

Review

Imaging Biomarkers in Alzheimer's Disease: A Practical Guide for Clinicians

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Abstract. Although recent developments in imaging biomarkers have revolutionized the diagnosis of Alzheimer's disease at early stages, the utility of most of these techniques in clinical setting remains unclear. The aim of this review is to provide a clear stepwise algorithm on using multitier imaging biomarkers for the diagnosis of Alzheimer's disease to be used by clinicians and radiologists for day-to-day practice. We summarized the role of most common imaging techniques and their appropriate clinical use based on current consensus guidelines and recommendations with brief sections on acquisition and analysis techniques for each imaging modality. Structural imaging, preferably MRI or alternatively high resolution CT, is the essential first tier of imaging. It improves the accuracy of clinical diagnosis and excludes other potential pathologies. When the results of clinical examination and structural imaging, assessed by dementia expert, are still inconclusive, functional imaging can be used as a more advanced option. PET with ligands such as amyloid tracers and ¹⁸F-fluorodeoxyglucose can improve the sensitivity and specificity of diagnosis particularly at the early stages of the disease. There are, however, limitations in using these techniques in wider community due to a combination of lack of facilities and expertise to interpret the findings. The role of some of the more recent imaging techniques including tau imaging, functional MRI, or diffusion tensor imaging in clinical practice, remains to be established in the ongoing and future studies.

Keywords: Alzheimer's disease, biomarkers, guidelines, imaging, magnetic resonance imaging, positron emission tomography

INTRODUCTION

Despite all the advances in imaging of Alzheimer's disease (AD) in the research setting, there is a lack of translation of these methodologies into the clinical practice. Most imaging biomarkers have not been validated in unselected patient cohorts and participants in large AD studies are not representative of the general population. These techniques require special facilities and expertise to perform and interpret. The paucity of standard acquisition and analysis methods

between different centers make the widespread adoption of them even more challenging. In addition, some of the new imaging modalities are still too expensive to be considered cost-effective in a community setting or in non-specialized centers.

Based on a growing body of evidence, the early and accurate diagnosis in preclinical stages of the disease is paramount to tackle AD [1, 2]. Therefore, an important goal of developing imaging biomarkers should be to bridge the gap between the research setting and clinical practice and to use these techniques as screening, diagnostic, or prognostic markers in the wider population.

In this review, we summarize the utility of different imaging biomarkers, review the current

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guidelines and recommendations, and present a stepwise algorithm for radiological evaluation of cognitive impairment for clinicians. We will also review some of the novel techniques, which may not come to clinical practice in the near future, to demonstrate their potential utility.

STRUCTURAL IMAGING

Current recommendation and guidelines

There is a consensus among all current guidelines that structural imaging, i.e., magnetic resonance imaging (MRI) or computerized tomography (CT), is required for evaluation of patients presenting with a cognitive/dementia syndrome (CDS) in the clinical setting [3–7]. The UK National Institute of Health and Care Excellence [3], European Federation of Neurological Societies (EFNS) [4, 8], and the Fourth Canadian Consensus Conference on Diagnosis and Treatment of Dementia [6] have all recommended using structural imaging to 1) exclude treatable causes of dementia and 2) assess the pattern of atrophy which could provide additional diagnostic or predictive values to clinical information. The presence of atrophy can improve the accuracy of a clinical diagnosis considerably. Scheltens et al. showed the incremental diagnostic gain of positive or negative MRI for any given clinical probability [5]. MRI is the preferable method because of its higher resolution and ability to delineate a spectrum of changes related to vascular pathologies and white matter diseases in the brain. High resolution CT can be used as a replacement when there is a contraindication for MRI. It has also been recommended to use a standard MRI protocols for dementia to facilitate the comparison with follow up imaging.

The characteristic pattern of atrophy in AD involves structures of the medial temporal lobe (MTL) including the hippocampi, entorhinal cortex, parahippocampal gyrus, amygdala, and also the cingulate cortex while the motor cortex is relatively spared [9–11]. Despite all the evidence supporting the presence of AD “signature” atrophy in large research studies based on group level analysis, it is still very challenging to quantify the degree of atrophy in a single case in clinical setting.

Acquisition and analysis techniques

The minimum sequences required to assess a patient with CDS are:

- 1) High resolution 3D/Volumetric T1 weighted images produced with fast spoiled gradient echo (FSPGR; GE Healthcare), magnetization prepared rapid acquisition GRE (MPRAGE; Siemens Healthineers) or other similar sequences in coronal and at least one more plane (axial or sagittal) which can be produced using reformatting software. This sequence is used to assess the anatomy, internal structure and degree of atrophy;
- 2) T2 weighted images;
- 3) Fluid-attenuated inversion recovery (FLAIR) sequence.

Both the latter are used to determine the severity of vascular changes. In addition, a coronal FLAIR is the best projection to evaluate the signal of hippocampi and MTL.

Other sequences such as 4) Gradient echo (GRE)/T2* /Susceptibility Weighted Imaging (SWI) and 5) Diffusion weighted imaging can provide additional valuable information. SWI can detect cerebral microbleeds, reflecting evidence of vascular amyloid pathology, which has both diagnostic role [12] and therapeutic implication in anti amyloid treatment [13]. Diffusion weighted images can exclude other pathologies such as recent vascular events and more importantly support the diagnosis of Creutzfeldt-Jakob disease as a cause of subacute cognitive impairment.

The following is a summary of different ways to assess the hippocampal and MTL atrophy:

- 1) Visual assessment: scoring atrophy from 0 to 4 based on the width of the choroid fissure, maximum width of the temporal horn, and height of hippocampal formation [14] (Fig. 1). This is a technique widely used in a clinical setting as it is not time consuming, does not require either specialist software, calculations or reformatting. The limitation of this method is the significant inter-observer variation in scaling atrophy [15–17]. Despite scoring variation there is, however, good inter-rater reliability when visual assessment is used to dichotomize the data into presence or absence of atrophy [15], which might be sufficient for clinical purposes.
- 2) Linear (1D): Different methods are used for the linear assessment of atrophy including measuring the radial width of the temporal horn [18], measurement of interuncal distance, [19] and a grading scale (0–4) for the degree of medial

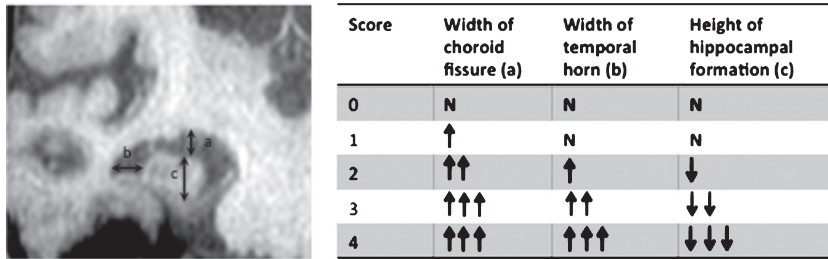


Fig. 1. Visual assessment of MTL and hippocampal atrophy. a) largest vertical width of choroid fissure; b) width of temporal horn; c) maximum vertical height of hippocampal formation. ↑, increase; ↓, decrease; N, normal.

temporal lobe atrophy using the area of the hippocampi and maximal transverse width of the temporal horns [20].

- 3) Planimetric (2D) measurements: These techniques comprise assessment of the degree of medial temporal lobe atrophy (MTA) by measuring the whole MTL region including the parenchyma and cerebrospinal fluid (CSF) spaces (A), the parenchyma in the MTL region (B) and the body of the ipsilateral lateral ventricle (C) in the coronal images at the level of interpeduncular fossa. Various parameters such as $2D\text{-MTA} = A - B$ or Medial Temporal Atrophy index ($MTAi = (A - B) \times 10 / C$) have been developed using these measurements [21–23]. Although all of the above techniques (linear and planimetric) are feasible and can be assessed on MRI images obtained in any clinical setting, there is a lack of normative values to distinguish normal from abnormal particularly at early stage of disease such as minimal cognitive impairment (MCI). Also, they have not been used on large cohorts of patients and, in comparison to volumetric measurements, they are less accurate [24].
- 4) Volumetric measurement (3D): There are manual and automated techniques for measuring the volume of hippocampi. The manual segmentation by an expert is considered as gold standard for hippocampal volumetry but it requires a high level of training, is very time consuming and costly. The other issue preventing it from being used in the clinical setting is significant variation in segmenting hippocampus [25]. To overcome the discrepancies in the segmentation of hippocampus between different centers, the European Alzheimer's Disease Consortium (EADC) and Alzheimer's Disease Neuroimaging Initiative (ADNI) with support

of the Alzheimer's Association have developed a consensual harmonisation protocol for manual segmentation of hippocampus [26, 27] which is available as a web-based protocol (<http://www.hippocampal-protocol.net>). The automated segmentation techniques can overcome some of the reliability issues of manual techniques, but extra software and expertise is required to operate them efficiently. In addition, there is limited data on their sensitivity and specificity and further validation is required before they can be used in clinical practice [25, 28].

In a multicenter European study, the accuracy of different structural MRI techniques to distinguish AD from healthy controls were compared. They found accuracy of visual rating (81%), manual hippocampal measurement (89%), and automated multivariate analysis of hippocampal and MTL atrophy (83%) were not significantly different [29].

Clinical implementation

The role of structural MRI differs based on the clinical presentation, stage of the disease, and possibility of an alternative diagnosis/mixed pathologies. Figure 2 demonstrates a stepwise approach in assessment of CDS in the clinic using structural MRI. If a patient presented with a clinical picture compatible with probable AD based on National Institute of Aging- Alzheimer's Association guideline (NIA-AA) [30], with clinical probability of 0.5 and above and MRI showed atrophy in AD signature regions, the diagnosis is confirmed with a probability of between 0.9–1 [5]. For such a case in clinical setting and assuming the patient will not be recruited to any research study or new therapeutic clinical trial, no further imaging will be required. Structural MRI can also help to determine the severity of vascular disease

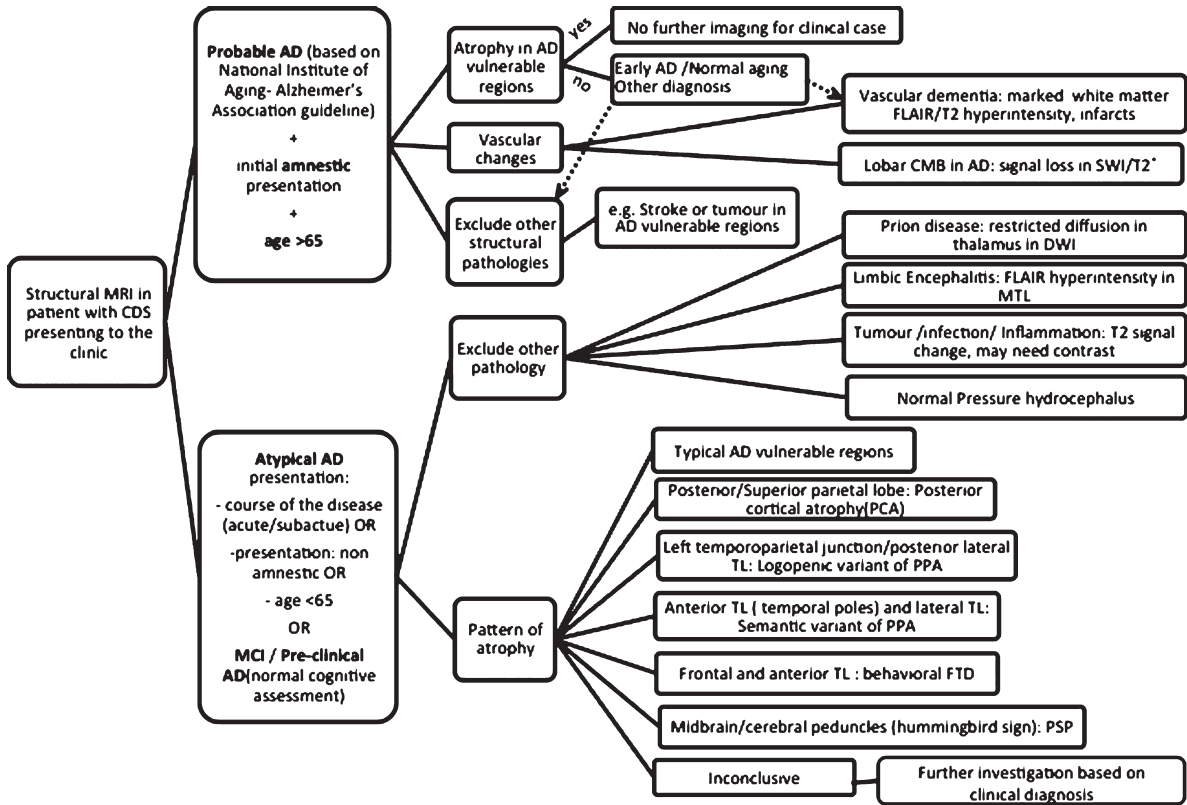


Fig. 2. Summary of structural MRI applications in assessment of AD based on current guidelines and literature review.

