

Evaluating the gray and white matter energy budgets of human brain function

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Abstract

The insatiable appetite for energy to support human brain function is mainly supplied by glucose oxidation ($CMR_{glc(ox)}$). But how much energy is consumed for signaling and nonsignaling processes in gray/white matter is highly debated. We examined this issue by combining metabolic measurements of gray/white matter and a theoretical calculation of bottom-up energy budget using biophysical properties of neuronal/glia cells in conjunction with species-exclusive electrophysiological and morphological data. We calculated a $CMR_{glc(ox)}$ -derived budget and confirmed it with experimental results measured by PET, autoradiography, ^{13}C -MRS, and electrophysiology. Several conserved principles were observed regarding the energy costs for brain's signaling and nonsignaling components in both human and rat. The awake resting cortical signaling processes and mass-dependent nonsignaling processes, respectively, demand $\sim 70\%$ and $\sim 30\%$ of $CMR_{glc(ox)}$. Inhibitory neurons and glia need 15–20% of $CMR_{glc(ox)}$, with the rest demanded by excitatory neurons. Nonsignaling demands dominate in white matter, in near opposite contrast to gray matter demands. Comparison between ^{13}C -MRS data and calculations suggests ~ 1.2 Hz glutamatergic signaling rate in the awake human cortex, which is ~ 4 times lower than signaling in the rat cortex. Top-down validated bottom-up budgets could allow computation of anatomy-based $CMR_{glc(ox)}$ maps and accurate cellular level interpretation of brain metabolic imaging.

Keywords

Aerobic glycolysis, spiking rate, astrocyte, glutamate, electroencephalography, lactate

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Introduction

The human brain is extremely energy demanding compared to other organs in the body¹ to support the conscious state.² The human brain is estimated to require 10–20 watts, while a few million watts is needed to power supercomputers.³ A highly energy efficient mammalian brain implies optimal organization of cortical circuits for cognitive functions.⁴ Recent experimental and theoretical research suggest that multiple factors such as optimized ion-channel kinetics, axon myelination, and warmer body temperature play key roles in minimizing the energy cost for action potential generation and propagation.^{5–11} It is also suggested that brain circuitries are organized in an energy-optimized way.⁴

These and other recent results highlight the importance of a more accurate brain energy budget calculation^{12–14} to promote a better understanding of the

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energetic basis of brain computation. Furthermore, understanding the relationship between energetics and function of the brain is crucial to the interpretation of functional magnetic resonance imaging (fMRI) studies of the human brain. The understanding of fMRI studies is based on experimental observations that brain energy metabolism from glucose oxidation ($CMR_{glc(ox)}$) changes in a correlated manner with signaling processes,^{15–18} which include action potential, synaptic transmission, glutamate or GABA recycling, calcium activities, and hemodynamics through neurovascular coupling.¹⁹ Nonsignaling processes are generally assigned to housekeeping needs (e.g. biosynthesis or proton leaks) and maintenance of resting cell membrane potentials, which generally do not change significantly with brain activity. At physiological temperatures, action potentials generally take around 2 ms duration. For low firing rates (<10 Hz), neuronal membranes spend less than 2% of the spiking duty cycle at the action potential level and are at the resting potential level for over 98% of the time. Therefore, for the range of firing rates modeled in this paper, we approximate the membrane potential during inter-signaling periods as being a constant at the resting potential value. However, there is still uncertainty regarding the cost for signaling and nonsignaling components in mammalian brain, especially in humans.²⁰ This uncertainty in nonsignaling processes also inherently implies ambiguity for signaling components, and thus makes quantitative interpretation of brain fMRI signals difficult.

Here, we construct a comprehensive bottom-up $CMR_{glc(ox)}$ -derived energy budget for human gray and white matter based on biophysical properties of neuronal and glial cells, together in conjunction with electrophysiological and morphological data from human and rat brain. While similar bottom-up approaches have been used previously,^{12,14,20–22} our approach differs in that we use extensive top-down validation, and associated our bottom-up calculations with a range of behavioral or metabolic states (e.g. anesthetized, awake, sleep, etc.) measured by various experimental techniques. Furthermore, we update the biophysical and physiological parameters previously used in bottom-up budgets for both rat and human brain. We compare $CMR_{glc(ox)}$ computed from the budget with $CMR_{glc(ox)}$ from experimental measurements (by [¹⁸F]-fluorodeoxyglucose (FDG) positron emission tomography (PET), ¹³C magnetic resonance spectroscopy (¹³C-MRS), [¹⁴C]-2-deoxyglucose (2DG) autoradiography). We also compare the neuronal activity computed from the budget with neuronal activity from experimental measurements (by firing rates from extracellular recordings in rats and bispectral index (BIS) from electroencephalography (EEG) recordings

in humans), allowing investigations of both signaling processes associated with ion flux and mass-dependent nonsignaling processes. This top-down validation of bottom-up energy budgets for gray and white matter may help provide solutions for several open issues, e.g. how the signaling and nonsignaling components interact with each other to underlie the metabolic data collected by methods like fMRI, PET, and ¹³C-MRS in the human brain.

Materials and methods

The budget details are in SI Text, Sections (a) to (f), where the important parameters are shown in Table 1. Note that all these parameters are based on more recent experimental reports and computational estimations, where we also did calculations to confirm some of these parameters. For example, recent *in vivo* ¹³C-MRS studies in rat and human brain suggest that 70–75% of the energy in the awake state supports cortical signaling functions, whereas the remaining 25–30% fraction is attributed to nonsignaling functions.¹³ Previous bottom-up budget estimates range from 20% to 50% of the ATP derived from $CMR_{glc(ox)}$ supporting rat cerebral cortex nonsignaling functions.²⁰ This large uncertainty in nonsignaling functions is mainly due to the lack of experimental evidence of specific cellular physiologic and metabolic processes. We used the allometry theory of metabolism for individual cells to estimate the housekeeping cost by computing averaged mass value of individual cells and mass-dependent metabolic rate^{23–25} (see SI Text, Section A.1).

In addition, to our knowledge, all the previous bottom-up budgets did not include glial activity.^{12,14,21,22} However, as reported in recent studies,^{26,27} astrocytes in gray matter as well as oligodendrocyte precursor cells (OPCs) and oligodendrocytes in white matter have active calcium responses related to spiking process through at least two different mechanisms, i.e. (i) transmembrane calcium influx via voltage-operated calcium channels, and (ii) calcium release from IP3-sensitive internal calcium stores. Hence, it is necessary to calculate energy cost for the calcium-related activities of glia. Furthermore, we accounted for billions of glial cells and unmyelinated axons in white matter²⁵ (see SI Text, Section B for details). Considering that the large number of glial cells may contribute to a non-negligible portion of the brain budget, we carefully considered glial cells in our bottom-up budget calculation for both gray and white matter (see SI Text, Sections A and B for details).

Moreover, the total axonal length of a human neuron – 7.56 cm – was calculated based on a more recently established relation of cross-species

Table 1. Comparison of energy budget “input” parameters, which include morphological (e.g. cell number, cell mass, cell size, etc.) and biophysical (e.g. membrane resistance and capacitance, etc.) components of individual cell types.

