

## Review



**Cite this article:** Blecher R, Heinemann-Yerushalmi L, Assaraf E, Konstantin N, Chapman JR, Cope TC, Bewick GS, Banks RW, Zelzer E. 2018 New functions for the proprioceptive system in skeletal biology. *Phil. Trans. R. Soc. B* **373**: 20170327. <http://dx.doi.org/10.1098/rstb.2017.0327>

Accepted: 6 August 2018

One contribution of 14 to a Theo Murphy meeting issue 'Mechanics of development'.

**Subject Areas:**  
biomechanics

**Keywords:**  
muscle spindle, Golgi tendon organs, adolescent idiopathic scoliosis, proprioception, musculoskeleton, fracture repair

**Author for correspondence:**  
Elazar Zelzer  
e-mail: [eli.zelzer@weizmann.ac.il](mailto:eli.zelzer@weizmann.ac.il)

†These authors contributed equally to this work.

# New functions for the proprioceptive system in skeletal biology

Ronen Blecher<sup>1,2,3,†</sup>, Lia Heinemann-Yerushalmi<sup>1,†</sup>, Eran Assaraf<sup>1,2,†</sup>, Nitzan Konstantin<sup>1</sup>, Jens R. Chapman<sup>3</sup>, Timothy C. Cope<sup>4</sup>, Guy S. Bewick<sup>5</sup>, Robert W. Banks<sup>6</sup> and Elazar Zelzer<sup>1</sup>

<sup>1</sup>Department of Molecular Genetics, Weizmann Institute of Science, Rehovot 76100, Israel

<sup>2</sup>Department of Orthopedic Surgery, Assaf HaRofeh Medical Center, Zerrifin 70300, Israel, affiliated to the Sackler Faculty of Medicine, Tel Aviv University, Tel Aviv 69978, Israel

<sup>3</sup>Swedish Neuroscience Institute, Seattle, WA 98122, USA

<sup>4</sup>Coulter Department of Biomedical Engineering, Georgia Institute of Technology and Emory University School of Medicine, Atlanta, GA 30332, USA

<sup>5</sup>Institute of Medical Sciences, University of Aberdeen, Aberdeen AB25 2ZD, UK

<sup>6</sup>Department of Biosciences, Durham University, Durham DH1 3LE, UK

EZ, 0000-0002-1584-6602

Muscle spindles and Golgi tendon organs (GTOs) are two types of sensory receptors that respond to changes in length or tension of skeletal muscles. These mechanosensors have long been known to participate in both proprioception and stretch reflex. Here, we present recent findings implicating these organs in maintenance of spine alignment as well as in realignment of fractured bones. These discoveries have been made in several mouse lines lacking functional mechanosensors in part or completely. In both studies, the absence of functional spindles and GTOs produced a more severe phenotype than that of spindles alone. Interestingly, the spinal curve phenotype, which appeared during peripubertal development, bears resemblance to the human condition adolescent idiopathic scoliosis. This similarity may contribute to the study of the disease by offering both an animal model and a clue as to its aetiology. Moreover, it raises the possibility that impaired proprioceptive signalling may be involved in the aetiology of other conditions. Overall, these new findings expand considerably the scope of involvement of proprioception in musculoskeletal development and function.

This article is part of the Theo Murphy meeting issue 'Mechanics of development'.

## 1. Introduction

Proprioception, in the original Sherringtonian concept of detection of mechanical stimuli arising within the musculoskeletal system itself, is a component of the sense of the relative position of one's own body parts as well as of the level of effort exerted by acting muscles. As such, it is a necessary part of the control of movement and posture. In the musculoskeletal system of humans and other terrestrial vertebrates, the two main types of mechanosensors involved are the muscle spindle [1] and the Golgi tendon organ (GTO) [2]. By virtue of their respective positions in parallel and in series with the force-producing muscle fibres that form the great bulk of skeletal muscles, they respond predominantly to length and changing length of the muscle (muscle spindles) and to actively generated muscle force (GTOs). Although muscle spindles and GTOs are often lumped together as proprioceptors, it is important to recognize that they respond to stimuli whether arising from internal or external (e.g. gravitational) forces. Despite differences in morphology, location, measured input, effect and other characteristics [3–5], these two organs share the ability to respond to mechanical conditions in their local muscle, initiate rapid response in specialized sensory afferent fibres, often loosely termed proprioceptive neurons, and

ultimately modulate local muscle tension through segmental and longer monosynaptic and polysynaptic reflexes [6,7]. True proprioception is typically short range, may be tonic or phasic, and is relatively weak compared with voluntary or externally evoked forces [8]. Nevertheless, here we present our recent evidence that constant disturbance in proprioception, as might occur during abnormal development or following a limb fracture, is extremely important. In order to understand some of the genetic and molecular aspects involved, we begin with a brief overview of the normal development of muscle spindles and GTOs.

Over the years, several molecular pathways have been identified to regulate proprioceptor formation, connectivity and function [9–13]. Proprioceptive neurons transmit mechanical sensations from muscles and tendons via the dorsal root ganglia (DRG) to the spinal cord. These neurons express the neurotrophic tyrosine tropomyosin receptor kinase C (*TrkC*; also known as neurotrophic tyrosine kinase receptor type 3 (*Ntrk3*)) [14] along with neurogenin 2 (*Ngn2*) and Runt related transcription factor 3 (*Runx3*), all essential for the generation and development of DRG sensory neurons [15,16]. In particular, *Runx3*, a member of the Runt domain-containing family of transcription factors, is highly expressed by DRG *TrkC*-positive neurons and is essential for their survival, axonal projection and connectivity to the spinal cord [17,18]. *Runx3*-knockout (KO) mice display severe limb ataxia, a phenotype that was recently recapitulated upon deletion of the genomic elements driving *Runx3* expression in DRG *TrkC* neurons in mice [19]. In skeletal muscles, differentiation of intrafusal (i.e. situated inside the muscle spindle) fibres begins with the establishment of neuromuscular connection between sensory afferent (Ia) neurons and primary myotubes, followed by induction from the sensory neurons. This process is regulated by neuregulin 1 (NRG1) and its receptor ErbB2 receptor tyrosine kinase 2 (ErbB2, also known as HER2) [20]. NRG1–ErbB2 signalling activates downstream targets such as early growth response 3 (*Egr3*), a member of the zinc-finger family of transcription factors [13,21,22] and the Ets transcription factors Pea3 and Erm [9]. Further developed intrafusal fibres express specific intrafusal molecules, including the *TrkC* ligand neurotrophin 3 (NT3) [23] and Ets transcription factor Er81 [9]. These molecules, along with *Egr3*, Pea3 and Erm, were shown to affect the survival of proprioceptive sensory neurons and maintain functional sensory axon-myotube connection during embryonic and early postnatal muscle spindle development [9,13].

