

# Quantitative MRI Helps to Detect Hip Ischemia: Preclinical Model of Legg-Calvé-Perthes Disease

Casey P. Johnson, PhD • Luning Wang, PhD • Ferenc Tóth, DVM, PhD • Olumide Aruwajoye, PhD • Cathy S. Carlson, DVM, PhD • Harry K. W. Kim, MD, MSc • Jutta M. Ellermann, MD, PhD

From the Center for Magnetic Resonance Research (C.P.J., L.W., J.M.E.) and Departments of Radiology (C.P.J., L.W., J.M.E.), Veterinary Population Medicine (F.T.), and Veterinary Clinical Sciences (C.S.C.), University of Minnesota, 2021 6th St SE, Minneapolis, MN 55455; Center for Excellence in Hip Disorders, Texas Scottish Rite Hospital, Dallas, Tex (O.A., H.K.W.K.); and Department of Orthopedic Surgery, UT Southwestern Medical Center, Dallas, Tex (H.K.W.K.). Received February 26, 2018; revision requested April 16; revision received May 16; accepted June 4. **Address correspondence to C.P.J.** (e-mail: [casey@cmrr.umn.edu](mailto:casey@cmrr.umn.edu)).

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**Purpose:** To determine whether quantitative MRI relaxation time mapping techniques can help to detect ischemic injury to the developing femoral head.

**Materials and Methods:** For this prospective animal study conducted from November 2015 to February 2018, 10 male 6-week-old piglets underwent an operation to induce complete right femoral head ischemia. Animals were humanely killed at 48 hours ( $n = 2$ ) or 4 weeks ( $n = 8$ ) after the operation, and the operated and contralateral-control femoral heads were harvested and frozen. Thawed specimens were imaged at 9.4-T MRI by using T1, T2, T1 in the rotating frame (T1 $\rho$ ), adiabatic T1 $\rho$ , relaxation along a fictitious field (RAFF), and T2\* mapping and evaluated with histologic analysis. Paired relaxation time differences between the operated and control femoral heads were measured in the secondary ossification center (SOC), epiphyseal cartilage, articular cartilage, and metaphysis and were analyzed by using a paired  $t$  test.

**Results:** In the SOC, T1 $\rho$  and RAFF had the greatest percent increases in the operated versus control femoral heads at both 48 hours (112% and 72%, respectively) and 4 weeks (74% and 70%, respectively). In the epiphyseal and articular cartilage, T2, T1 $\rho$ , and RAFF were similarly increased at both points (range, 24%–49%). At 4 weeks, T2, T1 $\rho$ , adiabatic T1 $\rho$ , and RAFF were increased in the SOC ( $P = .004, .018, <.001, \text{ and } .001$ , respectively), epiphyseal cartilage ( $P = .009, .008, .011, \text{ and } .007$ , respectively), and articular cartilage ( $P = .005, .016, .033, \text{ and } .018$ , respectively). Histologic assessment identified necrosis in SOC and deep layer of the epiphyseal cartilage at both points.

**Conclusion:** T2, T1 in the rotating frame, adiabatic T1 in the rotating frame, and relaxation along a fictitious field maps are sensitive in helping to detect ischemic injury to the developing femoral head.

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Legg-Calvé-Perthes disease (LCPD) is a childhood ischemic hip disorder caused by disruption of blood flow to the growing femoral head, which can lead to severe femoral head deformity and osteoarthritis (1). The disruption of blood flow produces extensive cell necrosis in the marrow and trabecular bone of the secondary ossification center (SOC) and the deep layer of the subarticular epiphyseal cartilage (2). Gradual revascularization of the necrotic epiphysis is accompanied by bone resorption, making the femoral head susceptible to deformation (2). Approximately half of patients with LCPD clinically present when the femoral head deformity is mild, providing a therapeutic window to protect against further deformity (1,3,4). However, clinically standard radiography and conventional morphologic MRI have limited ability to measure the early-stage ischemic injury, which delays treatment decisions (5).

To improve care of patients with LCPD, imaging methods are needed to quantify the severity of ischemic injury to the femoral head. Contrast agent-enhanced perfusion MRI can be used to help measure

the avascular SOC volume and determine whether revascularization has occurred (6–9). However, perfusion MRI does not directly measure SOC necrosis (10), nor does it measure cartilage injury. Furthermore, the long-term consequences of gadolinium deposition in the brain are a potential risk for children, particularly if contrast agents are used serially to monitor disease progression (11). Diffusion-weighted MRI is a complementary technique that can depict injury to the SOC and the surrounding cartilage (12–14), but it has relatively low spatial resolution and is sensitive to distortion and ghosting artifacts caused by magnetic susceptibility differences at air-to-tissue and bone-to-tissue interfaces and by motion (15).

To our knowledge, the role of relaxation time mapping techniques for assessment of bone and marrow ischemia has not been well established in the literature. Conversely, quantitative mapping of T1 in the rotating frame (T1 $\rho$ ) is sensitive in depicting ischemic injury in stroke and myocardial infarction (16–19). Furthermore, quantitative mapping of T2, T1 $\rho$ , adiabatic T1 $\rho$ , and relaxation along a fictitious field (RAFF) are sensitive in helping to detect

## Abbreviations

LCPD = Legg-Calvé-Perthes disease, RAFF = relaxation along a fictitious field, ROI = region of interest, SOC = secondary ossification center, T1 $\rho$  = T1 in the rotating frame

## Summary

Quantitative MR mapping techniques are sensitive in helping to detect ischemic injury to the developing femoral head.

