



HHS Public Access

Author manuscript

J Matern Fetal Neonatal Med. Author manuscript; available in PMC 2019 July 01.

Published in final edited form as:

J Matern Fetal Neonatal Med. 2019 July ; 32(13): 2113–2136. doi:10.1080/14767058.2018.1427058.

The Profiles of Soluble Adhesion Molecules in the “Great Obstetrical Syndromes”

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Abstract

Objective—The objective of this study was to determine the profiles of maternal plasma soluble adhesion molecules in patients with preeclampsia, small-for-gestational-age (SGA) fetuses, acute pyelonephritis, preterm labor with intact membranes (PTL), preterm prelabor rupture of the membranes (preterm PROM), and fetal death.

Materials and methods—A cross-sectional study was conducted to determine maternal plasma concentrations of sE-selectin, sL-selectin, and sP-selectin as well as sICAM-1, sVCAM-1, and sPECAM-1 in patients with 1) an uncomplicated pregnancy (control, n=100); 2) preeclampsia (n=94); 3) SGA fetuses (in women without preeclampsia/hypertension, n=45); 4) acute pyelonephritis (n=25); 5) PTL (n=53); 6) preterm PROM (n=24); and 7) fetal death (n=34). Concentrations of soluble adhesion molecules and inflammatory cytokines (tumor necrosis factor

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This study was presented at the 12th World Congress of Perinatal Medicine, November 3–6, 2015, in Madrid, Spain, as a poster presentation.

Conflict of Interest: The authors declare no conflicts of interest.

(TNF)- α and interleukin (IL)-8) were determined with sensitive and specific enzyme-linked immunoassays.

Results—In comparison to women with a normal pregnancy, 1) women with preeclampsia had higher median concentrations of sE-selectin, sP-selectin, and sVCAM-1, and a lower concentration of sL-selectin (all p-values <0.001); 2) patients with SGA fetuses had higher median concentrations of sE-selectin, sP-selectin, and sVCAM-1 (all p-values <0.05); 3) patients with a fetal death had higher median concentrations of sE-selectin and sP-selectin (all p-values <0.05); 4) patients with acute pyelonephritis had higher median plasma concentrations of sE-selectin, sICAM-1, and sVCAM-1 (all p-values < 0.001); 5) patients with preeclampsia and acute pyelonephritis, plasma concentrations of sVCAM-1, sE-selectin, and sP-selectin correlated with those of the pro-inflammatory cytokines TNF- α and interleukin (IL)-8 (all p-values <0.05); 6) patients with PTL had a higher median concentration of sP-selectin and a lower median concentration of VCAM-1 (all p-values <0.05); and 7) women with preterm PROM had lower median concentrations of sL-selectin and sVCAM-1 (all p-values <0.05).

Conclusions—The results of this study show that endothelial cell activation/dysfunction reflected by the plasma concentration of sE-selectin is not specific to preeclampsia but is present in pregnancies complicated by SGA fetuses, acute pyelonephritis, and fetal death. Collectively, we report that each obstetrical syndrome appears to have a stereotypical profile of soluble adhesion molecules in the peripheral circulation.

Keywords

preeclampsia; preterm labor; prelabor rupture of the membranes (PROM); pyelonephritis; small-for-gestational-age fetus

Introduction

Normal pregnancy is characterized by systemic intravascular inflammation as evidenced by phenotypic and functional activation of circulating granulocytes and monocytes measured by flow cytometry (1, 2) and also by an increase in plasma/serum concentrations of acute phase reactant proteins such as C-reactive protein (3, 4), fibrinogen (5–7), and complement split products (8–10). Central to the inflammatory process is endothelial activation (increased adhesion of endothelial cells to leukocytes), which may lead to endothelial cell dysfunction (decreased synthesis, release, and/or activity of endothelium-derived nitric oxide) (11–15). Changes of the endothelium in response to inflammation modify the leukocyte adhesion cascade, allowing leukocytes to attach to the endothelium and, subsequently, to transmigrate into the perivascular tissue (11, 13, 15–21).

Leukocyte-endothelial cell interactions are mainly mediated by cell adhesion molecules comprised of three families (selectins, integrins, and members of the immunoglobulin gene superfamily) (11, 13, 15–27). Pro-inflammatory cytokines change the expression of cell adhesion molecules that subsequently results in their shedding from the endothelium, leukocytes, or platelets (16, 22, 25, 28). Therefore, the soluble forms can be detected in the peripheral blood *in vivo* and in the supernatant during *in vitro* studies. The plasma

concentrations of these soluble adhesion molecules have been used as markers of endothelial, platelet, and leukocytic activation (29–32).

Intravascular inflammation is observed in the “great obstetrical syndromes,” which includes preeclampsia (1, 33–39), small-for-gestational-age (SGA) fetuses (40–48), acute pyelonephritis (2, 49–51), preterm labor with intact membranes (PTL) (52–54), and preterm prelabor rupture of the membranes (preterm PROM) (53–55). Previous studies have shown conflicting results about the profile of soluble adhesion molecules in preeclampsia (31, 32, 41–46, 56–110). Moreover, conclusions about the behavior of adhesion molecules require a comprehensive study of several obstetrical syndromes to ensure that changes are specific to each disease state. For example, the claim that preeclampsia is associated with endothelial cell activation based on an increase in plasma concentration of sE-selectin (31, 32, 43–45, 59, 62, 65, 68, 70, 71, 77, 80–82, 90, 94, 97, 103, 105, 110) would need to be revisited if similar findings could be observed in other obstetrical complications.

Therefore, this cross-sectional study was undertaken to determine whether the profiles of maternal plasma soluble adhesion molecules differ among patients with preeclampsia, SGA fetuses, acute pyelonephritis, PTL, preterm PROM, and fetal death.

Materials and Methods

Study design

A retrospective cross-sectional study was conducted. All samples were obtained from the Bank of Biological Materials of Wayne State University, the Detroit Medical Center, and the Perinatology Research Branch of the *Eunice Kennedy Shriver* National Institute of Child Health and Human Development (NICHD) (Detroit, MI). All patients were enrolled in the Labor and Delivery Unit or in the antenatal clinic at Hutzel Women’s Hospital (Detroit, MI). Women with the following characteristics were included: 1) normal pregnancy (n=100); 2) preeclampsia (n=94); 3) SGA fetuses (in women without preeclampsia/hypertension, n=45); 4) acute pyelonephritis (n=25); 5) PTL (n=53); 6) preterm PROM (n=24); and 7) fetal death (n=34). Women with multiple gestations and pregnancies affected by fetal chromosomal and/or structural anomalies were excluded. A subset of patients in this current study was included in a prior study (77).

All patients provided written informed consent, and the use of biological specimens and clinical data for research purposes was approved by the Institutional Review Boards of Wayne State University and NICHD.

Clinical definitions

Women who had a normal pregnancy met the following conditions: 1) venipuncture samples obtained between 20–42 weeks of gestation; 2) no medical, obstetrical, or surgical complications; and 3) delivery of a normal term (> 37 weeks of gestation) infant whose birth weight measured between the 10th and 90th percentiles for gestational age (114, 115).

Preeclampsia was defined as new-onset hypertension that developed after 20 weeks of gestation (systolic and/or diastolic blood pressure of \geq 140 and/or \geq 90 mmHg; measured on

at least two occasions, 4 hours to 1 week apart) and proteinuria (≥ 300 mg in a 24-hour urine collection, or two random urine specimens obtained 4 hours to 1 week apart containing ≥ 1+ protein by dipstick) (111). Severe preeclampsia was diagnosed according to criteria proposed by the American Congress of Obstetricians and Gynecologists (ACOG) (111, 112). Early-onset preeclampsia and late-onset preeclampsia were defined as cases diagnosed before and after 34 weeks of gestation, respectively (113).

An SGA fetus was defined as having a neonatal birth weight <10th percentile for gestational age (114). Patients with SGA fetuses were classified according to the umbilical artery Doppler velocimetry result (SGA fetus with an absent end-diastolic velocimetry (AEDV) and SGA fetus without an AEDV).

Acute pyelonephritis was diagnosed in the presence of fever (temperature ≥ 38°C), clinical signs or symptoms of an upper urinary tract infection (e.g., flank pain, costovertebral angle tenderness), pyuria, and a positive urine culture for microorganisms (115, 116). Women with acute pyelonephritis were classified as follows: 1) acute pyelonephritis with a positive urine culture only (without bacteremia) and 2) acute pyelonephritis with positive blood and urine cultures (with bacteremia).

Preterm labor with intact membranes was defined as the presence of premature uterine contractions leading to cervical changes and resulting in preterm delivery. Preterm PROM was diagnosed as amniorrhexis in a preterm gestation leading to preterm delivery. Fetal death was defined as death of the fetus after 20 weeks of gestation, confirmed by ultrasound (all fetal deaths were unexplained).

Sample collection and immunoassay

Venipuncture was performed upon the diagnosis of each pregnancy complication or upon enrollment at 20–42 weeks of gestation for women who had an uncomplicated pregnancy. Blood was collected in tubes containing EDTA. Following centrifugation, samples were stored at –70°C. The concentrations of soluble adhesion molecules were measured using an enzyme-linked immunosorbent assay (ELISA). The plasma concentrations of TNF- α and IL-8 were also determined using sensitive and specific immunoassays. All immunoassay kits were purchased from R&D Systems (Minneapolis, MN, USA), except sPECAM-1, which was purchased from Diaclone Research (Cedex, France). Table 1 displays the sensitivity and the inter-assay and intra-assay coefficients of variation for each assay.

Doppler velocimetry of the uterine and umbilical arteries

Pulse-wave and color Doppler ultrasound (Acuson, Sequoia, Mountain View, CA, USA) examinations of the uterine and umbilical arteries were performed by trained personnel for a subset of patients at the time of diagnosis of an SGA fetus. Uterine artery Doppler velocimetry was defined as abnormal if the mean resistance index (average of right and left) was above the 95th percentile for gestational age (117). Umbilical artery Doppler velocimetry was defined as abnormal if either the pulsatility index (PI) was above the 95th percentile for gestational age (118) or if waveforms were abnormal (absent or reverse end-diastolic velocities) (119). The inter- and intra-observer coefficients of variation of the

umbilical and uterine arteries were 0.81 (for both arteries), and 0.85 (umbilical artery) and 0.91 (uterine artery), respectively.

Statistical analysis

Normality of continuous variables was assessed using the Kolmogorov-Smirnov test and visual inspection of histograms. The chi-square test was used to evaluate the differences in proportions. Spearman's rank correlation coefficients were used to determine the relationship between the plasma soluble adhesion molecules and the plasma pro-inflammatory cytokine concentrations. For concentrations below the detection limit, 99% of the lowest detectable concentration across all samples was used. Group-level differences in concentrations were evaluated using the Kruskal-Wallis test, and pairwise comparisons were performed using the Mann-Whitney U test. An alternative analysis was performed using linear models with adjustment for gestational age at sampling. A p-value of <0.05 was considered significant. Analysis was performed using the R statistical language and environment (www.r-project.org).

Results

Clinical characteristics of the study population

The demographic, clinical, and obstetrical characteristics of the study population are displayed in Table 2. The median maternal age, rate of nulliparity, ethnicity, and median gestational age at sample collection and at delivery, as well as birth weight, differed among the study groups ($p < 0.05$ for all comparisons).

Among patients with preeclampsia, 76% (71/94) had preterm preeclampsia, 50% (47/94) had early-onset preeclampsia, and 38% (36/93) had preeclampsia with an SGA neonate. The rate of bacteremia was 52.2% (12/23) and the most common microorganism identified was *Escherichia coli* (7/12).

The soluble adhesion molecules, except sICAM-1, were detected in the maternal plasma of all patients. Soluble ICAM-1 was undetectable in 14 (3.7%) samples. Among women with uncomplicated pregnancies, maternal plasma sP-selectin, sL-selectin, sICAM-1, and sPECAM-1 did not correlate with gestational age at sampling (all p-values >0.05).

Plasma soluble adhesion molecules in patients with preeclampsia

The median maternal plasma concentrations of sE-selectin, sP-selectin, and sVCAM-1 were significantly higher among patients with preeclampsia compared to those with a normal pregnancy [preeclampsia (ng/mL): sE-selectin: 64.2 (46.1–81.4); sP-selectin: 116.2 (92.6–156.8); and sVCAM-1: 744.1 (641.1–986.7) versus normal pregnancy (ng/mL): sE-selectin: 45.1 (33.9–53.3); sP-selectin: 92.6 (72.4–111.1); and sVCAM-1: 507.8 (451.8–593.7); all p-values <0.001]. Additionally, patients with preeclampsia had a significantly lower median plasma sL-selectin concentration (ng/mL) compared to the controls [559.3 (444.9–671.2) versus 669.7 (583.4–830.4); p-value <0.001]. The median plasma concentrations of sICAM-1 and sPECAM-1 did not change significantly between these two groups (all p-values = 0.05) (Table 3A, Table 4, and Figures 1–6 and Supplementary Figure S1).

There were no significant differences in the median plasma concentration of soluble adhesion molecules among patients with preeclampsia based on the severity of preeclampsia (all p-values ≥ 0.05) (please see Supplementary material). The median plasma sE-selectin, sP-selectin, and sVCAM-1 concentrations in patients with preeclampsia were positively correlated with the plasma concentrations of the liver enzyme serum glutamic oxaloacetic transaminase (SGOT) [sP-selectin: Spearman's $\rho=0.22$, p-value=0.04; sE-selectin: Spearman's $\rho=0.24$, p-value =0.03; and sVCAM-1: Spearman's $\rho=0.30$, p-value =0.007]. By contrast, the plasma sVCAM-1 concentration had a significantly negative correlation with the platelet count [Spearman's $\rho=-0.24$, p-value=0.02].

Plasma soluble adhesion molecules in patients with small-for-gestational-age fetuses

The median maternal plasma concentrations of sE-selectin, sP-selectin, and sVCAM-1 were significantly higher in patients with SGA neonates compared to those with normal pregnancies [women with SGA neonates (ng/mL): sE-selectin: 54.5 (36.8–76.8); sP-selectin: 121.5 (94.1–145.4); and sVCAM-1: 595.3 (510.5–719.7) versus normal pregnancy (ng/mL): sE-selectin: 45.1 (33.9–53.3); sP-selectin: 92.6 (72.4–111.1); and sVCAM-1: 507.8 (451.8–593.7); all p-values <0.05]; (Table 3A, Table 4, and Figures 1–6, Supplementary Figure S2). However, the median plasma concentrations of sL-selectin, sICAM-1, and sPECAM-1 did not differ significantly between patients with SGA fetuses and those with normal pregnancies (all p-values ≥ 0.05). Among SGA patients, the median plasma concentration of sVCAM-1 was increased in those who had an AEDV than in patients without an AEDV, but this difference failed to reach statistical significance [SGA with AEDV: 670.1 (629.2–719.7) versus SGA without AEDV: 566.3 (460.6–730.2); p=0.05]. Moreover, a plasma sVCAM-1 concentration had a significant positive correlation with the mean uterine artery resistant index (RI) [Spearman's $\rho=0.5$; p-value <0.03].

Plasma soluble adhesion molecules in patients with a fetal death

Patients with a fetal death had higher median maternal plasma concentrations of sE-selectin and sP-selectin than women with normal pregnancies [fetal death (ng/mL): sE-selectin: 53.3 (37.2–64.0); sP-selectin: 119.2 (86.3–171.2) vs. normal pregnancy (ng/mL): sE-selectin: 45.1 (33.9–53.3); and sP-selectin: 92.6 (72.4–111.1); all p-values <0.05]. No significant differences were observed in the median concentrations of sL-selectin, sVCAM-1, sICAM-1, and sPECAM-1 between patients with a fetal death and a normal pregnancy (Table 3A, Table 4, and Figures 1–6, Supplementary Figure S3).

Plasma soluble adhesion molecules in patients with acute pyelonephritis

Patients with acute pyelonephritis had higher median plasma concentrations of sE-selectin, sVCAM-1, and ICAM-1 compared to the controls [acute pyelonephritis (ng/mL): sE-selectin 99.1 (76.8–175.2), sVCAM-1: 671.3 (543.7–713.4), sICAM-1 442.8 (332.6–622.2) versus normal pregnancy (ng/mL): sE-selectin 45.1 (33.9–53.3) ; sVCAM-1: 507.8 (451.8–593.7);sICAM-1: 266.7(217.6–312.8); all p-values < 0.001] (Table 3B, Table 4, and Figures 1–6, Supplementary Figure S4). The median concentration of sL-selectin was lower in patients with acute pyelonephritis compared to the controls, yet the result did not reach statistical significance (potentially due to a small sample size). No significant differences were observed in the concentrations of sP-selectin and sPECAM-1 between patients with

acute pyelonephritis and normal pregnancy (all p-values ≤ 0.05) (Table 3B, Table 4, and Figure 1–6, Supplementary Figure S4).

Among patients with acute pyelonephritis, those with bacteremia had a significantly high median maternal plasma concentration of sE-selectin, sVCAM-1, and sICAM-1 ($p < 0.01$) than those without bacteremia (Table 5).

The correlation of soluble adhesion molecules and pro-inflammatory cytokines in patients with preeclampsia and acute pyelonephritis

In patients with preeclampsia, plasma sVCAM-1 concentration had a significant positive correlation with TNF- α and IL-8 concentrations (Spearman's $\rho = 0.36$ and 0.29 ; p-value < 0.05 , for both groups). The correlation coefficients were stronger in patients with acute pyelonephritis (TNF- α : Spearman's $\rho = 0.57$; p-value = 0.003 ; and IL-8: Spearman's $\rho = 0.43$; p-value = 0.038) compared to those with preeclampsia.

Plasma sE-selectin and sP-selectin concentrations were positively correlated with plasma IL-8 concentrations in patients with both preeclampsia and acute pyelonephritis. Furthermore, the correlations were stronger in patients with acute pyelonephritis than in those with preeclampsia (preeclampsia: sE-selectin: Spearman's $\rho = 0.23$; p-value = 0.03 ; sP-selectin: Spearman's $\rho = 0.28$; p-value = 0.007 ; acute pyelonephritis: sE-selectin: Spearman's $\rho = 0.41$; p-value = 0.045 ; and sP-selectin: Spearman's $\rho = 0.5$; p-value = 0.02).

