



# Xpert Ultra Can Unambiguously Identify Specific Rifampin Resistance-Confering Mutations

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**KEYWORDS** Xpert MTB/RIF Ultra, rifampin-resistant tuberculosis, *rpoB* mutations, disputed mutations, Ultra probes, melt peak temperature ( $T_m$ ), melting temperature shift ( $\Delta T_m$ )

The deluge of data produced by the Xpert MTB/RIF test (Cepheid) can help improve global rifampin-resistant tuberculosis (RR-TB) control strategies through molecular epidemiological surveillance (1, 2). Recently, a new version of the test, Xpert Ultra (hereinafter called Ultra), was released (3). Determining the relationship between RR-confering *rpoB* mutations, Ultra probes, and melting temperature shifts ( $\Delta T_m$ ), i.e., the difference between mutant and wild-type melting temperatures, allows Ultra results to be utilized for rapid detection of RR-TB strains and related underlying *rpoB* mutations.

To determine the reliability of Ultra results for predicting specific mutations, we tested 13 rifampin-susceptible (RS)-TB strains and 104 RR-TB strains harboring 33 unique RR-confering mutations from the Belgian Coordinated Collections of Microorganisms in the Institute of Tropical Medicine Antwerp according to a protocol previously described (2) (see the supplemental material). Of note, the Glu250Gly ( $n = 2$ ) and Arg299Cys ( $n = 1$ ) mutations were among the RS-TB strains. We then compared Ultra raw results with available *rpoB* sequences of the strains.

Overall, 29/30 (97%) mutations inside the rifampin resistance-determining region (RRDR) were correctly identified by Ultra. Of concern, mutation His445Arg gave a “RIF Resistance Indeterminate” result among 3/4 strains tested, while it was reported as RR in the initial validation study (3). The silent mutation Thr444Thr was not reported as RR (Fig. 1). The RR-confering mutations on codons 170 and 491 situated outside the RRDR were not detected.

The probe reactions observed were largely in agreement with previous results (3), although we noted that mutations Met434Val, Met434Thr, and those in codon 435 were captured only by probe *rpoB2*, Ser450Leu and Ser450Trp were captured by both probe *rpoB3* and probe *rpoB4a*, His445Arg was captured only by probe *rpoB3*, and Lys446Gln was captured only by probe *rpoB4*.

All mutations except those in codon 450 were associated with a negative  $\Delta T_m$  (Fig. 2). The combination of  $\Delta T_m$  values with the capturing probes enabled us to differentiate mutations in codons 430, 431, 434, 435, 441, 446, 450, and 452, including disputed mutations (4) (Table 1). Mutation Asp435Tyr was unambiguously distinguished from Asp435Val with the  $|\Delta T_m|$  of probe *rpoB2*, while mutations Ser441Gln and Ser441Leu were discriminated from the rest by the  $|\Delta T_m|$  values of probes *rpoB2* and *rpoB3*. Mutations His445Asp and His445Tyr were distinguished from disputed mutations

Accepted manuscript posted online 20 June 2018

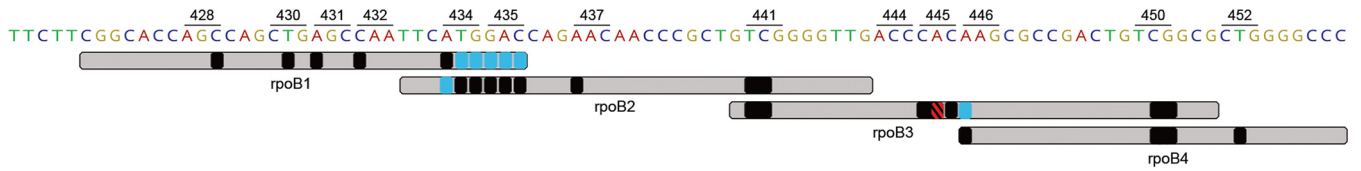
**Citation** Ng KCS, van Deun A, Meehan CJ, Torrea G, Driesen M, Gabriëls S, Rigouts L, André E, de Jong BC. 2018. Xpert Ultra can unambiguously identify specific rifampin resistance-confering mutations. *J Clin Microbiol* 56:e00686-18. <https://doi.org/10.1128/JCM.00686-18>.

