

Article

# Establishing Reliable Cu-64 Production Process: From Target Plating to Molecular Specific Tumor Micro-PET Imaging

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**Abstract:** Copper-64 is a useful radioisotope for positron emission tomography (PET). Due to the wide range of applications, the demand of <sup>64</sup>Cu with low metallic impurities is increasing. Here we report a simple method for the efficient production of high specific activity <sup>64</sup>Cu using a cyclotron for biomedical application. We designed new equipment based on the plating of enriched <sup>64</sup>Ni as the target, and used automated ion exchange chromatography to purify copper-64 efficiently after irradiation and dissolution of the target in good radiochemical and chemical yield and purity. The <sup>64</sup>Cu radionuclide produced using 99.32% enriched <sup>64</sup>Ni with a density of  $61.4 \pm 5.0$  mg/cm<sup>2</sup>, reaching a total radioactivity greater than 200 mCi, with specific activity up to 5.6 GBq/μmol. It was further incorporated into modified monoclonal antibody DOTA-rituximab to synthesize <sup>64</sup>Cu-DOTA-rituximab, which was used successfully for micro-PET imaging.

**Keywords:** copper-64; solid target; Rituximab; positron emission tomography (PET)

## 1. Introduction

Copper-64 (<sup>64</sup>Cu) is an attractive radionuclide of considerable interest for positron emission tomography (PET) imaging and radiotherapy due to its intrinsic physical and chemical properties. It has high spatial resolution comparable to <sup>18</sup>F radionuclide, with comparable average free travel distance for their generated positrons ( $R_{ave.}(\beta^+) = 0.70$  and  $0.69$  mm, respectively), due to their comparable positron energy ( $0.656$  MeV and  $0.635$  MeV, respectively) [1,2]. It has a relatively long half-life of 12.7 h, compared with fluorine-18 ( $t_{1/2} = 110$  min) and carbon-11 ( $t_{1/2} = 20.4$  min). In addition, <sup>64</sup>Cu also emits  $\beta^-$  and Auger electrons, enabling it to be useful for both PET imaging and radiotherapy. Moreover, the versatile coordination chemistry of <sup>64</sup>Cu allows for its reaction with a wide variety of chelator systems, such as DOTA, NOTA, TETA and CB-TE2A, that can be linked to antibodies, peptides and nanoparticles [3]. The <sup>64</sup>Cu radioisotope can be used for the design and synthesis of a wide range of radio-probes, providing attractive candidates for PET imaging.

A number of radiotracers involving <sup>64</sup>Cu as radionuclide have been applied in nuclear medicine as a means of studying their PET imaging [4–6]. Incorporation of <sup>64</sup>Cu into diacetyl-bis(*N*<sup>4</sup>-methylthiosemicarbazone) (ATSM) ligand was used for PET hypoxia imaging,

such as in head and neck cancer, and cardiac conditions [7,8]. More radiotracers are in clinical development [9–12], especially those with peptides and antibodies. For example,  $^{64}\text{Cu}$ -DOTA-trastuzumab was used to conduct PET imaging of HER2-positive lesions in patients with primary and metastatic breast cancer [13]. The PET image of  $^{64}\text{Cu}$ -DOTATATE provided superior image quality, and detected more lesions than  $^{111}\text{In}$ -DTPA-octreotide [14]. Grubmüller, B. et al. investigated the diagnostic potential of  $^{64}\text{Cu}$ -PSMA-617 in patients with prostate adenocarcinoma [15].

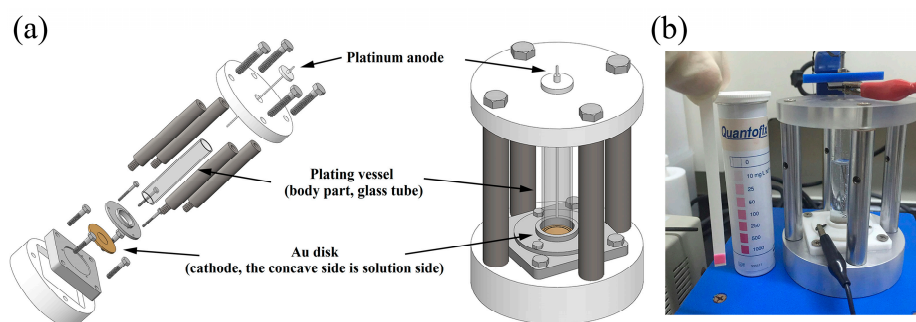
$^{64}\text{Cu}$  has been produced at many centers [16–19]. Among the nuclear reactions examined, the  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$  method is the best and widely used, since high production yield of the  $^{64}\text{Cu}$  can be obtained with low energy protons in this route [20,21]. At Washington University, an effective method was investigated to produce high specific activity  $^{64}\text{Cu}$  on a small biomedical cyclotron using the  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$  nuclear reaction, and  $^{64}\text{Cu}$  has been produced for more than 17 years by the irradiation of electroplated enriched  $^{64}\text{Ni}$  targets in this center [20,22]. The Turku PET Centre has been producing  $^{64}\text{Cu}$  since 2008 using  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$  reaction, and also handles the irradiated target, radioactive liquids and gases using automated equipment [23,24]. At the University of Wisconsin,  $^{64}\text{Cu}$  and  $^{61}\text{Co}$  radionuclides have been simultaneously produced using the  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$  nuclear reaction on a low energy proton-only cyclotron [25]. Ohya, T. et al. (2016) produced high-quality  $^{64}\text{Cu}$  for routine use via an electrodeposited  $^{64}\text{Ni}$  target, and successfully reduced the metallic impurities level of the  $^{64}\text{Cu}$  product, such as Co and Ni [21]. Other nuclear reactions examined include  $^{64}\text{Ni}$  (d, 2n)  $^{64}\text{Cu}$ ,  $^{64}\text{Zn}$  (d, 2p)  $^{64}\text{Cu}$ ,  $^{64}\text{Zn}$  (n, p)  $^{64}\text{Cu}$  [26,27]. In China, researchers also have a growing interest in  $^{64}\text{Cu}$ , and the demand of no-carrier added  $^{64}\text{Cu}$  has started to increase.

