

***Toxoplasma gondii*: Biological Parameters of the Connection to Schizophrenia**

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It is increasingly evident that the brain is not truly an immune privileged site and that cells of the central nervous system are sensitive to the inflammation generated when the brain is fighting off infection. Among the many microorganisms that have access to the brain, the apicomplexan protozoan *Toxoplasma gondii* has been one of the most studied. This parasite has been associated with many neuropsychiatric disorders including schizophrenia. This article provides a comprehensive review of the status of *Toxoplasma* research in schizophrenia. Areas of interest include (1) the limitations and improvements of immune-based assays to detect these infections in humans, (2) recent discoveries concerning the schizophrenia–*Toxoplasma* association, (3) findings of *Toxoplasma* neuropathology in animal models related to schizophrenia pathogenesis, (4) interactions of *Toxoplasma* with the host genome, (5) gastrointestinal effects of *Toxoplasma* infections, and (6) therapeutic intervention of *Toxoplasma* infections.

Key words: microorganism/psychiatric disorders/pathogenesis

Introduction

Converging evidence points to a central role of the immune system in the etiopathogenesis of schizophrenia and related psychiatric disorders. Involvement of the immune system is supported by genetic, neuropathological, neuroimaging, and metabolic analyses of multiple populations.^{1,2} The brain has been generally considered an immune-privileged environment due to the perceived exclusion of immune components and the blood–brain barrier. However, recent discoveries indicate that the brain is not truly immune privileged, and inflammatory events will occur when the brain is fighting off infection, potentially leading to profound structural and functional

changes. The inflammatory response is recognized as a causal factor in the pathology and chronic course of many central nervous system (CNS) diseases.³ Causes of inflammation have been related to a variety of cues, including infection, traumatic brain injury, toxic metabolites, or autoimmunity.⁴

Biological and Epidemiological Characteristics of Microbial Agents Likely to Be Associated With Schizophrenia

Schizophrenia is a persistent brain disease that is usually first evident in late adolescence or early adulthood and proceeds along a lifelong course. If a microbial agent contributes to schizophrenia pathogenesis, there are a number of biological and epidemiological requirements that the agent must fulfill to be a plausible candidate organism. For example, it might be expected that the candidate infectious agent or group of agents demonstrate a worldwide distribution as is similar for schizophrenia. Furthermore, given the schizophrenia is likely a disorder of neurodevelopment, the timing of exposure to a putative infectious agent is likely to occur during time periods important for neurodevelopment.^{5,6} The candidate agent must likewise persist in the brain, as numerous studies indicate some degree of immune dysregulation and inflammation in the brains of many individuals with schizophrenia.⁷ Since neurological destruction may not be present early in the course of schizophrenia, it is also likely that such organisms can persist in a nonreplicating (latent) or very slowly replicating (inactive) state for extended periods of time. Thus, the reactivation of a latent or slowly replicating infection would be consistent with early exposure and development of symptoms later in life. Causes of such reactivation might include normal endocrine and other developmental processes

associated with adolescence as well as exposure to infectious agents more commonly infecting individuals in this time period. In addition, although there are seasonal variations, the rates of occurrence of schizophrenia are relatively constant from year to year and do not differ greatly across geographic areas.^{8,9} With respect to schizophrenia, 2 models for endemic infections may be relevant. One scenario involves infectious agents with high prevalence that are spread person to person with inapparent symptoms on exposure. Another potential source of endemic agents would be ones transmitted through the environment by common sources such as food, water, or soil. In both cases the exposures are relatively constant and would show low rates of epidemic variation. Finally, family studies indicate a strong association of shared risk of schizophrenia within families¹⁰ and genes modulating the immune response are identified as significant components of the genetic risk for schizophrenia. It is thus likely that infectious agents associated with schizophrenia are ones that interact with host genetic factors. The interaction with genetic factors would also serve to explain how endemic infectious agents with relatively high prevalence might be associated with disorders that are less prevalent, because clinically apparent disease would be most likely to result from exposure to the infectious agent by genetically susceptible individuals. It is of note that these familial factors need not be limited to Mendelian inherited genes but would also include epigenetic modifications and microbial genes carried by a host as part of the inherited microbiome.

Seroepidemiological Studies of *Toxoplasma* and Psychiatric Disorders

Although there are several pathogenic agents that meet these criteria, the one which has been best studied in the context of schizophrenia is the apicomplexan protozoan *Toxoplasma gondii*. *Toxoplasma* is one of the most pervasive pathogens that infects approximately a billion people worldwide. Although initial infection with *Toxoplasma* is associated with few symptoms in immune-competent individuals, the host is often left with lifelong persistence of tissue cysts in the brain, retina, and muscles. When the immune response wanes, tissue cysts can become reactivated and invade any nucleated cell, as noted in immunocompromised patients. The parasite has a complex life cycle involving sexual replication in members of the cat family (Felidae) and asexual propagation in a wide variety of warm-blooded hosts.¹¹ There are 3 infectious stages in the life cycle of *Toxoplasma*: tachyzoites, which facilitate expansion during acute infection; bradyzoites (in tissue cysts), which maintain chronic infection; and sporozoites (in oocysts), which are disseminated in the environment.¹² For the purposes of this article, we will provide a comprehensive review of the status of *Toxoplasma* research in schizophrenia.

Multiple epidemiological studies and meta-analyses have documented that individuals with psychiatric disorders have increased odds of *Toxoplasma* exposure.¹³ However, there have also been studies that have not found this association. Many of the negative studies were performed in individuals with established schizophrenia living in populations with relatively low prevalence rates of *Toxoplasma* infection. This discrepancy was largely resolved by a recent study indicating that, in a low prevalence population, *Toxoplasma* exposure was associated with an increased risk in individuals with the recent onset of psychosis but not in individuals with established schizophrenia (figure 1).¹⁴ The difference is likely related to the timing of *Toxoplasma* exposure relative to the onset of symptoms. It is also possible that the antiparasitic effects of some of the medications commonly used to treat schizophrenia affect the levels of antibodies.¹⁵ Consistent with this possibility is the recent report of increased levels of *Toxoplasma* antibodies in individuals with treatment resistant forms of schizophrenia.¹⁶

Although most studies of *Toxoplasma* have examined associations with schizophrenia, increased rates of exposure to *Toxoplasma* have been found in a range of other psychiatric disorders including psychotic-like symptoms,¹⁷ bipolar disorder,^{18,19} self-directed violence²⁰ and suicide attempts,²¹ general anxiety disorders,²² mixed anxiety and depressive disorder,²³ obsessive-compulsive disorder,²⁴ Autism,²⁵ and depression during pregnancy.²⁶