**Editor** Karen C. Carroll, Johns Hopkins University School of Medicine

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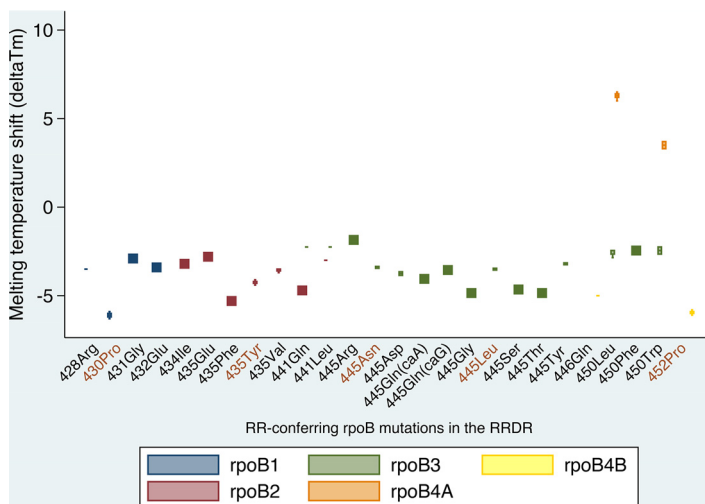
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**FIG 1** Overview of Xpert Ultra test results. The observed probe reactions for each RRDR mutation were laid over the claimed probe coverage (light gray). Shown in black are probe reactions concordant with manufacturer claims, in blue are probe reactions missed by one probe but captured by another probe, and in red is a probe reaction representing a “RIF Resistance Indeterminate” result from 3 out of 4 strains tested. Results in the hatched pattern were superimposed for greater visibility.

His445Leu and His445Asn through the  $|\Delta T_m|$  of probe rpoB3. Ser450Leu was distinguished from Ser450Trp by the  $|\Delta T_m|$  of probe rpoB4A. The indeterminate result associated with His445Arg may be caused by its  $|\Delta T_m|$  being equal to 1.8°C, unlike the  $|\Delta T_m|$  values for other mutations, which typically exceed 2°C. Our recent experience with Ultra on diagnostic sputum samples pertained only to the Ser450Leu and His445Asp mutations, for which the  $\Delta T_m$  corresponded exactly with the  $\Delta T_m$  that we observed for bacterial thermolysates. This should be validated more extensively, which is beyond the scope of our present study.

Our findings confirm the ability of Ultra to unambiguously identify a wide range of RRDR mutations. With the unprecedented rollout of Xpert MTB/RIF and associated connectivity solutions, such as DataToCare (Savics, Belgium) and GXAlert (SystemOne, USA) (2), Ultra results may allow us to rule out transmission between RR-TB patients in a specific setting (Fig. S1), distinguish relapse from reinfection (5) (Fig. S2), and resolve discordance between an RR Ultra result and a low-level RS phenotypic result due to a disputed mutation. For such applications, it is key that  $\Delta T_m$  values are included in the exported results.



**FIG 2** Melting temperature shifts ( $\Delta T_m$ s) observed upon detection of a rifampin resistance (RR)-conferring *rpoB* mutation in the RR-determining region (RRDR) by Xpert Ultra. The y axis reflects the melting temperature difference ( $\Delta T_m$ ) between mutant and wild-type probe-amplicon hybrids, while the x axis shows the mutations that we tested. The data points on the graph are  $\Delta T_m$  values grouped by their associated Ultra probes (differentiated by color), which correspond to a specific *rpoB* mutation. x axis labels in brown are disputed mutations.