and exclude other structural pathologies. However, if the clinical picture is atypical because of the course of the disease or presenting symptoms (non-amnesic), then the initial role of MRI is mainly to exclude other causes of cognitive impairment mimicking dementia some of which can be treated if diagnosed promptly. Examples of such conditions include limbic and paraneoplastic encephalitides, different infections, systemic and CNS inflammatory conditions, tumors, and stroke. Atypical presentations warrant a more thorough clinical assessment and detailed neuropsychological examination to diagnose other types of dementia and MRI can be useful in differentiating the pattern of atrophy in some of these cases (Fig. 3). Once non-degenerative causes are excluded, an atypical presentation should prompt consideration of non-amnesic variants of AD: posterior cortical atrophy presents with prominent visuospatial impairment while the logopenic variant of primary progressive aphasia (PPA) is characterized with impairment of single word retrieval and repetition of sentences. Moreover, the behavioral variant of AD presents with a dysexecutive syndrome and impaired

behavior. Non-AD dementia syndromes should always be a consideration in these atypical cases. Frontotemporal dementias (FTD) can present with abnormal behavior or impaired language. Evidence of Parkinsonism should lead to consideration of other diagnoses such as dementia with Lewy body (DLB)/ Parkinson's disease dementia (PDD), if accompanied by other clinical signs, namely supranuclear gaze palsy and prominent asymmetrical apraxia, progressive supranuclear palsy (PSP) and corticobasal degeneration (CBD) should be considered respectively. MRI can be helpful in identifying the pattern of atrophy associated with many less common forms of dementia as detailed in Fig. 3. One important caveat is that the absence of such atrophy patterns does not exclude the condition as in many instances, clinical syndromes might present without the "typical" atrophy pattern. In some diseases such as DLB/ PDD or CBD, no distinctive pattern of atrophy has been identified. Mixed pathologies are also extremely common especially with advancing age and the combination of different pathologies can contaminate the atrophy patterns of individual diseases. MRI is also

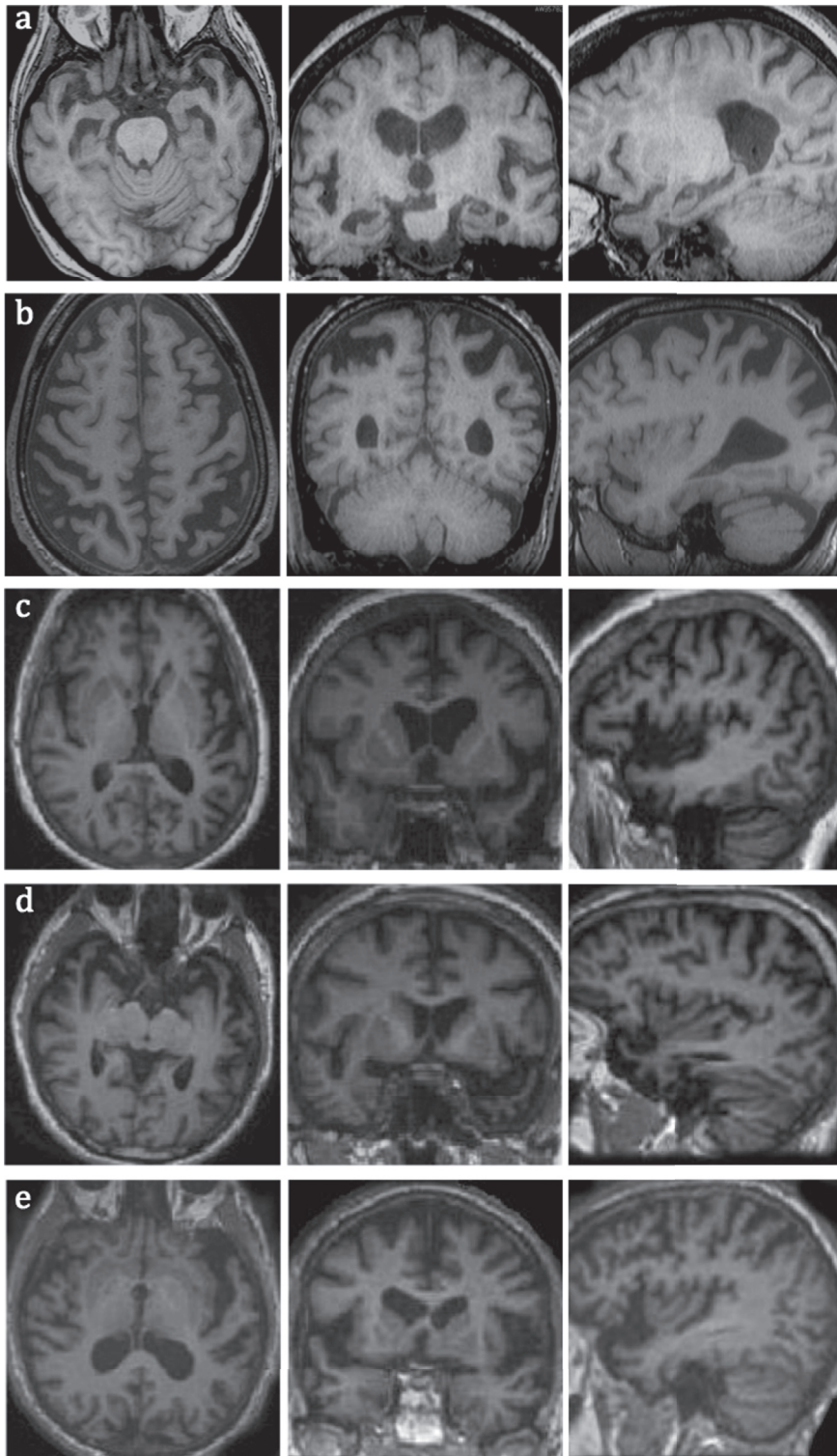


Fig. 3. MRI images of common types of dementia in axial, coronal, and sagittal planes, a) early AD with hippocampal and MTL atrophy; b) PCA with superior posterior parietal lobes atrophy; c) logopenic variant of PPA with atrophy in left temporoparietal junction; note more prominent left sided atrophy of posterior and lateral temporal lobes; d) semantic variant of PPA with atrophy in anterior and lateral temporal lobes; e) non-fluent variant of PPA with left frontal operculum and insular atrophy.

insensitive when CDS is at its early stages (e.g., MCI) and in these circumstances, it is advisable to move to the next tier of imaging depending on availability and clinical suspicion.

FUNCTIONAL IMAGING

Functional imaging includes positron emission tomography (PET) with different tracers, single photon emission computed tomography (SPECT), and advanced MRI techniques such as functional MRI (fMRI), diffusion tensor imaging (DTI), MR spectroscopy (MRS) and arterial spin labeling (ASL). Most of these imaging techniques have recently been developed and some are still far from being used in the routine clinical setting. For the purpose of this review, we focused mainly on the well-established methods such as ^{18}F -fluorodeoxyglucose (^{18}F -FDG) PET, SPECT, and amyloid PET. To make the algorithms more practical, we adapted the new classification scheme for AD biomarkers (A/T/N) introduced by Jack et al. [31]. The utility and role of functional imaging is hugely dependent on the clinical scenario and availability of facilities and expertise but as a general rule they can be used when the combination of clinical examination and structural imaging fail to determine the diagnosis.

FDG-PET

Cerebral hypometabolism is one of the main features of AD. PET with ^{18}F -FDG—a glucose analogue that provides information on the first stages of the glucose metabolism pathway, and a proxy for neuronal and synaptic activity—is a well-established imaging tool for the evaluation of brain function in dementia disorders, including AD.

Current recommendations and guidelines

The European Federation of the Neurological Societies [4, 8], The UK National Institute of Health and Care Excellence [3] and Fourth Canadian Consensus Conference on Diagnosis and Treatment of Dementia [6] all recommended using ^{18}F -FDG-PET in cases with dementia when despite clinical examination and structural imaging by a dementia specialist the diagnosis still remains in doubt, preventing adequate clinical management. There is partial consensus on using FDG in MCI cases. In these cases FDG is useful when the confirmation of a diagnosis can change a patient's clinical management [6]. FDG-PET can also be used as a prognostic biomarker. AD pattern

hypometabolism in FDG-PET in a patient with MCI can predict the conversion to AD within several years [8]. In the US, using FDG-PET for differentiating AD from other dementias (mostly FTD) is approved by Medicare and Medicaid services.

In typical AD, ^{18}F -FDG PET has consistently demonstrated metabolic deficits in the medial temporal lobe, parieto-temporal regions, precuneus, posterior cingulate gyrus, and/or frontal cortices, with sparing of the basal ganglia, thalamus, cerebellum, and visual and sensorimotor cortices [32–35] (Fig. 4). The extent of these hypometabolic patterns during disease progression has been found to correlate with patient performance on cognitive tests [36], and the severity of dementia [35]. In addition, cerebral hypometabolism depicted by ^{18}F -FDG PET has also been shown to correlate with clinical symptoms of cognitive impairment [37] and CSF markers of AD pathology, such as $\text{A}\beta_{42}$ and phosphorylated tau-protein levels [38, 39]. In longitudinal studies, ^{18}F -FDG PET results have shown predictive value in identifying healthy individuals progressing to MCI, and MCI subjects converting to AD [35, 40–42]. Overall sensitivity of FDG is around 76% and median specificity is 82% [43]. Additionally, ^{18}F -FDG-PET can provide differential diagnosis of dementias, allowing discrimination of AD from FTD with more than 85% sensitivity and specificity [35, 44], and AD from DLB with >90% sensitivity and 70% specificity [45, 46].

Acquisition and analysis techniques

Dynamic ^{18}F -FDG PET imaging with arterial blood sampling can provide regional estimates of cerebral glucose metabolic rate (CMR_{glc}) as the most accurate technique for assessing ^{18}F -FDG uptake. Arterial blood sampling, however, is a cumbersome procedure for routine clinical use, due to its invasive nature, associated discomfort and potential risk to the patient. For diagnostic purposes and in clinical settings, PET images can be assessed by visual inspection. This, however, is greatly dependent on the reader's experience. The lack of clear cut-off between normal and abnormal values can make it more challenging particularly in cases with mild disease. Thus, clinical ^{18}F -FDG dementia imaging usually utilizes standardized uptake value ratio (SUV_r) quantification of static imaging typically performed as a 30 min acquisition after an uptake period ranging from 30 to 60 min. During the uptake period, patients are placed in a quiet, dimly-lit room to avoid enhanced cerebral ^{18}F -FDG uptake from neuronal activation.

