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DOI: 10.3748/wjg.v24.i37.4217

World J Gastroenterol 2018 October 7; 24(37): 4217-4223

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

EDITORIAL

Focus on the gut-brain axis: Multiple sclerosis, the intestinal barrier and the microbiome

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Author contributions: Camara-Lemarroy CR conceived the study and drafted the manuscript; Metz LM and Yong VW revised and approved the final version of the article.

Supported by the Lejoie-Lake Fellowship (to Camara-Lemarroy CR) awarded by the Hotchkiss Brain Institute.

Conflict-of-interest statement: The authors have no conflict of interest to declare.

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Manuscript source: Invited manuscript

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Received: July 5, 2018 Peer-review started: July 5, 2018 First decision: July 25, 2018 Revised: July 30, 2018 Accepted: August 1, 2018 Article in press: August 1, 2018 Published online: October 7, 2018

Abstract

The brain-gut axis serves as the bidirectional connection between the gut microbiome, the intestinal barrier and the immune system that might be relevant for the pathophysiology of inflammatory demyelinating diseases. People with multiple sclerosis have been shown to have an altered microbiome, increased intestinal permeability and changes in bile acid metabolism. Experimental evidence suggests that these changes can lead to profound alterations of peripheral and central nervous system immune regulation. Besides being of pathophysiological interest, the brain-gut axis could also open new avenues of therapeutic targets. Modification of the microbiome, the use of probiotics, fecal microbiota transplantation, supplementation with bile acids and intestinal barrier enhancers are all promising candidates. Hopefully, pre-clinical studies and clinical trials will soon yield significant results.

Key words: Multiple sclerosis; Microbiome; Intestinal barrier; Bile acids; Gut-brain axis

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Core tip: Many studies suggest that the brain-gut connection can contribute to our knowledge of the pathophysiology of neurological conditions. Recent evidence suggests that people with multiple sclerosis have changes in their gut microbiome, their intestinal barrier and even in the metabolism of bile acids. All of these represent relevant therapeutic targets that could feasibly be addressed by pre-clinical and clinical studies. This knowledge acquired in the bench might soon be translated to the bedside.



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Camara-Lemarroy CR, Metz LM, Yong VW. Focus on the gutbrain axis: Multiple sclerosis, the intestinal barrier and the microbiome. *World J Gastroenterol* 2018; 24(37): 4217-4223 Available from: URL: http://www.wjgnet.com/1007-9327/full/ v24/i37/4217.htm DOI: http://dx.doi.org/10.3748/wjg.v24. i37.4217

INTRODUCTION

Clinical and preclinical studies have shown bidirectional interactions within the brain-gut axis and the gut microbiome, the intestinal barrier and the immune system, both in health and disease. These complex interactions might be relevant for the pathophysiology of inflammatory demyelinating diseases, and in particular, multiple sclerosis, where much interest has been placed in the recent literature.

THE GUT MICROBIOME

Much interest has been placed recently on the possible role of the gut microbiome in multiple sclerosis (MS) pathophysiology. Many review articles on this subject have recently been published^[1-3], perhaps more than original research articles that actually characterize the gut microbiome in patients with MS. This research is in keeping with the essential role that the gut microbiome has in regulating the development of the immune system^[4]. This area of research has also been the subject of recent symposia in international MS conferences^[5,6].

Much of the experimental evidence is derived from studies using the experimental autoimmune encephalomyelitis (EAE) mouse model of MS. Modifying the gut microbiota with either antibiotic cocktails or probiotics leads to EAE attenuation as well as a multitude of regulatory immune responses^[7-9]. Animals bred in germfree conditions are resistant to EAE induction and show an attenuated immunological response^[10,11], an effect lost when mice are repopulated with gut commensals^[11]. In recent intriguing experiments, transgenic mice prone to spontaneous brain autoimmunity developed severe disease when transplanted with fecal microbiota from MS patients, as opposed to mice that received fecal microbiota from healthy matching twins^[12]. Germ-free mice receiving similar transplants also developed severe EAE, while showing altered peripheral immune responses^[13].

From studies attempting to characterize the composition of the microbiome, it is clear there are some differences in people with MS compared to controls. People with relapsing-remitting MS (RRMS) have an abundance of *Anaerostipes, Faecalibacterium, Pseudomonas, Mycoplasma, Haemophilus, Blautia,* and *Dorea* and a relative decrease of *Bacteroides, Prevotella, Parabacteroides* and *Adlercreutzia*^[14-16]. In pediatric MS, patients have higher levels of members of *Desulfovibrionaceae* and depletion in *Lachnospiraceae* and *Ruminococcaceae*^[17,18]. Issues are further complicated by complex analyses at the taxa, phylum and species levels, and a myriad of microbes have been implicated. For example, studies have found a significant depletion in clostridial species^[15,19], *Butyricimonas*^[20], *Roseburia*^[21] and an increase in *Streptococcus*^[22], *Methanobrevibacter*, *Akkermansia* and *Coprococcus*^[14,20].

