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# Stress, inflammation, and eicosanoids: an emerging perspective

Sujanitha Umamaheswaran<sup>#1,2</sup>, Santosh K. Dasari<sup>#1</sup>, Peiying Yang<sup>3</sup>, Susan K. Lutgendorf<sup>4,5,6,7</sup>, and Anil K. Sood<sup>1,2,8</sup>

<sup>1</sup>Department of Gynecologic Oncology and Reproductive Medicine, The University of Texas MD Anderson Cancer Center, Unit 1362, 1515 Holcombe Blvd., Houston, TX 77030, USA

<sup>2</sup>Department of Cancer Biology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>3</sup>Department of Palliative, Rehabilitation and Integrative Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>4</sup>Department of Psychological and Brain Sciences, University of Iowa, Iowa City, IA, USA

<sup>5</sup>Department of Obstetrics and Gynecology, University of Iowa, Iowa City, IA, USA

<sup>6</sup>Department of Urology, University of Iowa, Iowa City, IA, USA

<sup>7</sup>Holden Comprehensive Cancer Center, University of Iowa, Iowa City, IA, USA

<sup>8</sup>Center for RNA Interference and Non-coding RNA, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>#</sup> These authors contributed equally to this work.

# Abstract

Clinical and experimental studies support the notion that adrenergic stimulation and chronic stress affect inflammation, metabolism, and tumor growth. Eicosanoids are also known to heavily influence inflammation while regulating certain stress responses. However, additional work is needed to understand the full extent of interactions between the stress-related pathways and eicosanoids. Here, we review the potential influences that stress, inflammation, and metabolic pathways have on each other, in the context of eicosanoids. Understanding the intricacies of such interactions could provide insights on how systemic metabolic effects mediated by the stress pathways can be translated into therapies for cancer and other diseases.

# Keywords

Eicosanoids; PGE<sub>2</sub>; Inflammation; Stress; Cancer

Correspondence to: Anil K. Sood.

Anil K. Sood: asood@mdanderson.org. Sujanitha Umamaheswaran and Santosh K. Dasari are co-first authors

# 1 Introduction

Psychological stress and environmental factors trigger a cascade of pathways in the central nervous system (CNS) and periphery, which subsequently activate stress responses in the autonomic nervous system (ANS) and hypothalamic-pituitary-adrenal (HPA) axis [1, 2]. Hypothalamic production of corticotrophin-releasing factor and arginine vasopressin mediates HPA responses by inducing adrenocorticotropic hormone (ACTH). ACTH mediates downstream release of glucocorticoids by the adrenal cortex, which plays a crucial role in regulating stress responses [3]. Activation of the sympathetic nervous system (SNS) and downstream release of catecholamines from sympathetic neurons and adrenal medulla form the ANS response cascade to stress. Activation of these pathways enable an individual to overcome threat and stressful situations. Stressed individuals are known to have higher levels of catecholamines along with ANS-associated effects on their respiratory, vascular, and cardiac systems [3]. However, chronic stress and the associated prolonged exposure to stress mediators (e.g., glucocorticoids and catecholamines) can have more profound effects on biological systems [4]. Epidemiological data show that social characteristics and chronic stress impact cancer progression. While some studies suggest that stressful life events and other psychosocial factors may correlate with increased risk of cancer [5], others have not found such links [6, 7]. Chronic stress and lack of social support increases breast cancer incidence by almost ninefold [8]. A meta-analysis of 165 studies concluded that psychosocial factors may be associated with cancer incidence and survival [9].

The role of psychosocial factors and stress in affecting disease outcome is an area of longstanding interest. In 1936, Hans Selye identified the activating influence of stress on physiological systems that not only transiently protect and restore the body, but also damage it. [10, 11]. Research over the years that followed has shown that stress impacts multiple biological processes, including inflammation, metabolism, and malignant progression [12]. Clinical and pre-clinical studies in various malignancies have shown that chronic psychological stress influences disease progression, relapse, and response to therapy [13– 18]. Sustained adrenergic signaling mediated by increased norepinephrine levels drives both tumor inflammation and aberrant arachidonic acid metabolism in cancer [19]. Stress responses occurring *via* the ANS are also known to directly affect immune cell number and function [20].

Stress mediators and their receptors influence various pathways that modulate tumor biology. cAMP responsive element-binding (CREB) protein is activated by signaling pathways in response to stress hormones and promotes proliferation, migration, angiogenesis, and apoptosis evasion [21]. Viral infections play a major role in tumor initiation, and adrenergic or glucocorticoid-induced signaling activates many tumorassociated viruses [1]. Phospholipase  $C\gamma$  and protein kinase Ca upregulation by norepinephrine promotes cell migration, and *in vitro* invasiveness of ovarian cancer and nasopharyngeal carcinoma also increases in the presence of norepinephrine [22, 23]. Catecholamines activate signal transducer and activator of transcription factor-3 (STAT3), which contributes to tumor growth, progression, angiogenesis, and apoptosis suppression [1, 24]. Neuroendocrine stress also activates hormones such as prolactin, which is known to promote tumor growth and cell survival in breast and other cancers [1].

Bioactive lipids have been recognized to promote processes associated with malignant progression. For example, up-regulation of sphingosine kinase 1 (SPHK1) in gastric cancer patients is associated with poor survival and increased progression [25]. The eicosanoid PGE<sub>2</sub> promotes colorectal cancer progression and inhibits activity of anti-tumor therapy like celecoxib in mouse models [26, 27]. Similarly, lipids are also known to promote chronic inflammation, a process that drives carcinogenesis. In this review, we discuss the complex interactions between inflammation, stress, and metabolic changes, with an emphasis on arachidonic acid metabolism and eicosanoids.

# 2 Stress and metabolism

Arachidonic acid is an essential dietary fatty acid and its derivatives have an important influence on cancer and inflammation. Arachidonic acid metabolism *via* cyclooxygenase (COX), lipoxygenase (LOX), and P450 epoxygenase pathways produces eicosanoids, including prostanoids, leukotrienes, and thromboxanes [28] (Fig. 1). Cyclooxygenase exists in two iso-forms, namely COX-1 and COX-2. While COX-1 is a constitutive enzyme in most cells in basal state, COX-2 is present in insignificant amounts which is inducible by cytokines and other inflammatory stimuli [28]. Importantly, COX-2 overexpression led to spontaneous development of pancreatic ductal adenocarcinoma in mice, which regressed completely upon COX-2 selective inhibitor (celecoxib) treatment [29].

