Airway Mucin 2 Is Decreased in Patients with Severe Chronic Obstructive Pulmonary Disease with Bacterial Colonization

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Abstract

Rationale: Mucins are essential for airway defense against bacteria. We hypothesized that abnormal secreted airway mucin levels would be associated with bacterial colonization in patients with severe chronic obstructive pulmonary disease (COPD)

Objectives: To investigate the relationship between mucin levels and the presence of potentially pathogenic micro-organisms in the airways of stable patients with severe COPD

Methods: Clinically stable patients with severe COPD were examined prospectively. All patients underwent a computerized tomography scan, lung function tests, induced sputum collection, and bronchoscopy with bronchoalveolar lavage (BAL) and protected specimen brush. Patients with bronchiectasis were excluded. Secreted mucins (MUC2, MUC5AC, and MUC5B) and inflammatory markers were assessed in BAL and sputum by ELISA.

Measurements and Main Results: We enrolled 45 patients, with mean age (\pm SD) of 67 (\pm 8) years and mean FEV₁ of 41 (\pm 10) % predicted. A total of 31% (n = 14) of patients had potentially pathogenic micro-organisms in quantitative bacterial cultures of samples obtained by protected specimen brush. Patients with COPD with positive cultures had lower levels of MUC2 both in BAL (P = 0.02) and in sputum (P = 0.01). No differences in MUC5B or MUC5AC levels were observed among the groups. Lower MUC2 levels were correlated with lower FEV₁ (r = 0.32, P = 0.04) and higher sputum IL-6 (r = -0.40, P = 0.01).

Conclusions: Airway MUC2 levels are decreased in patients with severe COPD colonized by potentially pathogenic micro-organisms. These findings may indicate one of the mechanisms underlying airway colonization in patients with severe COPD.

Clinical trial registered with www.clinicaltrials.gov (NCT01976117).

Keywords: chronic obstructive pulmonary disease; mucins; airway infection; airway inflammation.

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Chronic obstructive pulmonary disease (COPD) is a leading cause of morbidity, mortality, and resource use worldwide (1). Potentially pathogenic micro-organisms (PPMs) can be isolated from airway secretions in about 30–50% of clinically stable patients with severe COPD (2, 3). This bacterial colonization is associated with increased neutrophil-mediated airway inflammation (4, 5) and more frequent and severe episodes of acute exacerbation of COPD (6). Therefore, airway bacterial colonization plays an important negative role in the clinical course of the disease (7, 8).

Pathogenesis of airway bacterial colonization in COPD is poorly understood. One of the features of COPD is chronic hypersecretion of mucus, which is related to increased risk of airway infection (9, 10). Mucus is composed of water, salt, and proteins. The major macromolecular components of mucus are proteins called mucins (11, 12). Experimental studies have demonstrated that mucin secretion is required for defense against bacterial infections, linking mucin deficiency with chronic airway infections (13).

Several mucins have been described in the lower respiratory tract (14, 15), although mucin (MUC) 5AC, MUC5B, and MUC2 are the major secreted mucins detected in the airways from healthy individuals (16, 17). In moderate COPD, increases of MUC5AC and MUC5B have been detected compared with nonsmokers and smokers without airway obstruction (18, 19), although these findings have not been related to airway infection. In non-cystic fibrosis (CF) bronchiectasis, elevated MUC2 levels were found to be related to the presence of Pseudomonas aeruginosa and severe disease (20). However, no data regarding the role of mucins and its relationship with airway bacterial infection in COPD are available.

We postulated that mucin levels are protective against airway infection in COPD. Therefore, the aim of our study was to assess the association of airway mucin levels and airway bacterial colonization among patients with severe stable COPD.

Methods

Study Design and Ethics

This was a prospective cross-sectional study that included clinically stable patients with

severe COPD with and without airway bacterial colonization (n = 14 and n = 31, respectively). The institutional review board (IIBSP-MUC-200920) approved the study protocol, and all subjects gave signed, informed consent to participate in the study.

Participants

Patients were consecutively recruited from a specialist clinic at the Hospital de la Santa Creu i Sant Pau (Barcelona, Spain). The diagnosis of COPD was established according to the GOLD (Global Initiative for Chronic Obstructive Lung Disease) recommendations (1), and included patients with GOLD stages C and D. All participants were clinically stable, as defined by the absence of an exacerbation that required antibiotic or steroid treatment within 30 days before inclusion. All patients underwent a computerized tomography scan, and those with bronchiectasis, lung cancer, pneumonia, and/or interstitial lung diseases were excluded. Patients with active malignant disease and/or any type of immunosuppression were also excluded.