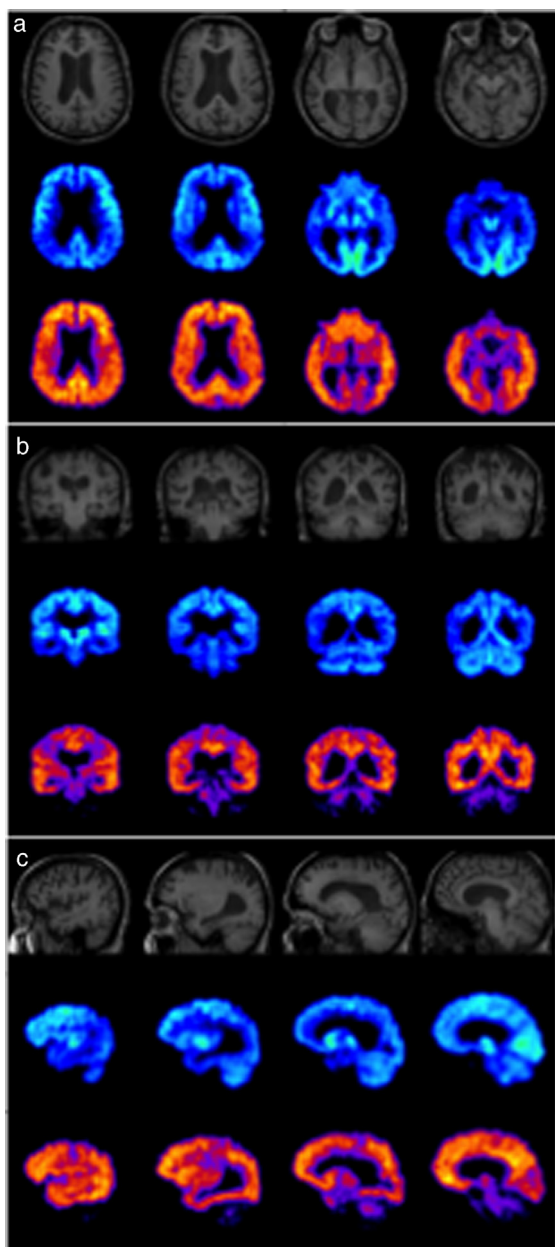


Fig. 4. MRI (top), FDG-PET (middle), and PiB-PET (bottom) images of a typical AD case in axial (a), coronal (b) and sagittal (c) planes showing hypometabolism in parietal and temporal lobes in FDG-PET and extensive cortical amyloid deposition in PiB-PET.

Furthermore, patients are required to fast for at least 4 h prior to tracer administration to minimize plasma glucose-related inhibition of ^{18}F -FDG delivery to tissue. SUVr is determined relative to a region whose metabolism is unaffected or mildly affected in AD; these regions include the cerebellar cortex, vermis or

whole cerebellum, pons, putamen, visual and motor cortices, or a composite of these [47, 48]. SUVr approaches have shown robust results in assessing metabolic deficits in AD, demonstrating good correlations with clinical symptoms, cognitive scores and dementia progression. Recently and in order to facilitate the ^{18}F -FDG-based assessment of AD, two cut points have been defined to dichotomize ^{18}F -FDG-PET data into normal and abnormal categories. The lenient cut point defined by an SUVr of 1.56 is based on the specificity, sensitivity and accuracy of ^{18}F -FDG distinguishing cognitively impaired cases from young healthy controls. The conservative cut point of an SUVr of 1.42 is based on cognitively impaired versus age-matched healthy controls [49]. It should be noted however that classification of an individual above a particular cut value, using either lenient or conservative criteria, does not necessarily preclude the absence of brain pathology.

Clinical implementation

Hypometabolism in ^{18}F -FDG-PET is a biomarker of neurodegeneration, neuronal dysfunction and synaptic disease, which is a relatively late finding in the course of AD [50]. Hence normal ^{18}F -FDG in a case with a suspicious diagnosis of dementia, although makes neurodegeneration very unlikely [8], it cannot exclude early CDS. The result of ^{18}F -FDG-PET in MCI/Preclinical AD can be significantly variable from AD pattern abnormality to cortical hypermetabolism [51, 52]. The pattern of abnormality can also be different according to the clinical presentation (amnesic or non amnesic) or underlying pathology [53]. The utility of ^{18}F -FDG-PET in diagnosis of CDS is summarized in Fig. 5. Although there are distinctive patterns of glucose hypometabolism in different dementia syndromes, i.e., AD, FTD, and DLB, these findings are based on group differences and not at the individual level. Interpretation of a single case can be more difficult as there are overlaps between areas of abnormality across these conditions. As a general rule, involvement of temporoparietal lobes, symmetric or more on the right [54] and posterior cingulate gyrus particularly is more in favor of AD. Anterior cerebral involvement in frontal, anterior temporal lobes and anterior cingulate gyrus is more common in FTD [44, 54], while posterior involvement in occipital lobes particularly primary visual cortex is more seen in DLB [46].

Amyloid-PET (^{11}C -Pittsburgh Compound B and ^{18}F -Labelled Amyloid Tracers)

^{11}C -Pittsburgh Compound B (^{11}C -PiB), a derivative of the amyloid-binding histological dye, thioflavin-T, is the best characterized PET tracer for imaging cerebral amyloid- β (A β) deposition [55, 56]. ^{11}C -PiB has been shown to bind selectively with high affinity to fibrillar A β aggregates, but not amorphous A β deposits or neurofibrillary tangles [57, 58].

PET studies have demonstrated increased cortical binding in brain areas known to be affected by fibrillar A β pathology, notably frontal, temporo-parietal, and posterior cingulate cortices [58, 59] (Fig. 4). Conversely, brain areas with low A β plaque density, such as the cerebellum, have minimal ^{11}C -PiB binding. *In vivo* patterns of ^{11}C -PiB binding to A β fibrils agree with the regional distribution of A β plaques observed by postmortem immunohistochemistry [60, 61] and autopsy [62, 63]. Imaging with ^{11}C -PiB has been reported to differentiate between AD patients and cognitively intact age-matched controls [59, 64], predict progression of MCI to symptomatic AD [65, 66], as well as allow differential diagnosis of dementias [67, 68]. ^{11}C -PiB retention has also been shown to be concordant with levels of CSF A β_{42} , underlining the potential of amyloid PET imaging as a marker of prodromal AD [69]. Compared to ^{18}F -FDG alone, amyloid imaging has demonstrated greater accuracy in the differentiation of patients with AD from healthy controls [70].

Despite the proven efficacy of ^{11}C -PiB in imaging A β -related neuropathology, the short half-life of the ^{11}C label (20 min) has restricted its use to centers with an on-site cyclotron, hampering its use for both research and clinical practice. To overcome the impracticalities of ^{11}C -PiB, several ^{18}F -labelled amyloid derivatives have emerged. Of these, the ^{18}F -labelled PiB analogue, ^{18}F -flutemetamol [71] (GE-067; VizamyTM, GE Healthcare), the stilbene derivative ^{18}F -florbetapir [72] (AV-45; AMYVIDTM, Eli Lilly), and the strylypyridine derivative ^{18}F -florbetaben [73] (BAY-94-9172; NeuraceqTM, Piramal) have all gained approval from the US Food and Drug Administration (FDA) and European Medicines Agency (EMA) for use as diagnostic agents of cerebral amyloid aggregation.

Similar to ^{11}C -PiB, all three compounds have exhibited high specific binding to fibrillar A β plaques *in vivo*, and strong correlations with autopsy and post-mortem pathological findings [74–79]. Although non-specific retention in white matter is higher for

these ^{18}F -labelled tracers than for ^{11}C -PiB, to date this has not presented a major impediment in the qualitative interpretation of images [71–73, 80–82].

Current recommendations and guidelines

The European Federation of the Neurological Societies [8], the Amyloid Imaging Task Force (Society for Nuclear Medicine and Molecular Imaging and the Alzheimer's Association) [83, 84], and the recent Canadian Consensus Conference on the Use of Amyloid Imaging [85] have similar recommendations for clinical utilities of amyloid PET. Similar to ^{18}F -FDG-PET, there is a general consensus that amyloid PET imaging should only be used in dementia expert centers. Amyloid PET is recommended for cases with objectively confirmed cognitive impairment for whom, despite comprehensive clinical/neuropsychological examinations and structural imaging, the diagnosis remains uncertain. Examples include 1) patients with a core clinical diagnosis of possible AD with an atypical course or presentation, early onset of symptoms (age < 65) or cases who present with mixed or heterogeneous syndromes; and 2) patients with persistent or progressive unexplained MCI in whom the knowledge of amyloid status could provide precise diagnosis or alter the management [8, 83–85]. It is inappropriate to use amyloid PET in cognitively normal individuals, based solely on positive family history, genetic testing, APOE4 status, or for nonmedical use [83]. The European Federation of the Neurological Societies also suggested that amyloid PET can be used for differentiating FTD from AD and cerebral amyloid angiopathy from other types of intracranial hemorrhages [8]. Importantly amyloid PET is not useful for staging the severity of a dementia syndrome [85].

Acquisition and analyzing techniques

Imaging with ^{11}C -PiB can be performed either by a dynamic PET acquisition beginning immediately upon tracer injection (typically of 90 min duration), or late static imaging of 20–30 min duration preceded by a tracer uptake period (40 or 50 min) [86] to produce an image that approximates the steady state distribution of tracer concentration in tissue relative to that in plasma. Dynamic imaging is, however, impractical for routine clinical practice. Comparison with kinetic analysis of dynamic ^{11}C -PiB PET, it has been demonstrated that reliable and reproducible quantification of ^{11}C -PiB binding can be obtained via simplified

analysis of static imaging using SUVr, with the cerebellar cortex acting as the reference region [59, 87].

In a clinical setting, static imaging with ^{18}F -florbetapir, ^{18}F -flutemetamol, and ^{18}F -florbetaben can be performed approximately 60, 90, and 130 min, respectively, after tracer administration [72, 88–90]. As for ^{11}C -PiB, reliable A β quantification can be achieved for these ^{18}F -labelled amyloid tracers with 20 min imaging and SUVr analysis relative to the cerebellar cortex. In a recently published meta-analysis of studies with ^{18}F -labelled A β tracers in AD, results indicated favorable specificity and sensitivity in the differentiation of AD patients from age-matched normal controls. Both ^{18}F -florbetapir and ^{18}F -florbetaben demonstrated sensitivity and specificity of over 87%, and though there were insufficient data for complete analysis for ^{18}F -flutemetamol, results from a phase II trial indicated greater than 92% sensitivity and specificity in differentiating AD from cognitively normal subjects [71, 91]. Comparisons between ^{11}C -PiB and ^{18}F -labelled amyloid ligands have also yielded high correlations for SUVr quantification of static imaging in cortical areas, although ^{11}C -PiB is superior in differentiating between controls and AD patients [80–82]. More recently, a common scale—namely Centiloid—for quantifying A β PET data has been developed, so as to allow standardization of results across differing A β radiotracers and imaging protocols [92]. It scales the outcome of each analysis method to a 0 to 100 scale anchored by young controls (≤ 45 years) and typical AD patients. A single cut point of 1.42 SUVr (Centiloid 19) has also been defined for amyloid PET based on the reliable worsening cut point methods [49]

Clinical implementation

While amyloid PET is considered the most sensitive and specific biomarker for AD, its utility in clinical setting is under vigorous evaluation. The most important role of clinical amyloid PET at this stage is to confirm the diagnosis of AD when the clinical presentation is atypical or to differentiate between amyloid associated dementia with non amyloid pathology. With the current therapeutic options, there is no clinical indication to perform amyloid imaging in straightforward advanced AD cases with high clinical probability as the result is most likely positive. Amyloid imaging also cannot differentiate between clinical variants of AD (typical amnesic AD versus posterior cortical atrophy versus logopenic variant of PPA) or AD from other amyloid associated

dementia (DLB or dementia with cerebral amyloid angiopathy). The role of amyloid PET for diagnosis of AD is summarized in Fig. 5. It is worth emphasizing that should AD specific disease modifying therapies become available in the future, the recommendation should change accordingly. This will expand the current application of amyloid PET in clinical settings.

