

Review

Allobetulin and Its Derivatives: Synthesis and Biological Activity

Wim Dehaen ^{1,*}, Anastasiya A. Mashentseva ^{1,2} and Talgat S. Seitembetov ³

¹ Molecular Design and Synthesis, Department of Chemistry, University of Leuven, Celestijnenlaan 200F, 3-3001 Leuven, Belgium

² Department of Chemistry, The L.N.Gumilev Eurasian National University, Munaitpasov str. 5, 010008 Astana, Kazakhstan; E-Mail: mashentseva.a@gmail.com

³ Department of Biochemistry, Medical University “Astana”, Beybetshilyk. 51, 010000 Astana, Kazakhstan; E-Mail: samankul46@mail.ru

* Author to whom correspondence should be addressed; E-Mail: wim.dehaen@chem.kuleuven.be.

Received: 22 February 2011; in revised form: 9 March 2011 / Accepted: 11 March 2011 /

Published: 14 March 2011

Abstract: This review covers the chemistry of allobetulin analogs, including their formation by rearrangement from betulin derivatives, their further derivatisation, their fusion with heterocyclic rings, and any further rearrangements of allobetulin compounds including ring opening, ring contraction and ring expansion reactions. In the last part, the most important biological activities of allobetulin derivatives are listed. One hundred and fifteen references are cited and the relevant literature is covered, starting in 1922 up to the end of 2010.

Keywords: triterpene; allobetulin; rearrangement; biological activity

1. Introduction

Triterpenes and triterpenoids are numerous and widely distributed in Nature. Biosynthetically, they are derived from squalene. Earlier studies have focused on the isolation and structural elucidation of the compounds, and there is still a lot of ongoing research in this area that has been regularly reviewed by Connolly and Hill [1]. During recent years, several interesting biological properties were found for this class of compounds, which in combination with their low toxicities lead to an increased research

effort [2,3]. More particularly, the oleanane group displays a number of significant pharmacological activities. Allobetulin (**2**) and its derivatives, obtained from the readily available lupane betulin (**1**), form a part of the oleanane group.

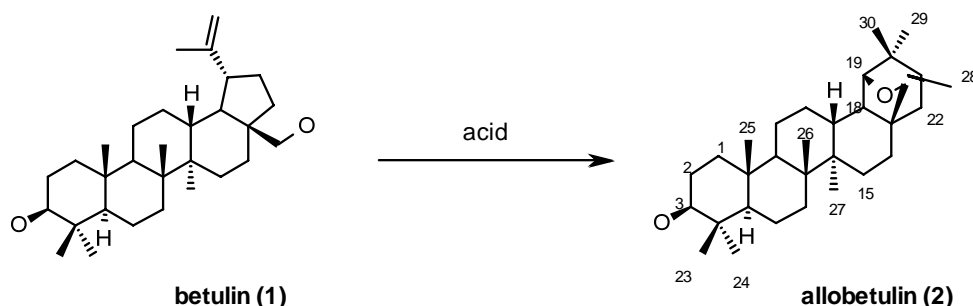
In this review, we summarize the chemistry of allobetulin analogs including: (1) their formation by rearrangement from betulin derivatives, (2) their further derivatisation, (3) their fusion with heterocyclic rings, and (4) the further rearrangements of allobetulin including ring opening, ring contraction and ring expansion reactions. In the final part (5), the most important biological activities of the allobetulin derivatives mentioned in sections 1–4 are listed.

There are also a number of allobetulin derivatives that are isolated from plant extracts. For a recent example see [4]. These will not be treated in this review. We also did not cover the chemistry of the ring contracted or *seco*-derivatives of allobetulin, other than their formation from allobetulin derivatives.

2. Betulin-Allobetulin Rearrangement

In 1922, Schulze and Pieroh reported that when betulin (**1**) was heated in formic acid, an unexpected formate ester product resulted, that gave an isomeric product after saponification that was named allobetulin (**2**) (Scheme 1) [5]. At that time, very little was known about the structure of (allo)-betulin due to the lack of adequate characterisation techniques, but the authors were able to conclude that the obtained product was a monoalcohol, containing an ether function and an otherwise strongly rearranged structure as compared to the dialcohol betulin (**1**). Dischendorfer *et al.* determined the correct molecular *bruto* formula of **2** not much later [6]. In the following years several authors carried out similar rearrangements and prepared derivatives of allobetulin (**2**), but breakthroughs regarding its structure came only after the work of Davy [7] who oxidized the acetate of allobetulin to the corresponding 28-oxo derivative, and then saponified it to the alcohol and oxidized this compound to oxyallobetulone (**3**). The latter was identical to a product (“ketone-lactone-A”) derived by rearrangement of betulonic acid. Only recently was an X-ray structure of allobetulin (**2**) reported [8].

Scheme 1. Rearrangement of betulin (**1**) to allobetulin (**2**).

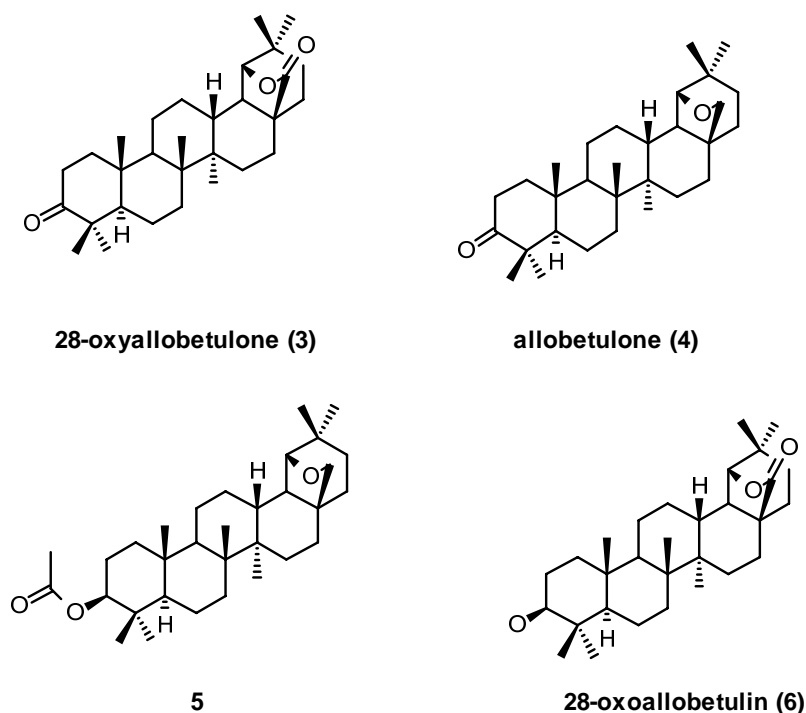


Various acidic conditions have been applied for this transformation, which is now known to belong to the class of Wagner-Meerwein rearrangements. Hydrobromic acid in chloroform [9], sulfuric acid in acetic acid [10], concentrated hydrochloric acid in ethanol [11,12] and even the highly toxic dimethyl sulfate [13] have been used for the transformation of **1** to **2** in moderate to good yields. The yield can be substantially improved by using acid reagents adsorbed on solid supports. Li *et al.* used “solid

acids” such as sulfuric acid or tosic acid on silica, Montmorillonite K10 and KSF, bleaching clays and kaolinite to obtain allobetulin and its derivatives in close to quantitative yield [14]. Pichette *et al.* have used ferric nitrate or ferric chloride absorbed on silica gel or alumina to convert betulin (**1**) into allobetulin (**2**) in excellent yield. Longer reaction times lead to the formation of allobetulone (**4**) or A-ring contracted products, respectively [15]. Ferric chloride hydrate itself (not supported) was also used for a larger scale reaction (approx. 5 g, 92% yield) [16]. Trifluoroacetic acid [17] or bismuth triflate (*via* triflic acid liberated by hydrolysis) [18] also give excellent results for this transformation. Russian researchers, including patent literature, mention the use of diluted sulfuric acid [19] and orthophosphoric acid [20] to combine the process of extraction of **1** from birch bark and rearrangement to **2**. This rearrangement can in fact be seen as an interesting undergraduate laboratory experiment [21].

Simple derivatives of betulin, such as betulone, 3-acetylbetulin, and betulinic acid have been transformed by the above methods to the corresponding allobetulin analogs allobetulone (**4**), 3-acetoxyallobetulin (**5**), and 28-oxoallobetulin (**6**). Betulinic acid is slower to rearrange in comparison to other betulin analogs and may give substantial amounts of side products. 28-Oxoallobetulin (**6**) may be prepared more effectively in two steps by rearrangement of the 3-acetylated betulinic acid, followed by hydrolysis [22]. As mentioned earlier, rearrangement of betulonic acid or its methyl ester [23] affords triterpene **3**, which can be reduced back to **6** (Figure 1) [24].

