



# Mesenchymal Stem Cell-Macrophage Choreography Supporting Spinal Cord Repair

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## Abstract

Spinal cord injury results in destructive events that lead to tissue loss and functional impairments. A hallmark of spinal cord injury is the robust and persistent presence of inflammatory macrophages. Mesenchymal stem cells (MSCs) are known to benefit repair of the damaged spinal cord often associated with improved functional recovery. Transplanted MSCs immediately encounter the abundance of inflammatory macrophages in the injury site. It is known that MSCs interact closely and reciprocally with macrophages during tissue healing. Here, we will review the roles of (transplanted) MSCs and macrophages in spinal cord injury and repair. Molecular interactions between MSCs and macrophages and the deficiencies in our knowledge about the underlying mechanisms will be reviewed. We will discuss possible ways to benefit from the MSC-macrophage choreography for developing repair strategies for the spinal cord.

**Keywords** Stem Cells · Bone Marrow · Immune Cells · Healing · Recovery · Paralysis · SCI

## Introduction

Traumatic spinal cord injury (SCI) typically results in immediate loss of nervous tissue followed by a phase of secondary damage. The initial trauma destroys neural cells and ruptures blood vessels in the injury epicenter, while in the surrounding

tissue (penumbra) the spinal cord-blood barrier (ScBB) of blood vessels is often breached and neural cells become necrotic. Secondary damage is perpetuated by the many cytotoxic factors associated with neural and epithelial cell death as well as by immune cells. The loss of nervous tissue ultimately leads to the formation of cystic cavities, which can be found at the injury epicenter and in adjacent segments.

A hallmark of SCI is a robust and persistent inflammatory response. The destructive events initiated by an injury to the spinal cord lead to a plethora of damage signals, which cause a massive infiltration of immune cells that initiate the inflammatory response [1–3]. In most tissues, intrinsic mechanisms regulate the evolution of macrophages from an inflammatory to an anti-inflammatory, reparative phenotype which supports recovery of homeostasis and tissue repair. However, in the damaged spinal cord, the majority of macrophages remain in their inflammatory phenotype resulting in persistent inflammation [4–8].

Mesenchymal stem cells (MSCs) have been investigated for treatment of SCI [9–11]. MSCs secrete growth factors and chemo- and cytokines, which mediate paracrine actions that support anatomical repair and functional recovery [12–16]. Their potential to create a reparative environment is the main motivation for exploring MSCs for repair of many types of tissues [17–21]. In animal models of SCI, MSC transplantation demonstrated promise for promoting repair [9–11, 22]. The mechanisms by which transplanted MSCs execute

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their reparative actions in the damaged spinal cord remain incompletely understood.

In numerous types of tissues, including skin and muscle, MSCs home into an injury and start communicating with immune cells thereby supporting tissue repair and remodeling. MSCs exert dual roles as “sensors” and “modulators” of the inflammatory response [23–26]. Macrophages, in turn, are known to mediate MSCs polarization to pro- or anti-inflammatory profiles [27–31]. These reciprocal actions between MSCs and macrophages have significant influence on the overall efficacy of the healing process. Homing of MSCs into the damaged spinal cord does not occur, which prevents such repair-promoting MSC-macrophage interactions.

In the injured spinal cord, a transplant of MSCs will encounter the abundantly present inflammatory macrophages. In this review, we focus on the role of transplanted MSCs, macrophages, and their reciprocal interactions in spinal cord injury and repair. We will give special attention to their roles in the formation of new blood vessels (i.e., angiogenesis) after SCI. Understanding the contributions of these two cell types and their interactions may be beneficial for the development of effective repair approaches for the spinal cord. We will also discuss potential approaches to benefit from their interactions for promoting repair and recovery after SCI.

## Inflammation after SCI

An injury to the spinal cord causes immediate and secondary destructive events, which together cause nervous tissue damage and, consequently, functional impairments. These destructive events, which include neural cell death, tissue destruction, and blood vessel rupture and leakage, result in an immune response which is characterized by enduring inflammation.