Non-steroidal anti-inflammatory drugs (NSAIDs) are effective inhibitors of the COX pathway, with most NSAIDs being non-selective COX-1 and COX-2 inhibitors. LOX pathway mainly operates in the lung, white blood cells, and platelets; its most significant metabolites are leukotrienes [30]. Epidemiological and clinical evidence implicates COX and LOX pathways in chronic inflammation and carcinogenesis caused by aberrant arachidonic acid metabolism [31, 32]. Many population-based epidemiological studies, focused mainly on colorectal cancer, showed a lower incidence upon long-term NSAID use [33, 34]. A comprehensive meta-analysis of published work from 1980 to 2008 showed an inverse correlation between long-term NSAID use and incidence of colon, breast, prostate, and lung cancers [35, 36]. Detailed analysis showed that regular intake of NSAIDs caused risk reductions of up to 43% in colon cancer, 28% in lung cancer, 27% in prostate cancer, and 25% in breast cancer [35]. Among the various arachidonic acid metabolites, prostaglandin  $E_2$  (PGE<sub>2</sub>) is the most widely investigated, especially in the context of the chronic stress. It is a bioactive arachidonic acid-derivative cyclooxygenase metabolite which exerts its activity via four G-protein-coupled receptors (EP1, EP2, EP3, and EP4) and produced abundantly in the brain in response to stress and COX-2 dysregulation. Studies have shown that  $PGE_2$  regulates stress responses, immunity, and inflammatory pathways [37, 38].

#### 2.1 Glucocorticoid influences

Studies have shown glucocorticoids to have a suppressive effect on eicosanoid synthesis. Therapeutic doses of the corticosteroid drug prednisone suppress eicosanoids synthesized by both COX and LOX, including PGE<sub>2</sub>, PGF<sub>2</sub>, LTB4, and TXB2 in human macrophageenriched bronchoalveolar lavage cells but not in other cells which express steroid receptors [39]. Endogenous glucocorticoids typically do not affect constitutive COX expression but

inhibit inducible COX which is expressed only in the context of inflammation. This suggests that there is a balance between glucocorticoids and COX levels in basal state and any disruptions to this balance could result in inflammation [40]. Similar effects are seen in glial cells and with the corticosteroid drug dexamethasone in human blood monocytes [41, 42]. Abrogation of inflammatory response by glucocorticoids is also observed in zebrafish in response to stress caused by tail vein amputation along with decrease in leukotriene B4 (LTB4) synthesis, an effect that is also seen upon treatment with the synthetic glucocorticoid beclomethasone [43]. Proteasome and eicosanoid profiling of human fibroblasts show that inflammation upregulates a number of eicosanoids synthesized by COX and LOX, such as PGE1, PGE2, TXB2, 11-HETE, and 12-HETE. The upregulated eicosanoid biosynthesis is inhibited by dexamethasone in both fibroblasts and human mesenchymal stem cells [44]. These findings suggest that certain stress mediators activated by components of the CNS suppress eicosanoid production in a pro-inflammatory environment.

#### 2.2 Activation of HPA axis

Excessive/prolonged physical or psychological strain (stress) induced by various stimuli (e.g., physical, mental, or emotional) can manifest itself physically by illness-associated stress responses such as fever, lethargy, anorexia, hyperalgesia, and glucocorticoid secretion [45]. The primary physical stimuli for illness-associated responses is stress induced systemic inflammation, which can be efficiently replicated in various animal models. Many stress stimuli at the end culminate in PGE<sub>2</sub> synthesis. Experimental findings support the involvement of the PGE<sub>2</sub>-synthesizing COX2 enzyme in response to illness as well as the upregulation of this enzyme by illness-associated stimuli [46, 47]. Studies in mice have shown that COX1 is involved in early responses to illness-related stress, particularly in HPA axis activation [46, 48, 49]. Systemic NSAID administration blocks HPA activation by illness-induced stress, substantiating the fact that HPA activation is associated with a COXdependent pathway [50]. Injection of COX1 inhibitor SC-560 into the CNS abrogates initial (ACTH) release in response to stress, suggesting the presence of a synergistic relationship between arachidonic acid metabolism and pituitary function [49]. Additionally, EP1 and EP3 deficient mice have impaired ACTH release, thereby establishing the importance of prostaglandins in HPA-mediated stress response [51]. EP3 is also expressed in catecholaminergic neuron groups and these sites are thought to be involved in illnessinduced HPA activation [52]. These findings collectively suggest that certain CNS responses to illness-induced stress are governed by eicosanoid-dependent mechanisms [45].

#### 2.3 Psychological stress response

Under psychological stress, PGE<sub>2</sub> signaling *via* its receptor EP1 plays an important role in regulating impulsive behavior *in vivo*, with EP1 deficiency resulting in impulsive aggression and deficits in social behavior in response to psychological stress. Some of these aggressive responses can be mimicked by administering EP1 antagonists to wild-type mice [53]. COX2 expression in cortical pyramidal neurons increases under psychological stressors like restraint and forced-swim stress [54, 55]. Increase in the catecholamine norepinephrine caused by psychosocial stress enhances PGE<sub>2</sub> synthesis *via* PTGS2 upregulation, which promotes formation of a pro-inflammatory and pro-metastatic microenvironment in ovarian cancer [19]. These studies indicate the presence of a direct relationship between

psychological stress and prostaglandin synthesis; the underlying mechanisms remain elusive but are being investigated. Similarly, high levels of PGE<sub>2</sub> correlate with poor prognosis in many malignancies including colon, lung, and breast cancer [56–58].

# 3 Stress, inflammation, and the role of eicosanoids

Apart from the well-studied stress-induced tumor angiogenesis program, another area of interest is the role of tumor infiltration of autonomous nervous system projections (tumor innervation) in tumor growth. The role of stress mediators and adrenergic signaling in inflammation has been widely studied, especially in the context of tumor angiogenesis. In addition to enhanced tumor angiogenesis, tumor innervation might further enhance tumorigenesis by enriching the tumor microenvironment with catecholamines released by the newly formed nerve endings. We and others have found that in response to catecholamines, tumor cells release neurotrophins (BDNF/NGF), which leads to increased tumor innervation and growth in restraint stress tumor models [59, 60]. There is evidence to show that behavioral stress contributes to an increase in catecholaminergic nerve fibers within lymphoid organs in primates, an indication of a long-term regulatory influence of stress on immune responses [61]. Norepinephrine, the ANS stress response mediator, possesses potent pro-inflammatory properties. High concentration of norepinephrine in circulation caused by cutaneous injury and skin damage leads to sustained inflammation and impaired wound healing [62, 63]. Post-ganglionic neurons innervate all primary and secondary lymphoid organs where norepinephrine is stored within the nerve terminals [64, 65]. In response to CNS activation by psychological stress, norepinephrine is released from the nerve terminals and interacts with immune cells via adrenergic receptors (AR) present on the immune cells. The level of receptor expression varies with immune cell type, activation state, and cytokines [20], which could explain the heterogeneity in response of different immune cells to stress mediators.

Stress-induced upregulation of catecholamines and glucocorticoids in general impairs antigen presentation, T cell proliferation, and attenuate humoral and cell-mediated immunity [2]. However, the actions of stress mediators on individual components of the immune system are quite varied. Norepinephrine increases dendritic cell migration in stress-induced CD8+ T cell response but antigen-presenting ability and delayed-type hypersensitivity are greatly reduced [66-68]. Adrenergic receptor stimulation of dendritic cells promotes Th2associated inflammation and Th17 differentiation [69, 70]. In vitro exposure to norepinephrine or ADRB2 agonists decreases IL-2 secretion from naïve murine CD4+ Tcells. Under Th1-promoting culture conditions and pathogens, norepinephrine induces naïve T cells to differentiate into Th1 cells which in turn produce high IFN $\gamma$  levels in response to norepinephrine [71, 72]. Suppressive phenotype of Treg and Th17 cells is enhanced by ADRB2 agonists and norepinephrine by upregulating CTLA-4, leading to decrease in IL-2 and inflammatory response [73, 74]. The effect of stress mediators on activity of CD8+ T cells remains unclear; however, psychological stress is known to upregulate CD8+ T cell number in humans [75, 76]. In macrophages, exposure to catecholamines and sympathomimetic molecules via β-adrenergic receptor 2 (ADRB2) abrogates production of pro-inflammatory TNF- $\alpha$  [77–80], whereas TNF- $\alpha$  production is greatly increased by stimulation of  $\alpha$ -adrenergic receptors [81–83].