Parameters	Budget parameters		SI text
	Current	Previous	
Number cells ($N_x^{y'}$, units of billions of cells) for human brain			
N_n^{gm}	13.64 (ref. ²⁵)		Section D
N_g^{gm}	19.15 (ref. ²⁵)		Section D
N_g^{wm}	41.69 (ref. ²⁵)		Section D
N_n^{wm} unmyelinated axon	2.71 (ref. ²⁵)		Section B
N_{axon}^{wm} myelinated axon	9.43 (ref. ²¹)		Section B
$N_{astrocyte}^{wm}$	1.48 (ref. ²¹)		Section B
N_{OPC}^{wm}	4.28 (ref. ²¹)		Section B
$N_{oligodendrocyte}^{wm}$	35.9 (ref. ²¹)		Section B
Mass of tissues (M_x) in units of g and cells (m_x) in units of ng for human brain			
M_{gm}	638.37 g (ref. ²⁵)	475 g (ref. ²²)	Section E
M_{wm}	593.89 g (ref. ²⁵)		Section E
m_n	29.5 ng (ref. ²⁵)		Section F
m_g	12.3 ng (ref. ²⁵)		Section F
Cellular anatomy by length (L_x) and diameter (D_x) in units of cm, mm, or μm			
L_{axon} human neuron	7.56 cm (ref. ²⁸)	10 cm (ref. ²²)	Section A.3
$L_{dendrite}$ human neuron	0.84 cm (ref. ²⁸)	1.1 cm (ref. ²²)	Section A.3
L_{axon} rat neuron	4 cm (ref. ¹⁴)	4 cm (ref. ¹⁴)	Section A.3
$L_{dendrite}$ rat neuron	0.44 cm (ref. ¹⁴)	0.44 cm (ref. ¹⁴)	Section A.3
D_{axon} human/rat neuron	0.3 mm (ref. ^{14,22})	0.3 mm (ref. ^{14,22})	Section A.3
$D_{dendrite}$ human/rat neuron	0.9 μm (ref. ^{14,22})	0.9 μm (ref. ^{14,22})	Section A.3
Cellular density (ρ) for gray matter (GM) in units of millions of cells/g			
Human GM neuronal ρ	21.4 (ref. ²⁵)	40 (ref. ²²)	18.3 (ref. ¹³) Section A.3
Rat GM neuronal ρ	40.3 (ref. ⁵⁶)	92 (ref. ²¹)	47.5 (ref. ^{13, 2}) Section A.3
Human GM glial ρ	30.0 (ref. ²⁵)	38 (ref. ²²)	18.3 (ref. ¹³) Section A.3
Rat GM glial ρ	59.4 (ref. ⁵⁶)	92 (ref. ²¹)	47.5 (ref. ^{13,32}) Section A.3
Membrane resistance (R_m , $M\Omega$) and capacitance (C_m , $\mu\text{F/cm}$) in gray (GM) and white (WM) matter			
R_m pyramidal cell	100 (ref. ⁵⁷)	74 (ref. ¹³)	200 (ref. ¹⁴) Section B
R_m interneuron	200 (ref. ^{58,59})	74 (ref. ¹³)	Section B
R_m glial cell	200 (ref. ⁶⁰)	74 (ref. ¹³)	10,500 (ref. ⁶¹) Section B
R_m astrocyte in WM	560 (ref. ²¹)	560 (ref. ²¹)	Section B
R_m OPC in WM	800 (ref. ²¹)	800 (ref. ²¹)	Section B
R_m oligodendrocyte in WM	200 (ref. ²¹)	200 (ref. ²¹)	Section B
R_m myelinated axon in WM	0.55×10^3 (ref. ²¹)	0.55×10^3 (ref. ²¹)	Section B
C_m for axon	1 (ref. ^{12,21})	1 (ref. ^{12,21})	Section B
C_m for myelinated axon	0.08 (ref. ^{12,21})	0.08 (ref. ^{12,21})	Section B
Na^+ entry ratio (ε) and oxygen-to-glucose (OGI) index; both without units			
ε pyramidal	1.5 (ref. ⁸)	1.3 (ref. ^{12,21})	Section A.3
ε interneuron	2.3 (ref. ⁶)	2.3 (ref. ^{12,21})	Section A.3
OGI	5.3 (ref. ¹³)	5.3 (ref. ¹³)	Section A.1

neuroanatomical data.^{25,28,29} Indeed, with this value (see Results), the calculated energy budget matches the PET data better than axon length of 10 cm, as reported by a previous study.³⁰ In addition, a new cell

counting method^{25,31} estimated that human gray matter neuronal density is 21.4 million/g and glial cell density is 30 million/g, which are both lower than 40 and 38 million/g, respectively, in previous rough estimations

which are based on extrapolation of cell counts from localized measurements.³⁰ These new methods were also able to estimate that rat cortical neuronal and glial cell densities are 40.3 and 59.4 million/g, which are slightly different from earlier reports.³²

The energy budgets for neuronal and glial cells in gray matter (SI Text, Section A) and white matter (SI Text, Section B) of the human brain were calculated separately, using cellular biophysical properties with electrophysiological and morphological data of human brain. The same principles, but scaled according to species differences, were then applied for energy budgets for neurons and glial cells in gray matter and white matter of the rat brain (SI Text, Section C). All the “output” parameters of the energy budget, both for rat and human, are listed in Table 2. Specifically for the human cerebrum, we estimated the number of neuronal and glial cells in gray and white matter (SI Text, Section D), masses of gray and white matter (SI Text, Section E), and masses of individual neuron and glial cells in gray and white matter (SI Text, Section F). We found that that gray matter and white matter weigh roughly the same, but there is a significantly larger number of glial cells (19.2 and 41.7 billion, respectively, in gray and white matter) than neurons (13.6 and 2.7 billion, respectively, in gray and white matter) and an individual neuron weighs more than twice the mass of a glial cell (SI Text, Sections D to F). The cerebellum was excluded in our budget.

Results

We first validated energy budgets for gray matter of rat and human brain to identify unifying principles of neuronal and glial cells. We then applied these validated components for estimating white matter energetics in rat and human brain.

Gray matter budget validation from *in vivo* PET, 2DG, and electrophysiology measurements

The total ATP consumption rate in human gray matter (E_{gm} ; SI Text, Section A; equations S1 to S12) contains nonsignaling ($E_{nonsignaling}^{gm}$) and signaling ($E_{signaling}^{gm}$) processes. $E_{nonsignaling}^{gm}$ includes energy demands for house-keeping mechanisms (E_{HK}^{gm}) and resting potential maintenance (E_{RP}^{gm}) of all cell types, whereas $E_{signaling}^{gm}$ depends on rate of neural firing (f) and includes energy demands for action potential conduction ($E_{AP}^{gm}(f)$), synaptic transmission ($E_{ST}^{gm}(f)$), glutamate or GABA recycling ($E_{glu}^{gm}(f)$, $E_{GABA}^{gm}(f)$), and calcium activity ($E_{Ca}^{gm}(f)$) of all the neuronal and glial cells, respectively. Based on the available metabolic and neurophysiologic data in rat (Table S1) and human (Table S2) brain, we calculated E_{HK}^{gm} and E_{RP}^{gm} of

1.1×10^{19} ATP/s and 1.3×10^{19} ATP/s, respectively (using equations S2 to S6), which together make $E_{nonsignaling}^{gm} = 2.4 \times 10^{19}$ ATP/s (Table 2). We assumed a fixed relationship for firing rates of excitatory (f_e ; see equation S2(a)) and inhibitory (f_i ; see equation S2(b)) neurons based on limited *in vivo* data of somatosensory cortex³³ (see SI Text, Section A), but this can be amended based on future f_e and f_i values measured across brain regions. Likewise, for $f = 1$ Hz, we calculated $E_{AP}^{gm}(f)$, $E_{ST}^{gm}(f)$, $E_{glu}^{gm}(f)$, and $E_{Ca}^{gm}(f)$ (using equations S3 to S6) of 0.37×10^{19} ATP/s, 2.28×10^{19} ATP/s, 0.2×10^{19} ATP/s, and 0.34×10^{19} ATP/s, respectively (Table 2). These terms together contribute to $E_{signaling}^{gm}$ (4.6×10^{19} ATP/s for $f = 1.15$ Hz). So the total gray matter cost (E_{gm}) for $f = 1$ Hz, which is given by sum of $E_{nonsignaling}^{gm}$ and $E_{signaling}^{gm}$, is 7.0×10^{19} ATP/s. Comparison of these terms show that $E_{ST}^{gm}(f)$ is by far the largest contributor to E_{gm} , where the vesicle transmitter releases probability also contributes to E_{glu}^{gm} (ρ_{ves} in equations S10 and S11). Given that ρ_{ves} is dependent on ambient temperature, we used more physiologically relevant ρ_{ves} values than used in previous studies¹⁴ (see SI Text, Section A.4). However, exocytic release of neurotransmitters from synaptic vesicles at the nerve terminal and subsequent retrieval of membranes that make up synaptic vesicles is likely to be energy free as lipids are completely recycled in this process.³⁴ Moreover, it is noted that $E_{nonsignaling}^{gm}$ is solely dependent on number of cells (i.e. mass of brain tissue), whereas $E_{signaling}^{gm}$ relies on both brain tissue mass and neuronal firing rate. We converted E_{gm} (units of ATP/s) to total glucose oxidation ($\text{calcCMR}_{glc(ox),T}$; units of $\mu\text{mol/g/min}$) by

$$\text{calcCMR}_{glc(ox),T} = hE_{gm} \quad (1a)$$

where h depends on the glucose-to-ATP transduction rate (GTR) which is given by $38 \times (\text{OGI}/6)$ and OGI is the measured oxygen-to-glucose index that is determined by the ratio of cerebral metabolic rates of oxygen (CMR_{O_2}) and glucose (CMR_{glc}) consumption (i.e. $\text{OGI} = \text{CMR}_{O_2}/\text{CMR}_{glc}$). Experimental measures indicate that glucose is not fully oxidized, resulting in $\text{GTR} = 33.6$ with $\text{OGI} = 5.3$.^{35,36} Thus, h is given by $6 \times 10^7 / (\text{GTR} M_{gm} A_{vo})$, where M_{gm} is the total mass of gray matter ($M_{gm} = 638$ g is used here,²⁵ Table 1) and A_{vo} is the Avogadro constant (6.023×10^{23} /mol). Because E_{gm} is the sum of $E_{nonsignaling}^{gm}$ and $E_{signaling}^{gm}$, then

$$\text{calcCMR}_{glc(ox),nonsignaling} = hE_{gm}^{nonsignaling} \quad (1b)$$

$$\text{calcCMR}_{glc(ox),signaling} = hE_{gm}^{signaling} \quad (1c)$$

Table 2. Comparison of energy budget “output” parameters, which include energies of individual cell types (e.g. neuronal and glial cells), energies of all cells (e.g. in gray and white matter), and neuronal firing rate.