Plasma soluble adhesion molecules in patients with preterm labor and intact membranes

Patients with PTL who delivered preterm had a higher median concentration of sP-selectin and a lower median concentration of sVCAM-1 compared to those with normal pregnancies [PTL (ng/mL): sP-selectin: 102.5 (83.2 – 133.3) and sVCAM-1: 454.0 (402.6 – 529.7) versus normal pregnancy (ng/mL): sP-selectin: 92.6 (72.4 – 111.1) and sVCAM-1: 507.8 (451.8 – 593.7); all p-values < 0.05]. No significant differences were observed in the median concentrations of sE-selectin, sL-selectin, and sICAM-1 between patients with preterm labor and intact membranes and the controls (all p-values ≥ 0.05) (Table 3B, Table 4, and Figure 1–6, Supplementary figure S5). The plasma concentration of sPECAM-1 was not measured in the PTL group.

Plasma soluble adhesion molecules in patients with preterm prelabor rupture of the membranes

Women with preterm PROM had lower median concentrations of sL-selectin and sVCAM-1 than patients who had a normal pregnancy [preterm PROM (ng/mL): sL-selectin: 518.1 (429.1 – 638.7) and sVCAM-1: 445.3 (376.2 – 472.6) versus normal pregnancy (ng/mL): sL-selectin: 669.7 (583.4 – 830.4) and sVCAM-1: 507.8 (451.8 – 593.7); all p-values < 0.05]. No significant differences were observed in the median concentrations of sE-selectin, sL-selectin, and sICAM-1 between patients with preterm PROM and the controls (Table 3B, Table 4, and Figures 1–6, Supplementary Figure S6). The plasma concentration of sPECAM-1 was not measured in patients with preterm PROM.

Discussion

Principal findings of the study

1) Women with preeclampsia had higher median concentrations of sE-selectin, sP-selectin, and sVCAM-1 and a lower median concentration of sL-selectin compared to women who had a normal pregnancy. This is evidence of endothelial, leukocytic, and platelet activation in this syndrome; 2) there was a significant positive correlation between the plasma sE-selectin, sP-selectin, and sVCAM-1 concentrations in patients with preeclampsia and the liver enzyme SGOT, a marker of multi-organ involvement in preeclampsia. This suggests that endothelial and platelet activation participate in the hepatic damage that occurs in preeclampsia; 3) patients with SGA fetuses had higher median concentrations of sE-selectin, sP-selectin, and sVCAM-1 compared to the controls; 4) patients with acute pyelonephritis had higher median plasma concentrations of sE-selectin, sVCAM-1, and sICAM-1 compared to the controls; 5) plasma sVCAM-1, sE-selectin, and sP-selectin concentrations had moderate correlations with pro-inflammatory cytokines (TNF- α and IL-8) in patients with preeclampsia and acute pyelonephritis. These correlations were stronger in acute pyelonephritis than in preeclampsia; 6) patients with preterm labor and intact membranes had a higher median concentration of sP-selectin and a lower median concentration of sVCAM-1 compared to the controls; 7) women with preterm PROM had lower median concentrations of sL-selectin and sVCAM-1 compared to the controls; and 8) patients with a fetal death had higher median concentrations of sE-selectin and sP-selectin compared to the controls.

Overall, the results of this study suggest that each obstetrical syndrome is characterized by a stereotypical profile of soluble adhesion molecules in the maternal plasma.

Soluble adhesion molecules and endothelial activation/dysfunction

The endothelium is a single cell layer that lines the interior surface of the blood and lymphatic vessels. Its fundamental functions include maintenance of vascular tone, hemostasis, fluid filtration, cell adhesion, and neutrophil recruitment (11, 13, 19, 21, 120). During an inflammatory response, the endothelium undergoes activation/dysfunction. Endothelial cell activation refers to increased adhesive properties of the endothelium to leukocytes in response to biomechanical stimuli (121–123) or cytokines (124–130). In endothelial cell dysfunction, there is an impaired endothelial cell-dependent relaxation (11, 13, 15), and this was initially observed in a human *in vivo* model of essential hypertension (131). Endothelial cell activation can lead to an endothelial dysfunction, and the major link between these processes is the leukocyte adhesion cascade (11, 13, 15–21).

The leukocyte adhesion cascade is central to the development of an inflammatory response and is a regulated, multi-step process consisting of capture, rolling, slow rolling, firm adhesion, adhesion strengthening, intramural crawling, and paracellular/transcellular migration of the leukocyte (11, 13, 16–21). The leukocyte-endothelial cell interactions are predominantly mediated by cell adhesion molecules comprised of three families (selectins, integrins, and members of the immunoglobulin gene superfamily) (16, 17, 19, 22–27). The selectin family is mainly responsible for the rolling of the leukocyte on the surface of the

endothelium and includes three members: E-selectin, L-selectin, and P-selectin (11, 13, 16–27). ICAM-1, VCAM-1, and PECAM-1 are ligands for leukocyte integrins and belong to the immunoglobulin superfamily. These molecules are important for adhesion, arrest, and transmigration of leukocytes (11, 13, 16–21, 23, 25).

In response to pro-inflammatory cytokines, specific adhesion molecules are shed from the surface of the endothelium, and they can be detected in the peripheral circulation (16, 22, 25, 28). High concentrations of such soluble forms of E-selectin have been interpreted as evidence of endothelial activation/dysfunction. In contrast, high concentrations of sL-selectin inhibit the attachment of leukocytes to the endothelium; therefore, low sL-selectin is indicative of leukocyte activation (132). Previous studies have shown that soluble adhesion molecules could be used as markers in various inflammatory conditions such as sepsis (14, 133, 134), acute pancreatitis (135, 136), rheumatoid arthritis (137–139), and cancer metastasis (140, 141).

Preeclampsia is associated with the activation of platelets, leukocytes, and endothelial cells

Preeclampsia is one of the great obstetrical syndromes (142–152) and is characterized by utero-placental insufficiency (151, 153–175), an imbalance between angiogenic and anti-angiogenic factors in the maternal plasma (37, 38, 46, 47, 151, 176–211), increased thrombin generation (212–221), platelet aggregation (214, 221–226), endothelial cell dysfunction (51, 110, 156, 161, 227–236), and exaggerated intravascular inflammation (1, 33–39).

Central to the inflammatory response are the adhesion molecules that mediate leukocyte-endothelial cell interactions (11, 13, 15–21). These molecules are also important for the development of early placentation and successful physiological transformation of the spiral arteries (237–246). However, in preeclampsia, the cytotrophoblasts fail to up-regulate expression of the immunoglobulin superfamily adhesion receptors (i.e., VCAM-1, PECAM-1); therefore, these cytotrophoblasts fail to mimic a vascular adhesion phenotype (168, 247–249). Additionally, the expression of adhesion molecules on cultured endothelial cells is stimulated by factors in the maternal circulation, e.g., pro-inflammatory cytokines that could be detected in patients with preeclampsia (29, 250). Due to the systemic inflammatory response (1, 33–39), endothelial activation/dysfunction (51, 110, 156, 161, 227–236), and the hypercoagulable state in preeclampsia (212–226), adhesion molecules are shed from the surface of cells, and their soluble forms can be detected in the peripheral circulation (16, 22, 25, 28).

The profiles of soluble adhesion molecules have been examined in patients with preeclampsia, yet the results are conflicting (31, 32, 41–46, 56–110). In this study, we simultaneously determined the behavior of the six adhesion molecules in preeclampsia and found higher median concentrations of sE-selectin, sVCAM-1 (both reflect activation of the endothelium), and sP-selectin (indicative of platelet activation) and a lower concentration of sL-selectin (represents leukocyte activation). Moreover, plasma sE-selectin, sP-selectin, and sVCAM-1 concentrations were correlated with the severity of liver injury in patients with preeclampsia as well as with pro-inflammatory cytokines (TNF- α and IL-8).

SGA pregnancy is associated with endothelial and platelet activation

A pregnancy with an SGA fetus is another great obstetrical syndrome (142, 149, 150) and shares common pathophysiological mechanisms with preeclampsia, such as utero-placental ischemia (155, 157, 158, 163, 169, 170, 173), abnormal placentation (111, 153, 154, 251–260), an imbalance between angiogenic and anti-angiogenic factors in the maternal plasma (46, 47, 185, 187, 190, 193–197, 199, 210, 261), platelet changes (262), and increased oxidative stress (232, 263–269). However, despite the shared pathophysiology, it is still unknown why some women go on to develop preeclampsia with or without an SGA fetus, whereas other women have only SGA fetuses. It was initially thought that endothelial involvement is systemic in preeclampsia and localized only to the utero-placental unit in SGA pregnancies (270). However, other investigators have reported enhanced endothelial (42–47) and neutrophil (40, 41, 48) activation in women with SGA fetuses, suggesting that there is a systemic maternal inflammatory response.

Herein, we report that patients with an SGA fetus had significantly higher plasma sE-selectin, sP-selectin, and sVCAM-1 concentrations than those who had a normal pregnancy. These results are indicative of the presence of endothelial and platelet activation in pregnancies with SGA fetuses. Our findings are similar to those reported by Johnson et al. (44) and Coata et al. (45) who showed that patients with SGA fetuses had higher sE-selectin and sVCAM-1 concentrations compared to the controls. Collectively, these observations suggest that patients with SGA fetuses have endothelial activation/dysfunction, and the profiles of the soluble adhesion molecules of these patients differ from those observed in pregnancies complicated by preeclampsia.

Pyelonephritis is associated with the activation of the endothelium

Acute pyelonephritis is a frequent cause of maternal systemic inflammation (271–276). Previous studies investigating the behavior of cytokines (277), chemokines (278), and complement (49, 279) in preeclampsia and acute pyelonephritis showed that both conditions were associated with intravascular inflammation (2, 34). However, transcriptomic analysis of the peripheral blood of patients with pyelonephritis (280) and preeclampsia (281) demonstrated differences in the inflammatory response (pyelonephritis was associated with increased expression of genes involved in innate immunity and decreased expression of genes involved in lymphocyte function (280), whereas the transcriptional profile of peripheral whole blood in preeclampsia demonstrated differential expression of genes involved in coagulation, immune regulation, the growth/developmental process, host defense, and tight junctions in the blood-brain barrier (281).

In the study herein, we present evidence indicating that acute pyelonephritis was associated with higher plasma concentrations of sE-selectin, sVCAM-1, and sICAM-1 compared to a normal pregnancy. The concentrations of sE-selectin, sVCAM-1, and sICAM-1 were associated with the degree of severity (bacteremia). Moreover, the magnitude of the differences was significantly greater in acute pyelonephritis than in preeclampsia. These findings demonstrate marked endothelial activation in acute pyelonephritis. Also, the dramatic rise in the sICAM-1 concentration was detected only in acute pyelonephritis, suggesting that sICAM-1 is more specific to acute pyelonephritis than to other obstetrical

conditions such as preeclampsia. In support of this finding, previous studies that used immunohistochemistry have shown increased expression of ICAM-1 in the kidneys with pyelonephritis (282, 283) and various glomerulonephritis (284–286). Additionally, elevated concentrations of sICAM-1 (a marker of endothelial activation/dysfunction) are well-documented in human models of sepsis (133, 134, 287–299). An unexpected finding was the lack of a demonstrable decrease in the plasma concentration of sL-selectin, a marker for leukocyte activation. We believe that the sample size of patients in the pyelonephritis group account for this observation, given the strong evidence for leukocyte activation derived from studies with flow-cytometry (2). Overall, more studies are needed to further explore the role of soluble adhesion molecules in patients with acute pyelonephritis.

The profiles of soluble adhesion molecules in preterm labor with intact membranes and preterm prelabor rupture of the membranes

We report herein that preterm labor was associated with a higher median concentration of sP-selectin but with a lower median concentration of sVCAM-1 than normal pregnancy. On the other hand, patients with preterm PROM had lower median concentrations of sL-selectin and sVCAM-1 compared to the controls. The profiles of the soluble adhesion molecules reflected platelet and leukocytic activation in preterm labor with intact membranes and in leukocyte activation in preterm PROM; however, under both conditions, no marked endothelial activity was observed. Previous studies that examined the behavior of soluble adhesion molecules in the maternal plasma/serum yielded conflicting results (300–304). For example, Chen et al, (304) showed that women with preterm delivery (spontaneous preterm labor or preterm PROM) had higher serum sICAM-1 and sVCAM-1 concentrations than patients with a normal term pregnancy, whereas no change was observed in the sE-selectin concentration. On the other hand, Bartha et al, (303) found no significant differences in the concentrations of sE-selectin, sVCAM-1, and sICAM-1 between patients who had spontaneous preterm labor and those with a normal term pregnancy. Even though the results from previous studies and of those reported herein are inconsistent, maternal intravascular inflammation (52–55) and platelet activation (305, 306) have been demonstrated in preterm labor and preterm PROM. Additionally, data from epidemiological studies showed that women with preterm deliveries are at an increased risk for future cardiovascular diseases (307–312), suggesting that there is some degree of endothelial dysfunction. Therefore, larger studies are needed to further examine the behavior of soluble adhesion molecules in different patient populations with preterm labor and preterm PROM.

The profiles of soluble adhesion molecules in patients with an unexplained fetal death

The observations of this study reflect platelet activation in the maternal circulation of cases with a fetal demise, as women with an unexplained fetal death had a higher median concentration of sP-selectin than those with a normal pregnancy. This is consistent with a previous study that reported a higher plasma sCD40L (marker of platelet activation) concentration in mothers with fetal demise than in those with normal pregnancies (313). Also, the concentrations of sP-selectin were higher in non-pregnant women with a history of venous thromboembolism and fetal death than in those with a history of thromboembolism but no fetal death (314). Collectively, these results suggest that there is a subclinical activation of the hemostatic system in women with fetal death.

Herein, we also report that patients with an unexplained fetal death had a higher median concentration of sE-selectin than women with a normal pregnancy, reflecting activation of the endothelium. Activation of the endothelium and platelets can occur in the setting of an inflammatory response (315, 316); yet, the role of inflammation in unexplained fetal death remains to be elucidated. A previous study reported that women with unexplained fetal death had a higher concentration of plasma C5a (a potent pro-inflammatory mediator (317, 318)) than women with a normal pregnancy (319). These observations are important as C5a can increase the expression of adhesion molecules on the endothelium and activate neutrophils (317, 320, 321), which, in turn, can activate platelets by secreting cathepsin G (322). Moreover, experimental evidence suggests that complement activation requires the stimulation of platelets, which is associated with the expression of P-selectin (323–326). Therefore, additional studies are needed to elucidate the link between inflammation and activation of the hemostatic system, endothelium, and leukocytes in patients with an unexplained fetal death.

Strengths and Limitations

This is the first study to simultaneously investigate the profile of six soluble adhesion molecules in pregnancies complicated by preeclampsia, SGA fetuses, acute pyelonephritis, preterm labor with intact membranes, preterm PROM, and fetal death.

Limitations include those stemming from the small sample size of some of the groups (i.e. pyelonephritis). Additionally, our study population is consistent with mainly African American patients, and there is a concern that our findings may not be applicable to other ethnic groups. However, in term of ethnic risk for pregnancy complications, African-American women are at markedly increased risk for preterm delivery, preeclampsia and cardiovascular diseases (327, 328). Second, there are differences in adhesions molecules concentrations in various ethnic groups. Indeed, Miller et al, found that females of African origin (Caribbean and West African) had significantly lower concentrations of sICAM-1, sVCAM-1, and sP-selectin than White women in England (329). Similar results were found by Akolekar et al, who reported that the concentrations of plasma P-selectin in women with a normal pregnancy between 11–13 weeks of gestation were lower in Blacks and Orientals than in White women (330). However, the effect of ethnicity on soluble adhesion molecules throughout pregnancy and in the great obstetrical syndromes was not studied yet, and it is beyond the scope of the current study. This may affect the generalization of our results to the entire obstetrical population; nevertheless, our findings are important and are relevant to the ethnic group with the highest risk for such complications. Further studies are needed to address the effect of ethnicity on the profile of the different.

Conclusions

Endothelial activation/dysfunction is present not only in preeclampsia but also in other obstetrical syndromes such as pregnancy complicated by an SGA fetus, acute pyelonephritis, and an unexplained fetal death. Collectively, we report that each obstetrical syndrome appears to have a stereotypical profile of soluble adhesion molecules in the peripheral circulation.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Funding

This research was supported, in part, by the Perinatology Research Branch, Program for Perinatal Research and Obstetrics, Division of Intramural Research, *Eunice Kennedy Shriver* National Institute of Child Health and Human Development, National Institutes of Health, U.S. Department of Health and Human Services (NICHD/NIH/DHHS); and, in part, with Federal funds from NICHD/NIH/DHHS under Contract No. HHSN275201300006C.

References

1. Sacks GP, Studena K, Sargent K, Redman CW. Normal pregnancy and preeclampsia both produce inflammatory changes in peripheral blood leukocytes akin to those of sepsis. *American journal of obstetrics and gynecology*. 1998; 179(1):80–6. [PubMed: 9704769]
2. Naccasha N, Gervasi MT, Chaiworapongsa T, Berman S, Yoon BH, Maymon E, et al. Phenotypic and metabolic characteristics of monocytes and granulocytes in normal pregnancy and maternal infection. *American journal of obstetrics and gynecology*. 2001; 185(5):1118–23. [PubMed: 11717644]
3. Watts DH, Krohn MA, Wener MH, Eschenbach DA. C-reactive protein in normal pregnancy. *Obstetrics and gynecology*. 1991; 77(2):176–80. [PubMed: 1988876]
4. Belo L, Santos-Silva A, Rocha S, Caslake M, Cooney J, Pereira-Leite L, et al. Fluctuations in C-reactive protein concentration and neutrophil activation during normal human pregnancy. *European journal of obstetrics, gynecology, and reproductive biology*. 2005; 123(1):46–51.
5. Romero R, Rickles FR, Matthews E, Scott D, Dinan C, Duffy T. Fibrinopeptide A during normal pregnancy. *American journal of perinatology*. 1988; 5(1):70–3. [PubMed: 3257391]
6. van Buul EJ, Steegers EA, Jongsma HW, Eskes TK, Thomas CM, Hein PR. Haematological and biochemical profile of uncomplicated pregnancy in nulliparous women; a longitudinal study. *The Netherlands journal of medicine*. 1995; 46(2):73–85. [PubMed: 7885525]
7. Manten GT, Franx A, Sikkema JM, Hameeteman TM, Visser GH, de Groot PG, et al. Fibrinogen and high molecular weight fibrinogen during and after normal pregnancy. *Thrombosis research*. 2004; 114(1):19–23. [PubMed: 15262480]
8. Gallery ED, Raftos J, Gyory AZ, Wells JV. A prospective study of serum complement (C3 and C4) levels in normal human pregnancy: effect of the development of pregnancy-associated hypertension. *Australian and New Zealand journal of medicine*. 1981; 11(3):243–5. [PubMed: 6945834]
9. Jagadeesan V. Serum complement levels in normal pregnancy and pregnancy-induced hypertension. *International journal of gynaecology and obstetrics: the official organ of the International Federation of Gynaecology and Obstetrics*. 1988; 26(3):389–91.
10. Richani K, Soto E, Romero R, Espinoza J, Chaiworapongsa T, Nien JK, et al. Normal pregnancy is characterized by systemic activation of the complement system. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2005; 17(4):239–45.
11. Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. *Nature reviews Immunology*. 2007; 7(10):803–15.
12. Zhang C. The role of inflammatory cytokines in endothelial dysfunction. *Basic research in cardiology*. 2008; 103(5):398–406. [PubMed: 18600364]
13. Pate M, Damarla V, Chi DS, Negi S, Krishnaswamy G. Endothelial cell biology: role in the inflammatory response. *Advances in clinical chemistry*. 2010; 52:109–30. [PubMed: 21275341]
14. Paulus P, Jennewein C, Zacharowski K. Biomarkers of endothelial dysfunction: can they help us deciphering systemic inflammation and sepsis? *Biomarkers : biochemical indicators of exposure, response, and susceptibility to chemicals*. 2011; 16(Suppl 1):S11–21.