### Different Phases in Inflammation After SCI

Damage to neural cells induces the expression of damage-associated molecular patterns (DAMPs), which are sets of small molecules and proteins released by cells present in the injury site that are responsible for several injury-related aspects of SCI, including homing of inflammatory cells [32, 33]. Neutrophils start accumulating within 6 h after injury, lymphocytes within 6–12 h, and macrophages within 12–24 h [7, 34, 35]. After a peripheral nerve injury, dorsal root ganglion neurons secrete CCL2, an attractant chemokine that mediates macrophage migration and activation for repair [36]. After SCI, increased concentrations of macrophage chemoattractant protein (MCP-1) and other cytokines are present in the damaged tissue [37, 38]. Immune cells also infiltrate an injury site through ruptured blood vessels to encounter the DAMP-positive microenvironment and exacerbate the inflammatory milieu [3]. The initial inflammatory

wave of the immune response is essential for debris clearance, phagocytosis of dying cells, and initiation of angiogenesis through the secretion of growth factors [1, 2, 6, 39].

In many types of tissue, the initial inflammatory phase is followed by a regulated induction of an anti-inflammatory, reparative, phase, which is governed by regulatory T lymphocytes (Tregs) and tissue remodeling/reparative macrophages [40]. For so far unknown reasons, wound healing and inflammation-mediated clearing of cellular debris in the damaged spinal cord differs from that in regenerative tissues, such as muscle and skin. After SCI, there is no efficiently regulated induction of an anti-inflammatory and reparative phase, resulting in a chronic cytotoxic inflammatory state that contributes to secondary degeneration, thereby limiting repair and functional recovery [4, 5, 41–43]. In the contused adult rat spinal cord, reparative macrophages can be found early after injury, during the initial inflammatory phase, but they disappear within a few days, while the inflammatory macrophages remain chronically present [5, 6]. In people with SCI, increased inflammatory markers were found in blood [44] and cerebrospinal fluid [45, 46]. Post-mortem analysis of human spinal cord tissue showed the persistent presence of inflammatory cells [4].

It has been shown that both systemic and local chronic inflammation are characterized by aggressive immune cell behavior in the injury site, accompanied by a continuous production of cytotoxic molecules, including reactive oxygen species (ROS), nitric oxide (NO), and apoptosis-inducing molecules, which are likely to contribute significantly to secondary nervous tissue degeneration. It is possible that suppression of inflammation in the chronic phase could allow remodeling cells to intervene and promote tissue repair more efficiently, after the beneficial acute inflammatory role has been executed [47].

### Macrophage Phenotypes

Macrophages are the most prevalent immune cell present in the spinal cord during the later phase of inflammation after injury. Macrophages, in the injury, originate initially from resident activated microglia, which tend to dissipate in the chronic phase, and later from infiltrated monocytes, which remain present in the chronic phase [1, 39, 48]. Macrophages sense the cellular and molecular composition of the microenvironment and in response alter their gene expression profile to secrete the appropriate effector molecules and express the necessary receptors on their surface [49, 50]. This dynamic transition in the phenotype of macrophages is also known as “polarization” or “functional state” [51]. Classically, macrophage polarization has been classified into two categories: (1) inflammatory, cytotoxic, M1-like macrophages, and (2) anti-inflammatory, reparative, M2-like macrophages. At present, it is widely accepted that macrophage phenotypes reflect a wide

spectrum of gene expression and protein secretion levels. Classifying macrophages in two categories is useful in a controlled environment, such as *in vitro* experiments, but is complicated in *in vivo* studies where it in fact may hinder repeatability and understanding of the actual mechanisms of macrophage polarization during wound healing [51–53]. Thus, it is more prudent to regard macrophage phenotypes *in vivo* as a balance between the two extremes of the inflammation spectrum, i.e., M1 and M2 macrophages. In this manuscript, we will refer to M1-like macrophages when the microenvironment is dominated by the presence of inflammatory macrophages and cytokines, and to M2-like macrophages when it is dominated by the presence of anti-inflammatory macrophages and cytokines.