## Implications for Patient Care

- Quantitative MRI of the hip may help to improve early detection and evaluation of ischemic injury, including severity of damage to the developing femoral head, which could lead to better clinical management of Legg-Calvé-Perthes disease and related ischemic bone and joint disorders.
- Quantitative relaxation time mapping provides a possible alternative to contrast-enhanced techniques for measuring the severity of bone and marrow necrosis in pediatric patients with Legg-Calvé-Perthes disease without intravenous administration of gadolinium contrast agents.
- The sensitivity of T1 in the rotating frame to detect necrosis of the secondary ossification center and cartilage can potentially aid in the development and evaluation of therapies for Legg-Calvé-Perthes disease.

ischemia-induced epiphyseal cartilage necrosis (20–22). T1 $\rho$  has also been shown to depict early articular cartilage damage overlying necrotic bone in adult patients with collapsed femoral head avascular necrosis (23). Thus, T1 $\rho$  and other relaxation times may be useful to help detect early ischemic injury to the bone, marrow, and cartilage of the growing femoral head. T1 $\rho$  and the related rotating frame methods, adiabatic T1 $\rho$  and RAFF, are of interest because they are sensitive to molecular interactions that are not detectable by using laboratory frame approaches (24). Whereas conventional T1, T2, and T2\* are sensitive to fast molecular interactions (on the order of megahertz, corresponding to the Larmor frequency), rotating frame methods use spin-lock radiofrequency pulses (ie, small magnetic fields) to probe slower molecular interactions (on the order of 10 Hz to 10 kHz) such as chemical exchange (24). Each rotating frame method probes a different range of slow molecular interactions (25), thus each may have a different sensitivity to tissue changes in response to ischemic injury.

The purpose of our study was to determine whether quantitative relaxation time mapping techniques can help to identify ischemic injury to the developing femoral head. Six relaxation times (T1, T2, T1 $\rho$ , adiabatic T1 $\rho$ , RAFF, and T2\*) were evaluated in a piglet model of ischemic osteonecrosis of the femoral head. We hypothesized that the relaxation time measurements can help to detect ischemic injury to the SOC and the subarticular epiphyseal cartilage as early as 48 hours after induction of ischemia.

## Materials and Methods

### Animals

Our prospective animal study was conducted with an institutional animal care and use committee–approved protocol from November 2015 to February 2018. Ten 5-week-old male

piglets were obtained from a commercial provider (Change of Pace, Aubrey, Tex). At 6 weeks old, the piglets underwent an operation on their right hips to induce ischemia of the entire femoral head by placing a tight ligature about the femoral neck and transecting the ligamentum teres femoris (26,27). Operations were performed by a board-certified pediatric orthopedic surgeon (H.K.W.K.) with 20 years of experience working with the model. The piglets were humanely killed at either 48 hours ( $n = 2$ ) or 4 weeks ( $n = 8$ ) following the operation. After they were killed, the right (operated) and left (contralateral control) femoral heads were surgically removed and immediately placed in a  $-20^{\circ}\text{C}$  freezer for storage until imaging at a later time. We focused on the 4-week period because this corresponds radiographically and histologically to a typical avascular stage at which children present with LCPD before onset of the reparative process (3,27,28). However, it is also of interest to study whether early ischemic damage is detectable. Thus, we also collected data at an acute point (48 hours after the operation). Only male piglets were studied to reduce experimental variability and because LCPD is five times more common in boys than in girls (29).

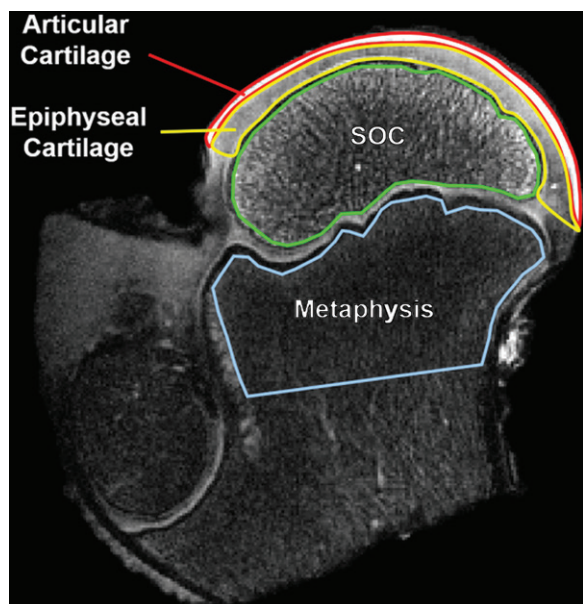
### MR Image Acquisition

The femoral head specimens were imaged individually by using a preclinical 31-cm-bore 9.4-T MRI system (Agilent Technologies, Santa Clara, Calif) equipped with a Varian console with VnmrJ software (Varian NMR Systems, Palo Alto, Calif). Imaging was performed by two MRI scientists (C.P.J. and L.W., with 12 and 10 years of experience, respectively). Immediately before imaging, the specimens were thawed at room temperature, secured with a holder to control positioning in the imager, and immersed in Fomblin (Specialty Fluids Company, Castaic, Calif) to reduce magnetic susceptibility–related artifacts. The specimens were placed in a Varian millipede radiofrequency coil (Varian NMR Systems), which was manually tuned and matched for each specimen. After the acquisition of scout images and manual second-order  $B_0$  shimming, images for quantitative relaxation time mapping were acquired. Each specimen was imaged for approximately 3 hours to provide high ( $0.16 \times 0.16 \text{ mm}^2$ ) in-plane spatial resolution. Imaging parameters are in Table 1. T1-weighted, T2-weighted, T1 $\rho$ -weighted, adiabatic T1 $\rho$ -weighted, and RAFF images for quantitative mapping were each acquired by using the same single-section two-dimensional fast-spin-echo sequence with different magnetization preparation blocks (Table 1). T2\*-weighted images were acquired by using a two-dimensional multiecho gradient-echo sequence on the same plane as the two-dimensional fast-spin-echo images. T2\* maps were not acquired for two of the 4-week postoperative femoral head pairs. Fat suppression was not applied in any imaging sequence. The imaging section used for quantitative mapping was oriented so that it bisected the femoral head while passing through the apex of the greater trochanter and 1.0 mm anterior to the insertion point of the ligamentum teres femoris. By using these anatomic landmarks, in combination with a holder to keep the femoral head in a fixed posi-

