kiisk, M.; Soopold, T.; Talvik, T. Serum total magnesium and ionized calcium concentrations in asphyxiated term newborn infants with hypoxic-ischaemic encephalopathy. *Acta Paediatr.* **2000**, *89*, 680–685. [[CrossRef](#)] [[PubMed](#)]
83. Bhat, M.A.; Charoo, B.A.; Bhat, J.I.; Ahmad, S.M.; Ali, S.W.; Mufti, M.U. Magnesium sulfate in severe perinatal asphyxia: A randomized, placebo-controlled trial. *Pediatrics* **2009**, *123*, e764–e769. [[CrossRef](#)] [[PubMed](#)]
84. Galinsky, R.; Bennet, L.; Groenendaal, F.; Lear, C.A.; Tan, S.; van Bel, F.; Juul, S.E.; Robertson, N.J.; Mallard, C.; Gunn, A.J. Magnesium is not consistently neuroprotective for perinatal hypoxia-ischemia in term-equivalent models in preclinical studies: A systematic review. *Dev. Neurosci.* **2014**, *36*, 73–82. [[CrossRef](#)] [[PubMed](#)]
85. Lingam, I.; Robertson, N.J. Magnesium as a neuroprotective agent: A review of its use in the fetus, term infant with neonatal encephalopathy, and the adult stroke patient. *Dev. Neurosci.* **2018**, *40*, 1–12. [[CrossRef](#)] [[PubMed](#)]
86. Tagin, M.; Shah, P.S.; Lee, K.S. Magnesium for newborns with hypoxic-ischemic encephalopathy: A systematic review and meta-analysis. *J. Perinatol.* **2013**, *33*, 663–669. [[CrossRef](#)] [[PubMed](#)]
87. Tagin, M.; Abdel-Hady, H.; ur Rahman, S.; Azzopardi, D.V.; Gunn, A.J. Neuroprotection for perinatal hypoxic ischemic encephalopathy in low- and middle-income countries. *J. Pediatr.* **2015**, *167*, 25–28. [[CrossRef](#)] [[PubMed](#)]

88. Filippi, L.; Poggi, C.; la Marca, G.; Furlanetto, S.; Fiorini, P.; Cavallaro, G.; Plantulli, A.; Donzelli, G.; Guerrini, R. Oral topiramate in neonates with hypoxic ischemic encephalopathy treated with hypothermia: A safety study. *J. Pediatr.* **2010**, *157*, 361–366. [[CrossRef](#)] [[PubMed](#)]
89. Filippi, L.; Fiorini, P.; Catarzi, S.; Berti, E.; Padrini, L.; Landucci, E.; Donzelli, G.; Bartalena, L.; Fiorentini, E.; Boldrini, A.; et al. Safety and efficacy of topiramate in neonates with hypoxic ischemic encephalopathy treated with hypothermia (NeoNATI): A feasibility study. *J. Matern. Fetal Neonatal. Med.* **2018**, *31*, 973–980. [[CrossRef](#)] [[PubMed](#)]
90. Amantea, D.; Certo, M.; Petrelli, F.; Tassorelli, C.; Micieli, G.; Corasaniti, M.T.; Puccetti, P.; Fallarino, F.; Bagetta, G. Azithromycin protects mice against ischemic stroke injury by promoting macrophage transition towards m2 phenotype. *Exp. Neurol.* **2016**, *275 Pt 1*, 116–125. [[CrossRef](#)] [[PubMed](#)]
91. Barks, J.L.Y.; Silverstein, F. *Repurposing Azithromycin for Neonatal Neuroprotection: Next Steps*; PAS: Toronto, ON, Canada, 2018.
92. Murray, D.M.; O'Connor, C.M.; Ryan, C.A.; Korotchikova, I.; Boylan, G.B. Early EEG grade and outcome at 5 years after mild neonatal hypoxic ischemic encephalopathy. *Pediatrics* **2016**, *138*, e20160659. [[CrossRef](#)] [[PubMed](#)]
93. Robertson, N.J.; Tan, S.; Groenendaal, F.; van Bel, F.; Juul, S.E.; Bennet, L.; Derrick, M.; Back, S.A.; Valdez, R.C.; Northington, F.; et al. Which neuroprotective agents are ready for bench to bedside translation in the newborn infant? *J. Pediatr.* **2012**, *160*, 544–552. [[CrossRef](#)] [[PubMed](#)]
94. Covey, M.V.; Loporchio, D.; Buono, K.D.; Levison, S.W. Opposite effect of inflammation on subventricular zone versus hippocampal precursors in brain injury. *Ann. Neurol.* **2011**, *70*, 616–626. [[CrossRef](#)] [[PubMed](#)]
95. Fancy, S.P.; Harrington, E.P.; Yuen, T.J.; Silbereis, J.C.; Zhao, C.; Baranzini, S.E.; Bruce, C.C.; Otero, J.J.; Huang, E.J.; Nusse, R.; et al. Axin2 as regulatory and therapeutic target in newborn brain injury and remyelination. *Nat. Neurosci.* **2011**, *14*, 1009–1016. [[CrossRef](#)] [[PubMed](#)]
96. Buser, J.R.; Maire, J.; Riddle, A.; Gong, X.; Nguyen, T.; Nelson, K.; Luo, N.L.; Ren, J.; Struve, J.; Sherman, L.S.; et al. Arrested preoligodendrocyte maturation contributes to myelination failure in premature infants. *Ann. Neurol.* **2012**, *71*, 93–109. [[CrossRef](#)] [[PubMed](#)]