**TABLE 1** Xpert Ultra raw results<sup>a</sup>

Mutation(s) <sup>b</sup>	No. of strains tested	Nucleotide change(s)	Xpert Ultra probe(s)	Wild-type $T_m$ range(s) (mean[s])	Mutant $T_m$ range(s)	$ \Delta T_m $ mean(s) or range(s)
Val170Phe	3	GTC→TTC	ND	ND	ND	ND
Glu250Gly <sup>#</sup>	2	GAG→GGG	ND	ND	ND	ND
Arg299Cys <sup>#</sup>	1	CGC→TGC	ND	ND	ND	ND
* <i>Leu430Pro</i>	8	CTG→CCG	<i>rpoB1</i>	69.1–69.5 (69.3)	63.0–63.4	5.9–6.3
<u><i>Leu430Pro</i></u> + *Met434Ile	1	CTG→CCG; ATG→ATA	<i>rpoB1</i> ; <i>rpoB2</i>	69.1–69.5 (69.3); 72.8–73.2 (73)	63.2; 69.8	6.1; 3.2
<u><i>Leu430Pro</i></u> + Met434Val	1	CTG→CCG; ATG→GTG	<i>rpoB1</i>	69.1–69.5 (69.3)	63.0	6.3
<u><i>Leu430Pro</i></u> + His445Gln	1	CTG→CCG; CAC→CAG	<i>rpoB1</i> ; <i>rpoB3</i>	69.1–69.5 (69.3); 75.5–76.0 (75.75)	63.5; 72.2	5.8; 3.6
<u><i>Leu430Pro</i></u> + His445Gln	1	CTG→CCG; CAC→CAA	<i>rpoB1</i> ; <i>rpoB3</i>	69.1–69.5 (69.3); 75.5–76.0 (75.75)	63.1; 71.7	6.2; 4.1
<u><i>Asp435Gly</i></u> + Met434Thr	1	GAC→GGC; ATG→ACG	<i>rpoB2</i>	72.8–73.2 (73)	69.7	3.3
* <i>Asp435Phe</i>	1	GAC→TTC	<i>rpoB2</i>	72.8–73.2 (73)	67.7	5.3
* <i>Asp435Tyr</i>	11	GAC→TAC	<i>rpoB2</i>	72.8–73.2 (73)	68.6–69.0	4.0–4.4
<u><i>Asp435Tyr</i></u> + Asn437Asp	1	GAC→TAC; AAC→GAC	<i>rpoB2</i>	72.8–73.2 (73)	66.6	6.4
<u><i>Asp435Tyr</i></u> + Met434Ile	1	GAC→TAC; ATG→ATT	<i>rpoB2</i>	72.8–73.2 (73)	68.5	4.5
* <i>Asp435Val</i>	5	GAC→GTC	<i>rpoB2</i>	72.8–73.2 (73)	69.3–69.5	3.5–3.7
<u><i>Asp435Val</i></u> + Gln432Glu	1	GAC→GTC; CAA→GAA	<i>rpoB2</i> ; <i>rpoB1</i>	72.8–73.2 (73); 69.1–69.5 (69.3)	70.5; 65.9	2.5; 3.4
* <i>Ser441Gln</i>	1	TCG→CAG	<i>rpoB2</i> ; <i>rpoB3</i>	72.8–73.2 (73); 75.5–76.0 (75.75)	68.3; 73.5	4.7; 2.3
* <i>Ser441Leu</i>	1	TCG→TTG	<i>rpoB2</i> ; <i>rpoB3</i>	72.8–73.2 (73); 75.5–76.0 (75.75)	70.0; 73.5	3.0; 2.3
His445Gly	1	CAC→GGC	<i>rpoB3</i>	75.5–76.0 (75.75)	70.9	4.9
His445Thr	1	CAC→ACC	<i>rpoB3</i>	75.5–76.0 (75.75)	70.9	4.9
His445Ser	1	CAC→AGC	<i>rpoB3</i>	75.5–76.0 (75.75)	71.1	4.7
His445Ser + * <i>Lys446Gln</i> + Thr444Thr	1	CAC→TCC; AAG→CAG; ACC→ACG	<i>rpoB4B</i>	67.0–67.6 (67.3)	62.3	5.0
His445Asp	3	CAC→GAC	<i>rpoB3</i>	75.5–76.0 (75.75)	71.9–72.1	3.7–3.9
<i>His445Leu</i>	2	CAC→CTC	<i>rpoB3</i>	75.5–76.0 (75.75)	72.2–72.3	3.5–3.6
<i>His445Asn</i>	2	CAC→AAC	<i>rpoB3</i>	75.5–76.0 (75.75)	72.3–72.4	3.4–3.5
<u><i>His445Asn</i></u> + * <i>Asp435Glu</i>	1	CAC→AAC; GAC→GAA	<i>rpoB3</i> ; <i>rpoB2</i>	75.5–76.0 (75.75); 72.8–73.2 (73)	72.4; 70.2	3.4; 2.8
