

Salmonella enterica serovar Typhimurium isolated from the urine of a dog undergoing treatment for immune-mediated polyarthritis

Stephen D. Cole,^{1,*} Shannon M. Palermo² and Shelley C. Rankin¹

Abstract

Introduction. In people, *Salmonella* is a common agent of gastroenteritis, but it can also cause extraintestinal disease such as urinary tract infections. In addition, *Salmonella* is often linked to the post-infection development of reactive arthritis. In canines, cases that document extraintestinal *Salmonella* infections or diseases similar to reactive arthritis have not been thoroughly described.

Case presentation. A case of a 5-year-old German shepherd dog with *Salmonella* bacteriuria during treatment for immune-mediated polyarthritis (IMPA) is described. The patient first suffered from a 3 month period of diarrhoea and presented for evaluation of a 2 month history of shifting-leg lameness. A diagnosis of IMPA was made based on cytological examination and negative synovial fluid culture. Treatment with immunosuppressive doses of prednisone lead to clinical resolution of lameness, but on a recheck abnormal urine was noted. *Salmonella enterica* serovar Typhimurium was isolated using standard culture methods. The patient was treated with enrofloxacin to control the bacteriuria.

Conclusion. This case report is, to the best of our knowledge, the first to describe *Salmonella* bacteriuria in a dog and suggests that *Salmonella* infection may be a potential inciting factor for IMPA.

INTRODUCTION

Salmonella enterica is a Gram-negative pathogen and a major cause of food-borne illness [1]. Non-typhoidal *Salmonella* serovars typically cause self-limiting diarrhoea in veterinary and human patients, but chronic infections have also been reported [2–4]. Extraintestinal infections are rare in both people and dogs, but may include endocarditis [5], pneumonia [6], discospondylitis [7] and urinary tract infection [8]. In addition, one of many possible sequelae of *Salmonella* enteritis in people is reactive arthritis that is characterized as post-infectious, suppurative inflammation that may affect one or multiple joints [9]. A systematic review calculated a weighted mean incidence of reactive arthritis cases of 12/1000 in human enteric salmonellosis patients [10].

A similar condition called immune-mediated polyarthritis (IMPA) occurs in dogs, but an underlying cause is rarely identified [11]. IMPA is considered a type III hypersensitivity reaction with immune complex deposition [11]. IMPA is divided into four types: idiopathic (I), reactive (II),

gastrointestinal (III) and neoplastic (IV) [11]. Specifically, type II has been linked to extra-articular infections (i.e. tick-borne disease, prostatitis, necrotizing pharyngitis) and type III to gastrointestinal or hepatic disease [11]. In the veterinary literature, to the authors' knowledge, there are no case reports of *Salmonella* as an agent of bacteriuria in a dog and no case reports that have linked IMPA with *Salmonella* infection.

CASE REPORT

A 5-year-old, castrated-male German shepherd dog was presented for an approximately 2 month history of alternating hindlimb and forelimb lameness. On physical examination, the carpi and elbows were warm, painful and effusive bilaterally. Cytological examination of synovial fluid obtained via arthrocentesis from the left and right elbows and carpi revealed suppurative inflammation, but no infectious agents were identified. Culture of synovial fluid was negative by aerobic culture. The dog also had intermittent, chronic diarrhoea for approximately 3 months and had lost approximately 23% (10 kg) of its overall body weight.

Received 22 November 2017; Accepted 27 April 2018

Author affiliations: ¹Department of Pathobiology, School of Veterinary Medicine, University of Pennsylvania, Philadelphia, PA, USA; ²Department of Clinical Studies - Philadelphia, School of Veterinary Medicine, University of Pennsylvania, Philadelphia, PA, USA.

***Correspondence:** Stephen D. Cole, scole@vet.upenn.edu

Keywords: *Salmonella*; bacteriuria; immune-mediated polyarthritis; diarrhoea.

Abbreviation: IMPA, immune-mediated polyarthritis.

