(Micro)Glia as Effectors of Cortical Volume Loss in Schizophrenia

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Contrary to the notion that neurology but not psychiatry is the domain of disorders evincing structural brain alterations, it is now clear that there are subtle but consistent neuropathological changes in schizophrenia. These range from increases in ventricular size to dystrophic changes in dendritic spines. A decrease in dendritic spine density in the prefrontal cortex (PFC) is among the most replicated of postmortem structural findings in schizophrenia. Examination of the mechanisms that account for the loss of dendritic spines has in large part focused on genes and molecules that regulate neuronal structure. But the simple question of what is the effector of spine loss, ie, where do the lost spines go, is unanswered. Recent data on glial cells suggest that microglia (MG), and perhaps astrocytes, play an important physiological role in synaptic remodeling of neurons during development. Synapses are added to the dendrites of pyramidal cells during the maturation of these neurons; excess synapses are subsequently phagocytosed by MG. In the PFC, this occurs during adolescence, when certain symptoms of schizophrenia emerge. This brief review discusses recent advances in our understanding of MG function and how these non-neuronal cells lead to structural changes in neurons in schizophrenia.

Key words: dendritic spine/inflammation/microglia/neur opathology/prefrontal cortex/pyramidal cell

Attempts to define structural brain alterations in schizophrenia during much of the 20th century failed to reveal consistent neuropathological changes. The state of affairs was so bad that the neurologist Plum¹ famously referred to the field as "the graveyard of neuropathologists," with Harrison² subsequently commenting that the field was noteworthy for "generating more heat than light and … memorable quotes rather than durable data." Fortunately, the last quarter of the 20th century saw the application of quantitative neuroanatomical methods to neuropathological studies and the advent of contemporary in vivo imaging methods. These advances allowed researchers to detect subtle, but consistent, anatomical changes in the brain in schizophrenia, and led to the claim that "no longer can there be doubt that there is underlying brain pathology,"³ fulfilling the view of Kraepelin that schizophrenia is a brain disorder. Although some suggest that this view may be a bit optimistic,⁵ meta-analyses of volumetric as well as longitudinal studies point to structural changes in schizophrenia (however, see also Heilbronner et al,⁶ Vita et al,⁷ Kambeitz et al,⁸ and Olabi et al.⁹).

Neuropathology of Schizophrenia

Neuron Pathology

A key finding by Eve Johnstone et al,¹⁰ using computed tomography, was ventricular enlargement in schizophrenia. Although intially thought to reflect disease progression,¹¹ subsequent studies noted ventricular enlargement in first-episode patients.^{12,13}

If the ventricles are enlarging, yet the brain is encased in an unyielding skull, what "gives"? Imaging studies have consistently revealed a decrease in gray matter volume in schizophrenia.^{12,14} These findings are corroborated by postmortem studies noting reduced cortical thickness.¹⁵ Although these changes are not seen in each subject, group differences in ventricular enlargement, gray matter volume, and cortical thickness, particularly in the prefrontal and medial temporal cortices, are consistently observed in schizophrenia, including in studies of firstepisode and antipsychotic drug (APD)-naïve patients.¹⁶⁻¹⁹ Such changes have even been reported in subjects deemed at high risk for developing the illness, although only a minority of high-risk patients subsequently develop schizophrenia,²⁰ leaving open the possibility that this may not be specific to schizophrenia.

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The loss of cortical volume and thickness suggests that there may be a loss of cortical neurons in schizophrenia. However, unbiased counts of total neocortical neuron number²¹ and the number of neurons in the prefrontal cortex (PFC)²² have uncovered no such difference. Instead, neuronal density is increased,^{23–25} leading Selemon and Goldman-Rakic²⁶ to propose the reduced neuropil hypothesis of schizophrenia. Their formulation posits that a decrease in cortical volume in the face of a normal complement of neurons occurs secondary to a decrease in neuropil, including dendrites and axons.

Relatively early studies of cortical gene expression were consistent with this hypothesis, reporting a loss of dendrite- and axon-associated genes.²⁷ Although anatomical and immunoblot studies of axonal markers in schizophrenia led to conflicting results.²⁷⁻³⁷ studies of dendrites consistently revealed dystrophic changes.³⁸⁻⁴² In particular, there is a decrease in the density of dendritic spines on PFC pyramidal cells (PCs) in schizophrenia,^{38,42-45} but not in samples from a psychiatric control group⁴⁴ (primarily mood disorders subjects treated with APDs). Some studies of changes in spine density revealed selective effects on deep layer 3 (L3) PFC PCs, consistent with a decrease in soma size of L3 PCs. 30,46,47 Prefrontal cortical PCs appear to be most vulnerable to spine loss; a less pronounced decrease in spine density has been reported in the primary auditory^{48,49} with no significant change in the visual cortex.⁴⁴

Because dendritic spines are the primary site of excitatory inputs to the PC, the loss of spines on PCs may lead to significant disruptions in excitatory signaling to corticofugal pathways. Unfortunately, there have been very few studies probing the correlation of dendritic spine density changes and cognitive performance (see Kim et al,⁵⁰ Cahill et al,⁵¹ and Hains et al⁵²), with none focusing on different times during development. Moreover, although it has been suggested that cognitive deficits are already present in first-episode schizophrenia,⁵³ this is an oversimplification, with deficits in performance of certain cognitive tasks (such as working memory) differing from those in processing speed. In addition, most studies of cognition in schizophrenia do not determine the degree to which such changes may be secondary to negative symptoms or other domains.⁵⁴ At this time, the functional impact of dendritic spine changes on specific symptom domains is unknown.

However, by comparing the shape of lost (vulnerable) and remaining dendritic spines, one may glean limited insight into function. Morphological parameters have long been used to categorize spines into different classes, including spines that hyperacute anatomists have fancifully described as thin-, stubby-, and mushroom-shaped.^{55,56} These adjectives are of limited utility: various parameters of spine shape (such as spine head diameter, which should be larger in mushroom than thin spines) show considerable overlap across different types of spines.⁵⁷ Nonetheless, thin spines, which are relatively long and lack a wide head, have been advanced as being immature and more likely to lack α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid AMPA receptors, ie, have functionally silent synapses^{58,59} (however, see also Busetto et al⁶⁰), whereas larger, mushroomshaped spines are thought to be mature.⁶¹ Only very recently has there been an assessment of the types of spines present in schizophrenia.⁶² This study reported that thin spines were preferentially lost in the auditory cortex, which was interpreted to suggest that there is a deficit in newly formed spines in the cortex. We await confirmation of these recent data, including in the PFC.

If dendritic spines are decreased in number, there may be corresponding decreases in the presynaptic partners of lost spines. However, postmortem studies of changes in presynaptic elements in schizophrenia, including proteins involved in vesicular trafficking and release, have yielded conflicting results.^{27–37} Several factors may contribute to the inconsistent results, including different dependent variables (mRNA vs protein levels), APD treatment, and differences in the areas and layers of the cortex being sampled. Still another reason for the conflicting data may be that most studies examining presynaptic changes have analyzed markers of synaptic release common to all neurons, thereby capturing both inhibitory and excitatory presynaptic elements. Because presynaptic axons forming synapses with spines are excitatory, more consistent results emerge when excitatory inputs are analyzed separately: there is a decrease in cortical levels of the glutamatergic marker vesicular glutamate transporter (VGluT) 1 (but not VGluT2, another glutamate transporter).^{63–65} Because VGluT1 and VGluT2 are mainly expressed by cortical and subcortical glutamatergic neurons, respectively,^{66–68} synapses formed by different afferent sources defining different circuits with distinct PCs may be compromised in schizophrenia.

