TRPV1 **and** *BDKRB2* **receptor polymorphisms can influence the exercise pressor reflex**

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Key points

- The mechanisms responsible for the high inter-individual variability in blood pressure responses to exercise remain unclear.
- Common genetic variants of genes related to the vascular transduction of sympathetic outflow have been investigated, but variants influencing skeletal muscle afferent feedback during exercise have not been explored.
- Single nucleotide polymorphisms in *TRPV1* rs222747 and *BDKRB2* rs1799722 receptors present in skeletal muscle were associated with differences in the magnitude of the blood pressure response to static handgrip exercise but not mental stress.
- The combined effects of *TRPV1* rs222747 and *BDKRB2* rs1799722 on blood pressure and heart rate responses during exercise were additive, and primarily found in men.
- Genetic differences in skeletal muscle metaboreceptors may be a risk factor for exaggerated blood pressure responses to exercise.

Abstract Exercise blood pressure (BP) responses demonstrate high inter-individual variability, which could relate to differences in metabolically sensitive afferent feedback from the exercising muscle. We hypothesized that single-nucleotide polymorphisms (SNPs) in genes encoding metaboreceptors present in group III/IV skeletal muscle afferents can influence the exercise pressor response. Two hundred men and women underwent measurements of continuous BP and heart rate at baseline and during 2 min of static handgrip exercise (30% maximal volitional contraction), post-exercise circulatory occlusion and mental stress (serial subtraction; internal control). Participants were genotyped for SNPs in *TRPV1* (rs222747; G/C), *ASIC3* (rs2288645; G/A), *BDKRB2* (rs1799722; C/T), *PTGER2* (rs17197; A/G) and *P2RX4* (rs25644; A/G). Exercise systolic BP (19 \pm 10 *vs.* 22 \pm 10 mmHg, *P* = 0.03) was lower in GG *versus* GC/CC minor allele carriers for *TRPV1* rs222747, while exercise diastolic BP (14 ± 7 *vs*. 17 ± 7 mmHg, $P = 0.007$) and

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heart rate (12 [±] ⁸ *vs*. 15 [±] 9 beats min−1, *^P* ⁼ 0.03) were lower in CC *versus* CT/TT minor allele carriers for *BDKRB2* rs1799722. Individuals carrying both minor alleles for *TRPV1* rs222747 and *BDKRB2* rs1799722 had greater systolic (22 \pm 11 *vs*. 17 \pm 10 mmHg, *P* = 0.04) and diastolic (18 \pm 7 *vs.* 14 ± 7 mmHg, $P = 0.01$) BP responses than those with no minor alleles; these differences were larger in men. No differences in BP or heart rate responses were detected during static handgrip with *ASIC3* rs2288645, *PTGER2* rs17197 or *P2RX4* rs25644. None of the selected SNPs were associated with differences during mental stress. These findings demonstrate that variants in *TRPV1* and *BDKRB2* receptors can contribute to BP differences during static exercise in an additive manner.

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Introduction

Blood pressure (BP) responses to exercise demonstrate high inter-individual variability (Ingelsson *et al.* 2007; Nunes *et al.* 2014; Notay *et al.* 2018), with exaggerated responses linked to an increased risk of future hypertension and cardiovascular mortality (Chaney & Eyman, 1988; Miyai *et al.* 2000, 2002; Weiss *et al.* 2010). The mechanisms responsible for the variability in exercise pressor responses are unclear. Activation of the sympathetic nervous system is critical for the maintenance of exercising BP and matching of cardiac output to skeletal muscle demand (Joyner & Casey, 2015), with accentuated sympathetic vasoconstriction considered a mechanism for a hypertensive response to exercise (Schultz & Sharman, 2014). During exercise, increases in sympathetic outflow occur primarily from feedforward signals from higher brain regions related to perception of effort (i.e. central command) and feedback signals from group III/IV skeletal muscle afferents sensitive to stretch and chemical stimuli, termed the muscle mechanoreflex and metaboreflex, respectively (Fisher *et al.* 2015). Stimulation of the muscle metaboreflex is known to potently activate the sympathetic nervous system (Mark *et al.* 1985), and its over-activation is implicated in mediating larger BP responses during exercise in patients with hypertension (Delaney *et al.* 2010; Greaney *et al.* 2015).

Evidence suggests that genetic influences can contribute to the variability in BP responses to exercise; however, to date, most research has focused on single-nucleotide polymorphisms (SNPs) in genes encoding adrenergic receptors (i.e. related to the vascular transduction of efferent sympathetic outflow), endothelial nitric oxide synthase, or components of the renin–angiotensin–aldosterone system (Eisenach *et al.* 2005; Ueno *et al.* 2005; Nieminen *et al.* 2006; Ingelsson *et al.* 2007; Dias *et al.* 2009; Nunes *et al.* 2014, 2016). SNPs are single-base-pair alterations in DNA that are prevalent throughout the human genome, and can influence, for example, gene expression or receptor function (Shastry, 2009). Whether genetic variations in metabolically sensitive receptors found in skeletal muscle group III/IV afferents contribute to the variability in BP responses to exercise is unknown. Functional differences in the chemical sensitivity of muscle metaboreceptors could modulate afferent feedback to brainstem regions controlling efferent sympathetic and parasympathetic outflow, and ultimately BP (Amann *et al.* 2015). Prior work has shown that group III/IV skeletal muscle afferents respond to a wide array of stimuli, and can contain acid sensing ion channel (ASIC), transient receptor potential vanilloid 1 (TRPV1), prostaglandin E2 (PTGER2), purinergic P2X (P2RX) and bradykinin B2 (BDKRB2) receptors (Greaney *et al.* 2015), though the specific contributions of individual receptor classes, particularly TRPV1 receptors, on the exercise pressor reflex remain controversial. Importantly, each of these genes possesses SNPs with common genetic variants (i.e. minor allele frequencies >10%).

The primary purpose of this study was to determine the effects of genetic variants of genes encoding metabolically sensitive receptors, present in group III/IV skeletal muscle afferents, on heart rate and BP responses during static handgrip exercise. This exercise mode was selected based on the large body of evidence that BP responses to static handgrip exercise are tightly linked to activation of the muscle metaboreflex (Mark *et al.* 1985; Delaney *et al.* 2010). To further isolate the actions of the muscle metaboreflex, BP responses were also assessed during post-exercise circulatory occlusion (PECO). In addition, as many of the metaboreceptors found in group III/IV afferents are not selective to skeletal muscle, heart rate and BP responses were measured during a mental stress task (serial subtraction) to serve as an internal control (i.e. a stimulus that increases BP without feedback from group III/IV afferents). We hypothesized that genetic variants in metabolically sensitive receptors found in skeletal muscle would contribute to the high inter-individual variability observed in BP responses to static handgrip exercise but not mental stress.

Methods

Ethics approval

The University of Guelph Research Ethics Board approved all procedures (REB no. 16-12-688/17-05-009) and the study conformed to the standards set by the *Declaration of Helsinki*, except for registration in a database. All participants provided informed, written consent prior to engaging in the study protocol.

Participants

Two hundred young healthy recreationally active men $(n = 91)$ and women $(n = 109)$ participated in the study between January 2017 and February 2018. All female participants were studied during the early follicular phase (between days 1 and 5) of the menstrual cycle and self-reported that they had a regular 28-day menstrual cycle; those on oral contraception $(n = 47)$ were studied within the first 5 days of their placebo pill phase. All participants were free of known cardiovascular or metabolic disease, and did not consume any chronic medications other than oral contraception. The present cohort consisted almost exclusively of non-Hispanic Caucasians with only five Hispanic and three black participants. A portion of the static handgrip blood pressure data from this study has been used previously to address an unrelated hypothesis (Notay *et al.* 2018).