Clinical and Functional Characterization

Demographic data, number of exacerbations in the previous year, time from last exacerbation, relevant comorbid conditions, and current treatments were recorded at inclusion using standardized questionnaires. Spirometry (Datospir-600; Sibelmed SA, Barcelona, Spain) was performed according to the Spanish Respiratory Society guidelines (21), using the predicted values for Mediterranean populations (22).

Microbiological Evaluation

The presence of airway colonization was determined by quantitative microbiological cultures obtained using a protected specimen brush (PSB). PSB samples were obtained from right intermedius bronchus using a flexible bronchoscope and processed using standard methodology. Samples were processed for qualitative and quantitative bacteriology, as previously described (23). Bacterial load was considered significant when it reached 10² colony-forming units/ml or greater (24).

Mucins Measurement

Bronchoalveolar lavage (BAL) samples were recovered using 150-ml saline lavage

with the bronchoscope wedged in the right middle lobe. Induced sputum was collected as previously described (25). Induced sputum was collected just before the bronchoscopy in all patients. BAL and induced sputum were centrifuged at $2,000 \times g$ for 10 minutes at 10°C. Proteases inhibitors (Calbiochem, San Diego, CA) were added to the samples during thawing.

MUC2, MUC5AC, and MUC5B were measured by validated, commercially available ELISA kits (USCN Life Science Inc., Wuhan, China), as we previously described (20)

Inflammatory Markers

IL-6 was measured in sputum by ELISA (R&D Systems, Abington, UK). $\alpha 1$ defensin was also measured in BAL and sputum by ELISA (BlueGene, Shangai, China)

Statistical Analyses

Statistical analysis was performed using the SPSS 17.0 software program (SPSS Inc., Chicago, IL). Results are presented as mean (\pm SD or SEM, as indicated) and frequency or percentage, as required. Continuous variables were analyzed using Student's *t* test and ANOVA, whereas categorical variables were analyzed using χ^2 tests. Mucin levels, airway inflammation, bacterial load, smoking status, tobacco habit (packs/year), and lung function were correlated by linear regression. Nonparametric tests were used when necessary. A *P* value of less than 0.05 was considered significant.

Results

A total of 45 patients with stable severe COPD were included in the study. Median age (\pm SD) was 67 (\pm 8) years, and median FEV₁ was 41 (\pm 10) % predicted; 37 of them (82%) were male, and 16 (35%) were current smokers. A total of 14 (31%) patients with stable, severe COPD had PPMs determined by PSB and were considered colonized.

Patient Characteristics

Table 1 shows the characteristics of the subjects, grouped according to the presence or absence of airway bacterial colonization. There were no statistically significant differences in sex, age, smoking status, lung function tests, pre-existing comorbid **Table 1.** Patient demographics, clinical characteristics, comorbid conditions, and prior treatments among colonized and noncolonized patients with chronic obstructive pulmonary disease

Variables	Colonized (<i>n</i> = 14)	Noncolonized (n = 31)	<i>P</i> Value
Age, yr Male/female, <i>n</i> BMI, kg/m ² Comorbid conditions	68.7 ± 10.8 11/3 25.1 ± 5.0	$\begin{array}{c} 66.4 \pm 7.3 \\ 26/5 \\ 26.0 \pm 3.9 \end{array}$	0.07 0.6 0.3
Smoker/ ex-smoker, n Pack-years Chronic cardiac disease, n (%) Prior malignancy, n (%) Chronic kidney disease, n (%) Diabetes mellitus, n (%) Hypertension, n (%) Depression, n (%) Stroke, n (%)	$5/9$ 59.4 ± 24.8 $3 (21.4)$ $2 (14.3)$ 0 $4 (28.6)$ $8 (57.1)$ $2 (14.3)$ $1 (7.1)$	$\begin{array}{c} 11/20\\ 52.0\pm17.5\\ 9\ (29.0)\\ 5\ (16.1)\\ 2\ (6.5)\\ 4\ (12.9)\\ 17\ (54.8)\\ 9\ (29.0)\\ 1\ (3.2)\end{array}$	0.6 0.3 0.5 0.8 0.3 0.2 0.8 0.2 0.5
Functional status FEV ₁ , L FEV ₁ , % Frequent exacerbator, <i>n</i> (%)* Time from last exacerbation, wk	$\begin{array}{c} 1.1 \pm 0.31 \\ 37.2 \pm 7.8 \\ 7 \ (50.0) \\ 12.7 \pm 4.7 \end{array}$	$\begin{array}{c} 1.3 \pm 0.44 \\ 43.1 \pm 10.7 \\ 5 \ (16.1) \\ 21.8 \pm 14.4 \end{array}$	0.1 0.2 0.02 0.03
Medications, <i>n</i> (%) ICS LABA LAMA Roflumilast	12 (85.7) 14 (100) 12 (85.7) 1 (7.1)	30 (96.8) 30 (96.8) 27 (87.0) 2 (6.5)	0.1 0.4 0.8 0.9

Definition of abbreviations: BMI = body mass index; ICS = inhaled corticosteroids; $LABA = long-acting \beta$ -agonist; LAMA = long-acting muscarinic antagonist.