15. Liao JK. Linking endothelial dysfunction with endothelial cell activation. *The Journal of clinical investigation*. 2013; 123(2):540–1. [PubMed: 23485580]
16. Radi ZA, Kehrli ME Jr, Ackermann MR. Cell adhesion molecules, leukocyte trafficking, and strategies to reduce leukocyte infiltration. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine*. 2001; 15(6):516–29.
17. Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nature reviews Immunology*. 2007; 7(9):678–89.
18. Zarbock A, Ley K. Mechanisms and consequences of neutrophil interaction with the endothelium. *The American journal of pathology*. 2008; 172(1):1–7. [PubMed: 18079440]
19. Nourshargh S, Alon R. Leukocyte migration into inflamed tissues. *Immunity*. 2014; 41(5):694–707. [PubMed: 25517612]
20. Gerhardt T, Ley K. Monocyte trafficking across the vessel wall. *Cardiovascular research*. 2015; 107(3):321–30. [PubMed: 25990461]
21. Muller WA. The regulation of transendothelial migration: new knowledge and new questions. *Cardiovascular research*. 2015; 107(3):310–20. [PubMed: 25987544]
22. Bevilacqua MP, Nelson RM. Selectins. *The Journal of clinical investigation*. 1993; 91(2):379–87. [PubMed: 7679406]
23. Petruzzelli L, Takami M, Humes HD. Structure and function of cell adhesion molecules. *The American journal of medicine*. 1999; 106(4):467–76. [PubMed: 10225251]
24. Kneuer C, Ehrhardt C, Radomski MW, Bakowsky U. Selectins--potential pharmacological targets? *Drug discovery today*. 2006; 11(21–22):1034–40. [PubMed: 17055414]
25. Smith CW. 3. Adhesion molecules and receptors. *The Journal of allergy and clinical immunology*. 2008; 121(2 Suppl):S375–9. [PubMed: 18241685]
26. Zarbock A, Ley K, McEver RP, Hidalgo A. Leukocyte ligands for endothelial selectins: specialized glycoconjugates that mediate rolling and signaling under flow. *Blood*. 2011; 118(26):6743–51. [PubMed: 22021370]
27. Telen MJ. Cellular adhesion and the endothelium: E-selectin, L-selectin, and pan-selectin inhibitors. *Hematology/oncology clinics of North America*. 2014; 28(2):341–54. [PubMed: 24589270]
28. Garton KJ, Gough PJ, Raines EW. Emerging roles for ectodomain shedding in the regulation of inflammatory responses. *Journal of leukocyte biology*. 2006; 79(6):1105–16. [PubMed: 16565325]
29. Abe E, Matsubara K, Ochi H, Ito M, Oka K, Kameda K. Elevated levels of adhesion molecules derived from leukocytes and endothelial cells in patients with pregnancy-induced hypertension. *Hypertension in pregnancy*. 2003; 22(1):31–43. [PubMed: 12648441]
30. Abe Y, El-Masri B, Kimball KT, Pownall H, Reilly CF, Osmundsen K, et al. Soluble cell adhesion molecules in hypertriglyceridemia and potential significance on monocyte adhesion. *Arteriosclerosis, thrombosis, and vascular biology*. 1998; 18(5):723–31.
31. Austgulen R, Lien E, Vince G, Redman CW. Increased maternal plasma levels of soluble adhesion molecules (ICAM-1, VCAM-1, E-selectin) in preeclampsia. *European journal of obstetrics, gynecology, and reproductive biology*. 1997; 71(1):53–8.
32. Kim SY, Ryu HM, Yang JH, Kim MY, Ahn HK, Lim HJ, et al. Maternal serum levels of VCAM-1, ICAM-1 and E-selectin in preeclampsia. *Journal of Korean medical science*. 2004; 19(5):688–92. [PubMed: 15483345]
33. Redman CW, Sacks GP, Sargent IL. Preeclampsia: an excessive maternal inflammatory response to pregnancy. *American journal of obstetrics and gynecology*. 1999; 180(2 Pt 1):499–506. [PubMed: 9988826]
34. Gervasi MT, Chaiworapongsa T, Pacora P, Naccasha N, Yoon BH, Maymon E, et al. Phenotypic and metabolic characteristics of monocytes and granulocytes in preeclampsia. *American journal of obstetrics and gynecology*. 2001; 185(4):792–7. [PubMed: 11641653]
35. Chaiworapongsa T, Gervasi MT, Refuerzo J, Espinoza J, Yoshimatsu J, Berman S, et al. Maternal lymphocyte subpopulations (CD45RA+ and CD45RO+) in preeclampsia. *American journal of obstetrics and gynecology*. 2002; 187(4):889–93. [PubMed: 12388971]
36. Redman CW, Sargent IL. Preeclampsia and the systemic inflammatory response. *Seminars in nephrology*. 2004; 24(6):565–70. [PubMed: 15529291]

37. Chaemsaitong P, Chaiworapongsa T, Romero R, Korzeniewski SJ, Stampalija T, Than NG, et al. Maternal plasma soluble TRAIL is decreased in preeclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2014; 27(3):217–27.
38. Chaiworapongsa T, Romero R, Korzeniewski SJ, Kusanovic JP, Soto E, Lam J, et al. Maternal plasma concentrations of angiogenic/antiangiogenic factors in the third trimester of pregnancy to identify the patient at risk for stillbirth at or near term and severe late preeclampsia. *American journal of obstetrics and gynecology.* 2013; 208(4):287e1–e15. [PubMed: 23333542]
39. Bouwland-Both MI, Steegers EA, Lindemans J, Russcher H, Hofman A, Geurts-Moespot AJ, et al. Maternal soluble fms-like tyrosine kinase-1, placental growth factor, plasminogen activator inhibitor-2, and folate concentrations and early fetal size: the Generation R study. *American journal of obstetrics and gynecology.* 2013; 209(2):121e1–11. [PubMed: 23583216]
40. Johnston TA, Greer IA, Dawes J, Calder AA. Neutrophil activation in small for gestational age pregnancies. *British journal of obstetrics and gynaecology.* 1991; 98(1):105–6. [PubMed: 1998619]
41. Sabatier F, Bretelle F, D'Ercole C, Boubli L, Sampol J, Dignat-George F. Neutrophil activation in preeclampsia and isolated intrauterine growth restriction. *American journal of obstetrics and gynecology.* 2000; 183(6):1558–63. [PubMed: 11120528]
42. Phocas I, Rizos D, Papoulias J, Xyni K, Sarandakou A, Salamalekis E. A comparative study of serum soluble vascular cell adhesion molecule-1 and soluble intercellular adhesion molecule-1 in preeclampsia. *Journal of perinatology : official journal of the California Perinatal Association.* 2000; 20(2):114–9. [PubMed: 10785888]
43. Bretelle F, Sabatier F, Blann A, D'Ercole C, Boutiere B, Mutin M, et al. Maternal endothelial soluble cell adhesion molecules with isolated small for gestational age fetuses: comparison with pre-eclampsia. *BJOG : an international journal of obstetrics and gynaecology.* 2001; 108(12):1277–82. [PubMed: 11843391]
44. Johnson MR, Anim-Nyame N, Johnson P, Sooranna SR, Steer PJ. Does endothelial cell activation occur with intrauterine growth restriction? *BJOG : an international journal of obstetrics and gynaecology.* 2002; 109(7):836–9. [PubMed: 12135223]
45. Coata G, Pennacchi L, Bini V, Liotta L, Di Renzo GC. Soluble adhesion molecules: marker of pre-eclampsia and intrauterine growth restriction. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2002; 12(1):28–34.
46. Crispi F, Dominguez C, Llurba E, Martin-Gallan P, Cabero L, Gratacos E. Placental angiogenic growth factors and uterine artery Doppler findings for characterization of different subsets in preeclampsia and in isolated intrauterine growth restriction. *American journal of obstetrics and gynecology.* 2006; 195(1):201–7. [PubMed: 16545329]
47. Chaiworapongsa T, Espinoza J, Gotsch F, Kim YM, Kim GJ, Goncalves LF, et al. The maternal plasma soluble vascular endothelial growth factor receptor-1 concentration is elevated in SGA and the magnitude of the increase relates to Doppler abnormalities in the maternal and fetal circulation. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2008; 21(1):25–40.
48. Ogge G, Romero R, Chaiworapongsa T, Gervasi MT, Pacora P, Erez O, et al. Leukocytes of pregnant women with small-for-gestational age neonates have a different phenotypic and metabolic activity from those of women with preeclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2010; 23(6):476–87.
49. Soto E, Richani K, Romero R, Espinoza J, Chaiworapongsa T, Nien JK, et al. Increased concentration of the complement split product C5a in acute pyelonephritis during pregnancy. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2005; 17(4):247–52.

50. Mazaki-Tovi S, Vaisbuch E, Romero R, Kusanovic JP, Chaiworapongsa T, Kim SK, et al. Maternal plasma concentration of the pro-inflammatory adipokine pre-B-cell-enhancing factor (PBEF)/visfatin is elevated in pregnant patients with acute pyelonephritis. *American journal of reproductive immunology*. 2010; 63(3):252–62. [PubMed: 20085562]
51. Adekola H, Romero R, Chaemsathong P, Korzeniewski SJ, Dong Z, Yeo L, et al. Endocan, a putative endothelial cell marker, is elevated in preeclampsia, decreased in acute pyelonephritis, and unchanged in other obstetrical syndromes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2015; 28(14):1621–32.
52. Gervasi MT, Chaiworapongsa T, Naccasha N, Blackwell S, Yoon BH, Maymon E, et al. Phenotypic and metabolic characteristics of maternal monocytes and granulocytes in preterm labor with intact membranes. *American journal of obstetrics and gynecology*. 2001; 185(5):1124–9. [PubMed: 11717645]
53. Assi F, Fruscio R, Bonardi C, Ghidini A, Allavena P, Mantovani A, et al. Pentraxin 3 in plasma and vaginal fluid in women with preterm delivery. *BJOG : an international journal of obstetrics and gynaecology*. 2007; 114(2):143–7. [PubMed: 17305891]
54. Cruciani L, Romero R, Vaisbuch E, Kusanovic JP, Chaiworapongsa T, Mazaki-Tovi S, et al. Pentraxin 3 in maternal circulation: an association with preterm labor and preterm PROM, but not with intra-amniotic infection/inflammation. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2010; 23(10):1097–105.
55. Gervasi MT, Chaiworapongsa T, Naccasha N, Pacora P, Berman S, Maymon E, et al. Maternal intravascular inflammation in preterm premature rupture of membranes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2002; 11(3):171–5.
56. Greer IA, Lyall F, Perera T, Boswell F, Macara LM. Increased concentrations of cytokines interleukin-6 and interleukin-1 receptor antagonist in plasma of women with preeclampsia: a mechanism for endothelial dysfunction? *Obstetrics and gynecology*. 1994; 84(6):937–40. [PubMed: 7526306]
57. Lyall F, Greer IA, Boswell F, Macara LM, Walker JJ, Kingdom JC. The cell adhesion molecule, VCAM-1, is selectively elevated in serum in pre-eclampsia: does this indicate the mechanism of leucocyte activation? *British journal of obstetrics and gynaecology*. 1994; 101(6):485–7. [PubMed: 7517182]
58. Meekins JW, McLaughlin PJ, West DC, McFadyen IR, Johnson PM. Endothelial cell activation by tumour necrosis factor-alpha (TNF-alpha) and the development of pre-eclampsia. *Clinical and experimental immunology*. 1994; 98(1):110–4. [PubMed: 7523006]
59. Fickling SA, Whitley GS, Nussey SS. The cell adhesion molecule, VCAM-1, is selectively elevated in serum in pre-eclampsia: does this indicate the mechanism of leucocyte activation? *British journal of obstetrics and gynaecology*. 1995; 102(2):173–4. [PubMed: 7538783]
60. Halim A, Kanayama N, el Maradny E, Nakashima A, Bhuiyan AB, Khatun S, et al. Plasma P selectin (GMP-140) and glycocalicin are elevated in preeclampsia and eclampsia: their significances. *American journal of obstetrics and gynecology*. 1996; 174(1 Pt 1):272–7. [PubMed: 8572020]
61. Djurovic S, Schjetlein R, Wisloff F, Haugen G, Berg K. Increased levels of intercellular adhesion molecules and vascular cell adhesion molecules in pre-eclampsia. *British journal of obstetrics and gynaecology*. 1997; 104(4):466–70. [PubMed: 9141584]
62. Krauss T, Kuhn W, Lakoma C, Augustin HG. Circulating endothelial cell adhesion molecules as diagnostic markers for the early identification of pregnant women at risk for development of preeclampsia. *American journal of obstetrics and gynecology*. 1997; 177(2):443–9. [PubMed: 9290466]
63. Airoidi L, Gaffuri B, Rossi G, Iurlaro E, Nozza A, Vigano P, et al. Soluble intercellular adhesion molecule-1 serum profile in physiologic and preeclamptic pregnancy. *American journal of reproductive immunology*. 1998; 39(3):183–8. [PubMed: 9526607]

64. Budak E, Madazli R, Aksu MF, Benian A, Gezer A, Palit N, et al. Vascular cell adhesion molecule-1 (VCAM-1) and leukocyte activation in pre-eclampsia and eclampsia. *International journal of gynaecology and obstetrics: the official organ of the International Federation of Gynaecology and Obstetrics*. 1998; 63(2):115–21.
65. Daniel Y, Kupfermanc MJ, Baram A, Jaffa AJ, Wolman I, Shenhav M, et al. Plasma soluble endothelial selectin is elevated in women with pre-eclampsia. *Human reproduction*. 1998; 13(12): 3537–41. [PubMed: 9886546]
66. Higgins JR, Papayianni A, Brady HR, Darling MR, Walshe JJ. Circulating vascular cell adhesion molecule-1 in pre-eclampsia, gestational hypertension, and normal pregnancy: evidence of selective dysregulation of vascular cell adhesion molecule-1 homeostasis in pre-eclampsia. *American journal of obstetrics and gynecology*. 1998; 179(2):464–9. [PubMed: 9731854]
67. Hubel CA, Lyall F, Weissfeld L, Gandley RE, Roberts JM. Small low-density lipoproteins and vascular cell adhesion molecule-1 are increased in association with hyperlipidemia in preeclampsia. *Metabolism: clinical and experimental*. 1998; 47(10):1281–8. [PubMed: 9781635]
68. Krauss T, Osmers R, Beran J, Diedrich F, Fleckenstein G, Kuhn W. Soluble adhesion molecules in patients with pre-eclampsia. *Zentralblatt fur Gynakologie*. 1998; 120(6):279–83. [PubMed: 9659698]
69. Daniel Y, Kupfermanc MJ, Baram A, Geva E, Fait G, Lessing JB. A selective increase in plasma soluble vascular cell adhesion molecule-1 levels in preeclampsia. *American journal of reproductive immunology*. 1999; 41(6):407–12. [PubMed: 10392229]
70. Heyl W, Handt S, Reister F, Gehlen J, Mittermayer C, Rath W. The role of soluble adhesion molecules in evaluating endothelial cell activation in preeclampsia. *American journal of obstetrics and gynecology*. 1999; 180(1 Pt 1):68–72. [PubMed: 9914581]
71. Heyl W, Handt S, Reister F, Gehlen J, Schroder W, Mittermayer C, et al. Elevated soluble adhesion molecules in women with pre-eclampsia. Do cytokines like tumour necrosis factor-alpha and interleukin-1beta cause endothelial activation. *European journal of obstetrics, gynecology, and reproductive biology*. 1999; 86(1):35–41.
72. Madazli R, Budak E, Calay Z, Aksu MF. Correlation between placental bed biopsy findings, vascular cell adhesion molecule and fibronectin levels in pre-eclampsia. *BJOG : an international journal of obstetrics and gynaecology*. 2000; 107(4):514–8. [PubMed: 10759271]
73. Acar A, Altinbas A, Ozturk M, Kosar A, Kirazli S. Selectins in normal pregnancy, pre-eclampsia and missed abortus. *Haematologia*. 2001; 31(1):33–8. [PubMed: 11345401]
74. Bowen RS, Moodley J, Dutton MF, Fickl H. Systemic inflammatory indices in pre-eclampsia and eclampsia. *Journal of obstetrics and gynaecology : the journal of the Institute of Obstetrics and Gynaecology*. 2001; 21(6):563–9. [PubMed: 12521769]
75. Oyama R. The relationship between the level of expression of intercellular adhesion molecule-1 in placenta and onset of preeclampsia. *The journal of obstetrics and gynaecology research*. 2001; 27(3):147–54. [PubMed: 11561831]
76. Aksoy H, Kumtepe Y, Akcay F, Yildirim AK. Correlation of P-selectin and lipoprotein(a), and other lipid parameters in preeclampsia. *Clinical and experimental medicine*. 2002; 2(1):39–43. [PubMed: 12049188]
77. Chaiworapongsa T, Romero R, Yoshimatsu J, Espinoza J, Kim YM, Park K, et al. Soluble adhesion molecule profile in normal pregnancy and pre-eclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2002; 12(1): 19–27.
78. Aliefendioğlu D, Erdem G, Tulek N, Yurdakok M. Neonatal and maternal serum levels of soluble ICAM-1 in preeclamptic and normal pregnancies. *American journal of perinatology*. 2002; 19(6): 333–9. [PubMed: 12357425]
79. Visser W, Beckmann I, Knook MA, Wallenburg HC. Soluble tumor necrosis factor receptor II and soluble cell adhesion molecule 1 as markers of tumor necrosis factor-alpha release in preeclampsia. *Acta obstetrica et gynecologica Scandinavica*. 2002; 81(8):713–9. [PubMed: 12174154]