Various studies have shown that stress promotes recruitment of tumor-associated macrophages which produce pro-inflammatory and pro-angiogenic factors in large quantities including vascular endothelial growth factor and matrix metalloproteinases [84, 85]. Stress can also activate immune suppressor cells like myeloid-derived suppressor cells (MDSCs) which can greatly hinder CD8+ T cell function [86–89]. Besides psychological stress, other forms of stress also adversely affect the immune system. For example, forced swim and surgical stress suppress NK cell activity in syngeneic F344 rat models of leukemia, mammary, and colon carcinomas while causing leukemia-related mortality and mammary carcinoma metastasis [90]. Excessive prostaglandin and catecholamine levels have been shown to contribute to postoperative immune suppression, suggesting the presence of an interaction between CNS activity and eicosanoid metabolism. Perioperative combination treatment with COX-2 inhibitors and  $\beta$ -blockers reduces lung tumor retention in mammary carcinoma models, while  $\beta$ -blockade prevents NK cell attenuation [91]. Studies on 38 earlystage breast cancer patients found that perioperative treatment with the ADRB2 antagonist propranolol and the COX-2 inhibitor etodolac inhibits metastasis-associated molecular pathways as well as recurrence of disease [92].

There is growing evidence to demonstrate that eicosanoids, mainly prostanoids and leukotrienes, are involved in crosstalk between immune components and epithelial cells in cancer. Under conditions of chronic inflammation, normal or transformed epithelial cells and resident immune cells secrete pro-inflammatory eicosanoids and cytokines which recruit additional leukocytes to the site. This leads to increased infiltration of dysregulated immune cells, loss of epithelial integrity, and upregulation of inflammatory mediators [93]. PGE<sub>2</sub> and elevated COX2 levels are known to increase neutrophil infiltration and exacerbate inflammation in inflammatory bowel disease (IBD), thereby increasing the risk for colon carcino-genesis. In IBD, inflammatory Th17 expansion is enhanced by PGE2 via IL-23 derived from dendritic cells [94]. IL-23 derived from peripheral blood mononuclear cells and naïve T cells promotes Th17 expansion in IBD, a process that is strongly mediated by PGE<sub>2</sub> [95, 96]. PGE<sub>2</sub> is also elevated in blood and tumor tissues of colon cancer patients, indicating a role for prostaglandins in inflammation-induced malignancy [97]. Murine colon cancer models with COX2 knockout or treated with COX2 inhibitors exhibit reduced carcinogenesis, suggesting that the COX2-PGE2 pathway could serve as a therapeutic target [98]. In line with this, the positive feedback between COX-2 and PGE2 is essential for expression of CXCR4 in cancer-associated MDSCs and the production of its ligand of CXCL2 in ovarian cancer [99]. Similarly, aspirin-PC inhibits cell proliferation, angiogenesis, and increased apoptosis in ovarian cancer cells and mouse models [100]. PGE<sub>2</sub> also functions as an immunosuppressive factor and contributed to the microenvironment of ovarian cancer by compromising the Toll-like receptor-mediated dendritic cell activation [101]. In animal models of chronic inflammation, PGE2 promotes inflammation via Th1 and Th17cells, thereby aggravating inflammation [102]. While most research on the involvement of eicosanoids in inflammation, immune system, and cancer is focused on COX-2-mediated pathways, LOXs and their metabolites also appear to play a role in inflammation, immune modulation, and associated cancer. The leukotriene LTB4 is also a potent attractant of neutrophils, eosinophils, T cells, and macrophages to sites of inflammation, via Rac-Erk signaling [103–106]. In BALB/c mouse models of asthma,

leukotrienes are known to mediate airway inflammation by increasing eosinophil infiltration [107]. Low affinity LTB4 receptor (BLT2) and LTB4 can also contribute to the chemoresistance; co-treatment with a BLT2 inhibitor substantially reduced resistance to cisplatin in the SKOV-3 cell tumor model [108]. Expression of 5-LOX was much higher in the human ovarian tumor tissues with highly infiltrated tumor-associated macrophages and macrophages treated with 5-LOX metabolites led to increased expression of MMP-7 through the p38 pathway [109].

There is considerable evidence to demonstrate the therapeutic potential of targeting the interaction between nerves with immune cells to improve immunity, particularly in cancer. Use of host cell-targeted immune therapies such as CTLA-4 and PD-1 blockade heavily rely on the presence of functional T cells, establishing the need for therapies which prevent immune suppression in the microenvironment. As described above, there is extensive experimental data to suggest that use of  $\beta$ -blockers could be an effective strategy to improve host immune responses. Similarly, NSAIDs that target COX and LOX pathways could be included in the therapeutic regimen to treat tumor-associated inflammation. Despite the seemingly independent pro-inflammatory actions of biobehavioral pathways and eicosanoids, there could be an underlying synergism between these processes in inducing inflammation, an interaction that has not been fully characterized yet due to biological complexity. Continued research in these areas is expected to uncover some of the mechanisms of interactions between CNS responses and eicosanoids in promoting inflammation.

# 4 Discussion

Multiple pre-clinical and clinical studies have shown that psychosocial factors, depression, and stress in humans affect many biological processes. Many of these studies specifically link SNS activation to malignant progression, metabolism dysregulation, and inflammation. Stress-inducible metabolic factors, such as COX2, modulate stress responses in illness and are also involved in activation of components of the CNS under stress. Stress mediators such as norepinephrine directly influence immune cell number and activity at a site, subsequently affecting inflammation. The studies outlined in this review show that stress can shape the immune environment and metabolism of tumors and normal cells, highlighting the importance of deeper understanding of the interactions between these processes in driving disease (Fig. 2).

While the effects of adrenergic signaling and stress on crucial tumor metabolic pathways have been extensively investigated [110, 111], interactions between stress and eicosanoids are not fully understood. Arachidonic acid pathway and eicosanoids respond to different types of stress stimuli and there could be responses to adrenergic signaling and psychological stress. It is important to uncover the underlying interactions between these processes in order to expand the understanding of disease development and progression.