Parameters	Budget parameters			SI Text
	Current	Previous		
Energy of individual cells (E_x^y , units of ATP/s) for gray (GM) and white (WM) matter; $f_e = 1$ Hz and $f_i = 2$ Hz				
$E_{\text{signaling}}^e$ Human GM	2.1×10^9	4.81×10^9 (ref. ¹³)	6.1×10^9 (ref. ²²)	Section A
$E_{\text{nonsignaling}}^e$ Human GM	1.22×10^9	0.92×10^9 (ref. ¹³)	0.86×10^9 (ref. ²²)	Section A
$E_{\text{signaling}}^i$ Human GM	2.87×10^9			Section A
$E_{\text{nonsignaling}}^i$ Human GM	0.88×10^9			Section A
E_{HK}^e Human GM	5.38×10^8			Section A.1
E_{HK}^i Human GM	3.55×10^8			Section A.1
E_{HK}^g Human GM	1.37×10^8			Section A.1
E_{RP}^e Human GM	6.82×10^8	3.42×10^8 (ref. ¹⁴)	8.6×10^8 (ref. ²²)	Section A.2
E_{RP}^i Human GM	3.41×10^8			Section A.2
E_{RP}^g Human GM	2.54×10^8	1.02×10^8 (ref. ¹⁴)	3.1×10^8 (ref. ²²)	Section A.2
E_{AP}^e Human GM	2.65×10^8		2.4×10^9 (ref. ²²)	Section A.3
E_{AP}^i Human GM	3.09×10^8			Section A.3
E_{ST}^e Human GM	15.8×10^8		3.18×10^9 (ref. ²²)	Section A.4
E_{ST}^i Human GM	20.5×10^8			Section A.4
E_{glu}^e Human GM	1.41×10^8		2.4×10^8 (ref. ²²)	Section A.5
E_{GABA}^i Human GM	1.82×10^8			Section A.5
E_{Ca}^e Human GM	1.16×10^8		3.05×10^8 (ref. ²²)	Section A.6
E_{Ca}^i Human GM	2.31×10^8			Section A.6
E_{Ca}^g Human GM	0.924×10^8			Section A.6
E_e Human GM	3.32×10^9	5.73×10^9 (ref. ¹³)	6.96×10^9 (ref. ²²)	Section A
E_i Human GM	3.75×10^9			Section A
E_g Human GM	5.69×10^8			Section A
$E_{\text{HK}}^{\text{astrocyte}}$ Human WM	2.23×10^8			Section B
$E_{\text{HK}}^{\text{OPC}}$ Human WM	2.23×10^8			Section B
$E_{\text{HK}}^{\text{oligodendrocyte}}$ Human WM	2.23×10^8			Section B
$E_{\text{RP}}^{\text{astrocyte}}$ Human WM	9.05×10^8			Section B
$E_{\text{RP}}^{\text{OPC}}$ Human WM	8.51×10^8			Section B
$E_{\text{RP}}^{\text{oligodendrocyte}}$ Human WM	3.4×10^8			Section B
$E_{\text{Ca}}^{\text{astrocyte}}$ Human WM	0.92×10^8			Section B
$E_{\text{Ca}}^{\text{oligodendrocyte}}$ Human WM	0.92×10^8			Section B
$E_{\text{signaling}}^e$ Rat GM	1.58×10^9	4.81×10^9 (ref. ¹³)	0.71×10^9 (ref. ¹⁴)	Section A, C
$E_{\text{nonsignaling}}^e$ Rat GM	1.17×10^9	0.92×10^9 (ref. ¹³)	0.34×10^9 (ref. ¹⁴)	Section A, C
E_e Rat GM	2.75×10^9	5.73×10^9 (ref. ¹³)	1.05×10^9 (ref. ¹⁴)	Section A, C
E_i Rat GM	2.78×10^9			Section A, C
E_g Rat GM	0.924×10^8			Section A, C
Energy of all cells (E_x^y , units of ATP/s) in whole cerebrum with awake resting firing rate.				
$E_{\text{nonsignaling}}^{\text{wm}}$	2.65×10^{19}			Section B
$E_{\text{signaling}}^{\text{wm}}$	1.36×10^{19}			Section B
$E_{\text{nonsignaling}}^{\text{gm}}$	2.4×10^{19}	5.18×10^{19} (ref. ²¹)		Section A
$E_{\text{signaling}}^{\text{gm}}$	4.6×10^{19}	7.77×10^{19} (ref. ²¹)		Section A
E_{wm}	4.01×10^{19}			Section B
E_{gm}	7.0×10^{19}	12.95×10^{19} (ref. ²¹)		Section A
Cortical firing rate of excitatory neurons in awake resting state (f_e , units of Hz)				
f_e Human	1.15 Hz	0.024 (ref. ²²)	0.5-2 (ref. ^{42,43})	Section A
f_e Rat	4.3 Hz	4 Hz (ref. ¹²)		Section C

where the total glucose oxidation is ($\text{calcCMR}_{\text{glc(ox),T}}$) given by the sums of signaling ($\text{calcCMR}_{\text{glc(ox),signaling}}$) and nonsignaling ($\text{calcCMR}_{\text{glc(ox),nonsignaling}}$) components

$$\text{calcCMR}_{\text{glc(ox),T}} = \text{calcCMR}_{\text{glc(ox),nonsignaling}} + \text{calcCMR}_{\text{glc(ox),signaling}} \quad (1d)$$

and neuronal ($\text{calcCMR}_{\text{glc(ox),N}}$) and astrocytic ($\text{calcCMR}_{\text{glc(ox),A}}$) components

$$\text{calcCMR}_{\text{glc(ox),T}} = \text{calcCMR}_{\text{glc(ox),N}} + \text{calcCMR}_{\text{glc(ox),A}} \quad (1e)$$

We compared $\text{calcCMR}_{\text{glc(ox),T}}$ with experimental measurements of CMR_{glc} in the rat (Table S1, by 2DG autoradiography) and human (Table S2, by FDG PET) brain. These were converted to total glucose oxidation rate ($\text{measCMR}_{\text{glc(ox),T}}$) by

$$\text{measCMR}_{\text{glc(ox),T}} = (\text{OGI}/6) \times \text{CMR}_{\text{glc}} \quad (2a)$$

$$\text{measCMR}_{\text{glc(ox),T}} = \text{measCMR}_{\text{glc(ox),N}} + \text{measCMR}_{\text{glc(ox),A}} \quad (2b)$$

where OGI was assumed to be 5.3 (see above) and the $\text{measCMR}_{\text{glc(ox),T}}$ is the sum of neuronal ($\text{measCMR}_{\text{glc(ox),N}}$) and astrocytic ($\text{measCMR}_{\text{glc(ox),A}}$) components, both of which can be measured by ^{13}C -MRS. Figure 1(a) compares values of $\text{calcCMR}_{\text{glc(ox),T}}$ and $\text{measCMR}_{\text{glc(ox),T}}$ for different behavioral states in both rat and human brain. The goodness of fit between $\text{calcCMR}_{\text{glc(ox),T}}$ and $\text{measCMR}_{\text{glc(ox),T}}$ is indicated by the line with an R^2 value of 0.99 for 10 behavioral states in rat and 6 behavioral states in human. We obtained the calculated firing rate (calcRate , unit of Hz) based on the relationship between $E_{\text{signaling}}^{\text{gm}}$ and its components (using equation S4(c); S1 Text, Section A)