**Table 1: Quantitative Relaxation Time Mapping Imaging Parameters**

Imaging Parameter	2D FSE	2D GRE
Field of view (mm <sup>2</sup> )	40 × 40	40 × 40
Sampling matrix	256 × 256	256 × 256
Resolution (mm <sup>2</sup> )	0.16 × 0.16	0.16 × 0.16
Section thickness (mm)	1.0	1.0
Repetition time/echo time (msec)	5000/5.1	30/5, 8, 11, 14, 17, 20, 25
Flip angle (degrees)	90/180	14
Bandwidth (Hz/px)	513	195
Echo train length	8	1
No. of signal averages	1	10
Imaging time (min:sec)	2:40	9:58
T1 map inversion times (msec)	200, 500, 800, 1100, 1400, 3000	NA
T2 map echo times (msec)	4, 20, 40, 60, 80, 100	NA
T1ρ map pulse type	Constant amplitude on-resonance spin lock pulse with $\gamma B_1$ of 1250 Hz	NA
T1ρ map TSLs (msec)	0, 24, 48, 96, 192	NA
Adiabatic T1ρ pulse type	Train of HS4 adiabatic pulses with duration of 6 msec and $\gamma B_{1,max}$ of 1250 Hz	NA
Adiabatic T1ρ map TSLs (msec)	0, 24, 48, 72, 96, 144	NA
RAFF pulse type	Train of RAFF2, pulses with duration of 4.53 msec, $\gamma B_{1,max}$ of 625 Hz, and phase cycling	NA
RAFF map TSLs (msec)	0, 36.2, 72.4, 108.6, 144.8	NA

Note.—2D = two-dimensional, FSE = fast spin echo, GRE = gradient-recalled echo, NA = not applicable, RAFF = relaxation along a fictitious field, T1ρ = T1 in the rotating frame, TSL = spin-lock time.



**Figure 1:** Region of interest (ROI) definitions for quantitative analysis of the relaxation time maps. A T2-weighted image of an operated femoral head 48 hours after induction of ischemia in a 6-week-old piglet is shown with the four ROIs outlined. In the growing femoral head, the vascularized subarticular epiphyseal cartilage is gradually replaced by the secondary ossification center (SOC) until the ossification front reaches the overlying articular cartilage. These three ROIs are potentially affected by the surgical induction of ischemia. The fourth ROI in the metaphysis was included in the analysis as an internal control as its blood supply is not altered by the surgical procedure.

tion, the imaging plane was co-registered for 9.4-T MRI and histologic analysis correlation.

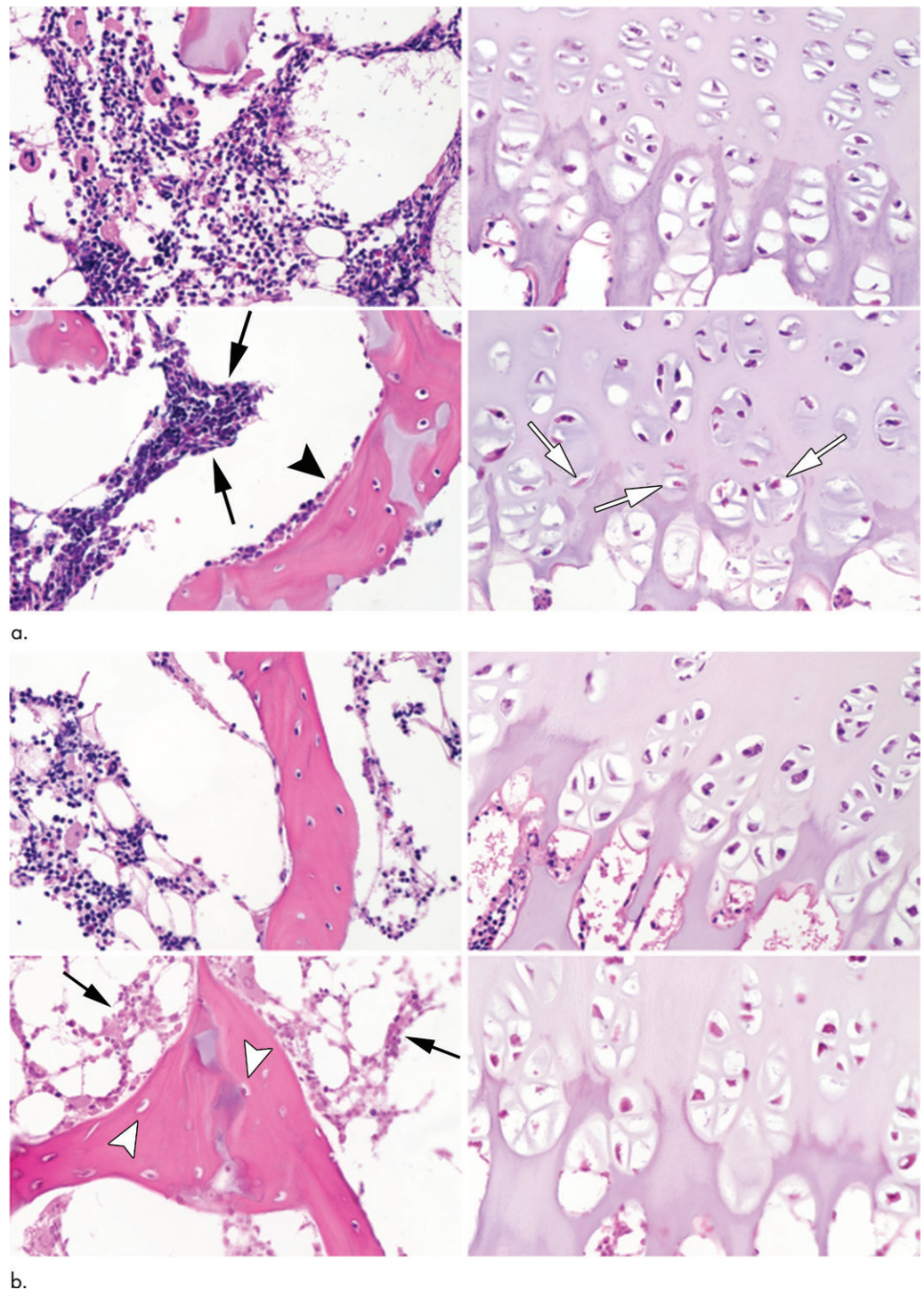
### Histologic Evaluation

Immediately after MRI, the femoral heads were bisected along the imaging plane with a high-speed bone saw (TechCut 5; Allied High Tech Products, Compton, Calif) and fixed in 10% neutral buffered formalin. Specimens were then decalcified by using 10% ethylenediaminetetraacetic acid and routinely processed into paraffin blocks for histologic sectioning. We stained 5- $\mu$ m-thick serial slices with hematoxylin-eosin and they were qualitatively evaluated to assess the morphologic structure of the SOC and overlying cartilage. Histologic evaluation was performed by a board-certified orthopedic veterinary pathologist (C.S.C., with 30 years of experience) who was blinded to treatment.