Most of the research of the proprioceptive system has focused on its well-known function in motor control. Yet, accumulated evidence shows that this system is also involved in non-autonomous regulation of skeletal development and function. In this review, we describe recent findings pertaining to the roles of proprioception in the maintenance of proper spinal alignment, as well as in morphological restoration of fractured bones.

## 2. The involvement of the proprioceptive system in maintaining spinal alignment

The vertebral column serves as the central axis of the body, playing essential roles in supporting weight and maintaining posture while allowing movement. As other skeletal elements,

the spine is subjected to high stresses created by body weight and by loads exerted by the attached muscles. The unique structure of the spine restricts movement between its parts to provide inherent stability and, thereby, reduces the need for continuous external stabilization by contraction of adjacent muscles [24]. The dynamic maintenance of body posture requires tight regulation of the position and orientation of numerous vertebrae and intervertebral discs. Yet, despite its importance, surprisingly little is known about this regulatory mechanism.

Scoliosis is a condition in which the spinal column is curved laterally by 10° or more [25]. The most common type of the disease is adolescent idiopathic scoliosis (AIS), which appears during puberty without a known cause or other skeletal anomalies in around 3% of school-aged children worldwide. Treatment includes back bracing, with the aim of stopping the progression of deformity. In severe or rapidly progressing curves, surgical correction may be required [26]. To date, despite substantial efforts to decipher the pathogenesis of acquired scoliosis, the mechanisms underlying this condition are still elusive [27].

Because the onset of AIS is not preceded by other skeletal abnormalities, uncovering the mechanisms underlying its pathogenesis has been particularly challenging. Yet, the appearance of a curve in spines comprising morphologically intact elements suggests the involvement of a non-autonomous regulatory mechanism in maintaining spinal alignment. Indeed, we recently reported that *Runx3*-KO mice, which lack *TrkC* neurons connecting between proprioceptors and spinal cord, developed peripubertal scoliosis without prior vertebral dysplasia or muscle asymmetry [28]. Similar results were obtained by conditional deletion of *Runx3* in peripheral nervous tissue or specifically in peripheral sensory neurons, but not in skeletal tissue. Moreover, deletion of enhancer elements driving *Runx3* expression in proprioceptive neurons induced a similar phenotype. A less severe phenotype was exhibited by *Egr3*-KO mice, which lack muscle spindles but not GTOs. Functional assays revealed a decrease in gait regularity, which was also more pronounced in *Runx3*-KO than in *Egr3*-KO mice. These findings implicate impaired proprioceptive signalling in acquired scoliosis and suggest that both receptor types are required for this regulatory mechanism.

Over the years, numerous attempts to develop genetic [29,30], neuroendocrine [31,32] or surgical perturbation [33,34] models for AIS have come short of producing a model that recapitulates the unique features of the disease [27]. Our mouse model displays several hallmark features of AIS, including apparently intact skeletons prior to the appearance of scoliosis, the temporal dynamics of the deformative process and an accentuated right-sided curve of the thoracic spine.

A large body of evidence supports the idea of neuromuscular involvement in the aetiology of scoliosis. These include abnormal morphology and function of neuromuscular elements identified in the central nervous system [35,36], the somatosensory [37] and vestibular [38] systems and in trunk muscles [39,40] of patients with AIS. Moreover, the association between neural insults, such as stroke [41] and cerebral palsy [42], and the development of trunk imbalance and deformity is well-established in clinical practice. In addition, both stroke [43,44] and cerebral palsy [45] have been shown to substantially impair proprioceptive functions,

suggesting a mechanistic cause for the acquired deformity. In animal models, removal of the spinal cord [46] or nerve roots [47–49] resulted in scoliosis, demonstrating the cross-species conservation of the association.

There are several observations that support the notion that proprioceptive function is impaired in patients with AIS. These include reported alterations in postural balance [50,51] and gait [52], as well as reduced numbers of muscle spindles in paravertebral muscles [53]. Additionally, abnormal neural proprioception-related responses, such as inability to reproduce joint angle [54], vibratory sensation [55] and the size-weight illusion, integrating proprioception and visual inputs [56] have also been seen in these patients. Also, the onset time of AIS during the second decade of life is consistent with the maturation of the proprioceptive system [57,58]. Indeed, it has been speculated that proprioception is involved in the control of spine stability, with muscle spindles acting as a regulatory feedback mechanism [59,60]; yet, direct evidence for this involvement has been lacking.

Our work may provide the missing link between proprioception and scoliosis. Our findings indicate that the proprioceptive system may not only provide dynamic control of spine alignment but also prevent progressive spinal deformation. Moreover, our data indicate that this unique relationship between proprioception and spinal alignment requires the synergistic action of both muscle spindles and GTOs. The clinical implication of this notion is that treatment should aim at restoring the balance between motor output and sensory feedback.

In our study, the appearance of spinal deformity (between mouse postnatal days 40 and 60) coincided with highly relevant anatomical and physiological changes, namely maturation of muscle mechanosensors [61,62] and substantial increases in muscle mass [63] and mobility. Thus, peripubertal scoliosis could result from the combination of increasing mechanical loads and a malfunctioning proprioceptive system. Interestingly, our proprioceptive-deficient mouse strains also developed ataxia, which is not seen in patients with AIS. Given the close functional and anatomical interactions between central and peripheral proprioceptive circuits, it is yet to be determined whether ataxia could contribute to the scoliotic phenotype.

From a genetic perspective, while our findings underscore the involvement of *Runx3* and *Egr3* in the mechanisms underlying AIS, the aetiopathogenesis of this disease has long been considered polygenic in nature [64,65]. To date, various loci have been identified as being associated with susceptibility to AIS [66–70]. On the basis of the aetiological explanation we propose, the search for the genetic background of AIS should focus on genes, loci and pathways associated with proprioception. This proposed aetiology may also promote development of evaluation and screening tests based on, for example, the performance of proprioception-dependent tasks. Altogether, the shift in focus in the research of AIS towards viewing it as a neuromuscular disease may lead to advances in diagnostics, in progression assessment and, possibly, to future treatment.