However, there are some limitations to these studies. The methods used to analyze the microbiome have been heterogeneous, with most (but not all) studies using a variation of 16S sequencing. There are differences in sample processing, DNA extraction, choice of primers, databases and hyper-variable regions analyzed across studies. Furthermore, close to two thirds of patients with MS have gastrointestinal symptoms such as constipation, dyspepsia and other functional gastrointestinal disorders^[23], and many of these have been also associated with an altered gut microbiota^[24]. Studies so far have not properly accounted for these symptoms or other relevant variables such as diet. An ongoing International MS Microbiome study aims to define a "core microbiome"^[3]. It might shine some light into this complicated field.

Nonetheless, there is mounting experimental evidence that the gut microbiome may play a role in MS pathophysiology and human studies suggest that patients have a different microbiome compared to controls. Of course, the true significance of the results obtained so far is unclear, considering that there has often been a failure to replicate microbiome animal studies in humans. But the next question that comes to mind is whether this can also constitute a relevant therapeutic target. Although this appears to be the case in experimental models, translation to clinical practice may prove challenging.

Modifying the microbiome through medications, possibly antibiotics, could be the simplest method, but several issues arise that question the feasibility of this approach. Targeting specific commensals might prove difficult and requires appropriate identification of these targets. The case of minocycline is an interesting example. Recently shown to delay the occurrence of a second demyelinating event in patients with a clinically isolated syndrome^[25], minocycline is an antibiotic known to alter the gut microbiome^[26]. Whether this is an additional mechanism of action remains unknown; it is noteworthy that the initial rationale for testing minocycline in early MS is based on its various immune-modifying properties^[27]. On the other hand, there is also evidence that MS disease modifying therapies (DMTs) may alter the microbiome directly^[26], and indeed, it also appears that a multitude of other medications such as antidepressants, antipsychotics and immune modulators may also do so^[28]. Issues such as the generation of resistant strains are also worthy of consideration.

Probiotics are a popular option but there are various issues with their practical implementation. Probiotics do not modify the host microbiome in a robust and persistent manner, although they are purported to be able to influence gut immunity and homeostasis. Despite success in showing a benefit for probiotics in animal models^[29,30], there are only a handful of clinical trials in MS. Results have been preliminary, with some modest beneficial trends in clinical variables and some biochemical markers of changes in peripheral immune function^[31-33]. However, they have included very small numbers of patients and the duration of these trials have been too short to shed any light onto clinically meaningful outcomes. There are many barriers to be overcome, such as selecting the appropriate formulation, dose and study design. There is also a lesson to be learned from the multiple clinical trials in inflammatory bowel disease (IBD), where despite a wealth of available studies (although heterogeneous in design and quality), the evidence supporting their clinical use is limited to carefully selected subpopulations^[34,35].

Fecal microbiota transplantation (FMT) would constitute the optimal strategy to modify the gut microbiome. It has proven to be remarkably effective in managing C. difficile colitis, and isolated case reports describe beneficial effects over MS disease course, through mechanisms that remain unclear^[36,37]. A clinical trial of FMT is underway^[38], but even before its completion, many questions arise. It is unclear which population should be studied and what characterizes an ideal donor, not to mention the dose, route of administration and dose scheduling (single dose vs multiple doses). Patient with C. diff colitis who undergo FMT have been previously treated with antibiotics such as vancomycin and metronidazole, and presumably, have had some of their microbiota depleted. Would patients with MS require "ablation" of their microbiome before FMT? DMTs have immune modulating properties and they may also directly alter the microbiome^[26], so their possible effects on the "engraftment" cannot be underestimated.

THE INTESTINAL BARRIER

The intestinal barrier is the physical and functional zone of interaction between the gut microbiome and the organism. It is a complex multi-layered structure that includes major portions of the gut immunological system and is essential for homeostasis^[26]. However, it has been comparatively ignored regarding its possible role in MS pathophysiology.

In experimental models, mice undergoing EAE show an altered intestinal barrier, with increased permeability and various gross morphological changes, as well as alterations in the expression of tight junction proteins in the intestinal mucosa^[39]. The peak of intestinal barrier dysfunction mirrors the peak of EAE dinical severity and preventing intestinal barrier breakdown leads to attenuation of EAE^[40]. These alterations have also been associated with several abnormal immunological responses.

Patients with MS also have an altered intestinal barrier. Almost 2 decades ago, investigators found that patients with MS had increased intestinal permeability when compared to controls, using an *in vivo* mannitol/lactulose ratio test^[41]. Increased intestinal permeability was also found to be associated with the number of peripheral CD45RO+ B cells^[41]. A more recent study confirmed this finding; up to 70% of MS patients had increased intestinal permeability^[42]. It has been hypothesized that an altered intestinal barrier might lead to bacterial translocation thus allowing the passage of noxious molecules such as microbial associated molecular patterns. This could then alter peripheral immune responses or allow these molecules to enter the CNS and alter neuroimmunity^[26,43].