Since deposition of amyloid plaques in brain is one of the first pathological changes in the course of AD [50], amyloid PET can detect brain abnormality at very early stages of the disease or in MCI/preclinical AD cases when the clinical diagnosis is less certain but the intervention will be more effective. The main caveat in using amyloid imaging in preclinical AD is that 10–30% of cognitively normal individuals can have positive amyloid PET [66, 93, 94]. The long-term prognosis in this group and the time scale that they may develop cognitive decline are not completely clear. Therefore, the result of amyloid imaging should be interpreted by dementia experts and based on the cognitive status and clinical presentation of the case. It is very important to remember positive amyloid PET alone does not confirm the diagnosis of AD or MCI.

The disclosure of the results to a patient and caregiver is another sensitive issue that needs special attention. It might be stressful news for patients and can pose legal and social constraints on their life. The Canadian consensus on the use of amyloid imaging [85] has recommended adopting the disclosure methods developed by Harkins et al. [95]. It includes educational session about the results of amyloid imaging and their meaning, assessment of mood and willingness of the participant to receive the results and then face-to-face disclosure session providing written report and explaining the implications with time for questions.

Single photon emission computed tomography

Perfusion hexamethylpropyleneamine oxime (HMPAO)-SPECT measures cerebral blood flow (CBF). It shows hypoperfusion in the regions that demonstrate low glucose metabolism on ^{18}F -FDG-PET such as temporoparietal cortex and posterior cingulate gyrus. A number of guidelines [3, 6, 8] advocate SPECT as an alternative to ^{18}F -FDG-PET but it has been shown that FDG has higher sensitivity and specificity for the diagnosis of AD and DLB [96]. In day-to-day practice, the utility of HMPAO-SPECT (CBF-SPECT) is dependent on

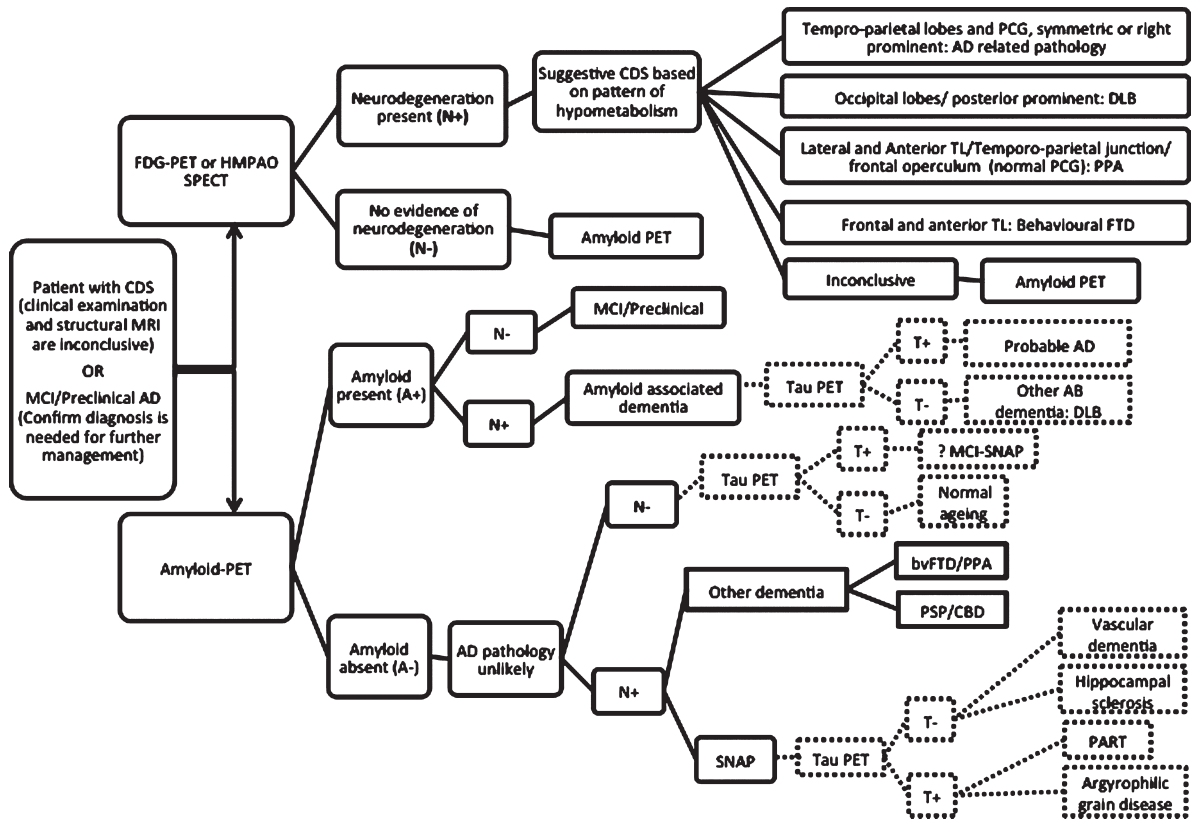


Fig. 5. Stepwise algorithm of PET imaging in the evaluation of AD. Patients who fulfill the criteria for PET imaging, can be imaged by FDG-PET or SPECT. If FDG-PET is inconclusive, amyloid imaging will be the next step. The dashed lines are demonstrating the role of tau PET when it becomes clinically available.

the availability of SPECT or ^{18}F -FDG-PET in each center. Currently the UK National Institute of Health and Care Excellence recommend HMPAO-SPECT to differentiated AD from vascular dementia and FTD prior to FDG [3]. While the Fourth Canadian Consensus Conference on Diagnosis and Treatment of Dementia [6] suggested using SPECT as an alternative to FDG when PET cannot be performed.

Dopaminergic iodine-123-radiolabelled 2₋-carbo methoxy-3₋-(4-iodophenyl)-N-(3-fluoropropyl) nortropane (FP-CIT) SPECT known commercially as DaTSCANTM (GE Healthcare, Waukesha, WI, USA) depicts striatal uptake of dopamine. Low dopamine transporter uptake in basal ganglia is a characteristic feature of DLB. Dopaminergic SPECT has very high sensitivity and specificity for diagnosing DLB [97], however it cannot distinguish DLB from other nigrostriatal neurodegenerative disorders associated with presynaptic dopaminergic deficiency such as PDD, PSP, CBD, or FTD with Parkinsonism. Both The UK National Institute of Health and Care Excellence [3] and The European Federation of

the Neurological Societies [8] recommend using dopaminergic SPECT in diagnosis of cases with suspected dementia with Lewy bodies.

Other imaging techniques

Tau-PET

Despite the high sensitivity of amyloid imaging in diagnosis of AD, there is a 15–20 year time lag between A β deposition and clinical symptoms in AD. By the time patients become symptomatic, the amyloid load has reached its plateau phase which means amyloid PET cannot be used as a staging or prognostic biomarker. On the other hand, the presence of neurofibrillary tangles, phosphorylated tau protein aggregates, in AD has close association with cognitive decline [98], severity of dementia symptoms [99], neuronal injury [100], and brain atrophy [101]. Therefore, tau imaging can be used as a surrogate marker to predict cognitive decline or disease progression in AD. Tau aggregation and deposition in AD occurs in a stereotypical spatiotemporal

pattern starting from transentorhinal/entorhinal cortex to hippocampus and then extending to the rest of temporal lobe and neocortical regions [102]. Widespread neocortical deposition is not common in cognitively normal individuals and this stepwise pattern of deposition can help to stage AD. The potential role of tau PET is depicted in Fig. 5. When patient is amyloid positive (A+) but doesn't have any evidence of neurodegeneration on FDG-PET or structural MRI (N-), presence of tau will increase the likelihood of AD or determine more advanced stages of the disease. On the other hand, if a patient is A+ and N+ without any evidence of tau, one should consider neurodegeneration as a consequence of non AD pathology and possibility of a mixed dementia syndrome.

Tau imaging might also be useful to facilitate the diagnosis of non-amyloid dependent tau related dementias including PSP, CBD, PPA, behavioral FTD, and a group of other pathologies defined as suspected non Alzheimer's disease pathologies (SNAP). Examples of SNAP conditions include argyrophilic grain disease and primary age-related tauopathy [103]. The distribution pattern of tau aggregates in different tauopathies is variable: widespread frontotemporal distribution in FTD, subthalamic nucleus and basal ganglia in PSP, neocortical areas and striatum in CBD, dentate gyrus and hippocampus in Pick's disease, amygdala and entorhinal cortex and hippocampus in argyrophilic grain disease, and mesial temporal lobe in primary age-related tauopathy [104].

In recent years, different tau selective PET tracers have been developed and been used for human studies: [¹⁸F]THK523, [¹⁸F]THK5117, [¹⁸F]THK5105 [¹⁸F]THK5351, [¹⁸F]AV1451(T807), and [¹¹C] PBB3 [105]. Although the initial results are promising, there is still long way before tau PET could be qualify for clinical use. If AD specific disease modifying therapies become available, then the indications of performing tau imaging may also change. Tau PET can be used as a follow up imaging tool to assess the response to treatment and predicting future clinical progression.

MR spectroscopy

MRS is a non-invasive method for measuring the concentration of certain chemicals in the brain. N-acetylaspartate (NAA), myoinositol (mI), Choline (Chol), Glutamate plus Glutamine (Glx), and Creatine (Cr) are the most commonly measured compounds. A number of studies in the past two decades have investigated the role of MRS in

neurodegenerative disorders. The rationale is that brain metabolites act as surrogate markers for different pathological processes such as neural damage, glial proliferation, loss of membrane integrity, and even inflammatory changes. MRS has the potential to be used as an imaging biomarker detecting the early metabolic changes in AD [106].

Since there is no significant change in the level of Cr in different conditions, it is commonly used as an internal reference to overcome any biases. Therefore, the result of MRS is usually reported as a ratio of other metabolites over Cr [107, 108]. Decrease in NAA or NAA/Cr, a marker of neuroaxonal density and viability, is one of the main reported findings in AD [109, 110]. Reduction of NAA has been reported in the medial temporal lobe [111], hippocampi [110], and posterior cingulate gyrus [112].

In addition to NAA reduction, increase in mI in several brain regions is considered another feature of AD [107, 109]. mI is a glial marker and increase in the level of mI can be a sign of increased glial content, size, and activation [108]. Increase in mI or mI/Cr is considered an early event in the course of AD pathology that can precede NAA reduction [109]. It has been shown that detection of NAA reduction and mI increase in suspected AD improves the specificity and accuracy of the clinical diagnosis significantly (100% for distinguishing AD from healthy controls) [113]. Based on the observed correlation between the metabolites' level and histopathological changes in AD, it has been suggested that reduction in NAA is related to early tau-mediated dynamic processes while increase in mI is associated with A β deposition [114].

Unlike NAA and mI, other metabolites including Chol [109, 113, 115] and Glx [112, 116] have shown inconsistent changes in AD. Lack of standardized techniques and choosing different brain regions as region of interest might be responsible for these contradictory results.

In comparison to other functional imaging techniques, MRS is more available and much less expensive with no radiation risk. It can be easily added to the structural MRI sequences and extract very useful functional information to help diagnosis. Moreover with the correlation between the metabolites level and pathological changes in AD, MRS can be utilized as a follow up imaging tool in therapeutic trials. However, to use MRS for clinical purpose, further large cohort research is required to standardize the techniques and compare the results of MRS with other functional biomarkers.

