ARTICLE



Expanding the phenotypic spectrum of variants in *PDE4D/PRKAR1A*: from acrodysostosis to acroscyphodysplasia

Caroline Michot¹ · Carine Le Goff¹ · Edward Blair² · Patricia Blanchet³ · Yline Capri⁴ · Brigitte Gilbert-Dussardier ⁵ · Alice Goldenberg⁶ · Alex Henderson⁷ · Bertrand Isidor⁸ · Hulya Kayserili⁹ · Esther Kinning¹⁰ · Martine Le Merrer¹ · Stanislas Lyonnet¹ · Sylvie Odent¹¹ · Pelin Ozlem Simsek-Kiper¹² · Chloé Quelin¹¹ · Ravi Savarirayan¹³ · Marleen Simon¹⁴ · Miranda Splitt⁵ · Judith M.A. Verhagen ¹⁵ · Alain Verloes ³ · Arnold Munnich¹ · Geneviève Baujat¹ · Valérie Cormier-Daire¹

Received: 7 October 2017 / Revised: 11 February 2018 / Accepted: 23 February 2018 / Published online: 13 July 2018 © European Society of Human Genetics 2018

Abstract

Acrodysostosis (MIM 101800) is a dominantly inherited condition associating (1) skeletal features (short stature, facial dysostosis, and brachydactyly with cone-shaped epiphyses), (2) resistance to hormones and (3) possible intellectual disability. Acroscyphodysplasia (MIM 250215) is characterized by growth retardation, brachydactyly, and knee epiphyses embedded in cup-shaped metaphyses. We and others have identified PDE4D or PRKAR1A variants in acrodysostosis; PDE4D variants have been reported in three cases of acroscyphodysplasia. Our study aimed at reviewing the clinical and molecular findings in a cohort of 27 acrodysostosis and 5 acroscyphodysplasia cases. Among the acrodysostosis cases, we identified 9 heterozygous de novo PRKAR1A variants and 11 heterozygous PDE4D variants. The 7 patients without variants presented with symptoms of acrodysostosis (brachydactyly and cone-shaped epiphyses), but none had the characteristic facial dysostosis. In the acroscyphodysplasia cases, we identified 2 PDE4D variants. For 2 of the 3 negative cases, medical records revealed early severe infection, which has been described in some reports of acroscyphodysplasia. Subdividing our series of acrodysostosis based on the disease-causing gene, we confirmed genotype-phenotype correlations. Hormone resistance was consistently observed in patients carrying *PRKAR1A* variants, whereas no hormone resistance was observed in 9 patients with *PDE4D* variants. All patients with PDE4D variants shared characteristic facial features (midface hypoplasia with nasal hypoplasia) and some degree of intellectual disability. Our findings of PDE4D variants in two cases of acroscyphodysplasia support that PDE4D may be responsible for this severe skeletal dysplasia. We eventually emphasize the importance of some specific assessments in the long-term follow up, including cardiovascular and thromboembolic risk factors.

Introduction

Acrodysostosis is a rare autosomal dominant condition first described by Pierre Maroteaux et al. in 1968 [1] and further reviewed by Robinow et al. in 1971 [2]. It consists in the association of (1) skeletal features characterized by short stature, facial dysostosis with nasal hypoplasia and peripheral dysostosis with severe brachymetatarsia,

Valérie Cormier-Daire valerie.cormier-daire@inserm.fr brachymetacarpia, brachydactyly, cone-shaped epiphyses, and advanced bone maturation, (2) inconstant resistance to multiple hormones including parathyroid hormone or thyrotropin, and (3) possible neurological involvement with moderate to mild intellectual disability [1, 2]. Differential diagnoses include Albright hereditary osteodystrophy (AHO) [MIM103580] and pseudopseudohypoparathyroid-ism [MIM 612463] due to loss of function variants in *GNAS* (α -stimulary subunit of the G-protein) and characterized by less severe skeletal involvement [3].

In 2011, the recurrent p.Arg368* variant in the *PRKAR1A* gene has been found in three individuals with acrodysostosis and resistance to multiple hormones [4]. *PRKAR1A* encodes the cyclic AMP (cAMP)-dependent regulatory subunit of protein kinase A. The mutated subunit impairs the protein kinase A response to cAMP [4],

Electronic supplementary material The online version of this article (https://doi.org/10.1038/s41431-018-0135-1) contains supplementary material, which is available to authorised users.

Extended author information available on the last page of the article

accounting for hormone resistance and skeletal abnormalities resembling those observed in AHO. We then identified *PDE4D* variants as another cause of acrodysostosis, most frequently without hormone resistance [5]. *PDE4D* is also involved in cAMP signaling pathway metabolism, encoding a class IV cAMP-specific phosphodiesterase, regulating cAMP concentration.

On the basis of these molecular basis, two distinct MIM references have been generated, namely ACRO1 (MIM 101800) for *PRKAR1A* variants and ACRO2 (MIM614613) for *PDE4D* variants. We previously reported some genotype–phenotype correlations. Indeed, patients with *PRKAR1A* variants tend to have less characteristic facial dysostosis, normal intelligence, and resistance to multiple hormones compared to patients with *PDE4D* variants who presented with characteristic facial dysostosis, intellectual disability, and subtle or absence of hormonal resistance [6, 7].

In recent studies, several acroscyphodysplasia cases [MIM 250215] [7, 8] were reported with a *PDE4D* variant. This entity consists of the association of severe growth retardation, micromelia predominating in the lower limbs, knee flexion, and severe brachydactyly altogether with a specific radiological appearance of the knees: cup-shaped metaphyses with embedded epiphyses [9]. This radiological aspect is reminiscent of the appearance of the cone-shaped epiphyses of small joints in acrodysostosis.

Studying 32 unrelated cases of acrodysostosis (n = 27) or acroscyphodysplasia (n = 5), we found *PDE4D* variants in 13 cases, including 2 cases of acroscyphodysplasia, and *PRKRA1A* variants in 9 acrodysostosis cases. We confirmed some genotype–phenotype correlations and involvement of *PDE4D* in acroscyphodysplasia. We also emphasize the importance of some specific assessments in the long-term follow up of these conditions.