80. Bersinger NA, Smarason AK, Muttukrishna S, Groome NP, Redman CW. Women with preeclampsia have increased serum levels of pregnancy-associated plasma protein A (PAPP-A), inhibin A, activin A and soluble E-selectin. Hypertension in pregnancy. 2003; 22(1):45–55. [PubMed: 12648442]
81. Anim-Nyame N, Sooranna SR, Johnson MR, Sullivan MH, Gamble J, Steer PJ. Impaired retrograde transmission of vasodilatory signals via the endothelium in pre-eclampsia: a cause of reduced tissue blood flow? Clinical science. 2004; 106(1):19–25. [PubMed: 12889986]
82. Aydin S, Benian A, Madazli R, Uludag S, Uzun H, Kaya S. Plasma malondialdehyde, superoxide dismutase, sE-selectin, fibronectin, endothelin-1 and nitric oxide levels in women with preeclampsia. European journal of obstetrics, gynecology, and reproductive biology. 2004; 113(1): 21–5.
83. Hanisch CG, Pfeiffer KA, Schlebusch H, Schmolling J. Adhesion molecules, activin and inhibin--candidates for the biochemical prediction of hypertensive diseases in pregnancy? Archives of gynecology and obstetrics. 2004; 270(2):110–5. [PubMed: 12898146]
84. Ramsay JE, Ferrell WR, Crawford L, Wallace AM, Greer IA, Sattar N. Divergent metabolic and vascular phenotypes in pre-eclampsia and intrauterine growth restriction: relevance of adiposity. Journal of hypertension. 2004; 22(11):2177–83. [PubMed: 15480103]
85. Vadachkoria S, Sanchez SE, Qiu C, Mui-Rivera M, Malinow MR, Williams MA. Hyperhomocyst(e)inemia and elevated soluble vascular cell adhesion molecule-1 concentrations are associated with an increased risk of preeclampsia. Gynecologic and obstetric investigation. 2004; 58(3):133–9. [PubMed: 15205565]
86. Donker RB, Molema G, Faas MM, Kallenberg CG, van Pampus MG, Timmer A, et al. Absence of in vivo generalized pro-inflammatory endothelial activation in severe, early-onset preeclampsia. Journal of the Society for Gynecologic Investigation. 2005; 12(7):518–28. [PubMed: 16202929]
87. Heyl W, Heintz B, Reister F, Harwig S, Witte K, Lemmer B, et al. Increased soluble VCAM-1 serum levels in preeclampsia are not correlated to urinary excretion or circadian blood pressure rhythm. Journal of perinatal medicine. 2005; 33(2):144–8. [PubMed: 15843265]
88. Vadachkoria S, Woelk GB, Mahomed K, Qiu C, Mui-Rivera M, Malinow MR, et al. Elevated soluble vascular cell adhesion molecule-1, elevated Homocyst(e)inemia, and hypertriglyceridemia in relation to preeclampsia risk. American journal of hypertension. 2006; 19(3):235–42. [PubMed: 16500507]
89. Lok CA, Nieuwland R, Sturk A, Hau CM, Boer K, Vanbavel E, et al. Microparticle-associated P-selectin reflects platelet activation in preeclampsia. Platelets. 2007; 18(1):68–72. [PubMed: 17365856]
90. Chavarria ME, Lara-Gonzalez L, Garcia-Paleta Y, Vital-Reyes VS, Reyes A. Adhesion molecules changes at 20 gestation weeks in pregnancies complicated by preeclampsia. European journal of obstetrics, gynecology, and reproductive biology. 2008; 137(2):157–64.
91. Laskowska M, Laskowska K, Leszczynska-Gorzela B, Oleszczuk J. sP-selectin in preeclamptic pregnancies with intrauterine normal growth and small-for-gestational-age foetus. Preliminary communication. Medycyna wieku rozwojowego. 2009; 13(3):212–7. [PubMed: 20081268]
92. Lok CA, Jebbink J, Nieuwland R, Faas MM, Boer K, Sturk A, et al. Leukocyte activation and circulating leukocyte-derived microparticles in preeclampsia. American journal of reproductive immunology. 2009; 61(5):346–59. [PubMed: 19341385]
93. Lewis DF, Canzoneri BJ, Gu Y, Zhao S, Wang Y. Maternal levels of prostacyclin, thromboxane, ICAM, and VCAM in normal and preeclamptic pregnancies. American journal of reproductive immunology. 2010; 64(6):376–83. [PubMed: 20482519]
94. Mori T, Shinohara K, Wakatsuki A, Watanabe K, Fujimaki A. Adipocytokines and endothelial function in preeclamptic women. Hypertension research : official journal of the Japanese Society of Hypertension. 2010; 33(3):250–4. [PubMed: 20075929]
95. Papakonstantinou K, Economou E, Hasiakos D, Vitoratos N. Antepartum and postpartum maternal plasma levels of E-selectin in pre-eclampsia, gestational proteinuria and gestational hypertension. European journal of obstetrics, gynecology, and reproductive biology. 2010; 153(1):112–3.

96. Robb AO, Din JN, Mills NL, Smith IB, Blomberg A, Zikry MN, et al. The influence of the menstrual cycle, normal pregnancy and pre-eclampsia on platelet activation. *Thrombosis and haemostasis*. 2010; 103(2):372–8. [PubMed: 20076841]
97. Strijbos MH, Snijder CA, Kraan J, Lamers CH, Gratama JW, Duvekot JJ. Levels of circulating endothelial cells in normotensive and severe preeclamptic pregnancies. *Cytometry Part B, Clinical cytometry*. 2010; 78(6):382–6.
98. Szarka A, Rigo J Jr, Lazar L, Beko G, Molvarec A. Circulating cytokines, chemokines and adhesion molecules in normal pregnancy and preeclampsia determined by multiplex suspension array. *BMC immunology*. 2010; 11:59. [PubMed: 21126355]
99. Molvarec A, Szarka A, Walentin S, Beko G, Karadi I, Prohaszka Z, et al. Serum heat shock protein 70 levels in relation to circulating cytokines, chemokines, adhesion molecules and angiogenic factors in women with preeclampsia. *Clinica chimica acta; international journal of clinical chemistry*. 2011; 412(21–22):1957–62. [PubMed: 21756887]
100. Molvarec A, Szarka A, Walentin S, Beko G, Karadi I, Prohaszka Z, et al. Serum leptin levels in relation to circulating cytokines, chemokines, adhesion molecules and angiogenic factors in normal pregnancy and preeclampsia. *Reproductive biology and endocrinology : RB&E*. 2011; 9:124. [PubMed: 21906313]
101. Veas CJ, Aguilera VC, Munoz IJ, Gallardo VI, Miguel PL, Gonzalez MA, et al. Fetal endothelium dysfunction is associated with circulating maternal levels of sE-selectin, sVCAM1, and sFlt-1 during pre-eclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2011; 24(11):1371–7.
102. Xiong Y, Zhou SF, Zhou R, Yang D, Xu ZF, Lou YT, et al. Alternations of maternal and cord plasma hemostasis in preeclampsia before and after delivery. *Hypertension in pregnancy*. 2011; 30(3):347–58. [PubMed: 21174589]
103. Carty DM, Anderson LA, Freeman DJ, Welsh PI, Brennand JE, Dominiczak AF, et al. Early pregnancy soluble E-selectin concentrations and risk of preeclampsia. *Journal of hypertension*. 2012; 30(5):954–9. [PubMed: 22441350]
104. Fei X, Hongxiang Z, Qi C, Daozhen C. Maternal plasma levels of endothelial dysfunction mediators including AM, CGRP, sICAM-1 and tHcy in pre-eclampsia. *Advances in clinical and experimental medicine : official organ Wroclaw Medical University*. 2012; 21(5):573–9. [PubMed: 23356193]
105. Mehrabian F, Jazi SM, Javanmard SH, Kaviani M, Homayouni V. Circulating endothelial cells (CECs) and E-selectin: Predictors of preeclampsia. *Journal of research in medical sciences : the official journal of Isfahan University of Medical Sciences*. 2012; 17(1):15–21. [PubMed: 23248652]
106. Farzadnia M, Ayatollahi H, Hasan-Zade M, Rahimi HR. A comparative study of vascular cell adhesion molecule-1 and high-sensitive C-reactive protein in normal and preeclamptic pregnancies. *Interventional medicine & applied science*. 2013; 5(1):26–30. [PubMed: 24265885]
107. Laskowska M, Laskowska K, Oleszczuk J. Elevated maternal serum sP-selectin levels in preeclamptic pregnancies with and without intrauterine fetal growth restriction, but not in normotensive pregnancies complicated by isolated IUGR. *Medical science monitor : international medical journal of experimental and clinical research*. 2013; 19:118–24. [PubMed: 23416763]
108. Wei SQ, Audibert F, Luo ZC, Nuyt AM, Masse B, Julien P, et al. Maternal plasma 25-hydroxyvitamin D levels, angiogenic factors, and preeclampsia. *American journal of obstetrics and gynecology*. 2013; 208(5):390e1–6. [PubMed: 23618499]
109. Nasrollahi S, Hoseini Panah SM, Tavilani H, Tavasoli S, Naderan M, Shoar S. Antioxidant status and serum levels of selectins in pre-eclampsia. *Journal of obstetrics and gynaecology : the journal of the Institute of Obstetrics and Gynaecology*. 2015; 35(1):16–8. [PubMed: 25280210]
110. Tuzcu ZB, Ascioglu E, Sunbul M, Ozben B, Arikan H, Koc M. Circulating endothelial cell number and markers of endothelial dysfunction in previously preeclamptic women. *American journal of obstetrics and gynecology*. 2015
111. Ishihara N, Matsuo H, Murakoshi H, Laoag-Fernandez JB, Samoto T, Maruo T. Increased apoptosis in the syncytiotrophoblast in human term placentas complicated by either preeclampsia

- or intrauterine growth retardation. *American journal of obstetrics and gynecology*. 2002; 186(1): 158–66. [PubMed: 11810103]
112. Sibai BM, Ewell M, Levine RJ, Klebanoff MA, Esterlitz J, Catalano PM, et al. Risk factors associated with preeclampsia in healthy nulliparous women. The Calcium for Preeclampsia Prevention (CPEP) Study Group. *American journal of obstetrics and gynecology*. 1997; 177(5): 1003–10. [PubMed: 9396883]
 113. von Dadelszen P, Magee LA, Roberts JM. Subclassification of preeclampsia. *Hypertension in pregnancy*. 2003; 22(2):143–8. [PubMed: 12908998]
 114. Alexander GR, Himes JH, Kaufman RB, Mor J, Kogan M. A United States national reference for fetal growth. *Obstetrics and gynecology*. 1996; 87(2):163–8. [PubMed: 8559516]
 115. Cunningham FG, Morris GB, Mickal A. Acute pyelonephritis of pregnancy: A clinical review. *Obstetrics and gynecology*. 1973; 42(1):112–7. [PubMed: 4720190]
 116. Gilstrap LC 3rd, Cunningham FG, Whalley PJ. Acute pyelonephritis in pregnancy: an anterospective study. *Obstetrics and gynecology*. 1981; 57(4):409–13. [PubMed: 7243084]
 117. Kurmanavicius J, Florio I, Wissler J, Hebisch G, Zimmermann R, Muller R, et al. Reference resistance indices of the umbilical, fetal middle cerebral and uterine arteries at 24–42 weeks of gestation. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 1997; 10(2):112–20.
 118. Arduini D, Rizzo G. Normal values of Pulsatility Index from fetal vessels: a cross-sectional study on 1556 healthy fetuses. *Journal of perinatal medicine*. 1990; 18(3):165–72. [PubMed: 2200862]
 119. Trudinger BJ, Cook CM, Giles WB, Ng S, Fong E, Connelly A, et al. Fetal umbilical artery velocity waveforms and subsequent neonatal outcome. *British journal of obstetrics and gynaecology*. 1991; 98(4):378–84. [PubMed: 2031896]
 120. Deanfield JE, Halcox JP, Rabelink TJ. Endothelial function and dysfunction: testing and clinical relevance. *Circulation*. 2007; 115(10):1285–95. [PubMed: 17353456]
 121. Rizzo V, McIntosh DP, Oh P, Schnitzer JE. In situ flow activates endothelial nitric oxide synthase in luminal caveolae of endothelium with rapid caveolin dissociation and calmodulin association. *The Journal of biological chemistry*. 1998; 273(52):34724–9. [PubMed: 9856995]
 122. Garcia-Cardena G, Comander J, Anderson KR, Blackman BR, Gimbrone MA Jr. Biomechanical activation of vascular endothelium as a determinant of its functional phenotype. *Proceedings of the National Academy of Sciences of the United States of America*. 2001; 98(8):4478–85. [PubMed: 11296290]
 123. Koo A, Nordsletten D, Umeton R, Yankama B, Ayyadurai S, Garcia-Cardena G, et al. In silico modeling of shear-stress-induced nitric oxide production in endothelial cells through systems biology. *Biophysical journal*. 2013; 104(10):2295–306. [PubMed: 23708369]
 124. Bevilacqua MP, Pober JS, Majeau GR, Cotran RS, Gimbrone MA Jr. Interleukin 1 (IL-1) induces biosynthesis and cell surface expression of procoagulant activity in human vascular endothelial cells. *The Journal of experimental medicine*. 1984; 160(2):618–23. [PubMed: 6332168]
 125. Bevilacqua MP, Pober JS, Wheeler ME, Cotran RS, Gimbrone MA Jr. Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes, and related leukocyte cell lines. *The Journal of clinical investigation*. 1985; 76(5): 2003–11. [PubMed: 3877078]
 126. Bevilacqua MP, Pober JS, Mendrick DL, Cotran RS, Gimbrone MA Jr. Identification of an inducible endothelial-leukocyte adhesion molecule. *Proceedings of the National Academy of Sciences of the United States of America*. 1987; 84(24):9238–42. [PubMed: 2827173]
 127. Yong K, Khwaja A. Leucocyte cellular adhesion molecules. *Blood reviews*. 1990; 4(4):211–25. [PubMed: 1706206]
 128. Jutila MA. Leukocyte traffic to sites of inflammation. *APMIS : acta pathologica, microbiologica, et immunologica Scandinavica*. 1992; 100(3):191–201.
 129. Williams TJ, Hellewell PG. Endothelial cell biology. Adhesion molecules involved in the microvascular inflammatory response. *The American review of respiratory disease*. 1992; 146(5 Pt 2):S45–50. [PubMed: 1443907]

130. Albelda SM, Smith CW, Ward PA. Adhesion molecules and inflammatory injury. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 1994; 8(8):504–12. [PubMed: 8181668]
131. Panza JA, Quyyumi AA, Brush JE Jr, Epstein SE. Abnormal endothelium-dependent vascular relaxation in patients with essential hypertension. *The New England journal of medicine*. 1990; 323(1):22–7. [PubMed: 2355955]
132. Schleiffenbaum B, Spertini O, Tedder TF. Soluble L-selectin is present in human plasma at high levels and retains functional activity. *J Cell Biol*. 1992; 119(1):229–38. [PubMed: 1382078]
133. Xing K, Murthy S, Liles WC, Singh JM. Clinical utility of biomarkers of endothelial activation in sepsis--a systematic review. *Critical care*. 2012; 16(1):R7. [PubMed: 22248019]
134. Zonneveld R, Martinelli R, Shapiro NI, Kuijpers TW, Plotz FB, Carman CV. Soluble adhesion molecules as markers for sepsis and the potential pathophysiological discrepancy in neonates, children and adults. *Critical care*. 2014; 18(1):204. [PubMed: 24602331]
135. Nakae H, Endo S, Sato N, Wakabayashi G, Inada K, Sato S. Involvement of soluble adhesion molecules in acute pancreatitis. *European surgical research Europäische chirurgische Forschung Recherches chirurgicales europeennes*. 2001; 33(5–6):377–82. [PubMed: 11805399]
136. Powell JJ, Siriwardena AK, Fearon KC, Ross JA. Endothelial-derived selectins in the development of organ dysfunction in acute pancreatitis. *Critical care medicine*. 2001; 29(3):567–72. [PubMed: 11373422]
137. Shingu M, Hashimoto M, Nobunaga M, Isayama T, Yasutake C, Naono T. Production of soluble ICAM-1 by mononuclear cells from patients with rheumatoid arthritis patients. *Inflammation*. 1994; 18(1):23–34. [PubMed: 7911453]
138. Krenn V, Schedel J, Doring A, Huppertz HI, Gohlke F, Tony HP, et al. Endothelial cells are the major source of sICAM-1 in rheumatoid synovial tissue. *Rheumatology international*. 1997; 17(1):17–27. [PubMed: 9194210]
139. Volin MV. Soluble adhesion molecules in the pathogenesis of rheumatoid arthritis. *Current pharmaceutical design*. 2005; 11(5):633–53. [PubMed: 15720279]
140. Makrilia N, Kollias A, Manolopoulos L, Syrigos K. Cell adhesion molecules: role and clinical significance in cancer. *Cancer investigation*. 2009; 27(10):1023–37. [PubMed: 19909018]
141. van Kilsdonk JW, van Kempen LC, van Muijen GN, Ruitter DJ, Swart GW. Soluble adhesion molecules in human cancers: sources and fates. *European journal of cell biology*. 2010; 89(6):415–27. [PubMed: 20227133]
142. Romero R. The child is the father of the man. *Prenat Neonat Med*. 1996; 1:8–11.
143. Ness RB, Roberts JM. Heterogeneous causes constituting the single syndrome of preeclampsia: a hypothesis and its implications. *American journal of obstetrics and gynecology*. 1996; 175(5):1365–70. [PubMed: 8942516]
144. Broughton Pipkin F, Roberts JM. Hypertension in pregnancy. *Journal of human hypertension*. 2000; 14(10–11):705–24. [PubMed: 11095161]
145. Sibai BM. Preeclampsia: an inflammatory syndrome? *American journal of obstetrics and gynecology*. 2004; 191(4):1061–2. [PubMed: 15507921]
146. Sibai B, Dekker G, Kupferminc M. Pre-eclampsia. *Lancet*. 2005; 365(9461):785–99. [PubMed: 15733721]
147. Roberts JM, Gammill HS. Preeclampsia: recent insights. *Hypertension*. 2005; 46(6):1243–9. [PubMed: 16230510]
148. Redman CW, Sargent IL. Latest advances in understanding preeclampsia. *Science*. 2005; 308(5728):1592–4. [PubMed: 15947178]
149. Romero R. Prenatal medicine: the child is the father of the man. 1996. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2009; 22(8):636–9.
150. Di Renzo GC. The great obstetrical syndromes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2009; 22(8):633–5.