97. Weaver, I.C.; Champagne, F.A.; Brown, S.E.; Dymov, S.; Sharma, S.; Meaney, M.J.; Szyf, M. Reversal of maternal programming of stress responses in adult offspring through methyl supplementation: Altering epigenetic marking later in life. *J. Neurosci.* **2005**, *25*, 11045–11054. [[CrossRef](#)] [[PubMed](#)]
98. Faraco, G.; Pancani, T.; Formentini, L.; Mascagni, P.; Fossati, G.; Leoni, F.; Moroni, F.; Chiarugi, A. Pharmacological inhibition of histone deacetylases by suberoylanilide hydroxamic acid specifically alters gene expression and reduces ischemic injury in the mouse brain. *Mol. Pharmacol.* **2006**, *70*, 1876–1884. [[CrossRef](#)] [[PubMed](#)]
99. Brogdon, J.L.; Xu, Y.; Szabo, S.J.; An, S.; Buxton, F.; Cohen, D.; Huang, Q. Histone deacetylase activities are required for innate immune cell control of th1 but not Th2 effector cell function. *Blood* **2007**, *109*, 1123–1130. [[CrossRef](#)] [[PubMed](#)]
100. Reynolds, A.D.; Banerjee, R.; Liu, J.; Gendelman, H.E.; Mosley, R.L. Neuroprotective activities of CD4⁺CD25⁺ regulatory t cells in an animal model of parkinson's disease. *J. Leukoc. Biol.* **2007**, *82*, 1083–1094. [[CrossRef](#)] [[PubMed](#)]
101. Ponomarev, E.D.; Veremeyko, T.; Barteneva, N.; Krichevsky, A.M.; Weiner, H.L. MicroRNA-124 promotes microglia quiescence and suppresses eae by deactivating macrophages via the c/ebp-alpha-pu.1 pathway. *Nat. Med.* **2011**, *17*, 64–70. [[CrossRef](#)] [[PubMed](#)]
102. Covey, M.V.; Jiang, Y.; Alli, V.V.; Yang, Z.; Levison, S.W. Defining the critical period for neocortical neurogenesis after pediatric brain injury. *Dev. Neurosci.* **2010**, *32*, 488–498. [[CrossRef](#)] [[PubMed](#)]
103. Van Velthoven, C.T.; Kavelaars, A.; van Bel, F.; Heijnen, C.J. Repeated mesenchymal stem cell treatment after neonatal hypoxia-ischemia has distinct effects on formation and maturation of new neurons and oligodendrocytes leading to restoration of damage, corticospinal motor tract activity, and sensorimotor function. *J. Neurosci.* **2010**, *30*, 9603–9611. [[CrossRef](#)] [[PubMed](#)]
104. Colton, C.A. Heterogeneity of microglial activation in the innate immune response in the brain. *J. Neuroimmune Pharmacol.* **2009**, *4*, 399–418. [[CrossRef](#)] [[PubMed](#)]
105. Gressens, P.; Marret, S.; Hill, J.M.; Brenneman, D.E.; Gozes, I.; Fridkin, M.; Evrard, P. Vasoactive intestinal peptide prevents excitotoxic cell death in the murine developing brain. *J. Clin. Investig.* **1997**, *100*, 390–397. [[CrossRef](#)] [[PubMed](#)]

106. Destot-Wong, K.D.; Liang, K.; Gupta, S.K.; Favrais, G.; Schwendimann, L.; Pansiot, J.; Baud, O.; Spedding, M.; Lelievre, V.; Mani, S.; et al. The ampa receptor positive allosteric modulator, s18986, is neuroprotective against neonatal excitotoxic and inflammatory brain damage through bdnf synthesis. *Neuropharmacology* **2009**, *57*, 277–286. [[CrossRef](#)] [[PubMed](#)]
107. Robertson, N.J.; Nakakeeto, M.; Hagmann, C.; Cowan, F.M.; Acolet, D.; Iwata, O.; Allen, E.; Elbourne, D.; Costello, A.; Jacobs, I. Therapeutic hypothermia for birth asphyxia in low-resource settings: A pilot randomised controlled trial. *Lancet* **2008**, *372*, 801–803. [[CrossRef](#)]
108. Pauliah, S.S.; Shankaran, S.; Wade, A.; Cady, E.B.; Thayyil, S. Therapeutic hypothermia for neonatal encephalopathy in low- and middle-income countries: A systematic review and meta-analysis. *PLoS ONE* **2013**, *8*, e58834. [[CrossRef](#)] [[PubMed](#)]
109. Bamji, M.S.; PV, V.S.M.; Williams, L.; Vardhana Rao, M.V. Maternal nutritional status & practices & perinatal, neonatal mortality in rural andhra pradesh, india. *Indian J. Med. Res.* **2008**, *127*, 44–51. [[PubMed](#)]
110. Osredkar, D.; Thoresen, M.; Maes, E.; Flatebo, T.; Elstad, M.; Sabir, H. Hypothermia is not neuroprotective after infection-sensitized neonatal hypoxic-ischemic brain injury. *Resuscitation* **2014**, *85*, 567–572. [[CrossRef](#)] [[PubMed](#)]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).