### 3. Proprioception and morphological repair of fractured bones

Correct bone morphology is essential for the function of the musculoskeletal system [71–75]. The evidence-based

textbook model for bone fracture repair describes four distinct stages, from haematoma formation through to bone modelling [76–84]. Yet, despite extensive clinical research into the association between fracture realignment and functional outcome [85–87], little attention has been paid to the mechanisms that restore the general shape of the fractured bone immediately after the injury and before union has been achieved. It is reasonable to assume that the ability to restore skeletal morphology after a traumatic insult to bone integrity would have granted vertebrates a considerable evolutionary advantage. Indeed, several pieces of evidence support the existence of a robust mechanism that rapidly restores bone morphology following injury. In human neonates, humeral birth fractures with severe angulations usually heal well without intervention and with little residual deformity [88]. Additionally, studies of primate skeletons have documented high rates (up to 30%) of well-healed fractures, mostly occurring in youth, which were also marked by minimal residual deformity, further indicating that effective morphological restoration occurs spontaneously and frequently [89–92]. These findings suggest that, during evolution, vertebrates have acquired a mechanism that realigns fractured bones [93].

Previously, we demonstrated the existence of such a mechanism by showing that fractured humeri of neonatal mice undergo realignment without any intervention [94]. The realignment process, which we dubbed natural reduction, involved substantial movement of the two fracture fragments. However, we did not identify the mechanism that senses the location and orientation of the fracture fragments to guide realignment. More recently, we found that muscle spindles and GTOs play this role together [95]. We showed that natural reduction failed in fractured bones of *Runx3*-KO mice. Conditional deletion of *Runx3* in peripheral nervous system, but not in limb mesenchyme, recapitulated the null phenotype, as did inactivation of muscles flanking the fracture site. *Egr3*-KO mice displayed a less severe phenotype, suggesting that both receptor types, as well as muscle contraction, are required for this regulatory mechanism.

Bones have long been known to possess autonomous mechanosensing capabilities [73–75,93,96]. To cope with a dynamic mechanical environment, bones adapt their morphology [97,98], mineral composition and density [99,100] in response to changes in mechanical loading. At the cell level, chondrocytes [101,102], osteoblasts [103] and osteocytes [104] have all been reported to be mechanosensitive. Fracture callus also has mechanosensing capabilities, as has been shown both clinically [105] and experimentally [106,107]. The finding of a proprioception-mediated mechanism that monitors and restores bone integrity adds a non-autonomous level of regulation to the current view of mechanosensing in fracture repair.

Interactions among different tissues regulate the development and growth of the musculoskeletal system [108–111]. For example, skeletal muscles have been shown to regulate the commitment of joint lineage cells [109] as well as the circumferential shape and mineral distribution of developing long bones [97]. In the same vein, it was recently suggested that muscle-derived satellite cells actively participate in fracture repair by expressing various growth factors [112]. The findings that proprioceptive circuitry and muscle activity regulate fracture repair further demonstrates the importance of such interactions between musculoskeletal tissues.

Interestingly, we showed that natural reduction becomes more effective with age. While this finding is at odds with the common knowledge on repair processes, it is consistent with the maturation of the proprioceptive system. In mice, the sensory endings of muscle spindles continue to develop until 30–40 days postnatally [61,62]. In humans, the ability to perform proprioception-specific tasks was shown to increase from childhood into adolescence [57,58]. These findings support the notion that proprioceptive efficiency improves with increased age, which would explain our observation.

The research of fracture repair has largely ignored the role of muscle pull in restoring bone alignment. Based on our findings, we suggest that muscle proprioceptors detect the position of the fracture fragments and guide natural reduction. According to this revised model, the breakage of the bone causes changes in length and tonus of attached muscles. Consequently, asymmetric muscle activation controlled by proprioceptive signals corrects the position of misaligned fracture fragments rapidly and effectively by pulling more strongly on the parts that are further away from their proper location. The activation of this mechanism immediately after the injury may optimize the healing process and its outcome substantially.

#### 4. Proprioception in ageing

Increased longevity in developed countries has long been considered an indication of great scientific and medical advancements. Nonetheless, it also poses considerable clinical and socioeconomic challenges, including a steep rise in healthcare expenditure. With advancing age, the musculoskeletal system undergoes several gradual changes leading to decline in function. For example, sarcopenia is defined as the loss of muscle mass that occurs with ageing, a process that includes reduction in the muscle cross-sectional area as well as a morphological change, ultimately resulting in a 60% reduction in muscle power [113]. A concurrent reduction in bone mineral content, known as osteopaenia or osteoporosis, further exposes the ageing skeleton to low-energy fragility fractures. Finally, accelerated denervation of motor neurons [114] may also contribute to increased fragility in advanced age. Similarly, various elements of the proprioceptive system also change during ageing. Muscle spindles in aged animals, for example, have been shown to possess fewer intrafusal fibres [115] as well as an altered morphology of their sensory endings [116]. In addition, electrophysiological studies showed that mature muscle spindles are altered, displaying a much lower dynamic response of primary endings compared with those of young animals [116]. Taken together, both primary alteration in neural and muscular elements of the musculoskeleton and proprioception-specific changes result in a gradual decline in proprioceptive function in elderly individuals.

One of the more substantial results of this decline is general fragility, manifested in an increased tendency to fall and sustain injuries, most notably hip fractures. By providing a better sense of position, proprioception training was shown to be highly useful in the prevention of falls [117] as well as in the rehabilitation of injured patients [118].

To summarize, similar to other elements of the neuromuscular axis, the proprioceptive system undergoes significant changes with advancing age, contributing to the increased

risk of sustaining a fragility fracture. Better understanding of proprioceptive pathways may assist in developing specific treatments directed at halting their functional decline, or regaining it during a rehabilitation process, thereby greatly improving the well-being of the mature population.

#### 5. The regulatory role of the proprioceptive system in musculoskeletal system: future directions

Proprioceptive mechanosensors provide constant regulation of skeletal muscle length and tension to coordinate motor control [119]. Our recent studies implicate the proprioceptive system in regulation of both maintenance and repair of the skeleton. This increases substantially the scope of known physiological functions of this system. Moreover, it raises the possibility that the proprioceptive system is involved in regulating other processes and that its dysfunction may contribute to the aetiology of various musculoskeletal pathologies.