151. Chaiworapongsa T, Chaemsaitong P, Yeo L, Romero R. Pre-eclampsia part 1: current understanding of its pathophysiology. *Nature reviews Nephrology*. 2014; 10(8):466–80. [PubMed: 25003615]
152. Myatt L, Roberts JM. Preeclampsia: Syndrome or Disease? *Current hypertension reports*. 2015; 17(11):83. [PubMed: 26362531]
153. Robertson WB, Brosens I, Dixon G. Maternal uterine vascular lesions in the hypertensive complications of pregnancy. *Perspectives in nephrology and hypertension*. 1976; 5:115–27. [PubMed: 1005030]
154. Brosens IA. Morphological changes in the utero-placental bed in pregnancy hypertension. *Clinics in obstetrics and gynaecology*. 1977; 4(3):573–93. [PubMed: 598186]
155. Sheppard BL, Bonnar J. An ultrastructural study of utero-placental spiral arteries in hypertensive and normotensive pregnancy and fetal growth retardation. *British journal of obstetrics and gynaecology*. 1981; 88(7):695–705. [PubMed: 7248226]
156. Roberts JM, Taylor RN, Musci TJ, Rodgers GM, Hubel CA, McLaughlin MK. Preeclampsia: an endothelial cell disorder. *American journal of obstetrics and gynecology*. 1989; 161(5):1200–4. [PubMed: 2589440]
157. Harrington KF, Campbell S, Bewley S, Bower S. Doppler velocimetry studies of the uterine artery in the early prediction of pre-eclampsia and intra-uterine growth retardation. *European journal of obstetrics, gynecology, and reproductive biology*. 1991; 42(Suppl):S14–20.
158. Harrington K, Cooper D, Lees C, Hecher K, Campbell S. Doppler ultrasound of the uterine arteries: the importance of bilateral notching in the prediction of pre-eclampsia, placental abruption or delivery of a small-for-gestational-age baby. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 1996; 7(3):182–8.
159. Conrad KP, Benyo DF. Placental cytokines and the pathogenesis of preeclampsia. *American journal of reproductive immunology*. 1997; 37(3):240–9. [PubMed: 9127646]
160. Dekker GA, Sibai BM. Etiology and pathogenesis of preeclampsia: current concepts. *American journal of obstetrics and gynecology*. 1998; 179(5):1359–75. [PubMed: 9822529]
161. Roberts JM. Endothelial dysfunction in preeclampsia. *Seminars in reproductive endocrinology*. 1998; 16(1):5–15. [PubMed: 9654603]
162. Albaiges G, Missfelder-Lobos H, Lees C, Parra M, Nicolaides KH. One-stage screening for pregnancy complications by color Doppler assessment of the uterine arteries at 23 weeks' gestation. *Obstetrics and gynecology*. 2000; 96(4):559–64. [PubMed: 11004359]
163. Papageorghiou AT, Yu CK, Bindra R, Pandis G, Nicolaides KH. Fetal Medicine Foundation Second Trimester Screening G. Multicenter screening for pre-eclampsia and fetal growth restriction by transvaginal uterine artery Doppler at 23 weeks of gestation. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2001; 18(5):441–9.
164. Granger JP, Alexander BT, Llinas MT, Bennett WA, Khalil RA. Pathophysiology of preeclampsia: linking placental ischemia/hypoxia with microvascular dysfunction. *Microcirculation*. 2002; 9(3):147–60. [PubMed: 12080413]
165. Myatt L. Role of placenta in preeclampsia. *Endocrine*. 2002; 19(1):103–11. [PubMed: 12583607]
166. Kadyrov M, Schmitz C, Black S, Kaufmann P, Huppertz B. Pre-eclampsia and maternal anaemia display reduced apoptosis and opposite invasive phenotypes of extravillous trophoblast. *Placenta*. 2003; 24(5):540–8. [PubMed: 12744931]
167. Papageorghiou AT, Yu CK, Nicolaides KH. The role of uterine artery Doppler in predicting adverse pregnancy outcome. *Best practice & research Clinical obstetrics & gynaecology*. 2004; 18(3):383–96. [PubMed: 15183134]
168. Fisher SJ. The placental problem: linking abnormal cytotrophoblast differentiation to the maternal symptoms of preeclampsia. *Reproductive biology and endocrinology : RB&E*. 2004; 2:53. [PubMed: 15236649]
169. Gilbert JS, Ryan MJ, LaMarca BB, Sedeek M, Murphy SR, Granger JP. Pathophysiology of hypertension during preeclampsia: linking placental ischemia with endothelial dysfunction.

American journal of physiology Heart and circulatory physiology. 2008; 294(2):H541–50. [PubMed: 18055511]

170. Ogge G, Chaiworapongsa T, Romero R, Hussein Y, Kusanovic JP, Yeo L, et al. Placental lesions associated with maternal underperfusion are more frequent in early-onset than in late-onset preeclampsia. *Journal of perinatal medicine*. 2011; 39(6):641–52. [PubMed: 21848483]
171. Espinoza J. Recent biomarkers for the identification of patients at risk for preeclampsia: the role of uteroplacental ischemia. *Expert opinion on medical diagnostics*. 2012; 6(2):121–30. [PubMed: 23480655]
172. Espinoza J. Uteroplacental ischemia in early- and late-onset pre-eclampsia: a role for the fetus? *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2012; 40(4):373–82.
173. George EM, Granger JP. Linking placental ischemia and hypertension in preeclampsia: role of endothelin 1. *Hypertension*. 2012; 60(2):507–11. [PubMed: 22566502]
174. Roberts JM. Pathophysiology of ischemic placental disease. *Seminars in perinatology*. 2014; 38(3):139–45. [PubMed: 24836825]
175. Bosco C, Diaz E, Gutierrez R, Gonzalez J, Parra-Cordero M, Rodrigo R, et al. A putative role for telocytes in placental barrier impairment during preeclampsia. *Medical hypotheses*. 2015; 84(1):72–7. [PubMed: 25499002]
176. Maynard SE, Min JY, Merchan J, Lim KH, Li J, Mondal S, et al. Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *The Journal of clinical investigation*. 2003; 111(5):649–58. [PubMed: 12618519]
177. Chaiworapongsa T, Romero R, Espinoza J, Bujold E, Mee Kim Y, Goncalves LF, et al. Evidence supporting a role for blockade of the vascular endothelial growth factor system in the pathophysiology of preeclampsia. *Young Investigator Award. American journal of obstetrics and gynecology*. 2004; 190(6):1541–7. [PubMed: 15284729]
178. Levine RJ, Maynard SE, Qian C, Lim KH, England LJ, Yu KF, et al. Circulating angiogenic factors and the risk of preeclampsia. *The New England journal of medicine*. 2004; 350(7):672–83. [PubMed: 14764923]
179. Chaiworapongsa T, Romero R, Kim YM, Kim GJ, Kim MR, Espinoza J, et al. Plasma soluble vascular endothelial growth factor receptor-1 concentration is elevated prior to the clinical diagnosis of pre-eclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2005; 17(1):3–18.
180. Bujold E, Romero R, Chaiworapongsa T, Kim YM, Kim GJ, Kim MR, et al. Evidence supporting that the excess of the sVEGFR-1 concentration in maternal plasma in preeclampsia has a uterine origin. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2005; 18(1):9–16.
181. Espinoza J, Romero R, Nien JK, Kusanovic JP, Richani K, Gomez R, et al. A role of the anti-angiogenic factor sVEGFR-1 in the ‘mirror syndrome’ (Ballantyne’s syndrome). *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2006; 19(10):607–13.
182. Levine RJ, Lam C, Qian C, Yu KF, Maynard SE, Sachs BP, et al. Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. *The New England journal of medicine*. 2006; 355(10):992–1005. [PubMed: 16957146]
183. Gotsch F, Romero R, Friel L, Kusanovic JP, Espinoza J, Erez O, et al. CXCL10/IP-10: a missing link between inflammation and anti-angiogenesis in preeclampsia? *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2007; 20(11):777–92.
184. Robinson CJ, Johnson DD. Soluble endoglin as a second-trimester marker for preeclampsia. *American journal of obstetrics and gynecology*. 2007; 197(2):174e1–5. [PubMed: 17689640]

185. Schlembach D, Wallner W, Sengenberger R, Stiegler E, Mortl M, Beckmann MW, et al. Angiogenic growth factor levels in maternal and fetal blood: correlation with Doppler ultrasound parameters in pregnancies complicated by pre-eclampsia and intrauterine growth restriction. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2007; 29(4):407–13.
186. Staff AC, Braekke K, Johnsen GM, Karumanchi SA, Harsem NK. Circulating concentrations of soluble endoglin (CD105) in fetal and maternal serum and in amniotic fluid in preeclampsia. *American journal of obstetrics and gynecology*. 2007; 197(2):176e1–6. [PubMed: 17689641]
187. Stepan H, Kramer T, Faber R. Maternal plasma concentrations of soluble endoglin in pregnancies with intrauterine growth restriction. *The Journal of clinical endocrinology and metabolism*. 2007; 92(7):2831–4. [PubMed: 17426082]
188. Stepan H, Unversucht A, Wessel N, Faber R. Predictive value of maternal angiogenic factors in second trimester pregnancies with abnormal uterine perfusion. *Hypertension*. 2007; 49(4):818–24. [PubMed: 17261644]
189. Vatten LJ, Eskild A, Nilsen TI, Jeansson S, Jenum PA, Staff AC. Changes in circulating level of angiogenic factors from the first to second trimester as predictors of preeclampsia. *American journal of obstetrics and gynecology*. 2007; 196(3):239e1–6. [PubMed: 17346536]
190. Wallner W, Sengenberger R, Strick R, Strissel PL, Meurer B, Beckmann MW, et al. Angiogenic growth factors in maternal and fetal serum in pregnancies complicated by intrauterine growth restriction. *Clinical science*. 2007; 112(1):51–7. [PubMed: 16928195]
191. Wikstrom AK, Larsson A, Eriksson UJ, Nash P, Norden-Lindeberg S, Olovsson M. Placental growth factor and soluble FMS-like tyrosine kinase-1 in early-onset and late-onset preeclampsia. *Obstetrics and gynecology*. 2007; 109(6):1368–74. [PubMed: 17540809]
192. Baumann MU, Bersinger NA, Mohaupt MG, Raio L, Gerber S, Surbek DV. First-trimester serum levels of soluble endoglin and soluble fms-like tyrosine kinase-1 as first-trimester markers for late-onset preeclampsia. *American journal of obstetrics and gynecology*. 2008; 199(3):266e1–6. [PubMed: 18771978]
193. Romero R, Nien JK, Espinoza J, Todem D, Fu W, Chung H, et al. A longitudinal study of angiogenic (placental growth factor) and anti-angiogenic (soluble endoglin and soluble vascular endothelial growth factor receptor-1) factors in normal pregnancy and patients destined to develop preeclampsia and deliver a small for gestational age neonate. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2008; 21(1):9–23.
194. Chaiworapongsa T, Romero R, Gotsch F, Espinoza J, Nien JK, Goncalves L, et al. Low maternal concentrations of soluble vascular endothelial growth factor receptor-2 in preeclampsia and small for gestational age. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2008; 21(1):41–52.
195. Gotsch F, Romero R, Kusanovic JP, Chaiworapongsa T, Dombrowski M, Erez O, et al. Preeclampsia and small-for-gestational age are associated with decreased concentrations of a factor involved in angiogenesis: soluble Tie-2. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2008; 21(6):389–402.
196. Erez O, Romero R, Espinoza J, Fu W, Todem D, Kusanovic JP, et al. The change in concentrations of angiogenic and anti-angiogenic factors in maternal plasma between the first and second trimesters in risk assessment for the subsequent development of preeclampsia and small-for-gestational age. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2008; 21(5):279–87.
197. Crispi F, Llubra E, Dominguez C, Martin-Gallan P, Cabero L, Gratacos E. Predictive value of angiogenic factors and uterine artery Doppler for early- versus late-onset pre-eclampsia and intrauterine growth restriction. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2008; 31(3):303–9.

198. De Vivo A, Baviera G, Giordano D, Todarello G, Corrado F, D'Anna R. Endoglin, PlGF and sFlt-1 as markers for predicting pre-eclampsia. *Acta obstetrica et gynecologica Scandinavica*. 2008; 87(8):837–42. [PubMed: 18607829]
199. Yinon Y, Nevo O, Xu J, Many A, Rolfo A, Todros T, et al. Severe intrauterine growth restriction pregnancies have increased placental endoglin levels: hypoxic regulation via transforming growth factor-beta 3. *The American journal of pathology*. 2008; 172(1):77–85. [PubMed: 18156205]
200. Kusanovic JP, Romero R, Chaiworapongsa T, Erez O, Mittal P, Vaisbuch E, et al. A prospective cohort study of the value of maternal plasma concentrations of angiogenic and anti-angiogenic factors in early pregnancy and midtrimester in the identification of patients destined to develop preeclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2009; 22(11):1021–38.
201. Chedraui P, Lockwood CJ, Schatz F, Buchwalder LF, Schwager G, Guerrero C, et al. Increased plasma soluble fms-like tyrosine kinase 1 and endoglin levels in pregnancies complicated with preeclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2009; 22(7):565–70.
202. Chaiworapongsa T, Romero R, Kusanovic JP, Mittal P, Kim SK, Gotsch F, et al. Plasma soluble endoglin concentration in pre-eclampsia is associated with an increased impedance to flow in the maternal and fetal circulations. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2010; 35(2):155–62.
203. Chaiworapongsa T, Romero R, Tarca AL, Kusanovic JP, Gotsch F, Mittal P, et al. A decrease in maternal plasma concentrations of sVEGFR-2 precedes the clinical diagnosis of preeclampsia. *American journal of obstetrics and gynecology*. 2010; 202(6):550e1–10. [PubMed: 20510958]
204. Chaiworapongsa T, Romero R, Savasan ZA, Kusanovic JP, Ogge G, Soto E, et al. Maternal plasma concentrations of angiogenic/anti-angiogenic factors are of prognostic value in patients presenting to the obstetrical triage area with the suspicion of preeclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2011; 24(10):1187–207.
205. Soto E, Romero R, Kusanovic JP, Ogge G, Hussein Y, Yeo L, et al. Late-onset preeclampsia is associated with an imbalance of angiogenic and anti-angiogenic factors in patients with and without placental lesions consistent with maternal underperfusion. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2012; 25(5):498–507.
206. Romero R, Chaiworapongsa T. Preeclampsia: a link between trophoblast dysregulation and an antiangiogenic state. *The Journal of clinical investigation*. 2013; 123(7):2775–7. [PubMed: 23934119]
207. Stampalija T, Chaiworapongsa T, Romero R, Chaemsaitong P, Korzeniewski SJ, Schwartz AG, et al. Maternal plasma concentrations of sST2 and angiogenic/anti-angiogenic factors in preeclampsia. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2013; 26(14):1359–70.
208. Szabo S, Xu Y, Romero R, Fule T, Karaszi K, Bhatti G, et al. Changes of placental syndecan-1 expression in preeclampsia and HELLP syndrome. *Virchows Archiv : an international journal of pathology*. 2013; 463(3):445–58. [PubMed: 23807541]
209. Chaiworapongsa T, Romero R, Korzeniewski SJ, Cortez JM, Pappas A, Tarca AL, et al. Plasma concentrations of angiogenic/anti-angiogenic factors have prognostic value in women presenting with suspected preeclampsia to the obstetrical triage area: a prospective study. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2014; 27(2):132–44.
210. Chaiworapongsa T, Romero R, Whitten AE, Korzeniewski SJ, Chaemsaitong P, Hernandez-Andrade E, et al. The use of angiogenic biomarkers in maternal blood to identify which SGA

fetuses will require a preterm delivery and mothers who will develop pre-eclampsia. The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet. 2015:1–15.