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# References

- Antoni MH, Lutgendorf SK, Cole SW, Dhabhar FS, Sephton SE, McDonald PG, Stefanek M, & Sood AK (2006). The influence of bio-behavioural factors on tumour biology: pathways and mechanisms. Nature Reviews. Cancer, 6(3), 240–248.
- Glaser R, & Kiecolt-Glaser JK (2005). Stress-induced immune dysfunction: implications for health. Nature Reviews. Immunology, 5(3), 243–251.
- Charmandari E, Tsigos C, & Chrousos G (2005). Endocrinology of the stress response. Annual Review of Physiology, 67, 259–284.
- McEwen BS (2002). Sex, stress and the hippocampus: allostasis, allostatic load and the aging process. Neurobiology of Aging, 23(5), 921–939. [PubMed: 12392796]
- 5. Penninx BW, et al. (1998). Chronically depressed mood and cancer risk in older persons. Journal of the National Cancer Institute, 90(24), 1888–1893. [PubMed: 9862626]
- Duijts SF, Zeegers MP, & Borne BV (2003). The association between stressful life events and breast cancer risk: a meta-analysis. International Journal of Cancer, 107(6), 1023–1029. [PubMed: 14601065]
- 7. Bleiker EM, et al. (2008). Personality factors and breast cancer risk: a 13-year follow-up. Journal of the National Cancer Institute, 100(3), 213–218. [PubMed: 18230799]
- Price MA, Tennant CC, Smith RC, Butow PN, Kennedy SJ, Kossoff MB, & Dunn SM (2001). The role of psychosocial factors in the development of breast carcinoma: part I. The cancer prone personality. Cancer, 91(4), 679–685. [PubMed: 11241234]
- Chida Y, Hamer M, Wardle J, & Steptoe A (2008). Do stress-related psychosocial factors contribute to cancer incidence and survival? Nature Clinical Practice. Oncology, 5(8), 466–475. [PubMed: 18493231]
- Selye H (1998). A syndrome produced by diverse nocuous agents. 1936. The Journal of Neuropsychiatry and Clinical Neurosciences, 10(2), 230–231. [PubMed: 9722327]
- Reiche EM, Nunes SO, & Morimoto HK (2004). Stress, depression, the immune system, and cancer. The Lancet Oncology, 5(10), 617–625. [PubMed: 15465465]
- 12. Cole SW, Nagaraja AS, Lutgendorf SK, Green PA, & Sood AK (2015). Sympathetic nervous system regulation of the tumour microenvironment. Nature Reviews. Cancer, 15(9), 563–572.
- Hassan S, Karpova Y, Baiz D, Yancey D, Pullikuth A, Flores A, Register T, Cline JM, D'Agostino R, Jr, Danial N, Datta SR, & Kulik G (2013). Behavioral stress accelerates prostate cancer development in mice. The Journal of Clinical Investigation, 123(2), 874–886. [PubMed: 23348742]
- Kruk J, & Aboul-Enein HY (2004). Psychological stress and the risk of breast cancer: a casecontrol study. Cancer Detection and Prevention, 28(6), 399–408. [PubMed: 15582263]
- Lutgendorf SK, de Geest K, Bender D, Ahmed A, Goodheart MJ, Dahmoush L, Zimmerman MB, Penedo FJ, Lucci III JA, Ganjei-Azar P, Thaker PH, Mendez L, Lubaroff DM, Slavich GM, Cole SW, & Sood AK (2012). Social influences on clinical outcomes of patients with ovarian cancer. Journal of Clinical Oncology, 30(23), 2885–2890. [PubMed: 22802321]
- Ramirez AJ, Craig TK, Watson JP, Fentiman IS, North WR, & Rubens RD (1989). Stress and relapse of breast cancer. BMJ, 298(6669), 291–293. [PubMed: 2493899]
- 17. Thaker PH, Han LY, Kamat AA, Arevalo JM, Takahashi R, Lu C, Jennings NB, Armaiz-Pena G, Bankson JA, Ravoori M, Merritt WM, Lin YG, Mangala LS, Kim TJ, Coleman RL, Landen CN, Li Y, Felix E, Sanguino AM, Newman RA, Lloyd M, Gershenson DM, Kundra V, Lopez-Berestein G, Lutgendorf SK, Cole SW, & Sood AK (2006). Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. Nature Medicine, 12(8), 939–944.
- Wang HM, Liao ZX, Komaki R, Welsh JW, O'Reilly MS, Chang JY, Zhuang Y, Levy LB, Lu C, & Gomez DR (2013). Improved survival outcomes with the incidental use of beta-blockers among patients with non-small-cell lung cancer treated with definitive radiation therapy. Annals of Oncology, 24(5), 1312–1319. [PubMed: 23300016]