Although the evidence linking the intestinal barrier with MS is much more limited than evidence linking MS with alterations of the gut microbiome, the question of whether it constitutes a viable therapeutic target is the same. Of course, the issue is complicated by the fact that the microbiome is essential in the regulation of intestinal barrier function, so it could be arbitrary to think of them as separate entities. An altered intestinal barrier is also a crucial aspect of the pathophysiology of IBD and celiac disease, so research from these fields has shed light on possible strategies to maintain intestinal barrier integrity.

One of the first components of the intestinal barrier is a thick mucus layer forming a protective film, enriched by secretory IgA and antimicrobial peptides and proteins. Oral supplementation with lecithin and phosphatidylcholine can adhere to the intestinal mucosa, strengthening the mucus layer and improving barrier function^[44-46]. Regulators of tight junctions, such as larazotide, are under development. Larazotide is a peptide able to re-arrange tight junctions and prevent intestinal barrier dysfunction. It has been studied in patients with celiac disease with promising results^[47-49]. Designing pre-clinical studies using these methods to enhance barrier function in the setting of autoimmune neurological disease should be straightforward.

Although probiotics may not be the ideal method to modify the microbiome, they have been suggested to play a significant role in modulating barrier function. E. coli strain nissle has been marketed in Europe for many years as a probiotic with beneficial effects on the intestinal barrier^[50]. It has moderate evidence from randomized trials showing it may lead to remission in ulcerative colitis^[51] and in the EAE mouse model of MS it reduced disease severity by maintaining intestinal barrier function^[40]. VSL#3 is another probiotic mixture with putative barrier-protecting properties^[52]. There is evidence of clinical effectiveness in the management of chronic pouchitis in patients with ulcerative colitis^[35,53]. VSL#3 administered to a small number of MS patients leads to an anti-inflammatory peripheral immune response^[33]. These two probiotic agents would be good candidates for a large, well-designed clinical trial.

Finally, we go full circle and return to FMT. It is believed that after successful modification of the microbiome, this strategy might lead to improved intestinal barrier function^[54]. The gut microbiome is essential for the regulation of intestinal barrier homeostasis^[55], partly through the production of short chain fatty acids (SCFA) such as butyrate, propionate and acetate. SCFAs can modulate tight junctions in the gut and modulate inflammatory responses in the intestinal mucosa^[44,55]. Other interesting alternatives have also recently been described Camara-Lemarroy CR et al. Gut-brain axis in multiple sclerosis



Figure 1 Alterations in intestinal homeostasis described in multiple sclerosis as therapeutic targets. Altered bile acid metabolism, altered microbiota and alterations in intestinal barrier function all lead to local and systemic alterations in immune responses that could negatively impact MS pathophysiology (grey squares). Bile acid supplementation, fecal microbiota transplantation, probiotics, antibiotics and barrier protectors are all possible therapeutic interventions (blue squares). MS: Multiple sclerosis; FMT: Fecal microbiota transplantation; PC: Paneth cells; EC: Epithelial cells; TJ: Tight junctions; L: Lymphocytes.

including the use of stool substitute preparations made from purified intestinal bacterial cultures derived from a single healthy donor^[56]. Of course, many questions would need to be settled before clinical trials as discussed above.

BILE ACIDS

Bile acids are the main regulators of fat and fat-soluble vitamins digestion. They also significantly affect gut physiology and homeostasis. Bile acids can modulate the intestinal barrier function through complex mechanisms^[57,58], and can shape the gut microbiota community. In turn, the microbiome can change bile acid metabolism^[59]. Remarkably, bile acids may also modulate inflammatory signaling in the central nervous system. Ursodeoxycholic acid can inhibit the inflammatory activity of microglia in vitro^[60], and tauroursodeoxycholic acid can shift microglia phenotypes towards an anti-inflammatory state through activation of the G protein-coupled bile acid receptor 1/Takeda G protein-coupled receptor 5^[61]. Bile acids are also agonists of the nuclear hormone receptor farnesoid X receptor. Bile acid farnesoid X agonism led to attenuation of EAE and modulation of neuroinflammatory responses^[62]. Mice fed a high-fat diet show dysregulated bile acid synthesis, gut dysbiosis and increased microglial activation^[63]. Furthermore, metabolomics studies have found alterations in bile acids in patients with MS compared to healthy controls^[64]. Conjugated bile acids such as ursodeoxycholic acid have been used in

the management of some gastrointestinal diseases for decades. A clinical trial of bile acid supplementation in MS is underway^[65].