Glial Pathology

One index of a shift in the targets of scientific inquiry in brain disorders over the past decade has been the introduction of neologisms such as gliotransmission. The scientific blinders that limited attention to neurons have been removed, leading to a broad interest in non-neuronal as well as neuronal elements of the nervous system. In particular, there is today a much greater appreciation of the diverse roles played by glia.

In 1982, Stevens⁶⁹ reported that reactive astrocytosis (an increase in astrocytes that occurs in response to cellular damage) was present in the thalamus, limbic areas, and periventricular sites in about ~70% of patients with schizophrenia; these observations were consistent with some earlier reports of gliosis in diencephalic and mesencephalic regions.^{70,71} However, these early studies of reactive astrocytosis did not use unbiased methods,

now *de rigueur*,^{72,73} to determine cell number or density, and lacked many methodological details and some control procedures, clouding interpretation of the results. Coupled with a lack of assessment of potential confounding variables,⁷⁴ substantial differences in the conclusions of early and more contemporary studies of glial changes in schizophrenia are not surprising.

Astrocytes. Roberts et al⁷⁵ examined 20 different brain areas for evidence of astrogliosis in the brains of schizophrenic subjects. They found no difference in the numbers of cells expressing the astrocytic marker glial fibrillary acidic protein (GFAP).^{76,77} Roberts et al⁷⁸ subsequently replicated their initial finding in a larger cohort, and most subsequent studies of schizophrenia also failed to detect an increase in the number or density of astrocytes.^{75,78–96} Studies of GFAP mRNA and protein levels largely corroborated the anatomical data.^{36,95,97–102}

Thus, available postmortem data suggest that there are probably no substantial changes in the number or density of astrocytes in the cortex in schizophrenia.¹⁰³ Future studies utilizing a different marker of astrocytes, aldehyde dehydrogenase 1 family, member L1 (Aldh1L1), which in contrast to GFAP appears to be an invariant marker of astrocytes¹⁰⁴ may reveal subtle, region-specific changes in astrocytes.

Microglia. There has long been considerable interest in the role of inflammation in promoting neuropathological changes in schizophrenia.^{103,105,106} Interest has piqued over the past decade with genetic studies revealing associations of schizophrenia with the major histocompatibility locus,¹⁰⁷ and more recently with specific variations in complement component 4 being strongly linked to the risk for developing schizophrenia.¹⁰⁸

Because microglia (MG) are the immune cells of the brain, potential changes in the number, density, and function of MG have been scrutinized. It should not be surprising to learn that this area of research is also littered with inconsistent results. Studies using various markers to label MG have resulted in reports of increased density of MG,^{85,109–111} increased MG activation,^{109,112} and degenerating MG cells.^{109,113} In contrast, other studies have found no change in these parameters.^{82,84,93,114–116} A recent metaanalysis of studies examining MG density in postmortem tissue concluded that the preponderance of evidence is consistent with a significant increase in MG density and a corresponding upregulation of MG-related proinflammatory genes in schizophrenia.¹¹⁷

Studies of MG in schizophrenia have in part been confounded by issues common to all postmortem studies, ranging from the use of APDs or other drugs to agonal state. However, there is another concern specific to MG: although MG occupy a restricted central nervous system (CNS) niche, virtually all MG markers are also present in (peripheral) monocytes and macrophages. The recent identification of transmembrane protein 119 (Tmem119)^{118,119} and potentially sialic acid–binding immunoglobulin-like lectin H (Siglec-H)¹²⁰ as selective markers of central MG but not peripherally derived cells should open the door for more accurate studies of MG number and density in schizophrenia.

An indirect approach to identifying changes in MG and inflammatory processes in schizophrenia has been through the development of radioligands for positron emission tomography studies of MG. Radioligands for the 18 kDa translocator protein (TSPO), a protein thought to be involved in steroidogenesis,^{121,122} have been proposed to be useful in monitoring inflammatory processes and MG activation in various disorders,^{123,124} including schizophrenia. Expression of TSPO is upregulated in inflammatory states and diseases¹²⁵⁻¹²⁸ and during MG activation.^{124,129} Early imaging studies with TSPO tracers generated conflicting results because the contribution of allelic variants in TSPO binding was not appreciated.¹³⁰ However, subsequent studies, conducted at various stages of the illness, were also inconsistent.¹³¹⁻¹⁴⁰ It has become clear that TSPO is not a specific marker of MG: the protein is also expressed in peripheral (and CNS-infiltrating) macrophages and monocytes, and has been reported to bind to astrocytes, endothelial cells, and perhaps even neurons.¹⁴¹⁻¹⁴⁴ Moreover, TSPO expression is increased substantially in response to a proinflammatory challenge in rodent, but not in human, MG.145 These considerations and others have cast doubt on the utility of TSPO as a marker of MG activation and inflammation.^{130,143,146–150}

Where Do the Lost Dendritic Spines Go in Schizophrenia?

During brain development, the number of synapses is not constant. In early postnatal development, synapses on neurons are formed in excess.¹⁵¹ Some of these supernumerary synapses are subsequently removed (pruned), while others are strengthened,¹⁵² optimizing the signal-tonoise ratio. The age at which mature neuron structure is achieved varies across brain regions. The PFC is the last area to mature, finally stabilizing in the third decade of life.¹⁵³

Peak density of PFC synapses^{153,154} occurs during adolescence, the period during which certain symptoms of schizophrenia typically emerge, leading Feinberg¹⁵⁵ to propose that schizophrenia may result from a defect in synaptic elimination programmed to occur during adolescence. This neurodevelopmental hypothesis was followed by several others,¹⁵⁶ which posit that the consequences of an insult during the second or third trimester of pregnancy lie dormant until manifest in adolescence.

Efforts to understand the process of synapse removal during development have revealed a critical role for MG, the innate immune cells of the CNS. Microglia are CNS macrophages derived from yolk-sac progenitors that migrate to the neural tube early in embryonic development,¹⁵⁷ and which locally renew by self-proliferation.¹⁵⁸ They are highly dynamic cells, extending and retracting their processes to surveil brain parenchyma for signs of insult or injury.^{159,160} Microglia also play key roles in the healthy brain (see Tremblay et al,¹⁶¹ Hong et al,¹⁶² and Kierdorf and Prinz¹⁶³), including in the modification of synaptic architecture in an experience-dependent manner, elimination of apoptotic neurons, and the formation of dendritic spines.^{164–166} Under physiological conditions, MG engulf excess synapses early in development in subcortical areas^{167,168} through complement-mediated pathways.^{168,169}

In the rat, MG transiently engulf dendritic spines on PFC PCs at postnatal day 39,¹⁷⁰ an age corresponding to late adolescence in humans (see Mallya et al¹⁷⁰). Presynaptic glutamatergic terminals synapsing with spines are also pruned by MG, but slightly later than spines, consistent with spine outgrowth preceding synapse formation.^{171–173} These data agree with Feinberg's¹⁵⁵ hypothesis, suggesting a deranged enhancement of physiological synapse pruning by MG during adolescence culminates in a reduced number of PFC PC dendritic spines.

