

## REVIEW ARTICLE

## GPCRs in context: sexual dimorphism in the cardiovascular system

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**Received** 20 October 2017; **Revised** 31 January 2018; **Accepted** 9 February 2018

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Cardiovascular disease (CVD) remains the largest cause of mortality worldwide, and there is a clear gender gap in disease occurrence, with men being predisposed to earlier onset of CVD, including atherosclerosis and hypertension, relative to women. Oestrogen may be a driving factor for female-specific cardioprotection, though androgens and sex chromosomes are also likely to contribute to sexual dimorphism in the cardiovascular system (CVS). Many GPCR-mediated processes are involved in cardiovascular homeostasis, and some exhibit clear sex divergence. Here, we focus on the G protein-coupled oestrogen receptor, endothelin receptors ET<sub>A</sub> and ET<sub>B</sub> and the eicosanoid G protein-coupled receptors (GPCRs), discussing the evidence and potential mechanisms leading to gender dimorphic responses in the vasculature. The use of animal models and pharmacological tools has been essential to understanding the role of these receptors in the CVS and will be key to further delineating their sex-specific effects. Ultimately, this may illuminate wider sex differences in cardiovascular pathology and physiology.

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## Abbreviations

ASMC, aortic smooth muscle cell; CVD, cardiovascular disease; CVS, cardiovascular system; DHT, dihydrotestosterone; E<sub>2</sub>, 17 $\beta$ -oestradiol; ECE, endothelin-converting enzyme; EETs, epoxyeicosatrienoic acids; EGFR, EGF receptor; ER $\alpha$ /ER $\beta$ , oestrogen receptor  $\alpha$  or  $\beta$ ; ERK1/2, extracellular-regulated kinases 1 and 2; ET-1/2/3, endothelin 1, 2 or 3; GPER, G protein-coupled oestrogen receptor; 20-HETE, 20-hydroxyeicosatetraenoic acid; LV, left ventricle/ventricular; MAP, mean arterial pressure; mPEGES1, microsomal PGE synthase 1; OVX, ovariectomized; PTX, *Pertussis* toxin; SHR, spontaneously hypertensive rat; VSMC, vascular smooth muscle cell

## Introduction

Cardiovascular disease (CVD) is collectively responsible for one in four deaths and is currently the leading cause of death worldwide (WHO, 2014). Premenopausal women have lower blood pressure (BP) and a reduced incidence of CVD and related mortality than age-matched men (Pilote *et al.*, 2007). During menopause, the ratio of sex hormones changes dramatically: circulating **oestrogen (17 $\beta$ -oestradiol)** levels decrease by >90%, **oestrone** by 70% and **testosterone** by 40% (Rothman *et al.*, 2011) (see Figure 1A for sex hormone synthesis). Despite this decline in oestrogen abundance, the female cardiovascular death rate does not increase at the age of menopause, suggesting that oestrogen is not the only cause of female cardioprotection (Liu *et al.*, 2003). Further, hormone replacement therapy with conjugated equine oestrogens plus **medroxyprogesterone** in postmenopausal women does not reduce primary and secondary cardiovascular events to premenopausal levels (Hulley *et al.*, 1998). Therefore, it is likely that the balance of sex hormones in addition to sex chromosome complement contributes to sexual dimorphism in the cardiovascular system (CVS).

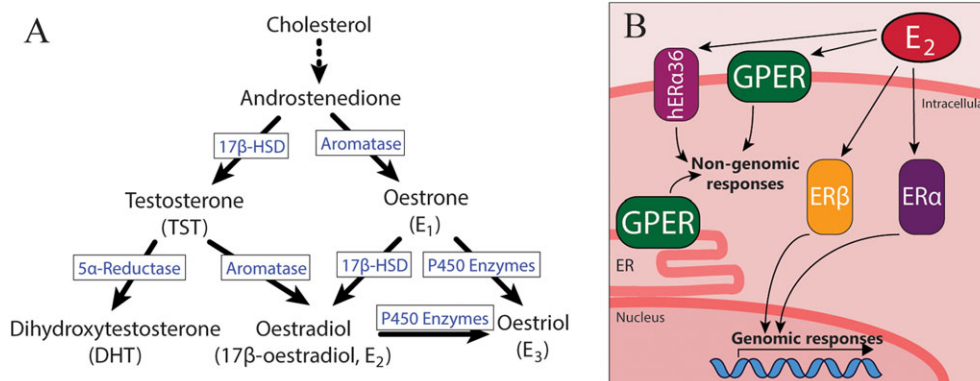
Genetic models of hypertension, including oestrogen-sensitive mRen2.Lewis (Chappell *et al.*, 2008), Dahl salt-sensitive (Hinojosa-Laborde *et al.*, 2000) and spontaneously hypertensive rats (SHRs) (Reckelhoff *et al.*, 2000), display exacerbated hypertension in males compared with females, an effect that appears to be partially mediated by sex hormones, either *via* oestrogenic protection or exacerbation by testosterone. 17 $\beta$ -Oestradiol (E<sub>2</sub>) is the principal endogenous oestrogen in females of reproductive age and is typically more physiologically potent than the other endogenous oestrogens, oestrone and **oestriol**. While E<sub>2</sub> is primarily produced by the ovaries of premenopausal women, oestradiol is also produced locally by aromatase conversion of androgens in other tissues including the vasculature, adipose tissue and the brain of both males and females (Nelson and Bulun, 2001), consistent with roles of oestrogen not only in female

reproduction but also in various physiological systems. In the CVS, E<sub>2</sub> is generally regarded as cardioprotective, with effects including vasodilation, reduction of vascular inflammation and inhibition of vascular smooth muscle cell (VSMC) proliferation. These effects of E<sub>2</sub> are both direct, through activation of cognate oestrogen receptors, and indirect, through modulation of other important receptor systems involved in cardiovascular homeostasis. When discussing differences between males and females, consistency is needed in the use of terms *sex* and *gender*. Based on definitions recommended by the Institute of Medicine (Pardue and Wizemann, 2001), the term 'gender' as used here refers to differences between men and women based on self-identity, while 'sex' is used to describe differences in animals based on chromosomal complement and reproductive organs.

In this review, we discuss the role of **GPCRs** in sexually dimorphic cardiovascular physiology and pathophysiology. GPCRs are the most successful family of drug targets to date (Rask-Andersen *et al.*, 2011) and are intimately involved in many physiological systems, positioning them as a research area of interest as novel therapeutic targets for disease. Sex differences have been noted in a number of key GPCR mediators in the CVS, most notably the angiotensin II **AT<sub>1</sub>** and **AT<sub>2</sub>** receptors of the renin-angiotensin system (Sullivan, 2008) and also vasopressin receptors (Bankir, 2001), relaxin receptors (Samuel *et al.*, 2017) and the adrenoceptors (Luzier *et al.*, 1998). For brevity, we focus on three key receptor systems that exhibit sexual dimorphism in their cardiovascular effects: G protein-coupled oestrogen receptor (GPER), endothelin (ET) receptors and eicosanoid receptors.

## G protein-coupled oestrogen receptor

The classical nuclear oestrogen receptors **ER $\alpha$**  and **ER $\beta$**  act as ligand-activated transcription factors and are responsible for the long-term effects of oestrogens (Prossnitz and Arterburn, 2015). Contrastingly, rapid oestrogenic signalling occurs *via*



**Figure 1**

Diagram of (A) sex hormone biosynthetic pathways and (B) oestrogen receptor localization within the cell. Oestrogens and androgens are both produced by sequential steps from cholesterol. Importantly, E<sub>2</sub> can be produced from both oestrogenic and androgenic precursors. E<sub>2</sub> can act on intracellular ER $\alpha$  and ER $\beta$  and their splice variants, including membrane localized human ER  $\alpha$ -36 (hER $\alpha$ 36). Additionally, the GPERs also respond to E<sub>2</sub> and is localized to either the plasma membrane or intracellular membranes depending on cell type. 17 $\beta$ -HSD, 17 $\beta$ -hydroxysteroid dehydrogenase.

membrane-associated oestrogen receptors (ERs), which include splice variants of ER $\alpha$  and a third member of the family, the GPER (Prossnitz and Arterburn, 2015) (Figure 1B).

GPER is expressed widely in vascular, lung, kidney, gastrointestinal, heart, adrenal, brain and nervous system tissues (Isensee *et al.*, 2008; Uhlen *et al.*, 2015) and is involved in the immune, nervous and cardiovascular systems, as well as cancer development, though is less important than the nuclear ERs for reproduction. The broader involvement of GPER in physiology has been reviewed extensively by Prossnitz and Arterburn (2015); here, we focus on the gender-specific effects of GPER in the CVS.

### GPER ligands and signalling

GPER (previously known as GPR30) was relatively recently deorphanized as an oestrogen receptor, with high affinity for 17 $\beta$ -oestradiol ( $pK_i = 8.2$ – $8.5$  nM) and low affinity for other endogenous oestrogens (Revankar *et al.*, 2005; Thomas *et al.*, 2005). GPER exhibits >1000-fold selectivity for E<sub>2</sub> over testosterone, **cortisol** and **progesterone** (Thomas *et al.*, 2005). In addition to endogenous estrogenic compounds, a number of phytoestrogens and xenoestrogens, including **resveratrol**, **quercetin** and **bisphenol A**, have also been shown to have agonist activity at GPER (Prossnitz and Arterburn, 2015). Ligands with high selectivity for GPER over ER $\alpha$  and ER $\beta$  have been developed to probe GPER physiology *in vivo* and *in vitro* and include agonist **G-1** (Bologa *et al.*, 2006) and antagonists **G15** (Dennis *et al.*, 2009) and **G36** (Dennis *et al.*, 2011). Interestingly, two compounds that are used clinically as negative modulators of nuclear ER activity, **tamoxifen** and **fulvestrant (ICI 182 780)**, are both full agonists at GPER (Thomas *et al.*, 2005).

Understanding the pharmacology of GPER has been complicated by seemingly discrepant results, and indeed, subcellular localization and cell-type specific signalling of GPER remain controversial. While GPER was initially identified as a membrane-associated oestrogen receptor mediating rapid signalling in response to E<sub>2</sub> (Filardo *et al.*, 2000; Thomas *et al.*, 2005), there is also evidence for the localization of GPER to intracellular sites, such as the Golgi and endoplasmic reticulum (Revankar *et al.*, 2005; Otto *et al.*, 2008). Additionally, changes in subcellular localization of GPER were observed throughout the oestrous cycle in mice (Cheng *et al.*, 2014); hence, sex hormone abundance may account for the discrepancies in cellular GPER distribution.

