

Cytoplasmic lattices are not linked to mouse 2-cell embryos developmental arrest

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Abstract

Cytoplasmic lattices are important regulators of oocyte maturation. They store components of the protein synthesis machinery including ribosomes and, among others, they are involved in the regulation of microtubule dynamics in both mouse and human. Cytoplasmic lattices undergo dramatic reorganizations at crucial stages of oocyte maturation, where they are abundantly present in the cytoplasm of developmentally competent oocytes named SN (Surrounded Nucleolus) while they are rare in the cytoplasm of 2-cell stage-arresting NSN (Not Surrounded Nucleolus) oocytes, suggestive of a requirement of cytoplasmic lattices for development past the 2-cell stage. Here, to elucidate this requirement, 2-cell mouse embryos derived from SN and NSN oocytes were analyzed by transmission electron microscopy. Contrary to what had been proposed hitherto, cytoplasmic lattices are present in 2-cell embryos derived not only from SN, but also from NSN oocytes, irrespective of the embryo production system (intracytoplasmic sperm injection, parthenogenesis). Hence our conclusion that cytoplasmic lattices do not count among the factor(s) responsible for the embryo arrest at this crucial stage of development.

Introduction

The acquisition of oocyte developmental competence is one of the most essential and fascinating biological processes, con-

sisting of several steps leading to meiosis progression and to the ability to sustain fertilization and the prospect embryo development.

Several factors and mechanisms are involved in each phase of this process like the control of oocyte development by the interaction with somatic cells, the arrangement of the chromosomes within the nucleus, the organelles rearrangement, the role played by epigenetic mechanisms, the chromatin conformation of the NSN oocytes and the down regulation of maternal genes/proteins are the most reasonable explanations of the embryo arrest at the 2/4-cell stage.¹⁻⁴

Among the organelles of the ooplasm, cytoplasmic lattices (CPLs) are ascribed with key roles in oocyte maturation, although their degree of involvement is not entirely understood. CPLs are causatively related to microtubule and organelle dynamics, ribosomal storage and maternal-zygotic transition by controlling the translational processes,^{3,5} they are also (apparently) required for the embryo development beyond the 2-cell stage;⁶ in other words, CPLs are functionally relevant to the housekeeper (basic) as well as luxury biological processes. Recent evidence showed that CPLs start to appear in primary oocytes and function as a storage site for aggregated maternal ribosomes.⁶ CPLs have also been found abundant in the cytoplasm of mouse and human germinal vesicle (GV) oocytes named Surrounded Nucleolus (SN), which account for 70-80% of GV oocytes that are able to develop beyond the 2/4-cell stage after maturation and fertilization; conversely, the remaining 20-30% of oocytes named Not Surrounded Nucleolus (NSN) are almost completely devoid of CPLs and also lack the ability to develop beyond the 2/4-cell stage.^{7,8} Based on these records, we hypothesized that presence/absence of CPLs in SN/NSN GV oocytes is linked to the occurrence of 2-cell stage arrest, as also suggested by recent studies on maternal proteins essential for embryo development beyond the 2/4-cell stage.⁴ Indeed, MATER, a main constituent of the subcortical maternal complex (SCMC) plays critical roles in CPLs formation and function and in embryo development beyond the 2-cell stage.⁴ It has been shown that the majority of GV oocytes isolated from animals with a transient mutation for MATER have an NSN phenotype with no CPLs in their cytoplasm⁷ and, as expected, are unable to support the embryonic development beyond the 2-cell stage. Interestingly, the depletion of other maternal proteins and/or members of the SCMC, like PADI6, probably leads to the formation of NSN GV oocytes (and/or prevents the transition from NSN to SN) with no CPLs

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and unable to complete the prospect embryo development.^{6,9,10} Based on these records, we sought to elucidate the requirement of CPLs for preimplantation development, by quantitatively analyzing the CPLs content in 2-cell embryos of SN and NSN oocytes; specifically, we tested the expectation that only SN-derived embryos have CPLs in their cytoplasm. Thanks to the use of two embryo production systems, namely parthenogenesis and fertilization (intracytoplasmic sperm injection, ICSI), we were able to exclude a differential response of SN and NSN oocytes to sperm, and to retain the pristine nature of the oocytes in respect to the cytoplasmic lattices, which are of maternal origin. The evidence here provided supports the view that cytoplasmic lattices are not linked to mouse 2-cell embryos developmental arrest.