The macrophage phenotype evolution has been characterized in damaged spinal cord nervous tissue in different injury models [6, 7, 41, 42, 47]. M1-like macrophages are mainly characterized by the secretion of inflammatory cytokines, such as interleukins (IL) 1 $\beta$ , 6, 12, tumor necrosis factor alpha (TNF $\alpha$ ), interferon gamma (IFN $\gamma$ ), and expression of the inducible nitric oxide synthase (iNOS), among other markers. M2-like macrophages are characterized by the secretion of anti-inflammatory molecules, such as IL4, IL10, transforming growth factor beta (TGF $\beta$ ), and growth factors that induce tissue remodeling [50, 51, 54, 55]. Evidence in the literature demonstrate that interventions resulting in depletion of M1-like macrophages in other tissues than spinal cord caused impairment of the healing process and lack of functional recovery [56, 57]. Interestingly, approaches that induce the shift from M1- to M2-like macrophages earlier than the natural timeline increase the chances of excessive fibrosis and, ultimately, failed tissue remodeling [58, 59]. The absence of reparative (M2-like) macrophages was also shown to result in a lack of neural tissue repair [60]. Together, these studies demonstrate the relevance of the role of each macrophage phenotype for successful tissue repair.

Different macrophage phenotypes have specific and crucial roles in tissue repair and remodeling. It has been proposed that modulating macrophages to a pro-reparative phenotype is a promising strategy to promote repair after SCI [8, 26, 47, 61, 62]. Their specific roles in the inflammation process need to be considered when designing macrophage-modulating strategies. Even though M1-like macrophages might appear to be the “bad” players in the immune response after injury being uncooperative of repair, they are essential for removing cellular/tissue debris before reparative events can successfully be executed. At the same time, M2-like macrophages appear to be the “good” players in the immune response after injury supporting repair, but when promoting this phenotype in the injured spinal cord, it will need to be precisely regulated to avoid counter-productive effects. The ultimate goal would be to design strategies to modulate the immune response after injury that respects the natural timeline of macrophage phenotype evolution. Promoting the M2-like phenotypes after the

window of necessary inflammation has passed is a promising strategy to enhance the natural immune-mediated tissue healing process that is lacking in the spinal cord [47].

## Macrophages Affect Angiogenesis

Angiogenesis is an essential process for wound healing. Cells involved in tissue repair are in need of oxygen and nutrients to survive and successfully contribute to the complex tissue repair process. For this, the formation of new blood vessels from existing capillaries at and near the injury site needs to be synchronized with the dynamic cellular aspects of repair. Macrophage polarization is tightly connected with the regulation of angiogenesis after injury [56, 63–65]. M1-like macrophages secrete enzymes that modify the extracellular matrix (ECM), as well as vascular endothelial growth factor (VEGF), which promotes proliferation of the vascular endothelial cells. M2-like macrophages secrete platelet-derived growth factor (PDGF) and various other factors that promote the proliferation of smooth muscle cells and pericytes, which are needed to stabilize newly formed blood vessels [35, 62, 63, 66].

The literature shows that the effects of macrophages on angiogenesis after injury are essential and in need of precise orchestration to facilitate proper repair. Thus, the sequential evolution of macrophage phenotypes needs to be considered cautiously when designing and exploring novel treatments for repair of the spinal cord. Clinically, treatments implemented in the acute/subacute phase of SCI will certainly need to take into account the different roles of the macrophage phenotypes in angiogenesis. When treatments are given to people in the chronic phase of SCI, the focus should be on reducing (persisting) inflammation and promoting the activation of the reparative macrophage phenotypes.

## MSCs for Spinal Cord Repair

An injury to the spinal cord causes tissue loss and functional impairments [67]. Tissue loss is reversely correlated with functional recovery after SCI [10, 63, 68]. One of the approaches to elicit repair after SCI is the transplantation of stem cells, which are known to secrete paracrine factors that influence repair and/or differentiate into neural cells to replace those that were lost [14, 21, 69, 70]. Using models of SCI, studies have shown that stem cell-based approaches elicit anatomical repair often accompanied by functional recovery [10, 71, 72]. An extensively explored type of stem cell for spinal cord repair is the MSC [10, 20, 73–75]. MSCs can be harvested from different types of (mesenchymal) tissues, including bone marrow, adipose tissue, and placenta. Obtaining the cells from these sources can be accomplished with relatively minor side effects, which adds to their clinical relevance. In most studies, the MSCs used are a heterogeneous population of

