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Dietary nitrate and skeletal muscle contractile function in heart failure

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Abstract

Heart failure (HF) patients suffer from exercise intolerance that diminishes their ability to perform normal activities of daily living and hence compromises their quality of life. This is due largely to detrimental changes in skeletal muscle mass, structure, metabolism, and function. This includes an impairment of muscle contractile performance, i.e., a decline in the maximal force, speed, and power of muscle shortening. Although numerous mechanisms underlie this reduction in contractility, one contributing factor may be a decrease in nitric oxide (NO) bioavailability. Consistent with this, recent data demonstrate that acute ingestion of NO₃⁻-rich beetroot juice, a source of NO via the NO synthase-independent enterosalivary pathway, markedly increases maximal muscle speed and power in HF patients. This review discusses the role of muscle contractile dysfunction in the exercise intolerance characteristic of HF, and the evidence that dietary NO₃⁻ supplementation may represent a novel and simple therapy for this currently-underappreciated problem.

Keywords

heart failure; dietary nitrate; nitric oxide; muscle speed; muscle power; beetroot juice

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Compliance with Ethical Guidelines

Conflict of Interest

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Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

Introduction

Heart failure (HF) is a deadly disease affecting nearly 6 million men and women in the United States and millions more worldwide (1). It is also a highly-disabling disease that markedly impairs the ability of patients to perform normal activities of daily living (e.g., walking) and hence significantly reduces their quality of life. Indeed, one of the major systems for characterizing the severity of HF, i.e., the New York Heart Association (NYHA) Functional Classification, categorizes HF patients based largely on their ability to undertake physical activity. Historically, the exercise intolerance accompanying HF was viewed as being strictly a function of the diminished ability of the failing heart to provide adequate blood flow to peripheral tissues (2,3). Over the last three decades, however, considerable evidence has accumulated that deleterious changes within skeletal muscle itself also majorly contribute to the diminished exercise capacity of patients with “classic” HF, i.e., HF with reduced ejection fraction (2,3). Much more recently, similar muscle abnormalities have been reported in patients with HF with preserved ejection fraction (3,4,5). In this brief review, we discuss these data, and in particular the evidence that alterations in intrinsic muscle contractile properties, i.e., in muscle strength, speed, and power, play an important role in limiting the functional capacity of patients with HF. From there we turn our attention to the possible significance of reductions in nitric oxide (NO) bioavailability in contributing to these detrimental changes. Finally, we describe recent research indicating that dietary nitrate (NO_3^-), a source of NO via the NO synthase (NOS) –independent enterosalivary pathway, may provide a simple but effective means of combating this problem.

Causes of exercise intolerance in HF

HF as a “cardiomuscular” disease

As indicated above, it had long been held that decreased perfusion of skeletal muscle was the singular determinant of the reduced exercise capacity of HF patients. Indeed, this seems to be a common misconception among many clinicians even today. It is now abundantly clear, however, that secondary effects of HF on skeletal muscle morphological and metabolic characteristics play a critical, if not dominant, role in this impairment. These HF-induced changes in muscle include, but are not limited to, an increased percentage of type II, or fast-twitch, fibers (7–11), an elevation in glycolytic enzyme activities (7,8), a decrease in capillarization (8,9), and a decline in mitochondrial respiratory capacity (7,8,11) resulting not only from a reduction in total muscle mitochondrial content but also from changes in the quality of mitochondria themselves (12). The sum total of these changes is a more fatigable phenotype characterized by a diminished ability to match aerobic ATP production to ATP demand during contractile activity, resulting in a greater decrease in phosphocreatine (PCr) and greater increases in inorganic phosphate (Pi) and H^+ levels during exercise at any given intensity (7,13,14,15). Importantly, these metabolic abnormalities appear to be independent of any reduction in blood flow and hence in O_2 delivery to muscle (13–19). In particular, increasing cardiac output and hence bulk limb blood flow via dobutamine infusion does not improve $\text{VO}_{2\text{peak}}$ or exercise capacity in patients with HF (13,14,19), nor does it correct the abnormalities in muscle metabolism observed during exercise (19). Thus, the primary limitation(s) to sustained exercise performance in HF patients appear(s) to reside outside of