Experimental protocol

The study consisted of a randomized crossover design examining BP responses to static handgrip exercise and mental stress. All participants underwent a familiarization visit to describe all aspects of the study protocol and practice performing maximal voluntary contractions on a handgrip dynamometer (Lafayette Instrument, Lafayette, LA, USA), as well as a single testing visit. Prior to the testing visit, participants were asked to refrain from caffeine, alcohol and vigorous exercise for a minimum of 24 h, and food or fluids for 1 h. Participants were studied in a light and temperature controlled laboratory. Following voiding and collection of anthropometric measurements, participants were positioned upright on a comfortable chair with their feet supported on an ottoman. Participants were asked to execute two maximal handgrip contractions in their left hand to determine maximal voluntary contraction (MVC); 16 participants were left-handed (7 men; 9 women). Each contraction lasted \sim 3 s and they were separated by at least 30 s of rest. If the MVC between the two contractions was $>$ 3 kg, a third MVC was required to be completed; however, this did not occur in any of the participants. The highest value was taken as MVC.

Next, participants were given 10 min of rest, after which continuous measures of heart rate and BP, as well as discrete minute-to-minute brachial BP, were

collected simultaneously over a 5 min resting baseline period. Following this baseline period, participants were randomized to begin collection during either the static handgrip or the mental stress protocol; a minimum of 10 min rest was provided between the two stressors to ensure that all cardiovascular measures returned to baseline values. Continuous measurements of heart rate and BP were collected throughout both stressors. During the static handgrip protocol, participants completed a 2 min resting baseline, 2 min static handgrip contraction at 30% MVC, and 2 min of PECO. To complete PECO, a manual sphygmomanometer (DS400 Aneroid Sphygmomanometer; D.E. Hokanson Inc., Bellevue, WA, USA) was inflated to 220 mmHg (i.e. suprasystolic) in the upper left arm immediately prior to the completion of the static handgrip contraction. During the mental stress task, participants were randomly assigned to subtract 11 or 13 from a three- or four-digit number, with a new number appearing on a personal computer every 5 s for a total of 2 min. The same 24 numbers were used for all participants. Answers were monitored to ensure that effort was given during the test; however, no verbal feedback was given to participants regarding correctness. Encouragement was also given to ensure participants continued to answer questions.

Measurements

Electrocardiography (Lead II) was used to continuously obtain beat-to-beat heart rate (ADInstruments Inc., Colorado Springs, CO, USA). Respiratory movements were monitored to ensure spontaneous breathing using a piezoelectric transducer positioned around the abdomen (Pneumotrace II, UFA, Morro Bay, CA, USA). To attain accurate recordings of blood pressure at rest, discrete left brachial blood pressure was logged on a minute-to-minute basis using an automated sphygmomanometer (BPTru Medical Devices, Coquitlam, Canada). A total of six discrete readings were taken, with the average of the last five recordings used for analysis. Both medium (cuff dimensions: 12×23 cm; $n = 132$) and large (cuff dimensions: 15×33 cm; $n = 68$) sized BP cuffs were used depending on the participant's arm circumference. Continuous beat-to-beat BP was recorded from the right middle finger using photoelectric plethysmography (Finometer MIDI, Finapres Inc., Enschede, Netherlands) to monitor haemodynamic responses during exercise and mental stress. Small (cuff fit: 45–55 mm; *n* = 27), medium (cuff fit: 55–65 mm; *n*=139) and large (cuff fit: 65–75 mm; $n = 34$) sized finger cuffs were used depending on the participant's middle finger circumference. All continuous data were digitized and stored with LabChart (PowerLab, ADInstruments, Colorado Springs, CO, USA). Heart rate, respiration and blood pressure were recorded at a sampling frequency of 1000 Hz.

Selection of genetic variants and genotyping

The five candidate SNPs were chosen based on prior literature demonstrating associations with differences in BP at rest (Ko *et al.* 2008), investigated for differences in pulse pressure (Stokes *et al.* 2011), higher prevalence in hypertensive populations (Cui*et al.* 2005; Sato *et al.* 2007), or alterations in receptor structure or sensitivity (Wang *et al.* 2016). All SNPs had a minor allele frequency (MAF) >10% in a European population according to dbSNP [\(https://www.ncbi.nlm.nih.gov/projects/SNP/\)](https://www.ncbi.nlm.nih.gov/projects/SNP/). DNA was extracted from either venous blood ($n = 90$) or saliva $(n = 110)$, as previously described (Klingel *et al.* 2017). Briefly, 2 mL of saliva was collected in an Oragene DNA collection kit and DNA was extracted according to the manufacturer's instructions (DNA Genotek, Ottawa, ON, Canada). For venous blood, DNA was extracted using the Qiagen Paxgene Blood DNA kit, according to the manufacturer's instructions (Qiagen, Toronto, ON, Canada). Participants were genotyped for SNPs in *TRPV1* (rs222747; G/C; MAF: 0.32), *ASIC3* (rs2288645; G/A; MAF: 0.20), *BDKRB2* (rs1799722; C/T; MAF: 0.43), *P2RX4* (rs25644; A/G; MAF: 0.16) and *PTGER2* (rs17197; A/G; MAF: 0.22) using the MassARRAY $\textcircled{\tiny R}$ Analyser 4 System (Agena Biosciences, San Diego, CA, USA) using iPlex gold chemistry and analysed using Typer 4.0 software at SickKid's Centre for Applied Genomics (Toronto, ON, Canada). Briefly, each locus was amplified by polymerase chain reaction and a third primer that flanks the polymorphism site was extended by one base. The extension reaction products were analysed using matrix-assisted laser desorption/ionization time of flight (MALDI-TOF) mass spectrometry to identify the amount and type of molecules present in the sample (Storm *et al.* 2003). Five DNA samples were randomly selected for replication and 100% concordance was achieved. Three quality control samples were also run and had 100% agreement.

Data and statistical analysis

Heart rate and BP measurements were averaged over a 2 min resting baseline and during the second minute of static handgrip exercise, PECO and mental stress. The change (Δ) from baseline was calculated for statistical analysis. The coefficients of variation were calculated to measure the absolute variability of BP and heart rate responses during static handgrip exercise. All haemodynamic data were collected and analysed prior to obtaining genotyping results (i.e. blind to group allocation).

Statistical analyses were performed using IBM SPSS Statistics 25 (IBM Corp., Armonk, NY, USA) and Prism (GraphPad Software Inc., La Jolla, CA, USA). Deviation from Hardy–Weinberg equilibrium was tested for each SNP using the χ^2 test. Due to the low number of

Table 1. Participant baseline characteristics

homozygotic minor alleles for most SNPs, we combined heterozygous and minor homozygous subjects into a single group called 'minor allele carriers' for each SNP (Klingel *et al.* 2017). Thus, associations between genes and study endpoints were investigated using a dominant genetic model (i.e. MM *vs*. Mm+mm). BP and heart rate responses between genotype groups were compared using a one-way analysis of covariance (ANCOVA). Age and sex were used as covariates for the comparison of resting baseline values, PECO and mental stress responses, while age, sex and MVC (Notay *et al.* 2018) were used as covariates during static handgrip exercise. Relative proportions of allele groups between men and women at baseline were tested using two-tailed Fisher's exact tests. Subgroup analysis was conducted to determine the influence of sex on heart rate and BP responses to static handgrip exercise, PECO and mental stress using two-way ANCOVAs. Age was used as a covariate for PECO and mental stress responses, while age and MVC were used during static handgrip exercise. Significant allele \times sex interactions were probed using Bonferroni *post hoc* tests.