Faecal culture was negative for *Campylobacter* and *Salmonella*, and a faecal float was free of parasites. The patient was antibody negative for *Ehrlichia*, *Anaplasma* and *Borrelia* by SNAP 4Dx (Idexx Laboratories) and *Bartonella* by Western blot (National Veterinary Laboratory, Franklin Lakes, NJ, USA). Urinalysis was not performed initially. The patient was diagnosed with IMPA and treated with immunosuppressive doses of prednisone (1.1 mg kg⁻¹ twice daily for approximately 5 months, including a tapered regimen). Tylosin powder [Elanco (Tylan Powder)] (1/2 teaspoon once daily for 21 days) and omeprazole [Procter and Gamble (Prilosec OTC)] (1 mg kg⁻¹ once daily pro re nata) were prescribed for the chronic diarrhoea. The owner also began a commercial, limited ingredient diet (Blue Buffalo).

At recheck, 10 weeks after diagnosis of IMPA, the patient's orthopaedic pain was well managed, the effusion had grossly resolved and the diarrhoea had subsided. While the patient did not exhibit any signs of lower urinary tract disease, malodorous urine was noted during the examination.

INVESTIGATIONS

A urinalysis revealed dilute urine with pyuria (6–10 per high powered field) and bacteriuria. Quantitative aerobic culture of urine obtained via cystocentesis yielded greater than 100 000 c.f.u. ml⁻¹ of a non-lactose-fermenting organism on MacConkey agar (Remel). The organism was biochemically identified as *Salmonella* spp. with the MicroScan Walkway 40si NC31 panel (Dade-Behring, Deerfield, IL), and was sensitive to all antimicrobials tested, including amoxicillin, trimethoprim/sulfamethoxazole and enrofloxacin [12]. The organism was serotyped as *S. enterica* serovar Typhimurium (4,12:i:1,2) using the xMAP *Salmonella* serotyping assay (Luminex).

TREATMENT

The patient completed a 21 day course of oral enrofloxacin (Luminex) (10 mg kg⁻¹ SID). Although the animal may have had subclinical bacteriuria according the International Society for Companion Animal Infectious Disease guidelines, and therapy may not have been warranted, the clinicians felt it prudent to treat given the zoonotic potential of the pathogen and the immunocompromised nature of the patient [13]. A fluoroquinolone antimicrobial was selected based on previous reports in the human literature [14, 15]. The authors acknowledge that amoxicillin or trimethoprim-sulfamethoxazole may have been a more judicious choice and should be considered in future cases. At the time of treatment, appropriate at-home interventions were put in place to prevent zoonotic transfer to the owners. Specifically, gloves were recommended for handling faecal material, it was recommended that contact with urine be avoided and it was advised that immunocompromised individuals should not be allowed to come into contact with the dog.

OUTCOME AND FOLLOW-UP

Seven-day follow-up urine and faecal cultures were negative for *Salmonella*. At the time of writing (approximately 10 months from initial presentation) the patient was clinically normal.

DISCUSSION

It has been postulated that domestic animals can serve as a reservoir of enteric agents of zoonotic disease (i.e. *Salmonella*, *Campylobacter*, possibly *Clostridium difficile*). A recent multi-laboratory study reported a 3.8 % prevalence of *Salmonella* in the faeces of diarrhoeic dogs, which was significantly different from the 1.8 % in non-diarrhoeic dogs [16]. *S. enterica* serovar Typhimurium was not among the serovars isolated [17]. A major risk factor for *Salmonella* infection in dogs is known to be the feeding of raw diets [17, 18]. The patient in this study was fed a common, commercial dry-food diet and there was no history of eating raw food. A single case of *Salmonella* bacteriuria in a cat has been linked to feeding a raw diet [19]. *S. enterica* serovar Typhimurium has not been reported to be one of the most common serotypes from dogs [17, 20], but it has been linked to several outbreaks that involved people and domestic animals (dogs and cats). These outbreaks were linked to cats that hunt wild birds [21], veterinary patients undergoing dental procedures at the same hospital [22] and animal shelters [23].