The regulatory role of the proprioceptive system can be either non-autonomous or mediated by autonomous mechanisms. Our two recent reports [28,95] provide examples for non-autonomous regulation, where the proprioceptive system serves as the sensor that activates muscles to achieve skeletal integrity and alignment. Given that the skeleton is a mechanosensitive tissue, it is tempting to speculate that the proprioceptive system can also influence the autonomous response of the skeleton to a changing mechanical environment. By modulating muscle tonus and activity, the proprioceptive system can control the load exerted on bones, joints, tendons and ligaments. These loads can then be translated into molecular signals by mechanosensors installed within these tissues, thereby regulating both growth and steady state. The existence of such an axis implies that abnormal proprioceptive function could lead to musculoskeletal pathology.

Conceptually, there is a fundamental difference between these two modes of involvement. In mediated regulation, mechanosensors in the affected tissue convert the mechanical loads into biological input. By contrast, during non-autonomous regulation the mechanosensors within the muscle need to identify deviation in organization or morphology of skeletal tissue. The ability of the muscle via its intrinsic sensory organs to detect morphological abnormality in neighbouring tissues implies that this regulatory mechanism contains a 'setpoint' from which deviations are identified and that also signals the termination of the correction process.

One mechanism that may contribute to the setpoint is the fusimotor system. The motor innervation of intrafusal fibres by gamma neurons, which innervate the polar regions of these fibres and regulate their contractile states, allows the central nervous system to control muscle spindle responsiveness to a given length or length change. In particular, increased static gamma activity produces increased tonic firing in spindle afferents. Better understanding of the fusimotor system may resolve any potential involvement in determining the aforementioned setpoint. The mechanical properties of the different intrafusal fibres are also relevant here. There are three types of intrafusal fibres, namely bag1, bag2 and chain fibres. Most muscle spindles contain one bag1, one bag2 and several chain fibres, and the action of

static gamma neurons on the responsiveness of spindle afferents is due to their innervation of the bag2 and chain fibres. Better understanding of their mechanical properties and the molecular mechanism that control them may reveal important insight into the activity of the spindle.

Finally, we know relatively little on the molecular mechanisms that regulate the development, structure and activity of proprioceptive sensory organs. Their involvement in so many important functions should encourage efforts to uncover these mechanisms in order to better understand

how the proprioceptive system regulates processes such as skeletal maintenance, repair and function.

**Data accessibility.** This article has no additional data.

**Competing interests.** We declare we have no competing interests.

**Funding.** This review was supported by grants from the Israel Science Foundation MORASHA Biomedical Research Program in Neurodegenerative Diseases, Genetic Disorders and Metabolic Diseases (no. 2147/17) and from the estate of Bernard Bishin for the WIS-Clalit Program (to E.Z.).