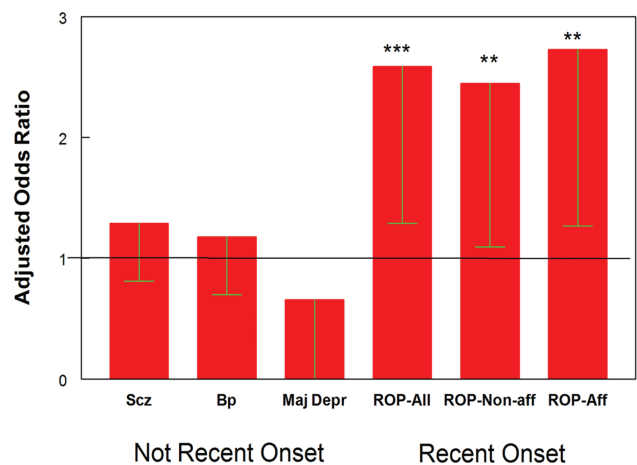


Fig. 1. Odds ratios associated with *Toxoplasma* exposure by clinical diagnosis. Bars represent odds ratios (Mean-95% confidence interval) associated with the indicated clinical diagnosis calculated by logistic regression as described in the text with age, gender, race, maternal education, and place of birth as covariates. The abbreviations used are as follows: Scz = nonrecent schizophrenia; Bp = nonrecent bipolar disorder; Maj Depr = major depressive disorder without recent onset psychosis; ROP-All = recent onset psychosis, all cases; ROP-Non-aff = recent onset psychosis, individuals with nonaffective psychosis; ROP-Aff = recent onset psychosis, individuals with affective psychosis. *** $P < .003$, ** $P < .03$. Not recent onset refers to individuals who did not have recent onset of psychosis. Reprinted from *PLoS Negl Trop Dis*. 2017 Nov; 11(11): e0006040.

It is worth mentioning that there are some controversies about the role of *Toxoplasma* in major depression.²⁷ *Toxoplasma* seropositivity has also been associated with alterations in cognitive functioning and overall cognitive decline in the elderly.^{28,29} At the other end of the age spectrum, exposure to *Toxoplasma* has been associated with decreased reading skills and memory capacity in school-aged children.³⁰ In some cases, psychiatric disorders such as suicide attempts were associated with specific levels of antibodies rather than just seroprevalence as in the case of suicide attempts³¹ and personality disorders.³² Moreover, studies have shown that latent *Toxoplasma* infections could have serious impacts on human health.³³ These findings suggest that exposure to *Toxoplasma* is not only a risk factor for schizophrenia but for a range of other disorders as well. These findings are consistent with a recently increased understanding of the shared genetic susceptibilities among psychiatric disorders and the overlapping of clinical phenotypes.

Limitations and Improvements of Immune-Based Assays for Characterization of *Toxoplasma* Infection

Because *Toxoplasma* persists largely in the form of tissue cysts in the brain, it is often not possible to directly detect organisms in infected individuals. Most studies of human infection rely almost entirely on the measurement of anti-*Toxoplasma* antibodies. The IgG class antibodies to *Toxoplasma* provide a marker of past exposure to the organism. However, with an increased understanding of the biology of the parasite, information related to the timing, the strain type, the infectious stage, the existence, or persistence of infection have become increasingly important.^{34–36}

Recent studies have revealed that *Toxoplasma* populations are geographically structured and have genetic compositions that vary among strains.³⁴ However, currently available solid phase immunoassays were developed in the 1970s to detect strains that were circulating at that time.³⁷ This raises concern whether these immunoassays have sufficient sensitivity to detect contemporary strains in all populations. We have found that many individuals who are “seronegative” using standard enzyme-linked immunosorbent assay (ELISA) nevertheless have evidence of reactivity to *Toxoplasma* proteins visualized by Western blotting techniques. For example, in a pilot study of 34 individuals, 25 of whom had psychiatric disorders (bipolar disorder, schizophrenia, recent onset psychosis) only 3 individuals (8.2 %) were IgG seropositive to *Toxoplasma* as defined by a commercially available assay (obtained from IBL America). On the other hand, 12 individuals (35.3%) showed clear reactivity to different *Toxoplasma* proteins by Western blot (figure 2). These findings indicate that standard assays may substantially underestimate the prevalence of *Toxoplasma* infection in a population and its effect on health and disease. These

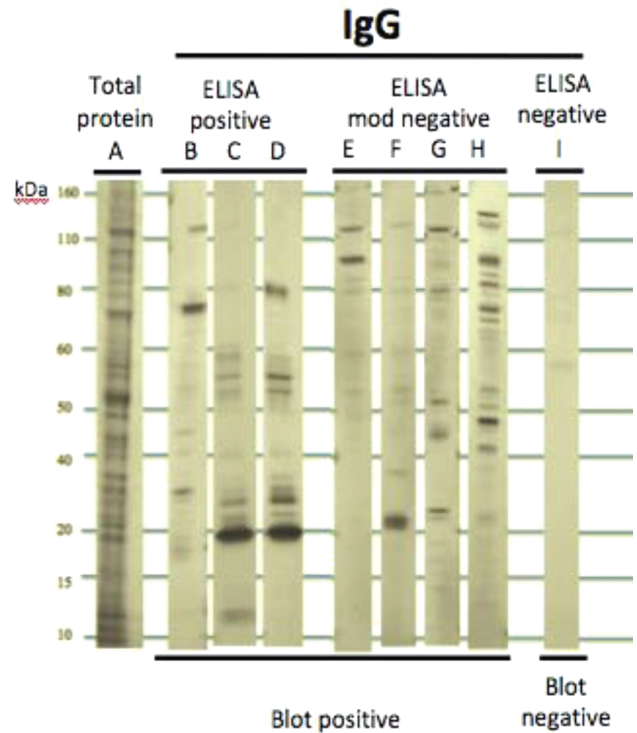


Fig. 2. Underestimates of *Toxoplasma* seropositivity revealed by immunoblotting. We performed a pilot study of 34 individuals, 25 of whom had psychiatric disorders. Cellular lysate from *Toxoplasma gondii* strain RH were homogenized and subjected to sodium dodecyl sulfate–polyacrylamide gel electrophoresis (SDS–PAGE) and Western blotting using the human sera characterized in ELISAs. A total protein stain of the cellular lysate is shown in the first lane. Standards listed in the far left of the diagram refer to kilodaltons. ELISA mod negative refers to an absorbance value that was slightly elevated from a true negative value. ELISA positive refers to an absorbance value that was slightly below the positivity cutoff value. Twelve of 34 individuals were positive for IgG via immunoblotting compared to 3 who were positive based on ELISAs.

findings are also of interest in light of studies purporting decreasing levels of antibodies to *Toxoplasma* in individuals living in the United States³⁸ and Europe³⁹ because such studies might be reflective of antigenic variation rather than truly decreased prevalence. Recently, much progress has been made in immune-based assays to characterize *Toxoplasma* infection.

