

## Review Article

# Intraparenchymal Neural Stem/Progenitor Cell Transplantation for Ischemic Stroke Animals: A Meta-Analysis and Systematic Review

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Intraparenchymal transplantation of neural stem/progenitor cells (NSPCs) has been extensively investigated in animal models of ischemic stroke. However, the reported therapeutic efficacy was inconsistent among studies. To evaluate this situation, PubMed, Embase, and Web of Science databases were searched for preclinical studies using NSPC intraparenchymal transplantation in ischemic stroke animals. Data of study quality score, neurobehavioral (mNSS, rotarod test, and cylinder test) and histological (infarct volume) outcomes, cell therapy-related serious adverse events, and related cellular mechanisms were extracted for meta-analysis and systematic review. A total of 62 studies containing 73 treatment arms were included according to our criterion, with a mean quality score of 5.10 in 10. Among these studies, almost half of the studies claimed no adverse events of tumorigenesis. The finally pooled effect sizes for neurobehavioral and histological assessments were large (1.27 for mNSS, 1.63 for the rotarod test, 0.71 for the cylinder test, and 1.11 for infarct volume reduction). With further analysis, it was found that the administration time poststroke, NSPC donor species, and transplantation immunogenicity had close correlations with the degree of infarct volume reduction. The NSPC dosage delivered into the brain parenchyma was also negatively correlated with the effect of the cylinder test. Intriguingly, endogenous apoptosis inhibition and axonal regeneration played the most critical role in intraparenchymal NSPC transplantation among the cellular mechanisms. These results indicate that intraparenchymal NSPC transplantation is beneficial for neurobehavioral and histological improvement and is relatively safe for ischemic stroke animals. Therefore, intraparenchymal NSPC transplantation is a promising treatment for stroke patients.

## 1. Introduction

Ischemic stroke is one of the leading causes of death and disability around the world [1, 2]. After stroke, approximately 90% of survivors experience motor dysfunction [3],

which lasts for the rest of their life and affects their daily life quality severely. However, there are few effective treatments for the ischemic stroke. Stem cell transplantation to rescue the motor function deficits poststroke has attracted a growing interest [4–6].

Among the various cell populations used for stroke cell therapy, neural stem/progenitor cells (NSPCs) and mesenchymal stem cells (MSCs) are both investigated extensively. Meta-analysis indicates that MSC therapy lies mainly in the time-dependent bystander mediators poststroke [7–9] as MSCs possess the poor potential of neural lineage differentiation. In contrast, NSPCs were regarded as a more appropriate source because of their capabilities of differentiation to neural cell phenotypes *in vitro* and *in vivo* [10, 11]. Transplanted NSPCs can migrate to peri-ischemic areas and ameliorate functional deficits [11–13]. Multiple but inconsistent mechanisms by which NSPCs enhance functional recovery were proposed, from neuroprotection [14] to neuroregeneration [15]. Similarly, the curative benefit is conflicting among studies when the following factors are involved, including donor cell states, dosage, immunogenicity, administration time after stroke onset, and immunosuppressive medicine usage. Therefore, there is a need for systematic analysis of the studies on intraparenchymal NSPC transplantation.

We conducted a meta-analysis to determine whether intraparenchymal NSPC transplantation is beneficial in pre-clinical studies based on neurobehavioral tests (mNSS, rotarod test, and cylinder test) and histological outcome (infarct volume). In addition, we pooled results about the cellular mechanisms and serious adverse events (SAEs). We hope that this analysis provides information for potential future clinical trials involving stem cell transplantation in stroke.

## 2. Methods

This meta-analysis was carried out following the guidelines of Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA, <http://www.prisma-statement.org/>, Table S1 in Supplementary Materials available online) [16].

**2.1. Search Strategy.** We searched for correlative studies about neural stem/progenitor cell (NSPC) transplantation in animal models of ischemic stroke in three databases (PubMed, Embase, and Institute for Scientific Information Web of Science databases, up until July 11, 2018) by independent investigators. The search strategy was as follows: ((neural stem cells) OR (stem cell, neural) OR (neural progenitor cell) OR (neural precursor cell) OR NSPC) AND (stroke OR ischemic stroke OR brain ischemia OR brain infarction OR cerebral ischemia OR intracranial ischemia OR cerebrovascular OR middle cerebral artery OR anterior cerebral artery). The default language for all included studies was English. After studies were extracted, the titles, abstracts, and the secondary references were reviewed carefully. If controversy existed in whether the study is eligible, the study would be examined again and all investigators would discuss to reach a consensus.

**2.2. Inclusion and Exclusion Criteria.** This meta-analysis included controlled studies claiming that NSPCs and other vehicles (culture medium, PBS, or saline) were delivered intraparenchymally in ischemic stroke animals (nonhuman). For all included studies, neurobehavioral function

assessments must be served as one kind of outcome indicators. We excluded studies with brain lesions other than cerebral ischemia or with cell therapy paradigm using mature cells or genetically modified cells other than those for labeling or tracing *in vivo* or preconditioned cell transplantation. We also excluded studies without precise animal numbers or outcome values for individual comparison.

**2.3. Study Quality Assessment.** To estimate methodological quality for all included studies, the CAMARADES (Collaborative Approach to Meta-Analysis and Review of Animal Data from Experimental Studies) checklist was applied as follows: (1) publication on a peer-reviewed journal, (2) control of temperature, (3) randomization to treatment groups, (4) allocation concealment, (5) blinded assessment of outcomes, (6) avoidance of neuroprotective anesthetics (mostly known as ketamine), (7) use of animals with relevant comorbidities, (8) sample size calculation, (9) compliance with animal welfare regulations, and (10) statement of conflict of interest. We endowed each item one point and got the total score of every study after retrieving the full text, as well as the supplementary data and secondary references.