cells, which deserves some caution because the source as well as the selection and expansion methods can result in cell populations with different repair potential [76]. For example, MSCs obtained from human olfactory mucosa showed a stronger potential in promoting *in vitro* remyelination than those obtained from bone marrow [77]. The MSCs from adipose tissue and bone marrow were shown to differentially benefit repair-related events, such as differentiation, angiogenesis, or immunomodulation [78]. Therefore, further characterization of each MSC's origin and behavior is important in order to select the source and processing method that best applies to the desired functional application.

MSCs migrate from their source to injured tissues through the blood stream, in response to numerous different chemoattractants. According to the type and site of injury, the circulating factors vary and the needs for repair are different. MSCs are known to activate their migratory mechanisms upon injury-mediated stimulation with VEGF, substance P, hepatocyte growth factor (HGF), the stromal derived factor 1 (SDF-1)/CXCR4 axis, and hyaluronic acid (HA), among others [79–81]. After SCI, natural homing into the damaged area is inefficient as is migration to the injury site after systemic transplantation. MSCs are often “trapped” in other highly vascularized tissues, such as the lungs, or they are transformed by the circulation environment. Also, many MSCs may not survive during the migratory event, resulting in low cell concentrations at the injury site (reviewed in [80, 81]). The delivery method needs to be considered, with the source and processing strategy, as an additional variable in the choice and guided by the desired end application.

### Reparative Properties of MSCs in the Injured Spinal Cord

MSCs are known to secrete factors that initiate and regulate specific actions contributing to the overall anatomical repair and functional recovery seen after their transplantation into the injured spinal cord [14, 16, 18, 74, 82, 83]. Many of these paracrine MSC-mediated events result in neuroprotection, which may be associated with functional recovery [10, 83, 84]. Evidence in the literature shows that an intraspinal MSC transplant leads to a decrease in apoptotic neural cell death [16, 85] and in overall neurotrophic support of the damaged tissue [74, 86]. Other studies demonstrated that a transplant of MSCs results in angiogenesis within the injured spinal cord segment and stabilization of the ScBB of breached blood vessels in the penumbra [10, 15, 87]. Also, MSC transplants resulted in reduced breakdown of ECM, which typically occurs after SCI and contributes significantly to tissue disorganization and destruction [88].

Besides the abovementioned events, MSCs are known to affect the profile of the inflammatory response through modulation of the macrophage phenotype, which has important

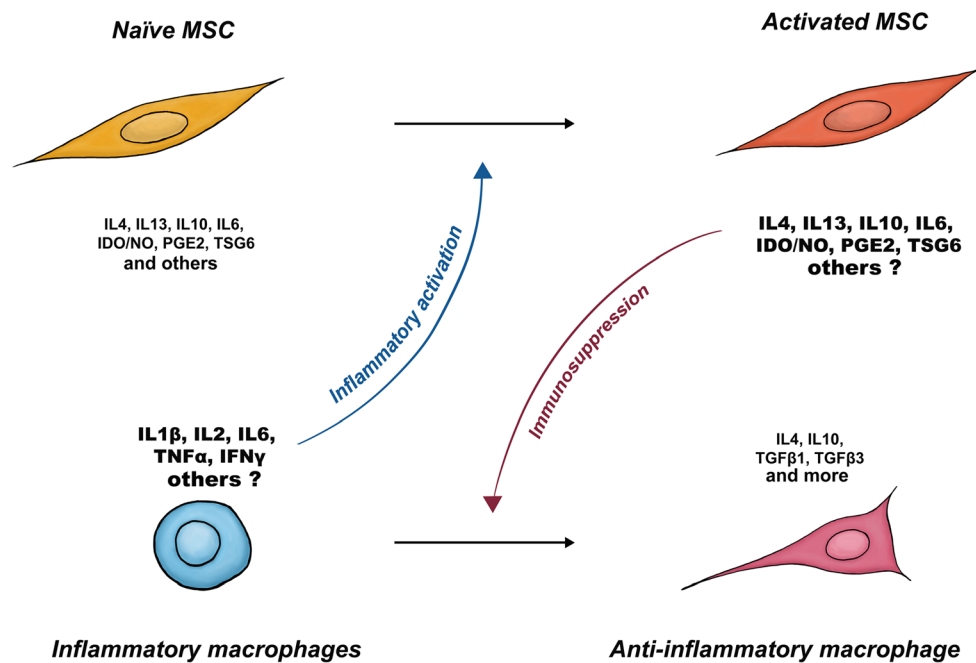
consequences for repair [25, 89–92]. MSCs have dual roles as “sensors” and “modulators” of the inflammatory response [23]. Inflammatory cues in an injury microenvironment are sensed by MSCs that, in response, produce cytokines that modulate macrophages to express their anti-inflammatory, pro-reparative, phenotype, which supports restoration of homeostasis and promote tissue repair [23, 24, 93] (Fig. 1).