Serological Determination of Cyst Burden

Because, as noted earlier, tissue cysts are predominantly located in the brain, measurement of parasite burden is difficult to achieve in living humans and animals. To overcome this limitation, we have developed a noninvasive, highly sensitive, and specific method for detecting tissue cysts in any disease state. This assay measures humoral immune response against a *Toxoplasma* matrix antigen called MAG1 using synthetic peptides.^{40,41} MAG1 is abundantly expressed within the cyst and in the cyst wall

surrounding the bradyzoites.⁴² Tissue cysts of *Toxoplasma* were found in brains of all MAG1-seropositive mice, whereas no cysts were detected in MAG1-seronegative mice. We have shown that MAG1 antibody level is strongly correlated with the number of brain cysts in the infected host ($r = 0.82$, $P = 0.0021$). For mice that remained MAG1 seronegative after exposure, we performed a bioassay to examine the possibility of latent infection below the detection limit. One month after the feeding of brains of MAG1 seronegative mice, sera from recipient mice were tested for the presence of *Toxoplasma* antibodies. None of mice ($n = 5$) generated the specific antibody confirming that MAG1 seronegative mice did not become latently infected and that they were free from brain cysts. These data suggest that MAG1 antibody can serve as a serologic marker for evidence of chronic infection and for cyst burden.

Serological Detection of Infecting Serotype and Form

Given *Toxoplasma* strains vary in their pathogenic potential, we and others have developed peptide-based assays to distinguish infection with the 3 main recognized types of *Toxoplasma*.^{43,44} These assays are capable of distinguishing the type of *Toxoplasma* infections in many individuals. We used this assay to identify serotypes of *Toxoplasma* in 219 pregnant women whose children developed schizophrenia and psychotic illnesses in adult life and 618 matched unaffected control mothers. We found that the offspring of mothers with virulent strain of *Toxoplasma* infection were at significantly increased risk for the development of psychoses as compared with the matched unaffected control mothers.⁴⁴ However, there were also a substantial number of cases who were cross-reactive or indeterminate, probably as a result of infection with more than one type or from a type variant from one of the recognized strains. The development and widespread application of assays capable of accurately identifying infecting type, as well as ones capable of distinguishing infections caused by oocysts as opposed to tissue cysts, would represent major tools for an improved understanding of *Toxoplasma* biology and epidemiology.

Heterogeneity in Infection State and Behavioral Effect

There is marked variation in the human response to *Toxoplasma* infection. Apart from a human genetic component, heterogeneity of host response is likely to arise through differences in parasite genotype/serotype,⁴⁵ the infection state,⁴¹ the infectious form at exposure,⁴⁶ and the timing of first exposure. The present review focuses on the necessity to distinguish infection states between persistent infection with brain-dwelling tissue cysts and resolved infection with specific anti-*Toxoplasma* antibodies (failed to progress to chronic stage). In a study reported from the Sapolsky laboratory,⁴⁷ changes in

behavior were found to be associated with cyst presence. In their study, all *Toxoplasma*-exposed rats tested positive for serum anti-*Toxoplasma* IgG, but only a subset of *Toxoplasma*-exposed rats developed chronic infection with cysts in the forebrain. The presence of behavioral changes including predator odor aversion and anxiety-related behavior were all linked with cyst presence. In another study, Afonso et al.⁴⁸ demonstrated that cysts, as well as manifestations of infection, were only evident in a subpopulation of infected C57BL/6J mice. The mice with brain cysts, versus those without cysts, displayed a plethora of behavioral alterations.

Toxoplasma type I strains do not readily develop tissue cysts or latent infection in laboratory mice. Recent studies from our group, which utilized a chronic model of type I infection, further addresses why abnormalities occur in some mice, but not in others. In our study,⁴⁰ we found that outbred mice following exposure to the virulent type I strain (GT1) displayed variable outcomes ranging from aborted to severe infections, characterized by antibody profiles, alterations in cytokine and gene expression, and behavior changes. The extent of most changes was directly correlated with levels of cyst burden determined by MAG1 antibody. We found that mice with high levels of MAG1 antibody (MAG1 > 0.5) exhibited reduced locomotor and exploratory activity, impaired object recognition memory, and lack of response to amphetamine induced activity. These changes were not found in mice with a lower level of cyst burden (MAG1 < 0.5) or mice that were acutely but not chronically infected (lack of MAG1 antibodies).

These variable phenotypes displayed in the rodent models may be analogous to the diversity of human responses. Previously, we measured MAG1 antibody responses in serum samples from 22 patients with clinical toxoplasmosis and from 26 patients with serological evidence of past exposure to *Toxoplasma* (more than 1 year infection).⁴¹ We found that not all *Toxoplasma* seropositive individuals develop MAG1 antibodies, and MAG1 antibody level can differentiate active from inactive human toxoplasmosis. The finding of tissue cysts and altered behavior in only a subset of infected mice is consistent with several human studies. For example, Luft and Remington⁴⁹ reported that 50% of *Toxoplasma*-seropositive HIV/AIDS patients showed no symptoms of toxoplasmic encephalitis despite being highly immune compromised.

Neuropathogenesis Is Crucial to Explain the Behavioral Changes

Most studies on rodents have shown that *Toxoplasma* cysts have a widespread distribution within the brain.⁵⁰ In vivo studies have revealed that these cysts are located within neurons but not other cell types of the CNS.^{51,52} To analyze which parts of neurons are infected by cysts, Haroon et al.⁵³ performed a detailed immunohistochemical

investigation. They found that parasitic antigens are not sequestered in cysts but reside in all major parts of neurons including the neuronal soma, dendrites, and axons. Moreover, recent analysis of cyst biology reveals a dynamic and replicating bradyzoite population.⁵⁴ This is in contrast to the usual view that bradyzoites are dormant, poorly replicating, or nonreplicating entities. These findings implicate that tissue cysts have extraordinary potential to alter the structure of neurons and their connections, thereby affecting behavior.

Persistence of tissue cysts requires a continuous immune response provided by resident CNS and/or infiltrating peripheral immune cells to prevent parasite reactivation and encephalitis. Previous data demonstrate that a low-grade inflammation is often triggered throughout the brain. For example, activated microglia and astrocytes often physically encircle parasitic cysts, complement factor C1q locates in the vicinity of tissue cysts, and brain magnetic resonance imaging showed mild to moderate ventricular dilatation.^{47,55–57} This inflammatory milieu may influence the function and morphology of neurons, leading to brain activity and connectivity changes.

The Morphology of Neurons is Altered by Tissue Cysts

In a study focused on the position of *Toxoplasma*-induced lesions, Parlog et al.⁵⁸ reveal impaired local connectivity evidenced by altered fiber density, loss of fiber continuity, and reduced levels of synaptic proteins PSD95 and synaptophysin. In a study to characterize CNS pathology with chronic infection, David et al.⁵⁹ reported alterations in neuronal morphology (a reduction in dendritic spines) and network activity. To address the effect of latent *Toxoplasma* infection in schizophrenia, Horacek et al.⁶⁰ studied whole-brain voxel-based morphometry of gray and white matter. They found a significant reduction of gray matter volume in *Toxoplasma* positive compared with *Toxoplasma*-negative patients.