**2.4. Data Extraction.** Besides the study quality score, we extracted the following data from each study: first author, published year, recipient species, animal models, donor species, cell characteristics (intervention time relative to stroke onset, graft sites, and cell dose), administration of immunosuppressive drugs, and sensorimotor or histological outcomes at the final time point recorded (showing the four most used data of mNSS, rotarod test, cylinder test, and infarct volume). For the safety of intraparenchymal NSPC transplantation, data of treatment-related serious adverse events (tumor/teratoma formation, seizure, infection, or death) were considered. In addition, the data of cellular treatment effects [17] (Table S2 in Supplementary Materials available online) were extracted for further analysis. The outcome values of the rotarod test were multiplied by  $-1$  given its positive relationship with the behavioral outcome in contrast to the other three measures [7]. GetData Graph Digitizer (version 2.24) was used to quantify the mean value and standard deviation (SD) or standard error (SE) from figures, if no detailed data was referred to in the text. When all data were shown in SE, SD could be recalculated with the following formula [18]:

$$SD = \sqrt{N} \times SE, \quad (1)$$

where  $N$  means the group size.

**2.5. Statistical Analysis.** All statistical analyses were performed using Stata (ver. 12.0, StataCorp). We evaluated the standard mean difference (SMD), 95% confidence interval (CI), and significance in a DerSimonian and Laird random-effects model by the statistics of Hedges'  $g$  across all studies. The therapeutic effects of intraparenchymally transplanted NSPCs in ischemic stroke were determined by obtaining mean effect sizes, with a value of  $<0.2$  defined as a small effect,  $0.2$ – $0.8$  as a medium effect, and

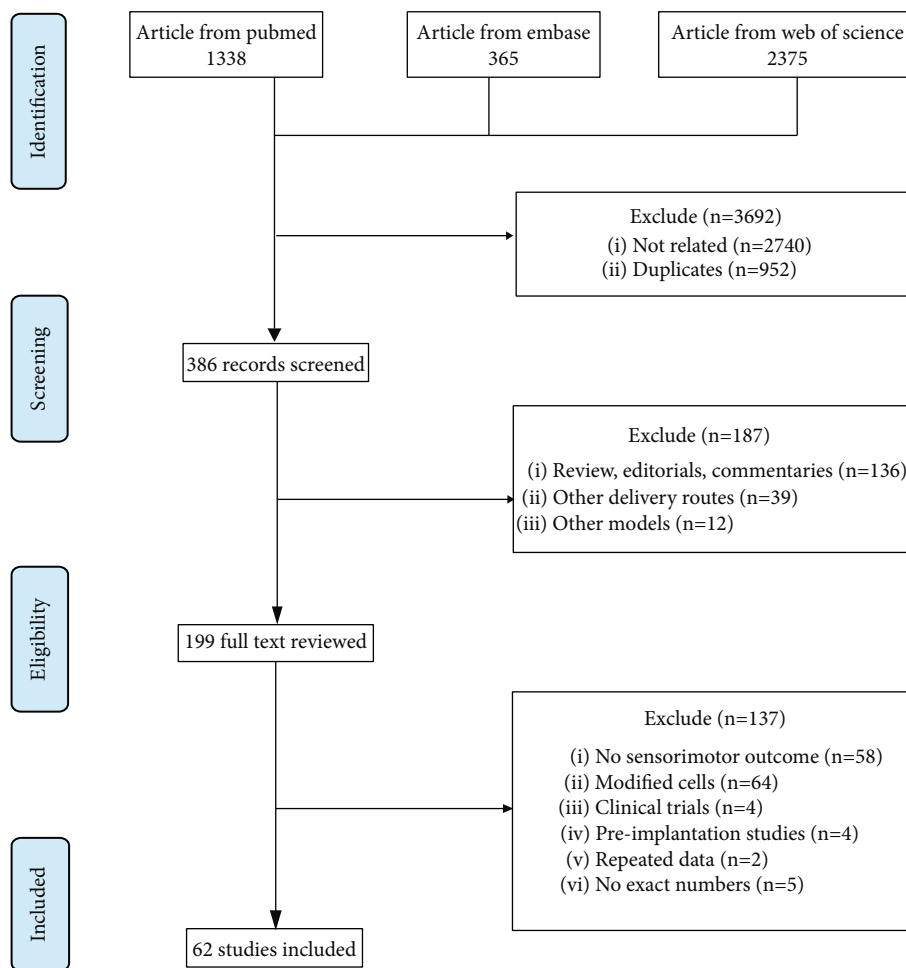


FIGURE 1: PRISMA flow diagram of including studies for this meta-analysis.

>0.8 as a large effect [19]. If heterogeneity defined by the  $I^2$  metric exists among different studies, with a value of 25%, 50%, and 75% considered to be low, moderate, and considerable heterogeneity, sensitivity analysis was used for heterogeneity analysis. Subgroup analysis and metaregression with the following clinical parameters were used for further evaluation.

