

Systematic analysis of transcriptomic profiles of COPD airway epithelium using next-generation sequencing and bioinformatics

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Introduction: COPD is a chronic inflammatory disease of lung. The inflammatory response in COPD is associated with neutrophils, macrophages, T lymphocytes, and bronchial epithelial cells, and occurs mainly in the small airway, leading to irreversible airflow limitation.

Methods: In order to investigate the microRNA–mRNA interaction in the microenvironment of the COPD airway, we used next-generation sequencing and bioinformatics in this study.

Results: We identified four genes with microRNA–mRNA interactions involved in COPD small-airway bronchial epithelial cells: *NT5E*, *SDK1*, *TNSI*, and *PCDH7*. Furthermore, miR6511a-5p–*NT5E* interaction was found to be involved in small-airway bronchial epithelial cells, large-airway bronchial epithelial cells, and alveolar macrophages.

Conclusion: Our results showed that miR6511a-5p–*NT5E* interaction plays an important role in COPD, which might be associated with cell–cell contact, activation of leukocytes, activation of T lymphocytes, and cellular homeostasis. These findings provide new information for further investigations of the COPD microenvironment, and may help to develop new diagnostic or therapeutic strategies targeting the bronchial epithelium for COPD.

Keywords: bioinformatics, COPD, epithelium, miR6511a-5p, next-generation sequencing, *NT5E*

Introduction

COPD affected 174.5 million people and caused 3.2 million deaths worldwide in 2015.¹ It is progressive and usually causes major disability.² COPD is a chronic inflammatory disease of the lung. The inflammatory response of COPD is associated with inhaled noxious substances, such as smoke and air pollution.³ Neutrophils, macrophages, and T lymphocytes are increased in the lung with COPD.³ As a defensive barrier of airways, bronchial epithelial cells are also affected in the inflammatory process of COPD.⁴ After being stimulated by inhaled insults, bronchial epithelial cells initiate immune and inflammatory responses, including recruiting macrophages, neutrophils, and dendritic cells.⁴ The inflammatory response occurs mainly in the small airway and results in significant airflow limitation, which is not fully reversible. With the progression of COPD, the inflamed small airways undergo remodeling processes involving airway-wall thickening, reduced airway diameter, and increasing resistance to respiratory flow.⁵ Recently, epithelial–mesenchymal transition caused by repeated stimulation was identified as a potential source of fibroblasts and myofibroblasts that could contribute to the chronic remodeling of airways.⁶

MicroRNAs have been recognized as important regulators in the gene regulation of COPD.⁷ Downregulated miR128 in smokers, downregulated miR181d and miR30a-3p in COPD lung tissue, and increased miR223 in lungs of smokers have



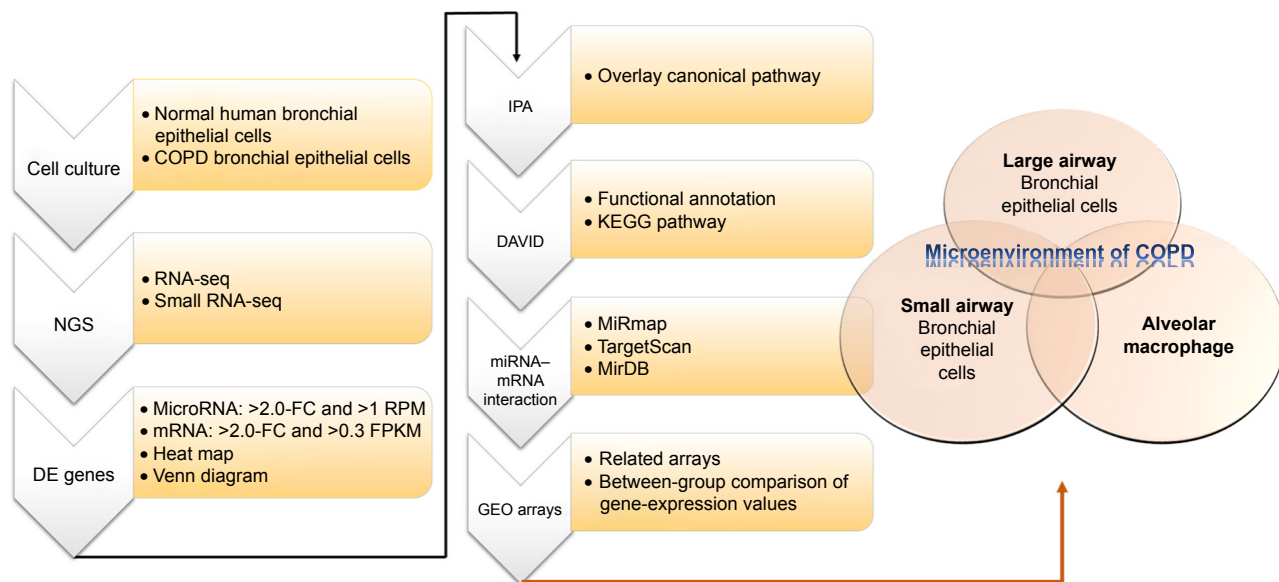


Figure 1 Flowchart of study design.

Notes: In order to investigate the roles of microRNA–mRNA interactions in the microenvironment of COPD, we used normal human bronchial epithelial cells and COPD bronchial epithelial cells for next-generation sequencing (NGS). Then, we analyzed the NGS data of with several bioinformatic tools, including MiRmap, Ingenuity Pathway Analysis (IPA), the Database for Annotation, Visualization, and Integrated Discovery (DAVID), MirDB, TargetScan, and the Gene Expression Omnibus (GEO) database.

Abbreviations: DE, differentially expressed; FC, fold change; RPM, reads per million; FPKM, fragments per kilobase of transcript per million mapped reads; KEGG, Kyoto Encyclopedia of Genes and Genomes.

been demonstrated.⁷ In a study investigating muscle wasting of COPD patients, the results showed that increased miR424-5p was associated with muscle wasting.⁸ These studies showed that microRNA–mRNA interactions may play important roles in COPD, as we have demonstrated in asthma.⁹ In order to investigate microRNA–mRNA interactions in the microenvironment of COPD airways, we used COPD bronchial epithelial (DHBE) cells and normal bronchial epithelial cells to perform next-generation sequencing (NGS) and bioinformatic analyses (Figure 1).

