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RIPK1-TRAF2 interplay on the TNF/NF- κ B signaling, cell death, and cancer development in the liver

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Hepatocellular carcinoma (HCC) is one of the most common causes of cancer-related death worldwide, with an estimated 782,000 new cases and 745,000 deaths in 2012 (1). According to the SEER Cancer Statistic Review (2), the prognosis of liver cancer patients is still poor with 5-year survival rate of 17.5%, and incidents of new cases have been rising each year at ~3% on average over the last 10 years. The prominent factors associated with HCC include chronic infection of hepatitis viruses (HVB, HVC), chronic alcohol consumption, non-alcoholic steatohepatitis, and exposure to hepatotoxins (3). Chronic liver damage following inflammation and cell death, accompanied by regenerative processes, is a feature of HCC development (3), suggesting that understanding the molecular machinery involved in cell death and regeneration in the liver may help identify molecular mechanisms of HCC.

Receptor-interacting protein kinase 1 (RIPK1), a member of the receptor-interacting serine/threonine kinase family, has pivotal roles in inflammatory signaling and activation of multiple cell death pathways (4). The physiological functions of RIPK1 are well documented in inflammatory signaling; however, its role in cell death and HCC development accompanied by liver inflammation remains unclear. RIPK1-mediated signaling is activated upon binding of tumor necrosis factor (TNF) secreted by macrophages and other immune-inflammatory cells to the TNF receptor 1 (TNFR1) (5). Subsequently, TNFR1 recruits a membrane-associated complex I containing TNFR1-associated death domain protein (TRADD), RIPK1, and TNFR-associated factor 2 (TRAF2) (6). In this complex, RIPK1 functions as a scaffold by rapidly being polyubiquitinated, and this linear ubiquitination chain serves as a binding site to NF- κ B essential modulator (NEMO), which activates NF- κ B signaling following phosphorylation of I κ B (Figure 1). Meanwhile, disassembly of the complex I through activation of deubiquitylation enzymes (A20, CYLD) and subsequent deubiquitination of RIPK1 occurs to reform complex IIa and IIb (6,8,9). Complex IIa, consisting of FADD, caspase-8, and RIPK1, mediates apoptosis in a manner dependent on the kinase activity of RIPK1. RIPK1 kinase activity also plays a role in the activation of necroptosis by binding with RIPK3 as a complex IIb (Figure 1). To better understand the role of RIPK1-mediated signaling in HCC genesis, it is imperative to distinguish the dual functions of RIPK1 (as a scaffold and a kinase) in hepatocytes. TRAF2, which binds with RIPK1 to form complex I and is stabilized by RIPK1 in mouse embryonic fibroblasts

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(MEFs) (10), also plays a crucial role in mediating multiple pathways including NF- κ B, JNK, and TNFR1-induced apoptosis pathways (11). However, detailed mechanisms of how TRAF2 regulates these pathways in hepatocytes and its contribution to HCC development have not been investigated.

In the recent issue of *Cancer Cell*, Schneider *et al.* (7) address the *in vivo* physiological functions of RIPK1 and TRAF2 using liver parenchymal cell (LPC)-specific knockout mice for *RIPK1* (*RIPK1^{LPC-KO}*), as well as *TRAF2* (*TRAF2^{LPC-KO}*), since mice deficient for *RIPK1* and *TRAF2* die soon after birth due to systemic inflammation (12,13). *RIPK1^{LPC-KO}* mice have normal liver function without spontaneous HCC development; however, upon LPS administration, *RIPK1^{LPC-KO}* mice show dramatic induction of apoptosis and liver damage with normal response to NF- κ B, as compared with wild-type mice (Table 1). *TRAF2^{LPC-KO}* mice also do not spontaneously develop HCC but show slight increase in NF- κ B activity, apoptosis, and liver damage (Table 1). On the other hand, double knockout mice for *RIPK1^{LPC-KO}/TRAF2^{LPC-KO}* and *TRAF2^{LPC-KO}/IKK β ^{LPC-KO}* spontaneously show apoptosis and damage in the liver with HCC development; NF- κ B activation by TNF stimulation in primary hepatocytes from these mice is impaired (Table 1). Moreover, concomitant deletion of caspase-8 completely rescues liver damage observed in *RIPK1^{LPC-KO}* mice, as well as spontaneous liver damage and HCC development in *RIPK1^{LPC-KO}/TRAF2^{LPC-KO}* mice (Table 1). They additionally show that loss of RIPK1 promotes ubiquitination and degradation of TRAF2 in mouse hepatocytes in a manner independent of kinase activity of RIPK1 using primary cells from mice lacking the kinase activity of RIPK1 (*RIPK1^{KD}*). These results indicate that RIPK1's scaffold function is required for stabilization of TRAF2 and that RIPK1 collaborates with TRAF2 for preventing caspase-8-mediated apoptosis and HCC development in mice.