Materials and Methods

Ethics statement

All mice (20 females and 4 males) used were housed in individually ventilated cages in the animal facility of the MPI Münster, with a controlled temperature of 22°C, with a 14/10 h light/dark photoperiod and with free access to water and food (Harlan Teklad 2020SX). Experimental procedures followed the ethical guidelines of the European Laboratory Animal Science Associations (FELASA) and the regulatory guidelines of European Directive 63/2010. On the local regulatory level, mice were used for experiments according to the ethical permit issued by the Landesamt für Natur, Umwelt und Verbraucherschutz (LANUV) of the state of North Rhine-Westphalia, Germany (permit number 84-02.05.20.13.022).

Collection of oocytes and spermatozoa

Six to eight week-old B6C3F1 female mice were used as donors of GV-stage oocytes for *in vitro* maturation (IVM) and metaphase II (MII)-stage oocytes for developmental activation (intracytoplasmic sperm injection, ICSI; parthenogenesis). Twelve week-old CD1 male mice were used as sperm donors for ICSI. GV-oocytes were isolated by puncturing the surface of the ovary 48 h after PMSG stimulation (10 IU), MII oocytes were recovered from the oviductal ampullae 15 h after hCG and PMSG stimulation injected 48 h apart (10 IU each), and freed of cumulus cells using hyaluronidase (50 IU/mL). Caudae epididymis were punctured with a 27G needle, and overlaid with pre-equilibrated Whittingham medium (3% BSA) on the bottom of a 1.5 mL Eppendorf tube so as to allow for sperm to swim-up at 37°C under 5% CO₂ in air.

Sorting and IVM of GV-stage oocytes

GV oocytes were scored for the presence/absence of a Hoechst-positive ring of heterochromatin surrounding the nucleolus, leading to allocation in two classes (Surrounded Nucleolus, SN; Not Surrounded Nucleolus, NSN). Scoring was performed while holding the oocytes in HEPES-buffered CZB medium in the presence of 0.1 µg/mL Hoechst 33342 on the stage of an inverted microscope (Nikon TE 2000) fitted with an EXFO light source. After sorting, oocytes were allowed to progress to MII by culturing them in α -MEM medium (Sigma cat.no. M4526) supplemented with 0.2% BSA (w/v) and 50 mg/mL gentamicin. Incubation took place at 37°C under 5% CO₂ and 5% O₂ (balance N₂). This low-oxygen atmosphere was

achieved in a homemade plastic chamber positioned on the shelf of the incubator and connected to a 5/5/90 CO₂/O₂/N₂ tank.

Parthenogenesis

Parthenogenetic activation of MII oocytes was performed for 6 hours in Ca-free α -MEM medium supplemented with 10 mM SrCl₂. Cytochalasin B (5 µg/mL; Calbiochem cat. No. 250233) was also added to the medium so as to prevent the extrusion of a pseudo polar body and thereby obtain diploid parthenotes.

Intracytoplasmic sperm injection

The upper layer (100 µL) of the sperm swim-up (30') suspension was collected and incubated as a drop under pre-equilibrated mineral oil for 2-3 h to allow for capacitation and acrosome reaction. Single sperm heads were injected into MII oocytes using a piezo drill (PrimeTech).¹¹

Embryo culture

Parthenogenetic and ICSI-fertilized embryos were cultured 4 days in 500 µL α -

MEM (supplemented with 0.2 % BSA (w/v) and 50 mg/mL gentamicin) in 4-well plates (Nunc) without oil overlay, under 5% CO₂ in air at 37°C.