Exposure to IFN $\gamma$  induces MSCs to enhance their secretion of indoleamine 2,3-dioxygenase, which inhibits the proliferation of activated natural killer (NK) cells and T lymphocytes [99]. The innate ability of MSCs to modulate inflammation could be a powerful tool to limit secondary nervous tissue degeneration, and thus providing a larger tissue platform for recovery strategies. For developing such approaches a better understanding of the mechanisms by which transplanted MSCs execute these immunomodulatory functions and communication with the macrophages is needed.

The literature so far provides evidence that the paracrine activities of MSCs transplanted in the injured spinal cord cause direct or indirect events that lead to neuroprotection. The direct events are through releasing anti-apoptotic and neurotrophic molecules that rescue neural cells from injury-induced death. The indirect actions are through the release of molecules that limit ECM breakdown, promote angiogenesis, stabilize breached ScBB, and modulate the immune response. Together these events can exert neuroprotection and therefore limit the loss of neural tissue. Evidence has shown that limiting the (secondary) loss of nervous tissue after SCI is associated with functional recovery [10].

Similar trends are observed when applying MSCs to central and peripheral nervous system injuries. The underlying pathophysiology of traumatic brain injury (TBI) follows largely similar patterns as observed in SCI. TBI results in immediate tissue damage, vascular damage, and inflammation, which all contribute to secondary degeneration [10]. Comparable to the injured spinal cord, transplantation of MSCs in the traumatically injured brain results in neuronal regeneration and tissue sparing associated with improved functional recovery [100, 101]. The main mechanisms underlying MSC-mediated repair of the brain are thought to be neural regeneration and immunomodulation through secreted factors [83, 100, 101], which are analogous to the mechanisms of MSC-mediated spinal cord repair. The pathophysiology of peripheral nerve injury differs from that in CNS injuries [102] with less vascular damage and less impediments to neuronal regeneration. However, peripheral nerve injuries are characterized by secondary degeneration and macrophage infiltration as is observed in CNS injuries. The reparative mechanisms of MSCs in the peripheral nerve are also thought to be based on paracrine support and immunomodulation. MSCs are also being explored for their ability to differentiate into Schwann cell-like cells for supporting repair [102, 103].





**Fig. 1** MSCs and macrophages reciprocally influence each other. Naïve MSCs secrete basal levels of anti-inflammatory cytokines [13, 88]. Inflammatory macrophages secrete numerous molecules that act as inflammatory stimuli modulating a shift from naïve MSC to “activated” MSC (“inflammatory activation”). The activated MSCs have switched on their immunomodulatory program, which results in the secretion of

similar molecules as naïve MSCs but in greater levels. These high levels of anti-inflammatory cytokines drive the switch from inflammatory to anti-inflammatory macrophages (“immunosuppression”), which are important in tissue repair and remodeling. The specific stimuli and molecular hierarchy involved in these cells regulation is still incompletely characterized [11, 23, 27, 92, 94–98]

## Reciprocal Communication Between MSCs and Macrophages

It is known that MSCs modulate the immune response, while macrophages influence the behavior of MSCs [27, 95, 104]. The directionality of the interactions between MSCs and macrophages suggests a tightly controlled reciprocal cooperation for regulating repair in regenerative tissues. In fact, evidence in the literature showed that macrophages establish a bidirectional crosstalk with stem cells promoting healing in various tissue types [95, 96, 105].