CONCLUSION

Exciting research suggests that the brain-gut axis, once an almost esoteric concept, might yield novel therapeutic targets in neuroimmunological diseases such as MS (Figure 1). The often-symbiotic roles of the gut microbiome, intestinal barrier and even bile acids in the regulation of neuroimmune responses is beginning to be elucidated. If future pre-dinical and clinical studies confirm the relevance of intestinal barrier dysfunction, bile acid metabolism and the gut microbiome in the pathophysiology of MS, the next step will be to translate these findings into therapeutics. Only well designed clinical trials will answer whether interventions such as FMT, probiotics or barrier protectors yield clinically meaningful results. The time is right to assess whether the gut-brain axis can be transferred from the bench to the bedside.

REFERENCES

- Freedman SN, Shahi SK, Mangalam AK. The "Gut Feeling": Breaking Down the Role of Gut Microbiome in Multiple Sclerosis. *Neurotherapeutics* 2018; 15: 109-125 [PMID: 29204955 DOI: 10.1007/s13311-017-0588-x]
- 2 Calvo-Barreiro L, Eixarch H, Montalban X, Espejo C. Combined therapies to treat complex diseases: The role of the gut microbiota in multiple sclerosis. *Autoimmun Rev* 2018; 17: 165-174 [PMID:



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29191793 DOI: 10.1016/j.autrev.2017.11.019]

- 3 Pröbstel AK, Baranzini SE. The Role of the Gut Microbiome in Multiple Sclerosis Risk and Progression: Towards Characterization of the "MS Microbiome". *Neurotherapeutics* 2018; 15: 126-134 [PMID: 29147991 DOI: 10.1007/s13311-017-0587-y]
- 4 Thaiss CA, Zmora N, Levy M, Elinav E. The microbiome and innate immunity. *Nature* 2016; 535: 65-74 [PMID: 27383981 DOI: 10.1038/ nature18847]
- 5 Hot Topic 6: Gut microbiota in multiple sclerosis. Symposium held at the 7th Joint ECTRIMS-ACTRIMS meeting, 25-28 October, Paris, France. Available from: URL: https://www.ectrims-congress. eu/2017/scientific-programme/scientific-programme.html
- 6 Brain-Gut Axis: a focus on finding a cure for Multiple Sclerosis. University of Alberta MS Centre Research Symposium, held May 4, 2018, Edmonton, Alberta, Canada. Available from: URL: https:// cloudfront.ualberta.ca/-/media/medicine/ms-centre/2018-ualbertams-centre-program-25apr18-pdfa.pdf
- 7 Ochoa-Repáraz J, Mielcarz DW, Haque-Begum S, Kasper LH. Induction of a regulatory B cell population in experimental allergic encephalomyelitis by alteration of the gut commensal microflora. *Gut Microbes* 2010; 1: 103-108 [PMID: 21326918 DOI: 10.4161/ gmic.1.2.11515]