Univariate metaregression analysis was carried out according to 12 variables, including (1) timing of NSPC intervention after stroke onset, (2) NSPC dose, (3) ischemic stroke models (transient, permanent), (4) graft sites (focal or global), (5) degree of NSPC immunogenicity (allogeneic or xenogeneic), (6) species of NSPC recipients, (7) species of NSPC donors, (8) state of donor NSPCs (pluripotent stem cell derivatives or primary cells), (9) administration of immunosuppressive drugs, (10) design in blindness, (11) randomization, and (12) study quality score. Afterwards, funnel plots were used to check the potential of publication bias for a visual impression. The data of significant publication bias was reassessed using a 2-tailed Egger regression intercept method. If publication bias existed, trim and fill analysis was used to adjust the bias and check whether the effect size affects the final results.

### 3. Results

**3.1. Selection and Description of Involved Studies.** The search procedure and strategies are described in Figure 1. A total of 4078 potentially relevant studies were extracted from three databases. After excluding 3692 irrelevant studies and 187 nonstandard papers, 199 studies with full text were reviewed. According to the inclusion and exclusion criteria, 137 studies dissatisfying the eligibility criteria were excluded. Finally, 62 studies containing 73 treatment arms without duplicate data description were included in this meta-analysis (Table S3 in Supplementary Materials available online). Among these identified studies, the ischemic stroke models were made transiently (46 of 62 studies) or permanently (16 of 62 studies). Rats (40 of 62 studies) and mice (19 of 62 studies) were used as recipient species, while three studies used Mongolian gerbil [20, 21] and pig [22]. The gender of all included animals was mainly male, except for seven studies with no statements [22–28], one study using female rats [29], and one study stating nonsex difference [30]. Meanwhile, human (28 of 62 studies), rat (14 of 62 studies), and mouse (20 of 62 studies) cells were chosen as the NSPC donors. The transplanted NSPCs were either from primary

cultures (45 of 73 comparisons) or from pluripotent stem cell derivatives (28 of 73 comparisons). There were also 21 studies stating the usage of immunosuppression drugs and 24 studies without it; the rest did not make a clear statement.

For SAEs related to intraparenchymal NSPC transplantation, tumor formation was most reported in 30 of the 62 studies (shown with an asterisk symbol in Table S3 in Supplementary Materials available online), but with no quantitative data.

**3.2. Study Quality Assessment.** The mean quality score across all studies was 5.10, ranging from 2 to 8 (Table S4 in Supplementary Materials available online). After being standardized by CAMARADES checklists, all the included studies were published in peer-reviewed journals and had claimed compliance with animal welfare regulations. No animals with relevant comorbidities were used for any of the studies. In addition, only thirteen studies did not describe the control of temperature, three studies had allocation concealment stressed, about 27 studies declared randomization to treatment, 39 studies described blinded assessment of outcomes, 38 studies avoided ketamine as anesthetics, and 35 studies stated conflict of interest (Table 1).

**3.3. Meta-Analysis and Effect Size.** Random-effects meta-analysis was performed for four considered measurements, and the pooled effect sizes ranged from 0.73 to 2.02 (2.02 (95% CI: 1.50–2.55) for mNSS, 1.54 (95% CI: 0.92–2.15) for the rotarod test, 0.73 (95% CI: 0.44–1.03) for the cylinder test, and 1.24 (95% CI: 0.83–1.65) for infarct volume) (Figure S1 in Supplementary Materials available online). However, the heterogeneity between studies for mNSS was large ( $\chi^2 = 84.74$ ,  $p \leq 0.001$ ,  $I^2 = 75.2\%$ ,  $df = 21$ ) (Figure S1(a) in Supplementary Materials available online). By sensitivity analysis, seven treatment arms whose effect size was greater than 3.0 were removed and the heterogeneity decreased ( $\chi^2 = 20.82$ ,  $p = 0.106$ ,  $I^2 = 32.8\%$ ), with the corrected effect size of 1.27 (95% CI: 0.94–1.60) (Figure 2(a)). For the rotarod test, the heterogeneity among studies was big ( $\chi^2 = 169.97$ ,  $p \leq 0.001$ ,  $I^2 = 87.1\%$ ,  $df = 22$ ) (Figure S1(b) in Supplementary Materials available online). When sensitivity analysis was performed, the heterogeneity went down ( $\chi^2 = 25.97$ ,  $p = 0.038$ ,  $I^2 = 42.2\%$ ) (Figure 2(b)) after three treatment arms with effect size greater than 3.0 and four other inadaptable comparisons were removed (Table S5 in Supplementary Materials available online). The corrected effect size was 1.69 (95% CI: 1.34–2.04) (Figure 2(b)). The heterogeneity among studies for the cylinder test was moderate ( $\chi^2 = 19.38$ ,  $p = 0.197$ ,  $I^2 = 22.6\%$ ,  $df = 15$ ) (Figure S1(c) in Supplementary Materials available online; Figure 2(c)). As for the infarct volume, the heterogeneity among studies was high ( $\chi^2 = 135.19$ ,  $p \leq 0.001$ ,  $I^2 = 77.1\%$ ,  $df = 31$ ) (Figure S1(d) in Supplementary Materials available online). After four inadaptable comparisons were removed by sensitivity analysis, the heterogeneity was slightly altered ( $\chi^2 = 107.27$ ,  $p \leq 0.001$ ,  $I^2 = 74.8\%$ ), with the corrected effect size of 1.11 (95% CI: 0.73–1.50) (Figure 2(d)).

Effect sizes for cellular mechanisms related to NSPC transplantation in ischemic stroke animals, including apoptosis inhibition, immunomodulation, neurotrophic factor release, angiogenesis, neurogenesis, gliosis reduction, white matter function, enzyme supplementation, and axonal function, were evaluated (Table 2). Among these mechanisms, apoptosis reduction and axonal function were the most obvious effects following NSPC intraparenchymal transplantation (Table 2).