Subjects and methods

Clinical ascertainment

Overall, 32 patients were recruited for this study, including 27 cases of acrodysostosis (comprising 10 patients previously described in Michot et al. [5]) and 5 cases of acroscyphodysplasia. Two patients had an affected parent. Inclusion criteria for acrodysostosis cases were a peripheral dysostosis with severe generalized brachydactyly, affecting metacarpals and phalanges, associated with cone-shaped epiphyses. Short stature, facial dysostosis, resistance to multiple hormones, and intellectual disability were not considered as mandatory. The acroscyphodysplasia cases were recruited on the sole criterion of a characteristic radiological aspect of the knees.

Among the 32 recruited patients, 18 were females and 14 were males. Thirty patients were sporadic cases, and in 2

cases, acrodysostosis was inherited from an affected mother.

Informed consent for participation, sample collection, and photograph publication were obtained using protocols approved by the Necker Hospital ethics board committee.

Clinical data collection

According to a literature review, a questionnaire with selected medical items was sent out to the referring physician, to collect details on the clinical and biological symptoms of the cases diagnosed with acrodysostosis and acroscyphodysplasia. Available data, photographs, and radiographs (when authorized by the patients or their legal representatives) were collected.

Hormonal screening

For complete screening of mineral metabolism, blood levels of creatinine, calcium, phosphorus, thyroxin (T4), thyrotropin, 25-hydroxyvitamin D, 1-25-dihydroxyvitamin D, parathyroid hormone (PTH), and fibroblast growth factor 23 (FGF23) levels were measured, along with urinary creatinine, calcium, and phosphorus excretion.

PRKAR1A and PDE4D molecular screening

Genomic DNA was obtained from peripheral blood leukocytes using standard procedures. The exons and exon-intron boundaries of *PRKAR1A* (GenBank NM_ 002734.4) and *PDE4D* (GenBank NM_001104631.1) were amplified using specific primers (available upon request). Amplification products were purified by ExoSapIT (Amersham, Buckinghamshire, UK) and directly sequenced using the Big Dye Terminator Cycle Sequencing Ready Reaction kit v.1.1 on an automatic sequencer (ABI3130x1; PE Applied Biosystems, Foster City, CA). Sequence analyses were performed using Seqscape software v2.5 (Applied Biosystems).

Results

Sequencing data

De novo heterozygous *PRKAR1A* variants were identified in 9 cases, including the p.Arg368* stop variant [4] in 5/9 and missense variants (c.786G>C (p.(Trp262Cys)), c.968A>C (p.(Tyr323Ser)), c.968A>G (p.(Tyr323Cys)), c.1117T>C (p.(Tyr373His))) in 4/9. All these variants were predicted as probably damaging using PolyPhen2 software and were not identified in 200 control chromosomes, nor indexed in gnomAD database. They altered a conserved amino acid,

located either in the catalytic domain or in the cAMPbinding domain B (Fig. 1).

PDE4D variants were identified in 11 acrodysostosis cases, including the two familial cases. They were all missense variants spread throughout the whole *PDE4D* coding sequence (Fig. 2), altering a conserved amino acid located in the catalytic domain and were predicted as probably damaging using PolyPhen2 software, except for the c.568T>G (p.(Ser190Ala)) and c.568T>C (p.(Ser190Pro)) predicted as benign. These last 2 variants occurred de novo and affected a serine residue predicted to be phosphorylated (UniProt database) and were absent from alleles in 200 ethnicity-matched controls and from gnomAD database.

De novo *PDE4D* variants (c.995T>C (p.(Phe332Ser)) and c.99T>G (p.(Ile333Met))) were identified in 2/5 patients with acroscyphodysplasia.

Clinical, biological, and radiological data of acrodysostosis cases

The clinical and available biological details are summarized in Table 1 for patients with identified variants.

Intrauterine growth retardation (IUGR) was observed in 8/9 with PRKAR1A variants, but only in a few patients with *PDE4D* variants (n = 5/11), but no available data in three cases). Postnatally, 8/9 patients with PRKAR1A variants developed growth deficiency, ranging from -2 SD to -6.7SD, whereas only 4/11 individuals with PDE4D variants had postnatal short stature. However, patients with PDE4D variants reported herein were young at time of the study (6 were below 10 years of age). Data on pubertal development were not available for these young patients, but one boy had cryptorchidism treated by testosterone and one girl a female hypospadias. One boy with PRKAR1A variant (out of 3) also had cryptorchidism, which required surgery. In four adult women with PRKAR1A variants, one experienced a miscarriage and an intrauterine fetal death, whereas another one had an absence of the mammary glands development.

Brachydactyly involved all rays of fingers and toes, associated with cone-shaped epiphyses. Three patients also had single palmar crease, including two with *PRKAR1A* variants and one without any identified variant. A relative hyperplasia of the first ray of the feet was historically described and was indeed observed in 6/9 patients with *PRKAR1A* variants and 2/11 with *PDE4D* variants. One patient with *PDE4D* variant (patient 20) developed a macrodactyly of second and third rays of one foot that required amputation. Elder patients with *PRKAR1A* variants developed stiffness of the elbows and one underwent corrective surgery for carpal tunnel syndrome.