$$\text{calcRate} = E_{\text{signaling}}^{\text{gm}} / (E_{\text{gm}}^{\text{AP}}(f) + E_{\text{gm}}^{\text{ST}}(f) + E_{\text{gm}}^{\text{glu}}(f) + E_{\text{gm}}^{\text{Ca}}(f)) \quad (3)$$

where the denominator terms were estimated for $f = 1$ Hz (Table 2) so that we could compare with measured neural activity rates (measRate for rat, measActivity for human). Recordings of neural activity were available for the rat (Table S1) and human (Table S2) brain under the same behavioral states as the metabolic measurements (see above). In the rat and human brain, respectively, the measRate for rat (f in units of Hz in Table S1) was represented by neuronal firing rates (f in units of Hz) and BIS-measured EEG recordings (f_{BIS} in units of BIS for human in Table S2), Figure 1(b) shows good agreement between

calcRate and measRate for 10 different behavioral states in rats (i.e. Pearson's correlation coefficient of 0.79 and $R^2 = 0.62$ with fit slope equal to 0.955). Human brain EEG, which measures summed activity of post-synaptic currents,³⁷ indicates that brain activity changes with different behavioral states. Figure 1(c) shows excellent agreement between calcRate with measActivity for six human behavioral states (i.e. Pearson's correlation coefficient of 0.98 and $R^2 = 0.96$). Overall, both in rat and human gray matter, Figures 1(a) to (c) emphasize the need for ATP yielded from glucose oxidation to support higher signaling activity in cerebral cortex.

Figures 1(d) to (f) are quite similar in the sense that the values of $\text{calcCMR}_{\text{glc(ox),T}}$, $\text{calcCMR}_{\text{glc(ox),N}}$, and $\text{calcCMR}_{\text{glc(ox),A}}$ (but each normalized to the value of the awake resting state $\text{calcCMR}_{\text{glc(ox),T}}$) of different behavioral states for both rat and human are plotted against measured neuronal activity rates (normalized to the awake resting state activity). The intercepts in Figure 1(d) to (f) are all nonzero as activity approached zero (i.e. absence of signaling): around 34% of awake resting state $\text{calcCMR}_{\text{glc(ox),T}}$ for total cellular non-signaling cost (Figure 1(d); Pearson's correlation coefficient 0.98 and $R^2 = 0.97$); 22% of awake resting state $\text{calcCMR}_{\text{glc(ox),T}}$ for neuronal nonsignaling cost (Figure 1(e); Pearson's correlation coefficient 0.98 and $R^2 = 0.97$), and 9% of awake resting state $\text{calcCMR}_{\text{glc(ox),T}}$ for astrocytic nonsignaling cost (Figure 1(f); Pearson's correlation coefficient 0.93 and $R^2 = 0.87$). The slopes in Figure 1(d) and (e) were significantly greater than the slope in Figure 1(f), suggesting that energy demand of neuronal activity is about 6 times greater than cost of glial activity.

Gray matter budget validation from in vivo ^{13}C -MRS measurements

^{13}C -MRS provides independent measures of neuronal ($\text{CMR}_{\text{glc(ox),N}}$) and astrocytic ($\text{CMR}_{\text{glc(ox),A}}$) components of glucose oxidation, but also independent measures of neuronal activity of glutamatergic (excitatory) and GABAergic (inhibitory) synaptic activity and their independent energy demands.³⁶ Thus, the gray matter energetics calculated for rat and human brain can also be validated by comparison with prior ^{13}C -MRS data acquired in rat and human brain (Tables S3 to S5).

Figure 2(a) is similar to Figure 1(e) except that the normalizing factor of the y-axis differs (i.e. divided by $\text{calcCMR}_{\text{glc(ox),N}}$ and $\text{calcCMR}_{\text{glc(ox),T}}$, respectively, in Figures 2(a) and 1(e)). The intercept on the vertical axis is ~25% of awake resting state $\text{calcCMR}_{\text{glc(ox),N}}$ for both species (Figure 2(a); Pearson's correlation coefficient = 0.96 and $R^2 = 0.92$). This indicates that the total energy for signaling related demands is 75–78% of