Definitive diagnosis of the aetiology of the gastrointestinal disease was never made; therefore, classifying the IMPA type is difficult. This patient could traditionally be classified as having type III (gastrointestinal) IMPA. The corticosteroids administered for IMPA may have also treated concurrent inflammatory bowel disease, leading to the resolution of diarrhoea [24]. The authors also suggest that there may be crossover with type II (reactive), since it is possible that *Salmonella* was responsible for the gastrointestinal disease. Although the faecal culture was negative for *Salmonella*, recovery of *Salmonella* from faeces can be variable depending on the technique used [25] and due to intermittent shedding [26]. To rule out *Salmonella* as an agent of enteritis, the American College of Veterinary Internal Medicine consensus statement recommends multiple faecal cultures if molecular detection (PCR) is not performed [16]. In retrospect, an approach such as this may have been helpful to confirm *Salmonella* as the cause of enteritis in this dog. Further research could evaluate the prevalence of *Salmonella* and other enteric pathogens in dogs with IMPA. Infectious agents that may be associated with IMPA in dogs are often vector-borne agents (i.e. *Ehrlichia*, *Borrelia*, *Bartonella*) [27], but greater understanding of all pathogens associated with IMPA may help elucidate its pathogenesis. It is likely that corticosteroid-mediated immunosuppression predisposed this patient to ascending bacteriuria [28].

In summary, this case report identifies and describes *Salmonella* from the urinary tract of a dog with chronic diarrhoea and IMPA. Given this report and the relationship of

Salmonella with reactive arthritis in people, owners should be asked about historical diarrhoea when patients have suspected or diagnosed IMPA. In patients with current or previous diarrhoea, the authors suggest thorough diagnostics such as serial faecal cultures or PCR. Identification of *Salmonella* in patients with IMPA could have important therapeutic implications and allow for implementation of appropriate measures to prevent zoonotic transfer. This report also suggests that the phenomenon of reactive arthritis following *Salmonella* infection may not be limited to people.

Funding information

The authors received no specific grant from any funding agency.

Acknowledgements

The authors acknowledge Kathleen Boyajian for performing molecular serotyping of the isolate.

Conflicts of interest

The authors declare that there are no conflicts of interest.

Ethical statement

The authors declare that they have adhered to all ethical guidelines required by their institution.