Methods

Primary cells

Primary normal human bronchial epithelial (NHBE) and DHBE cells were purchased from Lonza (Basel, Switzerland). Cell cultures were performed according to the manufacturer's protocol. Cells were grown in bronchial epithelial cell basal media (BEGM™ Bronchial Epithelial Cell Growth Medium BulletKit™; Lonza), supplemented with 2 mL bovine pituitary extract, 0.5 mL hydrocortisone, 0.5 mL human epidermal growth factor (hEGF), 0.5 mL epinephrine, 0.5 mL transferrin, 0.5 mL insulin, 0.5 mL retinoic acid, 0.5 mL triiodothyronine, and 0.5 mL GA1000. Cells were maintained at 37°C in a 5% CO₂ incubator and passaged with Reagent-Pack (Lonza) containing trypsin–EDTA (EDTA, trypsin neutralizing solution, and HEPES [4-(2-hydroxyethyl)-1-

piperazineethanesulfonic acid]-buffered solution). Cells were harvested for NGS analysis after cultivation from primary cells for one generation.

Next-generation sequencing

Expression profiles of microRNAs and mRNAs were examined using NGS. Total RNA of both normal and DHBE cells was extracted using Trizol reagent (Thermo Fisher Scientific, Waltham, MA, USA) according to the manufacturer's protocol. Purified RNA was then quantified at OD₂₆₀ using an ND-1000 spectrophotometer (NanoDrop; Thermo Fisher Scientific) and quality assessed with a 2,100 bioanalyzer with RNA 6000 LabChip kit (Agilent Technologies, Santa Clara, CA, USA) at Welgene Biotech (Taipei, Taiwan).

For small-RNA library construction and deep sequencing, samples were prepared using an Illumina sample-preparation kit according to the TruSeq small-RNA library sample-preparation guide. In summary, total RNA was ligated with 3' and 5' adaptors and reverse-transcribed into cDNA followed by polymerase chain-reaction amplification. The harvested cDNA constructs were fractionated by size and purified with 6% polyacrylamide-gel electrophoresis, and bands containing 18–40-nucleotide RNA fragments (140–155 nucleotides in length with both adapters) were selected. Libraries were then sequenced on an Illumina instrument (75SE cycle, single-end) and sequencing results processed with Illumina

software. For small-RNA-sequence analysis, sequencing data were applied to go through a filtering process to obtain qualified reads. Trimmomatic was used to trim or remove reads according to quality scores.¹⁰ Qualifying reads were then analyzed using miRDeep2 to clip the 3' adapter sequence and shorter reads (<18 nucleotides) removed, before aligning reads with the human genome from the University of California, Santa Cruz.¹¹ Because microRNAs are usually mapped to several genomic locations, only reads mapped perfectly to the genome five or more times were used for microRNA detection. MiRDeep2 was used to estimate expression levels of microRNAs. The criteria for microRNA selection were fold change >2, and reads per million >1.

For transcriptome sequencing, a library was constructed with Agilent's SureSelect Strand specific RNA-library-preparation kit for 75SE (single-end or paired-end) and sequencing performed on the Solexa platform. Sequences were determined directly using sequencing-by-synthesis technology via the TruSeq SBS kit. Raw sequences were obtained from Illumina Pipeline software bcl2fastq version 2.0 and expected to generate 30 million reads per sample. The sequences generated went through a filtering process to obtain qualifying reads. Trimmomatic was implemented to trim or remove the reads according to the quality score.¹⁰ Qualifying reads were analyzed using TopHat/Cufflinks to estimate gene-expression level, calculated as fragments per kilobase of transcript per million mapped reads.¹² For differential-expression analysis, the Cumberbund statistical package was employed to perform statistical analyses of gene-expression profiles. The reference genome and gene annotations were retrieved from the Ensembl database.

MiRmap database analysis

MiRmap is an open-source software library providing comprehensive microRNA-target prediction (<http://mirmap.ezlab.org>).¹³ Putative target genes can be identified by calculating the complementary ability of microRNA-mRNA interactions. The predictor also estimates mRNA-repression strength for ranking potential candidate targets by a combination of thermodynamic, evolutionary, probabilistic, and sequence-based features. The prediction results provide a list of putative target genes with MiRmap score, which is a predictive reference value. In this study, the criterion for selection of putative microRNA targets was MiRmap score ≥ 99 .

MirDB database analysis

MirDB is an online database for predicting microRNA-target and functional annotations. In MirDB, MirTarget was used for

predicting all the targets. MirTarget was designed by analyzing microRNA-target interactions from high-throughput sequencing experiments. MirDB can predict microRNA targets in five species, including human, mouse, rat, dog, and chicken.^{14,15}

TargetScan database analysis

TargetScan is an online database for predicting biological targets of microRNAs. It searches for the presence of conserved 8mer, 7mer and 6mer sites, matching the seed region of each microRNA. Predictions are ranked based on the predicted efficacy of targeting or by their probability of conserved targeting.¹⁶

DAVID database analysis

The Database for Annotation, Visualization, and Integrated Discovery (DAVID), which integrates multiple functional annotation databases, including Gene Ontology, biological processes, or Kyoto Encyclopedia of Genes and Genomes pathways, is a powerful tool for gene-function classification (<https://david.ncifcrf.gov>).¹⁷ A list of interesting genes can be classified into clusters of related biological functions, signaling pathways, or diseases by calculating the similarity of global annotation profiles with an agglomeration algorithm. It also provides an Ease score, which is a modified Fisher's exact *P*-value. The reference score represents how specifically the user genes are involved in the category (eg, signaling pathways). In this study, we selected an Ease score of 0.1 as default and 1 to extend clustering range.

Gene Expression Omnibus database analysis

The Gene Expression Omnibus (GEO) is a web database that collects submitted high-throughput gene-expression data of microarrays, chips, or NGS (<https://www.ncbi.nlm.nih.gov/geo>).¹⁸ Microarrays of accession numbers GSE4498 (small-airway bronchial epithelial cells), GSE5056 (large-airway bronchial epithelial cells), and GSE2125 (alveolar macrophages) were used in this study. GSE4498 assessed gene expression (HG133 Plus 2.0 array) in ten phenotypically normal smokers compared to 12 matched nonsmokers.¹⁹ GSE5056 assessed gene expression in 13 phenotypically normal smokers and nine normal nonsmokers.²⁰ GSE2125 assessed gene expression in 15 cigarette smokers and 15 nonsmokers.²¹ Raw data extracted from GEO were replotted and statistically analyzed with Student's *t*-test using GraphPad Prism 7 software (GraphPad Software, La Jolla, CA, USA).