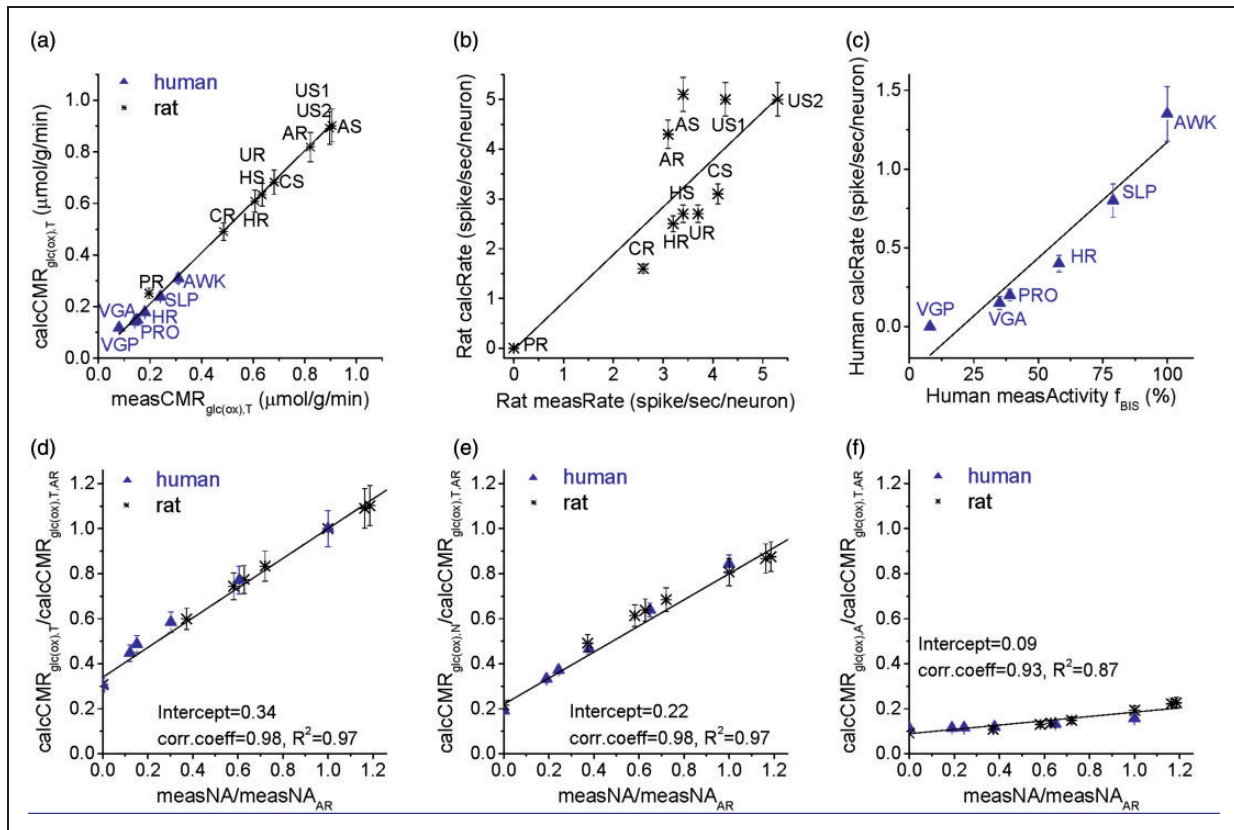


Figure 1. Relationship between $CMR_{glc(ox)}$ and neuronal activity. (a) Comparison between cortical values of calculated total $CMR_{glc(ox)}$ ($calcCMR_{glc(ox),T}$) and measured total $CMR_{glc(ox)}$ ($measCMR_{glc(ox),T}$), where $CMR_{glc(ox),T}$ is defined as the sum neuronal ($CMR_{glc(ox),N}$) and glial ($CMR_{glc(ox),A}$) components. Values of $measCMR_{glc(ox),T}$ for rat (asterisk) and human (blue triangles) brain were derived from 2-deoxyglucose (2DG) autoradiography and fluoro-2-deoxyglucose (FDG) PET, respectively, in rat (Table S1) and human (Table S2) brain. The abbreviated labels are: PR: pentobarbital; US: urethane stimulation; AR: awake rest; AS: awake stimulation; UR: urethane rest; US2: urethane stimulation; CR: α -chloralose rest; CS: α -chloralose stimulation; HR: halothane rest; HS: halothane stimulation; for human data, VGP: persistent vegetative; VGA: acute vegetative; PRO: propofol; SEV: sevoflurane; HR: halothane rest; SLP: non-REM sleep; AWK: awake. The relationship between $calcCMR_{glc(ox),T}$ and $measCMR_{glc(ox),T}$ is fitted by $y = -0.013 + 0.99x$, with an R^2 value of 0.99. (b) In rat somatosensory cortex, comparison between calculated and measured neuronal activity for the rat (asterisk), where the calculated cortical mean firing rates per neuron ($calcRate$) for each value of $calcCMR_{glc(ox),T}$ in Figure 1(a) is plotted versus measured average neuronal firing rate per neuron ($measRate$). Experimental conditions and values for $measRate$ in the rat are listed in Table S1. The relationship between $calcRate$ and $measRate$ is fitted by $y = -0.03 + 0.955x$, with an R^2 value of 0.62. Note that the $calcRate$, similar to $measRate$, range is from 0 to 5 Hz for all conditions shown in the rat. (c) In human visual cortex, comparison between calculated ($calcRate$) and measured ($measActivity$) neuronal activity (triangles), where the values of $calcRate$ for each value of $calcCMR_{glc(ox),T}$ in Figure 1(a) is plotted versus $measActivity$ given by EEG-derived bispectral index (BIS, f_{BIS}). Experimental conditions and values for human $measActivity f_{BIS}$ are listed in Table S2. The relationship between $calcRate$ and $measActivity$ is fitted by $y = -0.297 + 0.0198x$, with an R^2 value of 0.96. Note that the $calcRate$ in human, different from to $calcRate$ in rat, range is 0–2 Hz for all conditions shown. (d) Values of $calcCMR_{glc(ox),T}$ for both rat (asterisk) and human (triangles) as a function of measured neuronal activity ($measNA$), which is fitted by a linear function $y = 0.34 + 0.66x$, with an R^2 value of 0.97. Values of $measNA$ for rat and human brain are from Tables S1 to S2, respectively. The horizontal and vertical axes are normalized to the awake resting state values of $measNA$ and $calcCMR_{glc(ox),T}$, respectively (i.e. $measNA_{AR}$ and $calcCMR_{glc(ox),T,AR}$). (e) Values of calculated neuronal $CMR_{glc(ox)}$ ($calcCMR_{glc(ox),N}$) in both rat (asterisk) and human (triangles) brain as a function of $measNA$, which is fitted by a function $0.22 + 0.58x$, with an R^2 value of 0.97. The horizontal and vertical axes are normalized to the awake resting state values of $measNA$ and $calcCMR_{glc(ox),T}$, respectively (i.e. $measNA_{AR}$ and $calcCMR_{glc(ox),T,AR}$). (f) Values of calculated astrocytic $CMR_{glc(ox)}$ ($calcCMR_{glc(ox),A}$) in both rat (asterisk) and human (triangles) brains as a function of $measNA$, which is fitted by a function $y = 0.09 + 0.096x$, with an R^2 value of 0.87. The horizontal and vertical axes are normalized to the awake resting state values of $measNA$ and $calcCMR_{glc(ox),T}$, respectively (i.e. $measNA_{AR}$ and $calcCMR_{glc(ox),T,AR}$). See Figure S2(a) and (b) for separation of the signaling and nonsignaling components of $calcCMR_{glc(ox),T}$ for all of the behavioral states in both species.

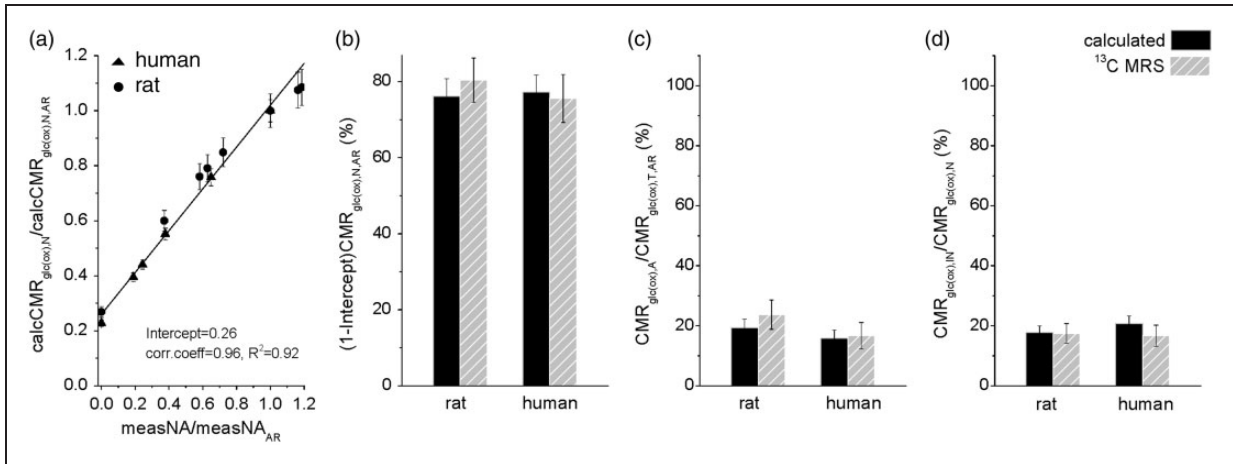


Figure 2. Comparison of $CMR_{glc(ox)}$ calculated from energy budget with experimentally measured $CMR_{glc(ox)}$ by ^{13}C -MRS. (a) Calculated neuronal $CMR_{glc(ox)}$ ($calcCMR_{glc(ox),N}$) in both rat (asterisk) and human (triangles) brain as a function of measured neuronal activity ($measNA$), which is fitted by a function $0.26 + 0.76x$, with an R^2 value of 0.92. The horizontal and vertical axes are normalized to the awake resting state values of $measNA$ and $calcCMR_{glc(ox),N}$, respectively (i.e. $measNA_{AR}$ and $calcCMR_{glc(ox),T,AR}$). Since values above the intercept reflect signaling (i.e. $measNA > 0$), this plot suggests that nonsignaling-dependent energy demand of neurons is $\sim 26\%$ of $CMR_{glc(ox),N,AR}$. This intercept derived from 2DG autoradiography, FDG PET, and electrophysiology data (Tables S1 and S2) is very similar to prior observations with ^{13}C -MRS in rat and human brain (Figure S1 and Table S3). (b) The signaling-dependent $CMR_{glc(ox)}$ of neuron populations (i.e. $(1-\text{intercept}) \times CMR_{glc(ox),N,AR}$ from Figure 2(a)) for rat and human cortex calculated (black bars) and measured by ^{13}C -MRS (gray bars) shows that the signaling-dependent energy demand of neurons is 75–80% of $CMR_{glc(ox),N,AR}$. The $CMR_{glc(ox)}$ experimental values for ^{13}C -MRS in the rat and human are shown in Figure S1 and Table S3. (c) The fraction of astrocytic $CMR_{glc(ox)}$ ($CMR_{glc(ox),A}$) in relation to $CMR_{glc(ox),T,AR}$ (i.e. $CMR_{glc(ox),A}/CMR_{glc(ox),T,AR}$) for calculated (black bars) and measured by ^{13}C -MRS (gray bars) shows that the glial energy demand is $\sim 20\%$ of $CMR_{glc(ox),T,AR}$. The glial experimental values for ^{13}C -MRS in the rat and human are shown in Table S4. (d) The fraction of inhibitory (GABAergic) neuronal $CMR_{glc(ox)}$ ($CMR_{glc(ox),IN}$) in relation to $CMR_{glc(ox),N}$ (i.e. $CMR_{glc(ox),IN}/CMR_{glc(ox),N}$) calculated (black bars) and measured by ^{13}C -MRS (gray bars) shows that the inhibitory (GABAergic) neuronal energy demand is $\sim 20\%$ of $CMR_{glc(ox),N}$. The GABAergic experimental values for ^{13}C -MRS in the rat and human are shown in Table S5.

neuronal metabolism and nonsignaling neuronal components (including resting potential and housekeeping) occupy 22–25% of total neuronal metabolism (Figure 2(b), black). These results from non-MRS based methods match extremely well with results from ^{13}C -MRS (Table S3; Figure S1), where the signaling neuronal demands occupy 75–80% of total neuronal metabolism and the nonsignaling neuronal components demand about 20–25% of total neuronal metabolism (Figure 2(b), gray).