Toxoplasma Infection and Neurotransmission

Alterations in brain connectivity can have potential downstream effects in the pathways of neurotransmitters. Previous studies have reported several disrupted neurotransmitter pathways including dopamine, gamma-aminobutyric acid (GABA), and glutamate.^{59,61}

Glutamate is arguably the most important neurotransmitter in the brain and implicated in the pathogenesis of schizophrenia. Converging evidence support a significant disruption in the glutamate signaling following infection. First, David et al.⁵⁹ found a significant reduction in astrocyte glutamate transporter (GLT-1). One of the major roles of astrocytes is to remove extracellular glutamate to prevent neuroexcitotoxicity. As expected, downregulation of GLT-1 is linked to an increase in extracellular glutamate in infected animals. Second, *Toxoplasma* is a

tryptophan auxotroph. Interferon gamma (IFN γ) suppresses the growth of the parasite through acceleration of tryptophan degradation.⁶² IFN γ has the ability to regulate the critical step of the kynurenine pathway, the major pathway of tryptophan metabolism.⁶² Metabolism further along this pathway produces kynurenic acid (KYNA), an antagonist of the *N*-methyl-D-aspartate receptor (NMDAR). Elevated levels of KYNA can block the glutamate recognition site of the NMDAR. As rodents infected with *Toxoplasma* and patients with schizophrenia have increased KYNA levels in brain,⁶³ KYNA has been hypothesized to be a pathogenic link between *Toxoplasma* infection and schizophrenia. Third, glutamate signaling could potentially be disrupted by GABAergic signaling. The distribution of glutamic acid decarboxylase 67 (GAD67), the enzyme responsible for converting glutamate to GABA, exhibited profound changes in *Toxoplasma* infected animals.⁶¹ Finally, recent studies from our group reported that NMDAR autoantibodies are elevated in chronic murine toxoplasmosis.^{56,64} NMDARs are heteromers of GluN1 subunits that bind glycine and GluN2 subunits that bind glutamate. NMDAR autoantibodies are reported to decrease the availability of NMDAR binding sites through the process of internalization,⁶⁵ potentially leading to increase in glutamate levels, as noted by David et al.⁵⁹

In the context of schizophrenia, there has been the most interest in the effects of infection on levels of dopamine. The *Toxoplasma* genome contains 2 aromatic amino acid hydroxylase genes (AAH1 and AAH2) that encode proteins that can produce L-DOPA, the direct precursor of dopamine.⁶⁶ Therefore, an intriguing hypothesis has been put forth that *Toxoplasma* might influence dopamine production and hence dopamine synaptic transmission. As has been reviewed recently in detail,⁶⁷ many studies report changes in the dopamine pathway in both acute and chronic infection. For example, McConkey's group reported increased levels of dopamine in vitro and in vivo.⁶⁸ In human studies, the role of dopamine was suggested by decreased novelty seeking in *Toxoplasma*-infected patients and by a high level of positive symptoms in *Toxoplasma*-seropositive patients with schizophrenia.⁶⁹ Here, we focus on recent studies that evaluate the impact of AAH gene deficiency on biochemical and behavior changes. A study from the Sibley group reported that neither overexpression nor deletion of AAH2 from parasites altered dopamine levels in vitro or in vivo.⁷⁰ We and others found no effects of AAH2 deletion on the host's exploratory behaviors as well as dopamine-dependent behavioral alterations.^{71,72} Recently, studies found that the AAH genes appear to be critically involved in the sexual cycle of the parasite in the gut of the definitive feline host where they likely play a role in oocyst wall formation.⁷³ Although studies with Δ aaH2 parasites suggest that AAH2 is not required for biochemical and behavioral abnormalities observed in chronically

infected mice, dopamine remains an important question relating to *Toxoplasma* pathogenesis.

Interaction of *Toxoplasma* With the Host Genome

There is an increasing realization that susceptibility to common infectious agents can be heritable and that a significant portion of this heritability is based on genetic polymorphisms. In the case of *Toxoplasma*, studies of congenital infection have implicated a number of toll-like receptors and other immune response genes relating to increased risk.^{74,75} In regards to infection in later life, a population study of 1227 individuals from a Mexican American community in Texas found that seropositivity to *Toxoplasma* was heritable with a heritability factor (H2) of 0.21 ± 0.05 ($P < .001$). However, specific polymorphisms significantly associated with *Toxoplasma* seropositivity were not identified. A population study of Ashkenazi Jews found that the *Toxoplasma* seropositivity in individuals with schizophrenia was associated with polymorphisms located near genes encoding glucocorticoid-inducible kinase 1 (SGK1) and solute carrier family 2 member 12 (SLC2A12), although these associations did not achieve statistical significance on a genome-wide level.⁷⁶

CD8+ T cells and IFN γ are significant effectors mediating resistance to chronic *Toxoplasma* infection. However, recent studies found that chronic *Toxoplasma* infection results in CD8 \pm T cell exhaustion,⁷⁷ a state in which these cells lose their functional ability defined by their capacity to proliferate and secrete effector cytokines. As patients with schizophrenia have decreased levels of CD8 immunity,^{78–80} attenuated CD8 T cells have been proposed to play a role in mediating schizophrenia in *Toxoplasma*-seropositive patients.⁸¹ This hypothesis posits that the decreased CD8 functionality might result in reactivation of some of the quiescent *Toxoplasma* parasites, leading to focal necrosis and localized inflammation.

Nurr1 is an orphan nuclear receptor that is essential for the development and continued survival of mesencephalic dopamine neurons. Because rare mutations in Nurr1 have been reported in schizophrenia patients^{82,83} and were associated with attention deficits in patients with schizophrenia,⁸⁴ this receptor is implicated as a potential contributor to the development of schizophrenia. Eells et al.⁸⁵ tested how *Toxoplasma* infection interacts with the heterozygous deletion of the Nurr1 gene. Their results suggest that reduced Nurr1 function, as found in the Nurr1 +/- genotype, makes mice more susceptible to *Toxoplasma*-induced alterations in the dopamine-related behaviors (elevated open field activity and a trend toward disrupted sensorimotor gating). The data suggest that combination of the genetic mutations that predispose individuals to schizophrenia and *Toxoplasma* infection are capable of causing neurological effects.

Gastrointestinal Effects of *Toxoplasma* Infections

Although the main effects of *Toxoplasma* relating to schizophrenia are thought to be due to the development of an immune response and pathology within the brain, it is important to note that *Toxoplasma* generally infects humans through the gastrointestinal tract and, in doing so, can alter gastrointestinal structure and functioning.⁸⁶ On ingestion of the parasite by the host, *Toxoplasma* organisms migrate to the surface epithelium and lamina propria of the small intestine.⁸⁷ From here, organisms penetrate the blood–gut endothelial barrier and gain access to the circulatory system and host organs. Recent studies indicate that alterations in gastrointestinal functioning are common in schizophrenia and other serious psychiatric disorders and that some of the manifestations of these disorders may be related intestinal abnormalities through aberrations in the gut–brain axis.⁸⁸ In particular, primary studies have shown measures of intestinal inflammation correlate with exposure indices to the parasite in individuals with psychiatric disorders compared with controls.⁸⁹ Another way in which the gut–brain axis is affected is through changes in the composition of the intestinal microbiome because the intestinal microbiome has been found to be altered in individuals with recent onset psychosis.⁹⁰ It is thus of interest that acute infection with *Toxoplasma* has been shown to alter the intestinal microbiome in mice.⁹¹ We have also found that chronic *Toxoplasma* infection alters the composition of the intestinal microbiome in mice as long as 5 months following infection (figure 3). Changes in the host bacterial microflora in the upper intestinal tract may potentially contribute to behavior changes during chronic infection.