To assess the additive effects of significant SNPs, we computed a genetic risk score which labelled individuals as 0, 1 or 2, according to the number of minor alleles they were carrying for the *TRPV1* rs222747 and *BDKRB2* rs1799722 SNPs. Group 0 comprised individuals carrying no minor alleles, group 1 comprised individuals carrying one minor allele (irrespective of the gene) and group 2 comprised individuals carrying both minor alleles. BP and heart rate responses between genetic risk score groups were compared with an ANCOVA, using age, sex and MVC as covariates. Significant main effects between genetic risk score groups were probed using Bonferroni *post hoc* tests. Significance was considered $P < 0.05$. Data are presented as mean \pm SD, unless otherwise stated.

Results

Complete BP, heart rate and genotype data were obtained in 200 participants with no adverse effects. Resting baseline characteristics of the cohort are presented in Table 1. All of the investigated SNPs were in Hardy–Weinberg equilibrium ($P > 0.05$).

As expected, all participants experienced increases in both systolic and diastolic BP during static handgrip exercise. The majority of participants also had an increase in heart rate with the exception of three individuals who had a slight decrease (removal of these participants did not impact the results). There was no effect of testing order on any variable $(P > 0.05)$. Figure 1 displays the distributions and coefficients of variation for changes in BP and heart rate with static exercise. Overall, the mean increase in systolic and diastolic BP during static handgrip

Figure 1. Distributions and coefficients of variation (CV) for changes in systolic BP (*A***), diastolic BP (***B***) and heart rate (***C***) during static handgrip exercise**

Table 2. Baseline characteristics for *TRPV1* **rs222747**

Characteristic	GG	$CG + CC$	P
Age (years)	$22 + 2$	$22 + 3$	0.80
Sex (male/female)	43/53	48/56	0.89
Height (cm)	$172 + 9$	$170 + 9$	0.10
Weight (kg)	71 ± 13	$67 + 13$	0.05
Body mass index (kg m ⁻²)	$24 + 3$	$23 + 3$	0.14
Heart rate (beats min ⁻¹)	$66 + 11$	$68 + 11$	0.15
Systolic blood pressure (mmHq)	$105 + 8$	$104 + 9$	0.75
Diastolic blood pressure (mmHq)	$66 + 7$	66 ± 7	0.97
Mean arterial pressure (mmHq)	$79 + 7$	$79 + 7$	0.92
Maximal volitional contraction (kg)	$36 + 12$	$36 + 13$	0.92
Data are shown as mean \pm SD.			

exercise was 20 ± 10 and 16 ± 7 mmHg, respectively. These pressor responses were largely maintained during PECO, which had mean increases in systolic and diastolic BP of 18 ± 10 and 12 ± 6 mmHg, respectively. The mean increases in heart rate during static handgrip exercise and PECO were 14 \pm 9 and 0 \pm 6 beats min⁻¹, respectively.

TRPV1 **rs222747**

Baseline characteristics were not significantly different between major (GG) and minor ($CC + CG$) allele carriers (Table 2). Systolic BP responses were smaller in GG *versus* CC + CG individuals during static handgrip exercise (19 \pm 10 *vs*. 22 \pm 10 mmHg, *P* = 0.03), but not PECO (17 \pm 10 *vs*. 19 \pm 11 mmHg, *P* = 0.29) or mental stress (14 \pm 9 *vs*. 14 ± 12 mmHg, $P = 0.94$) (Fig. 2). Diastolic BP responses were not significantly different between GG *versus* CC + CG individuals during static handgrip exercise (16 ± 7 *vs*. 17 ± 7 mmHg, $P = 0.15$), PECO (12 ± 6 *vs*. 12 ± 6 mmHg, $P = 0.65$) or mental stress (10 \pm 6 *vs*. 9 \pm 5 mmHg, $P =$ 0.14). A trend for smaller heart rate responses to static handgrip exercise was found in GG compared to CC + CG (13 \pm 8 *vs.* 15 \pm 9 beats min⁻¹, *P* = 0.09) but not during PECO (0 \pm 5 *vs*. 0 \pm 7 beats min⁻¹, *P* = 0.80) or mental stress (9 [±] ⁹ *vs*. 9 [±] 9 beats min−1, *^P* ⁼ 0.95). Subgroup analysis did not detect any allele \times sex interactions for static handgrip exercise, PECO or mental stress (all $P > 0.05$; data not shown).

BDKRB2 **rs1799722**

Baseline diastolic BP was higher in major (CC) allele carriers than minor (CT + TT) (67 \pm 8 *vs*. 65 \pm 6 mmHg, $P = 0.02$); however, all other characteristics were not significantly different between groups (all *P* > 0.05) (Table 3). Adjusting for this small baseline difference in diastolic BP did not alter the results for static handgrip exercise, PECO or mental stress data. Systolic BP responses were not different between CC and CT + TT individuals during static handgrip exercise (19 ± 9 *vs*. 21 ± 10 mmHg,

Table 3. Baseline characteristics for *BDKRB2* **rs1799722**

Data are shown as mean \pm SD.

P = 0.16), PECO (16 \pm 9 *vs*. 18 \pm 11 mmHg, *P* = 0.26) or mental stress (15 \pm 11 *vs*. 14 \pm 11 mmHg, *P* = 0.46). In contrast, CC had smaller diastolic BP responses during static handgrip exercise (14 \pm 7 *vs.* 17 \pm 7 mmHg, *P* = 0.007) and PECO (11 \pm 6 *vs*. 13 \pm 7 mmHg, *P* = 0.05), but not mental stress (10 \pm 6 *vs*. 10 \pm 5 mmHg, *P* = 0.98). Heart rate exhibited similar attenuated responses in CC during static handgrip exercise (12 \pm 8 *vs*. 15 \pm 9 beats min⁻¹, *P* = 0.03) but not PECO (0 ± 5 *vs*. 1 ± 7 beats min⁻¹, *P* = 0.23) or mental stress (9 ± 10 *vs*. 8 ± 8 beats min⁻¹, *P* = 0.61) (Fig. 3). Subgroup analysis demonstrated significant allele \times sex interactions with attenuated systolic and diastolic BP responses during static handgrip exercise in CC $vs.$ CC + CT men (21 \pm 10 *vs*. 26 ± 9 mmHg, $P = 0.01$; and 15 ± 7 *vs*. 19 ± 7 mmHg,

Data are mean ± SEM. *P* values adjusted for age, sex and maximal volitional contraction (static handgrip exercise only)

Data are mean \pm SEM. *P* values adjusted for age, sex and maximal volitional contraction (static handgrip exercise only).

 $P = 0.001$, respectively) but no differences in women (all $P > 0.05$) (Fig. 4). There were no sex differences in heart rate responses during static handgrip exercise, or heart rate and BP responses during PECO or mental stress (all $P > 0.05$.