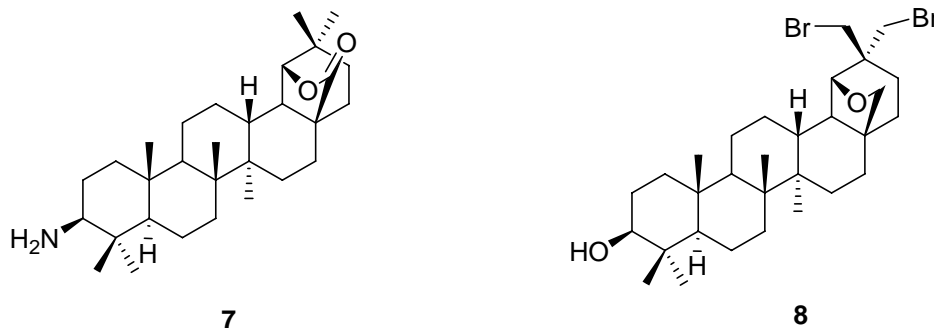
Figure 1. Structures of triterpenes 3-6.



Another example is the preparation of 3-amino-28-oxoallobetulin (**7**) after attempted trifluoroacetic acid deprotection of the corresponding Boc-protected betulinic acid derivative [25]. Treatment of betulin (**1**) with bromine was reported to give a good yield of the dibromoallobetulin (**8**) [26]. The structure of rearrangement product **8** was proven by X-ray crystallography. However, this good yield is difficult to reproduce so an efficient procedure towards this interesting product is still lacking. Pradhan

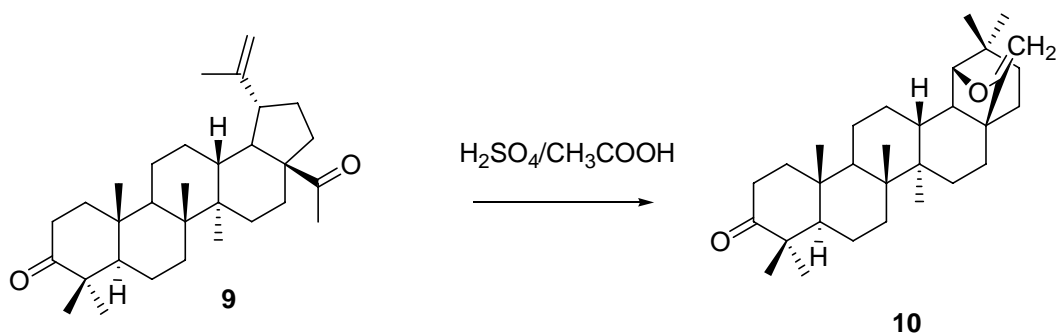
et al. likewise reported on the formation of the 3-acetylated 28-oxo analogs of **8** after treatment of the corresponding betulin derivative with *N*-bromosuccinimide in DMSO (Figure 2) [27-29].

Figure 2. Structures **7** and **8**.



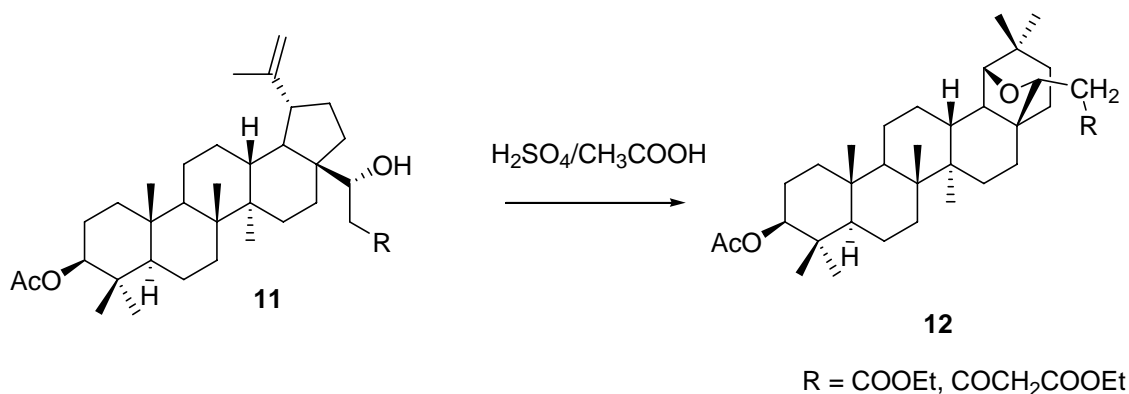
Davy *et al.* prepared an interesting enol ether analog **10** of allobetulin via rearrangement (20% sulfuric acid in acetic acid) of the acetyl derivative of betulone (**9**). Ozonolysis of compound **10** afforded 28-oxoallobetulone (**3**), proving the enol ether structure (Scheme 2) [30].

Scheme 2. Rearrangement of ketone **9** to enol ether **10**.



In the recent work of Czuk *et al.*, allobetulin homologues **12** were prepared in almost quantitative yields by trifluoroacetic acid induced rearrangement of secondary alcohols **11** that were prepared from 3-acetylated betulinic aldehyde by aldol condensation reactions (Scheme 3) [31].

Scheme 3. Rearrangement of alcohols **11** to allobetulin homologues **12**.

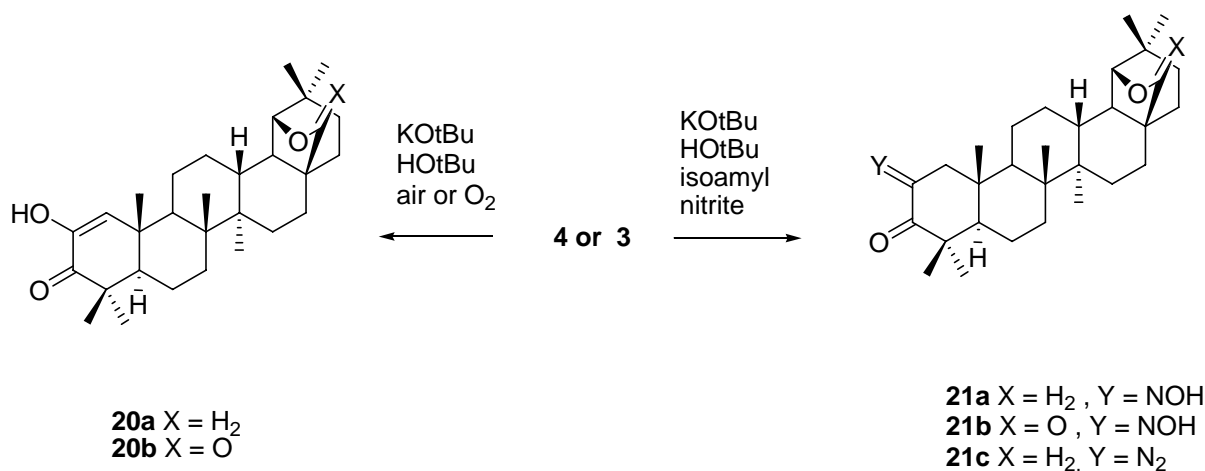


Allobetulone (**4**) was used to prepare the usual ketone analogues, such as the oxime **17a,b** (X = NOH, Y = H₂ or O) [5,37], and the phenylhydrazone **18** (X = N-NHPh) [5]. Epimeric spiro compounds **19a,b** were obtained (as a 1:2 mixture) from **4** by hetero-Diels-Alder cycloaddition reaction with benzoyl ketene generated *in situ* from 5-phenyl-2,3-dihydrofuran-2,3-dione (Figure 4). The two isomers **19a,b** were isolated and characterized by X-ray crystallography [38]. The effect of the substituents on the cumulene and aryl fragments on the stereoselectivity was studied [39].

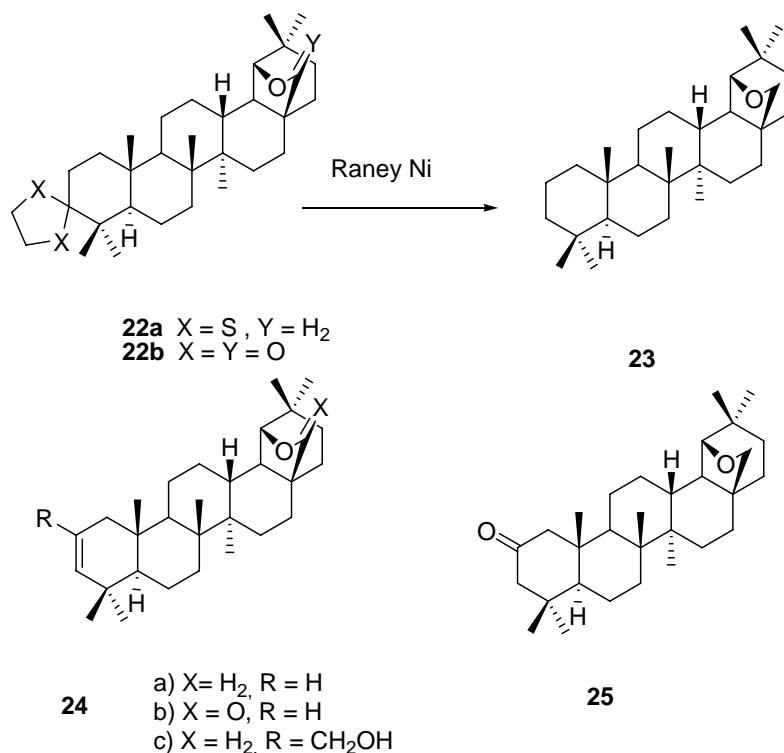
3-Acetoxyallobetulin (**5**) was oxidized to the lactone **6** with CrO₃ in acetic acid [5,40], similarly **3** was prepared starting from allobetulone (**4**) [6]. Zhang *et al.* succeeded to oxidize allobetulin (**2**) directly to 28-oxoallobetulone (**3**, 87% yield), using sodium periodate/ruthenium trichloride as the reagent [41]. The selective reduction of lactone **3** to 28-oxoallobetulin (**6**) is another viable alternative to prepare this compound [24].