Transmission electron microscopy

At the 2-cell stage, a total of 82 embryos (58 SN, 24 NSN) and 48 embryos (30 SN, 18 NSN) were collected after parthenogenesis and ICSI, respectively. They were fixed in 2% paraformaldehyde, 2% glutaraldehyde in 0.1 M cacodylate buffer, pH 7.2 for three hours at room temperature. Next the samples were adsorbed to poly-L-lysine coated cover slips.¹² Subsequently samples were post-fixed in 1% osmium tetroxide, 1.5% potassium ferrocyanide in 0.1 M cacodylate buffer, pH 7.2. Then they were stepwise dehydrated in ethanol, including *en bloc* 0.5% uranyl acetate staining during 70% ethanol. The coverslips were flat embedded in epon and polymerized. From the hardened sample the cover slip was removed and the epon bloc was ultrathin-sectioned at 70 nm. Sections were collected on copper grids and stained

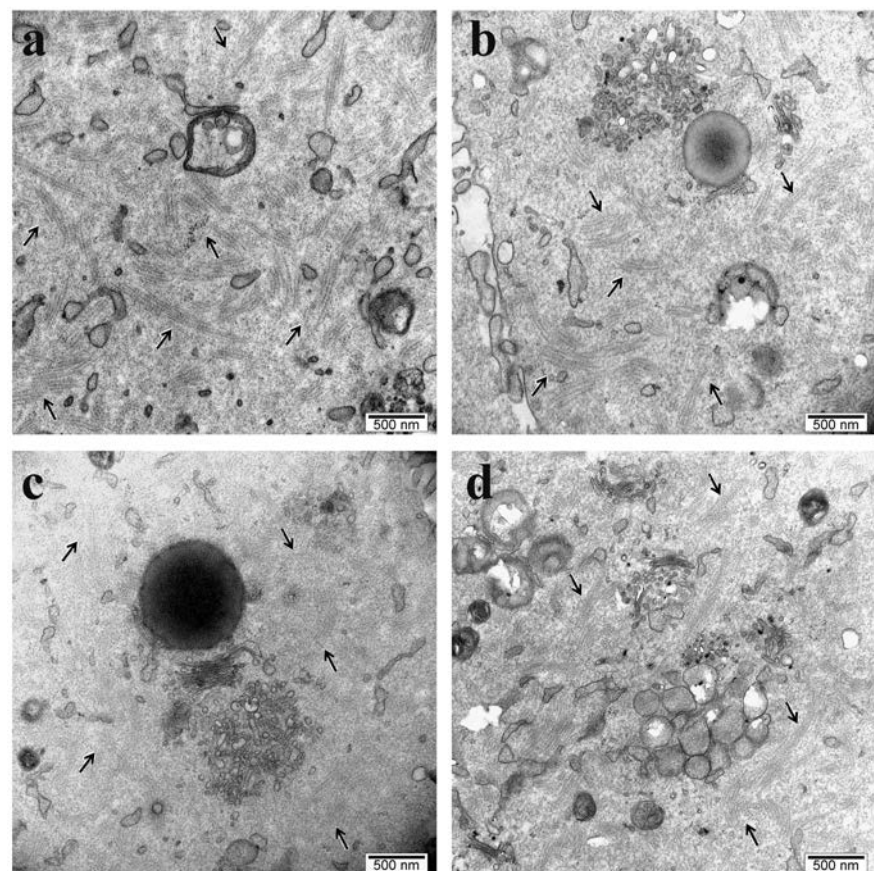


Figure 1. Representative TEM images of a) parthenogenetic NSN-derived 2-cell embryo and b) parthenogenetic SN-derived 2-cell embryo; c) NSN-derived 2-cell embryo obtained by ICSI; d) SN-derived 2-cell embryo obtained by ICSI. Arrows point to cytoplasmic lattices.

with lead. The samples were analyzed on a FEI-Tecnaï 12 electron microscope (FEI, Eindhoven, The Netherlands) and imaged on a side-view camera (Veleta, EMSIS, Münster, Germany).