## References

- Bewick GS, Banks RW. 2015 Mechanotransduction in the muscle spindle. *Pflugers Arch.* **467**, 175–190. (doi:10.1007/s00424-014-1536-9)
- Jami L. 1992 Golgi tendon organs in mammalian skeletal muscle: functional properties and central actions. *Physiol. Rev.* **72**, 623–666. (doi:10.1152/physrev.1992.72.3.623)
- Granit R. 1975 The functional role of the muscle spindles—facts and hypotheses. *Brain* **98**, 531–556. (doi:10.1093/brain/98.4.531)
- Maier A. 1997 Development and regeneration of muscle spindles in mammals and birds. *Int. J. Dev. Biol.* **41**, 1–17.
- Moore JC. 1984 The Golgi tendon organ: a review and update. *Am. J. Occup. Ther.* **38**, 227–236. (doi:10.5014/ajot.38.4.227)
- Chen HH, Hippenmeyer S, Arber S, Frank E. 2003 Development of the monosynaptic stretch reflex circuit. *Curr. Opin. Neurobiol.* **13**, 96–102. (doi:10.1016/S0959-4388(03)00006-0)
- Proske U, Gandevia SC. 2012 The proprioceptive senses: their roles in signaling body shape, body position and movement, and muscle force. *Physiol. Rev.* **92**, 1651–1697. (doi:10.1152/physrev.00048.2011)
- Evarts EV. 1981 Sherrington's concept of proprioception. *Trends Neurosci.* **4**, 44–46. (doi:10.1016/0166-2236(81)90016-3)
- Arber S, Ladle DR, Lin JH, Frank E, Jessell TM. 2000 ETS gene *Er81* controls the formation of functional connections between group Ia sensory afferents and motor neurons. *Cell* **101**, 485–498. (doi:10.1016/S0092-8674(00)80859-4)
- Cheret C *et al.* 2013 *Bace1* and *Neuregulin-1* cooperate to control formation and maintenance of muscle spindles. *EMBO J.* **32**, 2015–2028. (doi:10.1038/emboj.2013.146)
- Friese A, Kaltschmidt JA, Ladle DR, Sigrist M, Jessell TM, Arber S. 2009 Gamma and alpha motor neurons distinguished by expression of transcription factor *Err3*. *Proc. Natl Acad. Sci. USA* **106**, 13 588–13 593. (doi:10.1073/pnas.0906809106)
- Hippenmeyer S, Shneider NA, Birchmeier C, Burden SJ, Jessell TM, Arber S. 2002 A role for *neuregulin1* signalling in muscle spindle differentiation. *Neuron* **36**, 1035–1049. (doi:10.1016/S0896-6273(02)01101-7)
- Tourtellotte WG, Keller-Peck C, Milbrandt J, Kucera J. 2001 The transcription factor *Egr3* modulates sensory axon-myotube interactions during muscle spindle morphogenesis. *Dev. Biol.* **232**, 388–399. (doi:10.1006/dbio.2001.0202)
- Marmigere F, Ernfors P. 2007 Specification and connectivity of neuronal subtypes in the sensory lineage. *Nat. Rev. Neurosci.* **8**, 114–127. (doi:10.1038/nrn2057)
- Ma Q, Fode C, Guillemot F, Anderson DJ. 1999 *Neurogenin1* and *neurogenin2* control two distinct waves of neurogenesis in developing dorsal root ganglia. *Genes Dev.* **13**, 1717–1728. (doi:10.1101/gad.13.13.1717)
- Smeyne RJ, Klein R, Schnapp A, Long LK, Bryant S, Lewin A, Lira SA, Barbacid M. 1994 Severe sensory and sympathetic neuropathies in mice carrying a disrupted *Trk/NGF* receptor gene. *Nature* **368**, 246–249. (doi:10.1038/368246a0)
- Inoue K *et al.* 2003 *Runx3* is essential for the target-specific axon pathfinding of *trkC*-expressing dorsal root ganglion neurons. *Blood Cells Mol. Dis.* **30**, 157–160. (doi:10.1016/S1079-9796(03)00032-9)
- Levanon D *et al.* 2002 The *Runx3* transcription factor regulates development and survival of *TrkC* dorsal root ganglia neurons. *EMBO J.* **21**, 3454–3463. (doi:10.1093/emboj/cdf370)
- Appel E *et al.* 2016 An ensemble of regulatory elements controls *Runx3* spatiotemporal expression in subsets of dorsal root ganglia proprioceptive neurons. *Genes Dev.* **30**, 2607–2622. (doi:10.1101/gad.291484.116)
- Leu M, Bellmunt E, Schwander M, Farinas I, Brenner HR, Muller U. 2003 *ErbB2* regulates neuromuscular synapse formation and is essential for muscle spindle development. *Development* **130**, 2291–2301. (doi:10.1242/dev.00447)
- Fernandes MO, Tourtellotte WG. 2015 *Egr3*-dependent muscle spindle stretch receptor intrafusal muscle fiber differentiation and fusimotor innervation homeostasis. *J. Neurosci.* **35**, 5566–5578. (doi:10.1523/JNEUROSCI.0241-15.2015)
- Tourtellotte WG, Milbrandt J. 1998 Sensory ataxia and muscle spindle agenesis in mice lacking the transcription factor *Egr3*. *Nat. Genet.* **20**, 87–91. (doi:10.1038/1757)
- Ernfors P, Lee KF, Kucera J, Jaenisch R. 1994 Lack of neurotrophin-3 leads to deficiencies in the peripheral nervous system and loss of limb proprioceptive afferents. *Cell* **77**, 503–512. (doi:10.1016/0092-8674(94)90213-5)
- Rawls A, Fisher RE. 2010 Development and functional anatomy of the spine. In *The genetics and development of scoliosis* (eds K Kusumi, SL Dunwoodie), pp. 21–46. New York, NY, USA: Springer.
- Scoliosis Research Society (SRS). 2018 *Adolescent idiopathic scoliosis*. Milwaukee, WI: Scoliosis Research Society. (<https://www.srs.org/professionals/online-education-and-resources/conditions-and-treatments/adolescent-idiopathic-scoliosis>)
- Warner WC, Sawyer JR, Kelly DM. 2013 *Scoliosis and kyphosis*. In *Campbell's operative orthopedics* (eds ST Canale, JH Beaty), pp. 1703–1896, 12th edn. Philadelphia, PA: Elsevier.
- Ouellet J, Odent T. 2013 Animal models for scoliosis research: state of the art, current concepts and future perspective applications. *Eur. Spine J.* **22** (Suppl. 2), S81–S95. (doi:10.1007/s00586-012-2396-7)
- Blecher R *et al.* 2017 The proprioceptive system masterminds spinal alignment: insight into the mechanism of scoliosis. *Dev. Cell* **42**, 388–399 e383. (doi:10.1016/j.devcel.2017.07.022)
- Adham IM, Gille M, Gamel AJ, Reis A, Dressel R, Steding G, Brand-Saberi B, Engel W. 2005 The scoliosis (*sco*) mouse: a new allele of *Pax1*. *Cytogenet. Genome Res.* **111**, 16–26. (doi:10.1159/000085665)
- Blanco G *et al.* 2001 The *kyphoscoliosis (ky)* mouse is deficient in hypertrophic responses and is caused by a mutation in a novel muscle-specific protein. *Hum. Mol. Genet.* **10**, 9–16. (doi:10.1093/hmg/10.1.9)
- Machida M, Duboussset J, Imamura Y, Iwaya T, Yamada T, Kimura J. 1995 Role of melatonin deficiency in the development of scoliosis in pinealectomised chickens. *J. Bone Joint Surg. Br.* **77**, 134–138. (doi:10.1302/0301-620X.77B1.7822371)
- Thillard MJ. 1959 Vertebral column deformities following epiphysectomy in the chick. *C. r. hebd. Séanc. Acad. Sci., Paris* **248**, 1238–1240.
- Pal GP, Bhatt RH, Patel VS. 1991 Mechanism of production of experimental scoliosis in rabbits. *Spine* **16**, 137–142.
- Robin GC. 1996 Scoliosis induced by rib resection in chickens. *J. Spinal Disord.* **9**, 351. (doi:10.1097/00002517-199608000-00015)

