GENOME SEQUENCES





Complete Genome Sequence of a Vancomycin-Resistant Sequence Type 203 *Enterococcus faecium* Strain with *vanA* Belonging to Complex Type 859

Maria Hoffmann,^a Kuan Yao,^a Marc Allard,^a Maria Sanchez,^a Leif P. Andersen,^b Henrik Hasman,^c Anette M. Hammerum^c

^aU.S. Food and Drug Administration, Center for Food Safety and Applied Nutrition, College Park, Maryland, USA

^bDepartment of Clinical Microbiology, Rigshospitalet, Copenhagen, Denmark

^cDepartment of Bacteria, Parasites and Fungi, Statens Serum Institut, Copenhagen, Denmark

ABSTRACT In 2014, the first vancomycin-resistant (encoded by *vanA*) *Enterococcus faecium* isolate belonging to sequence type 203 (ST203) and complex type 859 (CT859) was detected in Denmark. In 2016, 64% of the Danish clinical *vanA E. faecium* isolates belonged to ST203 and CT859. Using Pacific Biosciences (PacBio) RS II sequencing, we describe the genome of ST203 CT859 *vanA E. faecium*.

n Denmark, *vanA Enterococcus faecium* is most prevalent among clinical vancomycinresistant *Enterococcus* (VRE) isolates (1).

Here, we present the complete genome sequence of a vancomycin-resistant *Entero-coccus faecium* (VREfm) isolate, including its three plasmids, isolated from urine from a patient hospitalized at Rigshospitalet in Copenhagen, Denmark, in October 2014. *E. faecium* VRE1589 was the earliest *vanA E. faecium* isolate belonging to sequence type 203 and complex type 859 (ST203-CT859) identified among the VRE isolates submitted to Statens Serum Institut as part of the Danish national surveillance program. Comparison to the cgMLST.org database (https://cgmlst.org/ncs), previous Danish studies, and personal communications with neighboring countries with whole-genome sequencing (WGS) data available suggests that the novel complex type (CT859) emerged in October 2014 in Copenhagen and spread to the rest of Denmark, the south of Sweden, and the Faroe Islands during 2015 (1). In 2016, 64% of the clinical *vanA E. faecium* isolates detected in Denmark belonged to ST203 and CT859 (2).

Previous analyses showed that the *E. faecium* isolate VRE1589 belonged to ST203 and to a new complex type, CT859, based on the MLST version 1.8 Web server (3) and SeqSphere+ version 3.4.0 (Ridom GmbH, Münster, Germany [http://www.ridom.de/ seqsphere/]) (3). For additional characterization of *E. faecium* VRE1589, we sequenced the isolate on a PacBio RS II sequencer. The genomic DNA was extracted with a DNeasy blood and tissue kit (Qiagen, Hilden, Germany) and sheared into approximately 20-kb fragments using g-TUBE (Covaris, Inc., Woburn, MA, USA). The library was prepared based on the 20-kb PacBio sample preparation protocol and sequenced using P6-C4 chemistry on three single-molecule real-time (SMRT) cells with a 240-min collection time. The continuous long reads were *de novo* assembled by the PacBio Hierarchical Genome Assembly Process 3.0 (HGAP3.0) program. Genomes were annotated using the National Center for Biotechnology Information (NCBI) Prokaryotic Genome Automated Pipeline version 2.9 (4) and on the RAST annotation server (http://rast.nmpdr.org/).

The presence of resistance genes was assessed using ResFinder version 2.1 (5), and plasmid replicons were detected using PlasmidFinder version 1.3 (5). The *vanA* gene was detected as part of Tn*1546* on a 49,542-bp plasmid (pVRE1589_p2) carrying three replicons similar to rep1, rep2, and rep7, with GenBank accession numbers AF007787,

Received 7 June 2018 Accepted 24 June 2018 Published 19 July 2018

Citation Hoffmann M, Yao K, Allard M, Sanchez M, Andersen LP, Hasman H, Hammerum AM. 2018. Complete genome sequence of a vancomycin-resistant sequence type 203 *Enterococcus faecium* strain with vanA belonging to complex type 859. Microbiol Resour Announc 7:e00815-18. https://doi.org/ 10.1128/MRA.00815-18.

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine

Copyright © 2018 Hoffmann et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Anette M. Hammerum, ama@ssi.dk. LT986680, and U01917, respectively. Besides *vanA*, three additional resistance genes were detected on pVRE1589_p2, *ant(6)-la*, *erm*(B), and *cat*(pC221). Furthermore, we detected three other resistance genes integrated on the chromosome, *msr*(C), *tet*(M), and *dfrG*. The isolate also contained a 217,014-bp plasmid (pVRE1589_p1) with a repUS15 replicon (accession number CP003586) and a 1,017-bp plasmid (pVRE1589_p3) with a rep14 replicon (GenBank accession number EFU01917); none of these plasmids contained any known resistance genes.

The origin of the ST203-CT859 vanA E. faecium isolate is unknown. Further studies might show why this complex type was so successful and where it originated.

Data availability. This complete genome project has been deposited at GenBank under accession numbers CP020484 to CP020487.

ACKNOWLEDGMENTS

We thank Karin Sixhøj Pedersen for her excellent technical assistance at SSI. This work was supported by the Danish Ministry of Health and the U.S. FDA, Center for Food Safety and Applied Nutrition.

REFERENCES

- Hammerum AM, Baig S, Kamel Y, Roer L, Pinholt M, Gumpert H, Holzknecht B, Røder B, Justesen US, Samulioniené J, Kjærsgaard M, Østergaard C, Holm A, Dzajic E, Søndergaard TS, Gaini S, Edquist P, Alm E, Lilje B, Westh H, Stegger M, Hasman H. 2017. Emergence of vanA Enterococcus faecium in Denmark, 2005–15. J Antimicrob Chemother 72:2184. https:// doi.org/10.1093/jac/dkx138.
- Høg BB, Korsgaard H, Sönksen UW (ed). 2016. DANMAP 2015: use of antimicrobial agents and occurrence of antimicrobial resistence in bacteria from food animals, food and humans in Denmark. DANMAP—The Danish Integrated Antimicrobial Monitoring and Research Programme, Copenhagen, Denmark.
- 3. de Been M, Pinholt M, Top J, Bletz S, Mellmann A, van Schaik W, Brouwer

E, Rogers M, Kraat Y, Bonten M, Corander J, Westh H, Harmsen D, Willems RJL. 2015. Core genome multilocus sequence typing scheme for high-resolution typing of *Enterococcus faecium*. J Clin Microbiol 53:3788–3797. https://doi.org/10.1128/JCM.01946-15.

- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (Meta)genomic annotation. OMICS 12:137–141. https://doi.org/ 10.1089/omi.2008.0017.
- Carattoli A, Bertini A, Villa L, Falbo V, Hopkins KL, Threlfall EJ. 2005. Identification of plasmids by PCR-based replicon typing. J Microbiol Methods 63:219–228. https://doi.org/10.1016/j.mimet.2005.03.018.