### Data Analysis

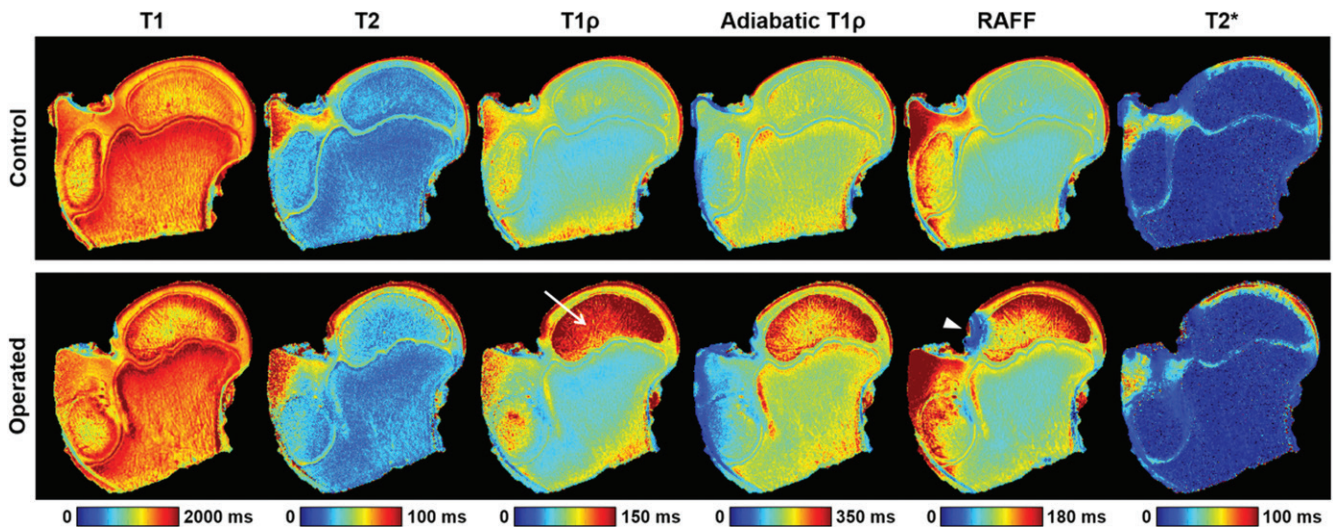
MRI postprocessing was performed by using Matlab (version R2013b; MathWorks, Natick, Mass). Quantitative relaxation time maps were generated by using a monoexponential fitting algorithm. Three regions of interest (ROIs) were manually drawn in the epiphysis for all operated and control femoral head images to encompass the SOC, the subarticular epiphyseal cartilage (ie, growth cartilage) overlying the SOC, and the overlying articular cartilage (Fig 1). A fourth ROI was also drawn in the metaphysis as an internal control because its intraosseous blood supply was unaltered by the surgical procedure (unlike the separate blood supply of the epiphysis via penetrating extraosseous blood vessels) (30). The ROIs were

**Figure 2:** Representative photomicrographs of histologic slices of the secondary ossification center (left-hand side in **a** and **b**) and epiphyseal cartilage (right-hand side in **a** and **b**) from paired control and operated femoral heads stained with hematoxylin-eosin at 40 $\times$  magnification. Images at the top of **a** and **b** are from control piglets and images at the bottom of **a** and **b** are from the piglets who underwent operation. (**a**) Images in a 6-week-old piglet 48 hours after the induction of ischemia. Hematopoietic cells in the bone marrow exhibited clumping and loss of distinctly identifiable cell margins in the operated femoral heads (black arrows). There was evidence of osteoblast necrosis (arrowhead). Many chondrocytes in the deep layer of the epiphyseal cartilage were eosinophilic and lacked distinct nuclei (white arrows). (**b**) Images in a second 6-week-old piglet 4 weeks after induction of ischemia. The bone marrow cells were diffusely necrotic, characterized by loss of nuclear staining and diffusely eosinophilic cytoplasm (arrows). There was also extensive evidence of osteocyte necrosis (loss of nuclear staining or the presence of empty osteocyte lacunae) in the trabecular bone (arrowheads). Diffuse chondronecrosis remained limited primarily to the deep layer of the epiphyseal cartilage.



defined by using T2-weighted images acquired as part of the T2 mapping sequence, which provided comparison between the epiphyseal cartilage and the overlying higher-intensity articular cartilage. Areas distorted by field inhomogeneity artifacts were excluded. The median relaxation time within each ROI was measured, and paired values for the operated and contralateral control femoral heads were compared. The median value was chosen because it is robust to outliers. For the 4-week postsurgical specimens, the paired values were statistically compared by using R statistical software (version

3.3.1; R Foundation for Statistical Computing, Vienna, Austria) with a parametric two-tailed paired *t* test. Two statistical significance thresholds were defined. We defined *P* values less than .002 as indicative of definitive significance with a type I error rate ( $\alpha = .05$ ) after conservative Bonferroni correction for 24 comparisons (six relaxation times  $\times$  four ROIs). Given the likelihood of measurements being correlated across both relaxation times and ROIs, we defined findings with *P* value less than .05 (and thus a minimal type II error rate) as hypothesis generating. A nonparametric Wilcoxon signed



**Figure 3:** Quantitative relaxation time maps acquired for one pair of operated and contralateral control femoral heads 48 hours after the induction of ischemia in a 6-week-old piglet. The operated femoral head had increased relaxation times in the secondary ossification center, epiphyseal cartilage, and articular cartilage compared with the control specimen. T1 in the rotating frame (*T1 $\rho$* ) relaxation times were markedly increased in the secondary ossification center (arrow). The metaphysis had similar relaxation times in both specimens, as expected. The relaxation along a fictitious field (*RAFF*) map for the operated femoral head had a region of susceptibility artifact because of the presence of a nearby air bubble (arrowhead).

rank test was also performed for comparison. Additionally, mean percent differences between the median relaxation time values for the paired operated and control femoral heads were calculated and compared for specimens with induced ischemia at postoperative 48 hours and 4 weeks.