Base catalyzed oxidation of allobetulone (**4**) with oxygen as the reagent affords the 2-hydroxy enone derivative of allobetulin (**20**) [34,40,42,43]. Similarly, the oximes **21a,b** (X = H₂, O) are prepared from **4** or **3** *via* condensation with isoamyl nitrite reagent [32]. Forster reaction of **21a** gave the corresponding 2-diazoallobetulone (**21c**, Scheme 4) [42].

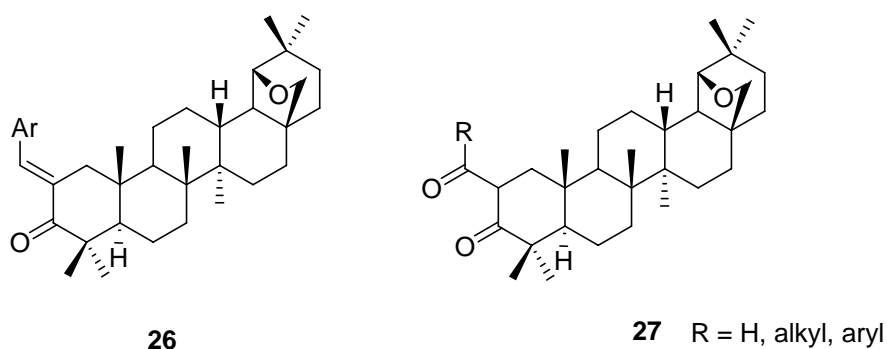
Scheme 4. Oxidation and oximation of ketones **3,4**.



Ethylenedithioketal **22a** (X = S, Y = H₂) was prepared from allobetulone (**4**) and reduced to allobetulane (**23**) with Raney nickel [44]. The 28-oxoallobetulone ketal **22b** (X = Y = O) was prepared in 86% yield from triterpene **3** and ethylene glycol [41]. Other ketals were also prepared from allobetulone (**4**) [45]. Allobetulenes **24a,b** that are of importance as biomarkers were prepared from allobetulin (**2**, X = H₂) or 28-oxoallobetulin (**6**, X = O) *via* tosylation in pyridine and elimination of toluenesulfonic acid [14]. The alkene **24a** is known from older work by the name of γ -allobetulin [46,47]. This alkene **24a** was subjected to the Prins reaction, leading to the alcohol **24c** (R = CH₂OH) [48]. Interestingly, allobetulone (**4**) was isomerized to the 2-keto triterpene **25** in the presence of sulfur and morpholine (Figure 5) [49].

Figure 5. Structures of compounds **22-25**.

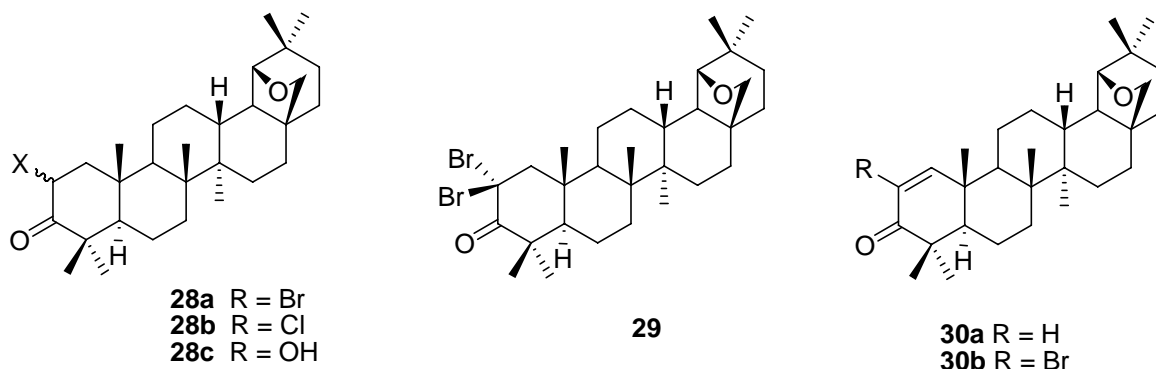
Aldol condensation reactions of allobetulone (**4**) with benzaldehydes and heterocyclic aldehydes lead to the α,β -unsaturated ketones **26** [6,50]. Similarly, Claisen condensations of **4** with formate and oxalic esters have been used to prepare the synthetically useful 1,3-diketones **27** or their enol tautomers (Figure 6) [16,51-53]. The formyl derivative **27** (R = H) was converted into the 2-fluoromethylidene derivative by treatment with DAST [54].

Figure 6. Structures of triterpenes **26** and **27**.

Both the 2-monobrominated (mixture of the 2 α - and 2 β -epimers) **28a,b** (X = Br) and the 2,2-dibrominated derivative **29** can be prepared by controlled reaction of allobetulone (**4**) with different brominating reagents [55-60]. The corresponding chlorinated derivatives **28a,b** (X = Cl) are also known [61]. The conformations of these brominated triterpene derivatives were studied in detail [58] and an X-ray structure of isolated 2 β -bromoallobetulone (**28b**) was reported. [62] Dehydrobromination of **28** or **29** gave unsaturated ketones **30a,b** [55,59]. Ketone **30a** (R = H) was also prepared directly via

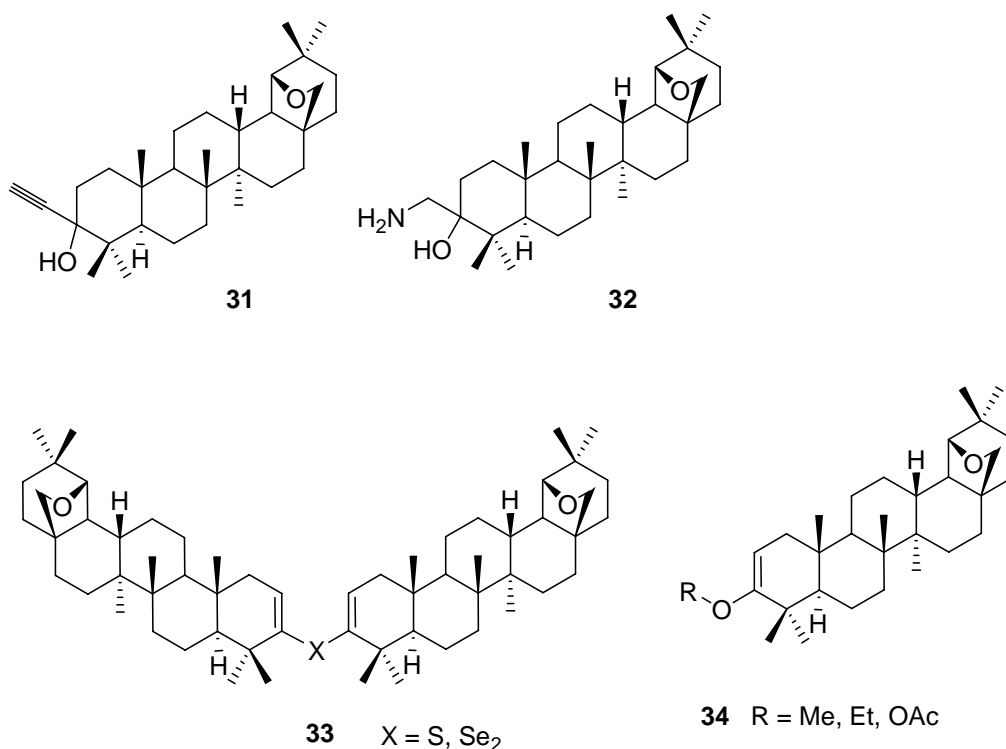
phenylselenenic anhydride oxidation of allobetulone **4** [39] and was used as a Michael acceptor for cyanide anion (Figure 7) [41].

Figure 7. Structures of triterpenes **28-30**.



Addition of acetylide to allobetulone (**4**) affords a monoacetylene **31** [63]. Trimethylsilylcyanide addition to **4** followed by reduction yields aminoalcohol **32** [35]. A dimeric bis(allobetulenyl) sulfide **33** (X = S) is isolated after treatment of allobetulone (**4**) with Lawesson's reagent [64] the corresponding diselenide **33** (X = Se₂) was isolated after attempted dehydrogenation of **4** with selenium dioxide [59]. Enol acetates and ethers **34** were also prepared starting from allobetulone (**4**) (Figure 8) [65,66].