As mentioned above, polyubiquitinated RIPK1 is shown to activate NF- κ B via NEMO-IKK-I κ B signaling. Their findings also suggest another scaffold function of RIPK1 in which RIPK1 stabilizes TRAF2 in hepatocytes, thereby inhibiting caspase-8-mediated apoptosis and liver damage (7). Intriguingly, deubiquitinated RIPK1, in turn, forms a complex IIa with FADD and caspase-8 to induce apoptosis or a complex IIb with RIPK3 to induce necroptosis (14-16) (Figure 1).

Kinase activity of RIPK1 is essential for its pro-apoptotic function in complex IIa and necroptosis through formation of complex IIb with RIPK3 (17). Previously, Kondylis *et al.* (15) show that deletion of *RIPK3* fails to suppress spontaneous liver damage and HCC development observed in *NEMO^{LPC-KO}* mice. To further examine whether necroptosis is involved in liver damage in the *RIPK1*-null background, Schneider *et al.* (7) administer LPS into *RIPK1^{LPC-KO}* or *RIPK1^{LPC-KO}/RIPK3^{-/-}* mice. However, *RIPK1^{LPC-KO}/RIPK3^{-/-}* mice show similar apoptosis and liver damage phenotypes to those in *RIPK1^{LPC-KO}* mice, although *RIPK1^{LPC-KO}/RIPK3^{-/-}* mice have delayed activation of caspase-8 and caspase-3 than *RIPK1^{LPC-KO}* mice. These results indicate that necroptosis by RIPK3 is not involved in liver damage induced by loss of RIPK1 or NEMO in mice, and RIPK3 has an unappreciated role as a regulator of apoptosis in addition to necroptosis.

To further understand functional association between RIPK1 and TRAF2 and the involvement of NF- κ B signaling in spontaneous HCC genesis, Schneider *et al.* (7) investigate liver damage and HCC development in *RIPK1^{LPC-KO}/TRAF2^{LPC-KO}* and *TRAF2^{LPC-KO}/IKK β ^{LPC-KO}* mice, in comparison with those in *TRAF2^{LPC-KO}* mice. Both *RIPK1^{LPC-KO}/TRAF2^{LPC-KO}* and *TRAF2^{LPC-KO}/IKK β ^{LPC-KO}* mice spontaneously develop liver damage and HCC with impaired NF- κ B activity; whereas, *TRAF2^{LPC-KO}* mice, which show functional NF- κ B activity and slightly increased apoptosis/damage in the liver, fail to develop HCC. These results might imply that RIPK1 and IKK β have anti-apoptotic or liver protective functions other than NF- κ B regulation. Data may also suggest that impaired NF- κ B function accelerates liver damage and HCC genesis induced by loss of TRAF2; however, it is unclear how exactly impaired NF- κ B function contributes toward spontaneous HCC development. Luedde *et al.* (18) previously show that *NEMO^{LPC-KO}* mice with impaired NF- κ B function spontaneously develop HCC following nonalcoholic steatohepatitis; whereas, *IKK β ^{LPC-KO}* mice which also have impaired NF- κ B function fail to develop steatohepatitis and HCC. Furthermore, complete deletion of NF- κ B by knocking out all the subunits of NF- κ B in mice (*NF- κ B^{LPC-KO}*) does not induce HCC (15). Thus, loss of NF- κ B alone does not induce HCC; however, loss of NF- κ B could collaborate with other genetic alterations toward liver damage and spontaneous HCC development. In contrast to the aforementioned possible tumor suppressive role of NF- κ B in HCC genesis, pro-carcinogenic function of NF- κ B is demonstrated in a model of inflammation-associated HCC, in which concomitant deletion of *IKK β* abolishes spontaneous HCC development observed in *Mdr2^{-/-}* mice (19). Thus, contributions of NF- κ B to HCC development are controversial and depend on physiological settings or cellular contexts.