Here we report a robust, reliable and user-friendly plating vessel, which can be used for the effective preparation of the  $^{64}\text{Ni}$  solid target. The production of  $^{64}\text{Cu}$  was performed on a Sumitomo HM-20 biomedical cyclotron (20 MeV) via the  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$  reaction. The  $\gamma$ -ray spectroscopy of the produced  $^{64}\text{Cu}$  solution was also measured to evaluate the radionuclide impurities. After  $^{64}\text{Cu}$  was purified, a labeling experiment to synthesize  $^{64}\text{Cu}$ -DOTA-Rituximab was performed to examine the quality of  $^{64}\text{Cu}$ , including labeling yield and radiochemical purity of the radiotracer, which targets the CD20 antigen, which is expressed on B cell lymphocytes and in the majority of non-Hodgkin's lymphoma (NHL) [28]. The synthesized radiotracer was further examined by micro-PET imaging using SCID (severe combined immune deficiency) mice bearing Ramos RA1 tumors which overexpress CD20 antigen.

## 2. Results and Discussion

### 2.1. Preparation of $^{64}\text{Ni}$ Target

In order to make  $^{64}\text{Cu}$  via the nuclear reaction of  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$ , we made the enriched  $^{64}\text{Ni}$  targets on a gold (Au) disk by electrodeposition of  $^{64}\text{Ni}$  from an aqueous solution of  $\text{Ni}(\text{NH}_3)_6^{2+}$  at pH = 9.05, using a robust, reliable and user-friendly apparatus (Figure 1). The Au disk (30.8 mm diameter  $\times$  1.5 mm thickness) was used as a cathode, and a platinum wire was used as an anode. A cavity of 12.1 mm diameter and 0.2 mm depth was milled into the Au disk and  $^{64}\text{Ni}$  (99.32%) was plated into this cavity. The  $^{64}\text{Ni}$  electroplating was performed with a constant current of 15–25 mA at 2.4–2.6 V in the aqueous solution for 48–72 h. During the electrodeposition, the green color of the  $^{64}\text{Ni}$  plating solution gradually faded away and the bubbling of  $\text{H}_2$  gas was clearly visible. When the  $^{64}\text{Ni}$  plating solution became colorless, the electrodeposition was finished. The absence of  $\text{Ni}^{2+}$  was also confirmed by analytical test strips. After cleaning and drying, the plated  $^{64}\text{Ni}$  on Au-disk was  $70.6 \pm 5.8$  mg and the density of the plated  $^{64}\text{Ni}$  was  $61.4 \pm 5.0$  mg/cm<sup>2</sup>, assuming a uniform thickness on the disk.



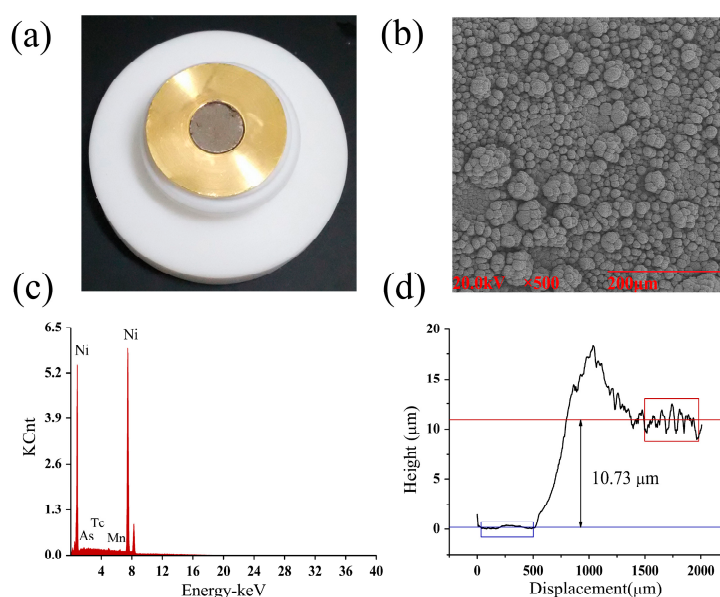
**Figure 1.** The  $^{64}\text{Ni}$  plating vessel. (a) Illustration of new  $^{64}\text{Ni}$  plating vessel by scheme; (b) Picture of the actual electric plating unit.

## 2.2. Quality Control of $^{64}\text{Ni}$ Target

The quality of the  $^{64}\text{Ni}$  solid target prepared above was evaluated by a number of physical techniques, to determine the uniformity, to characterize the metallic impurities, and to measure the thickness of  $^{64}\text{Ni}$  layer on Au-disk. The SEM (scanning electron microscopy) image (Figure 2b) of the  $^{64}\text{Ni}$  solid target showed the uniform layer of Ni on Au surface. The EDS (energy dispersive X-ray spectroscopy) (Table 1 and Figure 2c) results showed no significant amount of metallic impurities in the  $^{64}\text{Ni}$  layer. The  $^{64}\text{Ni}$  target thickness on Au-disk was measured to be  $10.73\ \mu\text{m}$  by alpha step apparatus (Figure 2d).