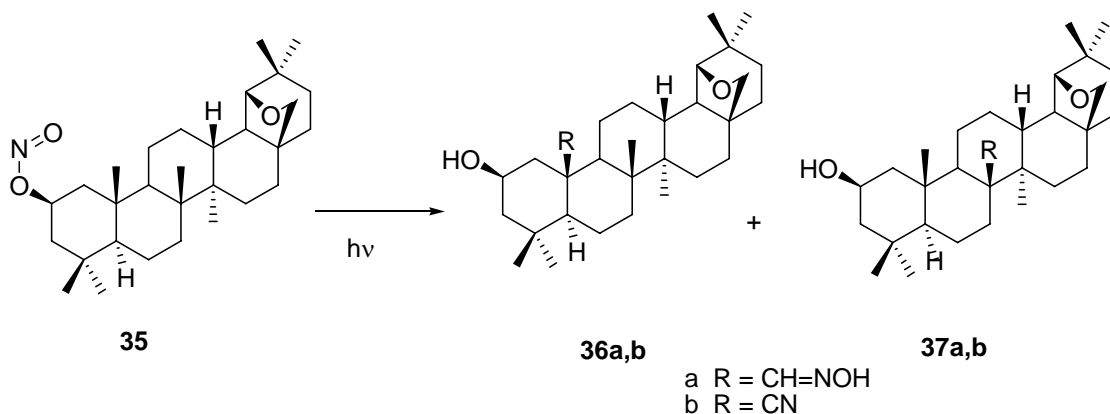
Figure 8. Structures of triterpenes **31-34**.



A remarkable photolytic transformation (Barton reaction) of 2-nitrite **35**, derived from the ketone **25** after reduction and esterification with nitrosyl chloride, lead to the formation of two regioisomeric

aldoximes **36** (40%) and **37** (40%) [67-69]. The remotely functionalized (C-25 and C-26) oximes **36a** and **37a** were further converted into nitriles **36b** and **37b** (Scheme 5).

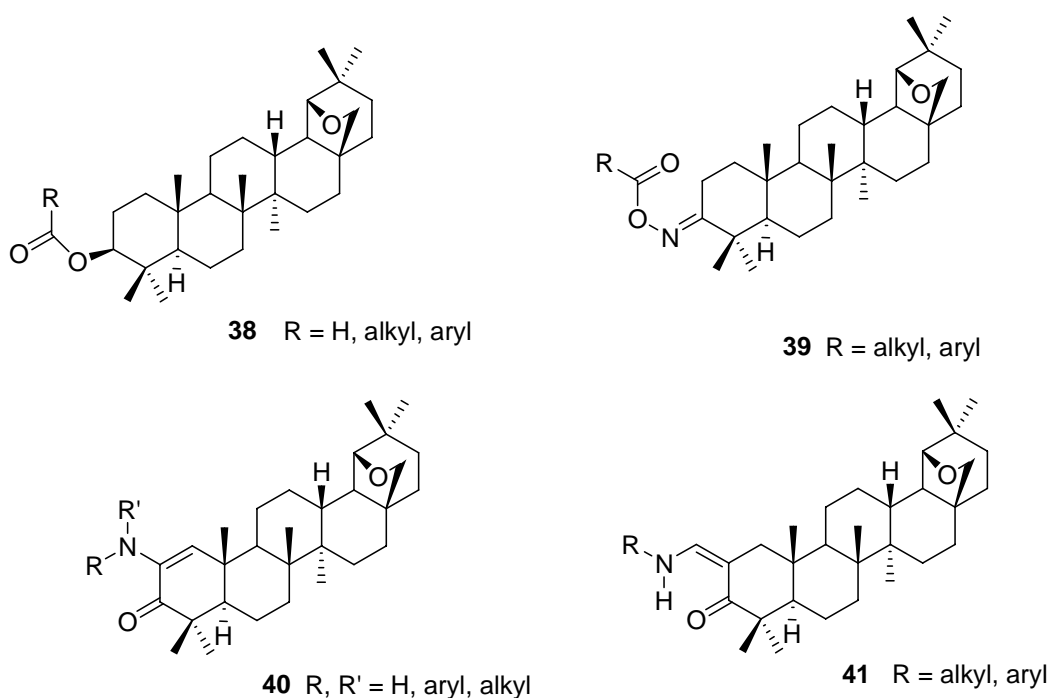
Scheme 5. Photolysis of nitrite **35**.



Simple ester derivatives **38** of allobetulin (**2**) or its 28-oxo analog **6** may be prepared *via* acylation of the 3- β OH function [5,70,71]. Acylation with cyclic anhydrides leads to monoacidic ester derivatives with improved water solubilities [71]. Alternatively, R groups containing ionic functionalities (ammonium, sulfonate) or polyethylene glycol solubilizers are attached [72]. Next to acylation, sulfonylation and phosphorylation reactions were also described [71]. The oximes **17a,b** may also be used to prepare O-acylated derivatives **39** [37,73].

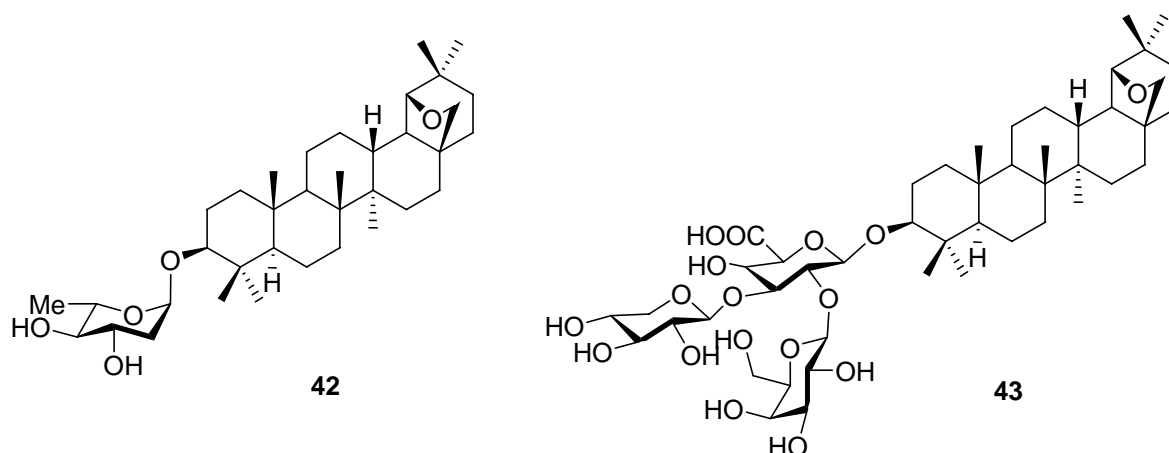
Enamines **40** are prepared from the enol acetate **34** (R = Ac) by epoxidation and condensation with primary or secondary amines. The reaction involves an oxidation, probably effected by adventitious oxygen [67]. 2-Alkylaminomethylene derivatives **41** of allobetulone (**4**) were prepared from the Claisen ester condensation product **27** (R = H) and primary amines (Figure 9) [53].

Figure 9. Structures of triterpenes **38-41**.



Glycosides and saponins with allobetulin (**2**) or its 28-oxo derivative **6** as aglycones were prepared by reacting the 3 β -hydroxy function with sugar derivatives, such as glycals [74-78] or a large collection of trichloroacetimidates [22,79-81]. This modification, see for instance glycoside **42** [78], saponin **43** [79] and the glycoside derivative of **20a** [40], greatly enhances water solubility and hence influences the biological properties (Figure 10).

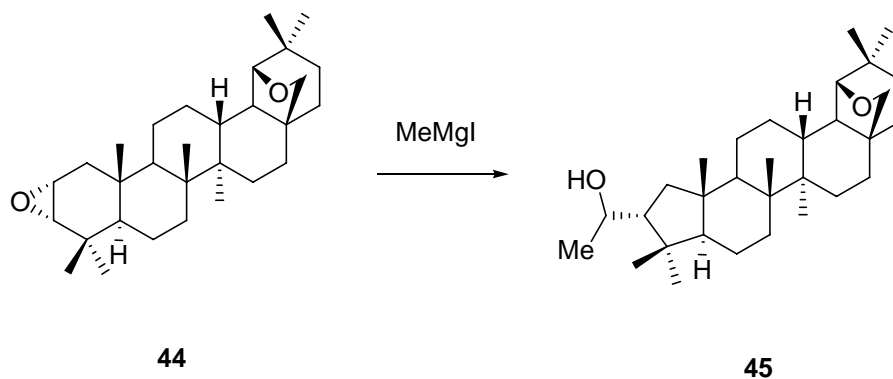
Figure 10. Structures of triterpene saponins **42** and **43**.