Functional MRI and diffuse tensor imaging

Neurodegenerative diseases are associated with synaptic dysfunction, axonal, and white matter tract loss with connectivity disruption, and brain network alteration. Advanced MRI techniques like fMRI and DTI can provide further information about these microstructure properties of the brain, its connectivity and networks.

fMRI can be performed as either rest-state (RS)- or task-based. RS-fMRI focuses on spontaneous low frequency fluctuations in blood-oxygen-level dependent signal. Around 10–20 RS networks have been described and the most common ones are default mode network (DMN), the salience network, and central executive network [117]. Different studies have shown that there are functional alteration in all these three networks, particularly DMN in AD and MCI when compared to healthy controls [118]. Also in a meta-analysis of task-based fMRI, MCI and AD patients presented different patterns of hypo-activity; frontoparietal and DMN were affected in MCI and visual network demonstrated low activity in AD cases relative to controls. This implies that pattern of neuronal network dysfunction might be a consequence of disease progression [119].

DTI measures the directionality of water diffusion. In the white matter tracts, water diffusion follows the orientation of the axonal bundles [120]. DTI studies in AD have shown white matter fiber alteration in temporal and frontal lobes and also corpus callosum and posterior cingulate gyrus [121] with a posterior to anterior gradient [122]. A meta-analysis showed DTI measurements of limbic regions have modest diagnostic power in discriminating AD from controls which was not significantly different to hippocampal atrophy [123].

These functional alterations appear prior to structural changes in MRI and could be used as an indicator of disease progression and to monitor response to disease modifying drugs. The caveat is that these techniques are at early stages of evaluation and require further large cohort studies. Moreover, there is significant variability of DTI-based diffusion metrics between MRI scanners that imposes a major restriction in multicenter studies [124]. These limitations mean there is no prospect of using these methods in clinical setting in the near future.

Arterial spin labeling

ASL is a non-invasive MRI technique for measuring tissue perfusion (CBF) by using magnetically labeled arterial blood water protons as an endogenous

tracer [125]. It is known that hypoperfusion of AD vulnerable regions starts well before clinical symptoms. There is high correlation between reduction in perfusion and glucose metabolism in ^{18}F -FDG-PET. Some take this further and suggest that vascular dysfunction might precede other abnormalities in AD and present even prior to A β aggregation [126].

Recently, several studies have investigated the correlation between ASL and FDG-PET/SPECT in AD. These have shown good concordance between hypoperfusion in ASL and other established techniques with comparable diagnostic accuracy [127]. The results in MCI cases however, were more contradictory as both hyper- and hypoperfusion have been reported [126].

Although ASL, with no radiation risk, lower cost and wider availability, can potentially be a valuable technique to measure brain perfusion, it still has some limitations for wide clinical application: ASL is sensitive to blood velocity and arterial transit time; images are dependent on the integrity of the cerebrovascular system and therefore, presence of steno-occlusive disease or other cerebrovascular pathology can interfere with the quality of image and lead to artefact or noise; it is also highly affected by head motion [127]. New methods have been developed to overcome some of these limitations but further research studies are required before ASL can be considered as a diagnostic biomarker for AD in clinical setting.

CONCLUSION

- 1) Imaging biomarkers are an essential part of workup in CDS. Appropriate and informed application of imaging can help clinicians in making early diagnosis or dealing with atypical cases of dementia.
- 2) Structural imaging (MRI or high resolution CT) should be the first tier of imaging in CDS. Using dementia specific protocol is very crucial for both diagnosis and follow up. Hippocampal and MTL atrophy can be evaluated visually or using quantitative techniques. Visual dichotomized assessment (atrophic vs not atrophic) has good reliability.
- 3) Functional imaging can provide further information about the pathophysiology of the AD but their utility in day-to-day practice is still under investigation. Currently, they are recommended when clinical examination and structural imaging cannot confirm the diagnosis.

- 4) Hypometabolism in ^{18}F -FDG-PET, an indicator of neurodegeneration, can differentiate dementia from normal aging. Absence of significant hypometabolism however, will not exclude AD pathology completely particularly at an early stage.
- 5) Although ^{18}F -FDG-PET is more available compared to amyloid PET, interpretation of ^{18}F -FDG PET can be more challenging. Standardized cut point and development of new analysis software might help wider group of dementia experts to use FDG in their routine practice.
- 6) Amyloid imaging will be more accessible in the near future with the new fluorinated amyloid tracers. Therefore, it is crucial for clinicians to know the indications of amyloid PET in the clinical setting. The result of PET imaging should always be interpreted by dementia experts and within the context of the clinical examination. Positive amyloid PET (A+) alone is not equal to AD while A- makes AD pathology very unlikely.
- 7) Other functional imaging techniques have shown promising results but are still far from being used in the clinical setting.

CONFLICT OF INTEREST

The authors have no conflict of interest to report.

REFERENCES

- [1] Siemers ER, Sundell KL, Carlson C, Case M, Sethuraman G, Liu-Seifert H, Dowsett SA, Pontecorvo MJ, Dean RA, Demattos R (2016) Phase 3 solanezumab trials: Secondary outcomes in mild Alzheimer's disease patients. *Alzheimers Dement* **12**, 110-120.
- [2] Falahati F, Westman E, Simmons A (2014) Multivariate data analysis and machine learning in Alzheimer's disease with a focus on structural magnetic resonance imaging. *J Alzheimers Dis* **41**, 685-708.
- [3] NICE (2007) National Collaborating Centre for Mental Health, National Institute for Health and Clinical Excellence: Guidance. In *Dementia: A NICE-SCIE Guideline on Supporting People With Dementia and Their Carers in Health and Social Care*. British Psychological Society, The British Psychological Society & The Royal College of Psychiatrists., Leicester (UK).
- [4] Hort J, O'Brien JT, Gainotti G, Pirttila T, Popescu BO, Rektorova I, Sorbi S, Scheltens P (2010) EFNS guidelines for the diagnosis and management of Alzheimer's disease. *Eur J Neurol* **17**, 1236-1248.
- [5] Scheltens P, Fox N, Barkhof F, De Carli C (2002) Structural magnetic resonance imaging in the practical assessment of dementia: Beyond exclusion. *Lancet Neurol* **1**, 13-21.
- [6] Soucy JP, Bartha R, Bocti C, Borrie M, Burhan AM, Laforce R, Rosa-Neto P (2013) Clinical applications of neuroimaging in patients with Alzheimer's disease: A review from the Fourth Canadian Consensus Conference on the Diagnosis and Treatment of Dementia 2012. *Alzheimers Res Ther* **5**, S3.
- [7] Jack CR, Albert M, Knopman DS, McKhann GM, Sperling RA, Carillo M, Thies W, Phelps CH (2011) Introduction to Revised Criteria for the Diagnosis of Alzheimer's Disease: National Institute on Aging and the Alzheimer Association Workgroups. *Alzheimers Dement* **7**, 257-262.
- [8] Filippi M, Agosta F, Barkhof F, Dubois B, Fox NC, Frisoni GB, Jack CR, Johannsen P, Miller BL, Nestor PJ, Scheltens P, Sorbi S, Teipel S, Thompson PM, Wahlund LO (2012) EFNS task force: The use of neuroimaging in the diagnosis of dementia. *Eur J Neurol* **19**, e131-140, 1487-1501.
- [9] Dickerson BC, Bakkour A, Salat DH, Feczko E, Pacheco J, Greve DN, Grodstein F, Wright CI, Blacker D, Rosas HD, Sperling RA, Atri A, Growdon JH, Hyman BT, Morris JC, Fischl B, Buckner RL (2009) The cortical signature of Alzheimer's disease: Regionally specific cortical thinning relates to symptom severity in very mild to mild AD dementia and is detectable in asymptomatic amyloid-positive individuals. *Cereb Cortex* **19**, 497-510.
- [10] Davis PC, Gearing M, Gray L, Mirra SS, Morris JC, Edland SD, Lin T, Heyman A (1995) The CERAD experience, Part VIII: Neuroimaging-neuropathology correlates of temporal lobe changes in Alzheimer's disease. *Neurology* **45**, 178-179.
- [11] Frisoni GB, Prestia A, Rasser PE, Bonetti M, Thompson PM (2009) In vivo mapping of incremental cortical atrophy from incipient to overt Alzheimer's disease. *J Neurol* **256**, 916-924.
- [12] Cordonnier C, van der Flier WM (2011) Brain microbleeds and Alzheimer's disease: Innocent observation or key player? *Brain* **134**, 335-344.
- [13] Rinne JO, Brooks DJ, Rossor MN, Fox NC, Bullock R, Klunk WE, Mathis CA, Blennow K, Barakos J, Okello AA, Rodriguez Martinez de Liano S, Liu E, Koller M, Gregg KM, Schenk D, Black R, Grundman M (2010) 11C-PiB PET assessment of change in fibrillar amyloid-beta load in patients with Alzheimer's disease treated with bapineuzumab: A phase 2, double-blind, placebo-controlled, ascending-dose study. *Lancet Neurol* **9**, 363-372.
- [14] Scheltens P, Leys D, Barkhof F (1992) Atrophy of medial temporal lobes on MRI in "probable" Alzheimer's disease and normal ageing: Diagnostic value and neuropsychological correlates. *J Neurol* **8**, 967-972.
- [15] Scheltens P, Launer LJ, Barkhof F (1995) Visual assessment of medial temporal lobe atrophy on magnetic resonance imaging: Interobserver reliability. *J Neurol* **242**, 557-560.
- [16] Scheltens P, Pasquier F, Weerts JGE, Barkhof F, Leys D (2016) Qualitative assessment of cerebral atrophy on MRI: Inter- and intra-observer reproducibility in dementia and normal aging. *Eur Neurol* **37**, 95-99.
- [17] Pereira JB, Cavallin L, Spulber G, Aguilar C, Mecocci P, Vellas B, Tsolaki M, Kloszewska I, Soinen H, Spenger C, Aarsland D, Lovestone S, Simmons A, Wahlund LO, Westman E (2016) Influence of age, disease onset and ApoE4 on visual medial temporal lobe atrophy cut-offs. *J Intern Med* **275**, 317-330.
- [18] Frisoni GB, Geroldi C, Beltramello A, Bianchetti A, Binetti G, Bordiga G, DeCarli C, Laakso MP, Soinen H,