**Table 1.** The metallic impurities in the plated  $^{64}\text{Ni}$  target, analyzed by EDS.

Element	Wt %	At %
AsL	00.00	00.00
TcL	00.15	00.09
MnK	00.07	00.07
NiK	99.78	99.84



**Figure 2.** The  $^{64}\text{Ni}$  target produced in this study. (a) Photo of the  $^{64}\text{Ni}$  target; (b) The SEM image of the  $^{64}\text{Ni}$  target; (c) The EDS spectrum of the  $^{64}\text{Ni}$  target; (d) The thickness measurement of the  $^{64}\text{Ni}$  solid target.

### 2.3. Preparation of $^{64}\text{Cu}$

After irradiation of 5 h, the  $^{64}\text{Ni}$  target was dissolved in 6 M hydrochloride acid, and then the solution was load to an anion exchange column to separate into different components. The  $^{64}\text{Ni}$  was washed out with 6 M HCl and collected for recycling. Due to the elevated cost of enriched  $^{64}\text{Ni}$ , recycling of the target material for re-use could reduce the production cost of  $^{64}\text{Cu}$ , without sacrificing the quality of subsequent  $^{64}\text{Cu}$  production. When the eluted was switched to 1 M HCl, the first band coming out was co-produced cobalt radioisotopes (approximately 1 mL), and the second was the  $^{64}\text{Cu}$ , which was collected and evaporated to dryness. The residue was dissolved in 0.1 M HCl for further use. The separation process of  $^{64}\text{Cu}$  took about 2.5 h after irradiation.

### 2.4. Quality of $^{64}\text{Cu}$

The quality of  $^{64}\text{Cu}$  produced was evaluated by analysis of its metallic impurities and measurement of half-life of its radioactivity. The inductively coupled plasma-mass spectrometry (ICP-MS) analysis was used to evaluate the amount of metallic impurities in a decayed  $^{64}\text{Cu}$  solution, and showed a concentration of 6.339 ppb of Ni (0.127  $\mu\text{g}$ ), 4.112 ppb of Cu (0.082  $\mu\text{g}$ ), 5.502 ppb of Zn (0.110  $\mu\text{g}$ ), 0.108 ppb of Fe (0.002  $\mu\text{g}$ ), 0.102 ppb of Co (0.002  $\mu\text{g}$ ), 0.669 ppb of Ga (0.013  $\mu\text{g}$ ). The gamma spectrum of the produced  $^{64}\text{CuCl}_2$  solution (Figure 3) showed that the  $^{64}\text{Cu}$  radionuclide purity was >99%. The half-life of the produced radioactivity was determined by the radioactivity measured at different time points into the following equations:

$$A = A_0 e^{-\lambda t}, \ln A = -\lambda t + \ln A_0, \ln \frac{A}{A_0} = -\lambda t \quad (1)$$

where A is the radioactivity of the  $^{64}\text{Cu}$  at time t,  $A_0$  is the radioactivity of the  $^{64}\text{Cu}$  at 0 h, and  $\lambda$  represents the constant. We obtained the following equation (Figure 4):

$$\ln A = -0.05456t + 5.438 \quad (2)$$

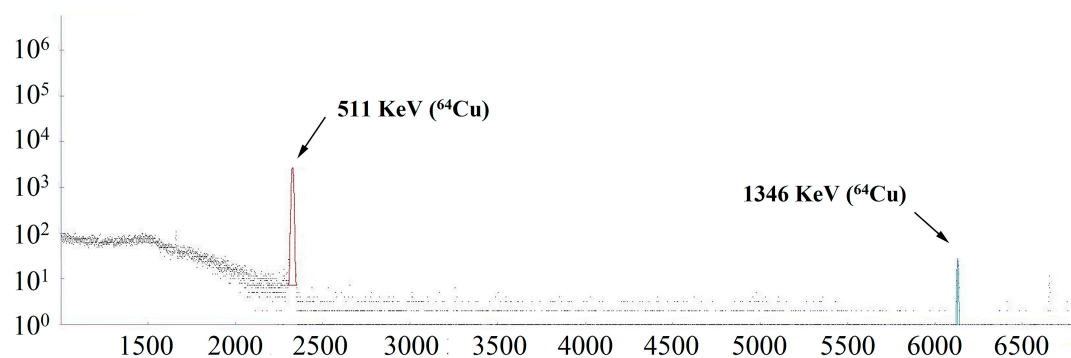
when  $t = T_{1/2}$ , the  $A = \frac{1}{2} A_0$

$$\ln \frac{1}{2} = -0.05456 T_{1/2} \quad (3)$$

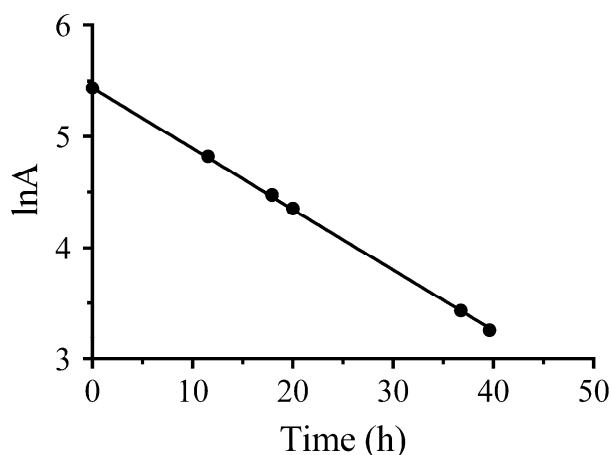
The half-life of the produced  $^{64}\text{Cu}$  was calculated:

$$T_{1/2} = \frac{\ln 2}{0.05456} = 12.704$$

The half-life calculated ( $T_{1/2} = 12.704$  h) of the produced  $^{64}\text{Cu}$  is in accordance with that of the radioisotope  $^{64}\text{Cu}$  ( $T_{1/2} = 12.7$  h).