His445Tyr	4	CAC→TAC	<i>rpoB3</i>	75.5–76.0 (75.75)	72.5–72.6	3.2–3.3
* <i>His445Arg</i>	4	CAC→CGC	<i>rpoB3</i>	75.5–76.0 (75.75)	73.9	1.9
<u><i>His445Arg</i></u> + Ser428Arg	1	CAC→CGC; AGC→AGG	<i>rpoB1</i>	69.1–69.5 (69.3)	65.8	3.5
Ser450Phe	1	TCG→TTC	<i>rpoB3</i>	75.5–76.0 (75.75)	71.8	4.0
* <i>Ser450Leu</i>	14	TCG→TTG	<i>rpoB3</i> ; <i>rpoB4A</i>	75.5–76.0 (75.75); 67.0–67.6 (67.3)	72.9–73.3; 73.3–73.8	2.5–2.9; 6.0–6.5
<u><i>Ser450Leu</i></u> + Thr482Asn	2	TCG→TTG; ACC→AAC	<i>rpoB2</i> ; <i>rpoB3</i> ; <i>rpoB4A</i>	72.8–73.2 (73); 75.5–76.0 (75.75); 67.0–67.6 (67.3)	69.2–69.5; 73.1–73.3; 73.6–73.7	3.5–3.8; 2.5–2.7; 6.3–6.4
<u><i>Ser450Leu</i></u> + Ile491Val	2	TCG→TTG; ATC→GTC	<i>rpoB2</i> ; <i>rpoB3</i> ; <i>rpoB4A</i>	72.8–73.2 (73); 75.5–76.0 (75.75); 67.0–67.6 (67.3)	70.0; 73.2–73.3; 73.6–73.7	3.0; 2.5–2.6; 6.3–6.4
* <i>Ser450Trp</i>	3	TCG→TGG	<i>rpoB3</i> ; <i>rpoB4A</i>	75.5–76.0 (75.75); 67.0–67.6 (67.3)	73.1–73.5; 70.6–71.0	2.3–2.7; 3.3–3.7
<u><i>Ser450Trp</i></u> + * <i>Ser431Gly</i>	1	TCG→TGG; AGC→GGC	<i>rpoB3</i> ; <i>rpoB4A</i> ; <i>rpoB1</i>	75.5–76.0 (75.75); 67.0–67.6 (67.3); 69.1–69.5 (69.3)	73.2; 70.7; 66.4	2.6; 3.4; 2.9
* <i>Leu452Pro</i>	12	CTG→CCG	<i>rpoB4B</i>	67.0–67.6 (67.3)	61.2–61.6	5.7–6.1
<i>Ile491Phe</i>	10	ATC→TTC	ND	ND	ND	ND

<sup>a</sup>Capturing probes, wild-type melt peak temperature ( $T_m$ ) ranges and means, mutant  $T_m$  ranges, and absolute values of melting temperature shift ( $\Delta T_m$ ) ranges associated with specific *rpoB* mutations in the strains tested and the corresponding nucleotide changes. ND, strains that harbored corresponding mutations outside the RRDR yielded a “RIF Resistance Not Detected” result. \*, rifampin resistance-determining region (RRDR) mutation unambiguously identified by unique combinations of Ultra probes and  $\Delta T_m$ s, including disputed ones (in italics). <sup>#</sup>, rifampin susceptible according to phenotypic testing.

<sup>b</sup>For double mutants, the high-confidence RR-conferring mutations are underlined (6, 7).

**SUPPLEMENTAL MATERIAL**

Supplemental material for this article may be found at <https://doi.org/10.1128/JCM.00686-18>.

**SUPPLEMENTAL FILE 1**, PDF file, 0.2 MB.

**ACKNOWLEDGMENT**

This work was supported by Erasmus Mundus Joint Doctorate Fellowship grant 2016-1346 to K.C.S.N.

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