- Testa C, Zanetti O, Trabucchi M (2002) Radial width of the temporal horn: A sensitive measure in Alzheimer disease. *AJNR Am J Neuroradiol* **23**, 35-47.
- [19] Dahlbeck JW, McCluney KW, Yeakley JW, Fenstermacher MJ, Bonmati C, Van Horn G, 3rd, Aldag J (1991) The interuncal distance: A new MR measurement for the hippocampal atrophy of Alzheimer disease. *AJNR Am J Neuroradiol* **12**, 931-932.
- [20] Erkinjuntti T, Lee DH, Gao F, Steenhuis R, Eliasziw M, Fry R, Merskey H, Hachinski VC (2016) Temporal lobe atrophy on magnetic resonance imaging in the diagnosis of early Alzheimer's disease. *Arch Neurol* **50**, 305-310.
- [21] Menendez-Gonzalez M, Lopez-Muniz A, Vega JA, Salas-Pacheco JM, Arias-Carrion O (2014) MTA index: A simple 2D-method for assessing atrophy of the medial temporal lobe using clinically available neuroimaging. *Front Aging Neurosci* **6**, 23.
- [22] Conejo Bayon F, Maese J, Fernandez Oliveira A, Mesas T, Herrera de la Llave E, Alvarez Avellon T, Menendez-Gonzalez M (2014) Feasibility of the Medial Temporal lobe Atrophy index (MTAI) and derived methods for measuring atrophy of the medial temporal lobe. *Front Aging Neurosci* **6**, 305.
- [23] Menéndez-González M, Oliveira AF, Bayón FC, Maese J, Uzal TM, Llave EHdl, Avellón TÁ (2015) Planimetry of the medial temporal lobe: A feasible method for supporting the diagnosis of Alzheimer's disease in clinical practice. *Neurol Neurosci*, doi: 10.3823-355
- [24] Menéndez-González M, Suárez-Sanmartin E, García C, Martínez-Cambor P, Westman E, Simmons A (2016) Manual planimetry of the medial temporal lobe versus automated volumetry of the hippocampus in the diagnosis of Alzheimer's disease. *Cureus* **8**, e544-e544.
- [25] Pini L, Pievani M, Bocchetta M, Altomare D, Bosco P, Cavedo E, Galluzzi S, Marizzoni M, Frisoni GB (2016) Brain atrophy in Alzheimer's Disease and aging. *Ageing Res Rev* **30**, 25-48.
- [26] Frisoni GB, Jack CR (2011) Harmonization of magnetic resonance-based manual hippocampal segmentation: A mandatory step for wide clinical use. *Alzheimers Dement* **7**, 171-174.
- [27] Jack CR Jr, Barkhof F, Bernstein MA, Cantillon M, Cole PE, Decarli C, Dubois B, Duchesne S, Fox NC, Frisoni GB, Hampel H, Hill DL, Johnson K, Mangin JF, Scheltens P, Schwarz AJ, Sperling R, Suhy J, Thompson PM, Weiner M, Foster NL (2011) Steps to standardization and validation of hippocampal volumetry as a biomarker in clinical trials and diagnostic criterion for Alzheimer's disease. *Alzheimers Dement* **7**, 474-485.e474.
- [28] Schröder J, Pantel J (2016) Neuroimaging of hippocampal atrophy in early recognition of Alzheimer's disease—a critical appraisal after two decades of research. *Psychiatry Res* **247**, 71-78.
- [29] Westman E, Cavallin L, Muehlboeck JS, Zhang Y, Mecocci P, Vellas B, Tsolaki M, Kloszewska I, Soininen H, Spenger C, Lovestone S, Simmons A, Wahlund LO (2011) Sensitivity and specificity of medial temporal lobe visual ratings and multivariate regional MRI classification in Alzheimer's disease. *PLoS One* **6**, e22506.
- [30] McKhann GM, Knopman DS (2011) The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging- Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* **7**, 263-269.
- [31] Jack CR, Bennett DA, Blennow K (2016) A/T/N: An unbiased descriptive classification scheme for Alzheimer disease biomarkers. *Neurology* **87**, 539-547.
- [32] Minoshima S, Giordani B, Berent S, Frey KA, Foster NL, Kuhl DE (1997) Metabolic reduction in the posterior cingulate cortex in very early Alzheimer's disease. *Ann Neurol* **42**, 85-94.
- [33] Silverman DH, Small GW, Phelps ME (1999) Clinical value of neuroimaging in the diagnosis of dementia. Sensitivity and specificity of regional cerebral metabolic and other parameters for early identification of Alzheimer's disease. *Clin Positron Imaging* **2**, 119-130.
- [34] de Leon MJ, Convit A, Wolf OT, Tarshish CY, DeSanti S, Rusinek H, Tsui W, Kandil E, Scherer AJ, Roche A, Imossi A, Thorn E, Bobinski M, Caraos C, Lesbre P, Schlyer D, Poirier J, Reisberg B, Fowler J (2001) Prediction of cognitive decline in normal elderly subjects with 2-[(18)F]fluoro-2-deoxy-D-glucose/positron-emission tomography (FDG/PET). *Proc Natl Acad Sci U S A* **98**, 10966-10971.
- [35] Mosconi L (2005) Brain glucose metabolism in the early and specific diagnosis of Alzheimer's disease. FDG-PET studies in MCI and AD. *Eur J Nucl Med Mol Imaging* **32**, 486-510.
- [36] Engler H, Forsberg A, Almkvist O, Blomquist G, Larsson E, Savitcheva I, Wall A, Ringheim A, Langstrom B, Nordberg A (2006) Two-year follow-up of amyloid deposition in patients with Alzheimer's disease. *Brain* **129**, 2856-2866.
- [37] Mielke R, Herholz K, Grond M, Kessler J, Heiss WD (1994) Clinical deterioration in probable Alzheimer's disease correlates with progressive metabolic impairment of association areas. *Dementia* **5**, 36-41.
- [38] Arlt S, Brassens S, Jahn H, Wilke F, Eichenlaub M, Apostolova I, Wenzel F, Thiele F, Young S, Buchert R (2009) Association between FDG uptake, CSF biomarkers and cognitive performance in patients with probable Alzheimer's disease. *Eur J Nucl Med Mol Imaging* **36**, 1090-1100.
- [39] Ceravolo R, Borghetti D, Kiferle L, Tognoni G, Giorgetti A, Neglia D, Sassi N, Frosini D, Rossi C, Petrozzi L, Siciliano G, Murri L (2008) CSF phosphorylated TAU protein levels correlate with cerebral glucose metabolism assessed with PET in Alzheimer's disease. *Brain Res Bull* **76**, 80-84.
- [40] Drzezga A, Lautenschlager N, Siebner H, Riemenschneider M, Willoch F, Minoshima S, Schwaiger M, Kurz A (2003) Cerebral metabolic changes accompanying conversion of mild cognitive impairment into Alzheimer's disease: A PET follow-up study. *Eur J Nucl Med Mol Imaging* **30**, 1104-1113.
- [41] Drzezga A, Grimmer T, Riemenschneider M, Lautenschlager N, Siebner H, Alexopoulos P, Minoshima S, Schwaiger M, Kurz A (2005) Prediction of individual clinical outcome in MCI by means of genetic assessment and (18)F-FDG PET. *J Nucl Med* **46**, 1625-1632.
- [42] Mosconi L, Brys M, Glodzik-Sobanska L, De Santi S, Rusinek H, de Leon MJ (2007) Early detection of Alzheimer's disease using neuroimaging. *Exp Gerontol* **42**, 129-138.
- [43] Smailagic N, Vacante M, Hyde C, Martin S, Ukoumunne O, Sachpekidis C (2015) (1)(8)F-FDG PET for the early diagnosis of Alzheimer's disease dementia and other dementias in people with mild cognitive impairment (MCI). *Cochrane Database Syst Rev* **1**, CD010632.

- [44] Rabinovici GD, Rosen HJ, Alkalay A, Kornak J, Furst AJ, Agarwal N, Mormino EC, O'Neil JP, Janabi M, Karydas A, Growdon ME, Jang JY, Huang EJ, Dearmond SJ, Trojanowski JQ, Grinberg LT, Gorno-Tempini ML, Seeley WW, Miller BL, Jagust WJ (2011) Amyloid vs FDG-PET in the differential diagnosis of AD and FTLD. *Neurology* **77**, 2034-2042.
- [45] Lim SM, Katsifis A, Villemagne VL, Best R, Jones G, Salting M, Bradshaw J, Merory J, Woodward M, Hopwood M, Rowe CC (2009) The 18F-FDG PET cingulate island sign and comparison to 123I-beta-CIT SPECT for diagnosis of dementia with Lewy bodies. *J Nucl Med* **50**, 1638-1645.
- [46] Kono AK, Ishii K, Sofue K, Miyamoto N, Sakamoto S, Mori E (2007) Fully automatic differential diagnosis system for dementia with Lewy bodies and Alzheimer's disease using FDG-PET and 3D-SSP. *Eur J Nucl Med Mol Imaging* **34**, 1490-1497.
- [47] Herholz K, Perani D, Salmon E, Franck G, Fazio F, Heiss WD, Comar D (1993) Comparability of FDG PET studies in probable Alzheimer's disease. *J Nucl Med* **34**, 1460-1466.
- [48] Herholz K (2014) The role of PET quantification in neurological imaging: FDG and amyloid imaging in dementia. *Clin Transl Imaging* **2**, 321-330.
- [49] Jack CR Jr, Wiste HJ, Weigand SD, Thorneau TM, Lowe VJ, Knopman DS, Gunter JL, Senjem ML, Jones DT, Kantarci K, Machulda MM, Mielke MM, Roberts RO, Vemuri P, Reyes DA, Petersen RC (2017) Defining imaging biomarker cut points for brain aging and Alzheimer's disease. *Alzheimers Dement* **13**, 205-216.
- [50] Jack CR Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, Shaw LM, Vemuri P, Wiste HJ, Weigand SD, Lesnick TG, Pankratz VS, Donohue MC, Trojanowski JQ (2013) Tracking pathophysiological processes in Alzheimer's disease: An updated hypothetical model of dynamic biomarkers. *Lancet Neurol* **12**, 207-216.
- [51] Cohen AD, Price JC, Weissfeld LA, James J, Rosario BL, Bi W, Nebes RD, Saxton JA, Snitz BE, Aizenstein HA, Wolk DA, DeKosky ST, Mathis CA, Klunk WE (2009) Basal cerebral metabolism may modulate the cognitive effects of A β in mild cognitive impairment: An example of brain reserve. *J Neurosci* **29**, 14770.
- [52] Johnson SC, Christian BT, Okonko OC, Oh JM, Harding S, Xu G, Hillmer AT, Wooten DW, Murali D, Barnhart TE, Hall LT, Racine AM, Klunk WE, Mathis CA, Bendlin BB, Gallagher CL, Carlsson CM, Rowley HA, Hermann BP, Dowling NM, Asthana S, Sager MA (2014) Amyloid burden and neural function in people at risk for Alzheimer's Disease. *Neurobiol Aging* **35**, 576-584.
- [53] Coutinho AM, Porto FH, Duran FL, Prando S, Ono CR, Feitosa EA, Spindola L, de Oliveira MO, do Vale PH, Gomes HR, Nitri R, Brucki SM, Buchpiguel CA (2015) Brain metabolism and cerebrospinal fluid biomarkers profile of non-amnesic mild cognitive impairment in comparison to amnesic mild cognitive impairment and normal older subjects. *Alzheimers Res Ther* **7**, 58.
- [54] Foster NL, Heidebrink JL, Clark CM, Jagust WJ, Arnold SE, Barbas NR, DeCarli CS, Turner RS, Koeppe RA, Higdon R, Minoshima S (2007) FDG-PET improves accuracy in distinguishing frontotemporal dementia and Alzheimer's disease. *Brain* **130**, 2616-2635.
- [55] Klunk WE, Wang Y, Huang GF, Debnath ML, Holt DP, Mathis CA (2001) Uncharged thioflavin-T derivatives bind to amyloid-beta protein with high affinity and readily enter the brain. *Life Sci* **69**, 1471-1484.
- [56] Mathis CA, Bacskai BJ, Kajdasz ST, McLellan ME, Frosch MP, Hyman BT, Holt DP, Wang Y, Huang GF, Debnath ML, Klunk WE (2002) A lipophilic thioflavin-T derivative for positron emission tomography (PET) imaging of amyloid in brain. *Bioorg Med Chem Lett* **12**, 295-298.
- [57] Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP, Bergstrom M, Savitcheva I, Huang GF, Estrada S, Aussen B, Debnath ML, Barletta J, Price JC, Sandell J, Lopresti BJ, Wall A, Koivisto P, Antoni G, Mathis CA, Langstrom B (2004) Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann Neurol* **55**, 306-319.
- [58] Klunk WE, Wang Y, Huang GF, Debnath ML, Holt DP, Shao L, Hamilton RL, Ikonovic MD, DeKosky ST, Mathis CA (2003) The binding of 2-(4'-methylaminophenyl)benzothiazole to postmortem brain homogenates is dominated by the amyloid component. *J Neurosci* **23**, 2086-2092.
- [59] Price JC, Klunk WE, Lopresti BJ, Lu X, Hoge JA, Ziolkowski SK, Holt DP, Meltzer CC, DeKosky ST, Mathis CA (2005) Kinetic modeling of amyloid binding in humans using PET imaging and Pittsburgh Compound-B. *J Cereb Blood Flow Metab* **25**, 1528-1547.
- [60] Ikonovic MD, Klunk WE, Abrahamson EE, Mathis CA, Price JC, Tsopelas ND, Lopresti BJ, Ziolkowski S, Bi W, Paljug WR, Debnath ML, Hope CE, Isanski BA, Hamilton RL, DeKosky ST (2008) Post-mortem correlates of in vivo PiB-PET amyloid imaging in a typical case of Alzheimer's disease. *Brain* **131**, 1630-1645.
- [61] Beckett TL, Webb RL, Niedowicz DM, Holler CJ, Matveev S, Baig I, LeVine H, 3rd, Keller JN, Murphy MP (2012) Postmortem Pittsburgh Compound B (PiB) binding increases with Alzheimer's disease progression. *J Alzheimers Dis* **32**, 127-138.
- [62] Bacskai BJ, Frosch MP, Freeman SH, Raymond SB, Augustinack JC, Johnson KA, Irizarry MC, Klunk WE, Mathis CA, DeKosky ST, Greenberg SM, Hyman BT, Growdon JH (2007) Molecular imaging with Pittsburgh Compound B confirmed at autopsy: A case report. *Arch Neurol* **64**, 431-434.
- [63] Leinonen V, Alafuzoff I, Aalto S, Suotunen T, Savolainen S, Nagren K, Tapiola T, Pirttila T, Rinne J, Jaaskelainen JE, Soininen H, Rinne JO (2008) Assessment of beta-amyloid in a frontal cortical brain biopsy specimen and by positron emission tomography with carbon 11-labeled Pittsburgh Compound B. *Arch Neurol* **65**, 1304-1309.
- [64] Archer HA, Edison P, Brooks DJ, Barnes J, Frost C, Yeatman T, Fox NC, Rossor MN (2006) Amyloid load and cerebral atrophy in Alzheimer's disease: An 11C-PIB positron emission tomography study. *Ann Neurol* **60**, 145-147.
- [65] Morris JC, Roe CM, Grant EA, Head D, Storandt M, Goate AM, Fagan AM, Holtzman DM, Mintun MA (2009) Pittsburgh compound B imaging and prediction of progression from cognitive normality to symptomatic Alzheimer disease. *Arch Neurol* **66**, 1469-1475.
- [66] Forsberg A, Engler H, Almkvist O, Blomqvist G, Hagman G, Wall A, Ringheim A, Langstrom B, Nordberg A (2008) PET imaging of amyloid deposition in patients with mild cognitive impairment. *Neurobiol Aging* **29**, 1456-1465.
- [67] Laforce R Jr, Rabinovici GD (2011) Amyloid imaging in the differential diagnosis of dementia: Review and potential clinical applications. *Alzheimers Res Ther* **3**, 31.