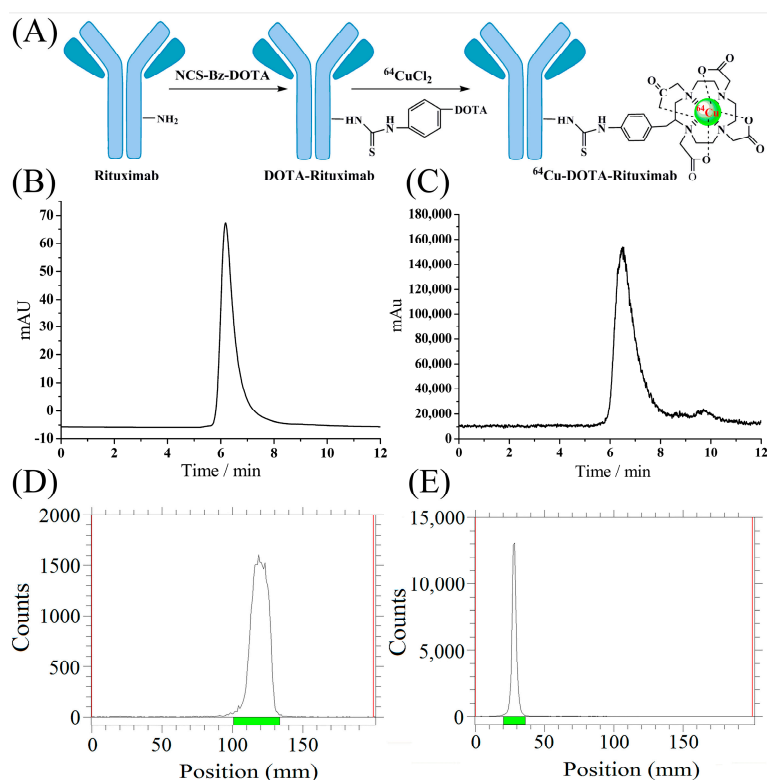
**Figure 3.** Gamma spectra of  $^{64}\text{CuCl}_2$  solution after purification.



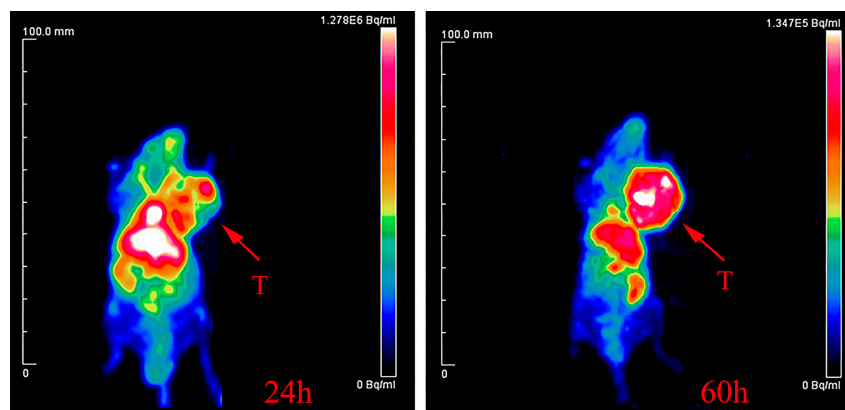
**Figure 4.** The radioactivity of the produced  $^{64}\text{Cu}$  measured at different time points vs. time, with fitted equation, on a logarithmic scale.

### 2.5. Radio-Synthesis of $^{64}\text{Cu}$ -DOTA-Rituximab and Micro-PET Imaging

To assess the quality and quantity of the produced  $^{64}\text{Cu}$ , we made  $^{64}\text{Cu}$ -DOTA-rituximab (Figure 5), with high chemical yield, high radiochemical purity, and high specific activity. Micro-PET imaging of the radiotracer in mice bearing Ramos RA1 tumors clearly showed the tumor at 24 h and 60 h post-injection, with excellent resolution and clarity at the latter (Figure 6).



**Figure 5.** Radio-synthesis of  $^{64}\text{Cu}$ -DOTA-rituximab. (A) Modification of Rituximab and radiolabeling by  $^{64}\text{Cu}$  radionuclide; (B) The Radio-HPLC (radioactive high performance liquid chromatography) chromatograph of rituximab; (C) The Radio-HPLC chromatograph of  $^{64}\text{Cu}$ -DOTA-rituximab after purification by PD-10 column; (D) The Radio-TLC (radioactive thin-layer chromatography) image of  $^{64}\text{CuCl}_2$ ; (E) The Radio-TLC image of  $^{64}\text{Cu}$ -DOTA-rituximab after purification by PD-10 column.



**Figure 6.** Micro-PET image of  $^{64}\text{Cu}$ -DOTA-rituximab in SCID mice bearing Ramos RA1 tumors at 24 h and 60 h post-intravenous injection. The arrows indicate the location of tumor.