Similarly, the *in vivo* ^{13}C -MRS experimental results of $CMR_{glc(ox),A}/CMR_{glc(ox),T}$ ratio in the awake state for both species (Table S4), showed that cortical glial cells in the awake state demand about 17–22% of total oxidative ATP (Figure 2(c), gray). The calculations provided matching results, where astrocytes in cortex demand 16–19% of total oxidative ATP (Figure 2(c), black). Finally, energy demand of GABAergic synapses ($CMR_{glc(ox),IN}$) in relation to $CMR_{glc(ox),N}$ has been measured in both rat and human brain (Table S5). The ^{13}C -MRS measured $CMR_{glc(ox),IN}/CMR_{glc(ox),N}$ ratio in the awake state for both species is 17–18% of oxidative ATP in neuronal populations (Figure 2(d), gray), while the

theoretical calculations provide comparative results that interneurons demand 18–21% of neuronal oxidative metabolism (Figure 2(d), black).

Energy demands of specific cellular components across species

In resting awake rat (Figure 3(a)), with an average neuronal firing rate of 4.3 Hz, nonsignaling processes (including housekeeping and resting potential) demand $\sim 27\%$ in excitatory neurons, $\sim 22\%$ in inhibitory neurons, and $\sim 47\%$ in glial cells. Synaptic transmission processes cost almost half of metabolism for both glutamatergic ($\sim 53\%$) and GABAergic ($\sim 47\%$) neurons. Action potential processes cost 10–15% of neuronal metabolism, while other signaling-related processes including calcium responses and glutamate/GABA recycling cost about 9% to 18% of neuronal metabolism. In halothane-anesthetized rat (Figure 3(b)), with an averaged neuronal firing rate of 2.5 Hz, nonsignaling processes demand a greater fraction of metabolism (i.e. $\sim 36\%$ in excitatory neurons, $\sim 28\%$ in inhibitory neurons, and $\sim 70\%$ in glial cells) and costs of other

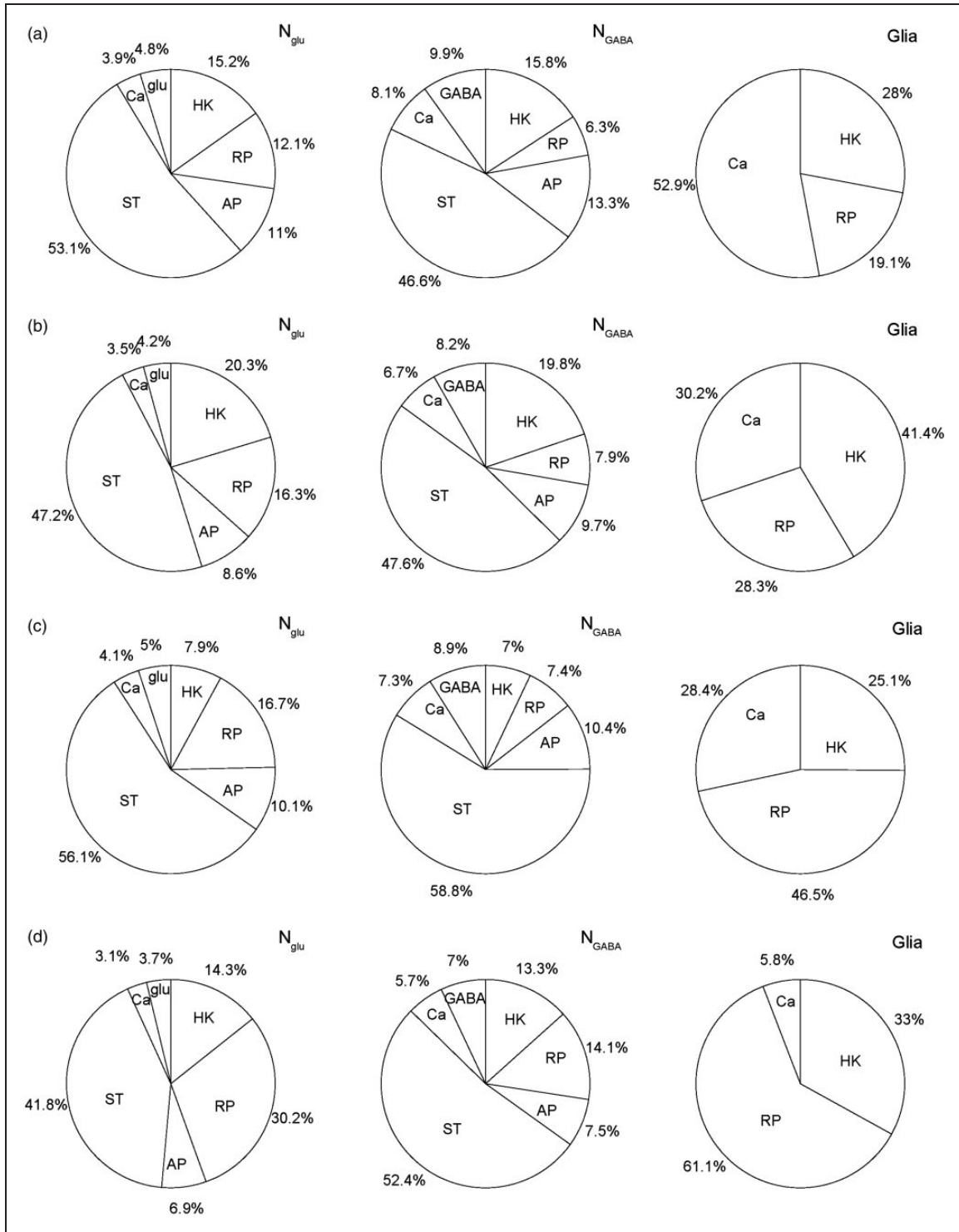


Figure 3. Distributions of ATP usage for different cellular mechanisms in excitatory glutamatergic neurons (N_{Glu}), inhibitory GABAergic interneurons (N_{GABA}), and glial cells (Glia) for the cerebral cortex of rat and human. The different cellular components represent housekeeping (HK), resting potential (RP), action potentials (APs), glutamate or GABA recycling (glu or GABA), presynaptic calcium (Ca), and synaptic transmission (ST). Distributions of energy budget for N_{Glu} , N_{GABA} , and Glia in rat cortex for (a) awake resting state and (b) halothane anesthesia compared to similar cellular components in human cortex for (c) awake resting state and (d) halothane anesthesia. Overall, in the cerebral cortex of rat and human, the HK and RP needs are greatest, while Ca need is quite small for Glia, and ST needs are nearly half of the total for N_{Glu} and N_{GABA} . From halothane anesthesia to awake resting states, the HK needs in N_{Glu} , N_{GABA} , and Glia are increased, whereas the APs, Ca, and glu needs in N_{Glu} and N_{GABA} are decreased relatively. See Figures S2 for further details of N_{Glu} , N_{GABA} , and Glia under awake resting and halothane anesthesia states for each species.

signaling-related processes are lower than in the awake state, but interestingly the fraction of energetics for GABAergic synaptic transmission is negligibly affected by halothane.

The calculated average neuronal firing rate in the resting awake human is about 1.15 Hz, much lower than that of rat. Nonsignaling processes in awake human demand $\sim 1/4$ of excitatory neuronal metabolism (i.e. similar to rat), $\sim 1/8$ of inhibitory neuronal metabolism (i.e., lower than rat), and $\sim 3/4$ of glial cell metabolism (i.e. higher than rat). Synaptic transmission processes in the awake human cost slightly more than half of neuronal metabolism (i.e. $\sim 56\%$ for glutamatergic and $\sim 59\%$ GABAergic activities). The similarities between rat and human are found for action potential processes (i.e. $\sim 10\%$ of neuronal metabolism) and signaling-related processes (i.e. 9% to 16% of neuronal metabolism for calcium responses and glutamate/GABA recycling). Upon induction with halothane anesthesia, as in the rat, a much lower average neuronal firing rate of 0.4 Hz is observed in human brain (Figure 3(d)), where demands of nonsignaling processes comprise a much greater fraction (i.e. $\sim 45\%$ in excitatory, $\sim 27\%$ in inhibitory neurons, and $\sim 94\%$ in glial cells) and costs of other signaling-related processes are dropped relatively because of weaker activity.

Recent experimental evidence indicates that astrocytes have very high calcium activity.^{26,27}

As Figures 1(f) and 2(c) indicate, if glial calcium cost was ignored, there will be a relatively large mismatch between calculated energy budget and measured glial metabolic cost measured by ^{13}C -MRS. As the right panels in Figure 3(a) to (d) demonstrate, calcium activities may cost significant fractions of glial metabolism, especially during non-anesthetized conditions when nearly half of glial metabolism is attributed to glial calcium signals.