A flat face with malar hypoplasia was noted in all patients (Fig. 3). Nevertheless the distinctive nasal hypoplasia was described in only 3/9 patients with *PRKAR1A* variants,

p.Arg335Leu p.Arg335Cys p.Arg335Pro p.Ala328Va p.Tyr373Cys p.lle327Thr p.Gln372* p.Gly289Gl p.Gln285Arg p.Tyr175Cys Arg368* p.Thr239Ala p.Alg213Thr 94100 143 AMP hind AMP bin dina 381 AA DD domain A omain B p.Tyr373His p.Trp262Cys p.Tyr323Cys p.Tvr323Sei

Fig. 1 Scheme of the *PRKAR1A* gene, modified from Linglart el al. [7]. Above are listed the previously published variants, including the recurrent p.Arg368* variant, which was found in four cases in this study. Below are the four new and unique variants described in this study



Fig. 2 Scheme of the *PDE4D* gene. Above are listed the previously published variants. Below are the eleven new and unique variants described in this study

whereas 11/11 patients with *PDE4D* variants had this typical facial feature. Noteworthy, the three patients with *PRKAR1A* variants and nasal hypoplasia harbored missense variants affecting the Tyrosine 323 (c.786G>C, c.968A>C and c.968A>G) and the tryptophane 262 residues, and not the recurrent stop variant affecting arginine 368 residue. Moreover, 3/9 patients with *PRKAR1A* variants and 5/11 with *PDE4D* had a prominent mandible. Two patients with *PRKAR1A* variants presented with delayed eruption of teeth and 1 with *PDE4D* variant had abnormal enamel.

Eight out of nine with *PRKAR1A* variant had normal intellectual development. Only patient 8 (who harbored a *PRKAR1A* variant affecting the tyrosine 323 residue (c.968A>C)) had a moderate intellectual disability with congenital axial hypotonia and triventricular cerebral dilatation. Noteworthy, the second patient with a variant affecting this residue, patient 9 (c.968A>G), was only 18 months old at the time of completion and besides he

Table 1 Clinical and biological data of the 20 acrodysostosis patients with identified variants described in this study

	PRKAR1A (n=9)	PDE4D (<i>n</i> =11)	Total
Paternal age at birth	34 to 40 y	19 to 50 y	mean : 35,2
Ethnicity	Caucasian, Asian, Maghreb	Caucasian, Turkish, South Arabia	-
Sex	6 F/3 M	4 F/7 M	10 F/10 M
Age	5,5 to 38 y	4,5 to 33 y	4,5 to 37 y
IUGR	8 (89%)	5 (45%)	13+ (65%)
Postnatal growth retardation	8 (89%)	4 (36%)	12+ (60%)
Actual height (SD)	-6,7 to -0,9 SD	-3 to +1,8 SD	-6,7 to +1,8 SD
Obesity (BMI)	3 (33%)	2 (18%)	5+ (25%)
Facial dysostosis			
Nasal hypoplasia	3 (33%)	10 (90%)	13+ (65%)
Depressed nasal bridge	8 (89%)	11 (100%)	19+ (95%)
Prominent mandibule	3 (33%)	5 (45%)	8+ (40%)
Peripheral dysostosis			
Severe brachydactyly	9 (100%)	11 (100%)	20+ (100%)
Short, broad metatarsals, metacarpals and phalanges	9 (100%)	10 (90%)	19+ (95%)
Cone-shaped epiphyses	8 (89%)	8 (72%)	16+ (80%)
Advanced bone age	4 (44%)	6 (54%)	10+ (50%)
Narrowing of caudal vertebral interpedicular distance	6 (66%)	2 (18%)	8+ (40%)
Hormonal screening			
Parathyroid	8 resistence to PTH (89%)	1 slight increased PTH (9%)	9 elevated (45%)
GH axis	2 abnormal (22%)	0	2 abnormal (10%)
External genitalia	1 cryptorchidism (33% M)	1 cryptorchidism (14% M); 1 feminine hypospadias	3 abnormal (15%)
Puberty	2 abnormal in female	0	2 abnormal (10%)
Thyroid	7 hypothyroidism (78%)	0	7 hypothyroidism (35%)
Mental retardation	1 (11%)	10 (90%)	11+ (55%)

See supplementary material for the full version of Table 1

had walked normally at 15 months of age, congenital hypotonia and poor sucking had been noticed before. Conversely almost all patients (10/11) with *PDE4D* variants had mild to moderate intellectual disability. Moreover, two patients with a *PDE4D* variant developed acute intracranial hypertension due to a thrombophlebitis (patients 11 and 12).

All patients with *PRKAR1A* variants had endocrine disorders: 8/9 had an increased level of parathyroid hormone (PTH), 7/9 presented with peripheral hypothyroidism, and 2/9 with growth hormone deficiency. Conversely, only 1 patient with *PDE4D* variant had slightly increased level of PTH and no patient experienced hypothyroidism.

Additional health issues were observed including (i) recurrent ENT infections (n = 2; both with *PRKAR1A* variants), (ii) chronic erysipelas with fixed edema of the lower limbs and hypogammaglobulinemia (n = 1; *PRKAR1A* variant), and (iii) deafness (n = 3; two *PRKAR1A* variants and one with *PDE4D*).

Moreover, both patients with *PRKAR1A* variant affecting the tyrosine 323 residue (c.968A>C and c.968A>G) had additional symptoms including cardiovascular malformations (one with external jugular vein stenosis and one with total anomalous pulmonary venous return with atrial septal defect). One (patient 8 (c.968A>C)) had severe laryngotracheomalacy with cricoid hemangioma, whereas the other (patient 9 (c.968A>G)) required the dilatation of a subglottic diaphragm in infancy.

Noteworthy, one of the elder patients (patient 5) with the p. Arg368* *PRKAR1A* variant died suddenly at 38 years of age, following severe bronchospasm during a surgical procedure. She had concomitant lactic acidosis, rhabdomyolysis, and anuric renal deficiency. No clear etiology was determined despite extensive metabolic and infectious work-up.