4. Ring Fusion to the A-Ring of Allobetulin

2,3-Epoxides may be formed by ring closure of the corresponding bromohydrins, available from reduction of 2-bromoallobetulones **28** (X = Br), by epoxidation of alkene **24a** [59,82] or by oxidation reactions of enol acetate **34** (R = Ac) as mentioned above [66]. The main feature of these epoxides is their propensity for ring opening reactions with nucleophiles [52,61]. Interestingly, 2 α ,3 α -epoxide **44** on treatment with a methyl Grignard reagent underwent rearrangement (see also next part 3) before addition of the organometal, affording nor-A alcohol derivative **45**. The 2 β ,3 β -isomeric epoxide underwent a similar rearrangement/addition sequence (Scheme 6) [83].

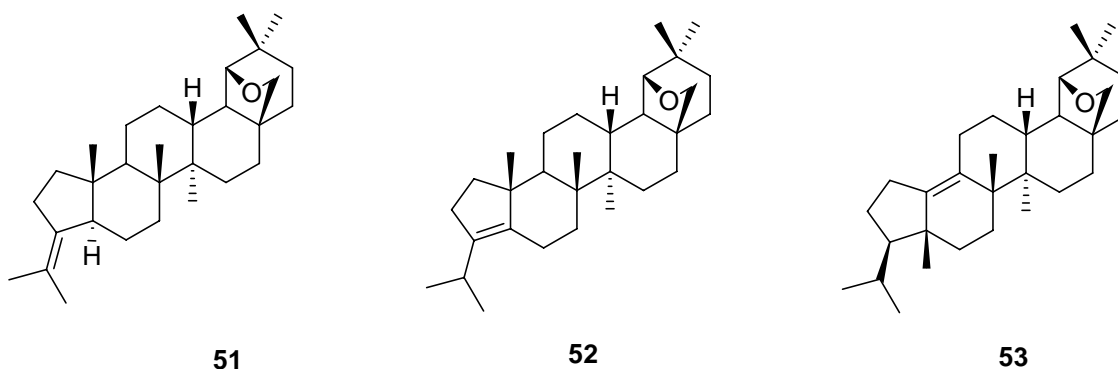
Scheme 6. Ring opening of epoxide **44** with Grignard reagent to afford **45**.



Allobetulone (**4**) and its substituted derivatives are the starting point for the annelation of allobetulin with heterocyclic rings. For instance, Fischer indole synthesis starting from arylhydrazine and ketone **4** gave the fused indole **46** [84,85]. 2-Hydroxyenone **20a** was condensed with diamines such as 1,2-

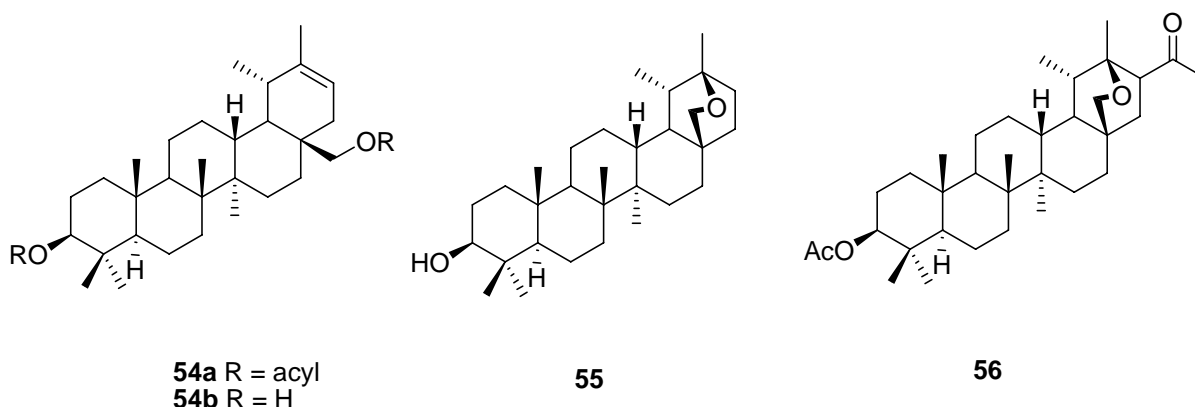
[15]. Silica- or alumina supported FeCl_3 hydrate gave a similar mixture (55:45 ratio) of **52** and **53** on extended reaction of betulin (**1**), via allobetulin (**2**) [16]. The reaction of allobetulin (**2**) with PCl_5 has been reinvestigated and was shown to lead directly to **52** at slightly higher temperatures (5–10 °C). At –10–0 °C, the expected **51** was formed [47]. The highest yields and selectivities of apoallobetulin isomers were obtained on treatment of betulin (**1**) with bismuth triflate. The relative amount of catalyst is important. Thus, heating **1** with 20 mol% catalyst for 40 h at reflux in dichloromethane gave 98% yield of **52**. On the other hand, heating of **1** or **52** for 8–15 h in the same solvent with 50 mol% bismuth triflate gave the isomer **53** almost quantitatively (96–98% yield) (Figure 13) [19].

Figure 13. Structures of apoallobetulins **51–53**.



Treatment of allobetulin (**2**) with acid chlorides in high boiling solvents leads to rearranged and ring opened diacylated products **54a**, that can be saponified to the so-called “heterobetulin” **54b**, which has an ursane framework [9,47,90,91]. “Alloheterobetulin” **55** is a ring closed isomer of the latter which can be obtained after treatment of **54b** with toluenesulfonic acid [92]. A remarkable rearrangement/O,C-diacylation was recently reported to occur (55% yield of **56**) when allobetulin (**2**) was treated with acetic anhydride and a few drops of perchloric acid (Figure 14) [93].

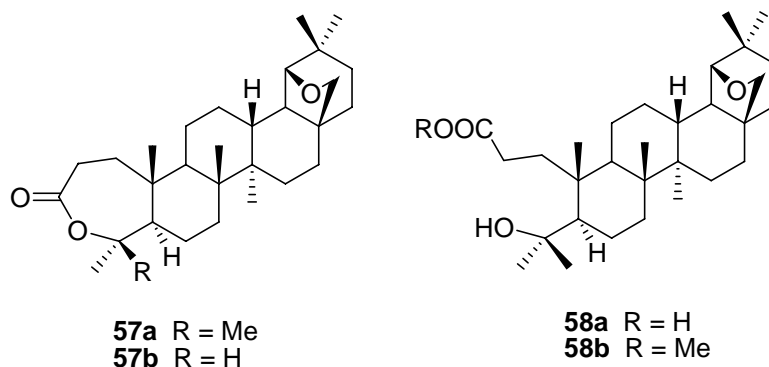
Figure 14. Structures of ring opened allobetulin derivatives **54–56**.



The Baeyer-Villiger oxidation of allobetulone (**4**) was investigated by different groups under different circumstances [40,94,95]. With MCPBA in dichloromethane, the main product (83%) is the ring-expanded lactone **57a**. Other peracids (performic, peracetic) give similar results. However, reaction of **4** with MCPBA in the presence of acid (acetic + sulfuric) leads to the formation of a nor-

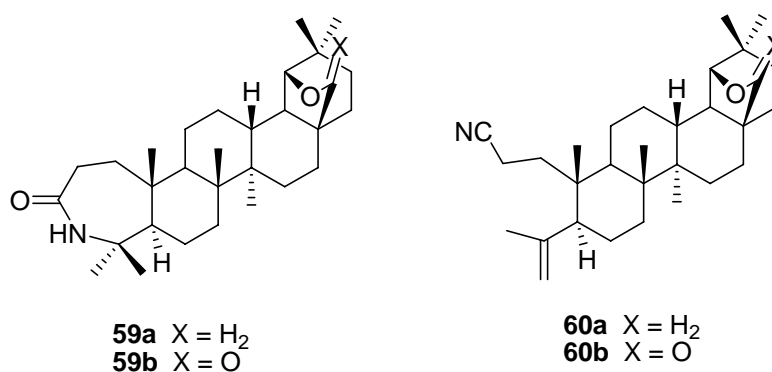
lactone **57b**. The 3,4-*seco* derivatives **58a,b** were obtained in good yield either from **57a** by alkaline hydrolysis or directly from **4**, carrying out the oxidation in methanol with a trace amount of sulfuric acid [95]. Larger amounts of acid (0.15%) lead to the formation of the 2 α -hydroxyallobetulone **28c** in good yield (86%) (Figure 15) [65].

Figure 15. Structures of A-*seco* derivatives **57** and **58**.

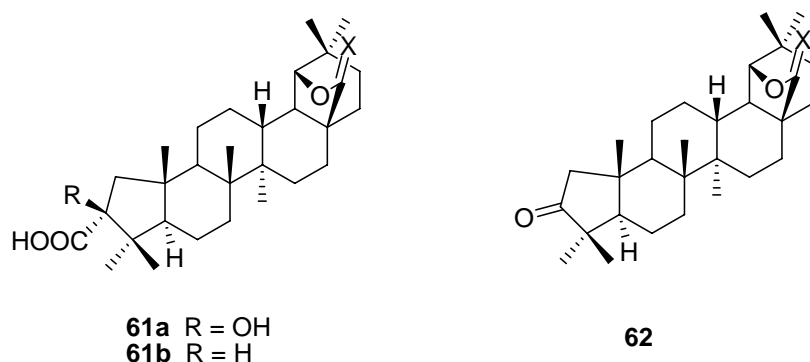


The Beckmann rearrangement of allobetulin oxime **17a**, induced by TsCl/pyridine or phosphoryl chloride, gave rise to the formation of a lactam **59a** (major product) and a 3,4-*seco*-triterpene nitrile **60** (minor product). The lactam **59a** could be transformed into the nitrile **60** on extended heating [96,97]. Upon Schmidt reaction of methyl betulonate or Beckmann rearrangement (POCl₃) of its oxime, the 28-oxo derivatives **59b** and **60b** were formed after two consecutive rearrangements [98]. Other 2,3-*seco*-derivatives were prepared via Beckmann fragmentation of allobetulin derivatives (Figure 16) [33,45,99,100].