- 8 Colpitts SL, Kasper EJ, Keever A, Liljenberg C, Kirby T, Magori K, Kasper LH, Ochoa-Repáraz J. A bidirectional association between the gut microbiota and CNS disease in a biphasic murine model of multiple sclerosis. *Gut Microbes* 2017; 8: 561-573 [PMID: 28708466 DOI: 10.1080/19490976.2017.1353843]
- 9 Wang Y, Begum-Haque S, Telesford KM, Ochoa-Repáraz J, Christy M, Kasper EJ, Kasper DL, Robson SC, Kasper LH. A commensal bacterial product elicits and modulates migratory capacity of CD39(+) CD4 T regulatory subsets in the suppression of neuroinflammation. *Gut Microbes* 2014; **5**: 552-561 [PMID: 25006655 DOI: 10.4161/gmic.29797]
- 10 Berer K, Mues M, Koutrolos M, Rasbi ZA, Boziki M, Johner C, Wekerle H, Krishnamoorthy G. Commensal microbiota and myelin autoantigen cooperate to trigger autoimmune demyelination. *Nature* 2011; 479: 538-541 [PMID: 22031325 DOI: 10.1038/nature10554]
- 11 Lee YK, Menezes JS, Umesaki Y, Mazmanian SK. Proinflammatory T-cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA* 2011; **108** Suppl 1: 4615-4622 [PMID: 20660719 DOI: 10.1073/pnas.1000082107]
- 12 Berer K, Gerdes LA, Cekanaviciute E, Jia X, Xiao L, Xia Z, Liu C, Klotz L, Stauffer U, Baranzini SE, Kümpfel T, Hohlfeld R, Krishnamoorthy G, Wekerle H. Gut microbiota from multiple sclerosis patients enables spontaneous autoimmune encephalomyelitis in mice. *Proc Natl Acad Sci USA* 2017; 114: 10719-10724 [PMID: 28893994 DOI: 10.1073/pnas.1711233114]
- 13 Cekanaviciute E, Yoo BB, Runia TF, Debelius JW, Singh S, Nelson CA, Kanner R, Bencosme Y, Lee YK, Hauser SL, Crabtree-Hartman E, Sand IK, Gacias M, Zhu Y, Casaccia P, Cree BAC, Knight R, Mazmanian SK, Baranzini SE. Gut bacteria from multiple sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. *Proc Natl Acad Sci USA* 2017; 114: 10713-10718 [PMID: 28893978 DOI: 10.1073/pnas.1711235114]
- 14 Cantarel BL, Waubant E, Chehoud C, Kuczynski J, DeSantis TZ, Warrington J, Venkatesan A, Fraser CM, Mowry EM. Gut microbiota in multiple sclerosis: possible influence of immunomodulators. *J Investig Med* 2015; 63: 729-734 [PMID: 25775034 DOI: 10.1097/JIM.00000000000192]
- 15 Miyake S, Kim S, Suda W, Oshima K, Nakamura M, Matsuoka T, Chihara N, Tomita A, Sato W, Kim SW, Morita H, Hattori M, Yamamura T. Dysbiosis in the Gut Microbiota of Patients with Multiple Sclerosis, with a Striking Depletion of Species Belonging to Clostridia XIVa and IV Clusters. *PLoS One* 2015; 10: e0137429 [PMID: 26367776 DOI: 10.1371/journal.pone.0137429]
- 16 Chen J, Chia N, Kalari KR, Yao JZ, Novotna M, Paz Soldan MM, Luckey DH, Marietta EV, Jeraldo PR, Chen X, Weinshenker BG, Rodriguez M, Kantarci OH, Nelson H, Murray JA, Mangalam AK. Multiple sclerosis patients have a distinct gut microbiota compared to healthy controls. *Sci Rep* 2016; 6: 28484 [PMID: 27346372