All patients had shortened tubular bones of the hands and feet, of all the digits, associated with cone-shaped epiphyses (Fig. 3). Besides these canonical signs, other radiological features were noticed: advanced bone age (in 4/9 with



Fig. 3 Patients presenting with acrodysostosis. **a**, **b** Patient 5; general aspect and hand X-ray. **c**–**e** Patient 9; face, hand, and hand X-ray. **f**–**h** Patient 15; face, hand, and hand X-ray. Please note on **c** and **f** the

typical facial gestalt with malar hypoplasia and nasal hypoplasia. Please note on **b**, **e**, and **h** the short broad metacarpals and phalanges, with cone-shaped epiphyses for patient 15

PRKAR1A variants and 6/11 with *PDE4D*), bilateral coxa valga (4/9 with *PRKAR1A* variants and 3/11 with *PDE4D*), irregular vertebral plates (2/9 with *PRKAR1A* variants and 2/11 with *PDE4D*), absence of widening of the interpedicular distance (3/9 with *PRKAR1A* variants and 2/11 with *PDE4D*). Moreover, 5/9 patients with *PRKAR1A* variants had skeletal abnormalities also affecting other epiphyses and one case had an important vertebral dysplasia in the first years of life, which progressively improved with age.

Clinical, biological, and radiological data of the five acroscyphodysplasia cases

All patients but one presented with postnatal growth retardation (-2 to -4 SD), after intrauterine growth retardation in only one case. Four patients have nasal hypoplasia, severe in two cases only; all five had a flat face with malar hypoplasia. They all had also micromelia and 3/5 had a severe brachydactyly with small phalanges and small metacarpals and metatarsals (two with moderate brachydactyly) (Fig. 4). Cone-shaped epiphyses were observed except in one young case. Two patients had knee stiffness. One patient presented with a peripheral hypothyroidism, but no other endocrine disorder was reported. All had an intellectual disability. Some non-specific MRI abnormalities were observed (one with thin corpus callosum, delayed myelinisation and cortical atrophy, another one with ventricular dilatation).

Among the 3 patients with no identified variants, 2 had a medical history of an early and severe infection and the last one of frequent infections.

Discussion

We describe here a large cohort of 27 patients with acrodysostosis. We identified variants in 74% of cases (20/ 27) including *PDE4D* variants in 11 and *PRKAR1A* in 9.

We further confirm genotype–phenotype correlations. Indeed, the 11 patients carrying *PDE4D* variants had the characteristic facial features with nasal hypoplasia, midface hypoplasia, and prominent mandible initially reported in acrodysostosis, and 10/11 had some degree of intellectual disability. This characteristic facial dysostosis and mild to moderate intellectual disability was observed, respectively, in only 3/9 and 1/9 cases carrying *PRKAR1A* variant. Noteworthy those patients had variants affecting the tryptophane 262 or the tyrosine 323 residues of PRKAR1A



Fig. 4 X-rays of the acroscyphodysplasia cases showing the whole legs, the zoom on the knee and a hand. **a–c** 1st case. **d–f** 2nd case. **g–i** 3rd case. **j–l** 4th case. **m–o** 5th case. Please note the cup-shaped

metaphyses with embedded epiphyses on knees and the aspect of the hands, similar to the acrodysostosis cases

(c.786G>C, c.968A>C, and c.968A>G), but not the recurrent p.Arg368* variant. Conversely, hormone resistance was consistently observed in patients carrying *PRKAR1A* variants (8/9 presenting with chronic resistance to parathyroid hormone and 7/9 had peripheral hypothyroidism), whereas only one case with *PDE4D* variant had a slightly increased level of PTH.

These results are in agreement with all cases reported so far (n = 49; 24 with *PRKARIA* variants and 25 with *PDE4D* variants) [4, 6, 7, 10–16] (cf Table 2). In the literature, the peripheral bone dysostosis is consistently observed in all patients. The severe brachymetacarpia/brachymetatarsia and brachydactyly affect all the digits. The lumbar stenosis is also described in patients with *PRKARIA* or *PDE4D* variants, but this data is scarcely specified in literature. Although short nose with flat face is consistently observed in all reported cases, severe nasomaxillary hypoplasia is more frequently reported in patients with *PDE4D* variants (n = 24 for whom data are specified), than in patients with *PRKAR1A* variants (n = 6 for whom data are specified). Intellectual disability is clearly reported in 24 cases of patients with *PDE4D* variants (of 25 reported in literature) but only in 5 with *PRKAR1A* variants (of 24 reported). Hormone resistance is conversely observed in only 6 patients carrying *PDE4D* variants (6/25 cases), whereas is present in 17 patients with *PRKAR1A* variants (of 24 cases (19 with the data available)) in literature.

We therefore confirm that molecular analysis shall be directed according to the symptoms: in case of facial dysostosis and moderate intellectual disability, first screen the *PDE4D* gene, and in case of less characteristic facial features and hormonal resistance first screen for *PRKAR1A* variants.

Our study also further highlights the risk of development of serious medical complications. Actually 3 patients with *PRKAR1A* and 2 with *PDE4D* variants developed progressive obesity which confirms previous publications [6]. cAMP signaling pathways, mediated by protein