- Nagaraja AS, Dorniak PL, Sadaoui NC, Kang Y, Lin T, Armaiz-Pena G, Wu SY, Rupaimoole R, Allen JK, Gharpure KM, Pradeep S, Zand B, Previs RA, Hansen JM, Ivan C, Rodriguez-Aguayo C, Yang P, Lopez-Berestein G, Lutgendorf SK, Cole SW, & Sood AK (2016). Sustained adrenergic signaling leads to increased metastasis in ovarian cancer via increased PGE2 synthesis. Oncogene, 35(18), 2390–2397. [PubMed: 26257064]
- 20. Sanders VM, & Straub RH (2002). Norepinephrine, the beta-adrenergic receptor, and immunity. Brain, Behavior, and Immunity, 16(4), 290–332.
- Jean D, & Bar-Eli M (2000). Regulation of tumor growth and metastasis of human melanoma by the CREB transcription factor family. Molecular and Cellular Biochemistry, 212(1–2), 19–28. [PubMed: 11108132]
- 22. Sood AK, Bhatty R, Kamat AA, Landen CN, Han L, Thaker PH, Li Y, Gershenson DM, Lutgendorf S, & Cole SW (2006). Stress hormone-mediated invasion of ovarian cancer cells. Clinical Cancer Research, 12(2), 369–375. [PubMed: 16428474]
- 23. Yang EV, Sood AK, Chen M, Li Y, Eubank TD, Marsh CB, Jewell S, Flavahan NA, Morrison C, Yeh PE, Lemeshow S, & Glaser R (2006). Norepinephrine up-regulates the expression of vascular endothelial growth factor, matrix metalloproteinase (MMP)-2, and MMP-9 in nasopharyngeal carcinoma tumor cells. Cancer Research, 66(21), 10357–10364. [PubMed: 17079456]
- Landen CN, Jr., et al. (2007). Neuroendocrine modulation of signal transducer and activator of transcription-3 in ovarian cancer. Cancer Research, 67(21), 10389–10396. [PubMed: 17974982]
- 25. Li W, Yu CP, Xia JT, Zhang L, Weng GX, Zheng HQ, Kong QL, Hu LJ, Zeng MS, Zeng YX, Li M, Li J, & Song LB (2009). Sphingosine kinase 1 is associated with gastric cancer progression and poor survival of patients. Clinical Cancer Research, 15(4), 1393–1399. [PubMed: 19228740]
- 26. Hansen-Petrik MB, McEntee M, Jull B, Shi H, Zemel MB, & Whelan J (2002). Prostaglandin E(2) protects intestinal tumors from nonsteroidal anti-inflammatory drug-induced regression in Apc(min/+) mice. Cancer Research, 62(2), 403–408. [PubMed: 11809688]
- 27. Yan M, Myung SJ, Fink SP, Lawrence E, Lutterbaugh J, Yang P, Zhou X, Liu D, Rerko RM, Willis J, Dawson D, Tai HH, Barnholtz-Sloan JS, Newman RA, Bertagnolli MM, & Markowitz SD (2009). 15-Hydroxyprostaglandin dehydrogenase inactivation as a mechanism of resistance to celecoxib chemoprevention of colon tumors. Proceedings of the National Academy of Sciences of the United States of America, 106(23), 9409–9413. [PubMed: 19470469]
- Wang D, & Dubois RN (2010). Eicosanoids and cancer. Nature Reviews. Cancer, 10(3), 181–193. [PubMed: 20168319]
- 29. Colby JK, et al. (2008). Progressive metaplastic and dysplastic changes in mouse pancreas induced by cyclooxygenase-2 overexpression. Neoplasia, 10(8), 782–796. [PubMed: 18670639]
- Lewis RA, Austen KF, & Soberman RJ (1990). Leukotrienes and other products of the 5lipoxygenase pathway. Biochemistry and relation to pathobiology in human diseases. The New England Journal of Medicine, 323(10), 645–655. [PubMed: 2166915]
- Agarwal S, Reddy GV, & Reddanna P (2009). Eicosanoids in inflammation and cancer: the role of COX-2. Expert Review of Clinical Immunology, 5(2), 145–165. [PubMed: 20477063]
- Harizi H, Corcuff JB, & Gualde N (2008). Arachidonic-acid-derived eicosanoids: roles in biology and immunopathology. Trends in Molecular Medicine, 14(10), 461–469. [PubMed: 18774339]
- Ruder EH, Laiyemo AO, Graubard BI, Hollenbeck AR, Schatzkin A, & Cross AJ (2011). Nonsteroidal anti-inflammatory drugs and colorectal cancer risk in a large, prospective cohort. The American Journal of Gastroenterology, 106(7), 1340–1350. [PubMed: 21407185]
- 34. Kuo CN, Pan JJ, Huang YW, Tsai HJ, & Chang WC (2018). Association between nonsteroidal anti-inflammatory drugs and colorectal cancer: a population-based case-control study. Cancer Epidemiology, Biomarkers & Prevention.
- 35. Harris RE (2009). Cyclooxygenase-2 (cox-2) blockade in the chemoprevention of cancers of the colon, breast, prostate, and lung. Inflammopharmacology, 17(2), 55–67. [PubMed: 19340409]
- Gurpinar E, Grizzle WE, & Piazza GA (2014). NSAIDs inhibit tumorigenesis, but how? Clinical Cancer Research, 20(5), 1104–1113. [PubMed: 24311630]
- 37. Narumiya S (2007). Physiology and pathophysiology of prostanoid receptors. Proceedings of the Japan Academy. Series B, Physical and Biological Sciences, 83(9–10), 296–319.

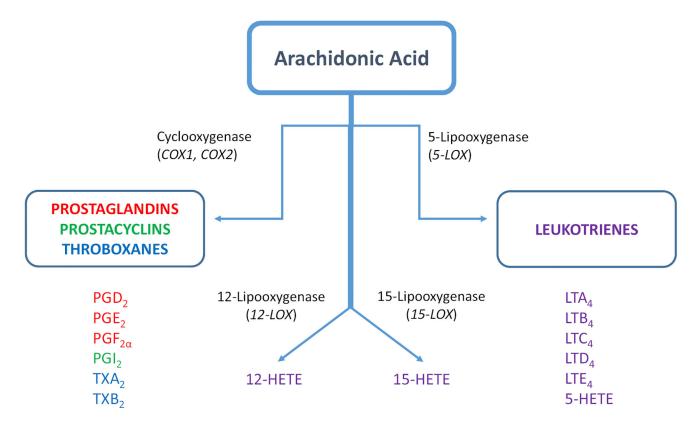
- Brown JR, & DuBois RN (2005). COX-2: a molecular target for colorectal cancer prevention. Journal of Clinical Oncology, 23(12), 2840–2855. [PubMed: 15837998]
- 39. Sebaldt RJ, Sheller JR, Oates JA, Roberts LJ, & FitzGerald GA (1990). Inhibition of eicosanoid biosynthesis by glucocorticoids in humans. Proceedings of the National Academy of Sciences of the United States of America, 87(18), 6974–6978. [PubMed: 2169616]
- 40. Masferrer JL, Seibert K, Zweifel B, & Needleman P (1992). Endogenous glucocorticoids regulate an inducible cyclooxygenase enzyme. Proceedings of the National Academy of Sciences of the United States of America, 89(9), 3917–3921. [PubMed: 1570314]
- Brenner T, Boneh A, Shohami E, Abramsky O, & Weidenfeld J (1992). Glucocorticoid regulation of eicosanoid production by glial cells under basal and stimulated conditions. Journal of Neuroimmunology, 40(2–3), 273–279. [PubMed: 1430157]
- Fu JY, Masferrer JL, Seibert K, Raz A, & Needleman P (1990). The induction and suppression of prostaglandin H2 synthase (cyclooxygenase) in human monocytes. The Journal of Biological Chemistry, 265(28), 16737–16740. [PubMed: 2120205]
- Chatzopoulou A, Heijmans JPM, Burgerhout E, Oskam N, Spaink HP, Meijer AH, & Schaaf MJM (2016). Glucocorticoid-induced attenuation of the inflammatory response in zebrafish. Endocrinology, 157(7), 2772–2784. [PubMed: 27219276]
- 44. Tahir A, Bileck A, Muqaku B, Niederstaetter L, Kreutz D, Mayer RL, Wolrab D, Meier SM, Slany A, & Gerner C (2017). Combined proteome and eicosanoid profiling approach for revealing implications of human fibroblasts in chronic inflammation. Analytical Chemistry, 89(3), 1945–1954. [PubMed: 28208246]
- 45. Furuyashiki T, & Narumiya S (2011). Stress responses: the contribution of prostaglandin E(2) and its receptors. Nature Reviews. Endocrinology, 7(3), 163–175.
- 46. Elander L, Engstrom L, Ruud J, Mackerlova L, Jakobsson PJ, Engblom D, Nilsberth C, & Blomqvist A (2009). Inducible prostaglandin E2 synthesis interacts in a temporally supplementary sequence with constitutive prostaglandin-synthesizing enzymes in creating the hypothalamicpituitary-adrenal axis response to immune challenge. The Journal of Neuroscience, 29(5), 1404– 1413. [PubMed: 19193887]
- Elmquist JK, Breder CD, Sherin JE, Scammell TE, Hickey WF, Dewitt D, & Saper CB (1997). Intravenous lipopolysaccharide induces cyclooxygenase 2-like immunoreactivity in rat brain perivascular microglia and meningeal macrophages. The Journal of Comparative Neurology, 381(2), 119–129. [PubMed: 9130663]
- Elander L, Ruud J, Korotkova M, Jakobsson PJ, & Blomqvist A (2010). Cyclooxygenase-1 mediates the immediate corticosterone response to peripheral immune challenge induced by lipopolysaccharide. Neuroscience Letters, 470(1), 10–12. [PubMed: 20034541]
- Garcia-Bueno B, Serrats J, & Sawchenko PE (2009). Cerebrovascular cyclooxygenase-1 expression, regulation, and role in hypothalamic-pituitary-adrenal axis activation by inflammatory stimuli. The Journal of Neuroscience, 29(41), 12970–12981. [PubMed: 19828811]
- Turnbull AV, & Rivier CL (1999). Regulation of the hypothalamic-pituitary-adrenal axis by cytokines: actions and mechanisms of action. Physiological Reviews, 79(1), 1–71. [PubMed: 9922367]
- 51. Matsuoka Y, Furuyashiki T, Bito H, Ushikubi F, Tanaka Y, Kobayashi T, Muro S, Satoh N, Kayahara T, Higashi M, Mizoguchi A, Shichi H, Fukuda Y, Nakao K, & Narumiya S (2003). Impaired adrenocorticotropic hormone response to bacterial endotoxin in mice deficient in prostaglandin E receptor EP1 and EP3 subtypes. Proceedings of the National Academy of Sciences of the United States of America, 100(7), 4132–4137. [PubMed: 12642666]
- Ericsson A, Arias C, & Sawchenko PE (1997). Evidence for an intramedullary prostaglandindependent mechanism in the activation of stress-related neuroendocrine circuitry by intravenous interleukin-1. The Journal of Neuroscience, 17(18), 7166–7179. [PubMed: 9278551]
- Matsuoka Y, Furuyashiki T, Yamada K, Nagai T, Bito H, Tanaka Y, Kitaoka S, Ushikubi F, Nabeshima T, & Narumiya S (2005). Prostaglandin E receptor EP1 controls impulsive behavior under stress. Proceedings of the National Academy of Sciences of the United States of America, 102(44), 16066–16071. [PubMed: 16247016]