the heart, and within the skeletal muscles themselves. Indeed, even what is perhaps the signature complaint of patients with HF – that is, of dyspnea on exertion – has been hypothesized to be due largely, if not entirely, to these changes in skeletal muscle characteristics (20). The resulting abnormal metabolic response to contractile activity is hypothesized to lead to greater stimulation of group III–IV afferent nerves in muscle and hence enhanced feedback stimulation of ventilation during exercise. Notably, the morphological and metabolic abnormalities found in the muscles of patients with HF do not improve following cardiac transplantation, which almost certainly contributes to the fact that even after transplant the exercise capacity of such patients remains significantly diminished compared to that of healthy control subjects (21,22).

Role of diminished muscle contractile function

The changes in skeletal muscle characteristics and thus metabolism and energetics described above are undoubtedly important in accounting for the reduced exercise tolerance found in patients with HF. Other factors, however, are also operative. In particular, a substantial number of studies have demonstrated that the muscles of HF patients are smaller and weaker than those of healthy individuals (23–30), with the plantar flexors in particular seeming to be adversely affected (31,32). Somewhat counterintuitively, given the changes in fiber type mentioned previously, HF also results in a significant decrease in the maximal velocity of muscle contraction, and hence an even greater reduction in maximal muscle power (which is the product of force and velocity) (30,33). These observations are salient for a number of reasons. First, many normal activities of daily living (e.g., getting out of a chair, climbing a flight of stairs, carrying groceries) are heavily dependent upon the ability of muscle to generate significant force and/or power. Second, HF-induced reductions in whole-body exercise capacity are closely associated with these alterations in muscle contractile performance (21,23,26,27,31,34,35). Senden et al. (36), for example, found that maximal power output during an incremental cycle ergometer test to fatigue in HF patients was significantly correlated with the strength and endurance of their knee extensors and flexors as assessed using isokinetic dynamometry. Other studies have demonstrated a significant relationship between the reduction in muscle function in HF patients and the reduction in VO_2peak (27) and/or the elevated ventilatory demand during exercise (35). In keeping with such data, resistance exercise training has been found to significantly improve walking ability and VO_2peak in HF patients (37,38), as well as to increase both objectively and subjectively measured physical function (39). Third, muscle function has been demonstrated to be a powerful predictor of long-term survival in patients with HF, even more so than VO_2peak (39). Clearly, alterations in muscle contractile characteristics play a critical role in the morbidity and mortality of HF.

Mechanisms

The mechanisms responsible for the HF-induced reductions in muscle force, speed, and power described above have not been fully elucidated. In part, they may be due to a disease-related reduction in habitual physical activity (33), and/or due to the muscle atrophy (“cardiac cachexia”) often found in patients with HF. Seminal research by Toth, Miller, and colleagues (34), however, has demonstrated that HF patients are weaker, slower, and less powerful than healthy control subjects even when both groups are carefully matched for age,

sex, level of physical activity, and medication use. Furthermore, such differences persist even when data are statistically adjusted for leg lean mass to account for possible muscle atrophy in the HF patients (34,41). Thus, physical inactivity and/or muscle wasting alone cannot account for the changes in muscle contractile function that are associated with, and contribute significantly to, the exercise intolerance characteristic of HF.

If the reductions in muscle force, speed, and power with HF cannot be fully explained by a more sedentary lifestyle or muscle atrophy, what factor(s) are responsible? The answer to that question would seem to reside at the molecular level itself. For example, using isolated single muscle fibers Szentesi et al. (42) have demonstrated that HF is accompanied by a reduction in isometric force per unit of cross-sectional area, which is paralleled by a decline in ATPase activity. These findings imply that HF results in a reduction in the number and/or rate of actomyosin crossbridge formation. Consistent with this, working in Toth's laboratory Miller found that HF was associated with a selective loss of myosin in type I, type IIA, and type IIA/X fibers (43). In a follow-up study, these same authors demonstrated a HF-induced slowing of myosin cross-bridge kinetics in both type I and type IIA fibers, as well as a reduction in Ca^{2+} sensitivity in the latter fiber type (44). These alterations were ascribed, at least in part, to a reduction in Akt phosphorylation, which was significantly correlated with the decline in myosin heavy-chain protein content (45). Studies by others have demonstrated abnormalities in Ca^{2+} -handling protein (i.e., ryanodine receptor type I, sarco(endo)plasmic reticulum Ca^{2+} ATPase (SERCA) 2a, phospholamban, and dihydropyridine receptor) regulation and/or content in the skeletal muscles of patients with HF (46,47), similar to those found in the failing heart (48). Importantly, changes in anabolic signaling and myosin heavy chain expression precede changes in whole-muscle function in HF patients, providing evidence of their cause-and-effect relationship (49).