Table 2 Com	parison of the	e clinical and	biological c	lata of the pa	atients of th	ne present	study to t	he literatu	re						
Patient	Linglatt et al. $[4]$ $(n = 3)$	Lee et al. [11] $(n = 2)$	Nagasaki et al. $[15]$ $(n = 1)$	Linglart et al. [7] $(n = 11$ news)	Muhn et al. [14] $(n = 4+\Pi)$	Kaname Kaname ($n = 1$) ($n = 1$)	Li et al. $[12]$ $(n = 1)$	Rhayem et al. [16] $(n = 1 $ new)	This study $(n = 9)$	Lee et al. [11] $(n = 3)$	Linglart et al. $[7]$ (n = 2 news)	Lynch et al. [6] $(n=8)$	Lindstrand et al. $[13]$ (n = 5)	Kaname et al. [10] $(n = 7)$	This study $(n = 11)$
Sex Age (death?) Ethnicity	Europe and mixed Europe/ west Africa	2M	Japanese	5M/6F 3 to 26 y	1M/3F 11.5 to 31 y Vietnamese / Caucasian	1M 3.5 to 39 y Japanese and Korean	12 y Chinese	9 y	6F/3M 5.5 to 37 y Caucasian, Maghreb, Chinese	2M/IF	2F 3 to 26 y	2.5 to 41 y	3.5 to 14.5 y	4M/3F 3.5 to 39 y Japanese and Korean	4F/7M 4.5 to 33 y Caucasian, Turkish, South
Gene sequencing PRKAR1A	p.Arg368* (x3)	p.Arg335Pro ; p lle327Thr	. P. Thr239Ala	p.Arg368* (x6); p. Gln285Arg ; Gln2859Glu ; p. Ala328Val; p. Ala328Val; p. P. P. P. P. D. Gln372*	p.Arg368*, P. Ala213Thr, Tyr373Cys, P. Arg335Cys	p.Arg368*]	p. Gly289- Glu	p. Tyr175- Cys	p.Arg368*, p. Tyr373His, p.323Ser, p. Trp262Cys, p. Tyr323Cys		~		~	~	Arabia /
PDE4D							~			p Gin22Glu; p. Giu590Ala; p. Giy673Asp	p. Ala227Ser; Glu590Ala Glu590Ala	p.Ala24.3Val (x4) ; p. (x1) ; p. Val268.Ma; p. Ser240Thr, pro164Leu Pro164Leu	p. Phe26Val; p.Met303Val; p.Vaf229Ala; p.Phe226Cys; p.Ile678Thr	p.Ile678Thr (x2); p. (x2); p. (a)673D; p. Glio228Phe (x2); p.Thr587Ala p.Thr587Ala	p. Phe226- Psc. p. Psc. p. Ser 190- Ser 190- Ala, p. Thr591As- Thr591As- n. p. Dr0, p. Pr0, p. Pr0, p. Leu316- Pr0, p. Leu316- Pr0, a. Pr0, a. Leu316- Pr0, a. Pr0, a. Leu316- Pr0, a. Pr0, a. Leu316- Pr0, a. Pr0, a. Leu316- Pr0, a. Leu316- Pr0, a. Pr0, a. Leu316- Pr0, a. Leu325- Pr0, a.
IUGR	+ 3/3	NA	I	+ 8/11	NA		+		8	NA	+1/2				Asn 4
Postnatal growth retardation	+ 3/3	+ 2/2	+	+ 7/11	+ 3/4	- 1/1	+	I	8	+ 2/2	+1/2	- (7/8)	- (3/5)	+477	3
Actual height (SD)	-2.5 to -4	Mild	-3.1	-1 to -4.6	-1.1 to -3.3	-1.7	-2	+1	-6.7 to -0.9	Mild	0.4 to -3		-0.7 to -2.9	-0.2 to -5.9	-3 to +1.8
Obesity Facial dysostosis	- 3/3	NA		+ 2/11	NA	- 1/1	I		3	NA	+ 2/2	+ (4/8)	- (4/5)	L/L	1
Nasal hypoplasia	+ 3/3	+ 2/2	+	+ 11/11	+ 2/4	+ 1/1	+		3	+ 3/3	+ 2/2	+ (1/8)	+ (5/5)	<i>L1L</i> +	6
Depressed nasal bridge	NA	Midface hypoplasia		+ 5/11 severe hypoplasia	+ 1/4	+ 1/1 mild NM hypoplasia	+		8	Midface hypoplasia	– 2/ 2 severe hypoplasia			+ 7/7 severe NM hypoplasia	11
Prominent mandibule	NA	NA		NA	+ 1/4	NA				NA	NA			NA	5
Peripheral dysostc	sis + 3/3	+ 2/2	+	+ 11/11	+ 4/4	+ 1/1 mild			6	+ 3/3	+ 2/2	+ (1/8)	+ (5/5)	+ 7/7 severe	11

SPRINGER NATURE

Table 2 (contir	nued)														
Patient	Linglart et al. [4] $(n = 3)$	Lee et al. [11] (<i>n</i> =2)	Nagasaki et al. [15] $(n = 1)$	Linglart et al. [7] $(n = 11$ news)	Muhn et al. [14] $(n = 4+/7)$	Kaname et al. $[10]$ $(n = 1)$	Li et al. $[12]$ $(n = 1)$	Rhayem et al. [16] (n = 1 new)	This study $(n = 9)$	Lee et al. [11] (<i>n</i> =3)	Linglart et al. [7] (n = 2 news)	Lynch et al. [6] $(n = 8)$	Lindstrand et al. $[13]$ $(n = 5)$	Kaname et al. [10] $(n = 7)$	This study $(n = 11)$
Severe brachydactyly Short, broad metaarsals, metacarpals and phalanges	+ 3/3	+ 2/2	+	+ 11/11	+ 4/4	+ 1/1	+	+	6	+ 3/3	+ 2/2	(9/9) +	+ (4/4)	L/L +	10
Cone-shaped epiphyses	+ 3/3	NA	+	+ 11/11	+ 4/4	NA		+	8	NA	+ 2/2	+ (2/6?)	NA	NA	٢
Advanced bone age X-ravs	+ 3/3	NA		+ 9/11	NA	+ 1/1	+		4	NA	+ 2/2	+ (2/2) and 1 N	NA	L/L +	9
Narrowing of caudal vertebral interpedicular distance	+ 3/3	+ 2/2			+ 1/4		NA	+	Ś	+ 3/3		I	+ (2/3)		-
Hormonal screening	16														
PTH (ng/L)	† 3/3		←	↑ 11/11	† (2/2)	N 1/1	←	←	8↑		N 2/2	N (8/8)	↑ (4/5)	<i>L/L</i> N	1 slightly↑
GH secretory response	Impaired 2/2		Z		↑ HGH (1/3)				1 complete deficiency						z
Gonadotropic axis	Mild gonadotropin resistance		Z		† FSH (1/3)				Z						z
OGE	2M: bilateral cryptorchidism —1F: normal	cryptorchidism: 1/2M		M: normal (5/ 5)—F: 2/6 with irregular cycles	Z				1 cryptorchidism	Unilateral undescended testis : 1/2	Normal	5M: 2 hypospadias; 3 cryptorchidism —3F: normal	4M:1 cryptorchidism		1 testicular ectopy; 1 feminine hypospa- dias
Free T4 (pmol/ L)	N 3/3	Congenital hypothyroidism 1/2	z	N or \downarrow	NA		z	z	N or (Congenital hypothyroidism 1/3	N or \downarrow	N (8/8)	N (4/4)		z
TSH (mUI/L) Neurology	† 3/3		←	† 9/11	† (2/3)	† 1/1	←	←	8↑		N 2/2	N (7/8)	N (4/4)	LIL N	z
Mental retardation		+ 2/2 (mild)	+ (mild)	- 11/11	+ 1/4 (mild)	+ 1/1 mild	I		1	+ 2/3 (1 mild; 1 significant)	+ 2/2	+ (8/8)	+ (5/5)	+ 7/7 (1 mild/ 6 severe)	6
Other	CNS calcifications 1/3	νA		+ 5/11 (behavioral troubles)	Coarse hair (1), sensineural deafness (1)				~	NA	 – 2/2 (behavioral troubles) 				Red hair, intracra- nial hyperten- sion