Quantitative evaluation of CPLs

Cytoplasmic lattices were quantitatively evaluated by Image J software (Rasband, W.S., ImageJ, U. S. National Institutes of Health, Bethesda, Maryland, USA, <https://imagej.nih.gov/ij/>, 1997-2016).

Two independent operators evaluated 10 randomly selected areas of the cytoplasm for each of the SN and NSN embryo-derived types (Parthenotes and ICSI). The instrumental settings for the CPLs grey level evaluations was done independently by the two operators; the calculated integrated density was reported in arbitrary units normalized to the selected area (mean value \pm standard deviation). The two-tail test was used to analyze the distribution of the data that have a Gaussian distribution. Means and standard deviations were compared with the Student's *t*-test; differences were considered statistically significant when $P < 0.05$.

Results

Our main goal in this study is to contribute to detail the underlying cytological and molecular events controlling the very early embryo development, by taking a close examination of SN and NSN derived embryos on the ultrastructural level: we envision that the one and same biological machinery controls both the prospective embryo development and the arrest at the 2/4-cell stage.

Transmission electron microscopy analyses showed that CPLs are distributed throughout the cytoplasm of 2-cell embryos derived from both SN and NSN oocytes, without any appreciable difference between the two embryo production systems, as shown in Figure 1 a-d. The quantitative evaluations of the CPLs showed no significant differences ($P > 0.05$) among any of the experimental conditions considered (Figure 2).

In addition to the CPLs we noticed one more feature that seems to differentiate between the SN and NSN-derived embryos, also in respect to apoptosis: the abundant presence of vesicular mitochondria known to be associated with the cytochrome c release during the initial phases of apoptosis. These organelles have been observed mainly in the cytoplasm of NSN-derived 2-cell embryos, irrespective of the experimental conditions used (Figure 3). It is known that partial down-regulation of aerobic

metabolism in both oocytes and embryos causes a reduced embryo viability pointing out the role exerted by mitochondria on the cytoplasmic regulation of preimplantation embryo development.¹³

Discussion

We previously reported that the cytoplasm of mouse and human SN and NSN GV oocytes is differentially enriched in CPLs,^{7,8} thus correlating this feature (low number) with the inability of NSN derived embryos to proceed beyond the 2/4-cell stage of development. The detailed ultra-

structural analysis of SN and NSN 2-cell embryos obtained by parthenogenesis and ICSI is consistent with a model in which CPLs can form *de novo* during a specific timeline occurring during or immediately after the transition GV-zygote.

The important implication of our observation is that CPLs alone are not responsible for 2/4-cell embryos developmental arrest, contrary to previous assertions ("CPLs do not form after fertilization" and "the absence of the CPLs ultimately results in the development arrest"⁶ and "oocytes lacking CFP1 showed defects in CPLs formation that, among others, are responsible for defects in the acquisition of developmental competence after fertilization").¹⁴ In