ASIC3 **rs2288645**

All baseline characteristics were similar between major (GG) and minor $(AA + AG)$ allele carrier groups, as were all BP and heart rate responses to static handgrip exercise, PECO, and mental stress (all *P* > 0.05; Table 4). Subgroup analysis did not yield any allele \times sex interactions during static handgrip exercise or mental stress (all $P > 0.05$); however, GG men exhibited a larger systolic BP response during PECO (22 \pm 11 *vs*. 17 \pm 9 mmHg, *P* = 0.03).

PTGER2 **rs17197**

Baseline heart rate was lower in major (AA) allele carriers compared to minor $(AG + GG)$ $(66 \pm 11 \text{ vs. } 69 \pm \text{)}$ 11 beats min⁻¹, $P = 0.01$), though no other characteristics

Figure 4. Mean changes (*-***) in systolic BP (***A***) and diastolic BP (***B***) during the second minute of static handgrip exercise in** *BDKRB2* **rs222747 within male (left) CC (***n* **= 25) and CT + TT** $(n = 66)$ allele carriers and within female (right) CC ($n = 40$) and $CT + TT$ ($n = 69$) allele carriers

Data are mean \pm SEM. P values adjusted for age and maximal volitional contraction.

Table 4. *PTGER2* **rs17197 polymorphism on baseline participant characteristics and BP and heart rate responses to static handgrip exercise, PECO and mental stress**

Data are shown as mean \pm SD

were different between groups (all *P* > 0.05), as were all BP and heart rate responses during static handgrip exercise, PECO and mental stress (Table 5). Subgroup analysis did not yield any allele \times sex interactions during static handgrip exercise or mental stress (all $P > 0.05$); however, AA men demonstrated larger systolic BP responses during PECO (23 \pm 12 *vs*. 17 \pm 7 mmHg, *P* = 0.01). A similar trend was observed for diastolic BP during PECO (interaction term, $P = 0.06$).

P2RX4 **rs25644**

All baseline characteristics were similar between major (AA) and minor $(AG + GG)$ allele carrier groups, as were all BP and heart rate responses to static handgrip exercise, PECO and mental stress (all *P* > 0.05; Table 6). Subgroup

Table 5. *ASIC3* **rs2288645 polymorphism on baseline participant characteristics and BP and heart rate responses to static handgrip exercise, PECO and mental stress**

	GG	$AA + AG$	P
Baseline characteristics			
Number (n)	132	68	
Age (years)	$21 + 2$	$22 + 3$	0.29
Sex (male/female)	56/76	35/33	0.23
Height (cm)	171 ± 9	172 ± 9	0.66
Weight (kg)	69 \pm 13	68 ± 13	0.64
Body mass index (kg m ⁻²)	24 ± 3	23 ± 3	0.30
Heart rate (beats min ⁻¹)	68 ± 11	66 ± 10	0.75
Systolic blood pressure (mmHq)	104 ± 9	105 ± 8	0.78
Diastolic blood pressure (mmHq)	$65 + 7$	$66 + 7$	0.24
Mean arterial pressure (mmHq)	$78 + 7$	$79 + 7$	0.35
Maximal volitional contraction (kq)	$35 + 13$	$36 + 13$	0.70
Static handgrip exercise			
\triangle Systolic blood pressure (mmHq)	20 ± 10	21 ± 11	0.69
Δ Diastolic blood pressure (mmHq)	16 ± 7	17 ± 7	0.34
Δ Heart rate (beats min ⁻¹)	14 ± 9	14 ± 8	0.76
PECO			
Δ Systolic blood pressure (mmHq)	18 ± 11	17 ± 10	0.45
Δ Diastolic blood pressure (mmHg)	12 ± 7	12 ± 6	0.51
Δ Heart rate (beats min ⁻¹)	1 ± 7	0 ± 6	0.50
Mental stress			
Δ Systolic blood pressure (mmHq)	14 ± 10	16 ± 12	0.20
Δ Diastolic blood pressure (mmHq)	10 ± 6	10 ± 6	0.37
Δ Heart rate (beats min ⁻¹)	9 ± 9	8 ± 9	0.50
Data are shown as mean \pm SD			

analysis did not yield any significant allele \times sex interactions (all $P > 0.05$) (data not shown).

*TRPV1***–***BDKRB2* **genotype risk score**

As shown in Fig. 5, individuals carrying both minor alleles (group 2) had larger systolic BP responses during static handgrip exercise than individuals carrying no minor alleles (group 0) $(22 \pm 11 \text{ vs. } 17 \pm 10 \text{ mmHg}, P = 0.04)$. Differences between group 2 and those carrying one minor allele (group 1) (22 \pm 11 *vs*. 20 \pm 9 mmHg, *P* = 0.45) or between group 1 and group 0 (20 \pm 9 *vs*. 17 \pm 10 mmHg, $P = 0.48$) did not reach statistical significance. Diastolic BP responses during static handgrip exercise were larger in group 2 *vs*. group 0 (18 \pm 7 *vs*. 14 \pm 7 mmHg, *P* = 0.01) and tended to be larger between group 1 *vs*. group **Table 6.** *P2RX4* **rs25644 polymorphism on baseline participant characteristics and BP and heart rate responses to static handgrip exercise, PECO and mental stress**

Data are shown as mean \pm SD

0 (16 \pm 6 *vs.* 14 \pm 7 mmHg, *P* = 0.08). No differences were found between group 2 *vs*. group 1 (18 ± 7 *vs*. 16 ± 6 mmHg, $P = 0.97$). Heart rate responses during static handgrip exercise were larger in group 2 *vs*. group 1 (16 ± 10 *vs*. 12 ± 8 beats min⁻¹, *P* = 0.04) and group 0 (16 ± ¹⁰ *vs*. 13 [±] 9 beats min−1, *^P* ⁼ 0.04). No differences were observed between group 1 *vs*. group 0 (13 \pm 9 *vs*. 12 \pm 8 beats min−1, *^P* ⁼ 0.99). Subgroup analysis demonstrated that within men those in group 2 had larger systolic BP responses during static handgrip exercise than group 0 $(27 \pm 11 \text{ vs. } 20 \pm 9 \text{ mmHg}, P = 0.01)$. Similarly, diastolic BP responses during static handgrip exercise were larger in group 2 *vs*. group 0 (19 \pm 7 *vs*. 13 \pm 7 mmHg, *P* = 0.001) and between group 1 *vs*. group 0 (18 \pm 6 *vs*. 13 \pm 7 mmHg, $P = 0.004$). Heart rate responses were larger in group 2 *vs*. group 0 (17 ± 11 *vs*. 11 ± 8 beats min⁻¹, $P = 0.02$). In contrast, all BP and heart rate responses during static handgrip exercise were not significantly different in women (all $P > 0.05$).

Discussion

The feedback of group III/IV skeletal muscle afferents is critical to maintaining the appropriate haemodynamic response during exercise (Amann *et al.* 2010). The present study sought to investigate whether genetic variants in metabolically sensitive receptors found in skeletal muscle afferents can influence BP responses to exercise. In support of our hypothesis, we observed that minor allele carriers of both *TRPV1* rs222747 and *BDKRB2* rs1799722 were associated with larger BP and/or heart rate responses during static handgrip exercise but not our internal control mental stress task. Further, the diastolic BP differences observed with *BDKRB2* rs1799722 during static handgrip exercise were also present during isolation of the muscle metaboreflex using PECO. Although independently these relative changes were small $(\Delta 2-3$ mmHg), the combination of these polymorphisms resulted in a \sim 22–23% greater difference in systolic and diastolic BP responses (Δ 4–5 mmHg) between individuals carrying both minor alleles than individuals carrying no minor alleles. Interestingly, subgroup analysis determined that the haemodynamic effects of these two genetic variants were primarily in men, with even larger differences (Δ 6–7 mmHg) in exercise BP responses between groups. No differences in haemodynamic responses during exercise or mental stress were detected for *PTGER2* rs17197, *ASIC3* rs2288645

or *P2RX4* rs25644. These results demonstrate that genetic variants in skeletal muscle metaboreceptors can independently and synergistically influence the magnitude of the BP response to exercise.