kinase A (PKA), have an important role in the metabolic homeostasis including the regulation of adiposity [17]. In mice, targeted disruption of the RII beta gene, coding for a regulatory subunit of PKA, leads to a lean phenotype with resistance to diet-induced obesity [18]. Cushing syndrome with bilateral adrenocortical hyperplasia may be caused by inactivating variants of *PRKAR1A* leading to an increase of PKA activation. The patients with these inactivating *PRKAR1A* variants have a lower BMI than the ones without variants, confirming a link between germline defects of PKA and human obesity phenotypes [19].

Similarly, PDE4D which converts cAMP to AMP, countering activation of PKA, is an insulin-responsive gene important for metabolism regulation, including adipocyte lipolysis [20]. Rat adipocytes treated with the PDE4 inhibitor rolipram showed a significant increase in lipolysis and reversed in part prostaglandin E2 antilipolysis [20]. Furthermore, cAMP-dependent PKA pathway is also involved in insulin secretion and resistance. Overexpression of PDE4D in α -cells reduced insulin secretion, whereas inhibition of PDE4 activity by rolipram or knockdown of PDE4D restored it [21]. One patient with acrodysostosis has clearly been described with a metabolic syndrome and hypertension [22]. Although the physiopathological link between PRKAR1A and PDE4D variants and human obesity and insulin resistance will require further study, follow-up of patients should be careful on the BMI, blood pressure, and glucose status.

We also report two patients with PDE4D variants who developed intracranial hypertension and thrombophlebitis. One adult patient with acrodysostosis and one child with PDE4D variant have also been described with a deep vein thrombosis [13, 23]. Actually, several case-control studies have assigned an association between PDE4D variants and risk of ischemic stroke among different ethnicities [24]. Moreover, PDE4D is associated with inflammation and reduced PDE4D is assumed to predispose individuals to atrial fibrillation, which increases the risk of cardiogenic stroke [25]. More studies are necessary to assess the functional and physiological effects of the described PDE4D SNPs. We advise to pay special attention to the thromboembolic risk during follow-up. Actually we emphasize the importance of a multidisciplinary long-term follow up, including cardiovascular and thromboembolic risk factors and we propose a short checklist for medical management (Table 3).

In this study, we also investigated 5 rare cases of acroscyphodysplasia initially described by Verloes et al. in 1991 [9]. All 5 patients had the characteristic metaphyseal changes, but only 3 patients had very short hands and feet, considered as a main criterion for this diagnosis [9, 26]. Among these 3 patients, *PDE4D* variants were identified in

the 2 with severe growth retardation (-3 and -4 SD), severe nasal hypoplasia and stiffness of the knees. This finding suggests that *PDE4D* screening should be considered in case of knee cup-shaped metaphyses with embedded epiphyses when associated with acrodysostosislike brachydactyly and nasal hypoplasia. As few observations of typical knee cup-shaped metaphyses with embedded epiphyses have been described after repeated injuries or meningococcemia [26], the reported infections could be at least a part of the cause of the aspect of acroscyphodysplasia in 2/3 patients without identified *PDE4D* variants. A recent study has described *GNAS* variants in 2 cases of acroscyphodysplasia, supporting the hypothesis of a genetic heterogeneity for this condition [8].

The majority of described cases of acrodysostosis have been sporadic with no familial medical history. In our study, we described two cases of mother-to-child transmission of PDE4D variants (patients 17 and 18). Prior to PRKAR1A and PDE4D identification, several reports have described autosomal dominant inheritance, from mother [27-31] or father [32]. Only one report described concordant siblings with seemingly unaffected parents [33], but it has been supported by the observation of Lynch et al. [6]. They described three affected children with clinically unaffected parents, but with a PDE4D variant-carrying father. They raise the hypothesis of an imprinting of PDE4D gene to explain the variable expressivity, as in the mouse genome, PDE4D showed a paternal bias in transmission [34]. However, the familial cases we described with a maternal transmission of PDE4D variants and clear phenotype in the kindred rather opposes this hypothesis. More familial cases will be needed to further accord the resulting phenotypes regarding the parental inheritance.