- [68] Rowe CC, Ng S, Ackermann U, Gong SJ, Pike K, Savage G, Cowie TF, Dickinson KL, Maruff P, Darby D, Smith C, Woodward M, Merory J, Tochon-Danguy H, O'Keefe G, Klunk WE, Mathis CA, Price JC, Masters CL, Villemagne VL (2007) Imaging beta-amyloid burden in aging and dementia. *Neurology* **68**, 1718-1725.
- [69] Fagan AM, Mintun MA, Mach RH, Lee SY, Dence CS, Shah AR, LaRossa GN, Spinner ML, Klunk WE, Mathis CA, DeKosky ST, Morris JC, Holtzman DM (2006) Inverse relation between in vivo amyloid imaging load and cerebrospinal fluid Abeta42 in humans. *Ann Neurol* **59**, 512-519.
- [70] Ng S, Villemagne VL, Berlangieri S, Lee ST, Cherk M, Gong SJ, Ackermann U, Saunderson T, Tochon-Danguy H, Jones G, Smith C, O'Keefe G, Masters CL, Rowe CC (2007) Visual assessment versus quantitative assessment of 11C-PIB PET and 18F-FDG PET for detection of Alzheimer's disease. *J Nucl Med* **48**, 547-552.
- [71] Vandenberghe R, Van Laere K, Ivanoiu A, Salmon E, Bastin C, Triau E, Hasselbalch S, Law I, Andersen A, Korner A, Minthon L, Garraux G, Nelissen N, Bormans G, Buckley C, Owenius R, Thurfjell L, Farrar G, Brooks DJ (2010) 18F-flutemetamol amyloid imaging in Alzheimer disease and mild cognitive impairment: A phase 2 trial. *Ann Neurol* **68**, 319-329.
- [72] Wong DF, Rosenberg PB, Zhou Y, Kumar A, Raymond V, Ravert HT, Dannals RF, Nandi A, Brasic JR, Ye W, Hilton J, Lyketsos C, Kung HF, Joshi AD, Skovronsky DM, Pontecorvo MJ (2010) In vivo imaging of amyloid deposition in Alzheimer disease using the radioligand 18F-AV-45 (florbetapir [corrected] F 18). *J Nucl Med* **51**, 913-920.
- [73] Rowe CC, Ackerman U, Browne W, Mulligan R, Pike KL, O'Keefe G, Tochon-Danguy H, Chan G, Berlangieri SU, Jones G, Dickinson-Rowe KL, Kung HP, Zhang W, Kung MP, Skovronsky D, Dyrks T, Holl G, Krause S, Friebe M, Lehman L, Lindemann S, Dinkelborg LM, Masters CL, Villemagne VL (2008) Imaging of amyloid beta in Alzheimer's disease with 18F-BAY94-9172, a novel PET tracer: Proof of mechanism. *Lancet Neurol* **7**, 129-135.
- [74] Zhang W, Kung MP, Oya S, Hou C, Kung HF (2007) 18F-labeled styrylpyridines as PET agents for amyloid plaque imaging. *Nucl Med Biol* **34**, 89-97.
- [75] Clark CM, Schneider JA, Bedell BJ, Beach TG, Bilker WB, Mintun MA, Pontecorvo MJ, Hefti F, Carpenter AP, Flitter ML, Krautkramer MJ, Kung HF, Coleman RE, Doraiswamy PM, Fleisher AS, Sabbagh MN, Sadowsky CH, Reiman EP, Zehntner SP, Skovronsky DM (2011) Use of florbetapir-PET for imaging beta-amyloid pathology. *JAMA* **305**, 275-283.
- [76] Choi SR, Schneider JA, Bennett DA, Beach TG, Bedell BJ, Zehntner SP, Krautkramer MJ, Kung HF, Skovronsky DM, Hefti F, Clark CM (2012) Correlation of amyloid PET ligand florbetapir F 18 binding with Abeta aggregation and neuritic plaque deposition in postmortem brain tissue. *Alzheimer Dis Assoc Disord* **26**, 8-16.
- [77] Clark CM, Pontecorvo MJ, Beach TG, Bedell BJ, Coleman RE, Doraiswamy PM, Fleisher AS, Reiman EM, Sabbagh MN, Sadowsky CH, Schneider JA, Arora A, Carpenter AP, Flitter ML, Joshi AD, Krautkramer MJ, Lu M, Mintun MA, Skovronsky DM (2012) Cerebral PET with florbetapir compared with neuropathology at autopsy for detection of neuritic amyloid-beta plaques: A prospective cohort study. *Lancet Neurol* **11**, 669-678.
- [78] Curtis C, Gamez JE, Singh U, Sadowsky CH, Villena T, Sabbagh MN, Beach TG, Duara R, Fleisher AS, Frey KA, Walker Z, Hunjan A, Holmes C, Escovar YM, Vera CX, Agronin ME, Ross J, Bozoki A, Akinola M, Shi J, Vandenberghe R, Ikonomic MD, Sherwin PF, Grachev ID, Farrar G, Smith AP, Buckley CJ, McLain R, Salloway S (2015) Phase 3 trial of flutemetamol labeled with radioactive fluorine 18 imaging and neuritic plaque density. *JAMA Neurol* **72**, 287-294.
- [79] Ong KT, Villemagne VL, Bahar-Fuchs A, Lamb F, Langdon N, Catafau AM, Stephens AW, Seibyl J, Dinkelborg LM, Reiningner CB, Putz B, Rohde B, Masters CL, Rowe CC (2015) Abeta imaging with 18F-florbetaben in prodromal Alzheimer's disease: A prospective outcome study. *J Neurol Neurosurg Psychiatry* **86**, 431-436.
- [80] Wolk DA, Zhang Z, Boudhar S, Clark CM, Pontecorvo MJ, Arnold SE (2012) Amyloid imaging in Alzheimer's disease: Comparison of florbetapir and Pittsburgh compound-B positron emission tomography. *J Neurol Neurosurg Psychiatry* **83**, 923-926.
- [81] Villemagne VL, Mulligan RS, Pejoska S, Ong K, Jones G, O'Keefe G, Chan JG, Young K, Tochon-Danguy H, Masters CL, Rowe CC (2012) Comparison of 11C-PiB and 18F-florbetaben for Abeta imaging in ageing and Alzheimer's disease. *Eur J Nucl Med Mol Imaging* **39**, 983-989.
- [82] Landau SM, Thomas BA, Thurfjell L, Schmidt M, Margolin R, Mintun M, Pontecorvo M, Baker SL, Jagust WJ (2014) Amyloid PET imaging in Alzheimer's disease: A comparison of three radiotracers. *Eur J Nucl Med Mol Imaging* **41**, 1398-1407.
- [83] Johnson KA, Minoshima S, Bohnen NI, Donohoe KJ, Foster NL, Herscovitch P, Karlawish JH, Rowe CC, Carrillo MC, Hartley DM, Hedrick S, Pappas V, Thies WH (2013) Appropriate use criteria for amyloid PET: A report of the Amyloid Imaging Task Force, the Society of Nuclear Medicine and Molecular Imaging, and the Alzheimer's Association. *Alzheimers Dement* **9**, e-1-16.
- [84] Johnson KA, Minoshima S, Bohnen NI, Donohoe KJ, Foster NL, Herscovitch P, Karlawish JH, Rowe CC, Hedrick S, Pappas V, Carrillo MC, Hartley DM (2013) Update on appropriate use criteria for amyloid PET imaging: Dementia experts, mild cognitive impairment, and education. Amyloid Imaging Task Force of the Alzheimer's Association and Society for Nuclear Medicine and Molecular Imaging. *Alzheimers Dement* **9**, e106-e109.
- [85] Laforce R, Rosa-Neto P, Soucy JP, Rabinovici GD, Dubois B, Gauthier S (2016) Canadian Consensus Guidelines on Use of Amyloid Imaging in Canada: Update and Future Directions from the Specialized Task Force on Amyloid imaging in Canada. *Can J Neurol Sci* **43**, 503-512.
- [86] McNamee RL, Yee SH, Price JC, Klunk WE, Rosario B, Weissfeld L, Ziolkowski S, Berginc M, Lopresti B, Dekosky S, Mathis CA (2009) Consideration of optimal time window for Pittsburgh compound B PET summed uptake measurements. *J Nucl Med* **50**, 348-355.
- [87] Lopresti BJ, Klunk WE, Mathis CA, Hoge JA, Ziolkowski SK, Lu X, Meltzer CC, Schimmel K, Tsopelas ND, DeKosky ST, Price JC (2005) Simplified quantification of Pittsburgh Compound B amyloid imaging PET studies: A comparative analysis. *J Nucl Med* **46**, 1959-1972.
- [88] Heurling K, Buckley C, Van Laere K, Vandenberghe R, Lubberink M (2015) Parametric imaging and quantitative analysis of the PET amyloid ligand [(18)F]flutemetamol. *Neuroimage* **121**, 184-192.