Significance of NO

Along with the molecular alterations described above, another factor potentially contributing to the HF-related impairment of skeletal muscle function may be a decrease in NO bioavailability. Although initially identified as a vasodilator, i.e., as "endothelium-derived relaxing factor", NO is in fact a key cellular signaling molecule with pleiotropic effects in many tissues. These include skeletal muscle, wherein among its other effects NO helps modulate contractile function (50,51). Specifically, during isometric contractions NO may (50,51), or may not (52), slightly suppress maximal force production. This may be due to nitrosation or S-nitrosylation of various proteins (53,54). During concentric activity, however, NO significantly increases the rate of force development, maximal shortening velocity, and maximal power of both single muscle fibers and isolated muscles (50,51). These stimulatory effects are thought to be mediated via activation of the classic NO-soluble guanyl cyclase (sGC)-cycle GMP (cGMP) pathway, and have been euphemistically described by Maréchal and Gailly (50) as a "slow-to-fast" shift qualitatively akin to the chronic transformation of muscle fiber type that occurs with, e.g., prolonged electrical stimulation. In failing *cardiac* muscle, however, increased production of reactive oxygen species (ROS) leads to more rapid destruction of NO, and hence reduced NO-sGC-cGMP signaling, which in turn is thought to contribute to reduced contractility in HF (55). Given other parallels between the effects of HF on cardiac and skeletal muscle (including,

somewhat paradoxically, an increase in inducible NOS, or iNOS, expression in both heart (56) and skeletal muscle (57,58)), it is possible that NO bioavailability is also diminished in the *skeletal* muscles of patients with HF, thus contributing to their reduced muscle function as described above.

That HF reduces NO signaling in skeletal muscle has not been directly demonstrated. There is, however, considerable evidence that HF leads to a decline in *whole-body* NO production and/or availability. Katz et al. (59), for example, have demonstrated that HF results in reduced urinary excretion of [¹⁵N]nitrate after infusion of L-[¹⁵N]arginine, indicative of an overall decline in NOS activity. In keeping with this, a number of studies have demonstrated that breath NO levels are lower in HF patients, both at rest (60) and especially during exercise (61,62,63), which is correlated with the magnitude of the impairment in exercise capacity (60,62). It is also well-established that HF leads to endothelial dysfunction in various tissues, including skeletal muscle, as a result of reduced NO production via endothelial NOS (64). These data, along with the central role that skeletal muscle myocytes are now known to play in NO/nitrite (NO₂⁻)/NO₃⁻ metabolism (65,66), strongly suggest that HF reduces NO bioavailability within skeletal muscle, just as it does in the heart.

Dietary NO₃⁻ as a source of NO

If an increase in ROS production and/or down-regulation of NOS-mediated NO synthesis with HF reduces skeletal muscle NO levels and thus contributes to muscle contractile dysfunction, what might be done to treat this problem? One intriguing possibility would be to attempt to increase NO production via the NOS-*independent* enterosalivary pathway (67). This pathway has been reviewed recently in this journal by Chirinos and Zamani (68), and hence will not be discussed in great detail here. In brief, however, in this pathway NO₃⁻ in the diet (found primarily in green leafy vegetables, e.g., spinach, arugula, chard, and also beetroot) is reduced to NO₂⁻ and then to NO. The initial conversion of NO₃⁻ to NO₂⁻ is performed by facultative anaerobic bacteria in the oral cavity, partially on first-pass but primarily after NO₃⁻ has been taken up via the gastrointestinal tract, circulated via the bloodstream, and then concentrated and secreted by the salivary glands. The NO₂⁻ that is formed is then also absorbed and circulated throughout the body, where it can be reduced to NO. A number of molecules have been found capable of catalyzing this second step, with deoxyhemoglobin/deoxymyoglobin, aldehyde oxidase, and xanthine oxidoreductase likely being particularly important in skeletal muscle (66,69,70). Of note, production of NO via this pathway does not require O₂ and proceeds more rapidly under acidic than neutral/basic conditions, which is precisely the opposite of the more well-known NOS pathway. This direct conversion of NO₃⁻ to NO₂⁻ and hence to NO therefore represents a very important mechanism for generating NO “on demand” under hypoxic/acidic conditions, such as commonly occur in contracting skeletal muscle. Indeed, recent research using homogenized rat skeletal muscle has directly demonstrated that acute exercise enhances NO production from NO₂⁻ (66).