DOI: 10.1038/srep28484]

- 17 Tremlett H, Fadrosh DW, Faruqi AA, Zhu F, Hart J, Roalstad S, Graves J, Lynch S, Waubant E; US Network of Pediatric MS Centers. Gut microbiota in early pediatric multiple sclerosis: a case-control study. *Eur J Neurol* 2016; 23: 1308-1321 [PMID: 27176462 DOI: 10.1111/ene.13026]
- 18 Tremlett H, Fadrosh DW, Faruqi AA, Hart J, Roalstad S, Graves J, Lynch S, Waubant E; US Network of Pediatric MS Centers. Gut microbiota composition and relapse risk in pediatric MS: A pilot study. *J Neurol Sci* 2016; 363: 153-157 [PMID: 27000242 DOI: 10.1016/j.jns.2016.02.042]
- 19 Rumah KR, Linden J, Fischetti VA, Vartanian T. Isolation of Clostridium perfringens type B in an individual at first clinical presentation of multiple sclerosis provides clues for environmental triggers of the disease. *PLoS One* 2013; 8: e76359 [PMID: 24146858 DOI: 10.1371/journal.pone.0076359]
- 20 Jangi S, Gandhi R, Cox LM, Li N, von Glehn F, Yan R, Patel B, Mazzola MA, Liu S, Glanz BL, Cook S, Tankou S, Stuart F, Melo K, Nejad P, Smith K, Topçuolu BD, Holden J, Kivisäkk P, Chitnis T, De Jager PL, Quintana FJ, Gerber GK, Bry L, Weiner HL. Alterations of the human gut microbiome in multiple sclerosis. *Nat Commun* 2016; 7: 12015 [PMID: 27352007 DOI: 10.1038/ncomms12015]
- 21 Swidsinski A, Dörffel Y, Loening-Baucke V, Gille C, Göktas Ö, Reißhauer A, Neuhaus J, Weylandt KH, Guschin A, Bock M. Reduced Mass and Diversity of the Colonic Microbiome in Patients with Multiple Sclerosis and Their Improvement with Ketogenic Diet. *Front Microbiol* 2017; 8: 1141 [PMID: 28702003 DOI: 10.3389/ fmicb.2017.01141]
- 22 Cosorich I, Dalla-Costa G, Sorini C, Ferrarese R, Messina MJ, Dolpady J, Radice E, Mariani A, Testoni PA, Canducci F, Comi G, Martinelli V, Falcone M. High frequency of intestinal TH17 cells correlates with microbiota alterations and disease activity in multiple sclerosis. *Sci Adv* 2017; **3**: e1700492 [PMID: 28706993 DOI: 10.1126/sciadv.1700492]
- 23 Levinthal DJ, Rahman A, Nusrat S, O'Leary M, Heyman R, Bielefeldt K. Adding to the burden: gastrointestinal symptoms and syndromes in multiple sclerosis. *Mult Scler Int* 2013; 2013: 319201 [PMID: 24163768 DOI: 10.1155/2013/319201]
- 24 De Palma G, Collins SM, Bercik P. The microbiota-gut-brain axis in functional gastrointestinal disorders. *Gut Microbes* 2014; 5: 419-429 [PMID: 24921926 DOI: 10.4161/gmic.29417]
- 25 Metz LM, Li DKB, Traboulsee AL, Duquette P, Eliasziw M, Cerchiaro G, Greenfield J, Riddehough A, Yeung M, Kremenchutzky M, Vorobeychik G, Freedman MS, Bhan V, Blevins G, Marriott JJ, Grand'Maison F, Lee L, Thibault M, Hill MD, Yong VW; Minocycline in MS Study Team. Trial of Minocycline in a Clinically Isolated Syndrome of Multiple Sclerosis. N Engl J Med 2017; 376: 2122-2133 [PMID: 28564557 DOI: 10.1056/NEJMoa1608889]
- 26 Camara-Lemarroy CR, Metz L, Meddings JB, Sharkey KA, Wee Yong V. The intestinal barrier in multiple sclerosis: implications for pathophysiology and therapeutics. *Brain* 2018; **141**: 1900-1916 [PMID: 29860380 DOI: 10.1093/brain/awy131]
- 27 Yong VW, Wells J, Giuliani F, Casha S, Power C, Metz LM. The promise of minocycline in neurology. *Lancet Neurol* 2004; 3: 744-751 [PMID: 15556807 DOI: 10.1016/S1474-4422(04)00937-8]
- 28 Maier L, Pruteanu M, Kuhn M, Zeller G, Telzerow A, Anderson EE, Brochado AR, Fernandez KC, Dose H, Mori H, Patil KR, Bork P, Typas A. Extensive impact of non-antibiotic drugs on human gut bacteria. *Nature* 2018; 555: 623-628 [PMID: 29555994 DOI: 10.1038/nature25979]
- 29 Kwon HK, Kim GC, Kim Y, Hwang W, Jash A, Sahoo A, Kim JE, Nam JH, Im SH. Amelioration of experimental autoimmune encephalomyelitis by probiotic mixture is mediated by a shift in T helper cell immune response. *Clin Immunol* 2013; 146: 217-227 [PMID: 23416238 DOI: 10.1016/j.clim.2013.01.001]
- 30 Lavasani S, Dzhambazov B, Nouri M, Fåk F, Buske S, Molin G, Thorlacius H, Alenfall J, Jeppsson B, Weström B. A novel probiotic mixture exerts a therapeutic effect on experimental autoimmune encephalomyelitis mediated by IL-10 producing regulatory T cells. *PLoS One* 2010; **5**: e9009 [PMID: 20126401 DOI: 10.1371/