- Garcia-Bueno B, et al. (2008). Stress mediators regulate brain prostaglandin synthesis and peroxisome proliferator-activated receptor-gamma activation after stress in rats. Endocrinology, 149(4), 1969–1978. [PubMed: 18079203]
- 55. Yamagata K, Andreasson KI, Kaufmann WE, Barnes CA, & Worley PF (1993). Expression of a mitogen-inducible cyclooxygenase in brain neurons: regulation by synaptic activity and glucocorticoids. Neuron, 11(2), 371–386. [PubMed: 8352945]
- 56. McLemore TL, et al. (1988). Profiles of prostaglandin biosyn-thesis in normal lung and tumor tissue from lung cancer patients. Cancer Research, 48(11), 3140–3147. [PubMed: 3130187]
- Rigas B, Goldman IS, & Levine L (1993). Altered eicosanoid levels in human colon cancer. The Journal of Laboratory and Clinical Medicine, 122(5), 518–523. [PubMed: 8228569]
- Wang D, & Dubois RN (2004). Cyclooxygenase-2: a potential target in breast cancer. Seminars in Oncology, 31(1 Suppl 3), 64–73.
- 59. Allen JK, Armaiz-Pena GN, Nagaraja AS, Sadaoui NC, Ortiz T, Dood R, Ozcan M, Herder DM, Haemerrle M, Gharpure KM, Rupaimoole R, Previs R, Wu SY, Pradeep S, Xu X, Dong Han H, Zand B, Dalton HJ, Taylor M, Hu W, Bottsford-Miller J, Moreno-Smith M, Kang Y, Mangala LS, Rodriguez-Aguayo C, Sehgal V, Spaeth EL, Ram PT, Wong STC, Marini FC, Lopez-Berestein G, Cole SW, Lutgendorf SK, diBiasi M, & Sood AK (2018). Sustained adrenergic signaling promotes intratumoral innervation through BDNF induction. In Cancer Res (p. canres.1701.2016).
- 60. Renz BW, et al. (2018). β2 Adrenergic-neurotrophin feedforward loop promotes pancreatic cancer. Cancer Cell, 33(1), 75–90 e7. [PubMed: 29249692]
- 61. Sloan EK, Capitanio JP, & Cole SW (2008). Stress-induced remodeling of lymphoid innervation. Brain, Behavior, and Immunity, 22(1), 15–21.
- Gosain A, Jones SB, Shankar R, Gamelli RL, & DiPietro LA (2006). Norepinephrine modulates the inflammatory and proliferative phases of wound healing. The Journal of Trauma, 60(4), 736– 744. [PubMed: 16612292]
- 63. Sivamani RK, Pullar CE, Manabat-Hidalgo CG, Rocke DM, Carlsen RC, Greenhalgh DG, & Isseroff RR (2009). Stress-mediated increases in systemic and local epinephrine impair skin wound healing: potential new indication for beta blockers. PLoS Medicine, 6(1), e12. [PubMed: 19143471]
- 64. Felten DL, et al. (1985). Noradrenergic and peptidergic inner-vation of lymphoid tissue. Journal of Immunology, 135(2 Suppl), 755s–765s.
- 65. Felten SY, & Olschowka J (1987). Noradrenergic sympathetic innervation of the spleen: II. Tyrosine hydroxylase (TH)-positive nerve terminals form synapticlike contacts on lymphocytes in the splenic white pulp. Journal of Neuroscience Research, 18(1), 37–48. [PubMed: 2890771]
- Maestroni GJ, & Mazzola P (2003). Langerhans cells beta 2-adrenoceptors: role in migration, cytokine production, Th priming and contact hypersensitivity. Journal of Neuroimmunology, 144(1–2), 91–99. [PubMed: 14597102]
- 67. Saint-Mezard P, Chavagnac C, Bosset S, Ionescu M, Peyron E, Kaiserlian D, Nicolas JF, & Berard F (2003). Psychological stress exerts an adjuvant effect on skin dendritic cell functions in vivo. Journal of Immunology, 171(8), 4073–4080.
- Seiffert K, Hosoi J, Torii H, Ozawa H, Ding W, Campton K, Wagner JA, & Granstein RD (2002). Catecholamines inhibit the antigen-presenting capability of epidermal Langerhans cells. Journal of Immunology, 168(12), 6128–6135.
- Manni M, Granstein RD, & Maestroni G (2011). β2-Adrenergic agonists bias TLR-2 and NOD2 activated dendritic cells towards inducing an IL-17 immune response. Cytokine, 55(3), 380–386. [PubMed: 21683614]
- Yanagawa Y, Matsumoto M, & Togashi H (2011). Adrenoceptor-mediated enhancement of interleukin-33 production by dendritic cells. Brain, Behavior, and Immunity, 25(7), 1427–1433.
- Alaniz RC, Thomas SA, Perez-Melgosa M, Mueller K, Farr AG, Palmiter RD, & Wilson CB (1999). Dopamine beta-hydroxylase deficiency impairs cellular immunity. Proceedings of the National Academy of Sciences of the United States of America, 96(5), 2274–2278. [PubMed: 10051631]
- 72. Swanson MA, Lee WT, & Sanders VM (2001). IFN-gamma production by Th1 cells generated from naive CD4+ T cells exposed to norepinephrine. Journal of Immunology, 166(1), 232–240.