The role of developmental synaptic pruning is not limited to MG. Astrocytes have been shown to participate in synapse elimination,¹⁷⁴ both directly (via recognition of an unidentified "eat-me" signal on a synapse destined for elimination through phagocytic pathways¹⁷⁵) and indirectly (in which release of transforming growth factor β regulates the expression of complement component Clq at synapses, recruiting MG to the site¹⁷⁶). Although the overall number of reactive astrocytes may not be increased in schizophrenia, there may be changes in one type of reactive astrocyte^{177,178} (the "A1" astrocyte, which are induced by activated MG and thought to be neurotoxic [see Liddelow et al¹⁷⁷ and Liddelow and Barres¹⁷⁸]).

Future Studies of Microglial Involvement in Schizophrenia

Microglia have historically been thought to adopt different morphologies based on their functional state. Surveilling MG are extensively branched and have a smaller somata than activated MG, which assume a large ball-like shape with few or no processes (see Hanisch and Kettenmann¹⁷⁹ and Ransohoff and Perry¹⁸⁰). In contrast to this traditional view, recent data indicate that during pruning of synapses MG do not assume a classic "activated" morphology, instead displaying many branched processes.^{168,170,181} These findings suggest that there are at least two different types of activated MG: those triggered in response to a pathological challenge, and those activated to engage in physiological neuronal sculpting. Furthermore, MG phenotype is governed by unidentified local cues, adding to the complex heterogeneity of MG.¹⁸² Disentangling the processes and signals that dictate the functional state of MG during developmental phagocytosis as opposed to those mediating inflammation- and pathology-based phagocytosis will be critical for future understanding of MG function in health and disease.

An issue not often addressed but critical to our understanding of MG is how to assess the relative contribution of MG that are intrinsic to the CNS from infiltrating macrophages generated in the periphery. Studies of developmental pruning have used markers common to both peripheral or central cells, such as ionized calciumbinding adapter molecule 1 (Iba1) and the fractalkine receptor CX₂CR1. However, in the healthy CNS, MG are the resident macrophages, while peripheral macrophages are mainly restricted to perivascular spaces, meninges, and the choroid plexus.^{183,184} The extent to which the blood-brain or cerebrospinal fluid-brain barriers may be porous under conditions such as inflammation or in illnesses such as schizophrenia is not known. Similarly, it is not clear to what degree peripherally derived monocytes enter the CNS at circumventricular sites,¹⁸⁵ and from there migrate to other areas.

Translating Microglial Dysfunction to Therapeutic Strategies

The elucidation of the mechanisms whereby developmentally specific synaptic pruning occurs may lead to new therapeutic targets for mitigating structural and functional changes in schizophrenia. Neuronal elements destined for elimination have undefined (but in part possibly complement-related) "find-me" signals that target MG to the neuron and "eat-me" signals that then cue the MG to phagocytose a particular spine or axonal element. There are also "don't-find-me" and "don't-eat-me" signals that help a spine evade detection and pruning, similar to those seen in apoptotic cells.¹⁸⁶ Pharmacological or molecular suppression of the former or amplification of the latter, particularly during adolescence, when spine pruning is active, might diminish excess pruning of spines on PFC PCs, thereby averting some of the behavioral pathology of schizophrenia. However, it is likely that too much suppression will result in too many spines, as seen in autism spectrum disorder (ASD) and Fragile X syndrome.¹⁸⁷ Notably, in both ASD and schizophrenia, social cognition is impaired, suggesting that there may be an optimal spine number above or below which negative consequences occur.

Concluding Remarks

Psychiatry has moved from questioning whether schizophrenia is a brain disease to determining how structural changes in certain brain areas and circuits lead to symptoms. Attempts to understand the pathophysiology of schizophrenia have become more challenging with the realization that schizophrenia is not a disease of neurons, but also critically involves non-neuronal cells. However, this added complexity may reveal important new drug targets for the treatment—or even prevention—of schizophrenia. Our appreciation of the many physiological roles played by MG is rapidly growing and points to the need for new methods to allow one to demonstrate conclusively if MG are effectors of the "excess synaptic elimination programmed to occur during adolescence" first posited by Feinberg¹⁵⁵ 35 years ago.

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References

- 1. Plum F. Prospects for research on schizophrenia. 3. neurophysiology. neuropathological findings. *Neurosci Res Program Bull*. 1972;10:384–388.
- 2. Harrison PJ. The neuropathology of schizophrenia. A critical review of the data and their interpretation. *Brain*. 1999;122(pt 4):593–624.
- 3. Weinberger DR. From neuropathology to neurodevelopment. *Lancet*. 1995;346:552–557.
- Kraepelin E, Robertson GM, Barclay RM. *Dementia Praecox* and Paraphrenia. Huntington, New York: R. E. Krieger Pub. Co.; 1971.
- 5. Fusar-Poli P, Meyer-Lindenberg A. Forty years of structural imaging in psychosis: promises and truth. *Acta Psychiatr Scand*. 2016;134:207–224.
- 6. Heilbronner U, Samara M, Leucht S, Falkai P, Schulze TG. The longitudinal course of schizophrenia across the lifespan: clinical, cognitive, and neurobiological aspects. *Harv Rev Psychiatry*. 2016;24:118–128.
- 7. Vita A, De Peri L, Deste G, Barlati S, Sacchetti E. The effect of antipsychotic treatment on cortical gray matter changes in schizophrenia: does the class matter? a meta-analysis and meta-regression of longitudinal magnetic resonance imaging studies. *Biol Psychiatry*. 2015;78:403–412.
- Kambeitz J, Kambeitz-Ilankovic L, Leucht S, et al. Detecting neuroimaging biomarkers for schizophrenia: a meta-analysis of multivariate pattern recognition studies. *Neuropsychopharmacology*. 2015;40:1742–1751.
- Olabi B, Ellison-Wright I, McIntosh AM, Wood SJ, Bullmore E, Lawrie SM. Are there progressive brain changes in schizophrenia? A meta-analysis of structural magnetic resonance imaging studies. *Biol Psychiatry*. 2011;70:88–96.
- Johnstone EC, Crow TJ, Frith CD, Husband J, Kreel L. Cerebral ventricular size and cognitive impairment in chronic schizophrenia. *Lancet*. 1976;2:924–926.