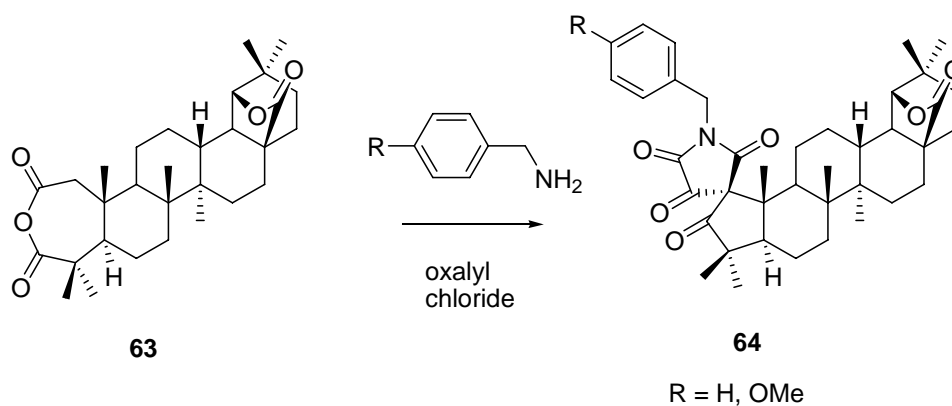
Figure 16. Structures of ring expanded and ring opened triterpenes **59** and **60**.



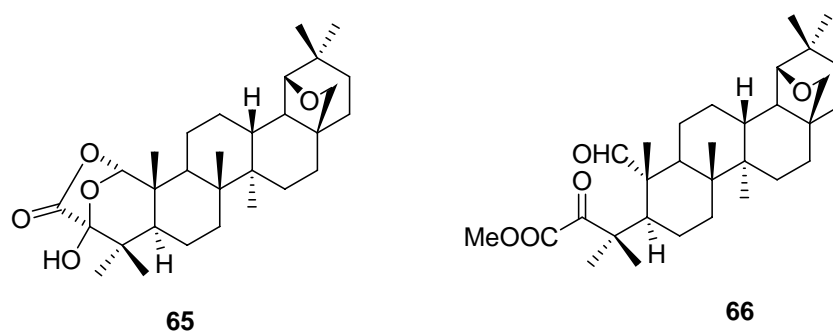
The dibromoallobetulin **29** underwent a *quasi*-Favorskii rearrangement on treatment with base, leading to the ring contracted product **61a**. Oxidative decarboxylation of the latter with lead tetraacetate gave the norketone **62** [45,56,96,101]. The latter is an interesting starting material that was used in many follow-up reactions that will not be discussed here. Benzilic acid rearrangement of diketone **20a** gives the same hydroxyacid **61a**. The photochemical Wolff rearrangement of diazo compound **21c** gave the ring contracted carboxylic acid **61b** (Figure 17) [45].

Figure 17. Structures of A-ring contracted triterpenes **61** and **62**.

Dischendorfer reported oxidation of the A-ring of 28-oxoallobetulone **3** to “allobetulinic acid” which formed a cyclic anhydride **63** [102,103]. This *seco*-derivative **63** was recently used to prepare spirocyclic derivatives **64** after treatment with benzylamines and oxalyl chloride [104]. Recently, the diacid analog of **63** was prepared by ozonolysis of the Claisen ester condensation product of **3** (*i.e.* the 3-oxo analog of **27**) [105]. This procedure has some similarity with earlier work by Ruzicka, who used chromic acid to prepare the diacid from hydroxymethyleneallobetulone (**27**, R = H) or directly from allobetulin (**2**) (Scheme 7) [106].

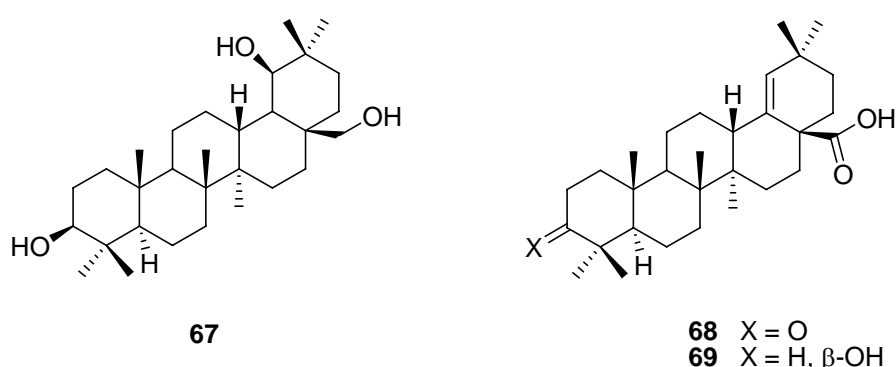
Scheme 7. Spirocyclic triterpenes **64**.

The nearly insoluble lactone **65** was formed in low yield (24%) on oxidation of allobetulone (**4**) with chromic acid. Treatment of **65** with diazomethane gave the 1,2-*seco* derivative **66** in good yield (Figure 18) [107].

Figure 18. Structures of oxidized triterpenes **65** and **66**.

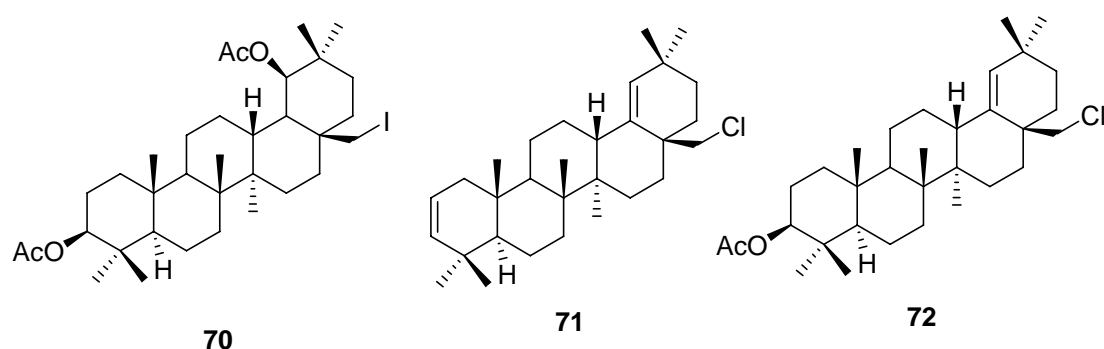
Another obvious position for ring cleavage is the lactone bridge of 28-oxoallobetulin derivatives. This bridge is quite stable towards saponification, but LiAlH_4 reduction of the 3β -acetoxy derivative of **5** [108] or 28-oxoallobetulone (**3**) [25,30,109] gives a germanicanetriol derivative **67** which was further transformed to different germanicanes by selective acylation, oxidation and dehydration reactions [108,109]. The lactone ring of the protected 28-oxoallobetulone **22b** was reductively cleaved with LiAlH_4 and after deprotection, 28-acetylation, dehydration with POCl_3 , saponification, and stepwise oxidation, moronic acid (**68**) and the reduced morolic acid **69** were obtained (Figure 19) [41]. In general, allobetulin and its derivatives are important starting materials for the synthesis of rare germanicanes and olean-18(19)-ene triterpenoids.

Figure 19. Structures of triterpenes **67-69**.



Treatment of allobetulin (**2**) with sodium iodide/acetyl chloride at reflux in acetonitrile lead to the formation of iodinated diacetate **70** [110]. Treatment of allobetulin (**2**) or its 3-acetate with POCl_3 in refluxing pyridine similarly gave dialkene **71** or the corresponding acetate **72** (Figure 20) [16].

Figure 20. Structures of triterpenes **70-72**.



6. Biological Properties of Allobetulin Analogs

The biological properties of betulin, betulinic acid and its derivatives are well known [2] and often activity studies of allobetulin derivatives are found back in the literature together with or in comparison to their betulin isomers. A wide spectrum of biological properties have been reported, including antiviral, antifeedant, immunotropic, antibacterial, antifungal, and anti-inflammatory activities, cytotoxicity and inhibition of glycogen phosphorylase activities.