## Results

Histologic analysis revealed that the femoral heads with induced ischemia at 48 hours after operation showed subtle evidence of necrosis (Fig 2a). Cells in the bone marrow of the SOC had reduced clarity of cytoplasmic and nuclear details, exhibited clumping, and were admixed with fibrillar material. Diffusely eosinophilic chondrocytes were present in the deep layer of the epiphyseal cartilage, indicative of cartilage necrosis. Seven of the eight femoral heads removed 4 weeks after operation showed evidence of extensive necrosis in the SOC (pyknosis and increased eosinophilia of nuclei, and cell shrinkage) and chondronecrosis in the deep layer of the epiphyseal cartilage (Fig 2b). There was no evidence of osteonecrosis or chondronecrosis in the femoral head control specimens. The histologic appearance of the SOC and cartilage of the eighth specimen removed 4 weeks after operation was similar to the contralateral (control specimen) femoral head from the same animal, which indicated a failure of the operation to induce ischemic injury. Thus, this was designated as a sham control specimen and was excluded from the statistical analyses. Additionally, one of the femoral heads obtained at 4 weeks after operation was categorically different than the others; the femoral head showed advanced deformation so that the SOC contained only necrotic tissue debris that lacked definition of tissue or bone structure. Unlike the other femoral heads that had changes resembling early-stage LCPD, the severe collapse of this latter femoral head represented the late fragmentation stage of the disease that is identifiable on radiographs and for which the

proposed MRI methodologic analysis is unnecessary. Thus, we also excluded this femoral head from the statistical analysis, leaving a group of six femoral head pairs at the 4-week point.

Despite the subtle appearance of histologic findings of tissue necrosis in the two femoral heads obtained at 48 hours after operation, relaxation times were increased in the SOC, epiphyseal cartilage, and articular cartilage for both specimens. Quantitative relaxation time maps for one of the operated femoral heads and its control specimen are shown in Figure 3, which demonstrate a pronounced increase in relaxation times, particularly T1 $\rho$ , in the SOC, epiphyseal cartilage, and articular cartilage. For comparison, conventional proton density- and T2-weighted images are shown in Figure E1 (online). Table 2 shows the average relaxation time percent increase in each of the four ROIs between the pairs of operated and control femoral heads. These values are also plotted in Figure E2 (online). T1 $\rho$  and RAFF relaxation times showed the greatest percent increase in the SOC 48 hours after induction of ischemia (112% and 72%, respectively). In the epiphyseal and articular cartilage, the greatest relaxation time increases were observed at T2 (increases of 24% and 32%, respectively), T1 $\rho$  (increases of 25% and 28%, respectively), adiabatic T1 $\rho$  (increases of 25% and 13%, respectively), and RAFF (increases of 25% and 28%, respectively). The internal control ROI in the metaphysis showed no notable changes in relaxation times between the operated and control femoral heads. Table 3 shows numerical data from the ROI analysis that reflect the percent increase findings.

The six 4-week postoperative femoral heads that exhibited extensive osteonecrosis in the SOC and chondronecrosis in the deep layer of the epiphyseal cartilage showed increased relaxation times in the SOC, epiphyseal cartilage, and articular cartilage compared with their control specimens. The average relaxation time percent increase in each of the four ROIs between the pairs of operated and control femoral heads are shown in Table 2

and Figure E2 (online). In the SOC, T1 $\rho$  and RAFF showed the greatest percent increases (74% and 70%, respectively). In the epiphyseal and articular cartilage, T2 (49% and 28%), T1 $\rho$  (38% and 25%), and RAFF (45% and 25%) had the greatest percentage increase. However, the relaxation times exhibited by the sham-operated femoral head were similar to those of the control specimens; the percent increases in T1 $\rho$  were 1.8%, 4.9%, 11.6%, and 1.4% for the SOC, epiphyseal cartilage, articular cartilage, and metaphysis, respectively. Detailed quantitative analyses of the ROI data are shown in Table 4. T1, T2, T1 $\rho$ , adiabatic T1 $\rho$ , and RAFF were all increased in the SOC ( $P = .040, .004, .018, <.001, \text{ and } .001$ , respectively) and epiphyseal cartilage ( $P = .034, .009, .008, .011, \text{ and } .007$ , respectively) in the operated femoral heads compared with the paired control specimens. Furthermore, T2, T1 $\rho$ , adiabatic T1 $\rho$ , and RAFF were all increased in the articular cartilage ( $P = .005, .016, .033, \text{ and } .018$ , respectively). T1 was not altered in the articular cartilage ROI ( $P = .24$ ) and T2\* was not altered in any of the ROIs ( $P = .09, .08, \text{ and } .06$  in the SOC, epiphyseal cartilage, and articular cartilage, respectively). None of the relaxation times was altered in the metaphysis ( $P = .49, .99, .46, .90, .72, \text{ and } .89$  for T1, T2, T1 $\rho$ , adiabatic T1 $\rho$ , RAFF, and T2\*, respectively). The statistical findings were consistent with the nonparametric test.

One of the 4-week postoperative specimens had increased relaxation times only in a well-circumscribed focal area of the SOC. Histologic assessment of this specimen confirmed that only this focal area of the SOC was necrotic, whereas the remaining SOC had undergone revascularization and new bone formation (Fig 4).

## Discussion

Quantitative MRI relaxation time mapping is a potential alternative approach to measure ischemic injury to the SOC and cartilage of the femoral head with high spatial resolution. Our findings support the hypothesis that quantitative relaxation time maps are sensitive in helping to detect ischemic injury to the developing femoral head. In particular, T1 $\rho$  and RAFF had the greatest percent increases in the SOC after injury at both 48 hours (112% and 72%, respectively) and 4 weeks (74% and 70%, respectively). T2, T1 $\rho$ , adiabatic T1 $\rho$ , and RAFF were also increased in the epiphyseal cartilage and articular cartilage at 48 hours and 4 weeks after the onset of ischemia. Our findings provide a basis that would support further investigations of MRI quantitative mapping techniques in children suspected of having or who have been diagnosed with LCPD.