**3.4. Subgroup Analysis and Metaregression Analysis.** Following the effect size evaluation and sensitivity analysis, the heterogeneity in infarct volume effect size was still detected. Then, subgroup analysis and metaregression analysis based on the 12 clinically related parameters were chosen to analyze their contribution to statistical heterogeneity for infarct volume effect size. We found that administration time post-stroke was negatively related to the infarct volume effect size ( $Adj R^2 = 26.92\%$ ,  $p = 0.008$ ), with an earlier NSPC transplantation and a bigger effect size (Table 3, Figure 3(a)). NSPC donor species also had a close correlation with infarct volume effect size ( $Adj R^2 = 20.00\%$ ,  $p = 0.025$ ), with mouse donors having more infarct volume reduction, indicating a preference for homologous transplantation (Table 3, Figure 3(b)). In addition, the transplantation immunity was correlated with the infarct volume reduction, with allograft indicating more reserved tissue ( $Adj R^2 = 15.24\%$ ,  $p = 0.044$ ) (Figure 3(c)). Meanwhile, for the cylinder test effect size, the NSPC dosage was discovered to be a negative correlative variable, with low dose for a better behavioral function ( $Adj R^2 = 100.00\%$ ,  $p = 0.063$ ) (Figure 3(d)).

**3.5. Publication Bias.** From the funnel plots (Figure 4), an approximately symmetrical distribution for mNSS and infarct volume was visualized, which means no publication bias. This was further confirmed by the Egger test in which the relative  $p$  values were 0.185 and 0.110 for mNSS and infarct volume, respectively, both of which exceeded 0.1. Although publication bias existed for the rotarod test and the cylinder test, with  $p$  values related to the Egger test equal to 0.094 and 0.073, respectively, these two recalibrated effect sizes were still larger than 1 (1.63 for the rotarod test and 0.71 for the cylinder test) after trim and fill analysis.

## 4. Discussion

This meta-analysis includes 62 controlled animal studies published from year 2004 to 2018. We found that intraparenchymal NSPC transplantation benefits neurobehavioral (mNSS, rotarod test, and cylinder test) and histological (infarct volume) outcomes poststroke with large effect sizes. The effect size of infarct volume reduction is correlated closely with administration time poststroke, NSPC donor species, and transplantation immunogenicity. The cylinder test effect size is correlated with NSPC donor dosage. No correlation was found between neurobehavioral or histological changes and other variables, such as stroke models, graft sites, species of NSPC recipients, state of donor NSPCs, immunosuppressive drugs, blinding and

TABLE 1: Distribution of included studies according to CAMARADES checklists.

| Criteria                                   | Number of studies | Per (%) |
|--|-------------------|---------|
| Publication on a peer-reviewed journal     | 62                | 100     |
| Control of temperature                     | 49                | 79.0    |
| Randomization to a treatment group         | 27                | 43.5    |
| Allocation concealment                     | 3                 | 4.8     |
| Blinded assessment of outcome              | 39                | 62.9    |
| Avoidance of neuroprotective anesthetics   | 38                | 61.3    |
| Use of animal with relevant comorbidities  | 0                 | 0       |
| Sample size calculation                    | 0                 | 0       |
| Compliance with animal welfare regulations | 62                | 100     |
| Statement of conflict of interest          | 35                | 56.5    |