Finally, neither PDE4D nor PRKAR1A variants were found in seven patients with characteristic skeletal features but no hormone resistance or facial dysostosis, supporting that other disease genes may account for these remaining acrodysostosis cases. Interestingly, both forms of acrodysostosis (ACRO1-MIM 101800-corresponding to variants of PRKAR1A and ACRO2-MIM614613-to variants of PDE4D) are due to variants of two genes of the same pathway of cAMP signaling. PRKAR1A is the cAMPdependent regulatory subunit of protein kinase A and PDE4D is a class IV cAMP-specific phosphodiesterase, regulating cAMP concentration. The involvement of PDE4D in acrodysostosis further supports the key role of cAMP signaling pathway in skeletogenesis, as previously shown for Albright hereditary osteodystrophy due to GNAS variants. The conditions share several clinical features (metacarpal abnormalities, obesity, and resistance to hormones depending on generation of cAMP for acrodysostosis with PRKAR1A variants), but some manifestations are variable in severity (degree and extent of brachydactyly and

Table 3 Proposed management recommandations for the follow-up of patients presenting with acrodysostosis

Assessments	At diagnosis	Minimal frequency
Physical examination		
Vital parameters (arterial blood pressure)	Х	Systematic per year
Physical examination (including facial gestalt, neurologic examination)	Х	
Anthropometrics measurements (BMI)	Х	
Joint mobility	Х	
Vertebral static assessment	Х	
Developmental milestones assessment	Х	
Endocrine screening		
Calcemia, phosphoremia, PTH, 25-hydroxy-vitamine D, FGF23, calciuria/creatininuria	Х	Systematic per year
T3, T4, TSH	Х	Systematic per year
IGF1 (+/- GH stimulation test)	Х	Once, if growth deficiency
Gonadotropic axis testing	Х	Depending on puberty
Blood glucose level	Х	Systematic per year
Others		
Genetics	Х	Systematic per year
Ear, nose, and throat (deafness? infections?)	Х	Systematic per year
Oral and maxillofacial (teeth? Nasal hypoplasia?)	Х	Systematic per year
Polygraphy	Х	If symptoms of sleep apnea syndrome
Thromboembolic risk factors	Х	Systematic per year
Heart ultrasound	Х	Once
X-rays	Х	Depending on orthopedics
Cerebral MRI	Х	Once, +/- depending on clinic
Psychological support	Х	Systematic

degree of parathyroid hormone resistance) and some others are quite similar. This clinical variability may be possibly linked to the tissue-specificity of the imprinting of GNAS or to the tissue-specific expression of alternative isoforms of protein kinase A or of phosphodiesterase 4 [4]. Finally, the dysregulation of cAMP intracellular signaling in response to a number of membrane-impermeable hormones could be the underlying common mechanism of acrodysostosis. Further study on the genomes of patients without identified variants is required to elucidate the molecular basis in these cases, but it might be hypothesized that the putative other genes involved in acrodysostosis are linked to regulation of cAMP signaling.

Acknowledgements We thank all patients and their families for their contribution to this work.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- 1. Maroteaux P, Malamut G. Acrodysostosis. Presse Med. 1968;76:2189–92.
- Robinow M, Pfeiffer RA, Gorlin RJ, et al. Acrodysostosis. A syndrome of peripheral dysostosis, nasal hypoplasia, and mental retardation. Am J Dis Child. 1971;121:195–203.
- Bastepe M, Juppner H. GNAS locus and pseudohypoparathyroidism. Horm Res. 2005;63:65–74.
- Linglart A, Menguy C, Couvineau A, et al. Recurrent PRKAR1A variant in acrodysostosis with hormone resistance. N Engl J Med. 2011;364:2218–26.
- Michot C, Le Goff C, Goldenberg A, et al. Exome sequencing identifies PDE4D variants as another cause of acrodysostosis. Am J Hum Genet. 2012;90:740–5.
- Lynch DC, Dyment DA, Huang L, et al. Identification of novel variants confirms PDE4D as a major gene causing acrodysostosis. Hum Mutat. 2013;34:97–102.
- Linglart A, Fryssira H, Hiort O, et al. PRKAR1A and PDE4D variants cause acrodysostosis but two distinct syndromes with or without GPCR-signaling hormone resistance. J Clin Endocrinol Metab. 2012;97:E2328–2338.
- Mitsui T, Kim OH, Hall CM, et al. Acroscyphodysplasia as a phenotypic variation of pseudohypoparathyroidism and acrodysostosis type 2. Am J Med Genet A. 2014;164A: 2529–34.

- 9. Verloes A, Le Merrer M, Farriaux JP, Maroteaux P. Metaphyseal acroscyphodysplasia. Clin Genet. 1991;39:362–9.
- Kaname T, Ki CS, Niikawa N, et al. Heterozygous variants in cyclic AMP phosphodiesterase-4D (PDE4D) and protein kinase A (PKA) provide new insights into the molecular pathology of acrodysostosis. Cell Signal. 2014;26:2446–59.
- Lee H, Graham JM Jr., Rimoin DL, et al. Exome sequencing identifies PDE4D variants in acrodysostosis. Am J Hum Genet. 2012;90:746–51.
- Li N, Nie M, Li M, et al. The first variant identified in a Chinese acrodysostosis patient confirms a p.G289E variation of PRKAR1A causes acrodysostosis. Int J Mol Sci. 2014;15: 13267–74.
- Lindstrand A, Grigelioniene G, Nilsson D, et al. Different variants in PDE4D associated with developmental disorders with mirror phenotypes. J Med Genet. 2014;51:45–54.
- Muhn F, Klopocki E, Graul-Neumann L, et al. Novel variants of the PRKAR1A gene in patients with acrodysostosis. Clin Genet. 2013;84:531–8.
- Nagasaki K, Iida T, Sato H, et al. PRKAR1A variant affecting cAMP-mediated G protein-coupled receptor signaling in a patient with acrodysostosis and hormone resistance. J Clin Endocrinol Metab. 2012;97:E1808–1813.
- 16. Rhayem Y, Le Stunff C, Abdel Khalek W, et al. Functional characterization of PRKAR1A variants reveals a unique molecular mechanism causing acrodysostosis but multiple mechanisms causing carney complex. J Biol Chem. 2015;290:27816–28.
- McKnight GS, Cummings DE, Amieux PS, et al. Cyclic AMP, PKA, and the physiological regulation of adiposity. Recent Prog Horm Res. 1998;53:139–59.
- Cummings DE, Brandon EP, Planas JV, Motamed K, Idzerda RL, McKnight GS. Genetically lean mice result from targeted disruption of the RII beta subunit of protein kinase A. Nature. 1996;382:622–6.
- London E, Rothenbuhler A, Lodish M, et al. Differences in adiposity in Cushing syndrome caused by PRKAR1A variants: clues for the role of cyclic AMP signaling in obesity and diagnostic implications. J Clin Endocrinol Metab. 2014;99: E303–10.
- Wang H, Edens NK. mRNA expression and antilipolytic role of phosphodiesterase 4 in rat adipocytes in vitro. J Lipid Res. 2007;48:1099–107.