To add further complexity, GPER appears to signal through both G $\alpha_s$  and G $\alpha_{i/o}$  G proteins, sometimes in the same cell type. Filardo *et al.* (2000) initially postulated GPER to be G $\alpha_{i/o}$  coupled, leading to transactivation of the **EGF receptor (EGFR)**, initiation of **extracellular-regulated kinase (ERK)** phosphorylation, **PI3K/Akt** signalling and calcium mobilization. Subsequently, the same group reported G $\alpha_s$  signalling, causing cAMP generation that quenched activation of ERK1/2 generated by the G $\alpha_{i/o}$  pathway (Filardo *et al.*, 2002). A GPER-mediated increase in intracellular cAMP was also reported by Thomas *et al.* (2005). However, while an independent group did confirm that EGFR transactivation is a downstream effect of GPER stimulation and that it occurs in both COS-7 and the MDA-MB-231 cells used by Filardo *et al.* (2000), they found no evidence of EGFR activation being sensitive to the G $\alpha_{i/o}$  inhibitor, *Pertussis* toxin (PTX), suggesting

that this element of GPER signalling does not involve coupling to G $\alpha_{i/o}$  (Revankar *et al.*, 2005). Interestingly, this report also showed that GPER-mediated calcium mobilization is partly inhibited by PTX and is also partly dependent on EGFR activation (Revankar *et al.*, 2005), suggesting multiplicity of signalling modes for GPER. These discordant results highlight the complexity of GPER pharmacology and the likelihood of cell type-specific signalling pathways.

### Tissue expression and role in the cardiovascular system

Direct vasodilatory effects of oestrogen in the vasculature are likely to be mediated by GPER. In isolated aortae of female mRen2.Lewis rats precontracted with phenylephrine, the magnitude of vasodilation produced by E<sub>2</sub> was identical to that produced by the agonist G-1 (Lindsey *et al.*, 2011a). Additionally, acute i.v. infusion of G-1 also caused a dose-dependent decrease in BP in normotensive male Sprague–Dawley rats of up to 15% within minutes (Haas *et al.*, 2009), while chronic infusion of G-1 for 2 weeks lowers mean arterial pressure (MAP) by 20% in the hypertensive ovariectomized (OVX) mRen2.Lewis rats (Lindsey *et al.*, 2009). However, in this study, G-1 infusion failed to reduce the BP of normotensive intact female and hypertensive male mRen2.Lewis rats (Lindsey *et al.*, 2009), potentially implicating GPER in BP control in males at baseline and in females during chronic hypertension. Furthermore, GPER plays an important role in maintaining basal tone in arteries, as shown by enhanced vasoconstriction and impaired vasodilation in response to the GPER antagonist G15 (Meyer *et al.*, 2012a). In addition to acute effects on the circulatory system, abundance of GPER in kidney (Kurt and Buyukafsar, 2013) and the CNS, particularly in the autonomic nuclei of the brainstem and the hypothalamic–pituitary axis (Brailoiu *et al.*, 2007), suggests a role for GPER in long-term maintenance of BP homeostasis. As such, GPER is important for both acute and chronic cardiovascular responses to oestrogen.

Despite uncertainty surrounding the coupling of GPER, as discussed above, the downstream effectors that mediate vasodilation are generally agreed upon. Within the vasculature, GPER-positive immunostaining was reported in both endothelial and VSMCs in rat thoracic aorta (Lindsey *et al.*, 2009), as well as rat carotid and middle cerebral arteries of both sexes (Broughton *et al.*, 2010), in agreement with a GPER-lacZ reporter mouse model (Isensee *et al.*, 2008) and consistent with both endothelium-dependent and endothelium-independent actions of GPER agonists. GPER-dependent vasodilation is mediated both by the endothelium and VSMCs, as endothelial denudation reduces, but does not completely abolish, G-1-stimulated vasodilation (Lindsey *et al.*, 2009). Activation of endothelial GPER triggers calcium mobilization and PI3K/Akt activation to produce nitric oxide (**NO**), as well as potassium efflux and membrane hyperpolarization, both of which lead to VSMC relaxation (Meyer *et al.*, 2012a; Lindsey *et al.*, 2014). G-1 also acts directly on VSMCs to induce vasodilation *via* cAMP accumulation and modulation of large conductance potassium channels (Lindsey *et al.*, 2014). Interestingly, GPER agonists produce similar degrees of vasorelaxation in isolated carotid arteries from male and female rats (Broughton *et al.*, 2010). Additionally, there

is evidence for antioxidant effects of GPER agonism with G-1 in the kidney (Lindsey *et al.*, 2011b), and G-1 itself may act directly as a scavenger of superoxide anions, further increasing its vasculoprotective actions (Broughton *et al.*, 2010).

There is evidence for regulation of GPER expression by female sex hormones, with higher protein abundance in the kidney during oestrus and pro-oestrus (Cheng *et al.*, 2014) and increased mRNA and protein expression in SKBR3 cells following prolonged treatment with progesterone or E<sub>2</sub> (Thomas *et al.*, 2005). However, GPER protein is expressed in rodent heart tissue at similar levels in both sexes (Deschamps and Murphy, 2009).

Genetic deletion studies have given further clues to the metabolic and cardiovascular effects of GPER, although debate remains due to discordant results. Female mice of the GPER knockout mouse line described by Martensson *et al.* (2009) had 23% higher MAP at 9 months, due to changes in resistance artery structure. The female GPER null mice also had impaired metabolism, with reduced body weight due to impaired skeletal growth (though no alterations in white adipose tissue relative to body weight) and hyperglycaemia due to glucose intolerance. No such metabolic or cardiovascular effects were seen in males. Contrastingly, Haas *et al.* (2009) observed increased body weight and adiposity in GPER<sup>-/-</sup> mice of both sexes. In further contrast to Martensson *et al.* (2009), no BP difference between genotypes was seen by Isensee *et al.* (2008), though this may be attributable to the animals being studied at a younger age or the mixed-strain background used by Martensson *et al.* (2009), or differing gene targeting strategies (cre/lox system used by Martensson *et al.* (2009), Neo<sup>+</sup>/LacZ cassette insertion into exon 3 used by Isensee *et al.* (2008)). Intriguingly, Wang *et al.* (2016) report that cardiomyocyte-specific deletion of GPER causes left ventricular (LV) dysfunction and remodelling in both sexes but a male-specific inflammatory response. This result is difficult to reconcile with the above studies, which show no deleterious cardiovascular effects in male mice with global GPER gene deletion, especially considering the similar expression of cardiac GPER between the sexes (Deschamps and Murphy, 2009). It is unlikely that these discrepancies are due to compensatory increases in the expression of classical ERs, as both Wang *et al.* (2016) and Martensson *et al.* (2009) reported no change in mRNA abundance of ER $\alpha$  and ER $\beta$ . Thus, despite GPER being a receptor for a primarily female sex hormone, there is surprisingly little conclusive evidence for sex differences in effects of this receptor, exacerbated by the fact that many initial BP characterization studies in rodents did not compare both sexes simultaneously (Haas *et al.*, 2009; Lindsey *et al.*, 2011a; Meyer *et al.*, 2012a).

### Interactions of GPER with the angiotensin system

Oestrogen is known to decrease expression of the prohypertensive AT<sub>1</sub> receptor, and it is likely that actions of GPER contribute to this. Treatment with GPER agonist, G-1, in OVX mRen2.Lewis rats reduced mRNA for AT<sub>1</sub> receptors in the aorta (Lindsey *et al.*, 2009) but had no effect on the abundance of AT<sub>2</sub> receptor transcript. In addition, the oestradiol metabolite, 2-methoxyestradiol, exerts similar effects on AT<sub>1</sub> receptor expression in a GPER-dependent and

EGFR-dependent manner (Koganti *et al.*, 2014), which may partially explain the female-specific cardioprotective effects of **CYP450 1B1**, which produces 2-methoxyestradiol (Pingili *et al.*, 2017).

### Interactions of GPER with the endothelin system

GPER is known to inhibit **ET-1**-induced contractions in isolated porcine coronary arteries (Meyer *et al.*, 2010). Similarly, isolated carotid arteries from GPER null mice had augmented contractile responses to ET-1, indicating that endogenous GPER dulls the vasoconstrictive response to ET-1, an effect that may be due to decreased sensitivity of contractile machinery to Ca<sup>2+</sup> release in VSMCs (Meyer *et al.*, 2012b). Additionally, GPER<sup>-/-</sup> mice do not exhibit an increase in **ET<sub>B</sub>** receptors and **endothelin-converting enzyme (ECE)-2** in the ageing myocardium, ET system components, which may be associated with heart failure and hypertrophy in older animals (Meyer *et al.*, 2016). Interestingly, there is also evidence that ET-1 increases mRNA expression of GPER and that GPER is necessary for some downstream actions of ET-1 in SKBR3 and HepG2 cell lines (Bartella *et al.*, 2016).

### Potential for GPER–mineralocorticoid receptor crosstalk

GPER appears to be necessary for some rapid actions of **aldosterone** (Gros *et al.*, 2013) which may be more efficacious than oestrogen to activate GPER-dependent downstream signalling (Gros *et al.*, 2011). However, competition binding assays indicate that aldosterone does not directly bind to GPER, at least not at the same site as E<sub>2</sub>, and does not cause [<sup>35</sup>S]GTP $\gamma$ S binding as E<sub>2</sub> does (Cheng *et al.*, 2014). This excludes GPER as a receptor for aldosterone but may imply functional crosstalk between GPER and the **mineralocorticoid receptor (NR3C2)**, the cognate receptor for aldosterone.

### Clinical relevance

The GPER maps to human chromosome 7p22.3, a locus that has been associated with hypertension due to familial hyperaldosteronism-II (Lafferty *et al.*, 2000), suggesting a role in BP homeostasis. A single nucleotide polymorphism (rs11544331) in the gene is associated with elevated BP in women but not men. Furthermore, in patients with resistant hypertension, females are almost twice as likely to carry the variant, with an allele frequency of 31% compared with 16% (Feldman *et al.*, 2014), suggesting that, particularly in women, GPER plays a role in BP regulation. In rat aortic VSMCs, this Phe16Leu variant showed hyporesponsiveness and a reduced ability to stimulate ERK phosphorylation and apoptosis in response to G-1 compared with animals expressing wild-type GPER (Feldman *et al.*, 2014).