Energy demands of specific cellular components in gray and white matter

There are some anatomical experimental data which show that white matter may contain a significant portion of unmyelinated axons and neurons even in adult human brain (see Figure 4 in Azevedo et al.²⁵), although most axons in nerves observed are myelinated in adult white matter.²¹ Hence, in our calculation for white matter, in addition to the 9.43 billion myelinated axons in white matter, we included 2.7 billion unmyelinated axons mainly from sparsely distributed neurons in white matter as reported by Azevedo et al.²⁵ These 2.7 billion neurons with unmyelinated axons may be actively signaling and will consume a significant portion of energy for adult white matter (Table 1; SI Text, Section B). These accounts in conjunction with nearly identical gray matter bottom-up descriptions for white

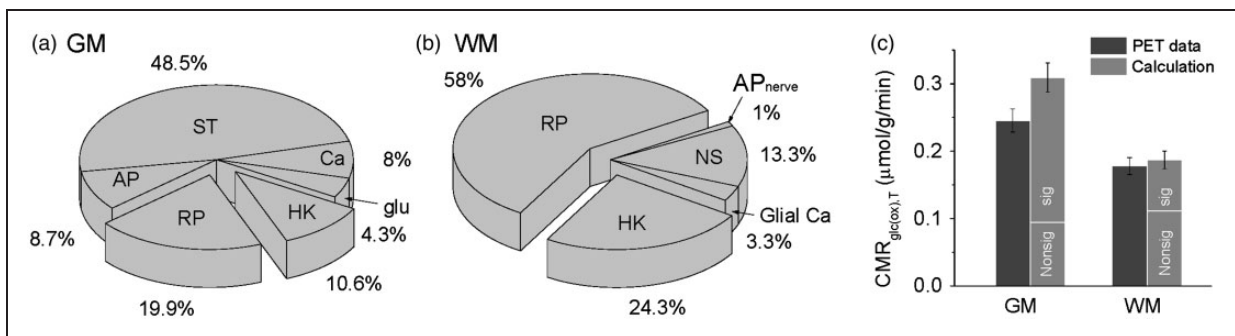


Figure 4. Distributions of ATP usage of different cellular mechanisms in gray matter (GM) and white matter (WM) of the human brain. The cellular components in GM represent housekeeping (HK), resting potential (RP), action potentials (AP), glutamate (and GABA) recycling (glu), presynaptic calcium (Ca), and synaptic transmission (ST). The cellular mechanisms in WM represent housekeeping (HK), resting potential (RP), neuron signaling (NS), action potentials in nerves (AP_{nerve}), and astrocyte calcium (glial Ca). Distributions of energy budget for (a) GM and (b) WM are shown in pie chart format where 100% represent the awake resting values of $\text{CMR}_{\text{glc(ox)}}$ in each tissue type. Nonsignaling costs (i.e. RP and HK) account for 30.5% and 82.3% of GM and WM demands respectively, suggesting that the total signaling costs in GM and WM are the remaining 69.5% and 17.7% portions, respectively. While the signaling costs in GM are assigned to energy needs of synaptic activity (i.e. ST, AP, Ca, glu), the signaling costs in WM are assigned to energy needs of 2.7 billion unmyelinated axons and 41.7 billion glial cells (i.e. NS, AP_{nerve} , Glial Ca). (see SI Text, Section B for details). (c) Bar plots of $\text{CMR}_{\text{glc(ox)}}$ for human brain PET data (dark gray^{38,55}) and calculated budget results (gray). For calculation, signaling (“sig” in the bar plot) and nonsignaling (“Nonsig” in the bar plot) components in GM and WM are shown in absolute value, and the nonsignaling component in GM is double the energy demand in WM. See Figure S3 for further details of GM and WM behavior at different levels of neuronal activity.

matter (E_{wm} ; using equations S13(a) to (d); SI Text, Section B) led us to compare distributions of cellular components in gray and white matter of human brain (Figure 4).

In gray matter, the nonsignaling and signaling processes respectively cost $\sim 30\%$ and $\sim 70\%$ (Figure 4(a)), whereas in white matter, the nonsignaling and signaling processes respectively cost $\sim 80\%$ and $\sim 20\%$ (Figure 4(b)). Nonsignaling costs, which primarily comprise of resting potential and housekeeping, account for 30.5% and 82.3% of gray and white matter demands, respectively. This suggests that signaling costs in gray matter (69.5% of total) are assigned to synaptic activity, whereas signaling costs in white matter (17.7% of total) meet the needs of billions of unmyelinated axons and glial cells. The $\text{calcCMR}_{\text{glc(ox),T}}$ values in gray and white matter of 0.31 and 0.19 $\mu\text{mol/g/min}$, respectively, in the awake state (Figure 4(c)) agree well with prior observations.³⁸

Both FDG PET and 2DG autoradiographic measures of metabolism only detect the usage of glucose molecules, whereas brain cells may also metabolize other substrates (e.g. lactate, ketone bodies) to provide the ATP they require.³⁹ In our calculations all ATP, which includes both aerobic and glycolytic pathways, are assumed to be generated through glucose. Thus, some of the deviations of our predictions from the experimental data could be explained by contributions from these alternate substrates. It should also be noted that the measured PET values for $\text{CMR}_{\text{glc(ox),T}}$ were averaged for gray and white matter, respectively, from the entire cerebrum. While absolute levels of metabolism differ significantly between gray and white matter, the absolute amount of energy attributed to nonsignaling in both tissues is roughly similar (Figure 4(c)).

Discussion

Although bottom-up gray matter energy budgets have existed since the early 2000s,^{12–14,21,22,40} experimental validation across behavioral states and/or species has been limited to largely comparison with a single experimental measurement from either 2DG autoradiography or FDG PET, usually just the awake resting value. We pooled a range of metabolic and physiologic measurements, using different methods, both in the rat and the human brain across several behavioral states (i.e. from awake to different anesthetized levels; Tables S1 to S5), to validate a comprehensive $\text{CMR}_{\text{glc(ox)}}$ -derived cellular level energy budget for gray and white matter (Figures 1 to 4). Based on our calculations, the higher metabolism in rat brain compared to human brain is primarily due to the higher firing rate and secondarily due to differences in synaptic density. Indeed, this issue was carefully formulated in a previous theoretical study,⁴¹ where energy consumption was reported to depend on many

parameters, but most of them were species independent neurophysiological and structural invariants, except firing rate. The excellent fit we found to neuroenergetic measurements across a wide range of brain activity strongly supports this previous theoretical conclusion and extends it to glia and GABAergic neurons.

While the majority of the morphological data were for rat brain (Table 1), upon scaling to human brain, the biophysical parameters were essentially unchanged (Table 2). This suggests that basic properties of action potential generation and synaptic transmission processes are conserved across mammalian species, consistent with the conclusion made from a pure top-down analysis.¹³ However, this current work differs from our previous work, which calculated microscopic parameters based on a top-down fit of the model to experimental data¹³ by demonstrating that excellent agreement with experimental data is obtained using an ab initio bottom-up approach incorporating current best experimental values of these parameters.

In cases where the microscopic experimental values were not well determined, the validation of theoretical budget calculations with experimental measurements provided a valuable way to predict some uncertain parameters. For example, although most of the human experimental data suggest that the averaged firing rate of awake human neurons is in the range of 0.5 to 2 Hz,^{42,43} an accurate estimation has been missing from both experimental and theoretical approaches. Lennie²² predicted the average neuronal firing rate to be ~ 0.024 Hz, whereas Karbowski^{41,44} predicted values closer to 1 Hz. By comparing our budget calculation with human brain ¹³C-MRS and PET metabolic data from different behavioral states, we predict that the resting average neuronal firing rate in resting awake human brain is ~ 1.15 Hz, which is in good agreement with the mean of the range reported in the human brain in vivo^{42,43} and theoretical estimation from Karbowski.^{41,44} Updated accounts of cerebellar budget¹² with cerebellar morphology⁴⁵ will be crucial to extending this budget.

Principles of neuronal and glial energetics in gray matter

The agreement between the measured and calculated values (of metabolism and firing rate) provided good validation of biophysical properties of neuronal and glial cells (Figure 1). As shown in Figure 1(a), varying the firing rate alone without changing the costs of different components can explain the majority of the change in energy consumption with state. However, the data leaves open the possibility that the variations seen from perfect agreement (Figure 1(b) and (c)) could be explained by small changes in membrane resistance,

synaptic transmission per spike, and/or housekeeping activities with each behavioral state. However, these changes cannot be large, indicating that the system maintains these factors close to constancy, which in turn may be important for maintaining fidelity of neuronal communication. The possibility of these factors having a dependence on the firing rate could be explored using the model presented by assessing if they provide a better fit to the experimental data. Despite differences in absolute metabolism and neuronal activity for rat (Table S1) and human (Table S2) brain, upon normalization to the awake state for each species, the energy demands for the total, neuronal, and astrocytic activities were nearly identical for both the species (Figures 1 to 3; Table 2).