- Weinberger DR. Implications of normal brain development for the pathogenesis of schizophrenia. *Arch Gen Psychiatry*. 1987;44:660–669.
- Shenton ME, Dickey CC, Frumin M, McCarley RW. A review of MRI findings in schizophrenia. *Schizophr Res.* 2001;49:1–52.
- Vita A, De Peri L, Silenzi C, Dieci M. Brain morphology in first-episode schizophrenia: a meta-analysis of quantitative magnetic resonance imaging studies. *Schizophr Res.* 2006;82:75–88.
- Lawrie SM, Abukmeil SS. Brain abnormality in schizophrenia. A systematic and quantitative review of volumetric magnetic resonance imaging studies. *Br J Psychiatry*. 1998;172:110–120.
- Goldman AL, Pezawas L, Mattay VS, et al. Widespread reductions of cortical thickness in schizophrenia and spectrum disorders and evidence of heritability. *Arch Gen Psychiatry*. 2009;66:467–477.
- Fusar-Poli P, Radua J, McGuire P, Borgwardt S. Neuroanatomical maps of psychosis onset: voxel-wise metaanalysis of antipsychotic-naive VBM studies. *Schizophr Bull*. 2012;38:1297–1307.
- 17. Borgwardt SJ, McGuire PK, Aston J, et al. Reductions in frontal, temporal and parietal volume associated with the onset of psychosis. *Schizophr Res.* 2008;106:108–114.
- Leung M, Cheung C, Yu K, et al. Gray matter in first-episode schizophrenia before and after antipsychotic drug treatment. Anatomical likelihood estimation meta-analyses with sample size weighting. *Schizophr Bull*. 2011;37:199–211.
- Song X, Quan M, Lv L, et al. Decreased cortical thickness in drug naïve first episode schizophrenia: in relation to serum levels of BDNF. J Psychiatr Res. 2015;60:22–28.
- Cannon TD, Cadenhead K, Cornblatt B, et al. Prediction of psychosis in youth at high clinical risk: a multisite longitudinal study in North America. *Arch Gen Psychiatry*. 2008;65:28–37.
- Pakkenberg B. Total nerve cell number in neocortex in chronic schizophrenics and controls estimated using optical disectors. *Biol Psychiatry*. 1993;34:768–772.
- 22. Thune JJ, Uylings HB, Pakkenberg B. No deficit in total number of neurons in the prefrontal cortex in schizophrenics. *J Psychiatr Res.* 2001;35:15–21.
- 23. Selemon LD, Rajkowska G, Goldman-Rakic PS. Elevated neuronal density in prefrontal area 46 in brains from schizophrenic patients: application of a three-dimensional, stereologic counting method. J Comp Neurol. 1998;392:402–412.
- 24. Selemon LD, Rajkowska G, Goldman-Rakic PS. Abnormally high neuronal density in the schizophrenic cortex. A morphometric analysis of prefrontal area 9 and occipital area 17. Arch Gen Psychiatry. 1995;52:805–18; discussion 819.
- 25. Selemon LD, Mrzljak J, Kleinman JE, Herman MM, Goldman-Rakic PS. Regional specificity in the neuropathologic substrates of schizophrenia: a morphometric analysis of Broca's area 44 and area 9. Arch Gen Psychiatry. 2003;60:69–77.
- 26. Selemon LD, Goldman-Rakic PS. The reduced neuropil hypothesis: a circuit based model of schizophrenia. *Biol Psychiatry*. 1999;45:17–25.
- 27. Mirnics K, Middleton FA, Marquez A, Lewis DA, Levitt P. Molecular characterization of schizophrenia viewed by microarray analysis of gene expression in prefrontal cortex. *Neuron*. 2000;28:53–67.
- Glantz LA, Lewis DA. Reduction of synaptophysin immunoreactivity in the prefrontal cortex of subjects with schizophrenia. Regional and diagnostic specificity. *Arch Gen Psychiatry*. 1997;54:660–669.

- Eastwood SL, Cairns NJ, Harrison PJ. Synaptophysin gene expression in schizophrenia. Investigation of synaptic pathology in the cerebral cortex. *Br J Psychiatry*. 2000;176:236–242.
- Glantz LA, Austin MC, Lewis DA. Normal cellular levels of synaptophysin mRNA expression in the prefrontal cortex of subjects with schizophrenia. *Biol Psychiatry*. 2000;48:389–397.
- Honer WG, Falkai P, Bayer TA, et al. Abnormalities of SNARE mechanism proteins in anterior frontal cortex in severe mental illness. *Cereb Cortex*. 2002;12:349–356.
- 32. Fung SJ, Sivagnanasundaram S, Weickert CS. Lack of change in markers of presynaptic terminal abundance alongside subtle reductions in markers of presynaptic terminal plasticity in prefrontal cortex of schizophrenia patients. *Biol Psychiatry*. 2011;69:71–79.
- Castillo MA, Ghose S, Tamminga CA, Ulery-Reynolds PG. Deficits in syntaxin 1 phosphorylation in schizophrenia prefrontal cortex. *Biol Psychiatry*. 2010;67:208–216.
- 34. Gil-Pisa I, Munarriz-Cuezva E, Ramos-Miguel A, Urigüen L, Meana JJ, García-Sevilla JA. Regulation of muncl8-1 and syntaxin-1A interactive partners in schizophrenia prefrontal cortex: down-regulation of muncl8-1a isoform and 75 kDa SNARE complex after antipsychotic treatment. *Int J Neuropsychopharmacol.* 2012;15:573–588.
- 35. Katrancha SM, Koleske AJ. SNARE complex dysfunction: a unifying hypothesis for schizophrenia. *Biol Psychiatry*. 2015;78:356–358.
- 36. Karson CN, Mrak RE, Schluterman KO, Sturner WQ, Sheng JG, Griffin WS. Alterations in synaptic proteins and their encoding mRNAs in prefrontal cortex in schizophrenia: a possible neurochemical basis for 'hypofrontality'. *Mol Psychiatry*. 1999;4:39–45.
- 37. Halim ND, Weickert CS, McClintock BW, et al. Presynaptic proteins in the prefrontal cortex of patients with schizophrenia and rats with abnormal prefrontal development. *Mol Psychiatry*. 2003;8:797–810.
- Glausier JR, Lewis DA. Dendritic spine pathology in schizophrenia. *Neuroscience*. 2013;251:90–107.
- Moyer CE, Shelton MA, Sweet RA. Dendritic spine alterations in schizophrenia. *Neurosci Lett.* 2015;601:46–53.
- 40. Kalus P, Müller TJ, Zuschratter W, Senitz D. The dendritic architecture of prefrontal pyramidal neurons in schizophrenic patients. *Neuroreport*. 2000;11:3621–3625.
- Broadbelt K, Byne W, Jones LB. Evidence for a decrease in basilar dendrites of pyramidal cells in schizophrenic medial prefrontal cortex. *Schizophr Res.* 2002;58:75–81.
- Konopaske GT, Lange N, Coyle JT, Benes FM. Prefrontal cortical dendritic spine pathology in schizophrenia and bipolar disorder. *JAMA Psychiatry*. 2014;71:1323–1331.
- Garey LJ, Ong WY, Patel TS, et al. Reduced dendritic spine density on cerebral cortical pyramidal neurons in schizophrenia. J Neurol Neurosurg Psychiatry. 1998;65:446–453.
- 44. Glantz LA, Lewis DA. Decreased dendritic spine density on prefrontal cortical pyramidal neurons in schizophrenia. Arch Gen Psychiatry. 2000;57:65–73.
- 45. Shelton MA, Newman JT, Gu H, et al. Loss of microtubule associated protein 2 immunoreactivity linked to dendritic spine loss in schizophrenia. *Biol Psychiatry*. 2015; 78:1–12.
- 46. Rajkowska G, Selemon LD, Goldman-Rakic PS. Neuronal and glial somal size in the prefrontal cortex: a postmortem morphometric study of schizophrenia and Huntington disease. *Arch Gen Psychiatry*. 1998;55:215–224.