Effect of dietary NO₃⁻ on muscle function in HF patients

To test the hypothesis that reduced NO bioavailability contributes to the diminished muscle function found in HF patients, and to evaluate dietary NO₃⁻ as a potential treatment, we recently studied nine middle-aged (average age 57±10 y) men (n=5) and women (n=4) with reduced systolic function (average ejection fraction = 28±11%) (71). Based on the NYHA criteria, the distribution of patients was 1/5/3/0 in classes I/II/III/IV, and all were treated with standard HF medications. Each was studied twice in the fasted state, using a double-blind, placebo-controlled, crossover design. In one trial, patients were tested 2 h after ingesting 140 mL of a concentrated beetroot juice (BRJ) supplement (Beet It Sport[®], James White Drinks, Ipswich, UK) containing 11.2 mmol of NO₃⁻, whereas in the other they were tested 2 h after ingesting the same volume of an NO₃⁻-depleted BRJ placebo that was indistinguishable in terms of color, texture, taste, and smell. Plasma NO₃⁻ and NO₂⁻ and breath NO levels along with heart rate and blood pressure were measured throughout each experiment. Muscle contractile function was assessed using isokinetic dynamometry, with each subject performing 3–4 maximal knee extensions using their dominant leg at angular velocities of 0 (isometric), 1.57, 3.14, 4.71, and 6.28 rad/s (0, 90, 180, 270, and 360 °/s, respectively). This range of velocities was chosen to span the ascending limb of the inverse parabolic power-velocity relationship, thus allowing us to determine (via curve fitting) the maximal velocity (V_{max}) and power (P_{max}) of knee extension based on the peak power (product of peak torque and velocity) achieved at each velocity. Following this testing, the patients also performed 50 maximal knee extensions at a velocity of 3.14 rad/s (180°/s), to determine the possible effects of dietary NO₃⁻ on fatigue resistance during repetitive, dynamic muscle contractions.

We found that ingestion of BRJ containing 11.2 mmol of NO₃⁻ resulted in a significant (P<0.001) elevation in plasma NO₃⁻ concentrations (average = 508±143 μmol/L in the NO₃⁻ trial vs. 29±11 μmol/L in the placebo trial). Average plasma NO₂⁻ concentration also tended to be higher following BRJ ingestion (i.e., 0.57±27 vs. 0.44±0.24 μmol/L), although this difference was not significant due to high variability. On the other hand, the difference in average breath NO levels (i.e., 29±13 vs. 16±5 ppb) was statistically significant (i.e., P<0.01). This increase in whole-body NO bioavailability was accompanied by 9% (P=0.07) and 11% (P<0.05) increases in peak knee extensor torque, and hence power, at the two highest velocities tested, i.e., 4.71 and 6.28 rad/s (270 and 360 °/s), respectively (Table 1). As a result, calculated V_{max} and P_{max} were 12% (P=0.08) and 13% (P<0.05) higher NO₃⁻ ingestion (Figure 1). Six of the nine HF patients were “responders”, with P_{max} increasing by 17±10%, whereas the other three showed minimal change (i.e., -1±5%). Although peak power was initially 6% higher (i.e., 1.78±0.46 vs. 1.68±0.39 W/kg; P<0.05) during the 50 contraction fatigue test, no difference was found in overall work performed (i.e., 28.8±9.8 vs. 28.1±9.5 J/kg; P=0.38) because the patients tended to fatigue somewhat more rapidly following NO₃⁻ ingestion. We also found no differences in heart rate (69±12 vs. 67±12 beats/min; P=0.40) or systolic (105±9 vs. 103±15 mmHg; P=0.76) or diastolic (66±8 vs. 66±8 mmHg; P=0.91) blood pressures.