Data are presented as n (%) unless otherwise indicated.

*Frequent exacerbator (>two acute exacerbations in the last 12 mo).

conditions, or prior medications used among groups. Patients with COPD with bacterial colonization had a higher rate of frequent exacerbations (P = 0.02) and shorter time from last exacerbation (P = 0.03)

Microbiology

Table 2 shows the identified PPMs and the bacterial load in PSB specimens. The most common pathogen collected from these patients with stable severe COPD were Haemophilus influenzae (n = 8), followed by Streptococcus pneumoniae (n = 2), Moraxella catarrhalis (n = 2), Neisseria meningitidis (n = 1), and Escherichia coli (n = 1), respectively. All colonized patients had only one species of PPMs in their PSB samples.

Bronchoalveolar Lavage Mucin Levels

MUC2 was the secreted mucin with the highest expression in the BAL, with a

Table 2. Potentially pathogenic micro-organisms and bacterial load isolated in protected specimen brush samples

Potentially Pathogenic Micro-organisms	n (%)*	≥10 ² CFU/ml <i>n</i> (%)	≥10 ³ CFU/ml <i>n</i> (%)	≥10 ⁴ CFU/ml <i>n</i> (%)
Haemophilus influenzae Streptococcus pneumoniae Moraxella catarrhalis Neisseria meningitidis Escherichia coli	8 (57) 2 (14) 2 (14)	2 (14)	7 (88) 1 (50) 2 (100) 1 (100)	1 (50)
	()			1 (100)

Definition of abbreviation: CFU = colony-forming units.

*% refers to the total number of micro-organisms.

mean (SD) of 8.1 (\pm 4.4) ng/ml. MUC5AC levels were 80.3 (\pm 11.2) pg/ml and MUC5B levels were 6.4 (\pm 4.0) ng/ml.

Stable patients with severe COPD and airway bacterial colonization had lower BAL MUC2 levels compared with those with no airway bacterial colonization ($6.3 \pm$ 1.3 vs. 8.9 ± 4.9 ng/ml, P = 0.02). No differences related to airway bacterial colonization status were observed for BAL MUC5AC levels (80.3 ± 4.3 vs. $80.3 \pm$ 13.0 pg/ml, P = 0.9) and BAL MUC5B levels (8.8 ± 10.8 vs. 4.2 ± 3.0 ng/ml, P = 0.1), respectively (Figure 1). No differences among BAL mucin levels and type of PPMs were found.

Sputum Mucin Levels

MUC2 was also the secreted mucin with highest expression in the sputum, with a mean (SD) of 52.4 (±32.8) ng/ml, followed by MUC5B (12.2 ng/ml [±14.9]) and MUC5AC (674.2 pg/ml [±179.1]), respectively. Similar to BAL results, only low sputum MUC2 levels were associated with airway bacterial colonization among stable patients with severe COPD (34.7 \pm 28.0 vs. 59.9 ± 30.0 ng/ml, P = 0.01). There were no differences in sputum MUC5AC levels $(304.3 \pm 114.4 \text{ vs. } 360.1 \pm 159.1 \text{ pg/ml},$ P = 0.5) and sputum MUC5B levels (14.2 ± 10.5 vs. 11.1 ± 6.9 ng/ml, P = 0.5) among colonized and noncolonized patients with stable severe COPD (Figure 2). There were no differences among sputum mucin levels and type of PPM.

A positive correlation was identified among MUC2 (r = 0.34, P = 0.03) and MUC5AC (r = 0.14, P = 0.02) sputum and BAL mucin samples from stable patients with severe COPD.

Inflammatory Markers

Sputum IL-6 and α 1 defensin levels were higher in colonized patients compared with noncolonized patients (1,948.5 ± 476.7 vs. 389.1 ± 127.2 ng/ml, *P* < 0.001 and 1,244.4 ± 250.2 vs. 498.8 ± 153.6 pg/ml, *P* = 0.01, respectively). BAL α 1 defensin levels were also higher in colonized patients, although differences were not statistically significant (143.2 ± 33.2 vs. 101.1 ± 12.2 pg/ml, *P* = 0.2).