The therapeutic potential of GPER in the CVS is fourfold. Firstly, GPER itself may be a useful target for hypertension, considering that G-1 has both acute and chronic antihypertensive actions in males and females (Haas *et al.*, 2009; Lindsey *et al.*, 2009), as well as protective effects on LV function and remodelling (Wang *et al.*, 2012), and will not activate potentially deleterious classical ER signalling in postmenopausal women (Hulley *et al.*, 1998). Secondly, GPER agonism has protective effects following myocardial

ischaemia/reperfusion injury, as prior treatment with G-1 reduces infarction size and improves recovery of contractile function in a rat model of ischaemic injury (Deschamps and Murphy, 2009). Thirdly, GPER antagonism by centrally administered G15 reduces stroke size and neurological deficit in male mice following a 1 h period of middle cerebral artery occlusion, even when administered up to 3 h after the ischaemic event (Broughton *et al.*, 2014). Lastly, GPER may also be an attractive candidate in atherosclerosis treatment and prevention; anti-inflammatory effects of oestrogen are likely partially mediated by GPER, and it is known that **raloxifene** (a selective oestrogen receptor modulator and GPER agonist) provides protection against atherosclerotic processes (Prossnitz and Barton, 2009). Agonism of GPER also has anti-inflammatory and anti-proliferative effects; in umbilical vein endothelial cells, G-1 abolished the pro-inflammatory effects of TNF- $\alpha$ , which is known to be involved in atherosclerosis and other inflammatory processes (Chakrabarti and Davidge, 2012). Similarly, G-1 and ICI 182780 inhibited serum-stimulated proliferation of human VSMCs (Haas *et al.*, 2009), which might be a valuable therapeutic approach for preventing excessive VSMC proliferation, as seen in atherosclerosis and following coronary stent implantation.

## Endothelins and endothelin receptors

ET-1 is well known as one of the most important biological vasoconstrictors, with an unusually potent and prolonged action (Yanagisawa *et al.*, 1988). ET-1, along with two other isoforms, **ET-2** and **ET-3**, comprise the family of biologically active ET peptides. ET-1 and ET-2 have equal affinity for both ET receptors, **ET<sub>A</sub>** and **ET<sub>B</sub>**, while ET-3 has higher affinity for **ET<sub>B</sub>** over **ET<sub>A</sub>** receptors (see Figure 2 for a schematic representation of ET receptor ligands mentioned in this review; for a more comprehensive list, please refer to Davenport *et al.*, 2016). All are produced from inactive precursors by the action of ECEs. There are established sex differences in the ET system that may partially explain sex differences in the development

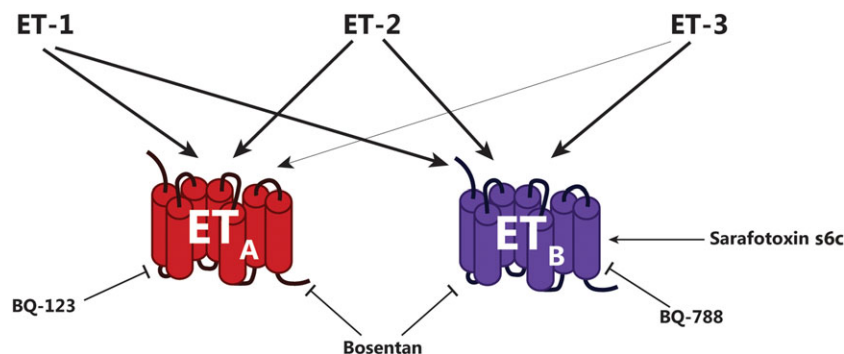
of hypertension and other CVD, previously reviewed by Gillis *et al.* (2016) but briefly summarized here.

Female gonadal hormones have depressant effects on circulating ET-1 levels. Ovariectomy of Sprague–Dawley rats increases plasma concentration of ET-1 and aortic expression of prepro-ET-1 (Tan *et al.*, 2003), indicating that female hormones favourably decrease pro-hypertensive ET system components. As well as  $E_2$ , oestrogen metabolites, 2-hydroxyestradiol and 2-methoxyestradiol, inhibited ET-1 release from porcine coronary artery endothelial cells, which was not blocked by the ER $\alpha$ /ER $\beta$  antagonist, ICI 182780, indicating a mechanism that is independent of nuclear ERs (Dubey *et al.*, 2001). Interestingly, there is evidence for involvement of GPER in the response to ET-1; GPER inhibits vasoconstrictor responses to ET-1, while in isolated arteries from GPER<sup>-/-</sup> mice, ET-1-induced vasoconstriction is enhanced due to sensitization of myofilaments to calcium release (Meyer *et al.*, 2012b).

### ET<sub>A</sub> receptors

ET<sub>A</sub>, a G $\alpha_{q/11}$ -coupled GPCR, is expressed ubiquitously in VSMCs and promotes potent vasoconstriction through intracellular calcium release. The ET<sub>A</sub> receptor is the predominant subtype expressed in cardiomyocytes, where it mediates positive inotropic effects (Davenport *et al.*, 2016). It is known that female sex hormones influence expression of ET<sub>A</sub> receptors. Hormone replacement with  $E_2$  or conjugated equine oestrogens decreases ET<sub>A</sub> receptor mRNA expression in the aorta of OVX New Zealand white rabbits (Pedersen *et al.*, 2009). Similarly,  $E_2$  treatment reduces ET<sub>A</sub> receptor mRNA expression in lungs of OVX Sprague–Dawley rats (Gohar *et al.*, 2016).

Intriguingly, women retain differential expression of ET receptor subtypes even after menopause. Saphenous veins isolated from men have 4–7 times higher ET receptor density than those from women, as well as a higher ratio of ET<sub>A</sub> to ET<sub>B</sub> receptors than women (approximately 3:1 compared with 1:1), favouring the constrictive actions of ET-1. This is reflected by the maximal vessel constriction to 1  $\mu$ M ET-1



**Figure 2**

Endothelin receptors and ligands. Relative potency of ET isoforms for respective receptors is indicated by thickness of arrows. Potency order of endogenous agonists for ET<sub>A</sub> receptors: ET-1 = ET-2 >> ET-3. Potency order of endogenous agonists for ET<sub>B</sub> receptors: ET-1 = ET-2 = ET-3. Compounds mentioned in the text as pharmacological modulators for the ET<sub>A</sub> and ET<sub>B</sub> receptors are shown: BQ-123 (selective ET<sub>A</sub> receptor antagonist), bosentan (unbiased ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist), BQ-788 (selective ET<sub>B</sub> receptor antagonist) and sarafotoxin s6c (selective ET<sub>B</sub> receptor agonist).

being nearly doubled in men (Ergul *et al.*, 1998). Consistent with higher abundance of vasoconstrictive ET<sub>A</sub> receptors in male blood vessels, selective blockade of ET<sub>A</sub> receptors (with **BQ-123**) increases forearm blood flow in males more than females, while dual blockade of ET<sub>A</sub> and ET<sub>B</sub> receptors (**BQ-123** and **BQ-788**) produced equal increases in blood flow between the genders, suggesting ET<sub>B</sub> receptors have a more significant role in women than men (Stauffer *et al.*, 2010). Both studies involved women of postmenopausal age that were not taking hormone replacement therapy, suggesting that oestrogen is not the sole contributor to sex-specific ET<sub>A</sub> receptor expression and that sex chromosomes or other developmental factors may play a role.

### ET<sub>B</sub> receptors

The ET<sub>B</sub> receptor is functionally distinct from the ET<sub>A</sub> receptor, mediating effects that generally oppose the prohypertensive actions of the ET<sub>A</sub> receptors. ET<sub>B</sub> receptors are expressed abundantly in lung and kidney tissue, where they function as clearance receptors to remove excess ET-1 from the circulation, preventing unnecessary activation of ET<sub>A</sub> receptors (Fukuroda *et al.*, 1994). The ET<sub>B</sub> receptor also promotes natriuresis and diuresis by direct actions on renal tubules (Nakano *et al.*, 2008; Kohan *et al.*, 2011). In the vasculature, ET<sub>B</sub> receptors are localized to the endothelium, where they promote NO production and release of vasorelaxant COX metabolites to induce endothelium-dependent vasodilation. VSMCs in certain vessel types also express the ET<sub>B</sub> receptor, which mediates sex-specific and tissue-specific vasoconstriction (Kellogg *et al.*, 2001; Schneider *et al.*, 2007).

Regarding the sex differences in the effects of this receptor, initial studies suggested that ET<sub>B</sub> receptor deficiency causes high BP only in males at baseline, while female rats deficient in ET<sub>B</sub> receptors develop more severe hypertension than males following a high-salt diet (Taylor *et al.*, 2003). However, this was later determined to be an artefact of the tail cuff method of BP measurement. Radiotelemetry measurements revealed that salt-induced hypertension in ET<sub>B</sub> receptor-deficient rats is comparable between the sexes, though ET<sub>B</sub> receptor-deficient females are more sensitive than controls to acute stress on both normal and high-salt diets, explaining the higher BP observed when measured by tail cuff (Speed *et al.*, 2015). The greater production of ROS in the female ET<sub>B</sub> receptor-deficient mice is a potential contributor to this more severe response (Sullivan *et al.*, 2006), indicating that ET<sub>B</sub> receptors are involved in protection against BP increase in females (Kittikulsuth *et al.*, 2013).

A possible explanation for these sex-specific effects of ET<sub>B</sub> receptors is that females have lower expression ratios of ET<sub>A</sub> to ET<sub>B</sub> receptors, compared with males, as mentioned above (Ergul *et al.*, 1998), and vascular mRNA abundance of ET<sub>B</sub> receptors is increased in male but not female DOCA-salt hypertensive rats (David *et al.*, 2002). This may be due to direct effects of female sex hormones, as E<sub>2</sub> treatment in OVX rabbits increased the mRNA for ET<sub>B</sub> receptors in coronary vessels and attenuated vasoconstriction by ET-1 (Pedersen *et al.*, 2008). Contrastingly, other studies have shown ET<sub>B</sub> receptor transcript abundance to be increased by loss of female hormones (OVX) and decreased LV and renal inner medulla following E<sub>2</sub> replacement (Nuedling *et al.*, 2003; Gohar *et al.*, 2016). While exogenous oestrogen may decrease ET<sub>B</sub> receptor

transcript levels in the inner medulla of females, receptor density between the sexes is comparable at baseline (Jin *et al.*, 2013), maintaining the lower ET<sub>A</sub> : ET<sub>B</sub> ratio of females. These disparate results demonstrate that ET<sub>B</sub> receptor expression and trafficking are likely mediated by sex hormones in a complex, tissue-specific manner and at present cannot explain the sexually divergent roles of this receptor.