The sharp increase in T1 $\rho$  and RAFF relaxation times in the SOC 48 hours after disruption of blood flow to the femoral head suggests that these methods are sensitive for detection of ischemic necrosis of the bone marrow. Per histologic analysis, the ischemic SOC had only mild changes in the marrow at 48 hours, which was consistent with a previous study (31). Our MRI findings were consistent with studies that showed that T1 $\rho$  is particularly sensitive in helping to detect ischemic injury to the brain and myocardium (16–19). In these studies, ischemia-induced increases in T1 $\rho$  were attributed to increased water uptake by affected cells and altered chemical exchange of labile

**Table 2: Average Increase in Relaxation Time**

ROI	Increase at 48 Hours (%)	Increase at 4 Weeks (%)
<b>SOC</b>		
T1	11 ± 11	16 ± 15
T2	32 ± 8	36 ± 20
T1 $\rho$	112 ± 10	74 ± 65
aT1 $\rho$	47 ± 3	45 ± 31
RAFF	72 ± 4	70 ± 41
T2*	11 ± 2	19 ± 15*
<b>Subarticular epiphyseal cartilage</b>		
T1	1 ± 3	12 ± 11
T2	24 ± 12	49 ± 34
T1 $\rho$	25 ± 3	38 ± 22
aT1 $\rho$	25 ± 0	35 ± 23
RAFF	25 ± 18	45 ± 27
T2*	4 ± 17	29 ± 21*
<b>Articular cartilage</b>		
T1	3 ± 3	7 ± 13
T2	32 ± 9	27 ± 17
T1 $\rho$	29 ± 1	25 ± 19
aT1 $\rho$	13 ± 6	20 ± 20
RAFF	29 ± 13	25 ± 20
T2*	-11 ± 10	24 ± 15*
<b>Metaphysis</b>		
T1	1 ± 0	2 ± 6
T2	2 ± 4	0 ± 10
T1 $\rho$	-2 ± 3	3 ± 10
aT1 $\rho$	-2 ± 7	1 ± 14
RAFF	-3 ± 12	3 ± 17
T2*	0 ± 3	0 ± 6*

Note.—Data are between paired operated and contralateral control femoral heads at 48 hours ( $n = 2$ ) and 4 weeks ( $n = 6$ ) after the induction of ischemia. Data are means  $\pm$  standard deviation. aT1 $\rho$  = adiabatic T1 $\rho$ , RAFF = relaxation along a fictitious field, ROI = region of interest, SOC = secondary ossification center, T1 $\rho$  = T1 in the rotating frame.

\* At the 4-week point, four T2\* maps were acquired (vs six).

protons between free water and macromolecules. Similar effects may be driving the relaxation time changes in the ischemic SOC. Our finding that T1 was relatively insensitive in helping to detect the SOC injury suggests that water content was not the dominant source of the relaxation time increases. The insensitivity of T2\* was likely because it is dominated by the magnetic susceptibility of the trabecular bone, which was not expected to be affected acutely by ischemia. The higher sensitivity of the spin-lock-based techniques compared with T2 was likely driven by the contributions of slow motional processes such as chemical exchange. Furthermore, the differences between T1 $\rho$ , adiabatic T1 $\rho$ , and RAFF may relate to the precise motional frequencies that were probed by each. However, the biologic correlates of these slow motional frequencies in the SOC are unknown. Intracellular changes that occur as a result of ischemia, including water uptake, decreased pH level, and macromolecular

changes (32), along with potential concomitant alterations to the interface between marrow and trabecular bone (33), may collectively contribute to the relaxation time increases. Our findings warrant further investigation of the acute effects of ischemia on SOC relaxation times to determine the primary biologic drivers of their sensitivities.

The sustained increase in SOC relaxation times 4 weeks after the onset of ischemia supports the hypothesis that quantitative mapping techniques can be used to determine the severity of ischemic injury to the SOC in early-stage LCPD and to help detect the early stages of repair. At the 4-week point, changes to the SOC were more variable as tissue repair changes in the SOC began to occur, including partial revascularization of the SOC, infiltration by fibrovascular tissue, clearance of necrotic marrow cells, bone resorption, and new bone formation (2). Our results suggest that relaxation times are increased only in the portion of the SOC that is necrotic and that the relaxation times are decreased as the necrotic tissue is replaced by viable repair tissue. The reduction of relaxation times in the partially-repaired femoral head may be attributed to the infiltration of dense fibrovascular tissue into the necrotic marrow space and the formation of new bone. Consistent with the 48-hour point, T1 and T2\* were relatively insensitive to changes in the SOC, which suggests that similar mechanisms are driving the increase in relaxation times throughout the ischemic stage of the disease. Further study is needed at additional points to determine how relaxation times respond to the dynamics of ischemic osteonecrosis, deformation, and repair of the femoral head.

The increase in relaxation times throughout the epiphyseal and articular cartilage of the injured femoral heads at both 48 hours and 4 weeks after induction of ischemia suggests that the cartilage composition was altered. Consistent with previous reports (26,34,35), histologic changes to the cartilage were primarily noted in the deep layer of the epiphyseal cartilage. However, our global MRI findings suggest that there were changes

**Table 3: Relaxation Time Differences between Paired Operated and Contralateral-Control Femoral Heads 48 Hours after Induction of Ischemia**

ROI	Control Hips (msec)	Operated Hips (msec)	Paired Difference (msec)
<b>SOC</b>			
T1	1570 ± 180	1760 ± 370	190 ± 190
T2	27.6 ± 0.9	36.5 ± 3.5	9.0 ± 2.6
T1ρ	66.7 ± 0.8	141.6 ± 5.2	74.9 ± 6.0
aT1ρ	200 ± 23	295 ± 39	94 ± 16
RAFF	90 ± 13	154 ± 19	64.4 ± 5.8
T2*	5.0 ± 0.3	5.6 ± 0.2	0.6 ± 0.1
<b>Subarticular epiphyseal cartilage</b>			
T1	1440 ± 95	1452 ± 56	12 ± 39
T2	48.1 ± 5.5	59.1 ± 0.8	11.0 ± 4.7
T1ρ	75.7 ± 2.9	94.7 ± 1.0	19.1 ± 1.9
aT1ρ	166.5 ± 3.8	207.5 ± 5.4	41.0 ± 1.6
RAFF	101 ± 14	124.6 ± 0.1	24 ± 15
T2*	39.6 ± 6.5	40.8 ± 0.2	1.2 ± 6.3
<b>Articular cartilage</b>			
T1	1921 ± 73	1971 ± 14	50 ± 59
T2	83.4 ± 0	109.9 ± 7.2	26.5 ± 7.2
T1ρ	115.7 ± 4.2	148.6 ± 4.2	33.0 ± 0.1
aT1ρ	296 ± 17	334.7 ± 2.6	38 ± 14
RAFF	162.6 ± 9.2	208.4 ± 9.5	456 ± 19
T2*	72.4 ± 2.1	64.5 ± 9.4	-7.9 ± 7.4
<b>Metaphysis</b>			
T1	1730 ± 170	1740 ± 170	12.9 ± 0.1
T2	25.7 ± 2.4	26.3 ± 1.5	0.6 ± 0.9
T1ρ	60.8 ± 3.3	59.4 ± 1.1	-1.4 ± 2.2
aT1ρ	200 ± 24	195 ± 10	-5 ± 14
RAFF	86 ± 16	82.4 ± 4.9	-4 ± 11
T2*	5.1 ± 0.4	5.1 ± 0.3	0 ± 0.1