### 3. Materials and Methods

#### 3.1. Materials and Reagents

High purity reagents were used for production of  $^{64}\text{Cu}$  in this study. Isotopically enriched  $^{64}\text{Ni}$  (99.32%  $^{64}\text{Ni}$ ; 0.13%  $^{58}\text{Ni}$ ; 0.07%  $^{60}\text{Ni}$ ; 0.01%  $^{61}\text{Ni}$ ; 0.47%  $^{62}\text{Ni}$ ) was from Isoflex Company (San Francisco, CA, USA);  $(\text{NH}_4)_2\text{SO}_4$  (99.999% metals basis) from Alfa Aesar (Ward Hill, MA, USA); concentrated HCl (99.999% trace metals basis),  $\text{HNO}_3$  (99.999% trace metals basis), and  $\text{NH}_4\text{OH}$  (99.99% trace metals basis) were purchased from Sigma-Aldrich (St. Louis, MO, USA); AG<sup>®</sup>1-X8 ion exchange resin was from Bio-Rad Laboratories (Hercules, CA, USA); platinum wire (99.997% metals basis) from Alfa Aesar (Ward Hill, MA, USA);  $\text{Ni}^{2+}$  analytical test strips from Qantofix (Sigma-Aldrich, St. Louis, MO, USA); disposable PD-10 desalting columns were from GE Healthcare (Piscataway, NJ, USA).

#### 3.2. Equipment

The alpha step apparatus (Alpha-step IQ, KLA-Tencor, Milpitas, CA, USA), scanning electron microscopy with energy dispersive X-ray spectroscopy (SEM-EDS) (1910FE, AMRAY, Pawtucket, RI, USA) were used to characterize the quality of the  $^{64}\text{Ni}$  solid target produced in this study. The irradiation experiments were performed using a Sumitomo HM-20 cyclotron (20 MeV, Sumitomo Heavy Industries, Ltd., Tokyo, Japan). The separation of  $^{64}\text{Cu}$  was performed using a  $^{64}\text{Cu}$  separation system (Industrial Equipment Division, Sumitomo Heavy Industries, Ltd., Tokyo, Japan). Inductively coupled plasma-mass spectrometry (ICP-MS) (ELEMENT XR mass spectrometer, Thermo Fisher, Bremen, Germany) was used to analyze the purity of the  $^{64}\text{Cu}$  sample. The Agilent Technologies 1200 series of high performance liquid chromatography (HPLC) system (Agilent, Lake Forest, CA, USA) equipped with both a UV absorption detector and a B-Fc-1000 HPLC radioactivity detector (Bioscan, Washington, DC, USA) and the radioactive thin-layer chromatography scanner (Radio-TLC) (Bioscan, IAR-2000, Washington, DC, USA) were used to analyze the radiochemical purity of tracers.

#### 3.3. Plating Solution

The  $^{64}\text{Ni}$  electroplating solution was prepared as reported previously with some modifications [25]. The enriched  $^{64}\text{Ni}$  metal (65–80 mg) was dissolved in 5 mL of warm 6 M  $\text{HNO}_3$ . After the metal was completely dissolved, the solution was then evaporated to dryness under vacuum. The green-colored residue was dissolved in 300  $\mu\text{L}$  of concentrated  $\text{H}_2\text{SO}_4$  and the solution then diluted with 2 mL of 18  $\text{M}\Omega\cdot\text{cm}$  water (Milli-Q Waters, Millipore Corporation, Billerica, MA, USA) slowly and carefully. The pH of the solution was adjusted to  $9.05 \pm 0.05$  by adding about 1.5 mL concentrated  $\text{NH}_4\text{OH}$ .

To this solution, ~300 mg of  $(\text{NH}_4)_2\text{SO}_4$  was added and the volume of the solution was adjusted to 5 mL with 18 M $\Omega$ ·cm water. The final solution was transferred to the electroplating cell for target plating.

### 3.4. Characterizations of Ni-64 Target

After electroplating, the  $^{64}\text{Ni}$  solid target was examined by measuring  $^{64}\text{Ni}$  thickness, composition and structure. The thickness was measured by alpha step apparatus, the composition was measured by scanning electron microscopy with energy dispersive X-ray spectroscopy (SEM-EDS), and the structure was characterized by scanning electron microscopy (SEM). Meanwhile, the identity of metallic impurities of the  $^{64}\text{Ni}$  target was further analyzed by energy dispersive X-ray spectroscopy (EDS).

### 3.5. Preparation of $^{64}\text{Ni}$ Target and Irradiation

The  $^{64}\text{Ni}$  targets were prepared by electrodeposition of the enriched  $^{64}\text{Ni}$  (99.32%) solution prepared as described above. Electroplating of  $^{64}\text{Ni}$  was realized using a plating vessel of our own design and manufacture (Figure 1), where the Au disk was used as a cathode and platinum wire as an anode. The electrodeposition of  $^{64}\text{Ni}$  was achieved at 2.4–2.6 V and 15–25 mA with the platinum anode at ~1 cm from the Au disk electrode. This process took 48–72 h. The enriched  $^{64}\text{Ni}$  target was irradiated at 12.5 MeV (which decreased by Al) with 20  $\mu\text{A}$  current on a Sumitomo HM-20 cyclotron (20 MeV) for about 5–7 h.  $^{64}\text{Cu}$  was produced from the  $^{64}\text{Ni}$  (p, n)  $^{64}\text{Cu}$  nuclear reaction.