Therapeutic Intervention of *Toxoplasma* Infections Relevant to Psychiatric Disorders

Although there is a great deal of information pointing to a role for *Toxoplasma* infections in the etiopathogenesis of schizophrenia, the ultimate documentation of this association would be the demonstration that anti-*Toxoplasma* medications alter the clinical symptoms or course of psychiatric disorders in infected individuals. This goal is hampered by the lack of effective anti-*Toxoplasma* therapy particularly for the tissue cyst form of the organism. Barriers to effective therapy of tissue cysts include a low level of replication and metabolism, making these forms relatively resistant to medications directed at folate metabolism and other pathways employed by rapidly replicating organisms. The need for a therapeutic medication to cross the blood–brain barrier constitutes another potential barrier. There have been 4 studies evaluating the therapeutic efficacy of adjunctive antiparasitics agents in patients with schizophrenia, as recently reviewed by Chorlton.⁹² No trials have demonstrated a change in psychopathology with adjunctive treatment. However,

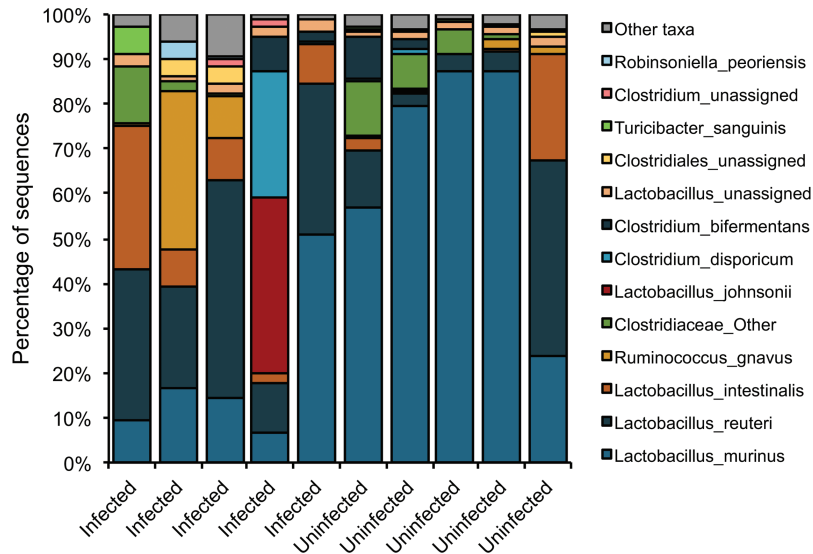


Fig. 3. Individual differences at the species level in chronic *Toxoplasma* infected and uninfected mice. Female *CD-1* mice were infected intraperitoneally with 500 GT1 tachyzoite, whereas uninfected controls received PBS only (5 per group). To establish a chronic infection, both control and infected mice were treated with sulfadiazine sodium (400 mg/L) from day 5 to 30. Mice were killed at 5 months postinfection. 16S rDNA libraries were sequenced using the MiSeq Illumina sequencing platform. Sequences were analyzed utilizing QIIME 1 focusing on v3–v4 region. The graph shows the relative bacterial composition of the intestinal microflora in each mouse. Species that has higher than 100 average counts are depicted individually, lower counts are combined in the “Other” taxa.

as noted earlier, possible reasons for treatment failure include selected drugs without evidence against chronic infection or used them at doses too low to reduce brain cyst burden. Our recent study showed, using a mouse model of chronic *Toxoplasma* infection, that abrogation of the inhibitory PD-1 signaling pathway that maintains T cells in an exhausted state significantly diminished brain cyst burden (77% lower).⁹³ The effect is likely mediated by brain leukocyte infiltration (CD3+ T cells, CD8+ T cells, and CD11b+ cells) through pathways connected to the cerebrospinal fluid-filled compartments such as ventricles and subarachnoid space. Our study provides proof of concept for blockade of immune checkpoint inhibitors as a therapy for chronic toxoplasmosis. Clinical trials will be necessary to document efficacy in humans and the potential role of this immunological approach in the treatment of psychiatric disorders.

Evolutionary Implications of *Toxoplasma* Infection

An etiological role for a zoonotic agent such as *Toxoplasma* can also provide an explanation for the persistence of psychiatric disorders such as schizophrenia within a human population. In the mouse model of *Toxoplasma* infection, the alterations in cognitive functioning and fear response result in behavioral changes associated with the likelihood of increased predation by felines, presumably, to facilitate the spread of the parasite to its definitive host. Although such behavior is detrimental to the host, it is beneficial to the organism in terms of sexual reproduction and the completion of the life cycle. Although vestigial in humans, who are unlikely

to be consumed by felines, the biological properties of the organism relating to altered cognition and fear response would be expected to be functioning regardless of the host. In fact, studies in humans have reported increased rates of risk assessment⁹⁴ (manifested as decreased levels of accident avoidance) and cognitive control⁹⁵ in some populations.

Conclusion

Numerous studies link exposure to *Toxoplasma* as a risk factor for schizophrenia and a range of other psychiatric disorders. The multifaceted effects of *Toxoplasma* infection on the gastrointestinal tract, neuroinflammation, neurodegeneration, and behavior are just starting to be understood. As a gastrointestinal pathogen, *Toxoplasma* might affect the brain through local and systemic inflammation or through changes in the intestinal microbiome. As a neurotropic pathogen, *Toxoplasma* might affect information processing within a wide variety of brain functional systems. Findings from epidemiology and psychiatric genetics have suggested a significant role of gene \times *Toxoplasma* interaction in psychiatric disorders. Experimental *Toxoplasma* infection provides a relevant model by which we can understand the complexity of human neurological diseases. Recent advances using human neurons derived from cellular reprogramming methods offer the opportunity to develop better models to study *Toxoplasma* in human neurons.⁹⁶ These studies may point the way to novel methods for the prevention and treatment of schizophrenia and other devastating psychiatric disorders.

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Conflict of Interest

The authors have declared that there are no conflicts of interest in relation to the subject of this study.