### Interactions between the endothelin and angiotensin systems

In the **angiotensin II** infusion model of hypertension, renal ET<sub>B</sub> receptor density is decreased in male hypertensive Sprague–Dawley rats compared with saline-infused controls, while density is preserved in females (Kittikulsuth *et al.*, 2011). As shown in this study and others, the angiotensin II hypertension model produces more profound increases in MAP in males than in females (Kittikulsuth *et al.*, 2011; Xue *et al.*, 2013), and the reduction in ET<sub>B</sub> receptor abundance in males specifically may partly explain this.

### Clinical relevance

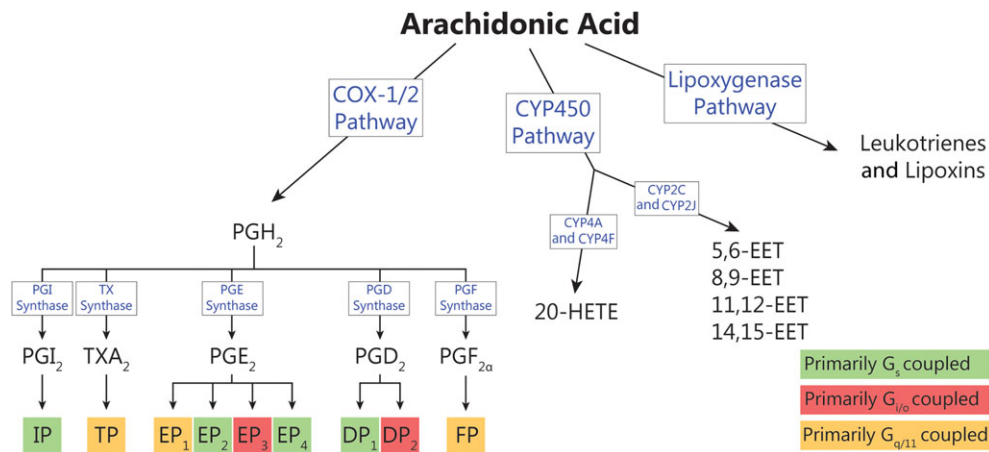
**Bosentan**, a dual ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist and the first clinically used drug to target the ET receptors, is one of several ET antagonists approved for the treatment of pulmonary hypertension, which primarily affects women (Seeland and Regitz-Zagrosek, 2012). A variant (Gly5565Thr) in the prepro-ET-1 gene previously shown to be associated with higher BP also affects patient outcomes to antihypertensive treatment in a gender-specific manner. In patients given either an angiotensin receptor blocker (**irbesartan**) or a beta-blocker (**atenolol**), male carriers of the T allele have greater reductions in systolic BP compared with those with GG genotype, while systolic BP reduction in females was not different between genotypes (Hallberg *et al.*, 2004).

In addition to this, elevated plasma ET-1 is associated with endothelial dysfunction and may be a clinical predictor of cardiovascular risk, particularly in women (Daka *et al.*, 2015). Higher circulating ET-1 levels are associated with coronary heart disease events in women but not men, independent of other risk factors and levels of circulating oestradiol (Daka *et al.*, 2015). Even in a young cohort of 2160 men and women aged between 25 and 41 without diabetes or body mass index scores above 35 kg·m<sup>-2</sup>, higher plasma ET-1 concentration was associated with elevated systolic BP and higher cardiovascular risk estimated by the Prospective Cardiovascular Münster and Framingham scores (Bossard *et al.*, 2015), though this was not reported by gender.

### Eicosanoids and eicosanoid GPCRs

Arachidonic acid is the metabolic precursor for eicosanoids, including prostanoids, epoxyeicosatrienoic acids (EETs) and leukotrienes. COX activity at arachidonic acid produces **PGH<sub>2</sub>** from which the primary bioactive prostanoids, **PGD<sub>2</sub>**, **PGE<sub>2</sub>** and **PGF<sub>2α</sub>**, **PGI<sub>2</sub>** and **TXA<sub>2</sub>** are derived. These act at their cognate GPCRs; DP<sub>1-2</sub>, EP<sub>1-4</sub>, **FP**, **IP** and **TP** receptors, as summarized in Figure 3.

Oestrogens have broad effects on the biosynthesis of prostanoids, including decreasing activity of **COX-1** and **COX-2**. There is evidence that many of these actions are



**Figure 3**

Metabolic pathways of arachidonic acid. COX-1/2 metabolism of arachidonic acid produces bioactive eicosanoid hormones; PGI<sub>2</sub>, TXA<sub>2</sub>, PGE<sub>2</sub>, PGD<sub>2</sub> and PGF<sub>2α</sub>. These prostanoids then act at nine cognate GPCRs: IP, TP, EP<sub>1-4</sub>, DP<sub>1,2</sub> and FP receptors. Actions of CYP450 enzymes on arachidonic acid produce 20-HETE and EETs, which may act through GPCRs. The final metabolic pathway of arachidonic acid produces leukotrienes and lipoxins *via* lipoxygenase activity, which are not discussed in this review.

mediated by GPER, as it inhibits prostanoid production and activity (Meyer *et al.*, 2015), while loss of GPER is associated with increased endothelial prostanoid-mediated vasoconstriction and increases in TP receptor-mediated contractions (Meyer *et al.*, 2012a). In the CVS, the prostanoid receptors producing significant sexually dimorphic phenotypes are the TP, IP and EP receptors, discussed below.

### TXA<sub>2</sub> (TP) receptors

The prostanoid TXA<sub>2</sub> is a potent activator of platelet aggregation, and inducer of vasoconstriction and vascular cell proliferation. This prostanoid is the primary agonist of the TP receptors, though other prostanoids including PGH<sub>2</sub> are also TP receptor agonists. This receptor is known to couple to G<sub>α<sub>q/11</sub></sub> proteins and activates Ca<sup>2+</sup>/DAG signalling (Kinsella *et al.*, 1997).

It has been long established that there are gender differences in response to TP receptor agonists. Following administration of **U46619**, a synthetic and stable TP receptor agonist, there was a 25% greater contraction in isolated aortae from male rats than from female, with no sex differences in contractile responses after **5-HT** and **noradrenaline** treatment (Karaniyan *et al.*, 1981). Furthermore, TP receptor activation increases synthesis of the vasoconstrictor **20-hydroxyeicosatetraenoic acid (20-HETE)** to three times that of basal levels in small porcine coronary arteries, potentially exacerbating effects of androgen-regulated 20-HETE synthesis in males (Randriamboavonjy *et al.*, 2005). *In vivo*, U46619 increases MAP by approximately 25 mmHg in male SHR, without any significant effect on MAP of females (Schirner and Taube, 1993).

Sex differences in sensitivity to TP receptor agonists may be due to changes in receptor abundance driven by the hormonal milieu. Testosterone increases TP receptor density in cultured male rat aortic smooth muscle cells (ASMCs) and male guinea pig coronary artery smooth muscle cells (Higashiura *et al.*, 1997). The more active testosterone metabolite, **dihydrotestosterone (DHT)**, also increased receptor

density in male rat ASMCs; though in females ASMCs, the increase was comparatively minor (Higashiura *et al.*, 1997). Acute administration of testosterone (two doses of 200 mg) to healthy men increased platelet TP receptor density and platelet aggregation response (Ajayi *et al.*, 1995). Androgens thus increase expression of TP receptors, and this effect appears to be augmented in males. Contrastingly, E<sub>2</sub> has no effect on TP receptor density in cultured rat ASMCs (Masuda *et al.*, 1991). However, pretreatment with E<sub>2</sub> desensitizes arteries to agonists of the TP receptors, causing decreased calcium influx and intracellular calcium release in response to U46619 in isolated porcine coronary arteries from both sexes in a dose-dependent manner (Han *et al.*, 1995), which may be attributable to changes in receptor trafficking mechanisms rather than changes in expression. The sensitizing effects of male gonadal hormones and converse effects of oestrogens partly explains why male arteries are more sensitive to contractile effects of TXA<sub>2</sub> and mimetics.

### Prostacyclin (IP) receptors

PGI<sub>2</sub> is produced in vascular endothelium and VSMCs from arachidonic acid *via* COX-1/2 and subsequent PGI<sub>2</sub> synthase activity. PGI<sub>2</sub> opposes the pro-thrombotic effects of TXA<sub>2</sub> to prevent platelet aggregation and reduce vascular proliferation, as well as acting as a potent vasodilator and promoting repair following vascular injury (Miggin and Kinsella, 2002). PGI<sub>2</sub> is the main endogenous ligand for the IP receptor, which primarily couples to G<sub>α<sub>s</sub></sub> and **adenylate cyclase**, though is also capable of signalling *via* G<sub>α<sub>i/o</sub></sub> and G<sub>α<sub>q/11</sub></sub> to inhibit adenylate cyclase and stimulate phospholipase C (**PLC**) respectively (Miggin and Kinsella, 2002). Studies in IP receptor-deficient mice indicate that perturbations in receptor function cause vulnerability to CVD such as ischaemic heart disease due to enhanced thrombosis, without changes in BP (Murata *et al.*, 1997), suggesting that the IP receptor is involved in acute responses rather than regulation of basal BP. In the LDL receptor<sup>-/-</sup> mouse model of atherosclerosis, female mice lacking the IP receptor developed more severe

lesions than controls, an effect that was gene dose-dependent and not seen in males. In addition to this, platelet activation associated with early atherogenesis was more severe in IP receptor-deficient females than males (Egan *et al.*, 2004). As such, atheroprotective actions of IP receptors are pertinent to both sexes, though loss of IP receptors is additionally deleterious in females.

Both the IP receptor and its ligand, PGI<sub>2</sub>, are regulated by sex hormones. Prostacyclin production in human endothelial cells was stimulated in a dose-dependent manner by E<sub>2</sub> and inhibited by selective oestrogen receptor modulator, tamoxifen (Mikkola *et al.*, 1995). This was reproduced in rat cerebral blood vessels, in which E<sub>2</sub> replacement after ovariectomy increases protein expression of COX-1 by five times and PGI<sub>2</sub> synthase by six times that of basal levels (Ospina *et al.*, 2002). Interestingly, both E<sub>2</sub> and DHT increased the abundance of IP receptor mRNA and protein *in vitro*, as a result of oestrogen and androgen response elements in the IP receptor gene promoter region (Eivers and Kinsella, 2016). The finding that the testosterone metabolite, DHT, can mediate expression of IP receptors may account for some of the cardioprotective effects of androgens in the CVS.