Ingenuity Pathway Analysis

Ingenuity Pathway Analysis (IPA) software (Ingenuity Systems, Redwood City, CA, USA) contains a large database with detailed and structured findings reviewed by experts. This IPA database was derived from thousands of biological, chemical, and medical studies, and provides researchers with quick searching. IPA also enables analysis, integration, and recognition of data from gene and single-nucleotide-polymorphism arrays, RNA and small-RNA sequencing, proteomics, and many other biological experiments. In addition, deeper understanding and identification of related signaling pathways, upstream regulators, molecular interactions, disease processes, and candidate biomarkers are also available in IPA.²²

Results

In order to identify the potential microRNA–mRNA interactions involved in the homeostasis of COPD epithelium,

we analyzed the expressions of genes and microRNAs using NGS data from DHBE and NHBE cells (Figure 2). The gene-expression heat map of differentially expressed genes revealed 685 genes with fold change >2, including 284 downregulated genes and 401 upregulated genes. The heat map of microRNA expression revealed 144 microRNAs with fold change >2. Based on the MiRmap web-based database, we predicted 543 mRNA as targets of these 144 microRNAs, including 282 targets of upregulated microRNAs and 261 targets of downregulated microRNAs. Venn diagrams of microRNA–mRNA interactions showed that 44 genes were downregulated and 36 genes upregulated in DHBE cells compared to NHBE cells. These 80 dysregulated genes with potential microRNA–mRNA interactions are listed in Table 1.

We then used IPA to analyze the function of these 80 dysregulated genes. Possible pathways related to these 80 genes are demonstrated in Figure 3. Figure 3A shows pathways

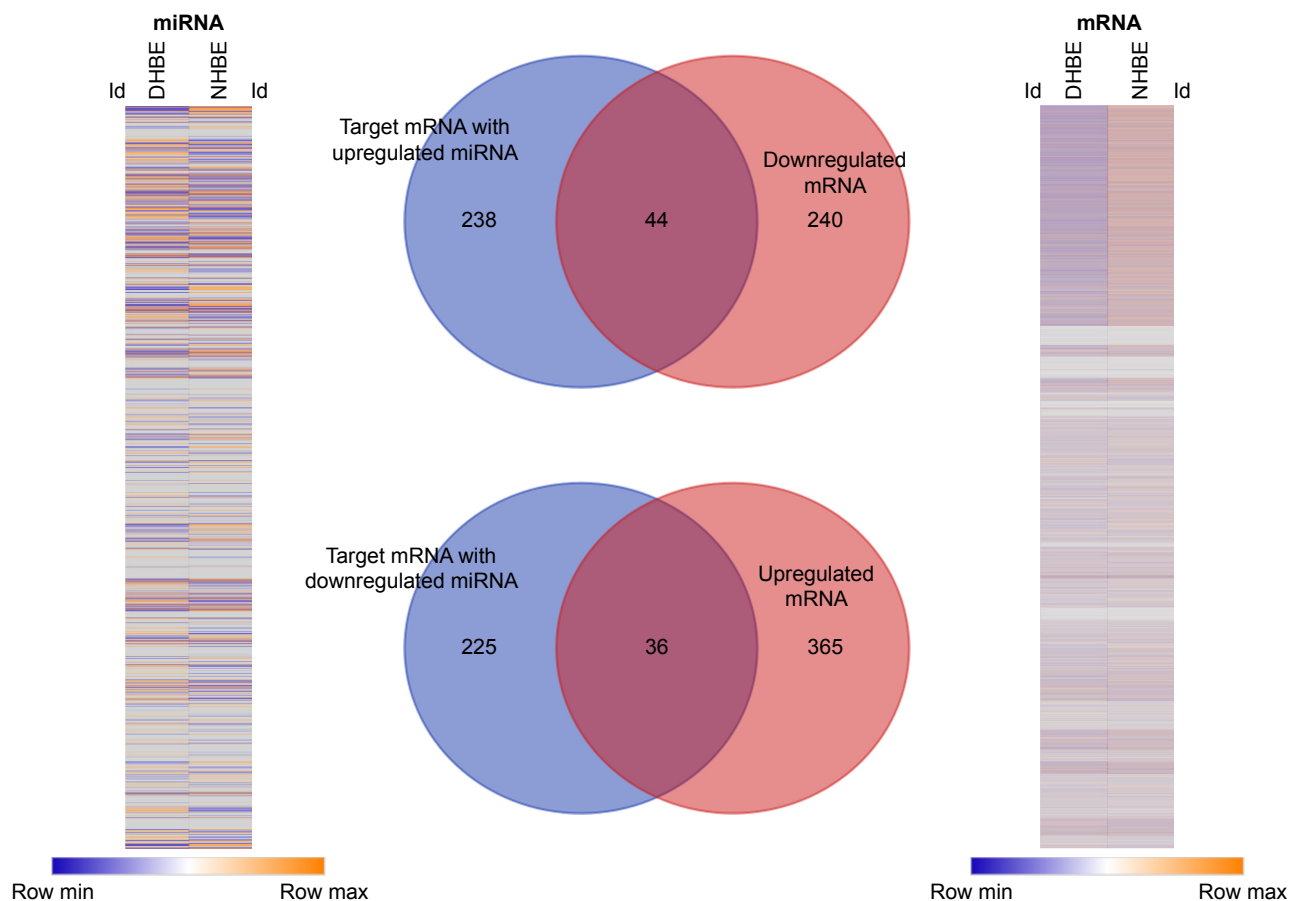


Figure 2 Identification of genes with potential microRNA–mRNA interactions in COPD bronchial epithelial cells.

Notes: Gene-expression heat map (right) of differentially expressed genes revealed 685 genes with fold change >2. The heat map (left) of differentially expressed microRNAs revealed 144 microRNAs with fold change >2 and reads per million (RPM) >1. According to the MiRmap web-based database, we predicted 543 mRNAs as targets of these 144 microRNAs. Venn diagrams of microRNA–mRNA interactions shows that 44 genes were downregulated and 36 genes upregulated in COPD bronchial epithelial (DHBE) cells compared to normal human bronchial epithelial (NHBE) cells. The selection threshold for microRNA-target prediction was MiRmap score ≥ 99 .