The clinical relevance of RIPK1 and TRAF2 to human HCC progression is also demonstrated in this paper (7). Low expression of RIPK1 and/or TRAF2 has strong correlation with worse prognosis in patients with HCC; however, detailed analyses of apoptosis and NF- κ B activity in tumors warrant further investigation. The mechanisms behind reduced expression of RIPK1 and TRAF2 in human HCC also remain to be elucidated. Since expression levels of RIPK1 and TRAF2 vary among cancer types (20), it would be important to inquire whether RIPK1 and/or TRAF2 inhibit apoptosis and tumor progression in other cancer types. At present, there is no strategy that can prevent development of HCC resulting from liver inflammation, apoptosis, and regeneration. Further molecular analyses underlying liver damage and HCC development would help identify novel targets for HCC therapy.

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References

1. Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015; 136:E359–86. [PubMed: 25220842]

2. Howlader N, Noone AM, Krapcho M, et al. National Cancer Institute; Bethesda, MD: 2016. SEER Cancer Statistics Review, 1975-2013. Available online: http://seer.cancer.gov/csr/1975_2013
3. Sherman M. Hepatocellular carcinoma: epidemiology, risk factors, and screening. *Semin Liver Dis.* 2005; 25:143–54. [PubMed: 15918143]
4. Saleh D, Degtrev A. Emerging Roles for RIPK1 and RIPK3 in Pathogen-Induced Cell Death and Host Immunity. *Curr Top Microbiol Immunol.* 2017; 403:37–75. [PubMed: 26385769]
5. Peltzer N, Darding M, Walczak H. Holding RIPK1 on the Ubiquitin Leash in TNFR1 Signaling. *Trends Cell Biol.* 2016; 26:445–61. [PubMed: 26877205]
6. Ofengeim D, Yuan J. Regulation of RIP1 kinase signalling at the crossroads of inflammation and cell death. *Nat Rev Mol Cell Biol.* 2013; 14:727–36. [PubMed: 24129419]
7. Schneider AT, Gautheron J, Feoktistova M, et al. RIPK1 Suppresses a TRAF2-Dependent Pathway to Liver Cancer. *Cancer Cell.* 2017; 31:94–109. [PubMed: 28017612]
8. Wertz IE, O'Rourke KM, Zhou H, et al. De-ubiquitination and ubiquitin ligase domains of A20 downregulate NF-kappaB signalling. *Nature.* 2004; 430:694–9. [PubMed: 15258597]