### 3.6. Radiochemical Separation

After irradiation, the  $^{64}\text{Ni}$  target was placed into the dissolving bath of 6 M HCl. The complete dissolution of the target material took about 40 min under heating, and then the solution was loaded on an anion exchange column (AG 1-X8) pretreated with 18 M $\Omega$ ·cm water and 6 M HCl, sequentially. The enriched  $^{64}\text{Ni}$  was eluted with 6 M HCl and collected for recycling, and the  $^{64}\text{Cu}$  fraction was eluted with 1 M HCl, which was further evaporated to dryness. The residue was finally dissolved in 0.1 M HCl for further use. All of these procedures were performed using an automated system and carried out in a hot cell with remote control (Figure 7, Sumitomo Heavy Industries, Ltd., Tokyo, Japan).

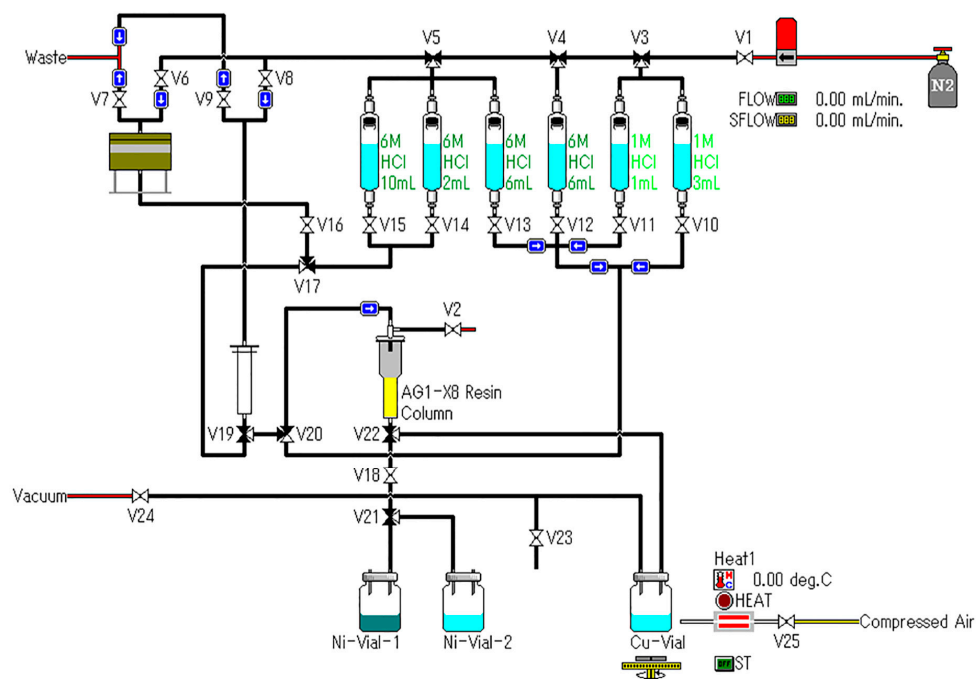


Figure 7. Schematic representation of the automated  $^{64}\text{Cu}$  separation system.

### 3.7. Radio-nuclide Analysis

The radionuclide identity and purity of the produced  $^{64}\text{CuCl}_2$  solution were measured using  $\gamma$ -ray spectroscopy (HTA Co., Ltd., Beijing, China). In addition, the identity of radioactivity of the produced  $^{64}\text{Cu}$  was further confirmed by measurement of its half-life, by measuring radioactivity at different time points. A decayed sample from the produced  $^{64}\text{Cu}$  was also analyzed by inductively coupled plasma-mass spectrometry (ICP-MS) for traces of metallic impurities.

### 3.8. Radiolabeling of DOTA-Rituximab and Micro-PET Imaging

To assess the quality of the produced  $^{64}\text{Cu}$  (quantity, specific activity and purity), the synthesis of a radiotracer,  $^{64}\text{Cu}$ -DOTA-rituximab, was performed. Here, the site-specific modification of monoclonal antibody DOTA-rituximab which contains two DOTA chelator in each antibody was used as we previously reported [29]. The  $^{64}\text{CuCl}_2$  solution prepared above was reacted with DOTA-rituximab in a solution of pH = 5.5 at RT for 30 min. After incubation, the radiotracer was purified using a PD-10 desalting column, and characterized by Radio-TLC and Radio-HPLC, to determine the labeling yield and radiochemical purity. The identity of the radiotracer,  $^{64}\text{Cu}$ -DOTA-rituximab, was further evaluated by PET imaging, which was carried out on a micro-PET rodent model scanner as reported earlier [29]. After formulation,  $^{64}\text{Cu}$ -DOTA-rituximab (0.5 mCi) was injected intravenously into the tail vein of the mice bearing Ramos RA1 tumors ( $n = 3$ ), which overexpress CD20 antigen, and the animals were imaged with micro-PET at both 24 and 60 h post-injection.

## 4. Conclusions

In this study, we presented the improved method for the preparation of  $^{64}\text{Cu}$ , especially the improved efficiency of electroplating of  $^{64}\text{Ni}$ . The  $^{64}\text{Ni}$  solid target on an Au-disk has a uniform surface, with the thickness of 10.73  $\mu\text{m}$ , and no metallic impurities. The total plated  $^{64}\text{Ni}$  was  $70.6 \pm 5.8$  mg and the density of the plated  $^{64}\text{Ni}$  was  $61.4 \pm 5.0$  mg/cm<sup>2</sup> (assuming a uniform thickness). After irradiation of the target and purification, the gamma spectrum of the produced  $^{64}\text{Cu}$  showed its radionuclide purity to be >99%, with both peaks at 511 keV and 1346 keV. In addition, the produced  $^{64}\text{CuCl}_2$  solution has high specific activity up to 5.6 GBq/ $\mu\text{mol}$ . It was further incorporated into the modified monoclonal antibody DOTA-rituximab to synthesize  $^{64}\text{Cu}$ -DOTA-rituximab, which was further used successfully for micro-PET imaging.