Table 1 Dysregulated genes with potential microRNA–mRNA interactions in COPD bronchial epithelial cells

44 downregulated genes	36 upregulated genes
<i>CCDC149, ENG, ACSS1, FOXP2, CECR1, FGFR1, C6orf223, SORL1, NOS1, ADM2, RAB3B, SDK1, PCDH7, IL24, QPRT, PAG1, CD40, PAIP2B, CDHRI, LARGE, NCALD, TGFBR3, GJC1, SCN4B, SV2A, IRAK3, CDH26, NT5E, PREX1, BEGAIN, SDK2, TNS1, KIF21B, KCNJ5, PLEKHH2, FZD10, UGT8, BEX4, ADCY9, AKT3, SLC7A8, NRSN2, OLFM2, ABCC9</i>	<i>LICAM, RTKN2, SYTI, SLC6A11, SUSDS, PCDH1, KRT80, LSP1, KIAA1644, CLDN11, CYGB, ADAMTS15, ADAM19, WDR72, LIF, SLC43A2, FAM84A, MAPIB, ALDH1A3, STC1, HHIP, RIMS3, SLC46A3, DMD, WNT5A, PLEKHA7, GPRC5A, TIMP2, FLII, GNG2, PEX5L, TFRC, MFI2, TTC9, C4orf26, CDH5</i>

related to 44 downregulated genes, and Figure 3B shows pathways related to 36 upregulated genes. In Figure 3A, epithelial adherens junction signaling, T-helper (T_H)-1- and T_H 2-activation pathways, and the T_H 2 pathway were involved in downregulation of the 44 genes. This implies that some of these genes may be associated with the function of bronchial epithelium adherence and immunity of the COPD lung. Because most microRNAs could have downregulated gene expression, we focused on the interaction between upregulated microRNA and downregulated mRNA. To analyze the possible mechanisms of these 44 downregulated genes in DHBE cells, we used DAVID (Figure 4). The results showed that these 44 genes were involved in the functions of membrane, transmembrane helix, transmembrane, nucleotide bonding, cell adhesion, calcium, disulfide bonds, and signals. We further analyzed the disease and function of 44 downregulated genes with IPA and then performed meta-analyses with MirDB and TargetScan. The meta-analyses showed 17 upregulated microRNA with interactions with 18 downregulated mRNAs (Table 2). To validate the identified 18 downregulated mRNAs in clinical COPD samples, we used the GEO database and selected a representative microarray (accession number GSE4498) that contains bronchial epithelial cells in small airways from 12 nonsmokers and ten smokers. Four genes with significant differences in mRNA expression between nonsmokers and smokers were found in GSE4498: *NT5E*, *SDK1*, *TNS1*, and *PCDH7* (Figure 5).

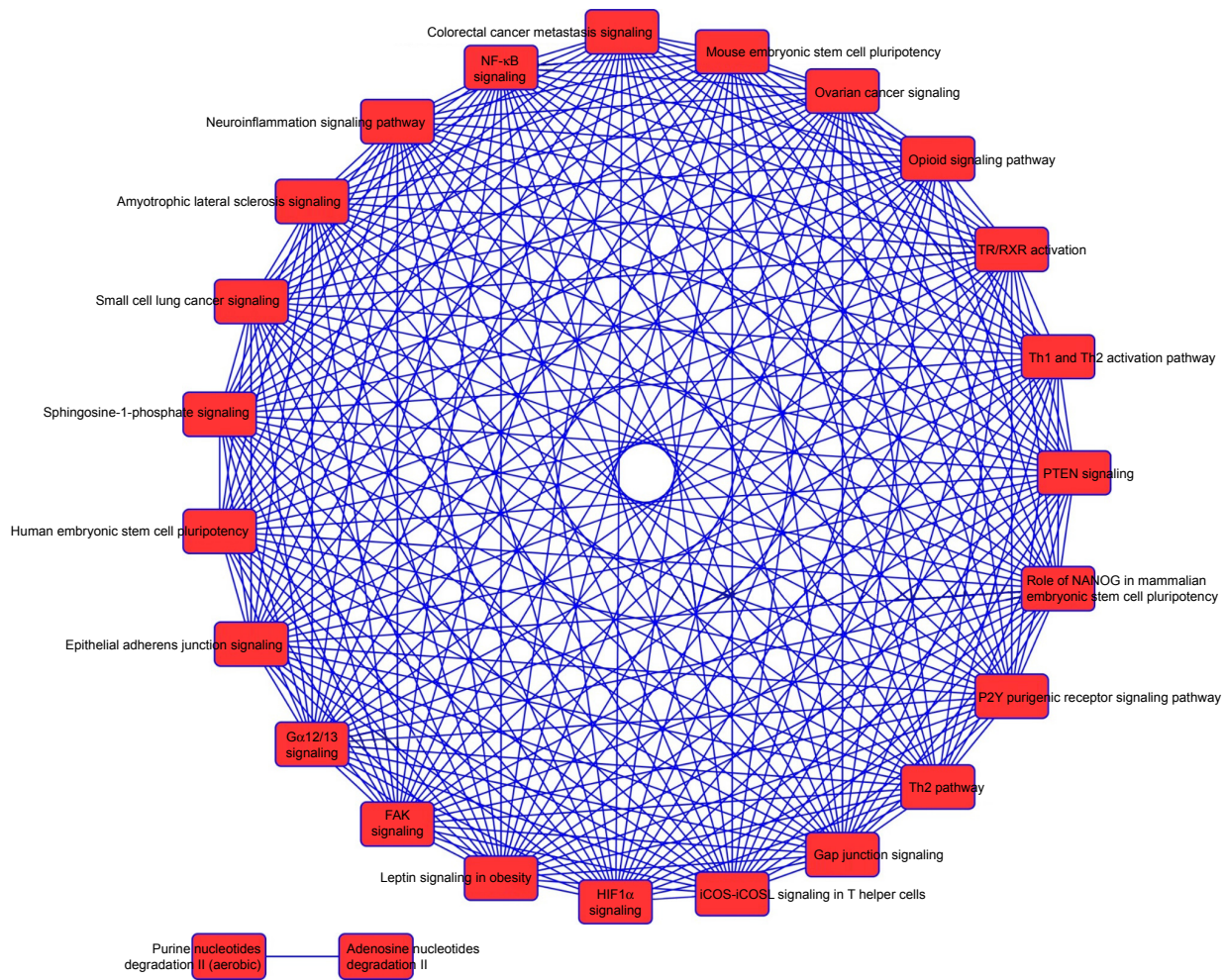
To investigate further the role of these four genes that were downregulated in the small airway of COPD, in the microenvironment of other parts of the COPD airways, we used the GSE5056 (large-airway bronchial epithelial cells) and GSE2125 (alveolar macrophages) databases for meta-analyses. The results showed *NT5E* was the only downregulated gene in bronchial epithelial cells of the small airway, large airway, and alveolar macrophages (Figure 6). Expression data for *TNS11*, *PCDH*, and *SDK1* were not available in the GSE5056 data set. Our data showed that hsa-miR6511a-5p was the possible upstream regulator for *NT5E* (Table 2). *NT5E* may be involved in cell–cell contact,

activation of leukocytes, activation of T lymphocytes, and cellular homeostasis. We also analyzed the 36 upregulated mRNAs and found only two interactions: miR20b-5p–*SLC46A3* and miR145-5p–*FLII*. However, of *SLC16A3* and *FLII* overexpression was not validated in the data sets of GSE4498, GSE5056, or GSE2125 (data not shown).