journal.pone.0009009]

- 31 Kouchaki E, Tamtaji OR, Salami M, Bahmani F, Daneshvar Kakhaki R, Akbari E, Tajabadi-Ebrahimi M, Jafari P, Asemi Z. Clinical and metabolic response to probiotic supplementation in patients with multiple sclerosis: A randomized, double-blind, placebo-controlled trial. *Clin Nutr* 2017; **36**: 1245-1249 [PMID: 27669638 DOI: 10.1016/j.clnu.2016.08.015]
- 32 Tamtaji OR, Kouchaki E, Salami M, Aghadavod E, Akbari E, Tajabadi-Ebrahimi M, Asemi Z. The Effects of Probiotic Supplementation on Gene Expression Related to Inflammation, Insulin, and Lipids in Patients With Multiple Sclerosis: A Randomized, Double-Blind, Placebo-Controlled Trial. J Am Coll Nutr 2017; 36: 660-665 [PMID: 28922099 DOI: 10.1080/0731572 4.2017.1347074]
- Tankou SK, Regev K, Healy BC, Cox LM, Tjon E, Kivisakk P, Vanande IP, Cook S, Gandhi R, Glanz B, Stankiewicz J, Weiner HL. Investigation of probiotics in multiple sclerosis. *Mult Scler* 2018; 24: 58-63 [PMID: 29307299 DOI: 10.1177/1352458517737390]
- 34 Abraham BP, Quigley EMM. Probiotics in Inflammatory Bowel Disease. Gastroenterol Clin North Am 2017; 46: 769-782 [PMID: 29173520 DOI: 10.1016/j.gtc.2017.08.003]
- 35 Derwa Y, Gracie DJ, Hamlin PJ, Ford AC. Systematic review with meta-analysis: the efficacy of probiotics in inflammatory bowel disease. *Aliment Pharmacol Ther* 2017; 46: 389-400 [PMID: 28653751 DOI: 10.1111/apt.14203]
- 36 Borody TJ, Leis SM, Campbell J, Torres M, Nowak A. Fecal microbiota transplantation (FMT) in multiple sclerosis (MS). Am J Gastroenterol 2011; 106: S352
- 37 Makkawi S, Camara-Lemarroy C, Metz L. Fecal microbiota transplantation associated with 10 years of stability in a patient with SPMS. *Neurol Neuroimmunol Neuroinflamm* 2018; 5: e459 [PMID: 29619403 DOI: 10.1212/NXI.00000000000459]
- 38 Kremenchutzky M. Fecal Microbial Transplantation in Relapsing Multiple Sclerosis Patients. cited 2018-05-22; In ClinicalTrials. gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: URL: https://clinicaltrials.gov/ct2/show/NCT03183869
- 39 Nouri M, Bredberg A, Weström B, Lavasani S. Intestinal barrier dysfunction develops at the onset of experimental autoimmune encephalomyelitis, and can be induced by adoptive transfer of autoreactive T cells. *PLoS One* 2014; 9: e106335 [PMID: 25184418 DOI: 10.1371/journal.pone.0106335]
- 40 Secher T, Kassem S, Benamar M, Bernard I, Boury M, Barreau F, Oswald E, Saoudi A. Oral Administration of the Probiotic Strain *Escherichia coli* Nissle 1917 Reduces Susceptibility to Neuroinflammation and Repairs Experimental Autoimmune Encephalomyelitis-Induced Intestinal Barrier Dysfunction. *Front Immunol* 2017; 8: 1096 [PMID: 28959254 DOI: 10.3389/fimmu.2017.01096]
- 41 Yacyshyn B, Meddings J, Sadowski D, Bowen-Yacyshyn MB. Multiple sclerosis patients have peripheral blood CD45RO+ B cells and increased intestinal permeability. *Dig Dis Sci* 1996; 41: 2493-2498 [PMID: 9011463 DOI: 10.1007/BF02100148]
- 42 Buscarinu MC, Cerasoli B, Annibali V, Policano C, Lionetto L, Capi M, Mechelli R, Romano S, Fornasiero A, Mattei G, Piras E, Angelini DF, Battistini L, Simmaco M, Umeton R, Salvetti M, Ristori G. Altered intestinal permeability in patients with relapsing-remitting multiple sclerosis: A pilot study. *Mult Scler* 2017; 23: 442-446 [PMID: 27270497 DOI: 10.1177/1352458516652498]
- 43 Buscarinu MC, Romano S, Mechelli R, Pizzolato Umeton R, Ferraldeschi M, Fornasiero A, Reniè R, Cerasoli B, Morena E, Romano C, Loizzo ND, Umeton R, Salvetti M, Ristori G. Intestinal Permeability in Relapsing-Remitting Multiple Sclerosis. *Neurotherapeutics* 2018; 15: 68-74 [PMID: 29119385 DOI: 10.1007/ s13311-017-0582-3]
- 44 Sun J, Shen X, Li Y, Guo Z, Zhu W, Zuo L, Zhao J, Gu L, Gong J, Li J. Therapeutic Potential to Modify the Mucus Barrier in Inflammatory Bowel Disease. *Nutrients* 2016; 8: [PMID: 26784223 DOI: 10.3390/nu8010044]
- 45 Merga Y, Campbell BJ, Rhodes JM. Mucosal barrier, bacteria and inflammatory bowel disease: possibilities for therapy. *Dig Dis*

2014; 32: 475-483 [PMID: 24969297 DOI: 10.1159/000358156]