- Kim MJ, Park SK, Lee JH, et al. Salt-inducible kinase 1 terminates cAMP signaling by an evolutionarily conserved negative-feedback loop in beta-cells. Diabetes. 2015;64:3189–202.
- Atabek ME, Pirgon O, Sert A. Metabolic syndrome manifestations in an adolescent with acrodysostosis. J Pediatr Endocrinol Metab. 2007;20:739–41.
- Sezer N, Sutbeyaz ST, Koseoglu F, Aras M, Akin C. Adult case of acrodysostosis with severe neurologic involvement. J Back Musculoskelet Rehabil. 2009;22:125–9.
- Das S, Roy S, Munshi A. Association between PDE4D gene and ischemic stroke: recent advancements. Int J Neurosci. 2016;126: 577–83.
- Jorgensen C, Yasmeen S, Iversen HK, Kruuse C. Phosphodiesterase-4D (PDE4D)—a risk factor for atrial fibrillation and stroke? J Neurol Sci. 2015;359:266–74.
- Dieux-Coeslier A, Moerman A, Holder M, et al. Metaphyseal chondrodysplasia with cone-shaped epiphyses: a specific form involving the lower limbs. Am J Med Genet A. 2004;124A: 60–6.
- Cantú JM, Hernández A, Panduro-Cerda A, et al. Autosomal dominant acrodysostosis. Hum Genet. 1979;47:345–6.
- Hernández RM, Miranda A, Kofman-Alfaro S. Acrodysostosis in two generations: an autosomal dominant syndrome. Clin Genet. 1991;39:376–82.
- 29. Niikawa N, Matsuda I, Ohsawa T, Kajii T. Familial occurrence of a syndrome with mental retardation, nasal hypoplasia, peripheral dysostosis, and blue eyes in Japanese siblings. Hum Genet. 1978;42:227–32.
- Sheela SR, Perti A, Thomas G. Acrodysostosis: autosomal dominant transmission. Indian Pediatr. 2005;42:822–6.
- Steiner RD, Pagon RA. Autosomal dominant transmission of acrodysostosis. Clin Dysmorphol. 1992;1:201–6.
- Davies SJ, Hughes HE. Familial acrodysostosis: can it be distinguished from Albright's hereditary osteodystrophy? Clin Dysmorphol. 1992;1:207–15.
- Taillet-Bellemère C, Maroteaux P. Acrodysostosis in a sister and brother born to normal parents. Ann Pediatr. 1991;38:31–6.
- Babak T, Deveale B, Armour C, Raymond C, Cleary MA, van der Kooy D, Johnson JM, Lim LP. Global survey of genomic imprinting by transcriptome sequencing. Curr Biol. 2008;18:1735–41.

Affiliations

Caroline Michot¹ · Carine Le Goff¹ · Edward Blair² · Patricia Blanchet³ · Yline Capri⁴ · Brigitte Gilbert-Dussardier ⁵ · Alice Goldenberg⁶ · Alex Henderson⁷ · Bertrand Isidor⁸ · Hulya Kayserili⁹ · Esther Kinning¹⁰ · Martine Le Merrer¹ · Stanislas Lyonnet¹ · Sylvie Odent¹¹ · Pelin Ozlem Simsek-Kiper¹² · Chloé Quelin¹¹ · Ravi Savarirayan¹³ · Marleen Simon¹⁴ · Miranda Splitt¹⁰ ⁷ · Judith M.A. Verhagen¹⁵ · Alain Verloes¹³ · Arnold Munnich¹ · Geneviève Baujat¹ · Valérie Cormier-Daire¹

- ¹ Department of Medical Genetics, INSERM UMR 1163, Paris Descartes-Sorbonne Paris Cité University, IMAGINE Institute, Necker Enfants Malades Hospital, Paris, France
- ² Oxford Centre for Genomic Medicine ACE Building, Nuffield Orthopaedic Centre Oxford University Hospitals NHS Foundation Trust Headington, Oxford OX3 7LE, UK
- ³ Department of Medical Genetics, CHRU de Montpellier—Arnaud de Villeneuve Hospital, Montpellier, France
- ⁴ Department of Medical Genetics, INSERM U676, Robert Debré

Hospital, Paris, France

- ⁵ Department of Genetics, C.H.U. La Milétrie, Poitiers, France, Poitiers University, EA3808 Poitiers, France
- ⁶ Department of Genetics, CHU de Rouen, Centre of Medical Genomics and of Personalized Medicine of Normandy, Rouen, France
- ⁷ Northern Genetics Service, Institute of Genetic Medicine, Newcastle upon Tyne, UK

- ⁸ Department of Medical Genetics, C.H.U. de Nantes, Nantes, France
- ⁹ Department of Medical Genetics, Istanbul University, Istanbul Faculty of Medicine, Istanbul, Turkey
- ¹⁰ The Ferguson-Smith Centre for Clinical Genetics Royal Hospital for Sick Children, Glasgow, UK
- ¹¹ Department of Clinical Genetics, CHU de Rennes, Rennes University, CNRS IGDR (institut de génétique et développement de Rennes), UMR6290 Rennes, France
- ¹² Pediatric Genetics Unit, Department of Pediatrics, Hacettepe University Faculty of Medicine, Ankara, Turkey
- ¹³ Victorian Clinical Genetics Service, Murdoch Children's Research Institute and University of Melbourne, Melbourne, Australia
- ¹⁴ Department of Medical Genetics, University Medical Centre Utrecht, Utrecht, The Netherlands
- ¹⁵ Department of Clinical Genetics, Erasmus University Medical Center, Rotterdam, The Netherlands