References

1. Leboyer M, Berk M, Yolken RH, Tamouza R, Kupfer D, Groc L. Immuno-psychiatry: an agenda for clinical practice and innovative research. *BMC Med.* 2016;14:173.
2. Dickerson F, Severance E, Yolken R. The microbiome, immunity, and schizophrenia and bipolar disorder. *Brain Behav Immun.* 2017;62:46–52.
3. Block ML, Zecca L, Hong JS. Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nat Rev Neurosci.* 2007;8:57–69.
4. Gendelman HE. Neural immunity: friend or foe? *J Neurovirol.* 2002;8:474–479.
5. Fineberg AM, Ellman LM, Buka S, Yolken R, Cannon TD. Decreased birth weight in psychosis: influence of prenatal exposure to serologically determined influenza and hypoxia. *Schizophr Bull.* 2013;39:1037–1044.
6. Severance EG, Gressitt KL, Buka SL, Cannon TD, Yolken RH. Maternal complement C1q and increased odds for psychosis in adult offspring. *Schizophr Res.* 2014;159:14–19.
7. Volk DW, Chitrapu A, Edelson JR, Roman KM, Moroco AE, Lewis DA. Molecular mechanisms and timing of cortical immune activation in schizophrenia. *Am J Psychiatry.* 2015;172:1112–1121.
8. Torrey EF, Miller J, Rawlings R, Yolken RH. Seasonality of births in schizophrenia and bipolar disorder: a review of the literature. *Schizophr Res.* 1997;28:1–38.
9. Simeone JC, Ward AJ, Rotella P, Collins J, Windisch R. An evaluation of variation in published estimates of schizophrenia prevalence from 1990 horizontal line 2013: a systematic literature review. *BMC Psychiatry* 2015;15:193.
10. Sørensen HJ, Nielsen PR, Pedersen CB, Benros ME, Nordentoft M, Mortensen PB. Population impact of familial and environmental risk factors for schizophrenia: a nationwide study. *Schizophr Res.* 2014;153:214–219.
11. Dubey JP. The history of *Toxoplasma gondii*—the first 100 years. *J Eukaryot Microbiol.* 2008;55:467–475.
12. Sibley LD, Khan A, Ajioka JW, Rosenthal BM. Genetic diversity of *Toxoplasma gondii* in animals and humans. *Philos Trans R Soc Lond B Biol Sci.* 2009;364:2749–2761.
13. Torrey EF, Bartko JJ, Yolken RH. *Toxoplasma gondii* and other risk factors for schizophrenia: an update. *Schizophr Bull.* 2012;38:642–647.
14. Yolken R, Torrey EF, Dickerson F. Evidence of increased exposure to *Toxoplasma gondii* in individuals with recent onset psychosis but not with established schizophrenia. *PLoS Negl Trop Dis.* 2017;11:e0006040.
15. Jones-Brando L, Torrey EF, Yolken R. Drugs used in the treatment of schizophrenia and bipolar disorder inhibit the replication of *Toxoplasma gondii*. *Schizophr Res.* 2003;62:237–244.
16. Vlatkovic S, Sagud M, Svob Strac D, et al. Increased prevalence of *Toxoplasma gondii* seropositivity in patients with treatment-resistant schizophrenia. *Schizophr Res.* 2018;193:480–481.
17. Lindgren M, Torniaainen-Holm M, Härkänen T, Dickerson F, Yolken RH, Suvisaari J. The association between toxoplasma and the psychosis continuum in a general population setting. *Schizophr Res.* 2018;193:329–335.
18. Dickerson F, Stallings C, Origoni A, et al. Antibodies to *Toxoplasma gondii* in individuals with mania. *Bipolar Disord.* 2014;16:129–136.
19. Hamdani N, Daban-Huard C, Lajnef M, et al. Relationship between *Toxoplasma gondii* infection and bipolar disorder in a French sample. *J Affect Disord.* 2013;148:444–448.
20. Pedersen MG, Mortensen PB, Norgaard-Pedersen B, Postolache TT. *Toxoplasma gondii* infection and self-directed violence in mothers. *Arch Gen Psychiatry.* 2012;69:1123–1130.
21. Okusaga O, Langenberg P, Sleemi A, et al. *Toxoplasma gondii* antibody titers and history of suicide attempts in patients with schizophrenia. *Schizophr Res.* 2011;133:150–155.
22. Markovitz AA, Simanek AM, Yolken RH, et al. *Toxoplasma gondii* and anxiety disorders in a community-based sample. *Brain Behav Immun.* 2015;43:192–197.
23. Alvarado-Esquivel C, Sanchez-Anguiano LF, Hernandez-Tinoco J, et al. *Toxoplasma gondii* infection and mixed anxiety and depressive disorder: a case-control seroprevalence study in Durango, Mexico. *J Clin Med Res.* 2016;8:519–523.
24. Flegr J, Horáček J. Toxoplasma-infected subjects report an obsessive-compulsive disorder diagnosis more often and score higher in obsessive-compulsive inventory. *Eur Psychiatry.* 2017;40:82–87.
25. Flegr J, Horacek J. Toxoplasmosis, but not borreliosis, is associated with psychiatric disorders and symptoms. *Schizophr Res* 2018.
26. Groer MW, Yolken RH, Xiao JC, et al. Prenatal depression and anxiety in *Toxoplasma gondii*-positive women. *Am J Obstet Gynecol* 2011;204:433 e431–437.
27. Gale SD, Berrett AN, Brown B, Erickson LD, Hedges DW. No association between current depression and latent toxoplasmosis in adults. *Folia Parasitol (Praha)* 2016;63:32–37.
28. Beste C, Getzmann S, Gajewski PD, Golka K, Falkenstein M. Latent *Toxoplasma gondii* infection leads to deficits in goal-directed behavior in healthy elderly. *Neurobiol Aging.* 2014;35:1037–1044.
29. Nimgaonkar VL, Yolken RH, Wang T, et al. Temporal cognitive decline associated with exposure to infectious agents in a population-based, aging cohort. *Alzheimer Dis Assoc Disord.* 2016;30:216–222.
30. Mendy A, Vieira ER, Albatineh AN, Gasana J. *Toxoplasma gondii* seropositivity and cognitive functions in school-aged children. *Parasitology.* 2015;142:1221–1227.
31. Alvarado-Esquivel C, Sánchez-Anguiano LF, Arnaud-Gil CA, et al. *Toxoplasma gondii* infection and suicide attempts: a case-control study in psychiatric outpatients. *J Nerv Ment Dis.* 2013;201:948–952.
32. Hinze-Selch D, Däubener W, Erdag S, Wilms S. The diagnosis of a personality disorder increases the likelihood for seropositivity to *Toxoplasma gondii* in psychiatric patients. *Folia Parasitol (Praha).* 2010;57:129–135.
33. Flegr J, Escudero DQ. Impaired health status and increased incidence of diseases in *Toxoplasma*-seropositive subjects: an explorative cross-sectional study. *Parasitology.* 2016;143:1974–1989.
34. Su C, Khan A, Zhou P, et al. Globally diverse *Toxoplasma gondii* isolates comprise six major clades originating from a