The universal principles for gray matter are: (i) Up to 30% of total ATP in the awake state is attributed to nonsignaling processes like maintenance of resting potential and housekeeping metabolism in neurons and astrocytes; (ii) ATP demand of neuronal activity during signaling processes dominates in comparison to energy demand of astrocytic activity; (iii) ATP demand increases almost linearly with increasing neuronal activity; however, glial activity demands increase in a non-linear manner beyond the awake state.

The finding that signaling and nonsignaling costs, respectively, occupy $\sim 80\%$ and $\sim 20\%$ of neuronal energy in the awake state (Figure 2(a)) was independently verified by comparison with ^{13}C -MRS measurements (Figure S1). In Figure 2(a), we plotted neuronal glucose oxidation vs. neuronal firing rate, as measured by PET, 2DG, and electrophysiology methods, whereas in Figure S1, we plotted neuronal glucose oxidation vs. neurotransmitter recycling, as measured by ^{13}C -MRS. The agreement between the non-MRS and ^{13}C -MRS results provides good validation of biophysical properties used in the calculations for neuronal and glial cells (Figure 2).

The agreement between these two plots reveals that synaptic transmission is indeed proportional to firing rate (electrical activity) or neurotransmitter recycling (chemical activity).⁴⁶ Additionally, we find that the amount of neurotransmitter being recycled at 1 Hz is effectively identical between species. For the awake rat, a value of $0.55 \mu\text{mol/g/min}$ has been measured by ^{13}C -MRS for neurotransmitter cycling (Table S3) at a neuronal firing rate of 4.3 Hz (Table 2). This corresponds to about $0.13 \mu\text{mol/g/min}$ recycling of neurotransmitter at 1 Hz firing. For the human cortex, where the neuronal density is exactly half that in the rat cortex (Table 1), we calculate nearly an identical value of neurotransmitter recycling at 1 Hz firing in the awake state (i.e. $0.34/1.15/2$) (Tables 2 and S3). This suggests that the amount of neurotransmitter packaged, released by vesicles, and then recycled

through glial cells is nearly proportional across species as a function of cortical firing rate, despite the different neuronal sizes across species. Given the similar slopes of Figures 2(a) and S1, incremental rise in firing by 1 Hz will increase glucose oxidation by $\sim 0.1 \mu\text{mol/g/min}$ (i.e. $0.13 \times 0.8 = 0.1$). In other words, in 1 g of cortical tissue ~ 1 quadrillion glutamate molecules are recycled through the synapse per firing event per second, and this is independent of species. The agreement between model predictions and experimental data (Figure 2(a)), which include ^{13}C -MRS where energy contributions of glutamatergic and GABAergic neurons were measured (Figure S1), suggests that our morphological estimates of different cell types are reasonable.

While signaling-associated functions of pyramidal cells dominate, energy demands of glial cells and inhibitory neurons are not negligible (Figure 3). The nonsignaling costs of glial cells are three times higher than that of neurons, and nearly half of glial metabolism is due to glial calcium signaling (Figure 3). We predict that glial energy possibly can increase quite significantly at very high activity levels (Figure 1(f)). Recent *in vivo* measurements indicate activity-dependent increase in glial function.^{27,47,48}

Principles of neuronal and glial energetics in white matter

In human gray matter, there are 32.8 billion total cells with glial cell to neuron ratio of about 3:2, whereas of the 44.4 billion total cells in human white matter an overwhelming majority are glial cells (Table 1). Despite these immense differences in number of glial cells between gray and white matter in the human brain, there is a two to three times difference in $\text{CMR}_{\text{glc(ox)}}$ between these tissues and the nonsignaling energy demands in gray and white matter are roughly of the same magnitude (Figure 4(c)). This indicates that the main difference between gray and white matter arises from the higher energy costs associated with neuronal signaling in the cerebral cortex (Figure 4(a)).

We estimated the cost for human white matter based on the same method used for rat data in a recent report,²¹ but here we included a small amount of non-glial cells in white matter (2.7 billion unmyelinated and 9.4 billion myelinated axons, Table 1).²⁵ Treating these non-glial cells as neurons would provide an upper limit of white matter energy demand. Since neurons fire spikes to induce synaptic transmission, these relatively few neurons in white matter may add non-ignorable costs to the white matter budget. This factor was not considered fully in the earlier report.²¹ Both glial calcium activity and activity of non-glial cells make energy cost in white matter increase slowly with rising cortical activity (e.g. switching from anesthesia to awake state, Figure S3). In

support of this possibility, studies show small but significant state-dependent changes in white matter metabolism,⁴⁹ an issue perhaps more relevant in specific disease conditions.⁵⁰ Furthermore, experimental evidence directly suggests that individual astrocytes^{27,48,49,51} or oligodendrocytes^{26,51,52} may also have large demands for maintenance of resting potentials (Figure 3).

Significant nonsignaling energetic demands

Recent *in vivo* ¹³C-MRS studies in rat and human brain suggest that majority of the energy in the awake state, as much as 70–75% of the total ATP derived from $CMR_{glc(ox)}$, support cortical signaling functions, whereas the remaining 25–30% fraction is attributed to nonsignaling functions.¹³ However, theoretical bottom-up budget estimates have varied from 20% to 50% of the ATP derived from $CMR_{glc(ox)}$ supporting rat cerebral cortex nonsignaling functions.²⁰ This uncertainty is mainly due to the lack of modeling and measurements of specific cellular physiologic and metabolic processes that occur in the nonsignaling state. Here, we take a new way to calculate the nonsignaling costs by computing the averaged mass value of individual neurons and glia, and mass-dependent basal metabolic rate^{23–25} (see SI Text Section A1 for details). This makes the calculated budget match the ¹³C-MRS results well. Moreover, budget results show that nonsignaling needs of white matter are more than double of that in the gray matter, presumably due to higher biosynthesis turnover demands (Figure 3). Together, the nonsignaling demands of white matter could be very large (Figure 4), as discussed below. Overall, our calculations show that in white matter, the energetic cost of the nonsignaling processes is 4 times higher than signaling processes, whereas in gray matter, the energetic cost of the signaling processes is 2.5 times higher than nonsignaling processes in the awake state.

Validation of $CMR_{glc(ox)}$ and neuronal activity in the gray matter budget (Figures 1 to 2) strengthened the synergy between “input” (Table 1) and “output” (Table 2) parameters for both rat and human brain to help narrow the recent debate on the magnitude of the nonsignaling cost²⁰ and provided evidence that signaling-dependent energy is a dominant part of the brain budget.^{13,53,54} In fact, we estimate that the signaling costs may increase up to 85% in awake and physiological stimulated conditions. However, significant fraction of $CMR_{glc(ox)}$ can now be assigned for housekeeping needs (i.e. ~11% in gray matter and ~24% in white matter, Figure 4) and maintaining resting potential (i.e. ~20% in gray matter and ~58% in white matter, Figure 4). These two factors together are assigned to nonsignaling costs, which were highest for glial cells and lowest for interneurons (Figure 3). While

our budget has not detailed different types of biosynthetic processes (e.g. protein, lipid, or myelin) or proton leaks for housekeeping needs, we have assigned a significant part of nonsignaling costs to maintaining resting potential.

In summary, this type of top-down validated bottom-up energy budget of the normal human brain provides exciting prospects to test the impact of microscopic variations or compartmentalized cellular perturbations on macroscopic $CMR_{glc(ox)}$, as measured *in vivo* by PET and ¹³C-MRS. The key to this top-down validated bottom-up energy budget was the rat data, which included independent measurements of electrical firing rate and ¹³C-MRS measurements of energy signaling and nonsignaling components. From the standpoint of using this model to interpret PET results, we believe the model allows us, within a linear approximation, to distinguish whether a change in regional activity is signaling or nonsignaling dependent, and whether it reflects changes in glutamatergic neurons, GABAergic neurons, or glial cells. In the near future, it may be possible to create both hemodynamic-based energy budgets of brain function, which could provide the basis for brain networks that are revealed by methods like fMRI and PET.

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Declaration of conflicting interests

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Authors' contributions

YY, PH, FH designed and performed research. YY, PH, DLR, DA, FH analyzed data and wrote paper.

Supplementary material

Supplementary material for this paper can be found at the journal website: <http://journals.sagepub.com/home/jcb>

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