- 46 Stremmel W, Gauss A. Lecithin as a therapeutic agent in ulcerative colitis. *Dig Dis* 2013; **31**: 388-390 [PMID: 24246994 DOI: 10.1159/000354707]
- 47 Pearce SC, Al-Jawadi A, Kishida K, Yu S, Hu M, Fritzky LF, Edelblum KL, Gao N, Ferraris RP. Marked differences in tight junction composition and macromolecular permeability among different intestinal cell types. *BMC Biol* 2018; 16: 19 [PMID: 29391007 DOI: 10.1186/s12915-018-0481-z]
- 48 Gopalakrishnan S, Tripathi A, Tamiz AP, Alkan SS, Pandey NB. Larazotide acetate promotes tight junction assembly in epithelial cells. *Peptides* 2012; 35: 95-101 [PMID: 22401910 DOI: 10.1016/ j.peptides.2012.02.016]
- 49 Khaleghi S, Ju JM, Lamba A, Murray JA. The potential utility of tight junction regulation in celiac disease: focus on larazotide acetate. *Therap Adv Gastroenterol* 2016; 9: 37-49 [PMID: 26770266 DOI: 10.1177/1756283X15616576]
- 50 Sonnenborn U. Escherichia coli strain Nissle 1917-from bench to bedside and back: history of a special Escherichia coli strain with probiotic properties. *FEMS Microbiol Lett* 2016; 363: [PMID: 27619890 DOI: 10.1093/femsle/fnw212]
- 51 Scaldaferri F, Gerardi V, Mangiola F, Lopetuso LR, Pizzoferrato M, Petito V, Papa A, Stojanovic J, Poscia A, Cammarota G, Gasbarrini A. Role and mechanisms of action of Escherichia coli Nissle 1917 in the maintenance of remission in ulcerative colitis patients: An update. *World J Gastroenterol* 2016; 22: 5505-5511 [PMID: 27350728 DOI: 10.3748/wjg.v22.i24.5505]
- 52 Dai C, Zhao DH, Jiang M. VSL#3 probiotics regulate the intestinal epithelial barrier in vivo and in vitro via the p38 and ERK signaling pathways. *Int J Mol Med* 2012; 29: 202-208 [PMID: 22089663 DOI: 10.3892/ijmm.2011.839]
- 53 Singh S, Stroud AM, Holubar SD, Sandborn WJ, Pardi DS. Treatment and prevention of pouchitis after ileal pouch-anal anastomosis for chronic ulcerative colitis. *Cochrane Database Syst Rev* 2015; CD001176 [PMID: 26593456 DOI: 10.1002/14651858. CD001176.pub3]
- 54 Reinisch W. Fecal Microbiota Transplantation in Inflammatory Bowel Disease. *Dig Dis* 2017; 35: 123-126 [PMID: 28147375 DOI: 10.1159/000449092]
- 55 Wells JM, Brummer RJ, Derrien M, MacDonald TT, Troost F, Cani PD, Theodorou V, Dekker J, Méheust A, de Vos WM, Mercenier A, Nauta A, Garcia-Rodenas CL. Homeostasis of the gut barrier and potential biomarkers. *Am J Physiol Gastrointest Liver Physiol* 2017; **312**: G171-G193 [PMID: 27908847 DOI: 10.1152/ ajpgi.00048.2015]
- 56 Petrof EO, Gloor GB, Vanner SJ, Weese SJ, Carter D, Daigneault MC, Brown EM, Schroeter K, Allen-Vercoe E. Stool substitute transplant therapy for the eradication of Clostridium difficile infection: 'RePOOPulating' the gut. *Microbiome* 2013; 1: 3 [PMID: 24467987 DOI: 10.1186/2049-2618-1-3]
- 57 Pavlidis P, Powell N, Vincent RP, Ehrlich D, Bjarnason I, Hayee B. Systematic review: bile acids and intestinal inflammation-luminal aggressors or regulators of mucosal defence? *Aliment Pharmacol Ther* 2015; 42: 802-817 [PMID: 26223936 DOI: 10.1111/apt.13333]
- 58 Keating N, Keely SJ. Bile acids in regulation of intestinal physiology. *Curr Gastroenterol Rep* 2009; 11: 375-382 [PMID: 19765365 DOI: 10.1007/s11894-009-0057-8]
- 59 Li Y, Tang R, Leung PSC, Gershwin ME, Ma X. Bile acids and intestinal microbiota in autoimmune cholestatic liver diseases. *Autoimmun Rev* 2017; 16: 885-896 [PMID: 28698093 DOI: 10.1016/ j.autrev.2017.07.002]
- 60 Joo SS, Won TJ, Lee DI. Potential role of ursodeoxycholic acid in suppression of nuclear factor kappa B in microglial cell line (BV-2). *Arch Pharm Res* 2004; 27: 954-960 [PMID: 15473667 DOI: 10.1007/ BF02975850]
- 61 Yanguas-Casás N, Barreda-Manso MA, Nieto-Sampedro M, Romero-Ramírez L. TUDCA: An Agonist of the Bile Acid Receptor GPBAR1/TGR5 With Anti-Inflammatory Effects in Microglial Cells. J Cell Physiol 2017; 232: 2231-2245 [PMID: 27987324 DOI: 10.1002/jcp.25742]

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- 62 **Ho PP**, Steinman L. Obeticholic acid, a synthetic bile acid agonist of the farnesoid X receptor, attenuates experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA* 2016; **113**: 1600-1605 [PMID: 26811456 DOI: 10.1073/pnas.1524890113]
- 63 Jena PK, Sheng L, Di Lucente J, Jin LW, Maezawa I, Wan YY. Dysregulated bile acid synthesis and dysbiosis are implicated in Western diet-induced systemic inflammation, microglial activation, and reduced neuroplasticity. *FASEB J* 2018; **32**: 2866-2877 [PMID:

29401580 DOI: 10.1096/fj.201700984RR]

- 64 Bhargava P, Mowry E, Calabresi P. Global metabolomics identifies perturbation of multiple metabolic pathways in Multiple Sclerosis (Abstract). *Neurology* 2015; 84: P5.242
- 65 Bhargave P. A Trial of Bile Acid Supplementation in Patients With Multiple Sclerosis. Cited 2018-05-22; In ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: URL: https://clinicaltrials.gov/ct2/show/NCT03423121

P- Reviewer: Abalo R, Quigley EM S- Editor: Gong ZM L- Editor: A E- Editor: Bian YN







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