Discussion

COPD is a disease with abnormal inflammatory airway processes and can lead to structural changes in lung parenchyma, airways, and vessels.²³ Macrophages in the lung are important immunoeffector cells. Lung macrophages provide innate and adaptive immunoresponses to inhaled foreign matters in the lung.²⁴ Bronchial epithelial cells of large and small airways and alveolar macrophages could be considered important components of the COPD microenvironment. Further investigation of gene regulations in this COPD microenvironment may provide potential therapeutic implications and knowledge of COPD pathogenesis. In this study, we analyzed the gene-expression profiles of microRNAs and mRNAs in NHBE and DHBE cells by NGS. We used bioinformatics to investigate the potential molecular mechanisms of gene regulation. Using MiRmap for predicting targets and Venn diagrams for intersection analysis, we focused on 44 potential microRNA–mRNA interactions (upregulated microRNA–downregulated mRNA). After meta-analysis with MirDB and TargetScan website, we focused on 17 upregulated microRNAs with interactions with 18 downregulated mRNAs. We further analyzed these 18 genes in the GEO database. The database we chose was GSE4498 (microarray data from small-airway bronchial epithelial cells), and four genes with significant and identical changes were found. In order to analyze the role of these four genes in the microenvironment of COPD, we further analyzed these four genes in the GSE5056 (large-airway bronchial epithelial cells) and GSE2125 (alveolar macrophages) databases. The results showed that *NT5E* was the only significantly downregulated gene. According to our NGS data, miR6511a-5p was the upstream regulator of *NT5E*. In the IPA analysis, *NT5E* was associated with

A



B

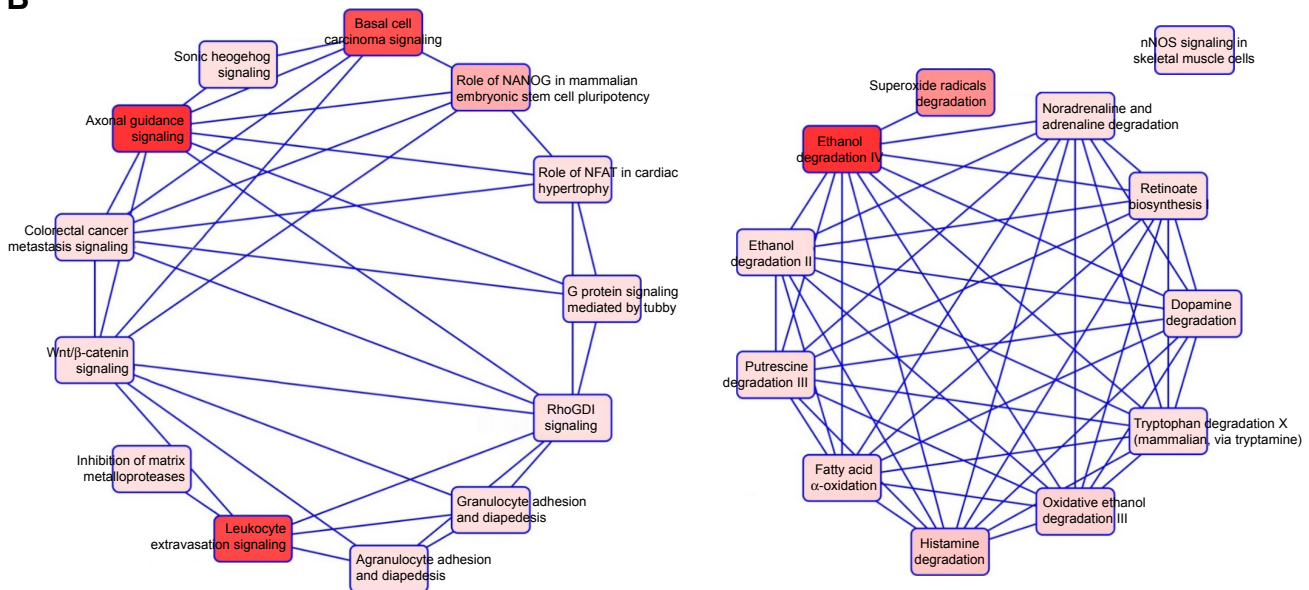


Figure 3 Functional analysis of dysregulated genes identified in COPD epithelial cells by Ingenuity Pathway Analysis (IPA).

Notes: The 80 identified dysregulated genes with potential microRNA–mRNA interactions were analyzed by IPA. **(A)** Pathways related to 44 downregulated genes; **(B)** pathways related to 36 upregulated genes.

Abbreviations: TR, thyroid hormone receptor; RXR, retinoid X receptor; PTEN, phosphatase and tensin homolog; FAK, focal adhesion kinase; NFAT, nuclear factor of activated T-cell.