- 47. Pierri JN, Volk CL, Auh S, Sampson A, Lewis DA. Decreased somal size of deep layer 3 pyramidal neurons in the prefrontal cortex of subjects with schizophrenia. *Arch Gen Psychiatry*. 2001;58:466–473.
- 48. Sweet RA, Henteleff RA, Zhang W, Sampson AR, Lewis DA. Reduced dendritic spine density in auditory cortex of subjects with schizophrenia. *Neuropsychopharmacology*. 2009;34:374–389.
- 49. Parker EM, Sweet RA. Stereological assessments of neuronal pathology in auditory cortex in schizophrenia. *Front Neuroanat*. 2017;11:131.
- 50. Kim IH, Racz B, Wang H, et al. Disruption of Arp2/3 results in asymmetric structural plasticity of dendritic spines and progressive synaptic and behavioral abnormalities. *J Neurosci.* 2013;33:6081–6092.
- Cahill ME, Xie Z, Day M, et al. Kalirin regulates cortical spine morphogenesis and disease-related behavioral phenotypes. *Proc Natl Acad Sci USA*. 2009;106:13058–13063.
- 52. Hains AB, Vu MA, Maciejewski PK, van Dyck CH, Gottron M, Arnsten AF. Inhibition of protein kinase C signaling protects prefrontal cortex dendritic spines and cognition from the effects of chronic stress. *Proc Natl Acad Sci USA*. 2009;106:17957–17962.
- 53. Bora E, Murray RM. Meta-analysis of cognitive deficits in ultra-high risk to psychosis and first-episode psychosis: do the cognitive deficits progress over, or after, the onset of psychosis? Schizophr Bull. 2014;40:744–755.
- Chan RC, Geng FL, Lui SS, et al. Course of neurological soft signs in first-episode schizophrenia: relationship with negative symptoms and cognitive performances. *Sci Rep.* 2015;5:11053.
- 55. Jones EG, Powell TP. Morphological variations in the dendritic spines of the neocortex. *J Cell Sci*. 1969;5:509–529.
- Peters A, Kaiserman-Abramof IR. The small pyramidal neuron of the rat cerebral cortex. The perikaryon, dendrites and spines. *Am J Anat*. 1970;127:321–355.
- 57. Arellano JI, Benavides-Piccione R, Defelipe J, Yuste R. Ultrastructure of dendritic spines: correlation between synaptic and spine morphologies. *Front Neurosci*. 2007;1:131–143.
- Kerchner GA, Nicoll RA. Silent synapses and the emergence of a postsynaptic mechanism for LTP. *Nat Rev Neurosci*. 2008;9:813–825.
- Nimchinsky EA, Sabatini BL, Svoboda K. Structure and function of dendritic spines. *Annu Rev Physiol.* 2002;64:313–353.
- 60. Busetto G, Higley MJ, Sabatini BL. Developmental presence and disappearance of postsynaptically silent synapses on dendritic spines of rat layer 2/3 pyramidal neurons. *J Physiol.* 2008;586:1519–1527.
- 61. Knott G, Holtmaat A. Dendritic spine plasticity-current understanding from in vivo studies. *Brain Res Rev.* 2008;58:282–289.
- MacDonald ML, Alhassan J, Newman JT, et al. Selective loss of smaller spines in schizophrenia. *Am J Psychiatry*. 2017;174:586–594.
- 63. Eastwood SL, Harrison PJ. Decreased expression of vesicular glutamate transporter 1 and complexin II mRNAs in schizophrenia: further evidence for a synaptic pathology affecting glutamate neurons. *Schizophr Res.* 2005;73:159–172.
- 64. Oni-Orisan A, Kristiansen LV, Haroutunian V, Meador-Woodruff JH, McCullumsmith RE. Altered vesicular glutamate transporter expression in the anterior cingulate cortex in schizophrenia. *Biol Psychiatry*. 2008;63:766–775.
- 65. Bitanihirwe BK, Lim MP, Kelley JF, Kaneko T, Woo TU. Glutamatergic deficits and parvalbumin-containing inhibitory

neurons in the prefrontal cortex in schizophrenia. BMC Psychiatry. 2009;9:71.

- 66. Fremeau RT Jr, Troyer MD, Pahner I, et al. The expression of vesicular glutamate transporters defines two classes of excitatory synapse. *Neuron*. 2001;31:247–260.
- 67. Kaneko T, Fujiyama F. Complementary distribution of vesicular glutamate transporters in the central nervous system. *Neurosci Res.* 2002;42:243–250.
- Fremeau RT Jr, Voglmaier S, Seal RP, Edwards RH. VGLUTs define subsets of excitatory neurons and suggest novel roles for glutamate. *Trends Neurosci*. 2004;27:98–103.
- 69. Stevens JR. Neuropathology of schizophrenia. Arch Gen Psychiatry. 1982;39:1131–1139.
- Nieto D, Escobar A. Major psychoses. In: Minkler J, ed. Pathology of the Nervous System. New York, NY: McGraw-Hill International Book Co; 1972:2654–2665.
- 71. Fisman M. The brain stem in psychosis. Br J Psychiatry. 1975;126:414–422.
- Sterio DC. The unbiased estimation of number and sizes of arbitrary particles using the disector. J Microsc. 1984;134:127–136.
- 73. Gundersen HJ, Bagger P, Bendtsen TF, et al. The new stereological tools: disector, fractionator, nucleator and point sampled intercepts and their use in pathological research and diagnosis. *APMIS*. 1988;96:857–881.
- 74. Dwork AJ. Postmortem studies of the hippocampal formation in schizophrenia. *Schizophr Bull*. 1997;23:385–402.
- Roberts GW, Colter N, Lofthouse R, Bogerts B, Zech M, Crow TJ. Gliosis in schizophrenia: a survey. *Biol Psychiatry*. 1986;21:1043–1050.
- Eng LF, Vanderhaeghen JJ, Bignami A, Gerstl B. An acidic protein isolated from fibrous astrocytes. *Brain Res.* 1971;28:351–354.
- Bignami A, Eng LF, Dahl D, Uyeda CT. Localization of the glial fibrillary acidic protein in astrocytes by immunofluorescence. *Brain Res.* 1972;43:429–435.
- Roberts GW, Colter N, Lofthouse R, Johnstone EC, Crow TJ. Is there gliosis in schizophrenia? Investigation of the temporal lobe. *Biol Psychiatry*. 1987;22:1459–1468.
- Stevens CD, Altshuler LL, Bogerts B, Falkai P. Quantitative study of gliosis in schizophrenia and Huntington's chorea. *Biol Psychiatry*. 1988;24:697–700.
- Casanova MF, Stevens JR, Kleinman JE. Astrocytosis in the molecular layer of the dentate gyrus: a study in Alzheimer's disease and schizophrenia. *Psychiatry Res.* 1990;35:149–166.
- 81. Arnold SE, Franz BR, Trojanowski JQ, Moberg PJ, Gur RE. Glial fibrillary acidic protein-immunoreactive astrocytosis in elderly patients with schizophrenia and dementia. *Acta Neuropathol.* 1996;91:269–277.
- 82. Arnold SE, Trojanowski JQ, Gur RE, Blackwell P, Han LY, Choi C. Absence of neurodegeneration and neural injury in the cerebral cortex in a sample of elderly patients with schizophrenia. *Arch Gen Psychiatry*. 1998;55:225–232.
- Falkai P, Honer WG, David S, Bogerts B, Majtenyi C, Bayer TA. No evidence for astrogliosis in brains of schizophrenic patients. A post-mortem study. *Neuropathol Appl Neurobiol*. 1999;25:48–53.
- 84. Falke E, Han LY, Arnold SE. Absence of neurodegeneration in the thalamus and caudate of elderly patients with schizophrenia. *Psychiatry Res.* 2000;93:103–110.
- 85. Radewicz K, Garey LJ, Gentleman SM, Reynolds R. Increase in HLA-DR immunoreactive microglia in frontal and temporal cortex of chronic schizophrenics. J Neuropathol Exp Neurol. 2000;59:137–150.