In summary, the results of this study clearly show that dietary NO₃⁻ ingestion can increase maximal muscle speed and power in patients with systolic HF. In fact, compared to healthy,

observations and to determine the effects of dietary NO_3^- on physical activity levels and quality of life in patients with HF. If proven effective in such studies, dietary NO_3^- supplementation would represent a novel therapy for muscle dysfunction in HF, an important symptom of the disease not presently addressed by any standard medications or therapies.

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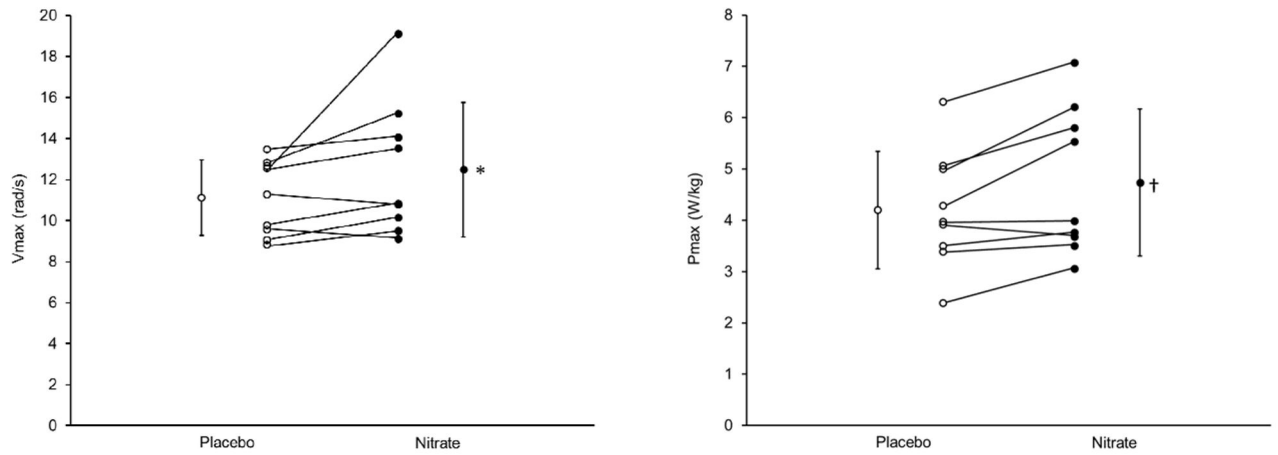


Figure 1. Effect of acute ingestion of 11.2 mmol of nitrate on the maximal speed (Vmax) and power (Pmax) of knee extension in patients with HF due to systolic dysfunction. Placebo; *open circles*. Nitrate; *closed circles*. Values are mean \pm SD; individual results are also shown. *P=0.08; †P<0.05. Data from Ref. 71.

Effect of acute ingestion of 11.2 mmol of nitrate on peak knee extensor torque and power at varying angular velocities in patients with HF due to systolic dysfunction.

Table 1

	<u>Placebo</u>	<u>Nitrate</u>
<u>Angular velocity</u>		
0.00 rad/s	1.98±0.48	1.99±0.31
1.57 rad/s	1.36±0.32	1.38±0.29
3.14 rad/s	1.05±0.25	1.09±0.25
4.71 rad/s	0.85±0.17	0.93±0.22 [*]
6.28 rad/s	0.64±0.22	0.71±0.24 [†]
<u>Angular velocity</u>		
0.00 rad/s	n/a	n/a
1.57 rad/s	2.14±0.51	2.17±0.46
3.14 rad/s	3.29±0.77	3.42±0.79
4.71 rad/s	4.03±0.80	4.38±1.03 [*]
6.28 rad/s	4.02±1.41	4.48±1.53 [†]
Peak torque (Nm/kg)		
Peak power (W/kg)		

Data are mean ± S.D. for n=9.

^{*} P<0.07;

[†] P<0.05.

Data from Ref. 71.