### PGE<sub>2</sub> receptors (EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub> and EP<sub>4</sub>)

PGE<sub>2</sub> is the major COX-2 metabolite in the kidney and is important for renal function through its vasodilatory and natriuretic functions (Breyer and Breyer, 2001). It is the principal endogenous ligand for the EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub> and EP<sub>4</sub> receptors, though each have distinct affinity profiles for other prostanoids. EP<sub>1</sub> receptors couple to G<sub>α<sub>q/11</sub></sub> causing increased cellular calcium, EP<sub>2</sub> and EP<sub>4</sub> receptors signal through G<sub>α<sub>s</sub></sub> to raise cAMP, while EP<sub>3</sub> receptors are G<sub>α<sub>i/o</sub></sub> coupled and inhibit cAMP production. EP<sub>4</sub> and EP<sub>2</sub> receptors are involved in vasorelaxation in response to PGE<sub>2</sub>, while EP<sub>1</sub> receptors are responsible for transient vasoconstriction (Purdy and Arendshorst, 2000).

Delineating the individual contributions of PGE receptor subtypes to male and female BP homeostasis has proven difficult and is currently understudied. One group investigated the roles of each subtype in male and female mice in response to acute i.v. administration of a bolus dose of PGE<sub>2</sub>, which caused an immediate reduction in MAP of both sexes, though this was greater in females (−30 mmHg compared with −23 mmHg in males) (Audoly *et al.*, 1999). Additionally, the maximal MAP reduction is blunted by EP<sub>2</sub> or EP<sub>4</sub> receptor knockout in females but not males, suggesting that EP<sub>2</sub> and EP<sub>4</sub> receptors, both G<sub>α<sub>s</sub></sub> coupled, are the primary subtypes mediating acute vasodilatory action of PGE<sub>2</sub> in females. Correspondingly, knockout of the G<sub>α<sub>i/o</sub></sub>-coupled EP<sub>3</sub> receptors augmented MAP reduction in response to PGE<sub>2</sub> in males only (Audoly *et al.*, 1999). Interestingly, the genetic deletion of EP<sub>1</sub> receptors results in a systolic BP reduction of approximately 10 mmHg in males only and was associated with compensatory effects including increased heart rate and a doubling of renin–angiotensin activity compared with controls (Audoly *et al.*, 1999). This suggests a role for EP<sub>1</sub> receptors in long-term BP homeostasis in males (Stock *et al.*, 2001).

A further point of sexual dimorphism in PGE receptor mechanisms may be the synthesis of the ligand, PGE<sub>2</sub>, produced by the sequential actions of COX-1/2 and PGE

synthases on PGH<sub>2</sub>. COX-2 is the main producer of PGE<sub>2</sub> in the kidney and shown to be higher in female DBA/1 mouse whole kidney homogenate, while ablation of the **microsomal PGE synthase 1 (mPGES1)** in mice causes a number of sex-specific effects on prostanoid synthesis and metabolism (Francois *et al.*, 2007). Further, females have higher expression of mPGES1 mRNA in the kidney at baseline and nearly two-fold higher PGE<sub>2</sub> levels, as indicated by urinary excretion of metabolites (Francois *et al.*, 2007). Higher PGE<sub>2</sub> production in the kidney of females may increase its beneficial natriuretic functions and provides further evidence for female-specific cardiovascular advantages.

### CYP450 metabolites of arachidonic acid (EETs and 20-HETE)

The cytochrome P450 arm of arachidonic acid metabolism produces EETs and 20-HETE. Both EETs and 20-HETE are vasoactive and have roles in regulation of renal tubular sodium transport and pressure–natriuresis (Fan *et al.*, 2015). **CYP2C** and **CYP2J epoxygenases** are primarily responsible for producing EETs, which have anti-inflammatory and vasodilator capacity. These effects may be *via* a cognate receptor, with some evidence for EETs binding to a G<sub>α<sub>s</sub></sub> coupled receptor with 11,12-EET increasing the binding of [<sup>35</sup>S]-GTPγS to G<sub>α<sub>s</sub></sub> but not G<sub>α<sub>i</sub></sub> proteins (Node *et al.*, 2001). However, a screen of 79 orphan receptors in HEK293 cells with a photoaffinity-labelled 14,15-EET analogue showed no evidence of binding (Chen *et al.*, 2011). An alternative possibility for the mechanistic basis for effects of EETs is their antagonist activity at TP receptors, relaxing arteries precontracted with TXA<sub>2</sub> and preventing binding of a specific TP receptor antagonist (Behm *et al.*, 2009). EETs may also act as secondary mediators of anti-inflammatory effects of the stimulation of AT<sub>2</sub> receptors (Rompe *et al.*, 2010). There is some evidence for gonadal hormone involvement in biosynthesis of EETs, as DHT treatment reduced renal mRNA for the primary epoxygenase CYP2C23 (Singh and Schwartzman, 2008).

Contrastingly, 20-HETE stimulates VSMC contraction and endothelial cell dysfunction and is produced by androgen-stimulated ω-hydroxylases of the CYP4A and CYP4F subfamilies. 20-HETE may contribute to the higher basal BP seen in males by androgen-induced ω-hydroxylase expression (Holla *et al.*, 2001; Wu and Schwartzman, 2011). Recent evidence suggests that a current orphan GPCR, **GPR75**, may be a receptor for 20-HETE (Garcia *et al.*, 2017). GPR75 had previously been reported as a receptor for the chemokine CCL5 (Ignatov *et al.*, 2006), although this pairing has not been confirmed (Southern *et al.*, 2013; Garcia *et al.*, 2017). In cultured human microvascular endothelial cells, 20-HETE induced inositol phosphate accumulation, consistent with GPR75 acting as a G<sub>α<sub>q/11</sub></sub>-coupled GPCR, and phosphorylation of EGFR, which is known to be activated by GPR75. Importantly, siRNA knockdown of GPR75 in these cells prevents induction of the mRNA for **angiotensin-converting enzyme**, which is a major effect of 20-HETE on the vasculature. Furthermore, GPR75 shRNA knockdown in mice completely attenuates hypertension and vascular remodelling caused by doxycycline-induced expression of CYP4A12 (a murine 20-HETE



synthase) (Garcia *et al.*, 2017). Alternatively, 20-HETE may exert its vascular effects by acting at TP receptors, as flow-induced constriction of rat and human arteries was abolished by an inhibitor of 20-HETE synthesis (HET0016) or a TP receptor antagonist (**SQ 29548**) but not COX inhibitor (**indomethacin**) or a specific TXA<sub>2</sub> synthase inhibitor (ozagrel) (Toth *et al.*, 2011), though direct binding of 20-HETE to the TP receptor has not been demonstrated.

There are significant effects of gender on levels of 20-HETE. The male ratio of 20-HETE to EETs in interlobar arteries is almost twice as high as females, and treatment with DHT increased this ratio in both sexes and eliminated sex difference (Singh and Schwartzman, 2008). DHT treatment also increased mRNA expression of CYP4A8, the primary hydroxylase responsible for 20-HETE production in the kidney (Singh and Schwartzman, 2008). Reciprocally, inhibitors of CYP4A activity and 20-HETE production reduce hypertension induced by androgen treatment (Wu and Schwartzman, 2011). Genetic deletion of CYP4A14 (a murine 20-HETE synthase) causes hypertension more prominently in males, which was attenuated by castration and rescued by androgen administration. CYP4A14<sup>-/-</sup> males also have associated higher serum androgen (testosterone and DHT) levels, suggesting a positive regulatory loop for sex hormones and CYP4A hydroxylase activity (Holla *et al.*, 2001) that may lead to higher BP in males.

### *Eicosanoids and the angiotensin system*

**Angiotensin-(1-7)**, a peptide ligand reportedly active at the **Mas** GPCR, is known for having generally opposing actions to the pro-hypertensive peptide, angiotensin II. Angiotensin-(1-7) acts as a vasodilator but additionally causes release of arachidonic acid and the production of prostanoids, including PGI<sub>2</sub> (Muthalif *et al.*, 1998). Furthermore, s.c. angiotensin-(1-7) infusion attenuates BP increase in both sexes of Dahl salt-sensitive rats following high salt diet and this effect was seen in females for up to 2 weeks (Eatman *et al.*, 2001).

### *Clinical relevance*

**Aspirin**, an irreversible inhibitor of COX-1 and a modulator of COX-2 function, is used as a primary and secondary preventative treatment against CVD. Sex bias in the actions of aspirin is well documented and is covered in a recent review by Pace *et al.* (2017). The first report of gender-divergent responses noted reduced antithrombotic effects in female rabbits compared with males (Kelton *et al.*, 1978), and multiple clinical trials have since noted gender differences in the efficacy of aspirin as a primary prevention for CVD, though men and women reap similar benefits from low-dose aspirin as a secondary prevention for CVD. In a randomized, placebo-controlled primary prevention trial of low-dose aspirin in 39 876 women, no effect was seen in reducing total cardiovascular risk, though there was a significant reduction in overall and ischaemic stroke rates of 26 and 33%, respectively, with no effect on risk of myocardial infarction (Ridker *et al.*, 2005). In the associated meta-analysis of randomized controlled trials, low-dose aspirin caused a 32% reduction in the risk of myocardial infarction for men with no effect on rate of stroke (Ridker *et al.*, 2005). In economic terms, aspirin treatment for the primary prevention of CVD is only cost-

effective in men with a greater than 10% 10 year CVD risk and women with greater than 15% risk, which occurs much later in life (Greving *et al.*, 2008).

In conclusion, this review has discussed three key GPCR-mediated systems that exhibit sexual dimorphism in the CVS and produce meaningful differences in homeostatic function and pathological dysfunction between the sexes. Acute sex-specific effects of GPER on the CVS are likely to be mediated through vascular action of GPER rather than effects on the heart, as males and females have similar cardiac GPER abundance. GPER in the CNS and the kidneys may also play a role in cardioprotection in females (Kurt and Buyukafsar, 2013), though these actions have not been extensively investigated for both sexes at present. Rodent knockout studies show sex-dependent cardiovascular and metabolic phenotypes due to genetic deletion of GPER, though controversy remains and more studies investigating both sexes are needed to confirm acute effects on the vasculature. This positions GPER as a potential therapeutic target, though further insight into the physiological mechanism of GPER is needed, specifically in relation to sex differences.

In the ET system, notable sex differences exist in receptor density and ligand abundance. Men have higher overall ET receptor density and a higher ratio of ET<sub>A</sub> to ET<sub>B</sub> receptors than women, encouraging the vasoconstrictive actions of ET-1. On the other hand, oestrogens suppress the release of ET-1, and females are more resistant to ET-1-mediated rises in BP, but it remains unclear how female sex steroids affect abundance of the generally anti-hypertensive ET<sub>B</sub> receptor, as effects appear to be tissue-specific and species-related.