- 86. Damadzic R, Bigelow LB, Krimer LS, et al. A quantitative immunohistochemical study of astrocytes in the entorhinal cortex in schizophrenia, bipolar disorder and major depression: absence of significant astrocytosis. *Brain Res Bull*. 2001;55:611–618.
- 87. Webster MJ, Knable MB, Johnston-Wilson N, Nagata K, Inagaki M, Yolken RH. Immunohistochemical localization of phosphorylated glial fibrillary acidic protein in the prefrontal cortex and hippocampus from patients with schizophrenia, bipolar disorder, and depression. *Brain Behav Immun*. 2001;15:388–400.
- 88. Rajkowska G, Miguel-Hidalgo JJ, Makkos Z, Meltzer H, Overholser J, Stockmeier C. Layer-specific reductions in GFAP-reactive astroglia in the dorsolateral prefrontal cortex in schizophrenia. *Schizophr Res.* 2002;57:127–138.
- Altshuler LL, Abulseoud OA, Foland-Ross L, et al. Amygdala astrocyte reduction in subjects with major depressive disorder but not bipolar disorder. *Bipolar Disord*. 2010;12: 541–549.
- Williams MR, Marsh R, Macdonald CD, et al. Neuropathological changes in the nucleus basalis in schizophrenia. *Eur Arch Psychiatry Clin Neurosci.* 2013;263:485–495.
- Williams MR, Hampton T, Pearce RK, et al. Astrocyte decrease in the subgenual cingulate and callosal genu in schizophrenia. *Eur Arch Psychiatry Clin Neurosci.* 2013;263:41–52.
- 92. Pantazopoulos H, Woo TU, Lim MP, Lange N, Berretta S. Extracellular matrix-glial abnormalities in the amygdala and entorhinal cortex of subjects diagnosed with schizophrenia. *Arch Gen Psychiatry*. 2010;67:155–166.
- Hercher C, Chopra V, Beasley CL. Evidence for morphological alterations in prefrontal white matter glia in schizophrenia and bipolar disorder. *J Psychiatry Neurosci.* 2014;39: 376–385.
- Benes FM, Davidson J, Bird ED. Quantitative cytoarchitectural studies of the cerebral cortex of schizophrenics. *Arch Gen Psychiatry*. 1986;43:31–35.
- 95. Williams M, Pearce RK, Hirsch SR, Ansorge O, Thom M, Maier M. Fibrillary astrocytes are decreased in the subgenual cingulate in schizophrenia. *Eur Arch Psychiatry Clin Neurosci*. 2014;264:357–362.
- Williams MR, Galvin K, O'Sullivan B, et al. Neuropathological changes in the substantia nigra in schizophrenia but not depression. *Eur Arch Psychiatry Clin Neurosci.* 2014;264:285–296.
- 97. Fatemi SH, Laurence JA, Araghi-Niknam M, et al. Glial fibrillary acidic protein is reduced in cerebellum of subjects with major depression, but not schizophrenia. *Schizophr Res.* 2004;69:317–323.
- Karson CN, Casanova MF, Kleinman JE, Griffin WS. Choline acetyltransferase in schizophrenia. *Am J Psychiatry*. 1993;150:454–459.
- 99. Katsel P, Byne W, Roussos P, Tan W, Siever L, Haroutunian V. Astrocyte and glutamate markers in the superficial, deep, and white matter layers of the anterior cingulate gyrus in schizophrenia. *Neuropsychopharmacology*. 2011;36:1171–1177.
- 100. Dean B, Gray L, Scarr E. Regionally specific changes in levels of cortical S100beta in bipolar 1 disorder but not schizophrenia. *Aust N Z J Psychiatry*. 2006;40:217–224.
- Tkachev D, Mimmack ML, Ryan MM, et al. Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. *Lancet*. 2003;362:798–805.
- 102. Perrone-Bizzozero NI, Sower AC, Bird ED, Benowitz LI, Ivins KJ, Neve RL. Levels of the growth-associated protein GAP-43 are selectively increased in association cortices in schizophrenia. *Proc Natl Acad Sci U S A*. 1996;93:14182–14187.

- 103. Trépanier MO, Hopperton KE, Mizrahi R, Mechawar N, Bazinet RP. Postmortem evidence of cerebral inflammation in schizophrenia: a systematic review. *Mol Psychiatry*. 2016;21:1009–1026.
- 104. Cahoy JD, Emery B, Kaushal A, et al. A transcriptome database for astrocytes, neurons, and oligodendrocytes: a new resource for understanding brain development and function. *J Neurosci.* 2008;28:264–278.
- 105. Nimgaonkar VL, Prasad KM, Chowdari KV, Severance EG, Yolken RH. The complement system: a gateway to geneenvironment interactions in schizophrenia pathogenesis. *Mol Psychiatry*. 2017;22:1554–1561.
- 106. Müller N. Inflammation in schizophrenia: pathogenetic aspects and therapeutic considerations [published online ahead of print April 10, 2018]. *Schizophr Bull.* doi:10.1093/ schbul/sby024.
- 107. Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophreniaassociated genetic loci. *Nature*. 2014;511:421–427.
- 108. Sekar A, Bialas AR, de Rivera H, et al; Schizophrenia Working Group of the Psychiatric Genomics Consortium. Schizophrenia risk from complex variation of complement component 4. *Nature*. 2016;530:177–183.
- 109. Wierzba-Bobrowicz T, Lewandowska E, Lechowicz W, Stepień T, Pasennik E. Quantitative analysis of activated microglia, ramified and damage of processes in the frontal and temporal lobes of chronic schizophrenics. *Folia Neuropathol.* 2005;43:81–89.
- 110. Fillman SG, Cloonan N, Catts VS, et al. Increased inflammatory markers identified in the dorsolateral prefrontal cortex of individuals with schizophrenia. *Mol Psychiatry*. 2013;18:206–214.
- 111. Busse S, Busse M, Schiltz K, et al. Different distribution patterns of lymphocytes and microglia in the hippocampus of patients with residual versus paranoid schizophrenia: further evidence for disease course-related immune alterations? *Brain Behav Immun.* 2012;26:1273–1279.
- 112. Bayer TA, Buslei R, Havas L, Falkai P. Evidence for activation of microglia in patients with psychiatric illnesses. *Neurosci Lett.* 1999;271:126–128.
- 113. Wierzba-Bobrowicz T, Lewandowska E, Kosno-Kruszewska E, Lechowicz W, Pasennik E, Schmidt-Sidor B. Degeneration of microglial cells in frontal and temporal lobes of chronic schizophrenics. *Folia Neuropathol.* 2004;42:157–165.
- 114. Steiner J, Mawrin C, Ziegeler A, et al. Distribution of HLA-DR-positive microglia in schizophrenia reflects impaired cerebral lateralization. *Acta Neuropathol.* 2006;112:305–316.
- 115. Steiner J, Bielau H, Brisch R, et al. Immunological aspects in the neurobiology of suicide: elevated microglial density in schizophrenia and depression is associated with suicide. *J Psychiatr Res.* 2008;42:151–157.
- Connor CM, Guo Y, Akbarian S. Cingulate white matter neurons in schizophrenia and bipolar disorder. *Biol Psychiatry*. 2009;66:486–493.
- 117. van Kesteren CF, Gremmels H, de Witte LD, et al. Immune involvement in the pathogenesis of schizophrenia: a metaanalysis on postmortem brain studies. *Transl Psychiatry*. 2017;7:e1075.
- 118. Bennett ML, Bennett FC, Liddelow SA, et al. New tools for studying microglia in the mouse and human CNS. *Proc Natl Acad Sci U S A*. 2016;113:E1738–E1746.
- 119. Satoh J, Kino Y, Asahina N, et al. TMEM119 marks a subset of microglia in the human brain. *Neuropathology*. 2016;36:39–49.