### MSCs Modulate Macrophage Polarization

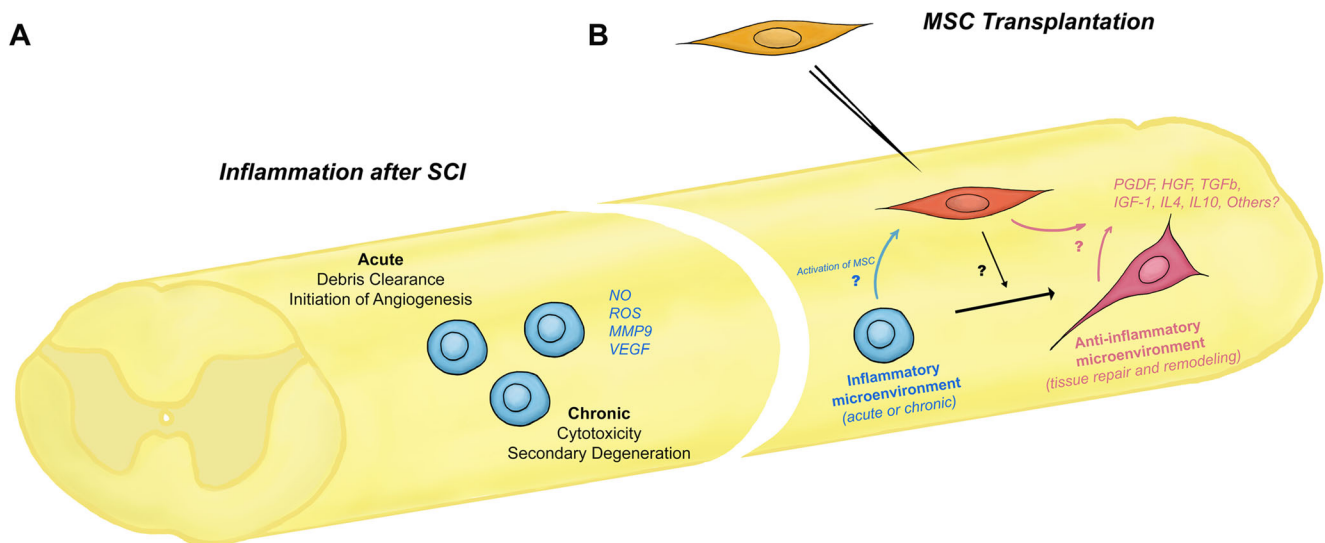
MSCs constitutively secrete immunosuppressive cytokines, such as TGF $\beta$ , IDO, NO, TNF-inducible gene-6 (TSG-6), prostaglandin-E2 (PGE2), and anti-inflammatory ILs that induce modifications in the metabolism of macrophages, resulting in a switch to the anti-inflammatory macrophage phenotypes [23, 24, 92, 96, 106] (Fig. 1). When cultured with MSC-conditioned medium, macrophages polarize showing increased expression of M2-like surface markers and reduced secretion of the inflammatory cytokines, IL1 $\beta$ , IL6, and TNF $\alpha$  [92]. Macrophages in culture with MSC spheroids, which secrete enhanced levels of PGE2, or with conditioned medium thereof, polarize from M1-like to M2-like macrophages [107, 108]. Systemic administration of umbilical

cord-derived MSCs to an animal model of an inflammation-related disorder, led to an increase in anti-inflammatory macrophages and alleviation of the associated symptoms [98]. The accumulated evidence in the literature so far reveals that MSCs support a shift among macrophages from the M1- to M2-like phenotypes (Fig. 2).