Eicosanoid receptors, most importantly the TP and IP receptors in female, play a significant role in gender divergence of the CVS. Male sex hormones increase expression of the pro-aggregatory, pro-hypertensive TP receptor. Interestingly, expression of the IP receptor is increased by both male and female sex hormones, potentially having implications for hormonal changes during ageing. Importantly, aspirin, a common preventative therapy for cardiovascular events, is not as effective in women as in men, and an alternative method for reducing incidence of CVD may be necessary for low-risk females.

Considering these functional differences in the CVS between the sexes, it is imperative that animal studies incorporate both sexes in order to delineate sex-dependent mechanisms of cardiovascular homeostasis (Blenck *et al.*, 2016). Looking forward, improving our knowledge of sex hormone contribution to cardiovascular sexual dimorphism will inform recommendations for clinical treatment of CVD between the genders.

### *Nomenclature of targets and ligands*

Key protein targets and ligands in this article are hyperlinked to corresponding entries in <http://www.guidetopharmacology.org>, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Harding *et al.*, 2018), and are permanently archived in the Concise Guide to PHARMACOLOGY 2017/18 (Alexander *et al.*, 2017a,b,c,d).

## Acknowledgements

This work was supported in part by a National Heart Foundation Future Leader Fellowship (N.J.S.), a St Vincent's Clinic Foundation grant-in-aid (N.J.S.) and Froulop Research Grant (N.J.S., J.L.J.C. and M.A.M.) and Australian Government Research Training Scholarships to M.A.M. and J.L.J.C. J.L.J.C. is additionally supported by a Simon and Michal Wilkenfeld Scholarship. The authors thank Bob Graham for critical reading of the manuscript and ongoing guidance.

## Conflict of interest

The authors declare no conflicts of interest.

## References

- Ajayi AAL, Mathur R, Halushka PV (1995). Testosterone increases human platelet thromboxane A<sub>2</sub> receptor density and aggregation responses. *Circulation* 91: 2742–2747.
- Alexander SPH, Cidowski JA, Kelly E, Marrion NV, Peters JA, Faccenda E *et al.* (2017a). The Concise Guide to PHARMACOLOGY 2017/18: Nuclear hormone receptors. *Br J Pharmacol* 174: S208–S224.
- Alexander SPH, Fabbro D, Kelly E, Marrion NV, Peters JA, Faccenda E *et al.* (2017b). The Concise Guide to PHARMACOLOGY 2017/18: Catalytic receptors. *Br J Pharmacol* 174: S225–S271.
- Alexander SPH, Christopoulos A, Davenport AP, Kelly E, Marrion NV, Peters JA *et al.* (2017c). The Concise Guide to PHARMACOLOGY 2017/18: G protein-coupled receptors. *Br J Pharmacol* 174: S17–S129.
- Alexander SPH, Fabbro D, Kelly E, Marrion NV, Peters JA, Faccenda E *et al.* (2017d). The Concise Guide to PHARMACOLOGY 2017/18: Enzymes. *Br J Pharmacol* 174: S272–S359.
- Audoly LP, Tilley SL, Goulet J, Key M, Nguyen M, Stock JL *et al.* (1999). Identification of specific EP receptors responsible for the hemodynamic effects of PGE<sub>2</sub>. *Am J Physiol* 277: H924–H930.
- Bankir L (2001). Antidiuretic action of vasopressin: quantitative aspects and interaction between V1a and V2 receptor-mediated effects. *Cardiovasc Res* 51: 372–390.
- Bartella V, De Francesco EM, Perri MG, Curcio R, Dolce V, Maggolini M *et al.* (2016). The G protein estrogen receptor (GPER) is regulated by endothelin-1 mediated signaling in cancer cells. *Cell Signal* 28: 61–71.
- Behm DJ, Ogbonna A, Wu C, Burns-Kurtis CL, Douglas SA (2009). Epoxyeicosatrienoic acids function as selective, endogenous antagonists of native thromboxane receptors: identification of a novel mechanism of vasodilation. *J Pharmacol Exp Therapeut* 328: 231–239.
- Blenck CL, Harvey PA, Reckelhoff JF, Leinwand LA (2016). The importance of biological sex and estrogen in rodent models of cardiovascular health and disease. *Circ Res* 118: 1294–1312.
- Bologa CG, Revankar CM, Young SM, Edwards BS, Arterburn JB, Kiselyov AS *et al.* (2006). Virtual and biomolecular screening converge on a selective agonist for GPR30. *Nat Chem Biol* 2: 207–212.
- Bossard M, Pumpol K, van der Lely S, Aeschbacher S, Schoen T, Krisai P *et al.* (2015). Plasma endothelin-1 and cardiovascular risk among young and healthy adults. *Atherosclerosis* 239: 186–191.
- Brailoiu E, Dun SL, Brailoiu GC, Mizuo K, Sklar LA, Oprea TI *et al.* (2007). Distribution and characterization of estrogen receptor G protein-coupled receptor 30 in the rat central nervous system. *J Endocrinol* 193: 311–321.
- Breyer MD, Breyer RM (2001). G protein-coupled prostanoid receptors and the kidney. *Annu Rev Physiol* 63: 579–605.
- Broughton BR, Miller AA, Sobey CG (2010). Endothelium-dependent relaxation by G protein-coupled receptor 30 agonists in rat carotid arteries. *Am J Physiol Heart Circ Physiol* 298: H1055–H1061.
- Broughton BRS, Brait VH, Ah Kim H, Lee S, Chu HX, Gardiner-Mann CV *et al.* (2014). Sex-dependent effects of G protein-coupled estrogen receptor activity on outcome after ischemic stroke. *Stroke* 45: 835–841.
- Chakrabarti S, Davidge ST (2012). G-protein coupled receptor 30 (GPR30): a novel regulator of endothelial inflammation. *PLoS one* 7: e52357.
- Chappell MC, Westwood BM, Yamaleyeva LM (2008). Differential effects of sex steroids in young and aged female mRen2.Lewis rats: a model of estrogen and salt-sensitive hypertension. *Genet Med* 5: S65–S75.
- Chen Y, Falck JR, Manthathi VL, Jat JL, Campbell WB (2011). 20-Iodo-14,15-epoxyeicosa-8(Z)-enoyl-3-azidophenylsulfonamide: photoaffinity labeling of a 14,15-epoxyeicosatrienoic acid receptor. *Biochemistry* 50: 3840–3848.
- Cheng SB, Dong J, Pang Y, LaRocca J, Hixon M, Thomas P *et al.* (2014). Anatomical location and redistribution of G protein-coupled estrogen receptor-1 during the estrus cycle in mouse kidney and specific binding to estrogens but not aldosterone. *Mol Cell Endocrinol* 382: 950–959.
- Daka B, Olausson J, Larsson CA, Hellgren MI, Råstam L, Jansson P-A *et al.* (2015). Circulating concentrations of endothelin-1 predict coronary heart disease in women but not in men: a longitudinal observational study in the Vara-Skövde Cohort. *BMC Cardiovasc Disord* 15: 146.
- Davenport AP, Hyndman KA, Dhaun N, Southan C, Kohan DE, Pollock JS *et al.* (2016). Endothelin. *Pharmacol Rev* 68: 357–418.
- David FL, Montezano ACI, Rebouças NA, Nigro D, Fortes ZB, Carvalho MHC *et al.* (2002). Gender differences in vascular expression of endothelin and ETA/ETB receptors, but not in calcium handling mechanisms, in deoxycorticosterone acetate-salt hypertension. *Braz J Med Biol Res* 35: 1061–1068.
- Dennis MK, Burai R, Ramesh C, Petrie WK, Alcon SN, Nayak TK *et al.* (2009). In vivo effects of a GPR30 antagonist. *Nat Chem Biol* 5: 421–427.
- Dennis MK, Field AS, Burai R, Ramesh C, Petrie WK, Bologa CG *et al.* (2011). Identification of a GPER/GPR30 antagonist with improved estrogen receptor counterselectivity. *J Steroid Biochem Mol Biol* 127: 358–366.
- Deschamps AM, Murphy E (2009). Activation of a novel estrogen receptor, GPER, is cardioprotective in male and female rats. *Am J Physiol Heart Circ Physiol* 297: H1806–H1813. <https://doi.org/10.1152/ajpheart.00283.2009>.
- Dubey RK, Jackson EK, Keller PJ, Imthurn B, Rosselli M (2001). Estradiol metabolites inhibit endothelin synthesis by an estrogen receptor-independent mechanism. *Hypertension* 37: 640–644.