- 120. Konishi H, Kobayashi M, Kunisawa T, et al. Siglec-H is a microglia-specific marker that discriminates microglia from CNS-associated macrophages and CNS-infiltrating monocytes. *Glia*. 2017;65:1927–1943.
- 121. Papadopoulos V, Baraldi M, Guilarte TR, et al. Translocator protein (18kDa): new nomenclature for the peripheral-type benzodiazepine receptor based on its structure and molecular function. *Trends Pharmacol Sci.* 2006;27:402–409.
- 122. Papadopoulos V, Fan J, Zirkin B. Translocator protein (18 kDa): an update on its function in steroidogenesis. *J Neuroendocrinol*. 2018;30:e12500.
- 123. Banati RB, Newcombe J, Gunn RN, et al. The peripheral benzodiazepine binding site in the brain in multiple sclerosis: quantitative in vivo imaging of microglia as a measure of disease activity. *Brain.* 2000;123(pt 11):2321–2337.
- 124. Banati RB. Visualising microglial activation in vivo. *Glia*. 2002;40:206–217.
- 125. Messmer K, Reynolds GP. Increased peripheral benzodiazepine binding sites in the brain of patients with Huntington's disease. *Neurosci Lett*. 1998;241:53–56.
- 126. Turner MR, Cagnin A, Turkheimer FE, et al. Evidence of widespread cerebral microglial activation in amyotrophic lateral sclerosis: an [11C](R)-PK11195 positron emission tomography study. *Neurobiol Dis.* 2004;15:601–609.
- 127. Batarseh A, Papadopoulos V. Regulation of translocator protein 18 kDa (TSPO) expression in health and disease states. *Mol Cell Endocrinol*. 2010;327:1–12.
- 128. McNeela AM, Bernick C, Hines RM, Hines DJ. TSPO regulation in reactive gliotic diseases. J Neurosci Res. 2018;96:978–988.
- 129. Vowinckel E, Reutens D, Becher B, et al. PK11195 binding to the peripheral benzodiazepine receptor as a marker of microglia activation in multiple sclerosis and experimental autoimmune encephalomyelitis. J Neurosci Res. 1997;50:345–353.
- 130. Owen DR, Yeo AJ, Gunn RN, et al. An 18-kDa translocator protein (TSPO) polymorphism explains differences in binding affinity of the PET radioligand PBR28. J Cereb Blood Flow Metab. 2012;32:1–5.
- 131. van Berckel BN, Bossong MG, Boellaard R, et al. Microglia activation in recent-onset schizophrenia: a quantitative (R)-[11C]PK11195 positron emission tomography study. *Biol Psychiatry*. 2008;64:820–822.
- 132. Doorduin J, de Vries EF, Willemsen AT, de Groot JC, Dierckx RA, Klein HC. Neuroinflammation in schizophrenia-related psychosis: a PET study. *J Nucl Med.* 2009;50:1801–1807.
- 133. Takano A, Arakawa R, Ito H, et al. Peripheral benzodiazepine receptors in patients with chronic schizophrenia: a PET study with [11C]DAA1106. *Int J Neuropsychopharmacol.* 2010;13:943–950.
- 134. Bloomfield PS, Selvaraj S, Veronese M, et al. Microglial activity in people at ultra high risk of psychosis and in schizophrenia: an [(11)C]PBR28 PET brain imaging study. Am J Psychiatry. 2016;173:44–52.
- 135. van der Doef TF, de Witte LD, Sutterland AL, et al. In vivo (R)-[(11)C]PK11195 PET imaging of 18kDa translocator protein in recent onset psychosis. *NPJ Schizophr*. 2016;2:16031.
- 136. Coughlin JM, Wang Y, Ambinder EB, et al. In vivo markers of inflammatory response in recent-onset schizophrenia: a combined study using [(11)C]DPA-713 PET and analysis of CSF and plasma. *Transl Psychiatry*. 2016;6:e777.
- 137. Di Biase MA, Zalesky A, O'keefe G, et al. PET imaging of putative microglial activation in individuals at ultra-high risk for psychosis, recently diagnosed and chronically ill with schizophrenia. *Transl Psychiatry*. 2017;7:e1225.

- 138. Hafizi S, Tseng HH, Rao N, et al. Imaging microglial activation in untreated first-episode psychosis: a PET Study With [¹⁸F]FEPPA. *Am J Psychiatry*. 2017;174:118–124.
- 139. Selvaraj S, Bloomfield PS, Cao B, Veronese M, Turkheimer F, Howes OD. Brain TSPO imaging and gray matter volume in schizophrenia patients and in people at ultra high risk of psychosis: an [¹¹C]PBR28 study. *Schizophr Res.* 2018;195:206–214.
- 140. Ottoy J, De Picker L, Verhaeghe J, et al. [¹⁸F]PBR111 PET imaging in healthy controls and schizophrenia: test–retest reproducibility and quantification of neuroinflammation[published online ahead of print January 11, 2018]. *J Nucl Med.* doi:10.2967/jnumed.117.203315.
- 141. Cosenza-Nashat M, Zhao ML, Suh HS, et al. Expression of the translocator protein of 18 kDa by microglia, macrophages and astrocytes based on immunohistochemical localization in abnormal human brain. *Neuropathol Appl Neurobiol*. 2009;35:306–328.
- 142. Lavisse S, Guillermier M, Hérard AS, et al. Reactive astrocytes overexpress TSPO and are detected by TSPO positron emission tomography imaging. J Neurosci. 2012;32:10809–10818.
- 143. Notter T, Coughlin JM, Gschwind T, et al. Translational evaluation of translocator protein as a marker of neuroinflammation in schizophrenia. *Mol Psychiatry*. 2018;23:323–334.
- 144. Varga B, Markó K, Hádinger N, et al. Translocator protein (TSPO 18kDa) is expressed by neural stem and neuronal precursor cells. *Neurosci Lett*. 2009;462:257–262.
- 145. Owen DR, Narayan N, Wells L, et al. Pro-inflammatory activation of primary microglia and macrophages increases 18kDa translocator protein expression in rodents but not humans. *J Cereb Blood Flow Metab.* 2017;37:2679–2690.
- 146. O'Donnell P. Microglia activation in subjects at risk for psychosis: fact or fiction? *Neuropsychopharmacology*. 2017;42:2472–2473.
- 147. Michell-Robinson MA, Touil H, Healy LM, et al. Roles of microglia in brain development, tissue maintenance and repair. *Brain*. 2015;138:1138–1159.
- 148. Turkheimer FE, Rizzo G, Bloomfield PS, et al. The methodology of TSPO imaging with positron emission tomography. *Biochem Soc Trans.* 2015;43:586–592.
- 149. Lockhart A, Davis B, Matthews JC, et al. The peripheral benzodiazepine receptor ligand PK11195 binds with high affinity to the acute phase reactant alpha1-acid glycoprotein: implications for the use of the ligand as a CNS inflammatory marker. *Nucl Med Biol.* 2003;30:199–206.
- 150. De Picker LJ, Morrens M, Chance SA, Boche D. Microglia and brain plasticity in acute psychosis and schizophrenia illness course: a meta-review. *Front Psychiatry*. 2017;8:238.
- 151. Innocenti GM, Price DJ. Exuberance in the development of cortical networks. *Nat Rev Neurosci*. 2005;6:955–965.
- 152. Katz LC, Shatz CJ. Synaptic activity and the construction of cortical circuits. *Science*. 1996;274:1133–1138.
- 153. Petanjek Z, Judaš M, Šimic G, et al. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc Natl Acad Sci USA*. 2011;108:13281–13286.
- 154. Huttenlocher PR. Synaptic density in human frontal cortex—developmental changes and effects of aging. *Brain Res.* 1979;163:195–205.
- 155. Feinberg I. Schizophrenia: caused by a fault in programmed synaptic elimination during adolescence? *J Psychiatr Res.* 1982;17:319–334.
- 156. Murray RM, Bhavsar V, Tripoli G, Howes O. 30 Years on: how the neurodevelopmental hypothesis of schizophrenia