35. Shi L, Wang D, Hui SC, Tong MC, Cheng JC, Chu WC. 2013 Volumetric changes in cerebellar regions in adolescent idiopathic scoliosis compared with healthy controls. *Spine J.* **13**, 1904–1911. (doi:10.1016/j.spinee.2013.06.045)
36. Wang D, Shi L, Chu WC, Burwell RG, Cheng JC, Ahuja AT. 2012 Abnormal cerebral cortical thinning pattern in adolescent girls with idiopathic scoliosis. *Neuroimage* **59**, 935–942. (doi:10.1016/j.neuroimage.2011.07.097)
37. Guo X, Chau WW, Hui-Chan CW, Cheung CS, Tsang WW, Cheng JC. 2006 Balance control in adolescents with idiopathic scoliosis and disturbed somatosensory function. *Spine* **31**, E437–E440. (doi:10.1097/01.brs.0000222048.47010.bf)
38. Shi L, Wang D, Chu WC, Burwell GR, Wong TT, Heng PA, Cheng JC. 2011 Automatic MRI segmentation and morphoanatomy analysis of the vestibular system in adolescent idiopathic scoliosis. *Neuroimage* **54** (Suppl. 1), S180–S188. (doi:10.1016/j.neuroimage.2010.04.002)
39. Acaroglu E, Akel I, Alanay A, Yazici M, Marcucio R. 2009 Comparison of the melatonin and calmodulin in paravertebral muscle and platelets of patients with or without adolescent idiopathic scoliosis. *Spine* **34**, E659–E663. (doi:10.1097/BRS.0b013e3181a3c7a2)
40. McIntire KL, Asher MA, Burton DC, Liu W. 2007 Trunk rotational strength asymmetry in adolescents with idiopathic scoliosis: an observational study. *Scoliosis* **2**, 9. (doi:10.1186/1748-7161-2-9)
41. Gillen G. 2015 Trunk control: supporting functional independence. In *Stroke rehabilitation: a function-based approach* (ed. G Gillen), pp. 360–393, 4th edn. St. Louis, MO: Elsevier.
42. Scoliosis Research Society (SRS). 2018 Neuromuscular Scoliosis. (<https://www.srs.org/patients-and-families/conditions-and-treatments/parents/scoliosis/early-onset-scoliosis/neuromuscular-scoliosis>)
43. Dukelow SP, Herter TM, Moore KD, Demers MJ, Glasgow JJ, Bagg SD, Norman KE, Scott SH. 2010 Quantitative assessment of limb position sense following stroke. *Neurorehabil. Neural Repair* **24**, 178–187. (doi:10.1177/1545968309345267)
44. Smith DL, Akhtar AJ, Garraway WM. 1983 Proprioception and spatial neglect after stroke. *Age Ageing* **12**, 63–69. (doi:10.1093/ageing/12.1.63)
45. Smorenburg AR, Ledebt A, Deconinck FJ, Savelsbergh GJ. 2012 Deficits in upper limb position sense of children with spastic hemiparetic cerebral palsy are distance-dependent. *Res. Dev. Disabil.* **33**, 971–981. (doi:10.1016/j.ridd.2012.01.006)
46. Barrios C, Tunon MT, De Salis JA, Beguiristain JL, Canadell J. 1987 Scoliosis induced by medullary damage: an experimental study in rabbits. *Spine* **12**, 433–439. (doi:10.1097/00007632-198706000-00003)
47. Liszka O. 1961 Spinal cord mechanisms leading to scoliosis in animal experiments. *Acta Med. Pol.* **2**, 45–63.
48. MacEwen GD. 1973 Experimental scoliosis. *Clin. Orthop. Relat. Res.* **93**, 69–74. (doi:10.1097/00003086-197306000-00009)
49. Pincott JR, Davies JS, Taffs LF. 1984 Scoliosis caused by section of dorsal spinal nerve roots. *J. Bone Joint Surg. Br.* **66**, 27–29. (doi:10.1302/0301-620X.66B1.6693473)
50. Gruber AH, Busa MA, Gorton Iii GE, Van Emmerik RE, Masso PD, Hamill J. 2011 Time-to-contact and multiscale entropy identify differences in postural control in adolescent idiopathic scoliosis. *Gait Posture* **34**, 13–18. (doi:10.1016/j.gaitpost.2011.02.015)
51. Lao ML, Chow DH, Guo X, Cheng JC, Holmes AD. 2008 Impaired dynamic balance control in adolescents with idiopathic scoliosis and abnormal somatosensory evoked potentials. *J. Pediatr. Orthop.* **28**, 846–849. (doi:10.1097/BPO.0b013e31818e1bc9)
52. Yang JH, Suh SW, Sung PS, Park WH. 2013 Asymmetrical gait in adolescents with idiopathic scoliosis. *Eur. Spine J.* **22**, 2407–2413. (doi:10.1007/s00586-013-2845-y)
53. Ford DM, Bagnall KM, Clements CA, McFadden KD. 1988 Muscle spindles in the paraspinal musculature of patients with adolescent idiopathic scoliosis. *Spine* **13**, 461–465. (doi:10.1097/00007632-198805000-00004)
54. Barrack RL, Whitecloud III TS, Burke SW, Cook SD, Harding AF. 1984 Proprioception in idiopathic scoliosis. *Spine* **9**, 681–685. (doi:10.1097/00007632-198410000-00005)
55. Wyatt MP, Barrack RL, Mubarak SJ, Whitecloud TS, Burke SW. 1986 Vibratory response in idiopathic scoliosis. *J. Bone Joint Surg. Br.* **68**, 714–718. (doi:10.1302/0301-620X.68B5.3782230)