- 73. Guereschi MG, Araujo LP, Maricato JT, Takenaka MC, Nascimento VM, Vivanco BC, Reis VO, Keller AC, Brum PC, & Basso AS (2013). Beta2-adrenergic receptor signaling in CD4+ Foxp3+ regulatory T cells enhances their suppressive function in a PKA-dependent manner. European Journal of Immunology, 43(4), 1001–1012. [PubMed: 23436577]
- 74. Vida G, et al. (2011). β2-Adrenoreceptors of regulatory lymphocytes are essential for vagal neuromodulation of the innate immune system. The FASEB Journal, 25(12), 4476–4485. [PubMed: 21840939]
- 75. Benschop RJ, Jacobs R, Sommer B, Schürmeyer TH, Raab JR, Schmidt RE, & Schedlowski M (1996). Modulation of the immunologic response to acute stress in humans by beta-blockade or benzodiazepines. The FASEB Journal, 10(4), 517–524. [PubMed: 8647351]
- Mathews PM, et al. (1983). Enhancement of natural cytotoxicity by beta-endorphin. Journal of Immunology, 130(4), 1658–1662.
- 77. Deng J, Muthu K, Gamelli R, Shankar R, & Jones SB (2004). Adrenergic modulation of splenic macrophage cytokine release in polymicrobial sepsis. American Journal of Physiology. Cell Physiology, 287(3), C730–C736. [PubMed: 15151906]
- Elenkov IJ, et al. (1996). Modulatory effects of glucocorticoids and catecholamines on human interleukin-12 and interleukin-10 production: clinical implications. Proceedings of the Association of American Physicians, 108(5), 374–381. [PubMed: 8902882]
- Hetier E, Ayala J, Bousseau A, & Prochiantz A (1991). Modulation of interleukin-1 and tumor necrosis factor expression by beta-adrenergic agonists in mouse ameboid microglial cells. Experimental Brain Research, 86(2), 407–413. [PubMed: 1684549]
- Panina-Bordignon P, Mazzeo D, Lucia PD, D'Ambrosio D, Lang R, Fabbri L, Self C, & Sinigaglia F (1997). Beta2-agonists prevent Th1 development by selective inhibition of interleukin 12. The Journal of Clinical Investigation, 100(6), 1513–1519. [PubMed: 9294119]
- Heijnen CJ, van der Voort CR, Wulffraat N, van der Net J, Kuis W, & Kavelaars A (1996). Functional alpha 1-adrenergic receptors on leukocytes of patients with polyarticular juvenile rheumatoid arthritis. Journal of Neuroimmunology, 71(1–2), 223–226. [PubMed: 8982123]
- 82. Spengler RN, et al. (1990). Stimulation of alpha-adrenergic receptor augments the production of macrophage-derived tumor necrosis factor. Journal of Immunology, 145(5), 1430–1434.
- Szelenyi J, Kiss JP, & Vizi ES (2000). Differential involvement of sympathetic nervous system and immune system in the modulation of TNF-alpha production by alpha2- and beta-adrenoceptors in mice. Journal of Neuroimmunology, 103(1), 34–40. [PubMed: 10674987]
- Madden KS, Szpunar MJ, & Brown EB (2011). Beta-Adrenergic receptors (beta-AR) regulate VEGF and IL-6 production by divergent pathways in high beta-AR-expressing breast cancer cell lines. Breast Cancer Research and Treatment, 130(3), 747–758. [PubMed: 21234673]
- 85. Sloan EK, Priceman SJ, Cox BF, Yu S, Pimentel MA, Tangkanangnukul V, Arevalo JMG, Morizono K, Karanikolas BDW, Wu L, Sood AK, & Cole SW (2010). The sympathetic nervous system induces a metastatic switch in primary breast cancer. Cancer Research, 70(18), 7042–7052. [PubMed: 20823155]
- 86. Waight JD, Netherby C, Hensen ML, Miller A, Hu Q, Liu S, Bogner PN, Farren MR, Lee KP, Liu K, & Abrams SI (2013). Myeloid-derived suppressor cell development is regulated by a STAT/ IRF-8 axis. The Journal of Clinical Investigation, 123(10), 4464–4478. [PubMed: 24091328]
- Gabrilovich DI, & Nagaraj S (2009). Myeloid-derived suppressor cells as regulators of the immune system. Nature Reviews. Immunology, 9(3), 162–174.
- Ostrand-Rosenberg S, Sinha P, Beury DW, & Clements VK (2012). Cross-talk between myeloidderived suppressor cells (MDSC), macrophages, and dendritic cells enhances tumor-induced immune suppression. Seminars in Cancer Biology, 22(4), 275–281. [PubMed: 22313874]
- Jin J, Wang X, Wang Q, Guo X, Cao J, Zhang X, Zhu T, Zhang D, Wang W, Wang J, Shen B, Gao X, Shi Y, & Zhang J (2013). Chronic psychological stress induces the accumulation of myeloid-derived suppressor cells in mice. PLoS One, 8(9), e74497. [PubMed: 24058577]
- Ben-Eliyahu S, et al. (1999). Evidence that stress and surgical interventions promote tumor development by suppressing natural killer cell activity. International Journal of Cancer, 80(6), 880–888. [PubMed: 10074922]