- small number of distinct ancestral lineages. *Proc Natl Acad Sci U S A*. 2012;109:5844–5849.
35. Xiao J, Jones-Brando L, Talbot CC Jr, Yolken RH. Differential effects of three canonical *Toxoplasma* strains on gene expression in human neuroepithelial cells. *Infect Immun*. 2011;79:1363–1373.
 36. Saeij JP, Boyle JP, Boothroyd JC. Differences among the three major strains of *Toxoplasma gondii* and their specific interactions with the infected host. *Trends Parasitol*. 2005;21:476–481.
 37. Voller A, Bidwell DE, Bartlett A, Fleck DG, Perkins M, Oladehin B. A microplate enzyme-immunoassay for toxoplasma antibody. *J Clin Pathol*. 1976;29:150–153.
 38. Jones JL, Kruszon-Moran D, Rivera HN, Price C, Wilkins PP. *Toxoplasma gondii* seroprevalence in the United States 2009–2010 and comparison with the past two decades. *Am J Trop Med Hyg* 2014;90:1135–1139.
 39. Nogareda F, Le Strat Y, Villena I, De Valk H, Goulet V. Incidence and prevalence of *Toxoplasma gondii* infection in women in France, 1980–2020: model-based estimation. *Epidemiol Infect* 2014;142:1661–1670.
 40. Xiao J, Li Y, Prandovszky E, et al. Behavioral abnormalities in a mouse model of chronic toxoplasmosis are associated with MAG1 antibody levels and cyst burden. *PLoS Negl Trop Dis*. 2016;10:e0004674.
 41. Xiao J, Viscidi RP, Kannan G, et al. The *Toxoplasma* MAG1 peptides induce sex-based humoral immune response in mice and distinguish active from chronic human infection. *Microbes Infect*. 2013;15:74–83.
 42. Ferguson DJ, Parmley SF. *Toxoplasma gondii* MAG1 protein expression. *Trends Parasitol*. 2002;18:482.
 43. Kong JT, Grigg ME, Uyetake L, Parmley S, Boothroyd JC. Serotyping of *Toxoplasma gondii* infections in humans using synthetic peptides. *J Infect Dis*. 2003;187:1484–1495.
 44. Xiao J, Buka SL, Cannon TD, et al. Serological pattern consistent with infection with type I *Toxoplasma gondii* in mothers and risk of psychosis among adult offspring. *Microbes Infect*. 2009;11:1011–1018.
 45. Xiao J, Yolken RH. Strain hypothesis of *Toxoplasma gondii* infection on the outcome of human diseases. *Acta Physiol (Oxf)*. 2015;213:828–845.
 46. Boyer K, Hill D, Mui E, et al.; Toxoplasmosis Study Group. Unrecognized ingestion of *Toxoplasma gondii* oocysts leads to congenital toxoplasmosis and causes epidemics in North America. *Clin Infect Dis*. 2011;53:1081–1089.
 47. Evans AK, Strassmann PS, Lee IP, Sapolsky RM. Patterns of *Toxoplasma gondii* cyst distribution in the forebrain associate with individual variation in predator odor avoidance and anxiety-related behavior in male Long-Evans rats. *Brain Behav Immun*. 2014;37:122–133.
 48. Afonso C, Paixão VB, Costa RM. Chronic *Toxoplasma* infection modifies the structure and the risk of host behavior. *PLoS One*. 2012;7:e32489.
 49. Luft BJ, Remington JS. Toxoplasmic encephalitis in AIDS. *Clin Infect Dis*. 1992;15:211–222.
 50. Berenreiterová M, Flegr J, Kuběna AA, Němec P. The distribution of *Toxoplasma gondii* cysts in the brain of a mouse with latent toxoplasmosis: implications for the behavioral manipulation hypothesis. *PLoS One*. 2011;6:e28925.
 51. Melzer TC, Cranston HJ, Weiss LM, Halonen SK. Host cell preference of *Toxoplasma gondii* cysts in murine brain: a confocal study. *J Neuroparasitology* 2010;1.
 52. Ferguson DJ, Hutchison WM. An ultrastructural study of the early development and tissue cyst formation of *Toxoplasma gondii* in the brains of mice. *Parasitol Res*. 1987;73:483–491.
 53. Haroon F, Händel U, Angenstein F, et al. *Toxoplasma gondii* actively inhibits neuronal function in chronically infected mice. *PLoS One*. 2012;7:e35516.
 54. Watts E, Zhao Y, Dhara A, Eller B, Patwardhan A, Sinai AP. Novel approaches reveal that *Toxoplasma gondii* bradyzoites within tissue cysts are dynamic and replicating entities in vivo. *MBio*. 2015;6:e011155–e01115.
 55. Xiao J, Li Y, Gressitt KL, et al. Cerebral complement C1q activation in chronic *Toxoplasma* infection. *Brain Behav Immun*. 2016;58:52–56.
 56. Kannan G, Crawford JA, Yang C, et al. Anti-NMDA receptor autoantibodies and associated neurobehavioral pathology in mice are dependent on age of first exposure to *Toxoplasma gondii*. *Neurobiol Dis*. 2016;91:307–314.
 57. Hermes G, Ajioka JW, Kelly KA, et al. Neurological and behavioral abnormalities, ventricular dilatation, altered cellular functions, inflammation, and neuronal injury in brains of mice due to common, persistent, parasitic infection. *J Neuroinflammation*. 2008;5:48.
 58. Parlog A, Harsan LA, Zagrebelsky M, et al. Chronic murine toxoplasmosis is defined by subtle changes in neuronal connectivity. *Dis Model Mech*. 2014;7:459–469.
 59. David CN, Frias ES, Szu JI, et al. GLT-1-dependent disruption of CNS glutamate homeostasis and neuronal function by the protozoan parasite *Toxoplasma gondii*. *PLoS Pathog*. 2016;12:e1005643.
 60. Horacek J, Flegr J, Tintera J, et al. Latent toxoplasmosis reduces gray matter density in schizophrenia but not in controls: voxel-based-morphometry (VBM) study. *World J Biol Psychiatry*. 2012;13:501–509.
 61. Brooks JM, Carrillo GL, Su J, Lindsay DS, Fox MA, Blader IJ. *Toxoplasma gondii* infections alter GABAergic synapses and signaling in the central nervous system. *MBio*. 2015;6:e01428–e01415.
 62. Pfefferkorn ER. Interferon gamma blocks the growth of *Toxoplasma gondii* in human fibroblasts by inducing the host cells to degrade tryptophan. *Proc Natl Acad Sci U S A*. 1984;81:908–912.
 63. Schwarcz R, Hunter CA. *Toxoplasma gondii* and schizophrenia: linkage through astrocyte-derived kynurenic acid? *Schizophr Bull*. 2007;33:652–653.
 64. Kannan G, Gressitt KL, Yang S, et al. Pathogen-mediated NMDA receptor autoimmunity and cellular barrier dysfunction in schizophrenia. *Transl Psychiatry*. 2017;7:e1186.
 65. Hughes EG, Peng X, Gleichman AJ, et al. Cellular and synaptic mechanisms of anti-NMDA receptor encephalitis. *J Neurosci*. 2010;30:5866–5875.
 66. Gaskell EA, Smith JE, Pinney JW, Westhead DR, McConkey GA. A unique dual activity amino acid hydroxylase in *Toxoplasma gondii*. *PLoS One*. 2009;4:e4801.
 67. Tedford E, McConkey G. Neurophysiological changes induced by chronic *Toxoplasma gondii* infection. *Pathogens* 2017;6.
 68. Prandovszky E, Gaskell E, Martin H, Dubey JP, Webster JP, McConkey GA. The neurotropic parasite *Toxoplasma gondii* increases dopamine metabolism. *PLoS One*. 2011;6:e23866.
 69. Flegr J. Schizophrenia and *Toxoplasma gondii*: an undervalued association? *Expert Rev Anti Infect Ther*. 2015;13:817–820.
 70. Wang ZT, Harmon S, O'Malley KL, Sibley LD. Reassessment of the role of aromatic amino acid hydroxylases and the effect of infection by *Toxoplasma gondii* on host dopamine. *Infect Immun*. 2015;83:1039–1047.
 71. Afonso C, Paixão VB, Klaus A, et al. Toxoplasma-induced changes in host risk behaviour are independent