Prior investigations focusing on the contribution of genetic variants to the variability in exercise BP responses have focused primarily on polymorphisms in genes related to adrenergic and renin–angiotensin–aldosterone system receptors (Eisenach *et al.* 2005; Ueno *et al.* 2005; Nieminen *et al.* 2006; Ingelsson *et al.* 2007; Dias *et al.* 2009; Nunes *et al.* 2014, 2016). However, while these studies have demonstrated that such SNPs can influence pressor responses to exercise, they may be confounded by not measuring or controlling for potential differences in central sympathetic outflow to the vasculature. Afferent signals arising from skeletal muscle provide important feedback to the brainstem to maintain necessary BP and heart rate responses during exercise (Amann *et al.* 2010). The exercise pressor reflex is known to consist of mechanically and metabolically sensitive afferents, which relate primarily to group III and IV afferents, respectively (Fisher *et al.* 2015). The contribution of this reflex is well studied and activation of the muscle metaboreflex is known to be a potent stimulator of sympathetic vasoconstrictor outflow and BP responses during static handgrip exercise (Mark *et al.* 1985). A large number of studies, primarily in animal models (Stebbins *et al.* 1986, 1990; Rotto & Kaufman, 1988; Rotto *et al.* 1989; Symons *et al.* 1991; Pan *et al.* 1993; Kindig *et al.* 2005; Hayes *et al.* 2008; Smith *et al.* 2010; Mizuno *et al.* 2011; Stone *et al.* 2015), have sought to identify the roles of specific metabolites responsible for engaging the

Figure 5. Genetic risk score investigating the additive effects of *TRPV1* **rs222747 and** *BDKRB2* **rs222747 on the mean changes (***-***) in systolic BP (***A***) and diastolic BP (***B***) during the second minute of static handgrip exercise across all participants (left), men (middle) and women (right)**

Group 0 comprised individuals carrying no minor alleles, group 1 comprised individuals carrying one minor allele (irrespective of the gene) and group 2 comprised individuals carrying both minor alleles. All participants, group 0 $(n = 34)$, group 1 $(n = 91)$, group 2 $(n = 75)$; in men, group 0 $(n = 13)$, group 1 $(n = 40)$ and group 2 $(n = 38)$; in women, group 0 ($n = 21$), group 1 ($n = 51$) and group 2 ($n = 37$). Data are mean \pm SEM. *P* values adjusted for age, sex and maximal volitional contraction.

exercise pressor reflex, focusing specifically on stimuli to TRPV1, BDKRB2, ASIC, P2RX4 and PTGER2 receptor pathways (Greaney *et al.* 2015). However, the physiological contributions or role of each receptor class remain controversial and difficult to study in humans. Examining common genetic variants, especially those with known functional consequences, may represent a novel model to uncover the involvement of specific receptors.

Direct data investigating the role of TRPV1 receptors on the exercise pressor reflex in humans are limited. Recently, topical forearm application of capsaicin (a selective TRPV1 agonist) was shown to attenuate muscle sympathetic nerve activity during static handgrip and BP and muscle sympathetic responses during PECO, hypothesized to be secondary to desensitization of TRPV1 receptors (Vianna *et al.* 2018). Similarly, animal data from rats demonstrate that infusion of a TRPV1 antagonist attenuates the mean BP and heart rate responses to a static contraction by 7 mmHg and 2 beats min−1, respectively, through a metabolic, not mechanical, pathway (Smith *et al.* 2010). Further, blockade of TRPV1 receptors attenuates the overactive exercise pressor response in hypertensive rats (Mizuno *et al.* 2011). The present results support a role for TRPV1 in mediating the pressor responses to exercise in humans and demonstrate that the *TRPV1* rs222747 polymorphism can contribute to the BP response to static handgrip exercise, with larger systolic BP responses seen in $CC + GC$ individuals (independent of sex). We did not investigate the functional consequences of *TRPV1* rs222747; however, in line with our findings, prior *ex vivo* culture work in human embryonic kidney suggests that this variant can be associated with increased calcium permeation and gain-of-function in response to acid or capsaicin (Wang *et al.* 2016).

Skeletal muscle contraction is associated with the release of bradykinin (Rett*et al.* 1989), and despite its known role as a vasodilator and potential regulator of exercise-induced hyperaemia (Langberg *et al.* 2002), bradykinin can also stimulate group III/IV afferents, particularly at high intensities (Stebbins *et al.* 1990; Pan *et al.* 1993). Although data are limited in humans, BDKRB2 antagonism in cats attenuates the BP and heart rate during a 30 s electrically stimulated static hindlimb contraction by ~41-50% (Pan et al. 1993). The present results support a role of bradykinin in activating the exercise pressor reflex in humans and demonstrate that *BDKRB2* rs1799722 $CT + TT$ individuals have greater diastolic BP responses during static handgrip exercise and PECO in comparison to major CC carriers. Subgroup analysis demonstrated that the systolic and diastolic BP responses to static handgrip exercise were impacted by sex, with differences present between CC v_s . CT + TT allele carriers in men but not women. These findings align with prior work demonstrating the influence of another *BDKRB*2 polymorphism, rs5810761, found to be associated with systolic BP responses during treadmill exercise in men but not in women (Nunes *et al.* 2014). Future work is required to understand the mechanisms responsible for sex-based differences in the influence of *BDKRB*2 polymorphisms on BP responses to exercise.

ASIC receptors have been shown to contribute to the exercise pressor reflex, as blunting lactic acid production during 20% MVC static handgrip exercise to failure decreases sympathetic and BP responses (Ettinger *et al.* 1991). However, it should be noted that the attenuated BP responses were observed primarily in the fourth to fifth minute of exercise immediately prior to fatigue (Ettinger *et al.* 1991). Thus, our 2 min 30% MVC static handgrip contraction may not have been of sufficient duration (or degree of fatigue) to activate this pathway. Similarly, systemic blockade of prostaglandin synthesis can attenuate mean BP and heart rate during the third minute of a 30% MVC static handgrip contraction (Fontana *et al.* 1995), while local blockade can attenuate the muscle sympathetic response prior to 30% MVC static contraction failure (Cui *et al.* 2007). However, not all work is consistent as prostaglandin blockade did not impact BP during a 40% MVC static handgrip to exhaustion (Davy *et al.* 1993). Again, it should be noted that prostaglandin accumulation during static exercise is greatest during high-intensity or ischaemic exercise (Symons *et al.* 1991) and thus may not have been activated by our moderate intensity exercise protocol. Finally, P2RX receptors have been shown to contribute to the exercise pressor reflex in humans as infusion of a non-specific P2 receptor agonist (pyridoxine hydrochloride) attenuates BP responses to a 30% MVC static handgrip contraction to failure (Cui *et al.* 2011). Again, this observation was seen in a static handgrip exercise response to failure (i.e. ischaemic model). Future work is required to establish the role of *ASIC3*, *P2RX4* and *PTGER2* polymorphisms using different exercise paradigms of varying intensity and contraction duration.