211. Meeme A, Buga GA, Mammen M, Namugowa AV. Angiogenic imbalance as a contributor to the pathophysiology of preeclampsia among black African women. The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet. 2016:1–7.
212. Cunningham FG, Pritchard JA. Hematologic considerations of pregnancy-induced hypertension. *Seminars in perinatology*. 1978; 2(1):29–38. [PubMed: 734445]
213. Weenink GH, Treffers PE, Vijn P, Smorenberg-School ME, Ten Cate JW. Antithrombin III levels in preeclampsia correlate with maternal and fetal morbidity. *American journal of obstetrics and gynecology*. 1984; 148(8):1092–7. [PubMed: 6711644]
214. Romero R, Mazor M, Lockwood CJ, Emamian M, Belanger KP, Hobbins JC, et al. Clinical significance, prevalence, and natural history of thrombocytopenia in pregnancy-induced hypertension. *American journal of perinatology*. 1989; 6(1):32–8. [PubMed: 2783368]
215. de Boer K, ten Cate JW, Sturk A, Borm JJ, Treffers PE. Enhanced thrombin generation in normal and hypertensive pregnancy. *American journal of obstetrics and gynecology*. 1989; 160(1):95–100. [PubMed: 2521425]
216. Cadroy Y, Grandjean H, Pichon J, Desprats R, Berrebi A, Fournie A, et al. Evaluation of six markers of haemostatic system in normal pregnancy and pregnancy complicated by hypertension or pre-eclampsia. *British journal of obstetrics and gynaecology*. 1993; 100(5):416–20. [PubMed: 8518239]
217. Chaiworapongsa T, Yoshimatsu J, Espinoza J, Kim YM, Berman S, Edwin S, et al. Evidence of in vivo generation of thrombin in patients with small-for-gestational-age fetuses and pre-eclampsia. The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet. 2002; 11(6):362–7.
218. Dekker G. Prothrombotic mechanisms in preeclampsia. *Thrombosis research*. 2005; 115(Suppl 1):17–21. [PubMed: 15790144]
219. Erez O, Romero R, Kim SS, Kim JS, Kim YM, Wildman DE, et al. Over-expression of the thrombin receptor (PAR-1) in the placenta in preeclampsia: a mechanism for the intersection of coagulation and inflammation. The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet. 2008; 21(6):345–55.
220. Erez O, Romero R, Hoppensteadt D, Than NG, Fareed J, Mazaki-Tovi S, et al. Tissue factor and its natural inhibitor in pre-eclampsia and SGA. The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet. 2008; 21(12):855–69.
221. Kenny LC, Baker PN, Cunningham, FG. Platelets, coagulation, and the liver. In: Lindheimer, MD, Roberts, JM, Cunningham, GC, editors. *Chesley's hypertensive disorders of pregnancy*. San Diego: Elsevier; 2009. 335–51.
222. Socol ML, Weiner CP, Louis G, Rehnberg K, Rossi EC. Platelet activation in preeclampsia. *American journal of obstetrics and gynecology*. 1985; 151(4):494–7. [PubMed: 3156500]
223. Romero R, Lockwood C, Oyarzun E, Hobbins JC. Toxemia: new concepts in an old disease. *Seminars in perinatology*. 1988; 12(4):302–23. [PubMed: 3065943]
224. Csaicsich P, Deutinger J, Tatra G. Platelet specific proteins (beta-thromboglobulin and platelet factor 4) in normal pregnancy and in pregnancy complicated by preeclampsia. *Archives of gynecology and obstetrics*. 1989; 244(2):91–5. [PubMed: 2523691]
225. Ahmed Y, van Iddekinge B, Paul C, Sullivan HF, Elder MG. Retrospective analysis of platelet numbers and volumes in normal pregnancy and in pre-eclampsia. *British journal of obstetrics and gynaecology*. 1993; 100(3):216–20. [PubMed: 8476825]

226. Major HD, Campbell RA, Silver RM, Branch DW, Weyrich AS. Synthesis of sFlt-1 by platelet-monocyte aggregates contributes to the pathogenesis of preeclampsia. *American journal of obstetrics and gynecology*. 2014; 210(6):547e1–7. [PubMed: 24440566]
227. Roberts JM, Taylor RN, Goldfien A. Clinical and biochemical evidence of endothelial cell dysfunction in the pregnancy syndrome preeclampsia. *American journal of hypertension*. 1991; 4(8):700–8. [PubMed: 1930853]
228. Clark BA, Halvorson L, Sachs B, Epstein FH. Plasma endothelin levels in preeclampsia: elevation and correlation with uric acid levels and renal impairment. *American journal of obstetrics and gynecology*. 1992; 166(3):962–8. [PubMed: 1532292]
229. Friedman SA, Schiff E, Emeis JJ, Dekker GA, Sibai BM. Biochemical corroboration of endothelial involvement in severe preeclampsia. *American journal of obstetrics and gynecology*. 1995; 172(1 Pt 1):202–3. [PubMed: 7847535]
230. Lyall F, Greer IA. The vascular endothelium in normal pregnancy and pre-eclampsia. *Reviews of reproduction*. 1996; 1(2):107–16. [PubMed: 9414447]
231. Taylor RN, de Groot CJ, Cho YK, Lim KH. Circulating factors as markers and mediators of endothelial cell dysfunction in preeclampsia. *Seminars in reproductive endocrinology*. 1998; 16(1):17–31. [PubMed: 9654604]
232. Cindrova-Davies T. Gabor Than Award Lecture 2008: pre-eclampsia - from placental oxidative stress to maternal endothelial dysfunction. *Placenta*. 2009; 30(Suppl A):S55–65. [PubMed: 19118896]
233. Lamarca B. Endothelial dysfunction. An important mediator in the pathophysiology of hypertension during pre-eclampsia. *Minerva ginecologica*. 2012; 64(4):309–20. [PubMed: 22728575]
234. Sandvik MK, Leirgul E, Nygard O, Ueland PM, Berg A, Svarstad E, et al. Preeclampsia in healthy women and endothelial dysfunction 10 years later. *American journal of obstetrics and gynecology*. 2013; 209(6):569e1–e10. [PubMed: 23899451]
235. Torrado J, Farro I, Zocalo Y, Farro F, Sosa C, Scasso S, et al. Preeclampsia Is Associated with Increased Central Aortic Pressure, Elastic Arteries Stiffness and Wave Reflections, and Resting and Recrutable Endothelial Dysfunction. *International journal of hypertension*. 2015; 2015:720683. [PubMed: 26351578]
236. Vinayagam V, Bobby Z, Habeebullah S, Chaturvedula L, Bharadwaj SK. Plasma markers of endothelial dysfunction in patients with hypertensive disorders of pregnancy: a pilot study in a South Indian population. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2015:1–6.
237. Damsky CH, Fitzgerald ML, Fisher SJ. Distribution patterns of extracellular matrix components and adhesion receptors are intricately modulated during first trimester cytotrophoblast differentiation along the invasive pathway, in vivo. *The Journal of clinical investigation*. 1992; 89(1):210–22. [PubMed: 1370295]
238. Labarrere CA, Faulk WP. Intercellular adhesion molecule-1 (ICAM-1) and HLA-DR antigens are expressed on endovascular cytotrophoblasts in abnormal pregnancies. *American journal of reproductive immunology*. 1995; 33(1):47–53. [PubMed: 7619233]
239. Gurtner GC, Davis V, Li H, McCoy MJ, Sharpe A, Cybulsky MI. Targeted disruption of the murine VCAM1 gene: essential role of VCAM-1 in chorioallantoic fusion and placentation. *Genes & development*. 1995; 9(1):1–14. [PubMed: 7530222]
240. Zhou Y, Genbacev O, Fisher SJ. The human placenta remodels the uterus by using a combination of molecules that govern vasculogenesis or leukocyte extravasation. *Annals of the New York Academy of Sciences*. 2003; 995:73–83. [PubMed: 12814940]
241. Liu Q, Yan X, Li Y, Zhang Y, Zhao X, Shen Y. Pre-eclampsia is associated with the failure of melanoma cell adhesion molecule (MCAM/CD146) expression by intermediate trophoblast. *Laboratory investigation; a journal of technical methods and pathology*. 2004; 84(2):221–8. [PubMed: 14688802]

242. Brown LM, Lacey HA, Baker PN, Crocker IP. E-cadherin in the assessment of aberrant placental cytotrophoblast turnover in pregnancies complicated by pre-eclampsia. *Histochemistry and cell biology*. 2005; 124(6):499–506. [PubMed: 16142450]
243. Cartwright JE, Balarajah G. Trophoblast interactions with endothelial cells are increased by interleukin-1beta and tumour necrosis factor alpha and involve vascular cell adhesion molecule-1 and alpha4beta1. *Experimental cell research*. 2005; 304(1):328–36. [PubMed: 15707597]
244. Lyall F. Mechanisms regulating cytotrophoblast invasion in normal pregnancy and pre-eclampsia. *The Australian & New Zealand journal of obstetrics & gynaecology*. 2006; 46(4):266–73. [PubMed: 16866784]
245. Blechschmidt K, Mylonas I, Mayr D, Schiessl B, Schulze S, Becker KF, et al. Expression of E-cadherin and its repressor snail in placental tissue of normal, preeclamptic and HELLP pregnancies. *Virchows Archiv : an international journal of pathology*. 2007; 450(2):195–202. [PubMed: 17149611]
246. McEwan M, Lins RJ, Munro SK, Vincent ZL, Ponnampalam AP, Mitchell MD. Cytokine regulation during the formation of the fetal-maternal interface: focus on cell-cell adhesion and remodelling of the extra-cellular matrix. *Cytokine & growth factor reviews*. 2009; 20(3):241–9. [PubMed: 19487153]
247. Zhou Y, Fisher SJ, Janatpour M, Genbacev O, Dejana E, Wheelock M, et al. Human cytotrophoblasts adopt a vascular phenotype as they differentiate. A strategy for successful endovascular invasion? *The Journal of clinical investigation*. 1997; 99(9):2139–51. [PubMed: 9151786]
248. Zhou Y, Damsky CH, Fisher SJ. Preeclampsia is associated with failure of human cytotrophoblasts to mimic a vascular adhesion phenotype. One cause of defective endovascular invasion in this syndrome? *The Journal of clinical investigation*. 1997; 99(9):2152–64. [PubMed: 9151787]
249. McMaster MT, Zhou Y, Fisher SJ. Abnormal placentation and the syndrome of preeclampsia. *Seminars in nephrology*. 2004; 24(6):540–7. [PubMed: 15529288]
250. Redman CW, Sargent IL. Placental debris, oxidative stress and pre-eclampsia. *Placenta*. 2000; 21(7):597–602. [PubMed: 10985960]
251. Gerretsen G, Huisjes HJ, Elema JD. Morphological changes of the spiral arteries in the placental bed in relation to pre-eclampsia and fetal growth retardation. *British journal of obstetrics and gynaecology*. 1981; 88(9):876–81. [PubMed: 7272259]
252. Hustin J, Foidart JM, Lambotte R. Maternal vascular lesions in pre-eclampsia and intrauterine growth retardation: light microscopy and immunofluorescence. *Placenta*. 1983; 4(Spec No):489–98. [PubMed: 6369298]
253. Labarrere C, Alonso J, Manni J, Domenichini E, Althabe O. Immunohistochemical findings in acute atherosclerosis associated with intrauterine growth retardation. *American journal of reproductive immunology and microbiology : AJRIM*. 1985; 7(4):149–55. [PubMed: 3893171]
254. Khong TY, De Wolf F, Robertson WB, Brosens I. Inadequate maternal vascular response to placentation in pregnancies complicated by pre-eclampsia and by small-for-gestational age infants. *British journal of obstetrics and gynaecology*. 1986; 93(10):1049–59. [PubMed: 3790464]
255. Pijnenborg R, Anthony J, Davey DA, Rees A, Tiltman A, Vercruyse L, et al. Placental bed spiral arteries in the hypertensive disorders of pregnancy. *British journal of obstetrics and gynaecology*. 1991; 98(7):648–55. [PubMed: 1883787]
256. Meekins JW, Pijnenborg R, Hanssens M, McFadyen IR, van Asshe A. A study of placental bed spiral arteries and trophoblast invasion in normal and severe pre-eclamptic pregnancies. *British journal of obstetrics and gynaecology*. 1994; 101(8):669–74. [PubMed: 7947500]
257. Brosens JJ, Pijnenborg R, Brosens IA. The myometrial junctional zone spiral arteries in normal and abnormal pregnancies: a review of the literature. *American journal of obstetrics and gynecology*. 2002; 187(5):1416–23. [PubMed: 12439541]
258. Madazli R, Benian A, Ilvan S, Calay Z. Placental apoptosis and adhesion molecules expression in the placenta and the maternal placental bed of pregnancies complicated by fetal growth

- restriction with and without pre-eclampsia. *Journal of obstetrics and gynaecology : the journal of the Institute of Obstetrics and Gynaecology*. 2006; 26(1):5–10. [PubMed: 16390700]
259. Brosens I, Pijnenborg R, Vercruyssen L, Romero R. The “Great Obstetrical Syndromes” are associated with disorders of deep placentation. *American journal of obstetrics and gynecology*. 2011; 204(3):193–201. [PubMed: 21094932]
260. Kim YM, Chaemsaihong P, Romero R, Shaman M, Kim CJ, Kim JS, et al. The frequency of acute atherosclerosis in normal pregnancy and preterm labor, preeclampsia, small-for-gestational age, fetal death and midtrimester spontaneous abortion. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2015:1–9.
261. Birdir C, Fryze J, Frolich S, Schmidt M, Koninger A, Kimmig R, et al. Impact of maternal serum levels of Visfatin, AFP, PAPP-A, sFlt-1 and PlGF at 11–13 weeks gestation on small for gestational age births. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2017; 30(6):629–34.
262. Missfelder-Lobos H, Teran E, Lees C, Albaiges G, Nicolaides KH. Platelet changes and subsequent development of pre-eclampsia and fetal growth restriction in women with abnormal uterine artery Doppler screening. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2002; 19(5):443–8.
263. Hubel CA. Oxidative stress in the pathogenesis of preeclampsia. *Proceedings of the Society for Experimental Biology and Medicine Society for Experimental Biology and Medicine*. 1999; 222(3):222–35.
264. Myatt L, Kossenjans W, Sahay R, Eis A, Brockman D. Oxidative stress causes vascular dysfunction in the placenta. *The Journal of maternal-fetal medicine*. 2000; 9(1):79–82. [PubMed: 10757441]
265. Madazli R, Benian A, Aydin S, Uzun H, Tolun N. The plasma and placental levels of malondialdehyde, glutathione and superoxide dismutase in pre-eclampsia. *Journal of obstetrics and gynaecology : the journal of the Institute of Obstetrics and Gynaecology*. 2002; 22(5):477–80. [PubMed: 12521411]
266. Vaughan JE, Walsh SW. Oxidative stress reproduces placental abnormalities of preeclampsia. *Hypertension in pregnancy*. 2002; 21(3):205–23. [PubMed: 12517328]
267. Burton GJ, Yung HW, Cindrova-Davies T, Charnock-Jones DS. Placental endoplasmic reticulum stress and oxidative stress in the pathophysiology of unexplained intrauterine growth restriction and early onset preeclampsia. *Placenta*. 2009; 30(Suppl A):S43–8. [PubMed: 19081132]
268. Potdar N, Singh R, Mistry V, Evans MD, Farmer PB, Konje JC, et al. First-trimester increase in oxidative stress and risk of small-for-gestational-age fetus. *BJOG : an international journal of obstetrics and gynaecology*. 2009; 116(5):637–42. [PubMed: 19298438]
269. Zhou X, Zhang GY, Wang J, Lu SL, Cao J, Sun LZ. A novel bridge between oxidative stress and immunity: the interaction between hydrogen peroxide and human leukocyte antigen G in placental trophoblasts during preeclampsia. *American journal of obstetrics and gynecology*. 2012; 206(5):447e7–16.
270. Friedman SA, de Groot CJ, Taylor RN, Golditch BD, Roberts JM. Plasma cellular fibronectin as a measure of endothelial involvement in preeclampsia and intrauterine growth retardation. *American journal of obstetrics and gynecology*. 1994; 170(3):838–41. [PubMed: 8141213]
271. Gilstrap LG 3rd, Hankins GD, Snyder RR, Greenberg RT. Acute pyelonephritis in pregnancy. *Comprehensive therapy*. 1986; 12(12):38–42.
272. Mabie WC, Barton JR, Sibai B. Septic shock in pregnancy. *Obstetrics and gynecology*. 1997; 90(4 Pt 1):553–61. [PubMed: 9380315]
273. Foxman B, Klemstine KL, Brown PD. Acute pyelonephritis in US hospitals in 1997: hospitalization and in-hospital mortality. *Annals of epidemiology*. 2003; 13(2):144–50. [PubMed: 12559674]
274. Barton JR, Sibai BM. Severe sepsis and septic shock in pregnancy. *Obstetrics and gynecology*. 2012; 120(3):689–706. [PubMed: 22914482]

275. Morgan J, Roberts S. Maternal sepsis. *Obstetrics and gynecology clinics of North America*. 2013; 40(1):69–87. [PubMed: 23466138]
276. Wing DA, Fassett MJ, Getahun D. Acute pyelonephritis in pregnancy: an 18-year retrospective analysis. *American journal of obstetrics and gynecology*. 2014; 210(3):219e1–6. [PubMed: 24100227]
277. Chaiworapongsa T, Romero R, Gotsch F, Kusanovic JP, Mittal P, Kim SK, et al. Acute pyelonephritis during pregnancy changes the balance of angiogenic and anti-angiogenic factors in maternal plasma. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2010; 23(2):167–78.
278. Gotsch F, Romero R, Espinoza J, Kusanovic JP, Mazaki-Tovi S, Erez O, et al. Maternal serum concentrations of the chemokine CXCL10/IP-10 are elevated in acute pyelonephritis during pregnancy. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2007; 20(10):735–44.
279. Soto E, Romero R, Vaisbuch E, Erez O, Mazaki-Tovi S, Kusanovic JP, et al. Fragment Bb: evidence for activation of the alternative pathway of the complement system in pregnant women with acute pyelonephritis. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2010; 23(10):1085–90.
280. Madan I, Than NG, Romero R, Chaemsaihong P, Miranda J, Tarca AL, et al. The peripheral whole-blood transcriptome of acute pyelonephritis in human pregnancy. *Journal of perinatal medicine*. 2014; 42(1):31–53. [PubMed: 24293448]
281. Chaiworapongsa T, Romero R, Whitten A, Tarca AL, Bhatti G, Draghici S, et al. Differences and similarities in the transcriptional profile of peripheral whole blood in early and late-onset preeclampsia: insights into the molecular basis of the phenotype of preeclampsia. *Journal of perinatal medicine*. 2013; 41(5):485–504. [PubMed: 23793063]
282. Yokoo A, Hirose T, Matsukawa M, Hotta H, Kunishima Y, Takahashi S. Expression of intercellular adhesion molecule-1 in mice with Pseudomonas-induced pyelonephritis. *The Journal of urology*. 1998; 160(2):592–6. [PubMed: 9679934]
283. Rui-Mei L, Kara AU, Sinniah R. In situ analysis of adhesion molecule expression in kidneys infected with murine malaria. *The Journal of pathology*. 1998; 185(2):219–25. [PubMed: 9713351]
284. Bishop GA, Hall BM. Expression of leucocyte and lymphocyte adhesion molecules in the human kidney. *Kidney international*. 1989; 36(6):1078–85. [PubMed: 2481060]
285. Lhotta K, Neumayer HP, Joannidis M, Geissler D, Konig P. Renal expression of intercellular adhesion molecule-1 in different forms of glomerulonephritis. *Clinical science*. 1991; 81(4):477–81. [PubMed: 1682080]
286. Brady HR. Leukocyte adhesion molecules and kidney diseases. *Kidney international*. 1994; 45(5):1285–300. [PubMed: 8072240]
287. Cowley HC, Heney D, Gearing AJ, Hemingway I, Webster NR. Increased circulating adhesion molecule concentrations in patients with the systemic inflammatory response syndrome: a prospective cohort study. *Critical care medicine*. 1994; 22(4):651–7. [PubMed: 7511496]
288. Endo S, Inada K, Kasai T, Takakuwa T, Yamada Y, Koike S, et al. Levels of soluble adhesion molecules and cytokines in patients with septic multiple organ failure. *Journal of inflammation*. 1995; 46(4):212–9. [PubMed: 8878795]
289. Sessler CN, Windsor AC, Schwartz M, Watson L, Fisher BJ, Sugerman HJ, et al. Circulating ICAM-1 is increased in septic shock. *American journal of respiratory and critical care medicine*. 1995; 151(5):1420–7. [PubMed: 7735595]
290. Boldt J, Muller M, Kuhn D, Linke LC, Hempelmann G. Circulating adhesion molecules in the critically ill: a comparison between trauma and sepsis patients. *Intensive care medicine*. 1996; 22(2):122–8. [PubMed: 8857119]