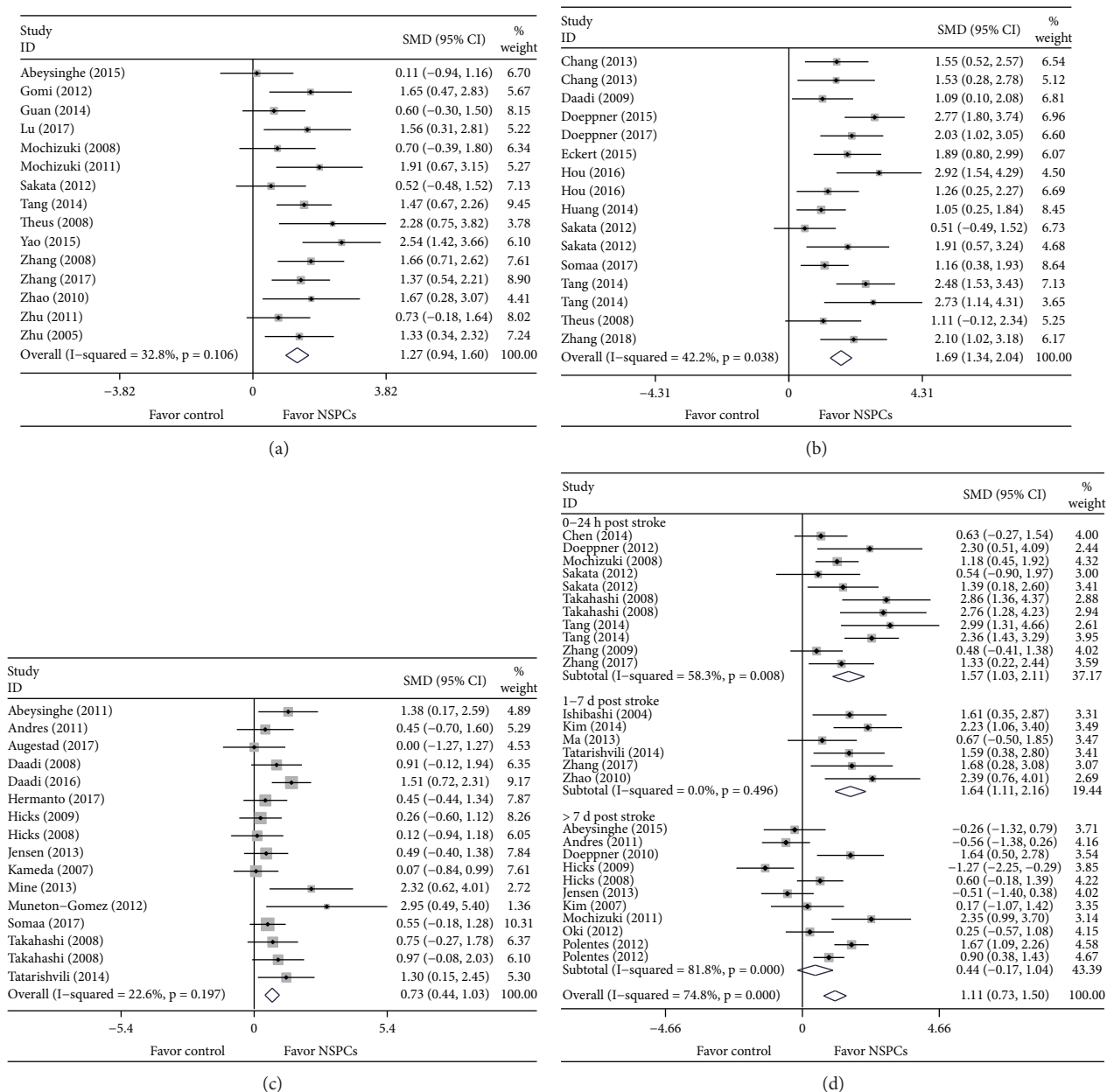


FIGURE 2: Effect sizes for intraparenchymal NPSC transplantation across related studies. Forest plot shows the mean effect sizes and 95% CI for (a) mNSS, (b) rotarod test, (c) cylinder test, and (d) infarct volume. SMD: standardized mean difference; CI: confidence interval; W: weight.

TABLE 2: Effect sizes for cellular treatment effects.

| Treatment effects      | Effect size | 95% CI     | Treatment arms |
|------------------------|-------------|------------|----------------|
| Apoptosis inhibition   | 3.09        | 1.94–4.25  | 13             |
| Immunomodulation       | 1.47        | 0.34–2.61  | 13             |
| Growth factors (BDNF)  | 1.37        | 0.57–2.17  | 14             |
| Growth factors (VEGF)  | 1.15        | 0.47–1.84  | 13             |
| Angiogenesis           | 1.18        | 0.42–1.94  | 11             |
| Neurogenesis           | 1.23        | 0.40–2.07  | 13             |
| Gliosis reduction      | 2.35        | 0.22–4.49  | 5              |
| White matter function  | 1.00        | 0.40–1.61  | 3              |
| Enzyme supplementation | 0.81        | –0.36–1.97 | 6              |
| Axonal function        | 2.47        | 1.36–3.57  | 1              |

TABLE 3: Subgroup meta-analysis and metaregression of variants correlated with infarct volume reduction.

| Clinical variants              | Effect size (95% CI) | $I^2$ , $p$ value   | df | Univariate analysis (Adj $R^2$ , $p$ ) |
|--------------------------------|----------------------|---------------------|----|--|
| Administration time poststroke |                      |                     |    | 26.92%, 0.008                          |
| $\leq 24$ h                    | 1.57 (1.03–2.21)     | 58.3%, 0.008        | 10 |  |
| 1–7 d                          | 1.64 (1.11–2.16)     | 0.0%, 0.496         | 5  |  |
| $\geq 7$ d                     | 0.44 (–0.17–1.04)    | 81.8%, $\leq 0.001$ | 10 |  |
| Cell dosage                    |                      |                     |    | 1.27%, 0.261                           |
| $\leq 1 \times 10^6$ cells/kg  | 1.45 (0.99–1.91)     | 57.4%, 0.012        | 9  |  |
| $1–5 \times 10^6$ cells/kg     | 0.82 (0.06–1.59)     | 83.1%, $\leq 0.001$ | 10 |  |
| $> 5 \times 10^6$ cells/kg     | 0.99 (0.25–1.73)     | 61.5%, 0.016        | 6  |  |
| Animal model                   |                      |                     |    | 0.79%, 0.280                           |
| Transient                      | 1.24 (0.82–1.67)     | 68.9%, $\leq 0.001$ | 19 |  |
| Permanent                      | 0.78 (–0.07–1.64)    | 82.7%, $\leq 0.001$ | 7  |  |