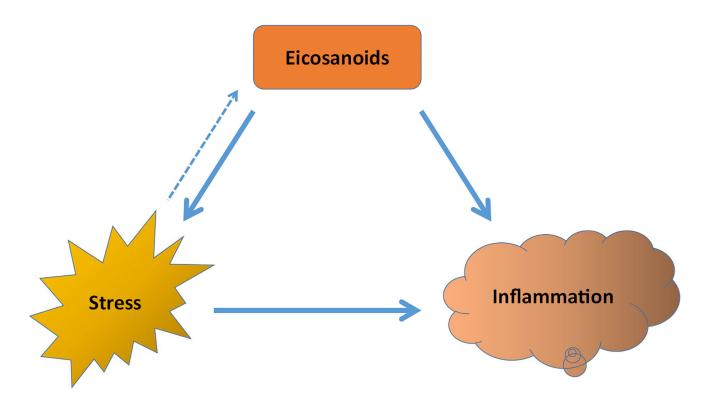
- Benish M, Bartal I, Goldfarb Y, Levi B, Avraham R, Raz A, & Ben-Eliyahu S (2008). Perioperative use of beta-blockers and COX-2 inhibitors may improve immune competence and reduce the risk of tumor metastasis. Annals of Surgical Oncology, 15(7), 2042–2052. [PubMed: 18398660]
- 92. Shaashua L, Shabat-Simon M, Haldar R, Matzner P, Zmora O, Shabtai M, Sharon E, Allweis T, Barshack I, Hayman L, Arevalo J, Ma J, Horowitz M, Cole S, & Ben-Eliyahu S (2017). Perioperative COX-2 and beta-adrenergic blockade improves metastatic biomarkers in breast cancer patients in a phase-II randomized trial. Clinical Cancer Research, 23(16), 4651–4661. [PubMed: 28490464]
- Noonan DM, de Lerma Barbaro A, Vannini N, Mortara L, & Albini A (2008). Inflammation, inflammatory cells and angio-genesis: decisions and indecisions. Cancer Metastasis Reviews, 27(1), 31–40. [PubMed: 18087678]
- 94. Sheibanie AF, Yen JH, Khayrullina T, Emig F, Zhang M, Tuma R, & Ganea D (2007). The proinflammatory effect of prostaglandin E2 in experimental inflammatory bowel disease is mediated through the IL-23->IL-17 axis. Journal of Immunology, 178(12), 8138–8147.
- 95. Boniface K, Bak-Jensen KS, Li Y, Blumenschein WM, McGeachy MJ, McClanahan TK, McKenzie BS, Kastelein RA, Cua DJ, & de Waal Malefyt R (2009). Prostaglandin E2 regulates Th17 cell differentiation and function through cyclic AMP and EP2/EP4 receptor signaling. The Journal of Experimental Medicine, 206(3), 535–548. [PubMed: 19273625]
- 96. Chizzolini C, Chicheportiche R, Alvarez M, de Rham C, Roux-Lombard P, Ferrari-Lacraz S, & Dayer JM (2008). Prostaglandin E2 synergistically with interleukin-23 favors human Th17 expansion. Blood, 112(9), 3696–3703. [PubMed: 18698005]
- Prescott SM, & Fitzpatrick FA (2000). Cyclooxygenase-2 and carcinogenesis. Biochimica et Biophysica Acta, 1470(2), M69–M78. [PubMed: 10722929]
- 98. Oshima M, Dinchuk JE, Kargman SL, Oshima H, Hancock B, Kwong E, Trzaskos JM, Evans JF, & Taketo MM (1996). Suppression of intestinal polyposis in Apc delta716 knockout mice by inhibition of cyclooxygenase 2 (COX-2). Cell, 87(5), 803–809. [PubMed: 8945508]
- Obermajer N, Wong JL, Edwards RP, Odunsi K, Moysich K, & Kalinski P (2012). PGE(2)-driven induction and maintenance of cancer-associated myeloid-derived suppressor cells. Immunological Investigations, 41(6–7), 635–657. [PubMed: 23017139]
- 100. Huang Y, Lichtenberger LM, Taylor M, Bottsford-Miller JN, Haemmerle M, Wagner MJ, Lyons Y, Pradeep S, Hu W, Previs RA, Hansen JM, Fang D, Dorniak PL, Filant J, Dial EJ, Shen F, Hatakeyama H, & Sood AK (2016). Antitumor and antiangiogenic effects of aspirin-PC in ovarian cancer. Molecular Cancer Therapeutics, 15(12), 2894–2904. [PubMed: 27638860]
- 101. Brencicova E, Jagger AL, Evans HG, Georgouli M, Laios A, Attard Montalto S, Mehra G, Spencer J, Ahmed AA, Raju-Kankipati S, Taams LS, & Diebold SS (2017). Interleukin-10 and prostaglandin E2 have complementary but distinct suppressive effects on Toll-like receptormediated dendritic cell activation in ovarian carcinoma. PLoS One, 12(4), e0175712. [PubMed: 28410380]
- 102. Yao C, Sakata D, Esaki Y, Li Y, Matsuoka T, Kuroiwa K, Sugimoto Y, & Narumiya S (2009). Prostaglandin E2-EP4 signaling promotes immune inflammation through Th1 cell differentiation and Th17 cell expansion. Nature Medicine, 15(6), 633–640.
- 103. Bomalaski JS, Dundee D, Brophy L, & Clark MA (1990). Leukotriene B4 modulates phospholipid methylation and chemo-taxis in human polymorphonuclear leukocytes. Journal of Leukocyte Biology, 47(1), 1–12. [PubMed: 2152937]
- 104. Haribabu B, Verghese MW, Steeber DA, Sellars DD, Bock CB, & Snyderman R (2000). Targeted disruption of the leukotriene B(4) receptor in mice reveals its role in inflammation and plateletactivating factor-induced anaphylaxis. The Journal of Experimental Medicine, 192(3), 433–438. [PubMed: 10934231]
- 105. Islam SA, Thomas SY, Hess C, Medoff BD, Means TK, Brander C, Lilly CM, Tager AM, & Luster AD (2006). The leukotriene B4 lipid chemoattractant receptor BLT1 defines antigenprimed T cells in humans. Blood, 107(2), 444–453. [PubMed: 16179368]
- 106. Woo CH, You HJ, Cho SH, Eom YW, Chun JS, Yoo YJ, & Kim JH (2002). Leukotriene B(4) stimulates Rac-ERK cascade to generate reactive oxygen species that mediates chemo-taxis. The Journal of Biological Chemistry, 277(10), 8572–8578. [PubMed: 11756405]

- 107. Henderson WR, Jr., et al. (1996). The importance of leukotrienes in airway inflammation in a mouse model of asthma. The Journal of Experimental Medicine, 184(4), 1483–1494. [PubMed: 8879219]
- 108. Park J, Park SY, & Kim JH (2016). Leukotriene B4 receptor-2 contributes to chemoresistance of SK-OV-3 ovarian cancer cells through activation of signal transducer and activator of transcription-3-linked cascade. Biochimica et Biophysica Acta, 1863(2), 236–243. [PubMed: 26597704]
- 109. Wen Z, Liu H, Li M, Li B, Gao W, Shao Q, Fan B, Zhao F, Wang Q, Xie Q, Yang Y, Yu J, & Qu X (2015). Increased metabolites of 5-lipoxygenase from hypoxic ovarian cancer cells promote tumor-associated macrophage infiltration. Oncogene, 34(10), 1241–1252. [PubMed: 24662827]
- 110. Fruhbeck G, et al. (2001). The adipocyte: a model for integration of endocrine and metabolic signaling in energy metabolism regulation. American Journal of Physiology. Endocrinology and Metabolism, 280(6), E827–E847. [PubMed: 11350765]
- 111. Fearon KC, Glass DJ, & Guttridge DC (2012). Cancer cachexia: mediators, signaling, and metabolic pathways. Cell Metabolism, 16(2), 153–166. [PubMed: 22795476]



#### Fig. 1.

Biosynthesis of eicosanoids. Arachidonic acid metabolism is initiated by phospholipase A which is activated by various chemical and mechanical stimuli, leading to arachidonic acid generation. Free arachidonic acid is metabolized by two main enzymes, cyclooxygenases (COX), and lipoxygenases (LOX). COX enzymes (COX-1 and COX-2) catalyze the formation of PGH2, which is further converted to various prostanoids (prostaglandins, prostacyclins, and thromboxanes). LOX enzymes (5-LOX, 12-LOX, and 15-LOX) mediate the synthesis of leukotrienes, hydroxyeicosatetraenoic acid (HETE) and lipoxins



# Fig. 2.

Stress and eicosanoids directly influence inflammation. However, the relationship between stress and eicosanoids is not well established. Eicosanoids activate certain components of CNS-mediated stress response pathway, while stress mediators are known to influence synthesis of certain eicosanoids. It is possible that stress mediators and eicosanoid-synthesizing enzymes influence each other, should their basal levels be disrupted. However, additional research is needed to elucidate the exact pathways and mediators involved in this interaction