Relationship between Mucin Levels and Lung Function and Airway Inflammation

A direct relationship among sputum MUC2 levels and FEV₁ % predicted was found

ORIGINAL RESEARCH

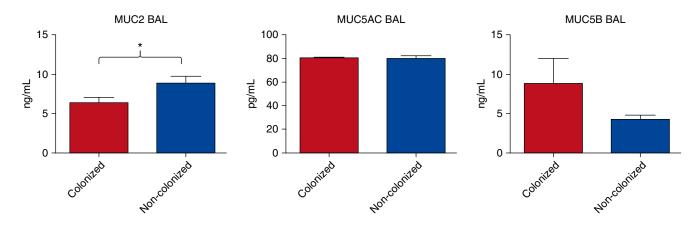


Figure 1. Mucin (MUC) 2, MUC5AC, and MUC5B bronchoalveolar lavage (BAL) levels in patients with severe chronic obstructive pulmonary disease (COPD) colonized and noncolonized by potentially pathogenic micro-organisms. Data are presented as median (±SEM). *P < 0.05.

(r = 0.355, P = 0.04; Figure 3A). In addition, there was a significant negative correlation between levels of MUC2 and IL-6 levels in sputum (r = -0.40, P = 0.01; Figure 3B). We found no other significant correlations among other mucin levels and inflammatory markers, including α 1 defensin. No correlation between mucin levels and airway bacterial load or smoking status was found.

Discussion

Low levels of airway MUC2 were associated with bacterial colonization and the presence of increased airway inflammation in patients with severe COPD. Although, in a crosssectional study, it is not possible to determine whether decreased MUC2 is a cause or a consequence of airway infection, in view of the experimental data published by Roy and colleagues (13) and others, these findings suggest a specific role for MUC2 in the pathogenesis of airway bacterial colonization in severe COPD. No differences in other airway-secreted mucins were found.

Mucins are proteins produced by respiratory epithelial cells essential for appropriate airway mucus formation (26, 27). Patients with COPD, especially those with chronic bronchitis, have increased mucus secretion (28). This mucus hypersecretion has been associated with an accelerated lung function decline and increased risk of exacerbations (9, 10, 29). Although it has been speculated that mucus hypersecretion is important in the pathogenesis of COPD, there are few data evaluating mucus properties in these patients. In our study, MUC2 was the predominant airway-secreted mucin both in BAL and in the sputum from patients with severe COPD, followed by MUC5B and MUC5AC.

Previous studies in asthma (16), CF (30–32), and chronic bronchitis (33) demonstrated higher levels of MUC5AC and MUC5B than MUC2. In COPD, Kirkham and colleagues (19) demonstrated that MUC5B was the major mucin in sputum in moderate patients with COPD as compared with smokers without airway obstruction. In this study, sputum with increased amount of MUC5B correlated with lower FEV₁. In our study, lower sputum MUC2 values correlate with lower FEV₁. Caramori and colleagues (18) demonstrated that MUC5AC expression was increased in bronchial submucosal glands of patients with stable moderate COPD. Recently,

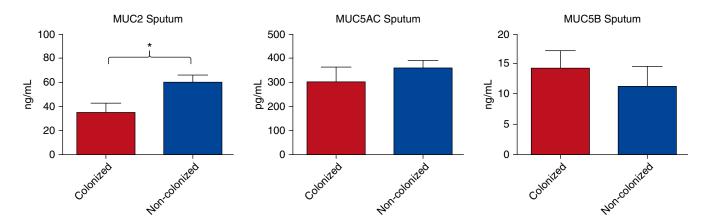


Figure 2. Mucin (MUC) 2, MUC5AC, and MUC5B sputum levels in patients with severe chronic obstructive pulmonary disease (COPD) colonized and noncolonized by potentially pathogenic micro-organisms. Data are presented as median (\pm SEM). *P < 0.05.

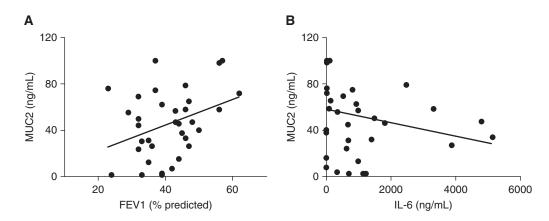


Figure 3. Relationship between (A) mucin (MUC) 2 sputum levels and FEV₁ % predicted and (B) sputum IL-6 levels.