In addition to examining the BP responses to static handgrip exercise, we employed PECO in an attempt to isolate the effects of the muscle metaboreflex. Our study aligns with prior work (e.g. Mark *et al.* 1985) showing that BP responses to static handgrip exercise are largely maintained during PECO and thus considered to be mediated primarily by stimulation of the muscle metaboreflex. We did observe that the effects of *BDKRB2* rs1799722 on diastolic BP responses demonstrated parallel group differences during static handgrip exercise and PECO; however, the effects of *TRPV1* rs222747 on systolic BP responses and *BDKRB2* rs1799722 on heart rate responses during static handgrip exercise were not accompanied by similar observations during PECO. The latter observations are not surprising given that heart rate responses during static handgrip exercise quickly return to baseline during PECO (present study: $\overline{\Delta}$ 0 ± 6 beats min⁻¹). While this could be taken

as evidence against a role of the muscle metaboreflex in controlling heart rate, other work suggests that metaboreflex-mediated cardiac sympathetic activation is largely masked by parallel cardiac parasympathetic reactivation at moderate intensities (Fisher *et al.* 2010). Interestingly, we observed that *ASIC3* rs2288645 and *PTGER2* rs17197 both impacted systolic BP during PECO, but not static handgrip, in men. This is consistent with the need for a larger ischaemic stimulus to activate these pathways, and with evidence that men have greater BP response to static handgrip and PECO (Notay *et al.* 2018; Parmar *et al.* 2018). It is also important to consider methodological limitations of using PECO to isolate the haemodynamic contributions of the muscle metaboreflex. Mainly, the PECO model assumes that neural reflexes (e.g. mechanoreflexes, metaboreflexes, central command) contribute additively and does not account for potential interactions. For example, it has been postulated that central command may be modified by perception of effort (Williamson, 2010), which itself can be influenced by feedback from group III/IV skeletal muscle afferents (Amann *et al.* 2008, 2015). Thus, during static handgrip exercise, alterations in muscle metaboreflex activation could be interacting with other neural mechanisms, such as central command, to further modify the BP response. Finally, the recent discovery of subtypes of group III/IV afferents which respond selectively to low or high (noxious) levels of intramuscular metabolites raise questions regarding the applicability of PECO, particularly as it relates to freely perfused exercise modes (Amann *et al.* 2015).

In addition to supporting a role for TRPV1 and BDKRB2 receptors in mediating the exercise pressor reflex in humans, the present study has a number of stimulating clinical implications. First, a substantial body of evidence demonstrates the prognostic value of exaggerated BP reactivity to exercise (Schultz & Sharman, 2014). That is, in otherwise healthy or diseased individuals, a larger BP response to dynamic (Miyai *et al.* 2000, 2002; Weiss *et al.* 2010) and static handgrip (Chaney & Eyman, 1988) exercise is associated with heightened risk for future hypertension or cardiovascular events. The elucidation of genetic risk factors for identifying such individuals would be beneficial for providing early monitoring or aggressive treatment to reduce these future risks. Second, over-activation of the muscle metaboreflex is considered to be a feature of the pathophysiology in both hypertension (Delaney *et al.* 2010; Greaney *et al.* 2015) and heart failure (Notarius *et al.* 2001). Older hypertensives display a \sim 30% larger systolic and diastolic peak BP response during a 40% MVC static handgrip exercise than age-matched normotensives (Delaney *et al.* 2010). In a cohort more comparable to the present study, young unmedicated prehypertensive men demonstrate \sim 30% higher peak mean arterial pressure responses during a 50% MVC static handgrip exercise than normotensive controls

(Choi *et al.* 2013). In the present study, the combination of the *TRPV1* rs222747 and *BDKRB2* rs1799722 polymorphisms in men resulted in individuals carrying minor alleles for both genes having 30% and 38% higher systolic and diastolic BP responses, respectively, during static handgrip exercise than those carrying no minor alleles. Thus, the BP differences caused by the *TRPV1* rs222747 and *BDKRB2* rs1799722 polymorphisms in men exhibit a similar magnitude difference as reported in those with prehypertension or hypertension. The functional consequences of an overactive muscle metaboreflex are more readily apparent in heart failure, where the result is exaggerated muscle sympathetic responses during static and dynamic exercise (Notarius *et al.* 2001, 2015), and restraint of exercising limb blood flow (Amann *et al.* 2014), providing a neural mechanism for exercise intolerance in this population. Whether the prevalence of *TRPV1* rs222747 and *BDKRB2* rs1799722 risk alleles in clinical populations further modifies BP or sympathetic responses warrants future investigation.

We acknowledge several limitations. First, the metabolically sensitive receptors found in group III/IV afferents are not specific to skeletal muscle and most can be found in several locations both centrally and peripherally. For example, TRPV1 receptors are expressed on both neural and non-neural cells, such as the brain, skin, mast cells, hair follicles, urinary bladder, lungs and inner ear, and are involved in a wide range of diverse physiological functions (Messeguer *et al.* 2006). Nevertheless, we selected each of these receptors on the basis of prior evidence that they are involved in mediating the exercise pressor response (Greaney *et al.* 2015) and employed mental stress to serve as an internal control to provide greater certainty that BP and heart rate differences during exercise were attributable to receptors present in skeletal muscle. Second, we examined the associations between polymorphisms and haemodynamic responses and did not determine how the structure or expression of these receptors was altered between major and minor allele carriers. In line with our findings, prior studies have demonstrated altered sensitivity of *TRPV1* rs222747 variants due to a Met315Ile missense mutation occurring in the exon region (Wang *et al.* 2016) and altered receptor expression of bradykinin receptors with *BDKRB2* rs1799722 variants due to the T allele (Braun *et al.* 1996). Third, we employed a 2 min static handgrip exercise completed in the seated posture and our results may not be generalizable to other postures, contraction durations or exercise modes. Fourth, our cohort consisted of young healthy individuals to avoid the confounding influences of background pharmacological therapy or co-morbidities; however, future research is necessary in those with hypertension shown to exhibit accentuated BP responses to exercise and heightened muscle metaboreflex activation (Delaney *et al.* 2010; Greaney *et al.* 2015). Fifth,

the coefficients of variation were larger during PECO than static handgrip, which likely impaired our ability to see parallel statistical changes in both tests; however, the magnitude and direction of differences between groups was similar. Finally, our sample size limited our capacity to investigate the interactions of all studied SNPs, and we consider these results to represent pilot findings to direct future investigations in larger cohorts.

Conclusion

Herein, we demonstrate that *TRPV1* and *BDKRB2* polymorphisms can influence the haemodynamic responses to exercise but not mental stress. Although the modest effects of individual genetic variants on BP and heart rate during exercise are not surprising given the likely redundancy of the muscle metaboreflex and the integrative complexity of BP regulation during exercise (Stone *et al.* 2015), we observed additive effects of*TRPV1* rs222747 and $BDKRB2$ rs1799722 resulting in \sim 22–23% differences in systolic and diastolic BP responses between individuals carrying both minor alleles compared to individuals carrying no minor alleles. Subgroup analysis determined that these differences were observed primarily in men, and corresponded similarly to reported differences between BP responses during static handgrip exercise in normotensive and hypertensive populations (Delaney *et al.* 2010; Choi *et al.* 2013). Future studies are required to test whether polymorphisms in both afferent and efferent pathways similarly produce additive responses and can explain a larger proportion of BP variance.