- Eatman D, Wang M, Socci RR, Thierry-Palmer M, Emmett N, Bayorh MA (2001). Gender differences in the attenuation of salt-induced hypertension by angiotensin (1–7). *Peptides* 22: 927–933.
- Egan KM, Lawson JA, Fries S, Koller B, Rader DJ, Smyth EM *et al.* (2004). COX-2-derived prostacyclin confers atheroprotection on female mice. *Science* 306: 1954–1957.
- Eivers SB, Kinsella BT (2016). Regulated expression of the prostacyclin receptor (IP) gene by androgens within the vasculature: combined role for androgens and serum cholesterol. *Biochim Biophys Acta* 1859: 1333–1351.
- Ergul A, Shoemaker K, Puett D, Tackett RL (1998). Gender differences in the expression of endothelin receptors in human saphenous veins in vitro. *J Pharmacol Exp Ther* 285: 511–517.
- Fan F, Muroya Y, Roman RJ (2015). Cytochrome P450 eicosanoids in hypertension and renal disease. *Curr Opin Nephrol Hypertens* 24: 37–46.
- Feldman RD, Gros R, Ding Q, Hussain Y, Ban MR, McIntyre AD *et al.* (2014). A common hypofunctional genetic variant of GPER is associated with increased blood pressure in women. *Br J Clin Pharmacol* 78: 1441–1452.
- Filardo EJ, Quinn JA, Bland KI, Frackelton AR Jr (2000). Estrogen-induced activation of Erk-1 and Erk-2 requires the G protein-coupled receptor homolog, GPR30, and occurs via trans-activation of the epidermal growth factor receptor through release of HB-EGF. *Mol Endocrinol* (Baltimore, Md) 14: 1649–1660.
- Filardo EJ, Quinn JA, Frackelton AR Jr, Bland KI (2002). Estrogen action via the G protein-coupled receptor, GPR30: stimulation of adenylyl cyclase and cAMP-mediated attenuation of the epidermal growth factor receptor-to-MAPK signaling axis. *Mol Endocrinol* (Baltimore, Md) 16: 70–84.
- Francois H, Facemire C, Kumar A, Audoly L, Koller B, Coffman T (2007). Role of microsomal prostaglandin synthase 1 in the kidney. *J Am Soc Nephrol* 18: 1466–1475.
- Fukuroda T, Fujikawa T, Ozaki S, Ishikawa K, Yano M, Nishikibe M (1994). Clearance of circulating endothelin-1 by ETB receptors in rats. *Biochem Biophys Res Commun* 199: 1461–1465.
- Garcia V, Gilani A, Shkolnik B, Pandey V, Zhang FF, Dakarapu R *et al.* (2017). 20-HETE signals through G protein-coupled receptor GPR75 (Gq) to affect vascular function and trigger hypertension. *Circ Res* 120: 1776–1788.
- Gillis EE, Sasser JM, Sullivan JC (2016). Endothelin, sex, and pregnancy: unique considerations for blood pressure control in females. *Am J Physiol Regul Integr Comp Physiol* 310: R691–R696.
- Gohar EY, Yusuf C, Pollock DM (2016). Ovarian hormones modulate endothelin A and B receptor expression. *Life Sci* 159: 148–152.
- Greving JP, Buskens E, Koffijberg H, Algra A (2008). Cost-effectiveness of aspirin treatment in the primary prevention of cardiovascular disease events in subgroups based on age, gender, and varying cardiovascular risk. *Circulation* 117: 2875–2883.
- Gros R, Ding Q, Liu B, Chorazyczewski J, Feldman RD (2013). Aldosterone mediates its rapid effects in vascular endothelial cells through GPER activation. *Am J Physiol Cell Physiol* 304: C532–C540.
- Gros R, Ding Q, Sklar LA, Prossnitz EE, Arterburn JB, Chorazyczewski J *et al.* (2011). GPR30 expression is required for the mineralocorticoid receptor-independent rapid vascular effects of aldosterone. *Hypertension* 57: 442–451.
- Haas E, Bhattacharya I, Brailoiu E, Damjanovic M, Brailoiu GC, Gao X *et al.* (2009). Regulatory role of G protein-coupled estrogen receptor for vascular function and obesity. *Circ Res* 104: 288–291.
- Hallberg P, Karlsson J, Lind L, Michaëlsson K, Kurland L, Kahan T *et al.* (2004). Gender-specific association between preproendothelin-1 genotype and reduction of systolic blood pressure during antihypertensive treatment – results from the Swedish irbesartan left ventricular hypertrophy investigation versus atenolol (SILVHIA). *Clin Cardiol* 27: 287–290.
- Han S-Z, Karaki H, Ouchi Y, Akishita M, Orimo H (1995). 17 $\beta$ -Estradiol inhibits Ca<sup>2+</sup> influx and Ca<sup>2+</sup> release induced by thromboxane A2 in porcine coronary artery. *Circulation* 91: 2619–2626.
- Harding SD, Sharman JL, Faccenda E, Southan C, Pawson AJ, Ireland S *et al.* (2018). The IUPHAR/BPS Guide to PHARMACOLOGY in 2018: updates and expansion to encompass the New Guide to IMMUNOPHARMACOLOGY. *Nucl Acids Res* 46: D1091–D1106.
- Higashiura K, Mathur RS, Halushka PV (1997). Gender-related differences in androgen regulation of thromboxane A2 receptors in rat aortic smooth-muscle cells. *J Cardiovasc Pharmacol* 29: 311–315.
- Hinojosa-Laborde C, Lange DL, Haywood JR (2000). Role of female sex hormones in the development and reversal of Dahl hypertension. *Hypertension* 35: 484–489.
- Holla VR, Adas F, Imig JD, Zhao X, Price E, Olsen N *et al.* (2001). Alterations in the regulation of androgen-sensitive Cyp 4a monooxygenases cause hypertension. *Proc Natl Acad Sci* 98: 5211–5216.
- Hulley S, Grady D, Bush T, Furberg C, Herrington D, Riggs B *et al.* (1998). Randomized trial of estrogen plus progestin for secondary prevention of coronary heart disease in postmenopausal women. Heart and Estrogen/progestin Replacement Study (HERS) Research Group. *JAMA* 280: 605–613.
- Ignatov A, Robert J, Gregory-Evans C, Schaller HC (2006). RANTES stimulates Ca<sup>2+</sup> mobilization and inositol trisphosphate (IP3) formation in cells transfected with G protein-coupled receptor 75. *Br J Pharmacol* 149: 490–497.
- Isensee J, Meoli L, Zazzu V, Nabzdyk C, Witt H, Soewarto D *et al.* (2008). Expression pattern of G protein-coupled receptor 30 in LacZ reporter mice. *Endocrinology* 150: 1722–1730.
- Jin C, Speed JS, Hyndman KA, O'Connor PM, Pollock DM (2013). Sex differences in ET-1 receptor expression and Ca<sup>2+</sup> signaling in the IMCD. *Am J Physiol Renal Physiol* 305: F1099–F1104.
- Karanian J, Moran F, Ramey E, Ramwell P (1981). Gender differences in prostaglandin receptors of rat aorta. *Br J Pharmacol* 72: 10–12.
- Kellogg DL, Liu Y, Pérgola PE (2001). Selected contribution: gender differences in the endothelin-B receptor contribution to basal cutaneous vascular tone in humans. *J Appl Physiol* 91: 2407–2411.
- Kelton J, Hirsh J, Carter C, Buchanan M (1978). Sex differences in the antithrombotic effects of aspirin. *Blood* 52: 1073–1076.
- Kinsella BT, O'Mahony DJ, Fitzgerald GA (1997). The human thromboxane A2 receptor alpha isoform (TP alpha) functionally couples to the G proteins Gq and G11 in vivo and is activated by the isopropane 8-epi prostaglandin F2 alpha. *J Pharmacol Exp Ther* 281: 957–964.
- Kittikulsuth W, Pollock JS, Pollock DM (2011). Sex differences in renal medullary endothelin receptor function in angiotensin II hypertensive rats. *Hypertension* 58: 212–218.
- Kittikulsuth W, Sullivan JC, Pollock DM (2013). ET-1 actions in the kidney: evidence for sex differences. *Br J Pharmacol* 168: 318–326.
- Koganti S, Snyder R, Gumaste U, Karamyan VT, Thekkumkara T (2014). 2-Methoxyestradiol binding of GPR30 down-regulates angiotensin AT<sub>1</sub> receptor. *Eur J Pharmacol* 723: 131–140.

- Kohan DE, Inscho EW, Wesson D, Pollock DM (2011). Physiology of endothelin and the kidney. *Compr Physiol*.
- Kurt AH, Buyukafsar K (2013). Vasoconstriction induced by G1, a G-protein-coupled oestrogen receptor1 (GPER-1) agonist, in the isolated perfused rat kidney. *Eur J Pharmacol* 702: 71–78.
- Lafferty AR, Torpy DJ, Stowasser M, Taymans SE, Lin JP, Huggard P *et al.* (2000). A novel genetic locus for low renin hypertension: familial hyperaldosteronism type II maps to chromosome 7 (7p22). *J Med Genet* 37: 831–835.
- Lindsey SH, Carver KA, Prossnitz ER, Chappell MC (2011a). Vasodilation in response to the GPR30 agonist G-1 is not different from estradiol in the mRen2.Lewis female rat. *J Cardiovasc Pharmacol* 57: 598–603.
- Lindsey SH, Cohen JA, Brosnihan KB, Gallagher PE, Chappell MC (2009). Chronic treatment with the G protein-coupled receptor 30 agonist G-1 decreases blood pressure in ovariectomized mRen2.Lewis rats. *Endocrinology* 150: 3753–3758.
- Lindsey SH, Liu L, Chappell MC (2014). Vasodilation by GPER in mesenteric arteries involves both endothelial nitric oxide and smooth muscle cAMP signaling. *Steroids* 81: 99–102.
- Lindsey SH, Yamaleyeva LM, Brosnihan KB, Gallagher PE, Chappell MC (2011b). Estrogen receptor GPR30 reduces oxidative stress and proteinuria in the salt-sensitive female mRen2.Lewis rat. *Hypertension* 58: 665–671.
- Liu PY, Death AK, Handelsman DJ (2003). Androgens and cardiovascular disease. *Endocr Rev* 24: 313–340.
- Luzier AB, Nawarskas JJ, Añonuevo J, Wilson MF, Kazierad DJ (1998). The effects of gender on adrenergic receptor responsiveness. *J Clin Pharmacol* 38: 618–624.
- Martensson UE, Salehi SA, Windahl S, Gomez MF, Sward K, Daszkiewicz-Nilsson J *et al.* (2009). Deletion of the G protein-coupled receptor 30 impairs glucose tolerance, reduces bone growth, increases blood pressure, and eliminates estradiol-stimulated insulin release in female mice. *Endocrinology* 150: 687–698.
- Masuda A, Mathur R, Halushka PV (1991). Testosterone increases thromboxane A2 receptors in cultured rat aortic smooth muscle cells. *Circ Res* 69: 638–643.
- Meyer MR, Amann K, Field AS, Hu C, Hathaway HJ, Kanagy NL *et al.* (2012a). Deletion of G protein-coupled estrogen receptor increases endothelial vasoconstriction. *Hypertension* 59: 507–512.
- Meyer MR, Baretella O, Prossnitz ER, Barton M (2010). Dilation of epicardial coronary arteries by the G protein-coupled estrogen receptor agonists G-1 and ICI 182,780. *Pharmacology* 86: 58–64.
- Meyer MR, Field AS, Kanagy NL, Barton M, Prossnitz ER (2012b). GPER regulates endothelin-dependent vascular tone and intracellular calcium. *Life Sci* 91: 623–627.
- Meyer MR, Fredette NC, Barton M, Prossnitz ER (2015). G protein-coupled estrogen receptor inhibits vascular prostanoid production and activity. *J Endocrinol* 227: 61–69.
- Meyer MR, Fredette NC, Sharma G, Barton M, Prossnitz ER (2016). GPER is required for the age-dependent upregulation of the myocardial endothelin system. *Life Sci* 159: 61–65.
- Miggin SM, Kinsella BT (2002). Investigation of the mechanisms of G protein: effector coupling by the human and mouse prostacyclin receptors. Identification of critical species-dependent differences. *J Biol Chem* 277: 27053–27064.
- Mikkola T, Turunen P, Avela K, Orpana A, Viinikka L, Ylikorkala O (1995). 17 beta-Estradiol stimulates prostacyclin, but not endothelin-1, production in human vascular endothelial cells. *J Clin Endocrinol Metab* 80: 1832–1836.
- Murata T, Ushikubi F, Matsuoka T, Hirata M (1997). Altered pain perception and inflammatory response in mice lacking prostacyclin receptor. *Nature* 388: 678–682
- Muthalif MM, Benter IF, Uddin MR, Harper JL, Malik KU (1998). Signal transduction mechanisms involved in angiotensin-(1–7)-stimulated arachidonic acid release and prostanoid synthesis in rabbit aortic smooth muscle cells. *J Pharmacol Exp Ther* 284: 388–398.
- Nakano D, Pollock JS, Pollock DM (2008). Renal medullary ETB receptors produce diuresis and natriuresis via NOS1. *Am J Physiol Renal Physiol* 294: F1205–F1211.
- Nelson LR, Bulun SE (2001). Estrogen production and action. *J Am Acad Dermatol* 45: S116–S124.
- Node K, Ruan X-L, Dai J, Yang S-X, Graham L, Zeldin DC *et al.* (2001). Activation of G $\alpha_s$  mediates induction of tissue-type plasminogen activator gene transcription by epoxyeicosatrienoic acids. *J Biol Chem* 276: 15983–15989.
- Nuedling S, van Eickels M, Allera A, Doevendans P, Meyer R, Vetter H *et al.* (2003). 17 beta-Estradiol regulates the expression of endothelin receptor type B in the heart. *Br J Pharmacol* 140: 195–201.
- Ospina JA, Krause DN, Duckles SP (2002). 17 $\beta$ -Estradiol increases rat cerebrovascular prostacyclin synthesis by elevating cyclooxygenase-1 and prostacyclin synthase. *Stroke* 33: 600–605.
- Otto C, Rohde-Schulz B, Schwarz G, Fuchs I, Klewer M, Brittain D *et al.* (2008). G protein-coupled receptor 30 localizes to the endoplasmic reticulum and is not activated by estradiol. *Endocrinology* 149: 4846–4856.
- Pace S, Sautebin L, Werz O (2017). Sex-biased eicosanoid biology: impact for sex differences in inflammation and consequences for pharmacotherapy. *Biochem Pharmacol* 145: 1–11.
- Pardue M-L, Wizemann TM (2001). *Exploring the Biological Contributions to Human Health: Does Sex Matter?* National Academies Press: Washington, DC.
- Pedersen SH, Nielsen LB, Mortensen A, Nilas L, Ottesen B (2008). Progesterins oppose the effects of estradiol on the endothelin-1 receptor type B in coronary arteries from ovariectomized hyperlipidemic rabbits. *Menopause* 15: 503–510.
- Pedersen SH, Nielsen LB, Pedersen NG, Nilas L, Ottesen B (2009). Hormone therapy modulates ETA mRNA expression in the aorta of ovariectomized New Zealand white rabbits. *Gynecol Endocrinol* 25: 175–182.
- Pilote L, Dasgupta K, Guru V, Humphries KH, McGrath J, Norris C *et al.* (2007). A comprehensive view of sex-specific issues related to cardiovascular disease. *CMAJ: Canadian Medical Association journal = journal de l'Association medicale canadienne* 176: S1–S44.
- Pingili AK, Davidge KN, Thirunavukkarasu S, Khan NS, Katsurada A, Majid DSA *et al.* (2017). 2-Methoxyestradiol reduces angiotensin II-induced hypertension and renal dysfunction in ovariectomized female and intact male mice. *Hypertension* 69: 1104–1112.
- Prossnitz ER, Arterburn JB (2015). International Union of Basic and Clinical Pharmacology. XCIV. G protein-coupled estrogen receptor and its pharmacologic modulators. *Pharmacol Rev* 67: 505–540.
- Prossnitz ER, Barton M (2009). Signaling, physiological functions and clinical relevance of the G protein-coupled estrogen receptor GPER. *Prostaglandins Other Lipid Mediat* 89: 89–97.