| Graft sites                    |                      |                     |    |  |
| Global                         | 0.24 (–0.59–1.08)    | 40.1%, 0.196        | 1  | 2.06%, 0.201                           |
| Focal                          | 1.19 (0.78–1.60)     | 75.2%, $\leq 0.001$ | 25 |  |
| Cell donor species             |                      |                     |    | 20.00%, 0.025                          |
| Mouse                          | 1.46 (0.84–2.08)     | 54.7%, 0.031        | 7  |  |
| Rat                            | 1.73 (1.15–2.31)     | 45.7%, 0.075        | 7  |  |
| Human                          | 0.51 (–0.08–1.10)    | 81.0%, $\leq 0.001$ | 11 |  |
| Cell recipient species         |                      |                     |    | –4.82%, 0.761                          |
| Mouse                          | 0.98 (0.40–1.56)     | 31.2%, 0.201        | 5  |  |
| Rat                            | 1.12 (0.64–1.60)     | 79.8%, $\leq 0.001$ | 20 |  |
| Others                         | 1.61 (0.35–2.87)     |                     | 0  |  |
| Immunoreactivity               |                      |                     |    | 15.24%, 0.044                          |
| No                             | 1.52 (1.11–1.93)     | 30.9%, 0.136        | 12 |  |
| Yes                            | 0.76 (0.21–1.31)     | 82.1%, $\leq 0.001$ | 14 |  |
| State of donor cells           |                      |                     |    | 5.14%, 0.115                           |
| PSC-NSPCs                      | 0.63 (–0.13–1.38)    | 84.1%, $\leq 0.001$ | 7  |  |
| WT-NSPCs                       | 1.33 (0.88–1.78)     | 68.2%, $\leq 0.001$ | 19 |  |
| Immunosuppression drugs        |                      |                     |    | –3.64%, 0.657                          |
| No                             | 1.17 (0.22–2.12)     | 78.6%, $\leq 0.001$ | 6  |  |
| Yes                            | 0.95 (0.32–1.58)     | 83.2%, $\leq 0.001$ | 11 |  |
| Unknown                        | 1.28 (0.82–1.74)     | 26.9%, 0.205        | 8  |  |
| Blinding                       |                      |                     |    | –5.59%, 0.758                          |
| No                             | 1.32 (0.17–2.46)     | 85.8%, $\leq 0.001$ | 6  |  |
| Yes                            | 1.06 (0.67–1.46)     | 69.1%, $\leq 0.001$ | 20 |  |
| Randomization                  |                      |                     |    | –3.47%, 0.483                          |
| No                             | 1.33 (0.61–2.05)     | 77.1%, $\leq 0.001$ | 10 |  |
| Yes                            | 0.99 (0.53–1.46)     | 74.6%, $\leq 0.001$ | 16 |  |