References

- Amann M, Blain GM, Proctor LT, Sebranek JJ, Pegelow DF & Dempsey JA (2010). Group III and IV muscle afferents contribute to ventilatory and cardiovascular response to rhythmic exercise in humans. *J Appl Physiol* **109**, 966–976.
- Amann M, Proctor LT, Sebranek JJ, Eldridge MW, Pegelow DF & Dempsey JA (2008). Somatosensory feedback from the limbs exerts inhibitory influences on central neural drive during whole body endurance exercise. *J Appl Physiol* **105**, 1714–1724.
- Amann M, Sidhu SK, Weavil JC, Mangum TS & Venturelli M (2015). Autonomic responses to exercise: group III/IV muscle afferents and fatigue. *Auton Neurosci Basic Clin* **188**, 19–23.
- Amann M, Venturelli M, Ives SJ, Morgan DE, Gmelch B, Witman MAH, Groot HJ, Walter Wray D, Stehlik J & Richardson RS (2014). Group III/IV muscle afferents impair limb blood in patients with chronic heart failure. *Int J Cardiol* **174**, 368–375.
- Braun A, Kammerer S, Maier E, Bohme E & Roscher AA ¨ (1996). Polymorphisms in the gene for the human B2-bradykinin receptor. New tools in assessing a genetic risk for bradykinin-associated diseases. *Immunopharmacology* **33**, 32–35.
- Chaney RH & Eyman RK (1988). Blood pressure at rest and during maximal dynamic and isometric exercise as predictors of systemic hypertension. *Am J Cardiol* **62**, 1058–1061.
- Choi H-M, Stebbins CL, Lee O-T, Nho H, Lee J-H, Chun J-M, Kim K-A & Kim J-K (2013). Augmentation of the exercise pressor reflex in prehypertension: roles of the muscle metaboreflex and mechanoreflex. *Appl Physiol Nutr Metab* **38**, 209–215.
- Cui J, Leuenberger UA, Blaha C, King NC & Sinoway LI (2011). Effect of P2 receptor blockade with pyridoxine on sympathetic response to exercise pressor reflex in humans. *J Physiol* **589**, 685–695.
- Cui J, McQuillan P, Momen A, Blaha C, Moradkhan R, Mascarenhas V, Hogeman C, Krishnan A & Sinoway LI (2007). The role of the cyclooxygenase products in evoking sympathetic activation in exercise. *Am J Physiol Heart Circ Physiol* **293**, H1861–H1868.
- Cui JA, Melista EA, Chazaro IA, Zhang YB, Zhou XB, Manolis AJE, Baldwin CTB, DeStefano ALC & Gavras HA (2005). Sequence variation of bradykinin receptors B1 and B2 and association with hypertension. *J Hypertens* **23**, 55–62.
- Davy KP, Herbert WG & Williams JH (1993). Effect of indomethacin on the pressor responses to sustained isometric contraction in humans. *J Appl Physiol* **75**, 273–278.
- Delaney EP, Greaney JL, Edwards DG, Rose WC, Fadel PJ & Farquhar WB (2010). Exaggerated sympathetic and pressor responses to handgrip exercise in older hypertensive humans: role of the muscle metaboreflex. *Am J Physiol Heart Circ Physiol* **299**, H1318–H1327.
- Dias RG, Alves M-JNN, Pereira AC, Rondon MUPB, dos Santos MR, Krieger JE, Krieger MH & Negrão CE (2009). Glu298Asp eNOS gene polymorphism causes attenuation in nonexercising muscle vasodilatation. *Physiol Genomics* **37**, 99–107.
- Eisenach JH, Barnes SA, Pike TL, Sokolnicki LA, Masuki S, Dietz NM, Rehfeldt KH, Turner ST & Joyner MJ (2005). Arg16/Gly β_2 -adrenergic receptor polymorphism alters the cardiac output response to isometric exercise. *J Appl Physiol* **99**, 1776–1781.
- Ettinger S, Gray K, Whisler S & Sinoway L (1991). Dichloroacetate reduces sympathetic nerve responses to static exercise. *Am J Physiol Heart Circ Physiol* **261**, H1653–H1658.
- Fisher JP, Seifert T, Hartwich D, Young CN, Secher NH & Fadel PJ (2010). Autonomic control of heart rate by metabolically sensitive skeletal muscle afferents in humans. *J Physiol* **588**, 1117–1127.
- Fisher JP, Young CN & Fadel PJ (2015). Autonomic adjustments to exercise in humans. *Compr Physiol* **5**, 475–512.
- Fontana GA, Pantaleo T, Bongianni F, Cresci F, Lavorini F, Guerra CT & Panuccio P (1995). Prostaglandin synthesis blockade by ketoprofen attenuates respiratory and cardiovascular responses to static handgrip. *J Appl Physiol* **78**, 449–457.
- Greaney JL, Wenner MM & Farquhar WB (2015). Exaggerated increases in blood pressure during isometric muscle contraction in hypertension: role for purinergic receptors. *Auton Neurosci Basic Clin* **188**, 51–57.

Hayes SG, McCord JL, Rainier J, Liu Z & Kaufman MP (2008). Role played by acid-sensitive ion channels in evoking the exercise pressor reflex. *Am J Physiol Heart Circ Physiol* **295**, H1720–H1725.

Ingelsson E, Larson MG, Vasan RS, O'Donnell CJ, Yin X, Hirschhorn JN, Newton-Cheh C, Drake JA, Musone SL, Heard-Costa NL, Benjamin EJ, Levy D, Atwood LD, Wang TJ & Kathiresan S (2007). Heritability, linkage, and genetic associations of exercise treadmill test responses. *Circulation* **115**, 2917–2924.

Joyner MJ & Casey DP (2015). Regulation of increased blood flow (hyperemia) to muscles during exercise: A hierarchy of competing physiological needs. *Physiol Rev* **95**, 549–601.

Kindig AE, Heller TB & Kaufman MP (2005). VR-1 receptor blockade attenuates the pressor response to capsaicin but has no effect on the pressor response to contraction in cats. *Am J Physiol Heart Circ Physiol* **288**, H1867–H1873.

Klingel SL, Roke K, Hidalgo B, Aslibekyan S, Straka RJ, An P, Province MA, Hopkins PN, Arnett DK, Ordovas JM, Lai C-Q & Mutch DM (2017). Sex differences in blood HDL-c, the total cholesterol/HDL-c ratio, and palmitoleic acid are not associated with variants in common candidate genes. *Lipids* **52**, 969–980.

Ko Y-L, Hsu L-A, Wu S, Teng M-S, Chang H-H, Chen C-C & Cheng C-F (2008). Genetic variation in the *ASIC3* gene influences blood pressure levels in Taiwanese. *J Hypertens* **26**, 2154–2160.

Langberg H, Bjørn C, Boushel R, Hellsten Y & Kjaer M (2002). Exercise-induced increase in interstitial bradykinin and adenosine concentrations in skeletal muscle and peritendinous tissue in humans. *J Physiol* **542**, 977–983.

Mark AL, Victor RG, Nerhed C & Wallin BG (1985). Microneurographic studies of the mechanisms of sympathetic nerve responses to static exercise in humans. *Circ Res* **57**, 461–469.

Messeguer A, Planells-Cases R & Ferrer-Montiel A (2006). Physiology and pharmacology of the vanilloid receptor. *Curr Neuropharmacol* **4**, 1–15.