morphed into the developmental risk factor model of psychosis. *Schizophr Bull*. 2017;43:1190–1196.

- 157. Ginhoux F, Greter M, Leboeuf M, et al. Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science*. 2010;330:841–845.
- 158. Ajami B, Bennett JL, Krieger C, Tetzlaff W, Rossi FM. Local self-renewal can sustain CNS microglia maintenance and function throughout adult life. *Nat Neurosci*. 2007;10:1538–1543.
- 159. Nimmerjahn A, Kirchhoff F, Helmchen F. Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo. *Science*. 2005;308:1314–1319.
- Davalos D, Grutzendler J, Yang G, et al. ATP mediates rapid microglial response to local brain injury in vivo. *Nat Neurosci*. 2005;8:752–758.
- 161. Tremblay MÈ, Lowery RL, Majewska AK. Microglial interactions with synapses are modulated by visual experience. *PLoS Biol.* 2010;8:e1000527.
- 162. Hong S, Dissing-Olesen L, Stevens B. New insights on the role of microglia in synaptic pruning in health and disease. *Curr Opin Neurobiol*. 2016;36:128–134.
- 163. Kierdorf K, Prinz M. Microglia in steady state. *J Clin Invest*. 2017;127:3201–3209.
- 164. Miyamoto A, Wake H, Ishikawa AW, et al. Microglia contact induces synapse formation in developing somatosensory cortex. *Nat Commun*. 2016;7:12540.
- 165. Weinhard L, di Bartolomei G, Bolasco G, et al. Microglia remodel synapses by presynaptic trogocytosis and spine head filopodia induction. *Nat Commun.* 2018;9:1228.
- 166. Parkhurst CN, Yang G, Ninan I, et al. Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor. *Cell*. 2013;155:1596–1609.
- 167. Paolicelli RC, Bolasco G, Pagani F, et al. Synaptic pruning by microglia is necessary for normal brain development. *Science*. 2011;333:1456–1458.
- 168. Schafer DP, Lehrman EK, Kautzman AG, et al. Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron*. 2012;74:691–705.
- 169. Stevens B, Allen NJ, Vazquez LE, et al. The classical complement cascade mediates CNS synapse elimination. *Cell*. 2007;131:1164–1178.
- 170. Mallya AP, Wang H-D, Lee HNR, Deutch AY. Microglial pruning of synapses in the prefrontal cortex during adolescence [published online ahead of print April 13, 2018]. *Cereb Cortex*. doi:10.1093/cercor/bhy061.
- 171. Miller M, Peters A. Maturation of rat visual cortex. II. A combined Golgi-electron microscope study of pyramidal neurons. *J Comp Neurol.* 1981;203:555–573.
- 172. Yuste R, Bonhoeffer T. Genesis of dendritic spines: insights from ultrastructural and imaging studies. *Nat Rev Neurosci*. 2004;5:24–34.
- 173. Knott GW, Holtmaat A, Wilbrecht L, Welker E, Svoboda K. Spine growth precedes synapse formation in the adult neocortex in vivo. *Nat Neurosci*. 2006;9:1117–1124.
- 174. Chung WS, Allen NJ, Eroglu C. Astrocytes control synapse formation, function, and elimination. *Cold Spring Harb Perspect Biol.* 2015;7:a020370.
- 175. Chung WS, Clarke LE, Wang GX, et al. Astrocytes mediate synapse elimination through MEGF10 and MERTK pathways. *Nature*. 2013;504:394–400.
- 176. Bialas AR, Stevens B. TGF- β signaling regulates neuronal C1q expression and developmental synaptic refinement. *Nat Neurosci.* 2013;16:1773–1782.

- 177. Liddelow SA, Guttenplan KA, Clarke LE, et al. Neurotoxic reactive astrocytes are induced by activated microglia. *Nature*. 2017;541:481–487.
- 178. Liddelow SA, Barres BA. Reactive astrocytes: production, function, and therapeutic potential. *Immunity*. 2017;46:957–967.
- 179. Hanisch UK, Kettenmann H. Microglia: active sensor and versatile effector cells in the normal and pathologic brain. *Nat Neurosci.* 2007;10:1387–1394.
- Ransohoff RM, Perry VH. Microglial physiology: unique stimuli, specialized responses. Annu Rev Immunol. 2009;27:119–145.
- 181. Sierra A, Encinas JM, Deudero JJ, et al. Microglia shape adult hippocampal neurogenesis through apoptosis-coupled phagocytosis. *Cell Stem Cell*. 2010;7:483–495.
- 182. De Biase LM, Schuebel KE, Fusfeld ZH, et al. Local cues establish and maintain region-specific phenotypes of basal ganglia microglia. *Neuron*. 2017;95:341–356.e6.

- 183. Prinz M, Priller J, Sisodia SS, Ransohoff RM. Heterogeneity of CNS myeloid cells and their roles in neurodegeneration. *Nat Neurosci.* 2011;14:1227–1235.
- 184. Prinz M, Priller J. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. *Nat Rev Neurosci.* 2014;15:300–312.
- 185. Pineau I, Lacroix S. Proinflammatory cytokine synthesis in the injured mouse spinal cord: multiphasic expression pattern and identification of the cell types involved. *J Comp Neurol.* 2007;500:267–285.
- 186. Hochreiter-Hufford A, Ravichandran KS. Clearing the dead: apoptotic cell sensing, recognition, engulfment, and digestion. *Cold Spring Harb Perspect Biol.* 2013;5:a008748.
- 187. Penzes P, Cahill ME, Jones KA, VanLeeuwen JE, Woolfrey KM. Dendritic spine pathology in neuropsychiatric disorders. *Nat Neurosci.* 2011;14:285–293.