9. Moquin DM, McQuade T, Chan FK. CYLD deubiquitinates RIP1 in the TNF α -induced necrosome to facilitate kinase activation and programmed necrosis. *PLoS One.* 2013; 8:e76841. [PubMed: 24098568]
10. Gentle IE, Wong WW, Evans JM, et al. In TNF-stimulated cells, RIPK1 promotes cell survival by stabilizing TRAF2 and cIAP1, which limits induction of non-canonical NF-kappaB and activation of caspase-8. *J Biol Chem.* 2011; 286:13282–91. [PubMed: 21339290]
11. Borghi A, Verstrepen L, Beyaert R. TRAF2 multitasking in TNF receptor-induced signaling to NF- κ B, MAP kinases and cell death. *Biochem Pharmacol.* 2016; 116:1–10. [PubMed: 26993379]
12. Kelliher MA, Grimm S, Ishida Y, et al. The death domain kinase RIP mediates the TNF-induced NF-kappaB signal. *Immunity.* 1998; 8:297–303. [PubMed: 9529147]
13. Yeh WC, Shahinian A, Speiser D, et al. Early lethality, functional NF-kappaB activation, and increased sensitivity to TNF-induced cell death in TRAF2-deficient mice. *Immunity.* 1997; 7:715–25. [PubMed: 9390694]
14. Dondelinger Y, Jouan-Lanhuet S, Divert T, et al. NF- κ B-Independent Role of IKK α /IKK β in Preventing RIPK1 Kinase-Dependent Apoptotic and Necroptotic Cell Death during TNF Signaling. *Mol Cell.* 2015; 60:63–76. [PubMed: 26344099]
15. Kondylis V, Polykratis A, Ehlken H, et al. NEMO Prevents Steatohepatitis and Hepatocellular Carcinoma by Inhibiting RIPK1 Kinase Activity-Mediated Hepatocyte Apoptosis. *Cancer Cell.* 2015; 28:582–98. [PubMed: 26555174]
16. Wang L, Du F, Wang X. TNF-alpha induces two distinct caspase-8 activation pathways. *Cell.* 2008; 133:693–703. [PubMed: 18485876]
17. He S, Wang L, Miao L, et al. Receptor interacting protein kinase-3 determines cellular necrotic response to TNF-alpha. *Cell.* 2009; 137:1100–11. [PubMed: 19524512]
18. Luedde T, Beraza N, Kotsikoris V, et al. Deletion of NEMO/IKKgamma in liver parenchymal cells causes steatohepatitis and hepatocellular carcinoma. *Cancer Cell.* 2007; 11:119–32. [PubMed: 17292824]
19. Haybaeck J, Zeller N, Wolf MJ, et al. A lymphotoxin-driven pathway to hepatocellular carcinoma. *Cancer Cell.* 2009; 16:295–308. [PubMed: 19800575]
20. Forbes SA, Beare D, Boutselakis H, et al. COSMIC: somatic cancer genetics at high-resolution. *Nucleic Acids Res.* 2017; 45:D777–83. [PubMed: 27899578]