Note.—Data are means ± standard deviation. aT1ρ = adiabatic T1ρ, RAFF = relaxation along a fictitious field, ROI = region of interest, SOC = secondary ossification center, T1ρ = T1 in the rotating frame.

to the cartilage matrix and/or intracellular composition that were undetected by the histologic analyses. It is well established that T2 and T1ρ can help to detect early degenerative changes in articular cartilage, such as increased water content, changes in collagen content and orientation, and loss of proteoglycans (36). It has also been shown (20–22) that T2, T1ρ, adiabatic T1ρ, and RAFF can help to detect focal cartilage lesions created by surgical induction of ischemia in epiphyseal cartilage. Our findings further support that T2, T1ρ, adiabatic T1ρ, and RAFF can help to detect early ischemic injury to the cartilage. Similar to the SOC, we found that T1 was relatively insensitive to ischemia-induced changes in the cartilage, which suggests that water content is not the dominant signal source. In terms of percent change, T2\* was equivalent to the other transverse relaxation times at the 4-week point; therefore, the lack of statistical findings may be because of the greater sensitivity to deviations in the magnetic field on the part of the technique. Collectively, our findings may be clinically significant because early detection will provide a more comprehensive assessment of ischemic damage to the growing femoral head than is currently possible by using routine clinical imaging methods.

**Table 4: Relaxation Time Differences between Paired Operated and Contralateral-Control Femoral Heads 4 Weeks after Induction of Ischemia**

ROI	Control Hips (msec)	Operated Hips (msec)	Paired Difference (msec)	No. of Specimens	<i>t</i> Value	<i>P</i> Value
<b>SOC</b>						
T1	1380 ± 220	1590 ± 230	210 ± 180 (10, 400)	6	2.76	.040*
T2	27.1 ± 2.8	36.5 ± 4.2	9.4 ± 4.6 (4.5, 14.3)	6	4.94	.004*
T1ρ	76 ± 12	129 ± 34	52 ± 37 (14, 91)	6	3.49	.018*
aT1ρ	193 ± 46	270 ± 28	76 ± 27 (48, 105)	6	6.99	<.001†
RAFF	80 ± 17	132 ± 19		6	6.51	.001†
T2*	5.0 ± 0.5	5.9 ± 0.4	0.9 ± 0.7 (−0.3, 2.1)	4	2.45	.09
<b>Subarticular epiphyseal cartilage</b>						
T1	1360 ± 56	1520 ± 110	160 ± 140 (20, 310)	6	2.90	.034*
T2	38.0 ± 2.5	56.2 ± 9.2	18 ± 11 (7, 30)	6	4.10	.009*
T1ρ	66.6 ± 3.0	92 ± 14	25 ± 14 (10, 40)	6	4.31	.008*
aT1ρ	154.4 ± 8.7	207 ± 30	52 ± 32 (18, 87)	6	3.97	.011*
RAFF	80.9 ± 6.2	117 ± 17	36 ± 20 (15, 56)	6	4.43	.007*
T2*	29.6 ± 2.8	38.5 ± 9.1	8.8 ± 6.8 (−2.0, 19.7)	4	2.58	.08
<b>Articular cartilage</b>						
T1	1780 ± 160	1890 ± 150	110 ± 200 (−100, 320)	6	1.35	.24
T2	66.4 ± 5.3	84.2 ± 8.4	17.3 ± 9.3 (8.0, 27.6)	6	4.69	.005*
T1ρ	95.8 ± 6.2	120 ± 14	23 ± 16 (6, 40)	6	3.56	.016*
aT1ρ	238 ± 27	283 ± 32	45 ± 38 (6, 85)	6	2.93	.033*
RAFF	128 ± 14	159 ± 23	31 ± 22 (8, 54)	6	3.44	.018*
T2*	52.8 ± 8.1	65 ± 14	12.6 ± 8.8 (−1.4, 26.6)	4	2.86	.06
<b>Metaphysis</b>						
T1	1560 ± 130	1590 ± 110	28 ± 93 (−69, 125)	6	0.74	.49
T2	25.4 ± 2.5	25.4 ± 4.0	0 ± 2.7 (−2.8, 2.9)	6	0.02	.99
T1ρ	62.1 ± 6.0	64 ± 10	2.1 ± 6.6 (−4.8, 9.0)	6	0.79	.46
aT1ρ	191 ± 30	192 ± 32	1 ± 234 (−24, 26)	6	0.13	.90
RAFF	79 ± 11	81 ± 15	2 ± 13 (−12, 16)	6	0.72	.72
T2*	5.2 ± 0.1	5.2 ± 0.4	0 ± 0.3 (−0.6, 0.5)	4	−0.15	.89

Note.—Data are means ± standard deviation unless otherwise noted; data in parentheses are 95% confidence intervals. aT1ρ = adiabatic T1ρ, RAFF = relaxation along a fictitious field, ROI = region of interest, SOC = secondary ossification center, T1ρ = T1 in the rotating frame.

\* Hypothesis generating ( $P < .05$ ).

† Definitely significant after Bonferroni correction ( $P < .002$ ).