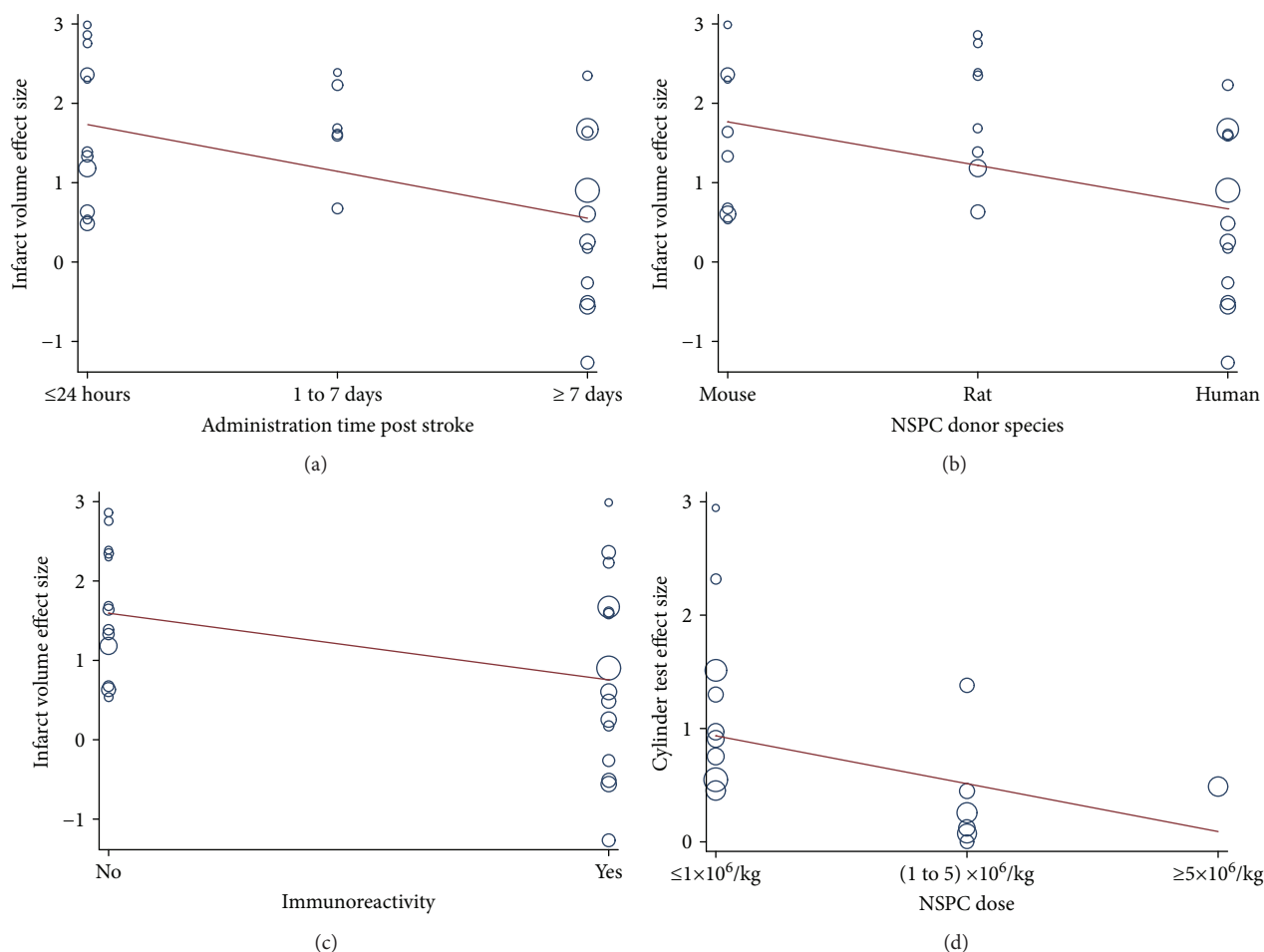


FIGURE 3: Metaregression analysis for related variables and effect sizes of infarct volume reduction and cylinder test. (a) An earlier intraparenchymal NSPC transplantation was associated with a smaller tissue loss ( $\text{Adj } R^2 = 26.92\%$ ,  $p = 0.008$ ). (b) Rodent cell donors were associated with greater residual tissue ( $\text{Adj } R^2 = 20.00\%$ ,  $p = 0.030$ ). (c) Allograft benefited the infarct volume reduction more ( $\text{Adj } R^2 = 15.24\%$ ,  $p = 0.044$ ). (d) A relatively lower NSPC dosage was associated with larger cylinder test performance ( $\text{Adj } R^2 = 100.00\%$ ,  $p = 0.063$ ). Values for effect sizes are Hedges'  $g$ .

randomization design, and study quality. The main mechanism by which intraparenchymal NSPC transplantation benefits ischemic stroke appears to lie in apoptosis inhibition and axonal function.

According to the STAIR guidance for methodological quality estimation [31], a mean quality score of 5.10 in this meta-analysis is higher than previous reports [32, 33]. However, the methodological quality of animal studies should be improved because the overestimation effect of low-quality studies potentially influences the results [34, 35]. After further analysis, we could not find direct proof in study quality score and neurobehavioral or histological effect size as was described in a similar study [36], and publication bias existed in neurobehavioral indices (rotarod and cylinder test), both of which encourage future research with rigorous design.

In this meta-analysis, the overall effect sizes for four considered indices are big, which would suggest an improvement in neurobehavioral and histological outcomes in adult ischemic stroke animals after intraparenchymal NSPC transplantation. However, because of the existence of heterogeneity among studies for mNSS, rotarod test, and infarct volume,

the output results should be interpreted with care. It also should be mentioned that there were several comparisons with an effect size more than 3.0, which contributes to the heterogeneity significantly [7, 17, 33]. After examining these studies, we found that there are some defects in the experimental design, including no blinding, no randomization, nonstatistical animal numbers, low-quality study, nonnormative procedure for assessments without pretraining, and low reliability for gained results. By sensitivity analysis to exclude inadequate comparisons, we obtained more homogenous data to draw a reliable conclusion for effect sizes of mNSS (1.27 SMD), rotarod test (1.63 SMD), and infarct volume.

The parameters used in this meta-analysis, including NSPC administration time, dosage, donor species, and transplantation immunogenicity, may be useful for future clinical application of stem cell therapy. Our metaregression analysis reveals that an earlier NSPC transplantation, that is, at the acute or subacute stage (within a period of 7 days), results in a greater reduction in infarct volume. This is partially consistent with the conclusions of a recent meta-analysis