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**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Nayak, T.K.; Brechbiel, M.W. Radioimmunoimaging with longer-lived positron-emitting radionuclides: Potentials and challenges. *Bioconjug. Chem.* **2009**, *20*, 825–841. [[CrossRef](#)] [[PubMed](#)]
2. Holland, J.P.; Sheh, Y.; Lewis, J.S. Standardized methods for the production of high specific-activity zirconium-89. *Nucl. Med. Biol.* **2009**, *36*, 729–739. [[CrossRef](#)] [[PubMed](#)]
3. Wadas, T.J.; Wong, E.H.; Weisman, G.R.; Anderson, C.J. Copper chelation chemistry and its role in copper radiopharmaceuticals. *Curr. Pharm. Des.* **2007**, *13*, 3–16. [[CrossRef](#)] [[PubMed](#)]
4. Natarajan, A.; Arksey, N.; Iagaru, A.; Chin, F.T.; Gambhir, S.S. Validation of  $^{64}\text{Cu}$ -dota-rituximab injection preparation under good manufacturing practices: A Pet tracer for imaging of B-cell non-Hodgkin lymphoma. *Mol. Imaging* **2015**, *14*, 1–11.



5. Kurihara, H.; Hamada, A.; Yoshida, M.; Shimma, S.; Hashimoto, J.; Yonemori, K.; Tani, H.; Miyakita, Y.; Kanayama, Y.; Wada, Y.; et al.  $^{64}\text{Cu}$ -DOTA-trastuzumab PET imaging and HER2 specificity of brain metastases in HER2-positive breast cancer patients. *EJNMMI Res.* **2015**, *5*, 8. [[CrossRef](#)] [[PubMed](#)]
6. Sanghera, B.; Wood, K.; Sonoda, L.I.; Gogbashian, A.; Lowe, G.; Nunes, A.; Stirling, J.; Shepherd, C.; Beynon, G.; Wong, W.L. Pre-clinical positron emission tomography reconstruction algorithm effect on Cu-64 ATSM lesion hypoxia. *Mol. Imaging Radionucl. Ther.* **2016**, *25*, 19–25. [[CrossRef](#)] [[PubMed](#)]
7. Grassi, I.; Nanni, C.; Cicoria, G.; Blasi, C.; Bunkheila, F.; Lopci, E.; Colletti, P.M.; Rubello, D.; Fanti, S. Usefulness of  $^{64}\text{Cu}$ -ATSM in head and neck cancer: A preliminary prospective study. *Clin. Nucl. Med.* **2014**, *39*, 59–63. [[CrossRef](#)] [[PubMed](#)]
8. Handley, M.G.; Medina, R.A.; Mariotti, E.; Kenny, G.D.; Shaw, K.P.; Yan, R.; Eykyn, T.R.; Blower, P.J.; Southworth, R. Cardiac hypoxia imaging: Second-generation analogues of  $^{64}\text{Cu}$ -ATSM. *J. Nucl. Med.* **2014**, *55*, 488–494. [[CrossRef](#)] [[PubMed](#)]
9. Pfeifer, A.; Knigge, U.; Binderup, T.; Mortensen, J.; Oturai, P.; Loft, A.; Berthelsen, A.K.; Langer, S.W.; Rasmussen, P.; Elema, D.; et al.  $^{64}\text{Cu}$ -DOTATATE PET for neuroendocrine tumors: A prospective head-to-head comparison with  $^{111}\text{In}$ -DTPA-octreotide in 112 patients. *J. Nucl. Med.* **2015**, *56*, 847–854. [[CrossRef](#)] [[PubMed](#)]
10. Anderson, C.J.; Dehdashti, F.; Cutler, P.D.; Schwarz, S.W.; Laforest, R.; Bass, L.A.; Lewis, J.S.; McCarthy, D.W.  $^{64}\text{Cu}$ -Teta-octreotide as a pet imaging agent for patients with neuroendocrine tumors. *J. Nucl. Med.* **2001**, *42*, 213–221. [[PubMed](#)]
11. Chang, A.J.; Grigsby, P.W. Differences in uptake and distribution of  $^{64}\text{Cu}$ -Atsm and  $^{18}\text{F}$ -FDG in primary tumors of patients with squamous cell carcinoma of the cervix. *Fuel Energy Abstr.* **2010**, *78*, S144.
12. Chang, A.J. Intratumoral heterogeneity of  $^{64}\text{Cu}$ -ATSM uptake is a prognostic indicator in patients with cervical cancer. *Omics J. Radiol.* **2013**, *2*, 545–553. [[CrossRef](#)]
13. Tamura, K.; Kurihara, H.; Yonemori, K.; Tsuda, H.; Suzuki, J.; Kono, Y.; Honda, N.; Kodaira, M.; Yamamoto, H.; Yunokawa, M.; et al.  $^{64}\text{Cu}$ -DOTA-trastuzumab PET imaging in patients with HER2-positive breast cancer. *J. Nucl. Med.* **2013**, *54*, 1869–1875. [[CrossRef](#)] [[PubMed](#)]
14. Pfeifer, A.; Knigge, U.; Mortensen, J.; Oturai, P.; Berthelsen, A.K.; Loft, A.; Binderup, T.; Rasmussen, P.; Elema, D.; Klausen, T.L.; et al. Clinical PET of neuroendocrine tumors using  $^{64}\text{Cu}$ -DOTATATE: First-in-humans study. *J. Nucl. Med.* **2012**, *53*, 1207–1215. [[CrossRef](#)] [[PubMed](#)]
15. Grubmüller, B.; Baum, R.P.; Capasso, E.; Singh, A.; Ahmadi, Y.; Knoll, P.; Floth, A.; Righi, S.; Zandieh, S.; Meleddu, C.; et al.  $^{64}\text{Cu}$ -PSMA-617 PET/CT imaging of prostate adenocarcinoma: First in-human studies. *Cancer Biother. Radiopharm.* **2016**, *31*, 277–286. [[CrossRef](#)] [[PubMed](#)]
16. Matarrese, M.; Bedeschi, P.; Scardaoni, R.; Sudati, F.; Savi, A.; Pepe, A.; Masiello, V.; Todde, S.; Gianolli, L.; Messa, C.; et al. Automated production of copper radioisotopes and preparation of high specific activity [ $^{64}\text{Cu}$ ]Cu-ATSM for PET studies. *Appl. Radiat. Isot.* **2010**, *68*, 5–13. [[CrossRef](#)] [[PubMed](#)]
17. Burke, P.; Golovko, O.; Clark, J.C.; Aigbirhio, F.I. An automated method for regular productions of copper-64 for PET radiopharmaceuticals. *Inorg. Chim. Acta* **2010**, *363*, 1316–1319. [[CrossRef](#)]
18. Rajec, P.; Csiba, V.; Leporis, M.; Štefečka, M.; Pataky, E.L.; Reich, M.; Ometáková, J. Preparation and characterization of nickel targets for cyclotron production of  $^{64}\text{Cu}$ . *J. Radioanal. Nucl. Chem.* **2010**, *286*, 665–670. [[CrossRef](#)]
19. Thisgaard, H.; Jensen, M.; Elema, D.R. Medium to large scale radioisotope production for targeted radiotherapy using a small PET cyclotron. *Appl. Radiat. Isot.* **2011**, *69*, 1–7. [[CrossRef](#)] [[PubMed](#)]
20. Kume, M.; Carey, P.C.; Gaehle, G.; Madrid, E.; Voller, T.; Margenau, W.; Welch, M.J.; Lapi, S.E. A semi-automated system for the routine production of copper-64. *Appl. Radiat. Isot.* **2012**, *70*, 1803–1806. [[CrossRef](#)] [[PubMed](#)]
21. Ohya, T.; Nagatsu, K.; Suzuki, H.; Fukada, M.; Minegishi, K.; Hanyu, M.; Fukumura, T.; Zhang, M.R. Efficient preparation of high-quality  $^{64}\text{Cu}$  for routine use. *Nucl. Med. Biol.* **2016**, *43*, 685–691. [[CrossRef](#)] [[PubMed](#)]
22. McCarthy, D.W.; Shefer, R.E.; Klinkowstein, R.E.; Bass, L.A.; Margeneau, W.H.; Cutler, C.S.; Anderson, C.J.; Welch, M.J. Efficient production of high specific activity  $^{64}\text{Cu}$  using a biomedical cyclotron. *Nucl. Med. Biol.* **1997**, *24*, 35–43. [[CrossRef](#)]
23. Elomaa, V.V.; Jurttila, J.; Rajander, J.; Solin, O. Automation of  $^{64}\text{Cu}$  production at turku PET centre. *Appl. Radiat. Isot.* **2014**, *89*, 74–78. [[CrossRef](#)] [[PubMed](#)]