56. Yekutiel M, Robin GC, Yarom R. 1981 Proprioceptive function in children with adolescent idiopathic scoliosis. *Spine* **6**, 560–566. (doi:10.1097/00007632-198111000-00006)
57. Goble DJ, Lewis CA, Hurvitz EA, Brown SH. 2005 Development of upper limb proprioceptive accuracy in children and adolescents. *Hum. Mov. Sci.* **24**, 155–170. (doi:10.1016/j.humov.2005.05.004)
58. Pickett K, Konczak J. 2009 Measuring kinaesthetic sensitivity in typically developing children. *Dev. Med. Child Neurol.* **51**, 711–716. (doi:10.1111/j.1469-8749.2008.03229.x)
59. Reeves NP, Narendra KS, Cholewicki J. 2007 Spine stability: the six blind men and the elephant. *Clin. Biomech.* **22**, 266–274. (doi:10.1016/j.clinbiomech.2006.11.011)
60. Bergmark A. 1989 Stability of the lumbar spine. A study in mechanical engineering. *Acta Orthop. Scand.* **230**, 1–54. (doi:10.3109/17453678909154177)
61. Maeda N, Osawa K, Masuda T, Hakeda Y, Kumegawa M. 1985 Postnatal development of the annulospiral endings of Ia fibres in muscle spindles of mice. *Acta Anat.* **124**, 42–46. (doi:10.1159/000146093)
62. Osawa K *et al.* 1988 Postnatal development of the annulospiral endings of Ia fibres in muscle spindles of the mouse temporal muscle. *Anat. Anz.* **167**, 253–257.
63. Griffin GE, Goldspink G. 1973 The increase in skeletal muscle mass in male and female mice. *Anat. Rec.* **177**, 465–469. (doi:10.1002/ar.1091770311)
64. Ikegawa S. 2016 Genomic study of adolescent idiopathic scoliosis in Japan. *Scoliosis Spinal Disord.* **11**, 5. (doi:10.1186/s13013-016-0067-x)
65. Ward K, Ogilvie J, Argyle V, Nelson L, Meade M, Braun J, Chettier R. 2010 Polygenic inheritance of adolescent idiopathic scoliosis: a study of extended families in Utah. *Am. J. Med. Genet. A* **152**, 1178–1188. (doi:10.1002/ajmg.a.33145)
66. Hayes M, Gao X, Yu LX, Paria N, Henkelman RM, Wise CA, Ciruna B. 2014 ptk7 mutant zebrafish models of congenital and idiopathic scoliosis implicate dysregulated Wnt signalling in disease. *Nat. Commun.* **5**, 4777. (doi:10.1038/ncomms5777)
67. Kou I *et al.* 2013 Genetic variants in GPR126 are associated with adolescent idiopathic scoliosis. *Nat. Genet.* **45**, 676–679. (doi:10.1038/ng.2639)
68. Ogura Y *et al.* 2015 A functional SNP in BNC2 is associated with adolescent idiopathic scoliosis. *Am. J. Hum. Genet.* **97**, 337–342. (doi:10.1016/j.ajhg.2015.06.012)
69. Sharma S *et al.* 2015 A PAX1 enhancer locus is associated with susceptibility to idiopathic scoliosis in females. *Nat. Commun.* **6**, 6452. (doi:10.1038/ncomms7452)
70. Takahashi Y *et al.* 2011 A genome-wide association study identifies common variants near *LBX1* associated with adolescent idiopathic scoliosis. *Nat. Genet.* **43**, 1237–1240. (doi:10.1038/ng.974)
71. Kettelkamp DB, Hillberry BM, Murrish DE, Heck DA. 1988 Degenerative arthritis of the knee secondary to fracture malunion. *Clin. Orthop. Relat. Res.* **234**, 159–169. (doi:10.1097/00003086-198809000-00029)
72. Ring D. 2005 Treatment of the neglected distal radius fracture. *Clin. Orthop. Relat. Res.* **431**, 85–92. (doi:10.1097/01.blo.0000152442.66083.ff)
73. Currey JD. 2003 The many adaptations of bone. *J. Biomech.* **36**, 1487–1495. (doi:10.1016/S0021-9290(03)00124-6)
74. Frost HM. 2001 From Wolff's law to the Utah paradigm: insights about bone physiology and its clinical applications. *Anat. Rec.* **262**, 398–419. (doi:10.1002/ar.1049)
75. Weiner S, Wagner HD. 1998 The material bone: structure-mechanical function relations. *Annu. Rev. Mater. Sci.* **28**, 271–298. (doi:10.1146/annurev.matsci.28.1.271)
76. Ai-Aql ZS, Alagl AS, Graves DT, Gerstenfeld LC, Einhorn TA. 2008 Molecular mechanisms controlling bone formation during fracture healing and distraction osteogenesis. *J. Dent. Res.* **87**, 107–118. (doi:10.1177/154405910808700215)
77. Bolander ME. 1992 Regulation of fracture repair by growth factors. *Proc. Soc. Exp. Biol. Med.* **200**, 165–170. (doi:10.3181/00379727-200-43410A)
78. Brighton C. 1984 Principles of fracture healing: Part I. The biology of fracture repair. In *Instructional course lectures XXXIII* (ed. JA Murray), pp. 60–82. Saint Louis, MO: CV Mosby.