References

- Painter JA, Hoekstra RM, Ayers T, Tauxe RV, Braden CR et al. Attribution of foodborne illnesses, hospitalizations, and deaths to food commodities by using outbreak data, United States, 1998–2008. *Emerg Infect Dis* 2013;19:407–415.
- Marks SL, Rankin SC, Byrne BA, Weese JS. Enteropathogenic bacteria in dogs and cats: diagnosis, epidemiology, treatment, and control. *J Vet Intern Med* 2011;25:1195–1208.
- Morse EV, Duncan MA. Canine salmonellosis: prevalence, epizootiology, signs, and public health significance. *J Am Vet Med Assoc* 1975;167:817–820.
- Keithlin J, Sargeant JM, Thomas MK, Fazil A. Systematic review and meta-analysis of the proportion of non-typhoidal *Salmonella* cases that develop chronic sequelae. *Epidemiol Infect* 2015;143:1333–1351.
- Ortiz D, Siegal EM, Kramer C, Khandheria BK, Brauer E et al. Nontyphoidal cardiac salmonellosis: two case reports and a review of the literature. *Tex Heart Inst J* 2014;41:401–406.
- Aguado JM, Obeso G, Cabanillas JJ, Fernández-Guerrero M, Alés J. Pleuropulmonary infections due to nontyphoid strains of *Salmonella*. *Arch Intern Med* 1990;150:54–56.
- Plessas IN, Jull P, Volk HA. A case of canine discospondylitis and epidural empyema due to *Salmonella* species. *Can Vet J* 2013;54:595–598.
- Abbott SL, Portoni BA, Janda JM. Urinary tract infections associated with nontyphoidal *Salmonella* serogroups. *J Clin Microbiol* 1999;37:4177–4178.
- Samuel MP, Zwillich SH, Thomson GT, Alfa M, Orr KB et al. Fast food arthritis – a clinico-pathologic study of post-*Salmonella* reactive arthritis. *J Rheumatol* 1995;22:1947–1952.
- Ajene AN, Fischer Walker CL, Black RE. Enteric pathogens and reactive arthritis: a systematic review of *Campylobacter*, *Salmonella* and *Shigella*-associated reactive arthritis. *J Health Popul Nutr* 2013;31:299–307.
- Rondeau MP, Walton RM, Bissett S, Drobatz KJ, Washabau RJ et al. Suppurative, nonseptic polyarthropathy in dogs. *J Vet Intern Med* 2005;19:654–662.
- Clinical and Laboratory Standards Institute. *Performance Standards for Antimicrobial Disk and Dilution Susceptibility Tests for Bacteria Isolated from Animals*, 4th ed. CLSI document Vet 01-A4. Wayne, PA: Clinical and Laboratory Standards Institute; 2016.
- Weese JS, Blondeau JM, Boothe D, Breitschwerdt EB, Guardabassi L et al. Antimicrobial use guidelines for treatment of urinary tract disease in dogs and cats: antimicrobial guidelines working group of the international society for companion animal infectious diseases. *Vet Med Int* 2011;2011:1–9.
- Tena D, González-Praetorius A, Bisquert J. Urinary tract infection due to non-typhoidal *Salmonella*: report of 19 cases. *J Infect* 2007;54:245–249.
- Jehangir A, Poudel D, Fareedy SB, Salman A, Qureshi A et al. Group D *Salmonella* urinary tract infection in an immunocompetent male. *Case Rep Infect Dis* 2015;2015:608632.
- Marks SL, Rankin SC, Byrne BA, Weese JS. Enteropathogenic bacteria in dogs and cats: diagnosis, epidemiology, treatment, and control. *J Vet Intern Med* 2011;25:1195–1208.
- Reimschuessel R, Grabenstein M, Guag J, Nemser SM, Song K et al. Multilaboratory survey to evaluate *Salmonella* prevalence in diarrheic and nondiarrheic dogs and cats in the United States between 2012 and 2014. *J Clin Microbiol* 2017;55:1350–1368.
- Leonard EK, Pearl DL, Finley RL, Janecko N, Peregrine AS et al. Evaluation of pet-related management factors and the risk of *Salmonella* spp. carriage in pet dogs from volunteer households in Ontario (2005–2006). *Zoonoses Public Health* 2011;58:140–149.
- Fauth E, Freeman LM, Cornjeo L, Markovich JE, Janecko N et al. *Salmonella* bacteriuria in a cat fed a *Salmonella*-contaminated diet. *J Am Vet Med Assoc* 2015;247:525–530.
- Leahy AM, Cummings KJ, Rodriguez-Rivera LD, Rankin SC, Hamer SA et al. Evaluation of faecal *Salmonella* shedding among dogs at seven animal shelters across Texas. *Zoonoses Public Health* 2016;63:515–521.
- Tauni MA, Osterlund A. Outbreak of *Salmonella typhimurium* in cats and humans associated with infection in wild birds. *J Small Anim Pract* 2000;41:339–341.
- Cherry B, Burns A, Johnson GS, Pfeiffer H, Dumas N et al. *Salmonella* Typhimurium outbreak associated with veterinary clinic. *Emerg Infect Dis* 2004;10:2249–2251.
- Wright JG, Tengelsen LA, Smith KE, Bender JB, Frank RK et al. Multidrug-resistant *Salmonella* Typhimurium in four animal facilities. *Emerg Infect Dis* 2005;11:1235–1241.
- Craven M, Simpson JW, Ridy AE et al. Canine inflammatory bowel disease. *J Small Anim Pract* 2004;45:336–342.
- Dusch H, Altwegg M. Evaluation of five new plating media for isolation of *Salmonella* species. *J Clin Microbiol* 1995;33:802–804.
- Tanaka Y, Katsube Y, Imaizumi K. Experimental carrier in dogs produced by oral administration of *Salmonella typhimurium*. *Nihon Juigaku Zasshi* 1976;38:569–578.
- Rhoades AC, Vernau W, Kass PH, Herrera MA, Sykes JE et al. Comparison of the efficacy of prednisone and cyclosporine for treatment of dogs with primary immune-mediated polyarthritis. *J Am Vet Med Assoc* 2016;248:395–404.
- Torres SM, Diaz SF, Nogueira SA, Jessen C, Polzin DJ et al. Frequency of urinary tract infection among dogs with pruritic disorders receiving long-term glucocorticoid treatment. *J Am Vet Med Assoc* 2005;227:239–243.