291. Austgulen R, Arntzen KJ, Haereid PE, Aag S, Dollner H. Infections in neonates delivered at term are associated with increased serum levels of ICAM-1 and E-selectin. *Acta paediatrica*. 1997; 86(3):274–80. [PubMed: 9099317]
292. Kayal S, Jais JP, Aguiñ N, Chaudiere J, Labrousse J. Elevated circulating E-selectin, intercellular adhesion molecule 1, and von Willebrand factor in patients with severe infection. *American journal of respiratory and critical care medicine*. 1998; 157(3 Pt 1):776–84. [PubMed: 9517590]
293. Baines PB, Marzouk O, Thomson AP, Sills JA, Riordan FA, Hart CA. Endothelial cell adhesion molecules in meningococcal disease. *Archives of disease in childhood*. 1999; 80(1):74–6. [PubMed: 10325765]
294. Weigand MA, Schmidt H, Pourmahmoud M, Zhao Q, Martin E, Bardenheuer HJ. Circulating intercellular adhesion molecule-1 as an early predictor of hepatic failure in patients with septic shock. *Critical care medicine*. 1999; 27(12):2656–61. [PubMed: 10628605]
295. Whalen MJ, Doughty LA, Carlos TM, Wisniewski SR, Kochanek PM, Carcillo JA. Intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 are increased in the plasma of children with sepsis-induced multiple organ failure. *Critical care medicine*. 2000; 28(7):2600–7. [PubMed: 10921602]
296. Gbadegesin RA, Cotton SA, Coupes BM, Awan A, Brenchley PE, Webb NJ. Plasma and urinary soluble adhesion molecule expression is increased during first documented acute pyelonephritis. *Archives of disease in childhood*. 2002; 86(3):218–21. [PubMed: 11861252]
297. Figueras-Aloy J, Gomez-Lopez L, Rodriguez-Miguel JM, Salvia-Roiges MD, Jordan-Garcia I, Ferrer-Codina I, et al. Serum soluble ICAM-1, VCAM-1, L-selectin, and P-selectin levels as markers of infection and their relation to clinical severity in neonatal sepsis. *American journal of perinatology*. 2007; 24(6):331–8. [PubMed: 17564956]
298. Shapiro NI, Schuetz P, Yano K, Sorasaki M, Parikh SM, Jones AE, et al. The association of endothelial cell signaling, severity of illness, and organ dysfunction in sepsis. *Critical care*. 2010; 14(5):R182. [PubMed: 20942957]
299. Schuetz P, Jones AE, Aird WC, Shapiro NI. Endothelial cell activation in emergency department patients with sepsis-related and non-sepsis-related hypotension. *Shock*. 2011; 36(2):104–8. [PubMed: 21522043]
300. Loukovaara M, Ylikorkala O. Serum nitric oxide metabolites and E-selectin in preterm premature rupture of membranes. *Acta obstetrica et gynecologica Scandinavica*. 2003; 82(7):616–9. [PubMed: 12790842]
301. Zou L, Zhang H, Zhu J. The value of the soluble intercellular adhesion molecule-1 levels in maternal serum for determination of occult chorioamnionitis in premature rupture of membranes. *J Huazhong Univ Sci Technolog Med Sci*. 2004; 24(2):154–7. [PubMed: 15315168]
302. Laudanski P, Raba G, Kuc P, Lemancewicz A, Kisielewski R, Laudanski T. Assessment of the selected biochemical markers in predicting preterm labour. *J Matern Fetal Neonatal Med*. 2012; 25(12):2696–9.
303. Bartha JL, Fernandez-Deudero A, Bugatto F, Fajardo-Exposito MA, Gonzalez-Gonzalez N, Hervias-Vivancos B. Inflammation and cardiovascular risk in women with preterm labor. *J Womens Health (Larchmt)*. 2012; 21(6):643–8. [PubMed: 22401498]
304. Chen X, Scholl TO. Maternal biomarkers of endothelial dysfunction and preterm delivery. *PLoS One*. 2014; 9(1):e85716. [PubMed: 24465662]
305. Saleh AA, Gerbasi FR, Mammen EF, Farag A. Increased platelet activation in preterm labor. *Thrombosis research*. 1992; 65(3):475–7. [PubMed: 1385910]
306. Erez O, Romero R, Hoppensteadt D, Fareed J, Chaiworapongsa T, Kusanovic JP, et al. Premature labor: a state of platelet activation? *Journal of perinatal medicine*. 2008; 36(5):377–87. [PubMed: 18958919]
307. Smith GD, Whitley E, Gissler M, Hemminki E. Birth dimensions of offspring, premature birth, and the mortality of mothers. *Lancet*. 2000; 356(9247):2066–7. [PubMed: 11145495]
308. Smith GC, Pell JP, Walsh D. Pregnancy complications and maternal risk of ischaemic heart disease: a retrospective cohort study of 129,290 births. *Lancet*. 2001; 357(9273):2002–6. [PubMed: 11438131]

309. Pell JP, Smith GC, Walsh D. Pregnancy complications and subsequent maternal cerebrovascular events: a retrospective cohort study of 119,668 births. *Am J Epidemiol.* 2004; 159(4):336–42. [PubMed: 14769636]
310. Catov JM, Newman AB, Roberts JM, Kelsey SF, Sutton-Tyrrell K, Harris TB, et al. Preterm delivery and later maternal cardiovascular disease risk. *Epidemiology.* 2007; 18(6):733–9. [PubMed: 17917602]
311. Bonamy AK, Parikh NI, Cnattingius S, Ludvigsson JF, Ingelsson E. Birth characteristics and subsequent risks of maternal cardiovascular disease: effects of gestational age and fetal growth. *Circulation.* 2011; 124(25):2839–46. [PubMed: 22124377]
312. Hastie CE, Smith GC, Mackay DF, Pell JP. Maternal risk of ischaemic heart disease following elective and spontaneous pre-term delivery: retrospective cohort study of 750 350 singleton pregnancies. *Int J Epidemiol.* 2011; 40(4):914–9. [PubMed: 21278195]
313. Erez O, Gotsch F, Mazaki-Tovi S, Vaisbuch E, Kusanovic JP, Kim CJ, et al. Evidence of maternal platelet activation, excessive thrombin generation, and high amniotic fluid tissue factor immunoreactivity and functional activity in patients with fetal death. *J Matern Fetal Neonat.* 2009; 22(8):672–87.
314. Ay C, Kaider A, Koder S, Husslein P, Pabinger I. Association of elevated soluble P-selectin levels with fetal loss in women with a history of venous thromboembolism. *Thrombosis research.* 2012; 129(6):725–8. [PubMed: 22169504]
315. Wagner DD. New links between inflammation and thrombosis. *Arteriosclerosis, thrombosis, and vascular biology.* 2005; 25(7):1321–4.
316. Ghasemzadeh M, Hosseini E. Platelet-leukocyte crosstalk: Linking proinflammatory responses to procoagulant state. *Thrombosis research.* 2013; 131(3):191–7. [PubMed: 23260445]
317. Ward PA. The dark side of C5a in sepsis. *Nature reviews Immunology.* 2004; 4(2):133–42.
318. Fernandez HN, Hugli TE. Primary structural analysis of the polypeptide portion of human C5a anaphylatoxin. Polypeptide sequence determination and assignment of the oligosaccharide attachment site in C5a. *The Journal of biological chemistry.* 1978; 253(19):6955–64. [PubMed: 690134]
319. Richani K, Romero R, Soto E, Espinoza J, Nien JK, Chaiworapongsa T, et al. Unexplained intrauterine fetal death is accompanied by activation of complement. *Journal of perinatal medicine.* 2005; 33(4):296–305. [PubMed: 16207114]
320. Jagels MA, Daffern PJ, Hugli TE. C3a and C5a enhance granulocyte adhesion to endothelial and epithelial cell monolayers: epithelial and endothelial priming is required for C3a-induced eosinophil adhesion. *Immunopharmacology.* 2000; 46(3):209–22. [PubMed: 10741901]
321. Foreman KE, Vaporciyan AA, Bonish BK, Jones ML, Johnson KJ, Glovsky MM, et al. C5a-induced expression of P-selectin in endothelial cells. *The Journal of clinical investigation.* 1994; 94(3):1147–55. [PubMed: 7521884]
322. Ferrer-Lopez P, Renesto P, Schattner M, Bassot S, Laurent P, Chignard M. Activation of human platelets by C5a-stimulated neutrophils: a role for cathepsin G. *The American journal of physiology.* 1990; 258(6 Pt 1):C1100–7. [PubMed: 2360620]
323. Del Conde I, Cruz MA, Zhang H, Lopez JA, Afshar-Kharghan V. Platelet activation leads to activation and propagation of the complement system. *The Journal of experimental medicine.* 2005; 201(6):871–9. [PubMed: 15781579]
324. Peerschke EI, Yin W, Grigg SE, Ghebrehiwet B. Blood platelets activate the classical pathway of human complement. *Journal of thrombosis and haemostasis : JTH.* 2006; 4(9):2035–42. [PubMed: 16961611]
325. Manthey HD, Woodruff TM, Taylor SM, Monk PN. Complement component 5a (C5a). *The international journal of biochemistry & cell biology.* 2009; 41(11):2114–7. [PubMed: 19464229]
326. Peerschke EI, Yin W, Ghebrehiwet B. Complement activation on platelets: implications for vascular inflammation and thrombosis. *Molecular immunology.* 2010; 47(13):2170–5. [PubMed: 20621693]
327. Caughey AB, Stotland NE, Washington AE, Escobar GJ. Maternal ethnicity, paternal ethnicity, and parental ethnic discordance: predictors of preeclampsia. *Obstet Gynecol.* 2005; 106:156–61. [PubMed: 15994632]

328. Martin JA, Hamilton BE, Osterman MJK. Births in the United States, 2016. NCHS Data Brief. 2017; (287):1–8.
329. Miller MA, Sagnella GA, Kerry SM, Strazzullo P, Cook DG, Cappuccio FP. Ethnic differences in circulating soluble adhesion molecules: the Wandsworth Heart and Stroke Study. *Clinical science*. 2003; 104:591–8. [PubMed: 12605595]
330. Akolekar R, Veduta A, Minekawa R, Chelemen T, Nicolaides KH. Maternal plasma P-selectin at 11 to 13 weeks of gestation in hypertensive disorders of pregnancy. *Hypertension in pregnancy*. 2011; 30(3):311–21. [PubMed: 20205626]

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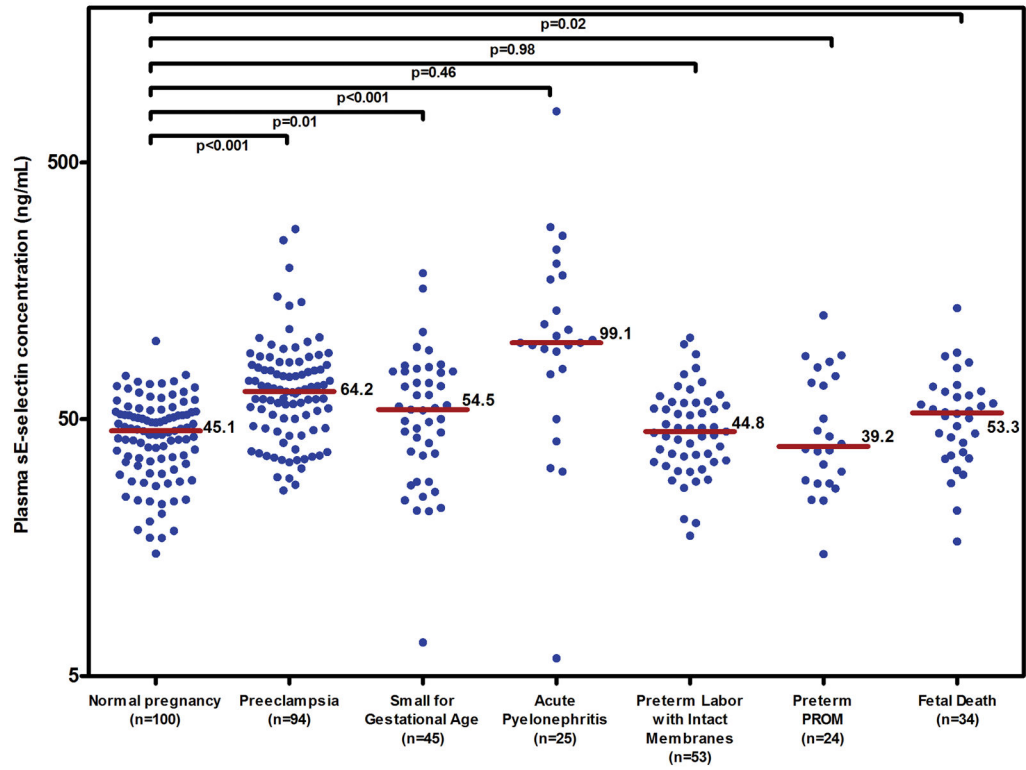


Figure 1.
Plasma sE-selectin concentration (ng/mL)

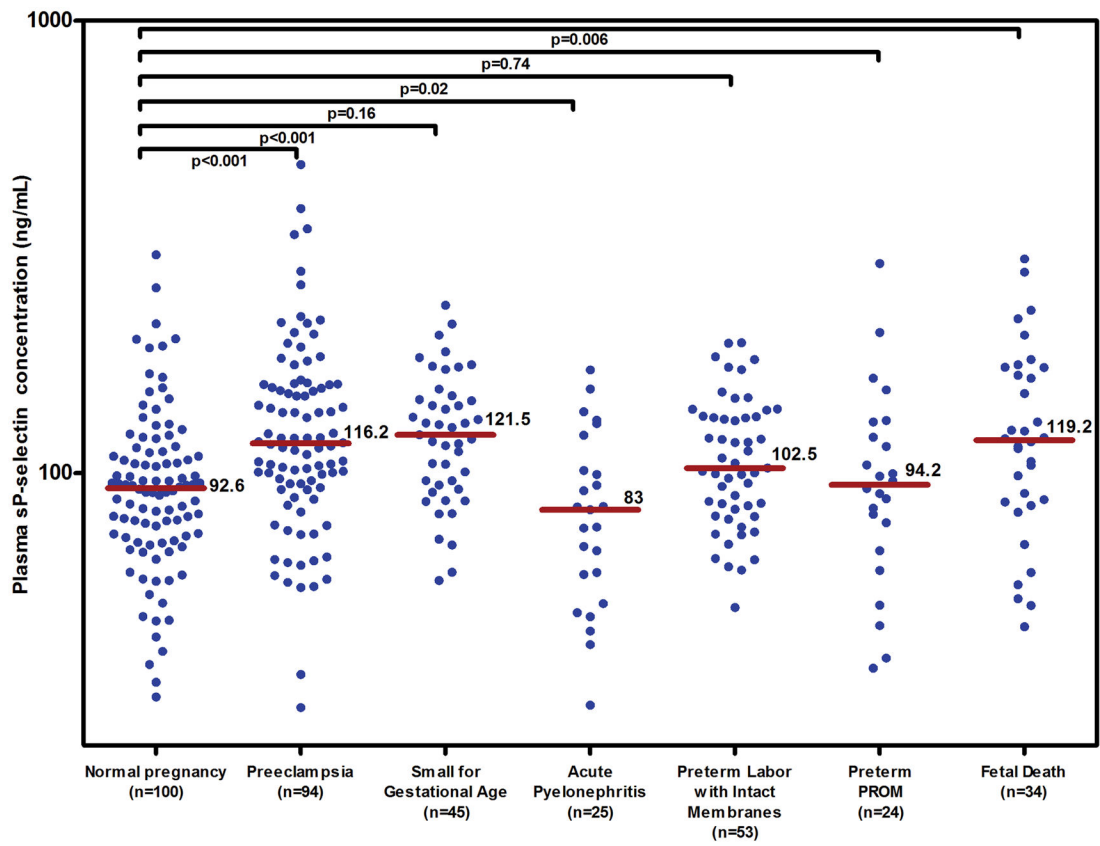


Figure 2.
Plasma sP-selectin concentration (ng/mL)