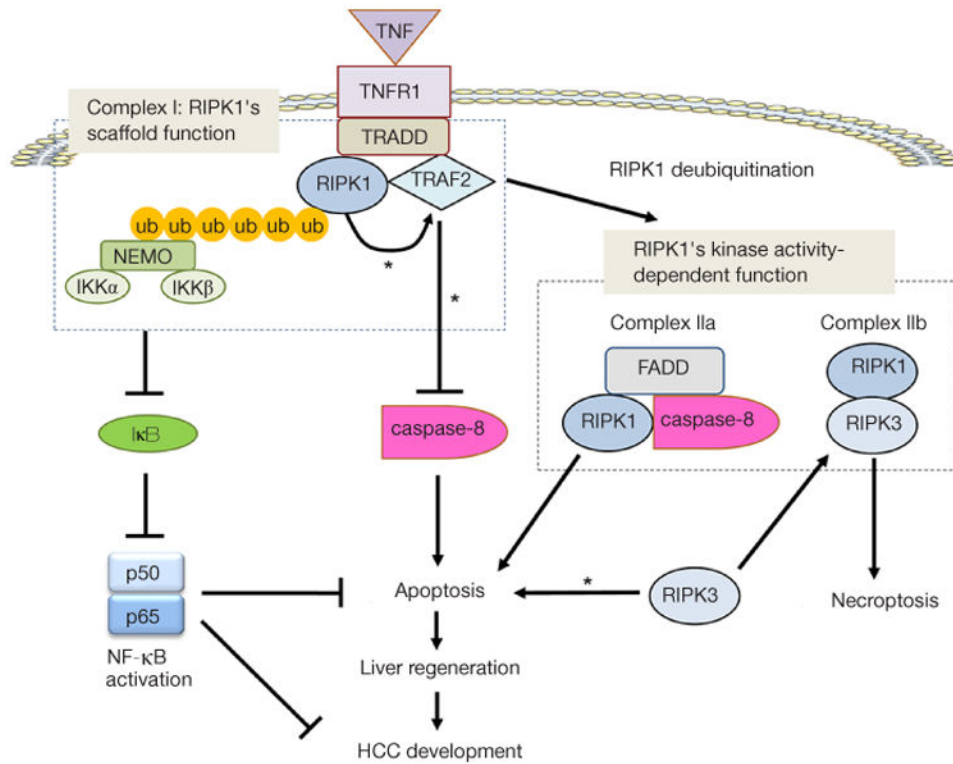


Figure 1. The TNF-RIPK1-mediated signaling and liver tumorigenesis. TNF ligation stimulates complex I formation containing TRADD, RIPK1, TRAF2, and NEMO to inhibit apoptosis via NF- κ B and caspase-8 (scaffold function of RIPK1). Deubiquitinated RIPK1 forms complex IIa with FADD and caspase-8 to induce apoptosis, while it can also form complex IIb with RIPK3 to induce necroptosis, depending on the cellular context (kinase function of RIPK1). *, discovered by Schneider *et al.* (7). TNF, tumor necrosis factor; TNFR1, tumor necrosis factor receptor 1; TRADD, TNFR1-associated death domain protein; RIPK1, receptor-interacting protein kinase 1; TRAF2, tumor necrosis factor receptor-associated factor 2; NEMO, NF- κ B essential modulator; IKK, inhibitor of nuclear factor kappa-B kinase; FADD, FAS-associated protein with death domain; HCC, hepatocellular carcinoma.

Table 1
Liver damage, NF- κ B activity, and HCC development in mouse models

Genotype	TNF/LPS stimulation	Apoptosis ¹	Liver damage ²	NF- κ B activity ³	HCC	Rescue by caspase- δ knockout
WT	-	-	-	+/-	-	
<i>RIPK1^{LPC-KO}</i>	+	-	-	++	N.D.	
<i>RIPK1^{LPC-KO}/RIPK3^{-/-}</i>	-	-	-	+/-	-	
	+	++++	+++	++	N.D.	Complete rescue
<i>RIPK1^{LPC-KO}/RIPK3^{-/-}</i>	-	N.D.	-	+/-	N.D.	
<i>TRAF2^{LPC-KO}</i>	+	++++ (delay)	+++	++	N.D.	
	-	+	+/-	+	-	
<i>RIPK1^{LPC-KO}/TRAF2^{LPC-KO}</i>	+	N.D.	N.D.	+++	N.D.	
	-	++	++	+/-	+++	Complete rescue
<i>TRAF2^{LPC-KO}/IKKβ^{LPC-KO}</i>	+	N.D.	N.D.	+/-	N.D.	
	-	+++	+	+/-	+	
<i>TRAF2^{LPC-KO}/IKKβ^{LPC-KO}</i>	+	N.D.	N.D.	+/-	N.D.	

¹ extent of apoptosis is determined based on cleaved caspase-3-positive cells in hepatocytes.

² extent of liver damage is determined based on the values of aspartate aminotransferase and alanine aminotransferase.

³ extent of NF- κ B activity is determined based on the results of electrophoretic mobility shift assay (EMSA).

N.D., not determined; HCC, hepatocellular carcinoma; TNF, tumor necrosis factor; LPS, lipopolysaccharide; WT, wild-type; RIPK1, receptor-interacting protein kinase 1; TRAF, tumor necrosis factor receptor-associated factor; IKK β , inhibitor of nuclear factor kappa-B kinase subunit beta.