- of parasite-derived AaaH2 tyrosine hydroxylase. *Sci Rep.* 2017;7:13822.
72. McFarland R, Wang ZT, Jouroukhin Y, et al. AAH2 gene is not required for dopamine-dependent neurochemical and behavioral abnormalities produced by *Toxoplasma* infection in mouse. *Behav Brain Res.* 2018;347:193–200.
 73. Wang ZT, Verma SK, Dubey JP, Sibley LD. The aromatic amino acid hydroxylase genes AAH1 and AAH2 in *Toxoplasma gondii* contribute to transmission in the cat. *PLoS Pathog.* 2017;13:e1006272.
 74. Wujcicka W, Gaj Z, Wilczyński J, Nowakowska D. Possible role of TLR4 and TLR9 SNPs in protection against congenital toxoplasmosis. *Eur J Clin Microbiol Infect Dis.* 2015;34:2121–2129.
 75. Shimokawa PT, Targa LS, Yamamoto L, Rodrigues JC, Kanunfre KA, Okay TS. HLA-DQA1/B1 alleles as putative susceptibility markers in congenital toxoplasmosis. *Virulence.* 2016;7:456–464.
 76. Avramopoulos D, Pearce BD, McGrath J, et al. Infection and inflammation in schizophrenia and bipolar disorder: a genome wide study for interactions with genetic variation. *PLoS One.* 2015;10:e0116696.
 77. Bhadra R, Gigley JP, Weiss LM, Khan IA. Control of *Toxoplasma* reactivation by rescue of dysfunctional CD8+ T-cell response via PD-1-PDL-1 blockade. *Proc Natl Acad Sci U S A.* 2011;108:9196–9201.
 78. Steiner J, Jacobs R, Panteli B, et al. Acute schizophrenia is accompanied by reduced T cell and increased B cell immunity. *Eur Arch Psychiatry Clin Neurosci.* 2010;260:509–518.
 79. Craddock RM, Lockstone HE, Rider DA, et al. Altered T-cell function in schizophrenia: a cellular model to investigate molecular disease mechanisms. *PLoS One.* 2007;2:e692.
 80. Riedel M, Spellmann I, Schwarz MJ, et al. Decreased T cellular immune response in schizophrenic patients. *J Psychiatr Res.* 2007;41:3–7.
 81. Bhadra R, Cobb DA, Weiss LM, Khan IA. Psychiatric disorders in toxoplasma seropositive patients—the CD8 connection. *Schizophr Bull.* 2013;39:485–489.
 82. Buervenich S, Carmine A, Arvidsson M, et al. NURR1 mutations in cases of schizophrenia and manic-depressive disorder. *Am J Med Genet.* 2000;96:808–813.
 83. Chen YH, Tsai MT, Shaw CK, Chen CH. Mutation analysis of the human NR4A2 gene, an essential gene for midbrain dopaminergic neurogenesis, in schizophrenic patients. *Am J Med Genet.* 2001;105:753–757.
 84. Ancín I, Cabranes JA, Vázquez-Álvarez B, et al. NR4A2: effects of an “orphan” receptor on sustained attention in a schizophrenic population. *Schizophr Bull.* 2013;39:555–563.
 85. Eells JB, Varela-Stokes A, Guo-Ross SX, et al. Chronic *Toxoplasma gondii* in Nurr1-null heterozygous mice exacerbates elevated open field activity. *PLoS One.* 2015;10:e0119280.
 86. Severance EG, Xiao J, Jones-Brando L, et al. *Toxoplasma gondii*-a gastrointestinal pathogen associated with human brain diseases. *Int Rev Neurobiol.* 2016;131:143–163.
 87. Dubey JP. Bradyzoite-induced murine toxoplasmosis: stage conversion, pathogenesis, and tissue cyst formation in mice fed bradyzoites of different strains of *Toxoplasma gondii*. *J Eukaryot Microbiol.* 1997;44:592–602.
 88. Severance EG, Prandovszky E, Castiglione J, Yolken RH. Gastroenterology issues in schizophrenia: why the gut matters. *Curr Psychiatry Rep.* 2015;17:27.
 89. Severance EG, Alaedini A, Yang S, et al. Gastrointestinal inflammation and associated immune activation in schizophrenia. *Schizophr Res.* 2012;138:48–53.
 90. Schwarz E, Maukonen J, Hyttiäinen T, et al. Analysis of microbiota in first episode psychosis identifies preliminary associations with symptom severity and treatment response. *Schizophr Res.* 2018;192:398–403.
 91. von Klitzing E, Ekmekci I, Kühl AA, Bereswill S, Heimesaat MM. Intestinal, extra-intestinal and systemic sequelae of *Toxoplasma gondii* induced acute ileitis in mice harboring a human gut microbiota. *PLoS One.* 2017;12:e0176144.
 92. Chorlton SD. *Toxoplasma gondii* and schizophrenia: a review of published RCTs. *Parasitol Res.* 2017;116:1793–1799.
 93. Xiao J, Li Y, Yolken RH, Viscidi RP. PD-1 immune checkpoint blockade promotes brain leukocyte infiltration and diminishes cyst burden in a mouse model of *Toxoplasma* infection. *J Neuroimmunol.* 2018;319:55–62.
 94. Flegr J. How and why *Toxoplasma* makes us crazy. *Trends Parasitol.* 2013;29:156–163.
 95. Stock AK, Dajkic D, Köhling HL, von Heinegg EH, Fiedler M, Beste C. Humans with latent toxoplasmosis display altered reward modulation of cognitive control. *Sci Rep.* 2017;7:10170.
 96. Halonen SK. Use of human neurons derived via cellular reprogramming methods to study host-parasite interactions of *Toxoplasma gondii* in neurons. *Cells* 2017;6:32–45.