Our study had limitations. First, imaging was performed ex vivo and specimens underwent freezing. Whereas freezing is known to cause an increase in articular cartilage relaxation times (37), the effect of freezing on relaxation times in the epiphyseal cartilage and bone marrow is unknown. In vivo imaging investigations of the piglet model would enable study of the effects of ischemia on the relaxation times at normal physiologic conditions. Second, we used a cross-sectional study design and included a limited sample size, and therefore the findings may not represent the range of the entire population. Furthermore, the effects of ischemia were more variable at later points, which is a source of statistical uncertainty. Additional studies are needed to characterize the longitudinal progression and variability of the femoral head relaxation times. Third, because of limited sample size and the large number of comparisons performed, the statistical analysis was primarily useful for hypothesis generation rather than definitive findings. Future studies can be designed to

focus on the most sensitive relaxation times and relevant ROIs. Fourth, a single two-dimensional section was acquired for quantitative mapping. Although this enabled high-spatial-resolution comparison with histologic analysis, it is of clinical interest to assess ischemic damage across the entire femoral head. Fifth, temperature was not controlled during the experiments, which could be a potential confound because of radiofrequency heating, particularly during the T1ρ sequence. However, the experiment consistently used the same imaging protocol, and therefore any temperature change would similarly affect all specimens. Sixth, we did not investigate the effect of lipid signals on the SOC relaxation times. It would be of interest to investigate how changes in lipid content with ischemia and aging affect the relaxation times. Finally, further study is needed to translate our findings to clinical 1.5-T and/or 3-T MR imagers, including optimization of pulse sequences to comply with radiofrequency power deposition limits and investigations at these lower field



strengths because rotating frame methods can have greater sensitivity to certain slow molecular interactions at higher fields (38).

In conclusion, T2, T1 $\rho$ , adiabatic T1 $\rho$ , and RAFF maps are sensitive in helping to detect ischemic injury to the developing femoral head. Regarding practical applications, quantitative relaxation time mapping techniques are worth investigating for detection and quantification of the severity of ischemic injury to the femoral head in children with early-stage LCPD and related ischemic bone and joint disorders.

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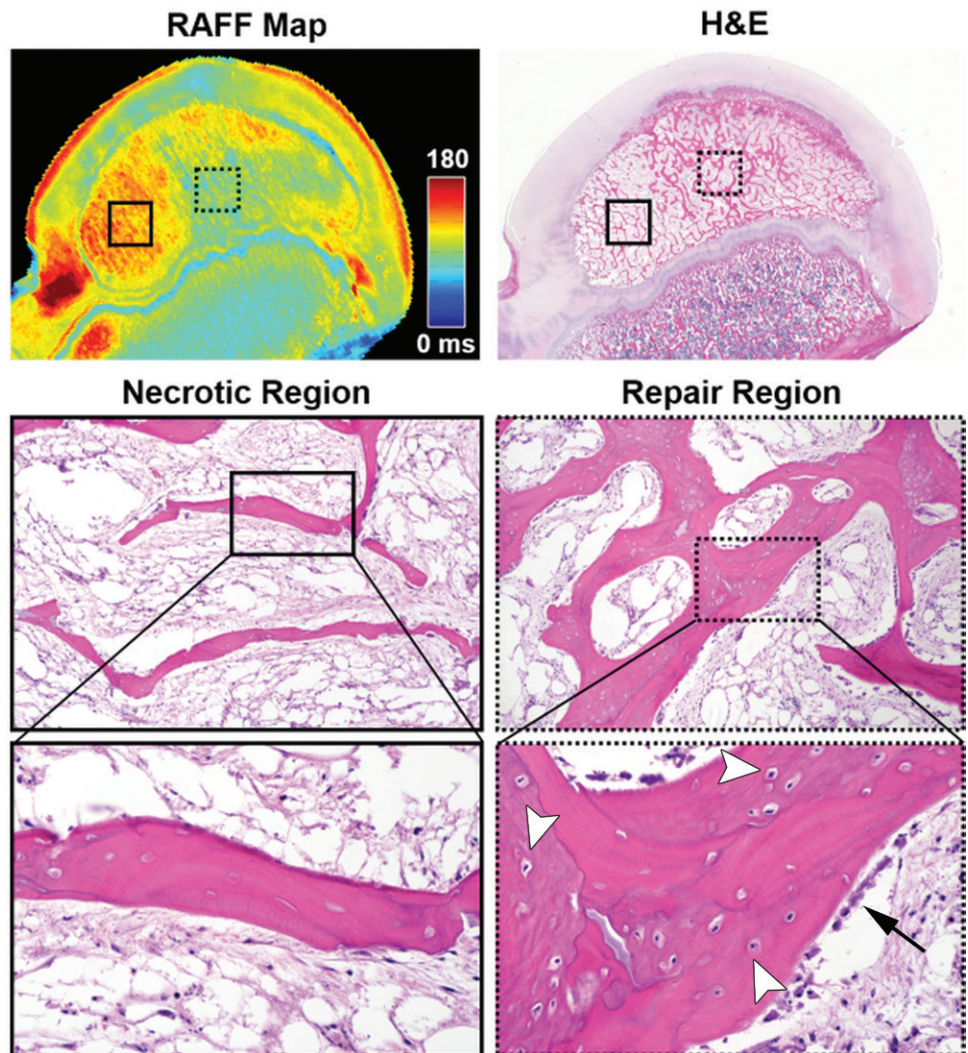
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**Figure 4:** Quantitative relaxation time mapping distinguished regions of secondary ossification center (SOC) necrosis and repair (validated at histologic analysis) in an operated femoral head 4 weeks after the induction of ischemia in a 6-week-old piglet. On a relaxation along a fictitious field (RAFF) map in the operated femoral head, an area of increased relaxation time in the lateral aspect of the SOC (red; solid box) corresponded to a region of diffuse bone necrosis (empty osteocyte lacunae and lack of viable surface osteoblasts) and marrow necrosis observed at histologic analysis (visible on the necrotic region images). An adjacent area of normalized relaxation times in the more medial aspect of the SOC (green; dotted box on the RAFF map) corresponded to a region with areas of new bone. In the repair region, viable osteoblasts (arrow) can be seen on the surface of new or woven bone containing basophilic (viable) osteocytes within lacunae (arrowheads); these areas of new bone surround an area of preexisting, necrotic lamellar bone containing empty osteocyte lacunae. Photomicrographs of the histologic sections stained with hematoxylin-eosin (H&E) were taken at 0.5 $\times$  (top), 10 $\times$  (middle), and 40 $\times$  (bottom) magnification.

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