- [89] Becker GA, Ichise M, Barthel H, Luthardt J, Patt M, Seese A, Schultze-Mosgau M, Rohde B, Gertz HJ, Reiningger C, Sabri O (2013) PET quantification of 18F-florbetaben binding to beta-amyloid deposits in human brains. *J Nucl Med* **54**, 723-731.
- [90] Barthel H, Gertz HJ, Dresel S, Peters O, Bartenstein P, Buerger K, Hiemeyer F, Wittemer-Rump SM, Seibyl J, Reiningger C, Sabri O (2011) Cerebral amyloid-beta PET with florbetaben (18F) in patients with Alzheimer's disease and healthy controls: A multicentre phase 2 diagnostic study. *Lancet Neurol* **10**, 424-435.
- [91] Yeo JM, Waddell B, Khan Z, Pal S (2015) A systematic review and meta-analysis of (18)F-labeled amyloid imaging in Alzheimer's disease. *Alzheimers Dement (Amst)* **1**, 5-13.
- [92] Klunk WE, Koeppe RA, Price JC, Benzinger TL, Devous MD, Sr., Jagust WJ, Johnson KA, Mathis CA, Minhas D, Pontecorvo MJ, Rowe CC, Skovronsky DM, Mintun MA (2015) The Centiloid Project: Standardizing quantitative amyloid plaque estimation by PET. *Alzheimers Dement* **11**, 1-15.e11-14.
- [93] Mintun MA, Larossa GN, Sheline YI, Dence CS, Lee SY, Mach RH, Klunk WE, Mathis CA, DeKosky ST, Morris JC (2006) [11C]PIB in a nondemented population: Potential antecedent marker of Alzheimer disease. *Neurology* **67**, 446-452.
- [94] Jagust WJ, Bandy D, Chen K, Foster NL, Landau SM, Mathis CA, Price JC, Reiman EM, Skovronsky D, Koeppe RA (2010) The Alzheimer's Disease Neuroimaging Initiative positron emission tomography core. *Alzheimers Dement* **6**, 221-229.
- [95] Harkins K, Sankar P, Sperling R, Grill JD, Green RC, Johnson KA, Healy M, Karlawish J (2015) Development of a process to disclose amyloid imaging results to cognitively normal older adult research participants. *Alzheimers Res Ther* **7**, 26.
- [96] O'Brien JT, Firbank MJ, Davison C, Barnett N, Bamford C, Donaldson C, Olsen K, Herholz K, Williams D, Lloyd J (2014) 18F-FDG PET and perfusion SPECT in the diagnosis of Alzheimer and Lewy body dementias. *J Nucl Med* **55**, 1959-1965.
- [97] O'Brien JT, McKeith IG, Walker Z, Tatsch K, Booij J, Darcourt J, Marquardt M, Reiningger C (2009) Diagnostic accuracy of 123I-FP-CIT SPECT in possible dementia with Lewy bodies. *Br J Psychiatry* **194**, 34-39.
- [98] Nelson PT, Alafuzoff I, Bigio EH, Bouras C, Braak H, Cairns NJ, Castellani RJ, Crain BJ, Davies P, Del Tredici K, Duyckaerts C, Frosch MP, Haroutunian V, Hof PR, Hulette CM, Hyman BT, Iwatsubo T, Jellinger KA, Jicha GA, Kovari E, Kukull WA, Leverenz JB, Love S, Mackenzie IR, Mann DM, Masliah E, McKee AC, Montine TJ, Morris JC, Schneider JA, Sonnen JA, Thal DR, Trojanowski JQ, Troncoso JC, Wisniewski T, Woltjer RL, Beach TG (2012) Correlation of Alzheimer disease neuropathologic changes with cognitive status: A review of the literature. *J Neuropathol Exp Neurol* **71**, 362-381.
- [99] Bierer LM, Hof PR, Purohit DP, Carlin L, Schmeidler J, Davis KL, Perl DP (1995) Neocortical neurofibrillary tangles correlate with dementia severity in Alzheimer's disease. *Arch Neurol* **52**, 81-88.
- [100] Bobinski M, Wegiel J, Wisniewski HM, Tarnawski M, Reisberg B, De Leon MJ, Miller DC (1996) Neurofibrillary pathology—correlation with hippocampal formation atrophy in Alzheimer disease. *Neurobiol Aging* **17**, 909-919.
- [101] Whitwell JL, Josephs KA, Murray ME, Kantarci K, Przybelski SA, Weigand SD, Vemuri P, Senjem ML, Parisi JE, Knopman DS, Boeve BF, Petersen RC, Dickson DW, Jack CR Jr (2008) MRI correlates of neurofibrillary tangle pathology at autopsy: A voxel-based morphometry study. *Neurology* **71**, 743-749.
- [102] Braak H, Braak E (1997) Frequency of stages of Alzheimer-related lesions in different age categories. *Neurobiol Aging* **18**, 351-357.
- [103] Jack CR Jr, Knopman DS, Chetelat G, Dickson D, Fagan AM, Frisoni GB, Jagust W, Mormino EC, Petersen RC, Sperling RA, van der Flier WM, Villemagne VL, Visser PJ, Vos SJ (2016) Suspected non-Alzheimer disease pathophysiology—concept and controversy. *Nat Rev Neurol* **12**, 117-124.
- [104] Dani M, Brooks DJ, Edison P (2016) Tau imaging in neurodegenerative diseases. *Eur J Nucl Med Mol Imaging* **43**, 1139-1150.
- [105] Okamura N, Harada R, Furukawa K, Furumoto S, Tago T, Yanai K, Arai H, Kudo Y (2016) Advances in the development of tau PET radiotracers and their clinical applications. *Ageing Res Rev* **30**, 107-113.
- [106] Gao F, Barker PB (2014) Various MRS application tools for Alzheimer disease and mild cognitive impairment. *AJNR Am J Neuroradiol* **35**, S4-s11.
- [107] Shonk TK, Moats RA, Gifford P, Michaelis T, Mandigo JC, Izumi J, Ross BD (1995) Probable Alzheimer disease: Diagnosis with proton MR spectroscopy. *Radiology* **195**, 65-72.
- [108] Ernst T, Chang L, Melchor R, Mehringer CM (1997) Frontotemporal dementia and early Alzheimer disease: Differentiation with frontal lobe H-1 MR spectroscopy. *Radiology* **203**, 829-836.
- [109] Kantarci K, Jack CR Jr, Xu YC, Campeau NG, O'Brien PC, Smith GE, Ivnik RJ, Boeve BF, Kokmen E, Tangalos EG, Petersen RC (2000) Regional metabolic patterns in mild cognitive impairment and Alzheimer's disease: A 1H MRS study. *Neurology* **55**, 210-217.
- [110] Schuff N, Amend D, Ezekiel F, Steinman SK, Tanabe J, Norman D, Jagust W, Kramer JH, Mastrianni JA, Fein G, Weiner MW (1997) Changes of hippocampal N-acetyl aspartate and volume in Alzheimer's disease. A proton MR spectroscopic imaging and MRI study. *Neurology* **49**, 1513-1521.
- [111] Schuff N, Capizzano AA, Du AT, Amend DL, O'Neill J, Norman D, Kramer J, Jagust W, Miller B, Wolkowitz OM, Yaffe K, Weiner MW (2002) Selective reduction of N-acetylaspartate in medial temporal and parietal lobes in AD. *Neurology* **58**, 928-935.
- [112] Kantarci K, Reynolds G, Petersen RC, Boeve BF, Knopman DS, Edland SD, Smith GE, Ivnik RJ, Tangalos EG, Jack CR Jr (2003) Proton MR spectroscopy in mild cognitive impairment and Alzheimer disease: Comparison of 1.5 and 3 T. *AJNR Am J Neuroradiol* **24**, 843-849.
- [113] Valenzuela MJ, Sachdev P (2001) Magnetic resonance spectroscopy in AD. *Neurology* **56**, 592-598.
- [114] Murray ME, Przybelski SA, Lesnick TG, Liesinger AM, Sychalla A, Zhang B, Gunter JL, Parisi JE, Boeve BF, Knopman DS, Petersen RC, Jack CR Jr, Dickson DW, Kantarci K (2014) Early Alzheimer's disease neuropathology detected by proton MR spectroscopy. *J Neurosci* **34**, 16247-16255.
- [115] Adalsteinsson E, Sullivan EV, Kleinhans N, Spielman DM, Pfefferbaum A (2000) Longitudinal decline of the

- neuronal marker N-acetyl aspartate in Alzheimer's disease. *Lancet* **355**, 1696-1697.
- [116] Antuono PG, Jones JL, Wang Y, Li SJ (2001) Decreased glutamate+glutamine in Alzheimer's disease detected in vivo with (1)H-MRS at 0.5 T. *Neurology* **56**, 737-742.
- [117] Lee MH, Smyser CD, Shimony JS (2013) Resting-state fMRI: A review of methods and clinical applications. *AJNR Am J Neuroradiol* **34**, 1866-1872.
- [118] Joo SH, Lim HK, Lee CU (2016) Three large-scale functional brain networks from resting-state functional MRI in subjects with different levels of cognitive impairment. *Psychiatry Investig* **13**, 1-7.
- [119] Li HJ, Hou XH, Liu HH, Yue CL, He Y, Zuo XN (2015) Toward systems neuroscience in mild cognitive impairment and Alzheimer's disease: A meta-analysis of 75 fMRI studies. *Hum Brain Mapp* **36**, 1217-1232.
- [120] Song SK, Sun SW, Ju WK, Lin SJ, Cross AH, Neufeld AH (2003) Diffusion tensor imaging detects and differentiates axon and myelin degeneration in mouse optic nerve after retinal ischemia. *Neuroimage* **20**, 1714-1722.
- [121] Sajjadi SA, Acosta-Cabronero J, Patterson K, Diaz-de-Grenu LZ, Williams GB, Nestor PJ (2013) Diffusion tensor magnetic resonance imaging for single subject diagnosis in neurodegenerative diseases. *Brain* **136**, 2253-2261.
- [122] Agosta F, Galantucci S, Filippi M (2017) Advanced magnetic resonance imaging of neurodegenerative diseases. *Neurol Sci* **38**, 41-51.
- [123] Clerx L, Visser PJ, Verhey F, Aalten P (2012) New MRI markers for Alzheimer's disease: A meta-analysis of diffusion tensor imaging and a comparison with medial temporal lobe measurements. *J Alzheimers Dis* **29**, 405-429.
- [124] Teipel S, Drzezga A, Grothe MJ, Barthel H, Chételat G, Schuff N, Skudlarski P, Cavado E, Frisoni GB, Hoffmann W, Thyrian JR, Fox C, Minoshima S, Sabri O, Fellgiebel A (2015) Multimodal imaging in Alzheimer's disease: Validity and usefulness for early detection. *Lancet Neurol* **14**, 1037-1053.
- [125] Deibler AR, Pollock JM, Kraft RA, Tan H, Burdette JH, Maldjian JA (2008) Arterial spin-labeling in routine clinical practice, part 1: Technique and artifacts. *AJNR Am J Neuroradiol* **29**, 1228-1234.
- [126] Hays CC, Zlatař ZZ, Wierenga CE (2016) The utility of cerebral blood flow as a biomarker of preclinical Alzheimer's disease. *Cell Mol Neurobiol* **36**, 167-179.
- [127] Zhang J (2016) How far is arterial spin labeling MRI from a clinical reality? Insights from arterial spin labeling comparative studies in Alzheimer's disease and other neurological disorders. *J Magn Reson Imaging* **43**, 1020-1045.