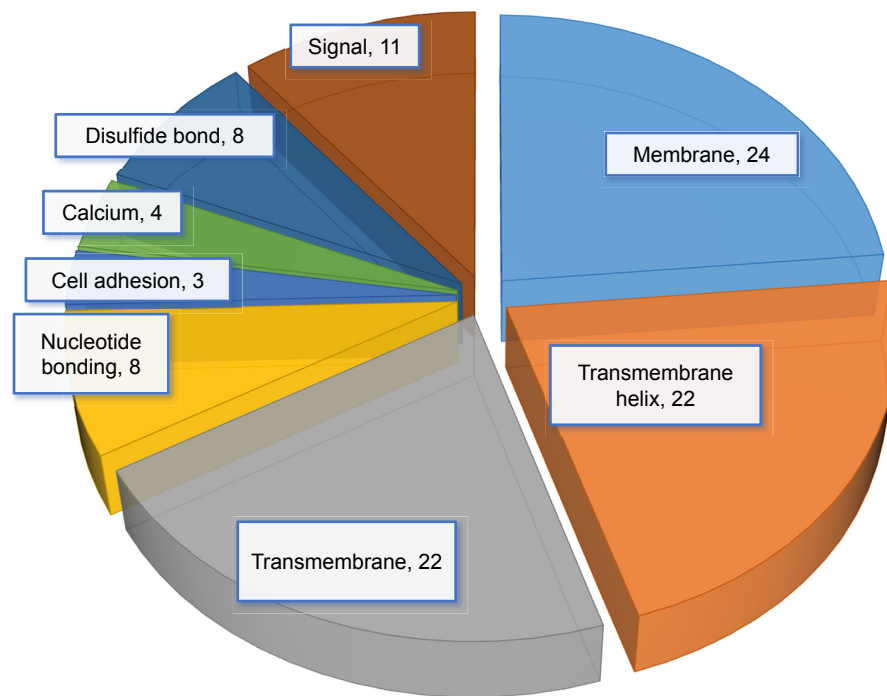


Figure 4 Possible mechanisms of dysregulated genes identified in COPD epithelial cells analyzed by Database for Annotation, Visualization, and Integrated Discovery (DAVID).

Notes: Functional annotation of the 44 downregulated genes was determined by gene ontology using DAVID. These 44 genes are involved in the functioning of membrane, transmembrane helix, transmembrane, nucleotide bonding, cell adhesion, calcium, disulfide bonds, and signals.

cell–cell contact, activation of leukocytes, activation of T lymphocytes, and cellular homeostasis.

The *NT5E* gene is associated with cellular functions dependent on G-protein-coupled receptors specific for adenosine. These functions include proliferation, apoptosis, and activation.²⁵ In a study of glioma cell lines, inhibition of ecto-5'-nucleotidase, the enzyme encoded by the *NT5E* gene, led to significant reduction in glioma-cell proliferation.²⁶ In a study of human airways, ecto-5'-nucleotidase was found to be responsible for the production of adenosine on the mucosal surface of human airway epithelial cells and was an important factor in the regulation of adenosine-mediated epithelial functions.²⁷ In a radiation-induced pulmonary fibrosis study, inhibition of ecto-5'-nucleotidase significantly reduced radiation-induced lung fibrosis.²⁸ In our study, the downregulation of *NT5E* discovered from NGS analysis was validated in the GEO databases of COPD small airway, large airway, and alveolar macrophages. Accompanying the results of IPA analysis, *NT5E*, regulated by miR6511a-5p, could be an important factor in the microenvironment of the COPD airway.

In our study, downregulation of *SDK1*, *TNS1*, and *PCDH1* discovered from NGS analysis was also validated in the GEO database of COPD small-airway epithelial cells. The *SDK1* gene encodes SDK1, which is an adhesion

molecule. SDK1 is activated by cellular stress, such as with reactive oxygen species.²⁹ SDK1 may be also associated with asbestos-exposure-related lung malignancies.²⁹ In a study investigating cross-species cancer genes, SDK1 was both significantly amplified and significantly deleted, which may indicate that the *SDK1* gene resides in unstable regions of the genome.³⁰ Therefore, the down-regulated *SDK1* shown in our study might have been related to tumorigenesis and associated with lung cancer in COPD patients.

The *TNS1* gene encodes tensin 1, which is involved in fibrillar adhesion formation. It might also be related to cell migration and cartilage development.³¹ A genome-wide association study of COPD also showed that patient forced expiratory volume in 1 second (FEV_1) or FEV_1 :forced vital capacity (FVC) ratio was associated with common variants at locus 2q35 in *TNS1*.³² A study on TNS1 function showed significant reduction in proliferation and migration in endothelial cells isolated from *TNS1*-knockout mice or those silenced with *TNS1* siRNA.³³ In our study, we found an opposite direction for *TNS1* expression in small-airway bronchial epithelial cells and alveolar macrophages of COPD lungs. These findings suggest that proliferation and migration might be decreased in DHBE cells and increased in COPD alveolar macrophages.

Table 2 Downregulated genes with microRNA–mRNA interactions and their functions

MicroRNA	mRNA	Function (IPA)	MiRmap score	MirDB (target score)	TargetScan (total context score)
hsa-miR1285-3p	<i>PLEKHH2</i>	Tumorigenesis of malignant tumor	99.6181	66	−0.17
hsa-miR195-5p	<i>FGFR1</i>	Proliferation of connective tissue cells, depressive disorder, angiogenesis, proliferation of lipoblasts, migration of cells, major depression	99.8560	72	−0.55
hsa-miR3173-3p	<i>CDH26</i>	Eosinophilia	99.1171	50	−0.87
	<i>SDK1</i>	Tumorigenesis of malignant tumors	99.9039	62	−0.58
hsa-miR3613-3p	<i>GJC1</i>	Cell–cell contact, muscle contraction	99.8337	89	−0.63
hsa-miR378g	<i>ENG</i>	Autophagy of vascular endothelial cells, proliferation of connective tissue cells, detachment of vascular endothelial cells, cellular homeostasis, angiogenesis, autophagy of cells, migration of cells, organism death	99.8158	80	−0.25
	<i>ADM2</i>	Respiration of adipocytes, angiogenesis, quantity of M2 macrophages, quantity of leukocytes	99.9646	67	−0.93
hsa-miR424-5p	<i>AKT3</i>	Autophagy of vascular endothelial cells, proliferation of connective tissue cells, autophagy of cells, migration of cells, cellular homeostasis	99.0670	99	−0.67
	<i>FGFR1</i>	Proliferation of connective tissue cells, depressive disorder, angiogenesis, proliferation of lipoblasts, migration of cells, major depression	99.8506	72	−0.55
hsa-miR4435	<i>TNS1</i>	Migration of cells	99.8208	74	−0.16
hsa-miR449a	<i>SDK2</i>	Tumorigenesis of malignant tumor	99.8641	83	−0.41
hsa-miR4532	<i>KIF21B</i>	Long-QT syndrome	99.5702	94	−0.46
hsa-miR4745-5p	<i>SLC7A8</i>	Digestive organ tumor	99.6903	54	−0.25
hsa-miR50015p	<i>TGFBR3</i>	Angiogenesis	99.7104	82	−0.29
	<i>KIF21B</i>	Long-QT syndrome	99.9772	95	−0.34
	<i>SDK1</i>	Tumorigenesis of malignant tumor	99.7629	78	−0.17
hsa-miR5190	<i>FOXP2</i>	Tumorigenesis of carcinoma	99.1872	93	−0.20
hsa-miR570-3p	<i>FOXP2</i>	Tumorigenesis of carcinoma	99.5231	95	−0.21
hsa-miR641	<i>PCDH7</i>	Neurological signs	99.4883	94	−0.47
hsa-miR6505-5p	<i>NCALD</i>	Digestive organ tumor	99.2089	96	−0.56
hsa-miR6511a-5p	<i>TNS1</i>	Migration of cells	99.9283	91	−0.13
	<i>OLFM2</i>	Secretion of protein	99.9239	100	−1.27
	<i>NT5E</i>	Cell–cell contact, activation of leukocytes, activation of T lymphocytes, cellular homeostasis	99.5498	95	−0.35
hsa-miR6724-5p	<i>NCALD</i>	Digestive organ tumor	99.7041	91	−0.75

Note: Predicted by Ingenuity Pathway Analysis (IPA) and meta-analysis with MiRmap, MirDB, and TargetScan.