- Purdy KE, Arendshorst WJ (2000). EP<sub>1</sub> and EP<sub>4</sub> receptors mediate prostaglandin E<sub>2</sub> actions in the microcirculation of rat kidney. *Am J Physiol Renal Physiol* 279: F755–F764.
- Randriamboavonjy V, Kiss L, Falck JR, Busse R, Fleming I (2005). The synthesis of 20-HETE in small porcine coronary arteries antagonizes EDHF-mediated relaxation. *Cardiovasc Res* 65: 487–494.
- Rask-Andersen M, Almen MS, Schioth HB (2011). Trends in the exploitation of novel drug targets. *Nat Rev Drug Discov* 10: 579–590.
- Reckelhoff JF, Zhang H, Srivastava K (2000). Gender differences in development of hypertension in spontaneously hypertensive rats: role of the renin–angiotensin system. *Hypertension* 35: 480–483.
- Revankar CM, Cimino DF, Sklar LA, Arterburn JB, Prossnitz ER (2005). A transmembrane intracellular estrogen receptor mediates rapid cell signaling. *Science* 307: 1625–1630.
- Ridker PM, Cook NR, Lee I-M, Gordon D, Gaziano JM, Manson JE *et al.* (2005). A randomized trial of low-dose aspirin in the primary prevention of cardiovascular disease in women. *N Engl J Med* 352: 1293–1304.
- Rompe F, Artuc M, Hallberg A, Alterman M, Ströder K, Thöne-Reineke C *et al.* (2010). Direct angiotensin II type 2 receptor stimulation acts anti-inflammatory through epoxyeicosatrienoic acid and inhibition of nuclear factor  $\kappa$ B. *Hypertension* 55: 924–931.
- Rothman MS, Carlson NE, Xu M, Wang C, Swerdloff R, Lee P *et al.* (2011). Reexamination of testosterone, dihydrotestosterone, estradiol and estrone levels across the menstrual cycle and in postmenopausal women measured by liquid chromatography-tandem mass spectrometry. *Steroids* 76: 177–182.
- Samuel CS, Royce SG, Hewitson TD, Denton KM, Cooney TE, Bennett RG (2017). Anti-fibrotic actions of relaxin. *Br J Pharmacol* 174: 962–976.
- Schirmer M, Taube C (1993). U 46619 induces different blood pressure effects in male and female spontaneously hypertensive rats (SHR). *Prostaglandins Leukot Essent Fatty Acids* 48: 469–473.
- Schneider MP, Boesen EI, Pollock DM (2007). Contrasting actions of endothelin ET<sub>A</sub> and ET<sub>B</sub> receptors in cardiovascular disease. *Annu Rev Pharmacol Toxicol* 47: 731–759.
- Seeland U, Regitz-Zagrosek V (2012). Sex and gender differences in cardiovascular drug therapy. In: Regitz-Zagrosek V (ed). *Sex and Gender Differences in Pharmacology*. Springer Berlin Heidelberg: Berlin, Heidelberg, pp. 211–236.
- Singh H, Schwartzman ML (2008). Renal vascular cytochrome P450-derived eicosanoids in androgen-induced hypertension. *Pharmacol Rep* 60: 29–37.
- Southern C, Cook JM, Neetoo-Isseljee Z, Taylor DL, Kettleborough CA, Merritt A *et al.* (2013). Screening beta-arrestin recruitment for the identification of natural ligands for orphan G-protein-coupled receptors. *J Biomol Screen* 18: 599–609.
- Speed JS, D'Angelo G, Wach PA, Sullivan JC, Pollock JS, Pollock DM (2015). High salt diet increases the pressor response to stress in female, but not male ETB-receptor-deficient rats. *Physiol Rep* 3: e12326.
- Stauffer BL, Westby CM, Greiner JJ, Van Guilder GP, DeSouza CA (2010). Sex differences in endothelin-1-mediated vasoconstrictor tone in middle-aged and older adults. *Am J Physiol Regul Integr Comp Physiol* 298: R261–R265.
- Stock JL, Shinjo K, Burkhardt J, Roach M, Taniguchi K, Ishikawa *Tet al.* (2001). The prostaglandin E<sub>2</sub> EP<sub>1</sub> receptor mediates pain perception and regulates blood pressure. *J Clin Invest* 107: 325–331.
- Sullivan JC (2008). Sex and the renin–angiotensin system: inequality between the sexes in response to RAS stimulation and inhibition. *Am J Physiol Regul Integr Comp Physiol* 294: R1220–R1226.
- Sullivan JC, Pollock JS, Pollock DM (2006). Superoxide-dependent hypertension in male and female endothelin B receptor-deficient rats. *Exp Biol Med (Maywood, NJ)* 231: 818–823.
- Tan Z, Wang T-H, Yang D, Fu X-D, Pan J-Y (2003). Mechanisms of 17 $\beta$ -estradiol on the production of ET-1 in ovariectomized rats. *Life Sci* 73: 2665–2674.
- Taylor TA, Garipey CE, Pollock DM, Pollock JS (2003). Gender differences in ET and NOS systems in ETB receptor-deficient rats: effect of a high salt diet. *Hypertension* 41: 657–662.
- Thomas P, Pang Y, Filardo EJ, Dong J (2005). Identity of an estrogen membrane receptor coupled to a G protein in human breast cancer cells. *Endocrinology* 146: 624–632.
- Toth P, Rozsa B, Springo Z, Doczi T, Koller A (2011). Isolated human and rat cerebral arteries constrict to increases in flow: role of 20-HETE and TP receptors. *J Cereb Blood Flow Metab* 31: 2096–2105.
- Uhlen M, Fagerberg L, Hallstrom BM, Lindskog C, Oksvold P, Mardinoglu A *et al.* (2015). Proteomics. Tissue-based map of the human proteome. *Science* 347: 1260419.
- Wang H, Jessup JA, Lin MS, Chagas C, Lindsey SH, Groban L (2012). Activation of GPR30 attenuates diastolic dysfunction and left ventricle remodeling in oophorectomized mRen2.Lewis rats. *Cardiovasc Res* 94: 96–104.
- Wang H, Sun X, Chou J, Lin M, Ferrario CM, Zapata-Sudo G *et al.* (2016). Cardiomyocyte-specific deletion of the G protein-coupled estrogen receptor (GPER) leads to left ventricular dysfunction and adverse remodeling: a sex-specific gene profiling analysis. *Biochim Biophys Acta* 1863: 1870–1882.
- WHO (2014). *Global Health Estimates: Deaths by Cause, Age, Sex and Country, 2000–2012*. World Health Organisation: Geneva.
- Wu CC, Schwartzman ML (2011). The role of 20-HETE in androgen-mediated hypertension. *Prostaglandins Other Lipid Mediat* 96: 45–53.
- Xue B, Johnson AK, Hay M (2013). Sex differences in angiotensin II- and aldosterone-induced hypertension: the central protective effects of estrogen. *Am J Physiol Regul Integr Comp Physiol* 305: R459–R463.
- Yanagisawa M, Kurihara H, Kimura S, Tomobe Y, Kobayashi M, Mitsui Y *et al.* (1988). A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature* 332: 411–415.