Miyai N, Arita M, Miyashita K, Morioka I, Shiraishi T & Nishio I (2002). Blood pressure response to heart rate during exercise test and risk of future hypertension. *Hypertension* **39**, 761–766.

Miyai N, Arita M, Morioka I, Miyashita K, Nishio I & Takeda S (2000). Exercise BP response in subjects with high-normal BP Exaggerated blood pressure response to exercise and risk of future hypertension in subjects with high-normal blood pressure. *J Am Coll Cardiol* **36**, 1626–1631.

Mizuno M, Murphy MN, Mitchell JH & Smith SA (2011). Antagonism of the TRPv1 receptor partially corrects muscle metaboreflex overactivity in spontaneously hypertensive rats. *J Physiol* **589**, 6191–6204.

Nieminen T, Lehtimäki T, Laiho J, Rontu R, Niemelä K, Kööbi T, Lehtinen R, Viik J, Turjanmaa V & Kähönen M (2006). Effects of polymorphisms in β_1 -adrenoceptor and α -subunit of G protein on heart rate and blood pressure during exercise test. The Finnish Cardiovascular Study. *J Appl Physiol* **100**, 507–511.

Notarius CF, Atchison DJ & Floras JS (2001). Impact of heart failure and exercise capacity on sympathetic response to handgrip exercise. *Am J Physiol Heart Circ Physiol* **280**, H969–H976.

Notarius CF, Millar PJ, Murai H, Morris BL, Marzolini S, Oh P & Floras JS (2015). Divergent muscle sympathetic responses to dynamic leg exercise in heart failure and age-matched healthy subjects. *J Physiol* **593**, 715–722.

Notay K, Lee JB, Incognito AV, Seed JD, Arthurs AA & Millar PJ (2018). Muscle strength influences pressor responses to static handgrip in men and women. *Med Sci Sports Exerc* **50**, 778–784.

Nunes RAB, Barroso LP, Pereira AdaC, Krieger JE & Mansur AJ (2014). Gender-related associations of genetic polymorphisms of α-adrenergic receptors, endothelial nitric oxide synthase and bradykinin B2 receptor with treadmill exercise test responses. *Open Heart* **1**, e000132.

Nunes RAB, Pereira Barroso L, da Costa Pereira A, Pinto Brandão Rondon MU, Negrão CE, Krieger JE & Mansur AJ (2016). Alpha2A-adrenergic receptor and eNOS genetic polymorphisms are associated with exercise muscle vasodilatation in apparently healthy individuals. *Int J Cardiol Heart Vasc* **13**, 14–18.

Pan HL, Stebbins CL & Longhurst JC (1993). Bradykinin contributes to the exercise pressor reflex: mechanism of action. *J Appl Physiol* **75**, 2061–2068.

Parmar HR, Sears J, Molgat-Seon Y, McCulloch CL, McCracken LA, Brown CV, Sheel AW, Dominelli PB (2018). Oral contraceptives modulate the muscle metaboreflex in healthy young women. *Appl Physiol Nutr Metab* **43**, 460–466.

Rett K, Wicklmayr M, Fink E, Maerker E, Dietze G & Mehnert H (1989). Local generation of kinins in working skeletal muscle tissue in man. *Biol Chem Hoppe Seyler* **370**, 445–449.

Rotto DM & Kaufman MP (1988). Effect of metabolic products of muscular contraction on discharge of group III and IV afferents. *J Appl Physiol* **64**, 2306–2313.

Rotto DM, Massey KD, Burton KP & Kaufman MP (1989). Static contraction increases arachidonic acid levels in gastrocnemius muscles of cats. *J Appl Physiol* **66**, 2721–2724.

Sato M, Nakayama T, Soma M, Aoi N, Kosuge K, Haketa A, Izumi Y, Matsumoto K, Sato N & Kokubun S (2007). Association between prostaglandin E2 receptor gene and essential hypertension. *Prostaglandins Leukot Essent Fatty Acids* **77**, 15–20.

Schultz MG & Sharman JE (2014). Exercise hypertension. *Pulse* **1**, 161–176.

Shastry BS (2009). SNPs: impact on gene function and phenotype. *Methods Mol Biol* **578**, 3–22.

Smith SA, Leal AK, Williams MA, Murphy MN, Mitchell JH & Garry MG (2010). The TRPv1 receptor is a mediator of the exercise pressor reflex in rats. *J Physiol* **588**, 1179–1189.

Stebbins CL, Carretero OA, Mindroiu T & Longhurst JC (1990). Bradykinin release from contracting skeletal muscle of the cat. *J Appl Physiol* **69**, 1225–1230.

Stebbins CL, Maruoka Y & Longhurst JC (1986). Prostaglandins contribute to cardiovascular reflexes evoked by static muscular contraction. *Circ Res* **59**, 645–654.

Stokes L, Scurrah K, Ellis JA, Cromer BA, Skarratt KK, Gu BJ, Harrap SB & Wiley JS (2011). A loss-of-function polymorphism in the human P2X4 receptor is associated with increased pulse pressure. *Hypertension* **58**, 1086–1092.

- Stone AJ, Copp SW, Kim JS & Kaufman MP (2015). Combined, but not individual, blockade of ASIC3, P2X, and EP4 receptors attenuates the exercise pressor reflex in rats with freely perfused hindlimb muscles. *J Appl Physiol* **119**, 1330–1336.
- Storm N, Darnhofer-Patel B, van den Boom D & Rodi CP (2003). MALDI-TOF mass spectrometry-based SNP genotyping. *Methods Mol Biol* **212**, 241–262.
- Symons JD, Theodossy SJ, Longhurst JC & Stebbins CL (1991). Intramuscular accumulation of prostaglandins during static contraction of the cat triceps surae. *J Appl Physiol* **71**, 1837–1842.
- Ueno LM, Frazzatto EST, Batalha LT, Trombetta IC, do Socorro Brasileiro M, Irigoyen C, Brum PC, Villares SMF & Negrão CE (2005). α_{2B} -Adrenergic receptor deletion polymorphism and cardiac autonomic nervous system responses to exercise in obese women. *Int J Obes* **30**, 214–220.
- Vianna LC, Fernandes IA, Barbosa TC, Teixeira AL & Claudio Lucas da Nóbrega A (2018). Capsaicin-based analgesic balm attenuates the skeletal muscle metaboreflex in healthy humans. *J Appl Physiol* **125**, 362–368.
- Wang S, Joseph J, Diatchenko L, Ro JY & Chung M-K (2016). Agonist-dependence of functional properties for common nonsynonymous variants of human transient receptor potential vanilloid 1. *Pain* **157**, 1515–1524.
- Weiss SA, Blumenthal RS, Sharrett AR, Redberg RF & Mora S (2010). Exercise blood pressure and future cardiovascular death in asymptomatic individuals. *Circulation* **121**, 2109–2116.
- Williamson JW (2010). The relevance of central command for the neural cardiovascular control of exercise. *Exp Physiol* **95**, 1043–1048.

Additional information

Competing interests

The authors declare no conflicts ofinterest relevant to the content of this study.

Author contributions

K.N. and P.J.M. conceived and designed the research; K.N., S.L.K., J.B.L., C.J.D., J.D.S. and P.J.M. performed the experiments; K.N. and M.S. analysed the data; K.N., D.M.M. and P.J.M. interpreted the results; K.N. prepared the figures and drafted the manuscript; K.N., D.M.M. and P.J.M. edited and revised the manuscript. All authors have read and approved the final version of this manuscript and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

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