The *PCDH7* gene encodes Pcdh7, which is a member of the cadherin superfamily.³⁴ The *PCDH7* gene product is an integral membrane protein involves in cell–cell recognition and adhesion.³⁵ A study of gastric cancer demonstrated that Pcdh7 inhibited cell migration and invasion.³⁵ Therefore, downregulation of *PCDH7* could promote cell migration. In the nervous system, protocadherins are widely expressed transmembrane proteins, and overexpression of Pcdh7 would cause intrinsic apoptotic pathways in primary cortical neurons.³⁶ Our IPA analysis also showed that *PCDH7* was associated with neurological signs. However, in the pathogenesis of COPD, Pcdh7 might be more related to migration and adhesion of bronchial epithelial cells of the small airway.

To date, there has been neither cure against COPD nor effective biomarkers for diagnosing COPD. In this study,

we identified potential microRNA–mRNA interactions that regulate the homeostasis of DHBE cells. Our results showed that the miR6511a-5p–*NT5E* interaction plays an important role in COPD and might be associated with cell–cell contact, activation of leukocytes, activation of T lymphocytes, and cellular homeostasis. We also found that miR3173-3p–*SDK1*, miR4435–*TNS1*, and miR641–*PCDH7* interactions might be associated with COPD pathogenesis in small-airway bronchial epithelial cells (Figure 7). However, there are several limitations in this study. Firstly, it is not easy to obtain bronchial epithelial cells from either healthy subjects or COPD patients. Potentially dysregulated microRNAs, mRNAs, and their interactions in COPD were generated from only one normal subject and one COPD patient. Although we used GEO to validate the differentially expressed genes and TargetScan

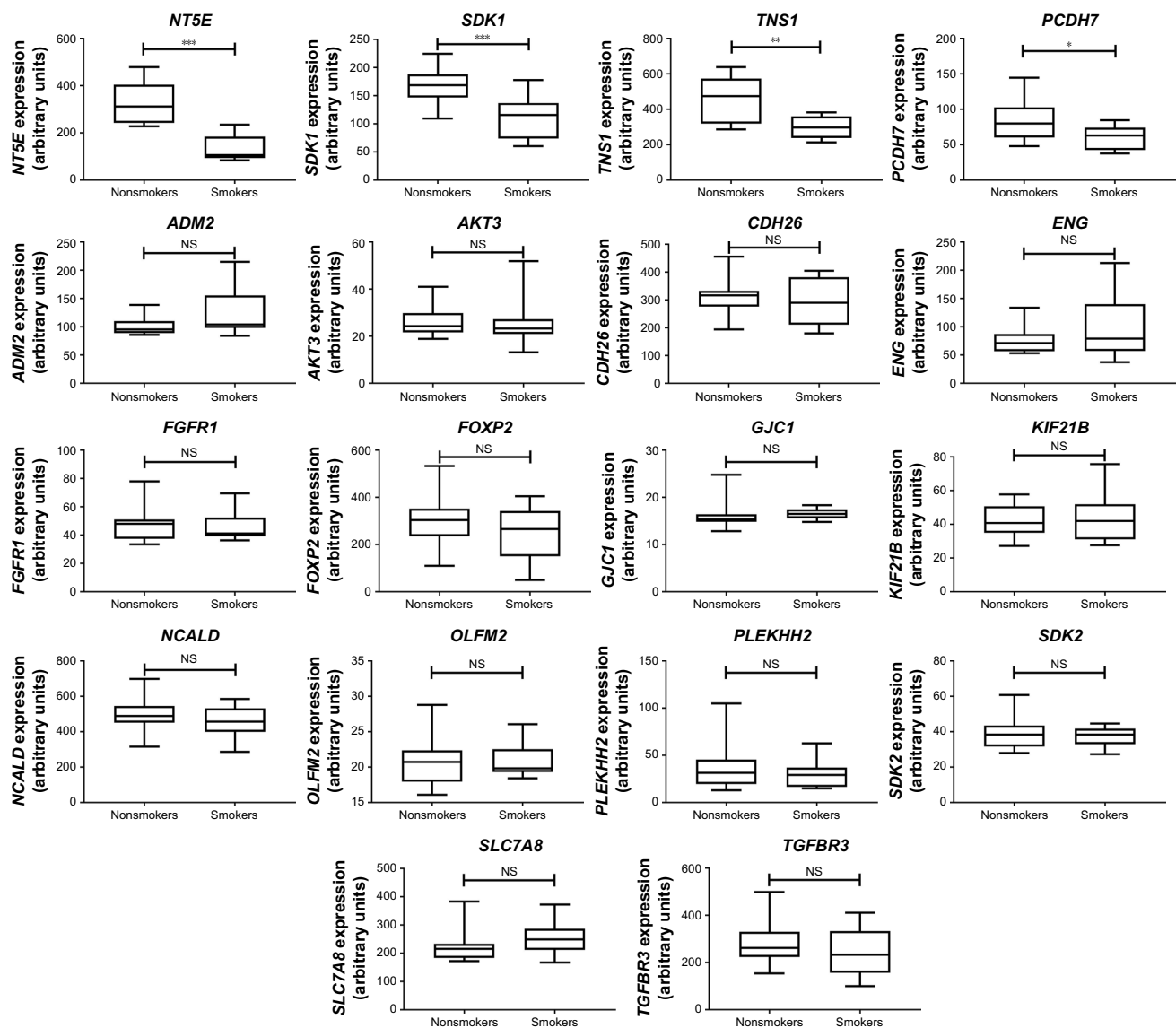


Figure 5 Gene Expression Omnibus (GEO) database analysis of 18 downregulated genes with potential microRNA–mRNA interactions in COPD small airway.