Transplantation of MSCs into the contused spinal cord in adult rats was found to result in a decrease in TNF $\alpha$ , IL6, and IL1 $\beta$  and an increase in IL4 and IL13 accompanied by tissue sparing, axon preservation, decreased scar formation, and improved functional outcomes [11]. Treatment of the adult mice contused spinal cord with conditioned medium of embryonic MSCs resulted in restoration of the macrophage phenotype after the inflammatory phase and improved locomotor recovery, indicating that soluble factors are important in MSC immunomodulation [111]. In general, promoting the shift to M2-like macrophages, via MSCs, may support repair after SCI [26].

### Macrophages Influence MSCs

Inflammation and macrophage-derived cytokines influence the secretory profile of MSCs [12, 24, 79]. It was found that *in vitro* exposure to macrophage-conditioned media, as well as co-culture with different phenotypes of macrophages, modifies the secretome of MSCs and their viability for cardiac tissue repair [27]. Crisostomo and colleagues used inflammatory



**Fig. 2** MSC and macrophages orchestrate spinal cord repair. **a** Injury to the spinal cord is characterized by robust infiltration of immune cells. During the (sub-)acute phase of injury, inflammatory (M1-like) macrophages help clearing debris by releasing NO, ROS, and ECM-modifying enzymes, such as MMP9. M1-like macrophages also release proangiogenic factors that promote the formation of new blood vessels, such as VEGF. In the spinal cord, macrophage polarization fails to shift to the M2-like phenotypes, causing chronic inflammation and, consequently, a continuous exposure to cytotoxic molecules that impede recovery and induce secondary degeneration [1, 7, 109, 110]. **b** Transplants of

MSCs may promote the shift of macrophages to the reparative (M2-like) phenotype. Exposure of MSCs to the inflammatory microenvironment activates their immunomodulatory program; mechanisms underlying this activation are still incompletely understood. The transition to reparative macrophages and the paracrine effect of MSCs supports repair through released reparative molecules, such as PDGF, HGF, IGF-1, or TGFb. The regulatory mechanisms need further study in order to benefit the development of targeted MSC therapies for spinal cord repair [6, 11, 23, 26, 27]

stimuli, such as  $\text{TNF}\alpha$  and lipopolysaccharides (LPS), to condition MSCs and found an increase in the secretion of growth factors that support tissue repair [12]. Other types of stem cells, including oligodendrocyte precursor cells [60] and hair stem cells [112], can also be modulated by macrophage-secreted molecules resulting in improved tissue regeneration. Considering the sequence of macrophage phenotypes during wound healing, it is clear that the immunomodulatory crosstalk between MSCs and macrophages within the injury site is crucial for the overall repair process and recovery [95]. Further investigations into these immunomodulatory mechanisms could support the development of therapies for the spinal cord that harness the reparative and modulatory potential of MSCs and macrophages to create a reparative milieu in the injury site.

### MSC Immunomodulation Requires Activation

Effective immunomodulation by MSCs occurs upon exposure to activating stimuli. Under stress conditions, MSCs are programmed to increase their secretion of growth factors. For instance, MSCs in culture under hypoxic conditions produce enhanced levels of growth factors [12]. Cultured MSCs exposed to inflammatory molecules increase the expression of receptors known to bind immune regulatory mediators and the production and secretion of anti-inflammatory factors [24, 113]. Despite the classical immunosuppressive potential of

MSCs, it is now also accepted that MSCs exhibit a spectrum-type of phenotypic polarizations, similar as that seen among macrophages, depending on their microenvironment. MSCs express toll-like receptors (TLRs), which represent one of the gates for determining their immunomodulatory activities [30, 31, 113]. LPS-induced activation of TLR4 on MSCs results in activation of the inflammatory pathway and the secretion of pro-inflammatory mediators. On the other hand, activation of TLR3 on MSCs induces the production of anti-inflammatory mediators [31]. These data indicate that MSCs need activation to exert their immunomodulatory properties. At present, the mechanisms involved in MSC activation remain partially known and further investigations are needed to potentially involve them in the development of reparative strategies that involve preconditioned MSCs.