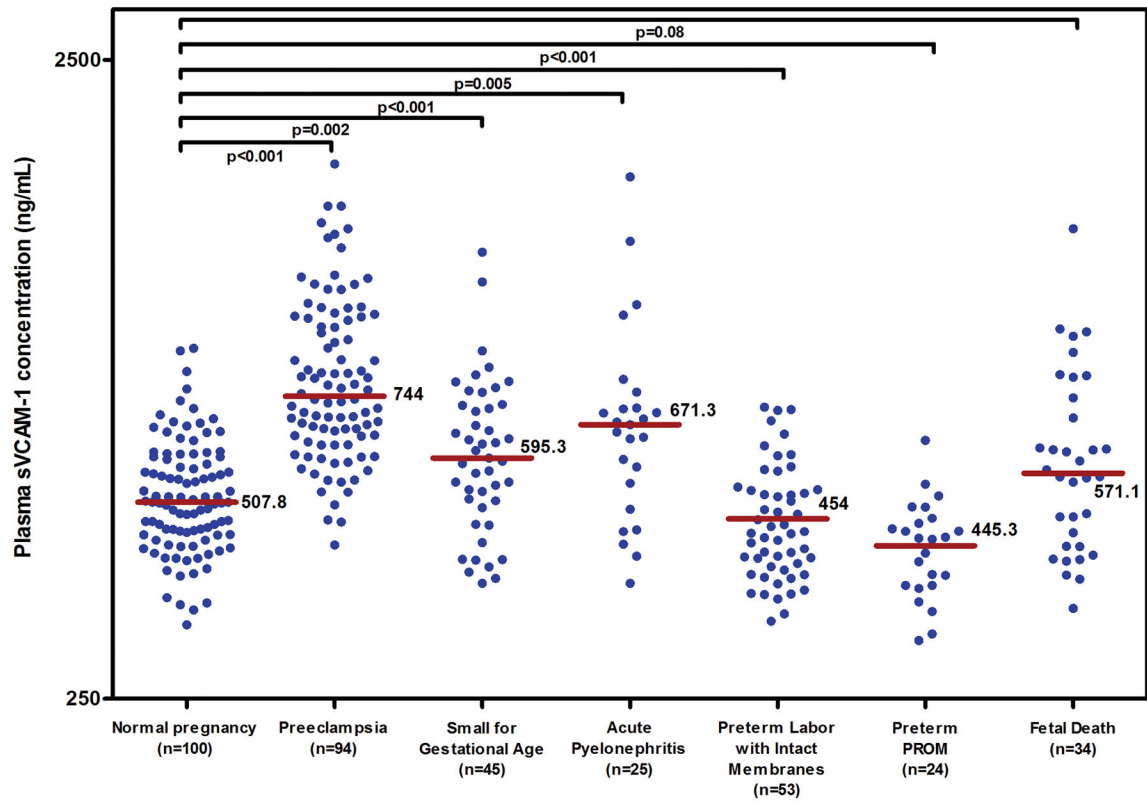


Figure 3.
Plasma sVCAM-1 concentration (ng/mL)

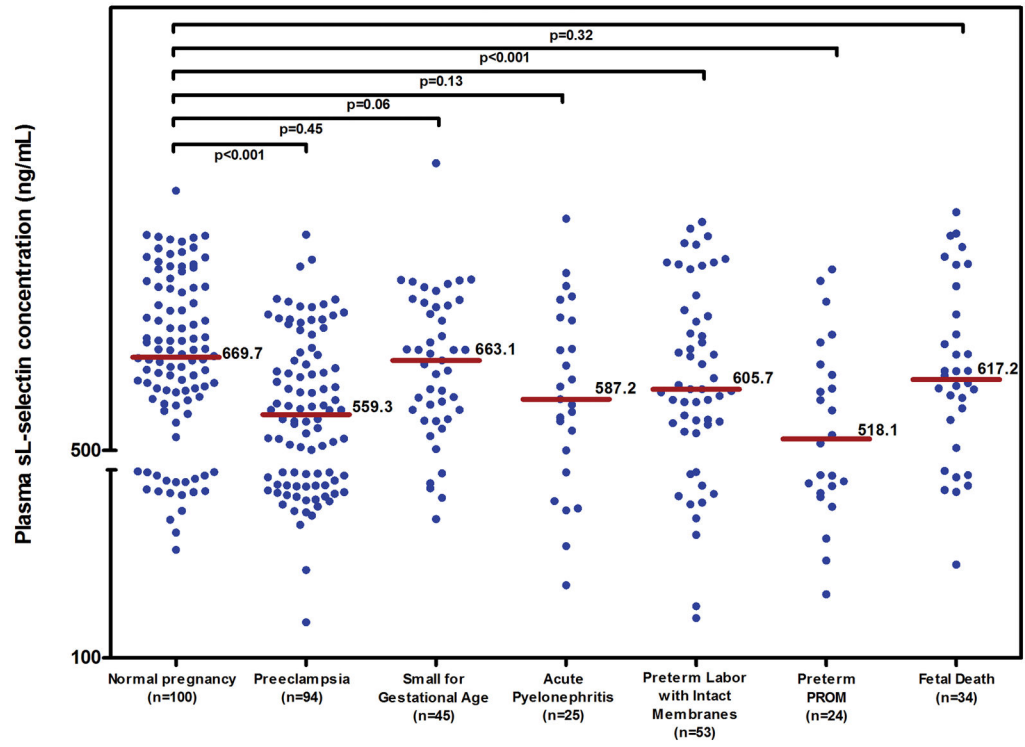


Figure 4.
Plasma sL-selectin concentration (ng/mL)

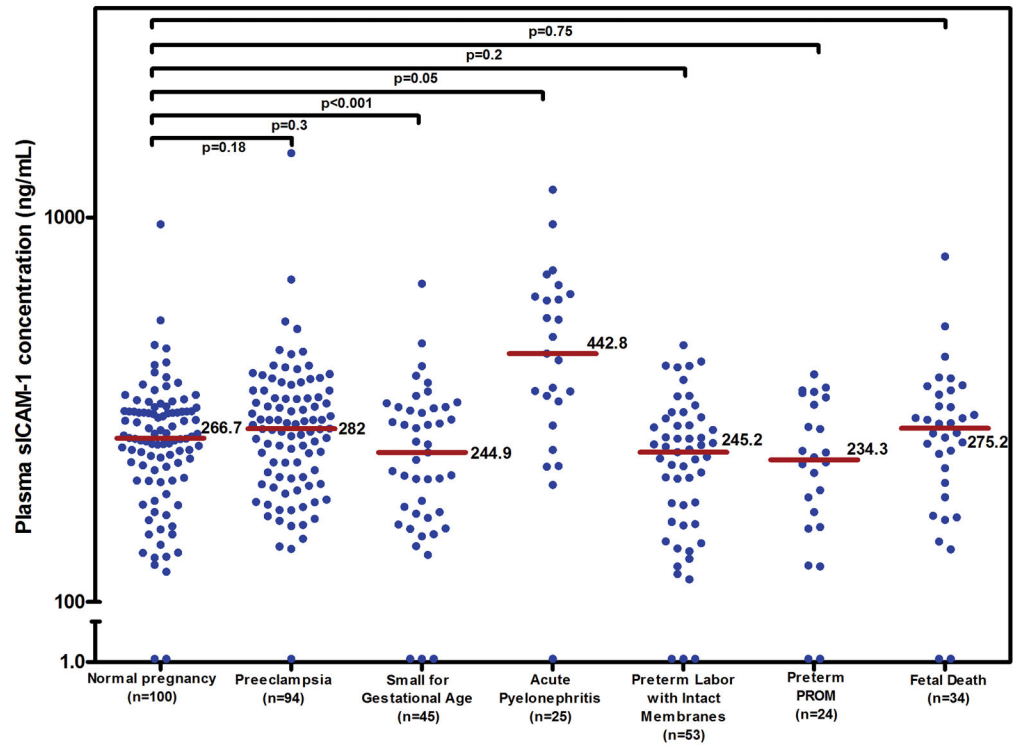


Figure 5.
Plasma sICAM-1 concentration (ng/mL)

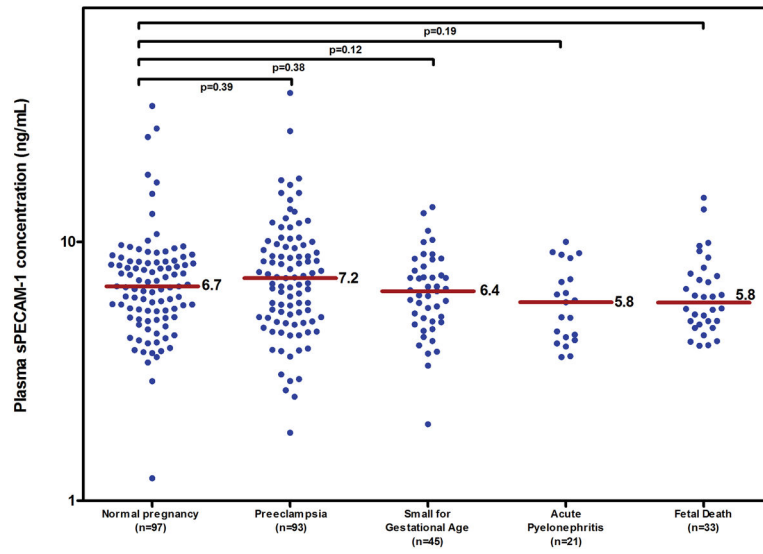


Figure 6.
Plasma sPECAM-1 concentration (ng/mL)

Table 1

Sensitivity and coefficient of variation (CV) of the immunoassay kits

Analyte	Sensitivity	Inter-assay CV	Intra-assay CV
soluble E-selectin	0.331(ng/mL)	9.07	2.77
soluble L-selectin	0.799(ng/mL)	4.54	1.46
soluble P-selectin	0.72 (ng/mL)	5.96	4.18
soluble VCAM-1	3.171(ng/mL)	8.66	5.00
soluble ICAM-1	1.058 (ng/mL)	6.89	2.85
soluble PECAM-1	0.049(ng/mL)	11.70	2.07
IL-8	5.82 (pg/mL)	6.27	5.36
TNF-α	0.329 (pg/mL)	6.60	5.95

ICAM-1: intercellular adhesion molecule-1; IL: interleukin; PECAM-1: platelet endothelial cell adhesion molecule-1; TNF: tumor necrosis factor; VCAM-1: vascular cell adhesion molecule-1

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Table 2

Demographic, clinical and obstetric characteristics of the study population

	Normal Pregnancy (n = 100)	Preeclampsia (n = 94)	Small for Gestational Age (n = 45)	Pyelonephritis (n = 25)	Preterm Labor with Intact Membranes (n=53)	Preterm PROM (n=24)	Fetal Death (n=34)	P value
Age (years)	25 (21–30)	24 (20.8–30)	25 (20–31)	22 (20.5–31)	21 (19–26.5)	28 (22–32.8)	24.5 (20–29.3)	0.04
Nulliparity	25% (25/100)	61.7% (58/94)	35.6% (16/45)	28% (7/25)	41.5% (22/53)	16.7% (4/24)	40.6% (13/32)**	<0.001
Race								
• African-American	63% (63/100)	64.9% (61/94)	80% (36/45)	72% (18/25)	88.7% (47/53)	91.7% (22/24)	88.2% (30/34)	
• Caucasian	11% (11/100)	11.7% (11/94)	15.6% (7/45)	16% (4/25)	9.4% (5/53)	8.3% (2/24)	2.9% (1/34)	0.007
• Hispanic	21% (21/100)	21.3% (20/94)	2.2% (1/45)	8% (2/25)	-	-	6% (2/34)	
• Other	5% (5/100)	2.1% (2/94)	2.2% (1/45)	4% (1/25)	1.9% (1/53)	-	2.9% (1/34)	
Gestational age at venipuncture (weeks)	34 (28.3–38.4)	33.8 (29.3–36.8)	33.5 (30.1–37.5)	31.5 (25.7–35.8)	30.4 (25.5–33)	30.1 (25.1–31.9)	30.1 (24.9–35.4)	<0.001
Gestational age at delivery (weeks)	39.1 (38.4–40.1)	34.2 (30.5–37.2)	35 (30.9–37.6)	39.2* (37.3–40.3)	35.1 (27.8–37.1)	30.2 (25.3–31.9)	30.1 (24.7–35.5)	<0.001
Birthweight (grams)	3,375 (3,158–3,800)	1,830 (1,320–2,733)	1,980 (1,045–2,325)	3,215* (2,770–3,583)	2,353 (1,005–2,775)	1,295 (655–1,988)	1,106 (513–2,031)	<0.001

* data not available in 5 cases;

** data not available in 2 cases;

PROM – prelabor rupture of membranes; numbers in () either expressed as percentage or Interquartile range

Data is presented as percent (number), median (interquartile range)

Table 3

The concentrations of soluble adhesion molecules in patients with preeclampsia, small for gestational age and fetal death compared to normal pregnancy

	Normal pregnancy (n=100)	Preeclampsia (n=94)	P value	Small for gestational age (n=45)	P value	Fetal Death (n=34)	P value
Soluble E-selectin (ng/mL)	45.1 (33.9–53.3)	64.2 (46.4–81)	<0.001	54.5 (36.8–76.8)	0.01	53.3 (37.2–64)	0.02
Soluble L-selectin (ng/mL)	669.7 (583.4–830.4)	559.3 (444.9–671.2)	<0.001	663.1 (567.8–787.3)	0.45	617.2 (503.9–765.8)	0.32
Soluble P-selectin (ng/mL)	92.6 (72.4–111.1)	116.2 (93.3–156.2)	<0.001	121.5 (94.1–145.4)	<0.001	119.2 (86.3–171.2)	0.006
Soluble VCAM-1 (ng/mL)	507.8 (451.8–593.7)	744 (644.7–983.8)	<0.001	595.3 (510.5–719.7)	0.002	571.1 (433.3–739.9)	0.08
Soluble ICAM-1 (ng/mL)	266.7 (217.6–312.8)	282.0 (211.1–340.4)	0.18	244.9 (169.7–319.1)	0.3	275.2 (204.2–323.2)	0.75
Soluble PECAM-1 (ng/mL)	6.7* (5.1–8.4)	7.2** (5.1–9.6)	0.39	6.4 (4.9–7.7)	0.38	5.8** (4.9–7.4)	0.19

NOTE: P value (obstetrical condition vs. normal pregnancy); ICAM-1: intercellular adhesion molecule; PECAM-1: platelet endothelial cell adhesion molecule; VCAM-1: vascular cell adhesion molecule;

* data not available in 3 cases;

** data not available in 1 case

Data is presented as median (interquartile range).

Summary of the differences in soluble adhesion molecules concentration in patients with normal pregnancy and those with preeclampsia, small for gestational age, acute pyelonephritis, preterm labor and intact membranes, preterm PROM and fetal death

Table 4

Soluble Adhesion Molecules	Preeclampsia (n=94)	Small for Gestational Age (n=45)	Acute Pyelonephritis (n=25)	Preterm Labor with Intact Membranes (n=53)	Preterm PROM (n=24)	Fetal Death (n=34)
Soluble E-selectin (ng/mL)	↑(+,+)	↑(+,+)	↑(+,+)	↔	↔	↑(+,+)
Soluble L-selectin (ng/mL)	↓(+,+) ¹⁾	↔	↓(-,+)	↓(-,+)	↓(-,+)	↔
Soluble P-selectin (ng/mL)	↑(+,+)	↑(+,+)	↔	↑(+,-)	↔	↑(+,+)
Soluble VCAM-1 (ng/mL)	↑(+,+)	↑(+,+)	↑(+,+)	↓(+,-)	↓(+,+) ²⁾	↑(+,+)
Soluble ICAM-1 (ng/mL)	↔	↔	↑(+,+)	↔	↔	↔
Soluble PECAM-1 (ng/mL)	↔	↔	↔	not measured	not measured	↔

ICAM-1: intercellular adhesion molecule-1; PECAM-1: platelet endothelial cell adhesion molecule-1; VCAM-1: vascular cell adhesion molecule-1; ↑ : a higher median concentration than in normal pregnancy, ↓ : a lower median concentration than in normal pregnancy, ↔: no significant change in the median concentration.

(+/-, +/-) positive/negative test by wilcoxon test (p-value <0.05) and positive/negative test by linear model with adjustment for gestational age as needed (p-value<0.05);

¹⁾ differences are higher earlier in gestation (significant interaction between gestational age and group);

²⁾ differences are higher later in gestation (significant interaction between gestational age and group).

Table 5

The concentrations of soluble adhesion molecules in patients with preterm labor and intact membranes, preterm PROM and acute pyelonephritis compared to normal pregnancy

	Normal Pregnancy (n=100)	Preterm Labor with Intact Membranes (n=53)	P value	Preterm PROM (n=24)	P value	Acute pyelonephritis (n=25)	P value
Soluble E-selectin (ng/mL)	45.1 (33.9–53.3)	44.8 (34.6–57.8)	0.46	39.2 (28.7–70.5)	0.98	99.1 (78.6–175.2)	<0.001
Soluble L-selectin (ng/mL)	669.7 (583.4–830.4)	605.7 (542.4–761.2)	0.13	518.1 (429.1–638.7)	<0.001	587.2 (500.2–750.9)	0.06
Soluble P-selectin (ng/mL)	92.6 (72.4–111.1)	102.5 (83.2–133.3)	0.02	94.2 (75.1–122.5)	0.74	83 (59.7–101.3)	0.16
Soluble VCAM-1 (ng/mL)	507.8 (451.8–593.7)	454 (402.6–529.7)	0.005	445.3 (376.2–472.6)	<0.001	671.3 (543.7–713.4)	<0.001
Soluble ICAM-1 (ng/mL)	266.7 (217.6–312.8)	245.2 (161.6–288.2)	0.05	234.3 (167.6–329.5)	0.2	442.8 (332.6–622.2)	<0.001
Soluble PECAM-1 (ng/mL)	6.7* (5.1–8.4)	not measured	-	not measured	-	5.8*** (4.3–7.2)	0.12

NOTE: P value (obstetrical condition vs. normal pregnancy); ICAM-1: intercellular adhesion molecule; PECAM-1: platelet endothelial cell adhesion molecule; PROM: prelabor rupture of membranes; VCAM-1: vascular cell adhesion molecule;

* data not available in 3 cases;

*** data not available in 4 cases.

Data is presented as median (interquartile range)

The concentrations of soluble adhesion molecules in patients with acute pyelonephritis in the presence or absence of bacteremia

Table 6

	Acute pyelonephritis without bacteremia (n=11)	Acute pyelonephritis with bacteremia (n=12)	P value
Soluble E-selectin (ng/mL)	78.6 (32.3–97.2)	178.4 (100.7–251.5)	<0.001
Soluble L-selectin (ng/mL)	587.2 (489.2–687.2)	631.3 (549.6–790.9)	0.61
Soluble P-selectin (ng/mL)	84.2 (49.1–101.3)	79.5 (62–119.9)	0.79
Soluble VCAM-1 (ng/mL)	576.4 (456.7–654.1)	734.1 (689.2–1025.2)	<0.001
Soluble ICAM-1 (ng/mL)	353.6 (332.6–442.8)	627.1 (562.9–724.6)	0.004
Soluble PECAM-1 (ng/mL)	4.8 (3.9–7.5)	6.3 (5.1–9.1)	0.13

ICAM-1: intercellular adhesion molecule; PECAM-1: platelet endothelial cell adhesion molecule; VCAM-1: vascular cell adhesion molecule

Data is presented as median (interquartile range).