6.1. Antiviral properties

In 1995, it was found that allobetulin (**2**) itself showed moderate inhibitory activity against the influenza B virus [111]. It was claimed in the patent literature that different derivatives of allobetulin, including **2** and its 3-*O*-acylated and phosphorylated derivatives, **4**, **30a**, **32**, exhibited significant antiviral activity and could be used to treat herpes virus (HSV-herpes simplex virus) infection [35]. Also in 2002, compound **3** was shown in cell culture to inhibit influenza A growth while being inactive against HSV and the enterovirus ECHO-6 [112]. Somewhat later, allobetulin derivatives **3**, **6**, and different *O*-acylated oximes **39** were tested against several viruses such as HSV, influenza and ECHO-6 [24,37]. In fact, the non-acetylated oxime **17a** had the largest effect against influenza virus A, while being only moderately active against enterovirus ECHO-6 and inactive with respect to HSV. The *N*-acetylated oximes **39** had a moderate activity towards HSV, but were inactive against the other viruses [37]. It was confirmed that 28-oxoallobetulone (**3**) strongly inhibited the influenza virus, but did not influence HSV reproduction [24]. Rearranged product **56** showed only moderate inhibition of the *Papilloma* virus [93].

6.2. Antifeedant properties

In 1990, Lugemwa *et al.* reported high antifeedant activity against the bollworm larvae, *Heliothis zea*, for the glycoside derivative of **30a**. Simple allobetulin derivatives such as **2**, **20a** and **30a** itself were not active. The antifeedant property was selective and the glycoside did not display high activity against either the Colorado potato beetle (*Leptinotarsa decemlineata*) or the fall armyworm (*Spodoptera frugiperda*) [40].

6.3. Immunotropic activities

Different 2-substituted allobetulone derivatives, including the formyl analog **27** (R = H) and different condensation products with amines **41** were screened [53]. In fact, compounds **27** and **41** (R = ⁱPr) had the most promising activity combined with low toxicity. In later work it was shown that these compounds had high biological activity on chronic administration and that their immunosuppressive activity was the result of toxic effect on the lymphocytes [113].

6.4. Antibacterial and antifungal activities

Compounds **2** and **3** were taken into a screening of 32 betulin derivatives against *Chlamydia pneumoniae*. Allobetulin (**2**) was equal to betulin (**1**) in antichlamydial activity (48%), but 28-oxoallobetulone (**3**) was inactive [114].

6.5. Anti-inflammatory and anti-ulcer properties

Biological tests on mice with the carrageenan and formalin edema models showed that acylated derivatives of allobetulin (**2**) possessed anti-inflammatory activity comparable to ortophen (diclofenac) [71,115]. Moderate antiulcer activity of **3** and 3-*O*-acylated allobetulin derivatives were observed in mice [112,115].

6.6. Cytotoxicity

The cytotoxicity of pyrazine and quinoxaline derivatives of allobetulin (**47**) was tested against a T-lymphoblastic leukemia cell line and found to be lower than the fused triterpene analogs based on unrearranged betulin or betulinic acid [86]. Pichette *et al.* did a study on the cytotoxicity of betulin- and allobetulin-derived 3 β -*O*-monodesmosidic saponins (such as **42**) with higher hydrosolubility and better pharmacokinetics. *In vitro* anticancer activity of saponins derived from **2** and **6** showed that the bioactivity for these glycosides was only moderate (IC₅₀ 30–40 μ M/L), as compared to the corresponding betulinic acid derivatives (IC₅₀ 7.3–10.1 μ M/L) [23]. The *in vitro* toxicity of **67** (human lung carcinoma or human colorectal adenocarcinoma assay) was comparable to that of betulinic acid or 5-fluorouracil [108]. Chacotrioside saponins such as **43** were fourfold superior to betulinic acid against human breast (MCF7) and prostate (PC-3) adenocarcinomas cell lines. Moreover, chacotriosides bearing non-polar functions at the C-28 position had a haemolytic activity against red blood cells [80]. Allobetulin derivatives **12** with 28-functionality were reported by Czuk *et al.* to have moderate cytotoxicity [31].

6.7. Inhibition of glycogen phosphorylase

Morolic acid (**69**) (IC₅₀ 70.3 μ M/L), its 3-epimer (IC₅₀ 34.5 μ M/L) and 3-*O*-acetylated derivative (IC₅₀ 32.7 μ M/L) were shown to cause moderate inhibitory activity against rabbit muscle glycogen phosphorylase [41].

7. Conclusions

Allobetulin and its analogues are easily accessible starting from the corresponding betulin derivatives. Although a large structural variety of allobetulin analogs is already available by functionalisation, ring fusion to the A ring, further rearrangements, ring contractions, ring expansions, and ring cleavages, there is still much chemical space unexplored. Further investigations are certainly worthwhile because of the interesting bioactivities.

Acknowledgements

W.D. thanks the University of Leuven, the F.W.O-Vlaanderen, and the I.U.A.P. for continuing financial support.

References

1. Connolly, J.D.; Hill, R.A. Triterpenoids. *Nat. Prod. Rep.* **2010**, *27*, 79-132 and the previous reviews in this series.
2. Tolstikov, G.A.; Flekhter, O.B.; Shul'ts, E.E.; Baltina, L.A.; Tolstikov, A.G. Betulin and its derivatives. Chemistry and biological activity. *Khim. Interes. Ust. Razv.* **2005**, *13*, 1-30.
3. Dzubak, P.; Hajdich, M.; Vydra, D.; Hustova, A.; Kvasnica, M.; Biedermann, D.; Markova, L.; Sarek, J. Pharmacological activities of natural triterpenoids and their therapeutic implications *Nat. Prod. Rep.* **2006**, *23*, 394-411.

4. Pathak, A.; Kulshreshtha, D.K.; Maurya, R. Coumaryl triterpene lactone, phenolic and naphthalene glycoside from stem bark of *Diospyros angustifolia*. *Phytochemistry* **2004**, *65*, 2153-2158.
5. Schulze, H.; Pieroh, K. Zur Kenntnis des Betulins. *Chem. Ber.* **1922**, *55*, 2322-2346.
6. Dischendorfer, O.; Juvan, H. Untersuchungen auf dem Gebiete der Phytochemie—Über das Allobetulin. *Monatsh. Chem.* **1930**, *52*, 272-281.
7. Davy, G.S.; Halsall, T.G.; Jones, E.R.H.; Meakins, G.D. The chemistry of the triterpenes. Part X. The structures of some isomerisation products from betulin and betulinic acid. *J. Chem. Soc.* **1951**, 2702-2705.
8. Santos, R.C.; Pinto, R.M.A.; Beja, A.M.; Salvador, J.A.R.; Paixão, J.A. 19 β ,28-Epoxy-18 α -olean-3 β -ol. *Acta Cryst.* **2009**, *E65*, o2088-o2089.
9. Dischendorfer, O. Phytochemical studies. I. Betulin. *Monatsh. Chem.* **1923**, *44*, 123-139.
10. Barton, D.H.R.; Holness, N.J.; Triterpenoids. V. Some relative configurations in rings C, D, and E of the β -amyrin and the lupeol group of triterpenoids. *J. Chem. Soc.* **1952**, 78-92.
11. Lawrie, W.; McLean, J.; Taylor, G.R. Triterpenoids in the bark of mountain ash *Sorbus aucuparia*. *J. Chem. Soc.* **1960**, 4303-4308.
12. Errington, S.G.; Ghisalberti, E.L.; Jefferies, P.R. The chemistry of the Euphorbiaceae. XXIV. Lup-20(29)-ene-3 β ,16 β ,28-triol from *Beyeria brevifolia* var *brevifolia*. *Aust. J. Chem.* **1976**, *29*, 1809-1814.
13. Linkowska, E. Triterpenoids. Part XI. Isomerization of betulin and its derivatives. *Pol. J. Chem.* **1994**, *68*, 875-876.
14. Li, T.S.; Wang, J.X.; Zheng, X.J. Simple synthesis of allobetulin, 28-oxyallobetulin and related biomarkers from betulin and betulinic acid catalysed by solid acids. *J. Chem. Soc. Perkin Trans. 1* **1998**, 3957-3965.
15. Lavoie S.; Pichette A.; Garneau F.-X.; Girard, M.; Gaudet, D. Synthesis of betulin derivatives with solid supported reagents. *Synth. Commun.* **2001**, *31*, 1565-1571.
16. Kazakova, O.B.; Khusnutdinova, E.F.; Tolstikov, G.A.; Suponitsky, K.Y. Synthesis of new olean-18-(19)-ene derivatives from allobetulin. *Russ. J. Bioorg. Chem.* **2010**, *36*, 512-515.
17. Medvedeva, N.I.; Flekhter, O.B.; Kukovinets, O.S.; Galin, F.Z.; Tolstikov, G.A.; Baglin, I.; Cavé, C. Synthesis of 19 β ,28-epoxy-23,24-dinor-A-neo-18 α -olean-4-en-3-one from betulin. *Russ. Chem. Bull.* **2007**, *56*, 835-837.
18. Salvador, J.A.R.; Pinto, R.M.A.; Santos, R.C.; Le Roux, C.; Beja, A.M.; Paixão, J.A. Bismuth triflate-catalyzed Wagner-Meerwein rearrangement in terpenes. Application to the synthesis of the 18 α -oleanone core and A-neo-18 α -oleanene compounds from lupanes. *Org. Biomol. Chem.* **2009**, *7*, 508-517.
19. Levdanskii, V.A.; Levdanskii, A.V.; Kuznetsov, B.N. Method of producing allobetulin from birch bark via isomerization of betulinol. *Russ. Patent RU 2374261*, 2009, [CA 2009, 151, 571219].
20. Kuznetsova, S.A.; Kuznetsov, B.N.; Red'kina, E.S.; Skvortsova, G.P. Method of allobetulin production. *Russ. Patent RU 2334759*, 2008, [CA 2008, 149, 378906].