24. Rajander, J.; Schlesinger, J.; Avila-Rodriguez, M.A.; Solin, O. Increasing specific activity in Cu-64 production by reprocessing the Ni-64 target material. *J. Label. Compd. Radiopharm.* **2009**, *52*, S234.
25. Avila-Rodriguez, M.A.; Nye, J.A.; Nickles, R.J. Simultaneous production of high specific activity  $^{64}\text{Cu}$  and  $^{61}\text{Co}$  with 11.4 MeV protons on enriched  $^{64}\text{Ni}$  nuclei. *Appl. Radiat. Isot.* **2007**, *65*, 1115–1120. [[CrossRef](#)] [[PubMed](#)]
26. Ometáková, J.; Rajec, P.; Csiba, V.; Leporis, M. Automated production of  $^{64}\text{Cu}$  prepared by 18 MeV cyclotron. *J. Radioanal. Nucl. Chem.* **2012**, *293*, 217–222. [[CrossRef](#)]
27. Kim, J.Y.; Park, H.; Lee, J.C.; Kim, K.M.; Lee, K.C.; Ha, H.J.; Choi, T.H.; An, G.I.; Cheon, G.J. A simple Cu-64 production and its application of Cu-64 ATSM. *Appl. Radiat. Isot.* **2009**, *67*, 1190–1194. [[CrossRef](#)] [[PubMed](#)]
28. Hagemester, F. Rituximab for the treatment of non-hodgkin's lymphoma and chronic lymphocytic leukaemia. *Drugs* **2010**, *70*, 261–272. [[CrossRef](#)] [[PubMed](#)]
29. Wu, Y.; Zhu, H.; Zhang, B.; Liu, F.; Chen, J.; Wang, Y.; Wang, Y.; Zhang, Z.; Wu, L.; Si, L.; et al. Synthesis of site-specific radiolabeled antibodies for radioimmunotherapy via genetic code expansion. *Bioconjug. Chem.* **2016**, *27*, 2460–2468. [[CrossRef](#)] [[PubMed](#)]

**Sample Availability:** Sample of the rituximab is available from the authors.



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