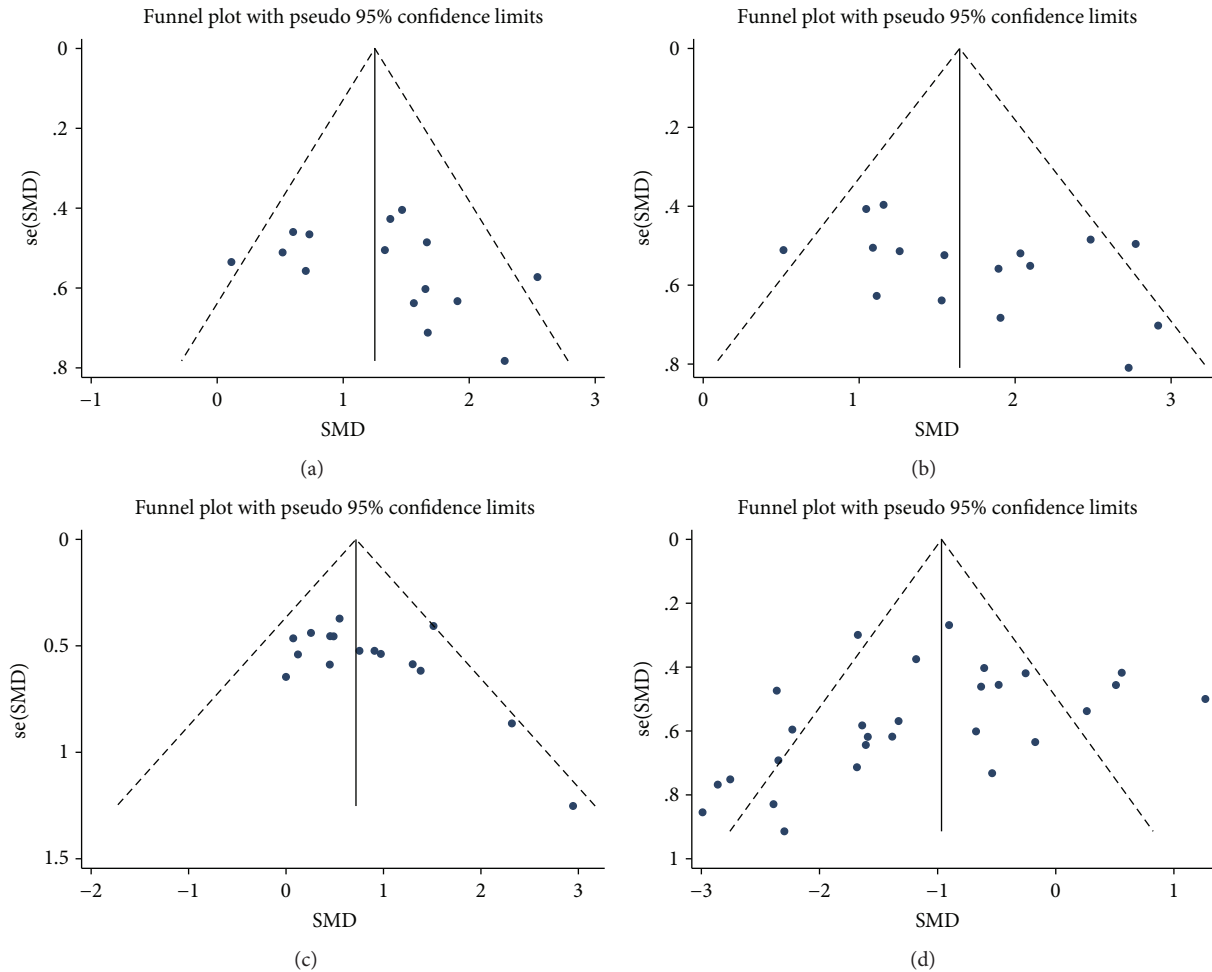


FIGURE 4: Funnel plots for publication bias of (a) mNSS, (b) rotarod test, (c) cylinder test, and (d) infarct volume. SMD: standardized mean difference.

[33]. Compared with the late NSPC transplantation, the early delivery may spare secondary damage to the brain tissue and preserve neural circuits more by neuroprotection effects [37, 38]. Interestingly, we found that a relatively low amount of NSPCs for intraparenchymal transplantation could improve the cylinder test more than the high-dose groups. The effective cell dose is less than  $1 \times 10^6$  cells/kg. This appears to contradict the perceived understanding that a better neurobehavioral outcome results from a higher cell dose. A relatively low dose of NSPCs delivered intraparenchymally could migrate to the lesion area directly without exacerbating the ischemic brain injury by a high dose (volume) of grafted cells and tumor formation. Our subgroup meta-analysis and metaregression analysis also suggest that NSPCs from mouse donors contribute to a larger effect size of infarct volume reduction than those from rats and humans. And a more favorable lesion reduction effect is achieved through allogeneic transplantation rather than xenotransplantation, that is, rodent NSPCs to rodents, consistent with the results of Chen et al. [33] and Yousefifard et al. [39] in NSPC transplantation for spinal cord injury. This is likely related to the immunological reactions. For human NSPC grafts, the transplantation paradigm of

mouse-mouse was within species, with low immune response to host tissue or cell grafts [40]. Finally, another factor that determines clinical translation is tumorigenesis. In our meta-analysis, about half of the included studies have claimed no malignant neoplasm formation from cell grafts. Since most of the NSPCs in the included studies are derived from pluripotent stem cells, this suggests that the cells used for transplantation are a reasonably appropriate source for stroke therapy [41].

The mechanisms underlying stem cell transplantation for ischemic stroke are not clear. Meta-analysis on MSC transplantation for stroke animal studies [7, 36, 42] suggests the involvement of apoptosis inhibition, immunomodulation, neurotrophic factors, angiogenesis, neurogenesis, gliosis reduction, white matter function, enzyme supplementation, and axonal function. We found that apoptosis inhibition is the main contributor to neurobehavioral and histological improvement in intraparenchymal NSPC transplantation. Axonal function also has a significant role in behavioral and histological amelioration. Thus, neuroprotection and cell replacement underlie the therapeutic effect of intraparenchymal NSPC transplantation in ischemic stroke animals.



## 5. Conclusions

This study suggests that intraparenchymal delivery of NSPCs is a promising candidate for ischemic stroke therapy, improving the functional deficits and histopathology in animal models of ischemic stroke. The possible cellular mechanisms associated with intraparenchymal NSPC transplantation include apoptosis inhibition and axonal function.

## Disclosure

Hailong Huang's current address is the Department of Rehabilitation Medicine, Zhongnan Hospital of Wuhan University, Wuhan, 430071, China.

## Conflicts of Interest

All the authors have declared that no conflict of interest exists.

## Authors' Contributions

H.L.H. is responsible for the conception and design, collection and/or assembly of the data, and manuscript writing. K.Q. is responsible for the collection and/or assembly of the data and data analysis and interpretation. X.H.H. is responsible for the data analysis and interpretation. X.L. and Y.F.Z. are responsible for the collection and/or assembly of the data. Z.S.C. is responsible for the administrative support. X.L.H. and H.C. are responsible for the conception and design, financial support, and data analysis and interpretation. H.L.H. and K.Q. contributed equally to this study. And all the authors approved the final version of this manuscript.

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## Supplementary Materials

*Supplementary 1.* Figure S1: effect sizes of all included studies for (a) mNSS, (b) rotarod test, (c) cylinder test, and (d) infarct volume.

*Supplementary 2.* Table S1: PRISMA 2009 checklist.

*Supplementary 3.* Table S2: extracted data associated with cellular mechanisms of NSPC intraparenchymal transplantation in ischemic stroke animals.

*Supplementary 4.* Table S3: characteristics for all included studies.

*Supplementary 5.* Table S4: study quality assessment referred to the CAMARADES checklists.

*Supplementary 6.* Table S5: reasons for excluding related comparisons.

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