In regenerative tissues, an injury leads to increases in chemokines and cytokines which induce the progressive homing and exposure of MSCs to activating stimuli that elicit their regulatory activities [79]. The SCI microenvironment contains many damage signals and inflammatory mediators that are also found in circulating cerebrospinal fluid and blood [44, 45]. Transplanting naïve MSCs in the injured spinal cord may therefore imply a stressful shock for the cells, which could compromise their survival and, thus, their effects on repair. Priming MSCs *in vitro* using inflammatory stimuli prior to transplantation could kick-start their machinery for increased production of anti-inflammatory cytokines before being

introduced to the injury microenvironment. Preconditioning of MSCs may enhance their survival and immunomodulatory capacity, potentially resulting in more efficient repair of the injured spinal cord.

In line with the need for priming before exposure to the injury environment, multipotent adult progenitor cells (MAPCs) are largely similar to MSCs in their immunomodulatory and reparative potential, while they have shown sensitivity to the injection time when transplanted after injury [114]. MAPCs modulate macrophage polarization, reduce axonal dieback, and elicit significant functional recovery, when injected systemically 1 day after a contusive SCI in rats [114]. The injury site also needs maturation to “allow” efficient actions of MSCs; immune cells need to remove debris, while endothelial cells and others need time to express adhesion molecules to bind MSCs. MSCs need anchoring to elicit their secondary signaling and repair-related expression profile [79–81]. This evidence supports the need to optimize the strategies for combining MSCs preconditioning with the optimal injection time-points, with the objective of preserving the active MSCs in the injury site while they promote repair.

### MSC Preconditioning

The preconditioning strategy has been explored in various models of disease or trauma. In a model of pyelonephritis, an infectious disease that derives from an excessive inflammation disorder, MSCs were injected intravenously after preconditioning *in vitro* with activated leukocytes. This approach resulted in primed MSCs secreting enhanced amounts of TGF $\beta$ , matrix metalloproteinase-2 (MMP2), and glycogen synthase kinase-3 $\beta$  (GSK3 $\beta$ ), which all are inflammation suppressors, and improving the disease outcome [115]. Another study compared the immunomodulatory potential of MSCs from bone marrow of human donors by measuring their secretion of PGE2 and capacity to induce the shift in macrophage phenotype, after pre-activating them with various inflammatory stimuli. Preconditioned MSCs showed an increase in their secretion of PGE2 compared to controls and a stronger ability to induce M2-like macrophages in culture. Interestingly, not all inflammatory stimuli resulted in the same outcome; preconditioning with IFN $\gamma$  caused MSC-mediated induction of more inflammatory macrophages [76].

An alternative approach to improve MSC immunomodulation may be genetic modification of the cells, which is known as “intrinsic preconditioning.” Transplantation of MSCs overexpressing IL13 in a mouse model of SCI resulted in a significant improvement in anatomical repair and functional recovery compared to transplantation of unmodified MSCs. In addition to the anatomical and functional effects, transplanting the modified MSCs also resulted in an increase in the population of M2-like macrophages, demonstrating successful immunomodulation [116].

The abovementioned results contribute to the evidence that inflammatory priming can result in more efficient MSC-mediated immunomodulation. However, these studies also raise awareness about the need to unravel the mechanisms and pathways involved in the different situations. When designing the preconditioning and transplantation experiments, considering the inflammation window within the immunomodulation timeline is necessary to allow the crucial reparative actions of all inflammatory cells.

### Conclusions and Remarks

Unraveling the mechanisms underlying the interactions between macrophages and MSCs in the context of wound healing may provide tools to modify spinal cord nervous tissue to improve repair. Using transplanted MSCs to target inflammation provides the opportunity of combining therapeutic approaches that so far have mostly been addressed individually. MSCs may modulate inflammation as well as secrete paracrine factors that elicit neuroprotection. The maturation state of the injury site, i.e., the degree of inflammation and presence of cell adhesion molecules, is crucial for determining the optimal time of MSC transplantation. It needs to be kept in mind that MSC preconditioning may be an integral aspect of these potentially powerful repair approaches. Future mechanistic studies are necessary to unravel the true potential of MSC preconditioning and MSC-mediated immunomodulation for spinal cord repair.

**Required Author Forms** [Disclosure forms](#) provided by the authors are available with the online version of this article.

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