Chillappagari and colleagues (34) showed higher levels of MUC5AC and MUC5B in sputum of subjects with moderate to severe COPD during a periexacerbation period compared with control subjects.

All of these findings suggest that the expression of airway-secreted mucins are altered in COPD and may be related to disease severity (especially to lung function) and clinical situation (stability or exacerbation). In addition, as has previously been emphasized (35), COPD has a different pathogenesis and different airway inflammatory profile to other chronic airway diseases. Specific roles of mucin subtypes and the impact of external stimuli (such as airway infection) in different airway diseases will require further study.

Bacterial colonization plays an important role in the pathogenesis and course of COPD (36). Chronic airway infection increases airway inflammation and leads to further impairment of local host defense (4, 5, 37). Several studies have demonstrated that patients with COPD with airway colonization by PPMs had worse clinical outcomes (6, 10).

The reasons why some patients with COPD become colonized are not well established. Mucins have been postulated as natural antimicrobial agents (17). In the gastrointestinal tract, mucins have demonstrated an antibiotic function against *Helicobacter pylori* (38), but few data are available regarding the relationship between mucin expression and airway infection. Recent experimental studies have demonstrated the crucial role of MUC5B as an airway defense mechanism (13).

In idiopathic pulmonary fibrosis, a polymorphism of the MUC5B gene has been associated with increased airway bacterial burden and risk of death (39). In patients with non-CF bronchiectasis, a recent study showed that stable patients with airway bacterial colonization had higher levels of sputum MUC2, especially those colonized with P. aeruginosa (20). In our study, we found that patients colonized by PPMs, primarily by H. influenzae, expressed lower levels of airway MUC2 in the airway from BAL and sputum samples compared with patients without airway bacterial colonization. These findings may suggest a specific role for MUC2 in the response to bacterial infection that may be airway disease specific and modulated by specific bacterial pathogens. In COPD, the presence of MUC2 might be protective against bacterial colonization, mainly due to H. influenzae.

Factors that influence mucin secretion may be a key to understanding these findings. Different studies have demonstrated that mucin secretion is regulated in relation to several factors, such as bacterial products (39) and inflammatory cytokines (40). MUC2 gene is up-regulated *in vitro* by inflammatory mediators present in the airway secretions of patients with chronic lung disease (12). However, using human bronchial epithelial cells, Fujisawa and colleagues (41) demonstrated that mucin expression is stimulated by inflammatory cytokines in a time- and dose-dependent manner. These authors showed that the persistence of inflammatory stimulus over time or the presence of excessive inflammation (both

features present in most patients with severe COPD) inhibits mucin induction (41). These findings are consistent with our study, in which we found that higher airway inflammation (detected by IL-6 levels) in patients with COPD were correlated with lower MUC2 levels. This observation may have important biological consequences, suggesting that those patients with severe COPD with elevated and persistent airway inflammation are those with lower MUC2 levels. No relationship among mucin levels and other factors that potentially may influence mucin secretion, such as airway bacterial load or current smoking, were found. Further studies are needed to interpret this complex pathway, which may contribute to a better understanding of the role of secreted mucins as natural antimicrobial agents in the airways of patients with COPD.

The strengths of our study include the use of gold standard bronchoscopic PPM identification with PSB quantitative microbiological cultures (24). In addition, the careful selection of stable patients with severe COPD without bronchiectasis or other structural lung diseases, as determined by computerized tomography scan, strengthen the validity of the results.

Our study also has limitations. First, although our sample size was larger than those previously reported in other studies of airway mucins in patients with COPD (16, 19), asthma (16), and CF (30–32), it was still small enough to limit the strength and generalizability of the results. Second, a control group was not included, and it might have informed the normal expression of mucin levels in patients without COPD.

Third, we did not assess several other factors that may influence mucin secretion, such as specific microbiological properties, pattern recognition receptors, airway cell counts, or neutrophil activity. Fourth, previous studies have suggested concerns about accurate measurements of MUC2 (12), indicating that some results should be considered with caution. Finally, although the differences we observed were clearly statistically significant, we do not know enough about the biology of specific mucin subtypes in the airway to determine the minimum clinically important difference in airway mucin levels. Further studies correlating mucin levels with clinical outcomes are needed.

In conclusion, we found that airway MUC2 levels are decreased in patients with severe COPD with bacterial colonization, which is related to higher airway inflammation. These observations may suggest that secreted mucins play a role in determining the presence of airway bacterial colonization in stable severe COPD. This justifies a larger, more detailed investigation of the role of mucins in airway defense in patients with COPD.

<u>Author disclosures</u> are available with the text of this article at www.atsjournals.org.

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