79. Cho TJ, Gerstenfeld LC, Einhorn TA. 2002 Differential temporal expression of members of the transforming growth factor  $\beta$  superfamily during murine fracture healing. *J. Bone Miner. Res.* **17**, 513–520. (doi:10.1359/jbmr.2002.17.3.513)
80. Einhorn TA. 1998 The cell and molecular biology of fracture healing. *Clin. Orthop. Relat. Res.* **355** (Suppl.), S7–S21. (doi:10.1097/00003086-199810001-00003)
81. Gerstenfeld LC, Cullinane DM, Barnes GL, Graves DT, Einhorn TA. 2003 Fracture healing as a post-natal developmental process: molecular, spatial, and temporal aspects of its regulation. *J. Cell. Biochem.* **88**, 873–884. (doi:10.1002/jcb.10435)
82. Schindeler A, McDonald MM, Bokko P, Little DG. 2008 Bone remodeling during fracture repair: the cellular picture. *Semin. Cell Dev. Biol.* **19**, 459–466. (doi:10.1016/j.semcdb.2008.07.004)
83. Shapiro F. 2008 Bone development and its relation to fracture repair. The role of mesenchymal osteoblasts and surface osteoblasts. *Eur. Cell. Mater.* **15**, 53–76. (doi:10.22203/eCM.v015a05)
84. Wilkins KE. 2005 Principles of fracture remodeling in children. *Injury* **36**(Suppl. 1), A3–S11. (doi:10.1016/j.injury.2004.12.007)
85. Ellsasser JC, Moyer CF, Lesker PA, Simmons DJ. 1975 Improved healing of experimental long bone fractures in rabbits by delayed internal fixation. *J. Trauma* **15**, 869–876. (doi:10.1097/00005373-197510000-00005)
86. Fogel GR, Morrey BF. 1987 Delayed open reduction and fixation of ankle fractures. *Clin. Orthop. Relat. Res.* **335**, 187–195. (doi:10.1097/00003086-199702000-00027)
87. Lam SJ. 1964 The place of delayed internal fixation in the treatment of fractures of the long bones. *J. Bone Joint Surg.* **46**, 393–397. (doi:10.1302/0301-620X.46B3.393)
88. Husain SN, King EC, Young JL, Sarwark JF. 2008 Remodeling of birth fractures of the humeral diaphysis. *J. Pediatr. Orthop.* **28**, 10–13. (doi:10.1097/BPO.0b013e3181558c67)
89. Bramblett CA. 1967 Pathology in the Darajani baboon. *Am. J. Phys. Anthropol.* **26**, 331–340. (doi:10.1002/ajpa.1330260308)
90. Duckworth WL. 1911 On the natural repair of fractures, as seen in the skeletons of anthropoid apes. *J. Anat. Physiol.* **46**, 81–85.
91. Schultz AH. 1939 *Notes on diseases and healed fractures of wild apes: and their bearing on the antiquity of pathological conditions in man*. Baltimore, MD: Johns Hopkins University Press.
92. Schultz AH. 1944 Age changes and variability in gibbons. A morphological study on a population sample of a man-like ape. *Am. J. Phys. Anthropol.* **2**, 1–129. (doi:10.1002/ajpa.1330020102)
93. Currey JD. 2002 *Bones; structure and mechanics*. Princeton, NJ, USA; Oxfordshire, England: Princeton University Press.
94. Rot C, Stern T, Blecher R, Friesen B, Zelzer E. 2014 A mechanical Jack-like mechanism drives spontaneous fracture healing in neonatal mice. *Dev. Cell* **31**, 159–170. (doi:10.1016/j.devcel.2014.08.026)
95. Blecher R, Krief S, Galili T, Assaraf E, Stern T, Anekstein Y, Agar G, Zelzer E. 2017 The proprioceptive system regulates morphologic restoration of fractured bones. *Cell Rep.* **20**, 1775–1783. (doi:10.1016/j.celrep.2017.07.073)
96. Burr DB, Allen MR. 2014 *Basic and applied bone biology*. San Diego, CA, USA: Elsevier - Academic Press.
97. Sharir A, Stern T, Rot C, Shahar R, Zelzer E. 2011 Muscle force regulates bone shaping for optimal load-bearing capacity during embryogenesis. *Development* **138**, 3247–3259. (doi:10.1242/dev.063768)
98. Robling AG, Castillo AB, Turner CH. 2006 Biomechanical and molecular regulation of bone remodeling. *Annu. Rev. Biomed. Eng.* **8**, 455–498. (doi:10.1146/annurev.bioeng.8.061505.095721)
99. Bach-Gansmo FL, Wittig NK, Bruel A, Thomsen JS, Birkedal H. 2016 Immobilization and long-term recovery results in large changes in bone structure and strength but no corresponding alterations of osteocyte lacunar properties. *Bone* **91**, 139–147. (doi:10.1016/j.bone.2016.07.005)
100. Ellman R, Spatz J, Cloutier A, Palme R, Christiansen BA, Boussein ML. 2013 Partial reductions in mechanical loading yield proportional changes in bone density, bone architecture, and muscle mass. *J. Bone Miner. Res.* **28**, 875–885. (doi:10.1002/jbmr.1814)
101. Lee HS, Millward-Sadler SJ, Wright MO, Nuki G, Salter DM. 2000 Integrin and mechanosensitive ion channel-dependent tyrosine phosphorylation of focal adhesion proteins and  $\beta$ -catenin in human articular chondrocytes after mechanical stimulation. *J. Bone Miner. Res.* **15**, 1501–1509. (doi:10.1359/jbmr.2000.15.8.1501)
102. Wann AK, Zuo N, Haycraft CJ, Jensen CG, Poole CA, McGlashan SR, Knight MM. 2012 Primary cilia mediate mechanotransduction through control of ATP-induced  $\text{Ca}^{2+}$  signalling in compressed chondrocytes. *FASEB J.* **26**, 1663–1671. (doi:10.1096/fj.11-193649)
103. Davidson RM, Tatakis DW, Auerbach AL. 1990 Multiple forms of mechanosensitive ion channels in osteoblast-like cells. *Pflugers Arch.* **416**, 646–651. (doi:10.1007/BF00370609)
104. Huiskes R, Ruimerman R, van Lenthe GH, Janssen JD. 2000 Effects of mechanical forces on maintenance and adaptation of form in trabecular bone. *Nature* **405**, 704–706. (doi:10.1038/35015116)
105. Aro HT, Chao EY. 1993 Bone-healing patterns affected by loading, fracture fragment stability, fracture type, and fracture site compression. *Clin. Orthop. Relat. Res.* **293**, 8–17. (doi:10.1097/00003086-199308000-00003)
106. Probst A, Spiegel HU. 1997 Cellular mechanisms of bone repair. *J. Invest. Surg.* **10**, 77–86. (doi:10.3109/08941939709032137)
107. Thompson Z, Mclaud T, Hu D, Helms JA. 2002 A model for intramembranous ossification during fracture healing. *J. Orthop. Res.* **20**, 1091–1098. (doi:10.1016/S0736-0266(02)00017-7)
108. Blitz E *et al.* 2009 Bone ridge patterning during musculoskeletal assembly is mediated through SCX regulation of Bmp4 at the tendon-skeleton junction. *Dev. Cell* **17**, 861–873. (doi:10.1016/j.devcel.2009.10.010)
109. Kahn J *et al.* 2009 Muscle contraction is necessary to maintain joint progenitor cell fate. *Dev. Cell* **16**, 734–743. (doi:10.1016/j.devcel.2009.04.013)
110. Schwartz Y, Farkas Z, Stern T, Aszodi A, Zelzer E. 2012 Muscle contraction controls skeletal morphogenesis through regulation of chondrocyte convergent extension. *Dev. Biol.* **370**, 154–163. (doi:10.1016/j.ydbio.2012.07.026)
111. Zelzer E, Blitz E, Killian ML, Thomopoulos S. 2014 Tendon-to-bone attachment: from development to maturity. *Birth Defects Res. C Embryo Today* **102**, 101–112. (doi:10.1002/bdr.21056)
112. Abou-Khalil R *et al.* 2015 Role of muscle stem cells during skeletal regeneration. *Stem Cells* **33**, 1501–1511. (doi:10.1002/stem.1945)
113. Thom JM, Morse CI, Birch KM, Narici MV. 2007 Influence of muscle architecture on the torque and power-velocity characteristics of young and elderly men. *Eur. J. Appl. Physiol.* **100**, 613–619. (doi:10.1007/s00421-007-0481-0)
114. Guillet C, Auguste P, Mayo W, Kreher P, Gascan H. 1999 Ciliary neurotrophic factor is a regulator of muscular strength in aging. *J. Neurosci.* **19**, 1257–1262. (doi:10.1523/JNEUROSCI.19-04-01257.1999)
115. Swash M, Fox KP. 1972 The effect of age on human skeletal muscle. Studies of the morphology and innervation of muscle spindles. *J. Neurol. Sci.* **16**, 417–432. (doi:10.1016/0022-510X(72)90048-2)
116. Kim GH, Suzuki S, Kanda K. 2007 Age-related physiological and morphological changes of muscle spindles in rats. *J. Physiol.* **582**, 525–538. (doi:10.1113/jphysiol.2007.130120)
117. Riva D, Bianchi R, Rocca F, Mamo C. 2016 Proprioceptive training and injury prevention in a professional men's basketball team: a six-year prospective study. *J. Strength Cond. Res.* **30**, 461–475. (doi:10.1519/JSC.0000000000001097)
118. Lephart SM, Pincivero DM, Giraldo JL, Fu FH. 1997 The role of proprioception in the management and rehabilitation of athletic injuries. *Am. J. Sports Med.* **25**, 130–137. (doi:10.1177/036354659702500126)
119. Windhorst U. 2007 Muscle proprioceptive feedback and spinal networks. *Brain Res. Bull.* **73**, 155–202. (doi:10.1016/j.brainresbull.2007.03.010)