Notes: Gene expression of the 18 downregulated genes with potential microRNA–mRNA interactions was analyzed using GSE4498 microarray data from the GEO database. The results showed that expression of *NT5E*, *SDK1*, *TNS1*, and *PCDH7* was significantly downregulated in patients with COPD compared to normal controls. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Abbreviation: NS, not significant.

and MirDB to validate the interaction between microRNA and gene expression predicted by MiRmap, these findings need to be validated in more clinical samples. Secondly, the DHBE cells, cultivated in the same normal epithelial culture medium, had already gone out of the actual microenvironment in COPD airways. As this was a screening and bioinformatic study, our findings need further validation.

Conclusion

In conclusion, this study showed that miR6511a-5p–*NT5E*, miR3173-3p–*SDK1*, miR4435–*TNS1*, and miR641–*PCDH7* interactions might play important roles in the bronchial epithelium of COPD. These findings provide new information

for further investigations of the COPD microenvironment, and may help in development of new diagnostic or therapeutic strategies targeting bronchial epithelium for COPD.

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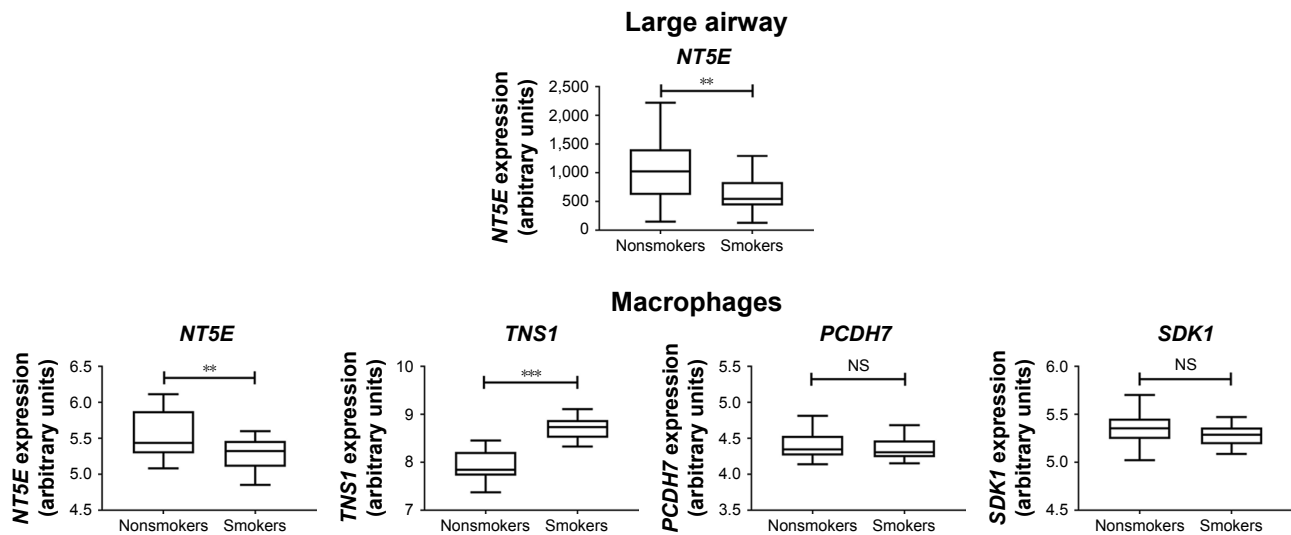


Figure 6 Gene Expression Omnibus database analysis of the four genes downregulated in COPD small airway, large airway, and alveolar macrophages.

Notes: The four downregulated gene with potential microRNA–mRNA interactions validated in the GSE4498 database (COPD small-airway bronchial epithelial cells) were further analyzed in the GSE5056 database (large airway) and GSE2125 database (alveolar macrophages). *NT5E* was the only significantly downregulated mRNA in all databases. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Abbreviation: NS, not significant.

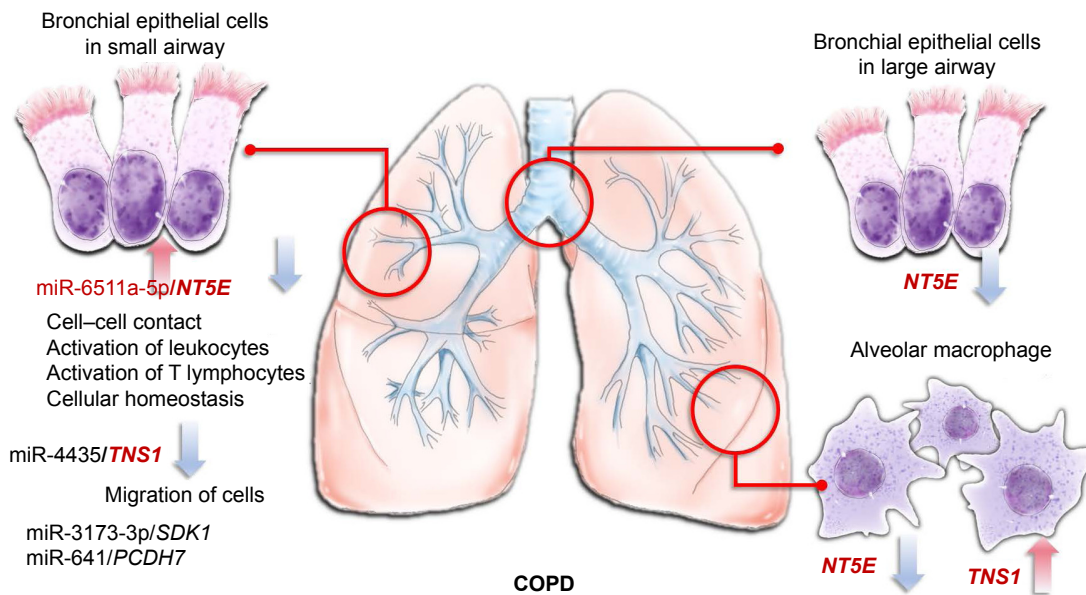


Figure 7 MicroRNA–mRNA interactions in the microenvironment of COPD.

Notes: The miR6511a-5p–*NT5E* interaction plays an important role in COPD, and may be associated with cell–cell contact, activation of leukocytes, activation of T lymphocytes, and cellular homeostasis. miR3173-3p–*SDK1*, miR4435–*TNS1*, and miR641–*PCDH7* interactions might also be associated with COPD pathogenesis in small-airway bronchial epithelial cells.

Disclosure

The authors report no conflicts of interest in this work.

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