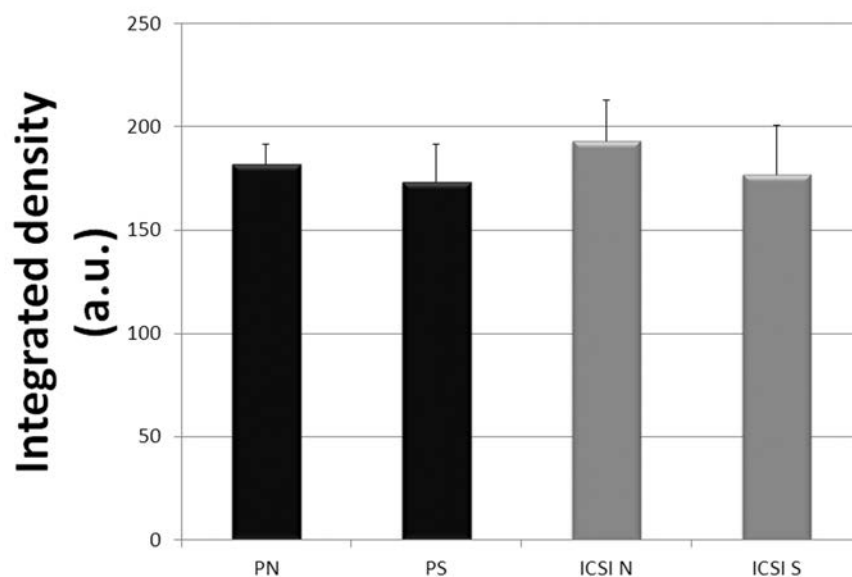


Figure 2. Integrated density evaluation (mean value \pm SD in arbitrary units, a.u.) of detectable CPLs in cytoplasmic representative areas (see materials and methods) of PN and PS (Parthenogenetic activated surrounded (S) and not-surrounded (N) nucleolus oocytes) and ICSI S and ICSI N (Intracytoplasmic Sperm Injected SN and NSN oocytes).

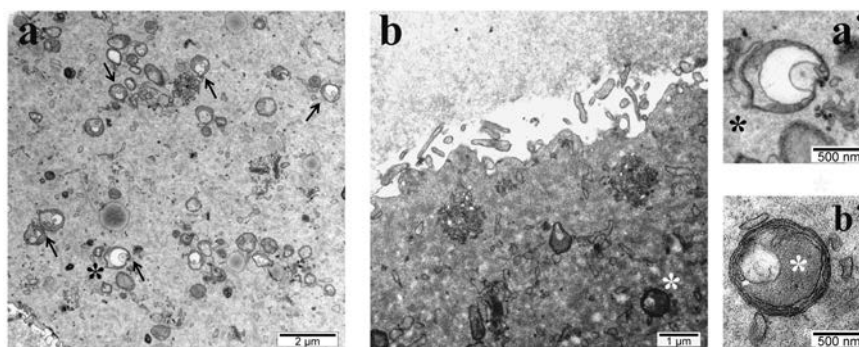


Figure 3. Representative TEM images of a) parthenogenetic NSN-derived 2-cell embryo with vesicular mitochondria (black arrows and black asterisk); b) SN-derived 2-cell embryo obtained by ICSI showing non-defective mitochondria (white asterisk); a' and b' magnification of a vesicular and normal mitochondria respectively.

order to link CPLs functions to the acquisition of oocyte genome architecture supportive of embryo development, we traced the development of embryos derived from SN and NSN GV oocytes, followed by two levels of inquiry. Firstly, we wished to confirm the well-known distinction whereby SN oocytes support development past the 2-cell stage, whereas NSN oocytes arrest at the 2-cell stage. We employed ICSI so as not to lose any oocytes to a failure of fertilization, which is not uncommon among *in vitro*-matured oocytes due to a hardening of the zona pellucida. Secondly, we examined the ultrastructure of these embryos, expecting to observe CPLs only in 2-cell embryos that were derived from SN oocytes. We employed chemical activation (resulting in diploid parthenogenesis) so as to preserve the pristine nature of the oocytes in respect to the CPLs (which are of maternal origin) and exclude any detrimental effects of the sperm injection on the ultrastructure. To our surprise, TEM analyses revealed the presence of CPLs in 2-cell embryos derived from both SN and NSN oocytes without any significant statistical difference (Figures 1 and 2).

We are aware that these preliminary observations require further investigations to fully understand this fascinating and challenging biological phenomenon. Nevertheless, we felt the urgent need to show that factors (*i.e.*, CPLs), hitherto considered responsible for embryo arrest at the 2-cell stage, are actually not related to this outcome. Instead, we wish to shift the attention to other different organelles/mechanisms (*i.e.*, abnormal mitochondria) that could, together with others, be the real player(s) of this intricate and extremely fascinating process that is the preimplantation

embryo development.

These results prelude to further research with the ultimate aim to find new markers for better (non-invasive) oocyte selection, toward preferential therapeutic use of SN oocytes in assisted reproductive technology procedures.

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