21. Green, B.; Bentley, M.D.; Chung, B.Y.; Lynch, N.G.; Jensen, B.L. Isolation of betulin and rearrangement to allobetulin. A biomimetic natural product synthesis. *J. Chem. Ed.* **2007**, *84*, 1985-1987.
22. Thibeault, D.; Gauthier, C.; Legault, J.; Bouchard, J.; Dufour, P.; Pichette, A. Synthesis and structure-activity relationship study of cytotoxic germanicane- and lupane-type 3 β -O-monodesmosidic saponins starting from betulin. *Bioorg. Med. Chem.* **2007**, *15*, 6144-6157.
23. Vystrčil, A.; Stejskalova-Vondraskova, E.; Cerny, J. Identity of gratiolone and betulic acid. *Collect. Czech. Chem. Commun.* **1959**, *24*, 3279-3286.
24. Flekhter, O.B.; Boreko, E.I.; Nigmatullina, L.R.; Pavlova, N.I.; Medvedeva, N.I.; Nikolaeva, S.N.; Ashavina, O.A.; Savinova, O.V.; Baltina, L.A.; Galin, F.Z.; Zarudii, F.S.; Tolstikov, G.A. Synthesis and antiviral activity of lupane triterpenoids and their derivatives. *Pharm. Chem. J.* **2004**, *38*, 355-358.
25. Giniyatullina, G.V.; Flekhter O.B.; Baikova, I.P.; Starikova, Z.A.; Tolstikov, G.A. Effective synthesis of methyl 3 β -amino-3-deoxybetulinate. *Chem. Nat. Comp.* **2008**, *44*, 603-605.
26. Lezhneva, M.Y.; Schultz, E.E.; Bagryanskaya, I.Y.; Gatilov, Y.V.; Shakirov, M.M.; Tolstikov, G.A.; Adekenov, S.M. Synthesis and crystal structure of 29,30-dibromoallobetulin. *Chem. Nat. Comp.* **2006**, *42*, 186-188.
27. Pradhan, B.P.; Chakraborty, D.K.; Roy, A. Novel method for reductive cleavage of triterpenoid lactones with lithium in ethylenediamine. *Ind. J. Chem. B Org. Chem. Med. Chem.* **1993**, *32B*, 721-725.
28. Pradhan, B.P.; Chakraborty, D.K.; Dutta, S.; Ghosh, R.; Roy, A. Studies on the reactions of N-bromosuccinimide in dimethyl sulfoxide: Part IV. Action on lupenyl acetate. *Ind. J. Chem. B Org. Chem. Med. Chem.* **1991**, *30B*, 32-37.
29. Pradhan, B.P.; Mukherjee, M.M.; Chakrabarti, D.K.; Shoolery, J.N. Action of N-bromosuccinimide on triterpene acids and esters in dimethyl sulfoxide. *Ind. J. Chem. B Org. Chem. Med. Chem.* **1983**, *22B*, 12-16.
30. Davy, G.S.; Halsall, T.G.; Jones, E.R.H. The chemistry of the triterpenes. Part IX. Elucidation of the Betulin-Oleanolic acid relationship. *J. Chem. Soc.* **1951**, 2696-2702.
31. Csuk, R.; Barthel, A.; Kluge, R.; Kommera, H. Synthesis and biological evaluation of antitumour-active betulin derivatives. *Bioorg. Med. Chem.* **2010**, *18*, 1344-1355.
32. Dračinský, M.; Richtr, V.; Křeček, V.; Sejbal, J.; Klinot, J.; Buděšínský, M. Preparation and conformational study of 19 β ,28-epoxy-18 α -olean-5-ene derivatives *Collect. Czech. Chem. Commun.* **2006**, *71*, 387-410.
33. Tolmacheva, I.A.; Nazarov, A.V.; Maiorova, O.A.; Grishko, V.V. Synthesis of lupane and 19 β ,28-epoxy-18 α -oleane 2,3-*seco*-derivatives based on betulin. *Chem. Nat. Comp.* **2008**, *44*, 606-611.
34. Carlson, R.M.; Gibson, D.J. Anti-fungal formulation of triterpene and essential oil. *PCT Int. Appl. WO 2004089357*, 2004, [CA 2004, 141, 370537].
35. Carlson, R.M. Preparation of allobetulin triterpenoids for therapeutic use in the treatment of herpes virus infection. *U.S. Patent US 6369101*, 2002, [CA 2002, 136, 279583].

36. Flekhter, O.B.; Ashavina, O.Y.; Smirnova, I.E.; Baltina, L.A.; Galin, F.Z.; Kabal'nova, N.N.; Tolstikov, G.A. Selective oxidation of triterpene alcohols by sodium hypochlorite. *Chem. Nat. Comp.* **2004**, *40*, 141-143.
37. Flekhter, O.B.; Boreko, E.I.; Nigmatullina, L.R.; Pavlova, N.I.; Medvedeva, N.I.; Nikolaeva, S.N.; Tret'yakova, E.V.; Savinova, O.V.; Baltina, L.A.; Karachurina, L.T.; Galin, F.Z.; Zarudii, F. S, Tolstikov, G.A. Synthesis and pharmacological activity of acylated betulonic acid oximes and 28-oxo-allobetulone. *Pharm. Chem. J.* **2004**, *38*, 148-152.
38. Rybalova, T.V.; Galitov, Y.V.; Nekrasov, D.D.; Rubtsov, A.E.; Tolstikov, A.G. Synthesis of spiro[(6-phenyl-3,4-dihydro-2H-1,3-dioxine)-2R(S), 3'-(19',28'-oxidooleanan)]-4-ones and X-ray diffraction analysis of their configuration. *J. Struct. Chem.* **2005**, *46*, 1126-1130.
39. Nekrasov, D.D.; Obukhova, A.S.; Lisovenko, N.Y.; Roubtsov, A.E. Effect of substituents in the cumulene and aryl fragments of aroylketenes on the stereoselectivity of Diels-Alder heteroreaction with mono-, bi, and polycyclic terpenoids containing a carbonyl group. *Chem. Het. Comp.* **2010**, *46*, 413-418.
40. Lugemwa, F.N.; Huang, F.-Y.; Bentley, M.D.; Mendel, M.J.; Alford, A.R. A heliothis-zea antifeedant from the abundant birchbark triterpene betulin. *J. Agric. Food Chem.* **1990**, *38*, 493-496.
41. Zhang, P.; Hao, J.; Liu, J.; Zhang, L.; Sun, H. Efficient synthesis of morolic acid and related triterpenes starting from betulin. *Tetrahedron* **2009**, *65*, 4304-4309.
42. Huneck, S. Triterpenes. XV. Preparation and Forster reaction of 19 β , 28-epoxy-3-oxo-2-anti-oximino-1 α -cyano-18 α H-oleanane and the bromination of 19 β ,28-epoxy-3-oxo-1 α -cyano-18 α H-oleanane. *Chem. Ber.* **1965**, *98*, 3204-3209.
43. Korovin, A.V.; Tkachev, A.V. Synthesis of quinoxalines fused with triterpenes, ursolic acid and betulin derivatives. *Russ. Chem. Bull.* **2001**, *50*, 304-310.
44. Protiva, J.; Ocadlik, R.; Klinotova, E.; Klinot, J.; Vystrčil, A. Triterpenes. Part LXXXI. Ethylene dithioketals in ring A of 19 β , 28-epoxy-18 α -oleanane; mass spectra and reduction with deuterated Raney nickel. *Coll. Czech Chem. Commun.* **1987**, *52*, 501-507.
45. Huneck, S. Triterpenes. X. Photochemical transformations. 2. Preparation of 19 β , 28-epoxy-3-oxo-2-diazo-18 α H-oleanane and its photochemical conversion to A-nor compounds. *Chem. Ber.* **1965**, *98*, 1837-1857.
46. Ruzicka, L.; Brungger, H.; Gustus, E.L. Polyterpenes and polyterpenoids. LXVIII. Betulin. *Helv. Chim. Acta* **1932**, *15*, 634-648.
47. Klinot, J.; Vystrčil, A. Side-products in the conversion of allobetulin to heterobetulin. *Collect. Czech. Chem. Commun.* **1964**, *29*, 516-530.
48. Rybina, A.V.; Shepelevich, I.S.; Talipov, R.F.; Galin, F.Z.; Spirikhin, L.V. Transformation of 19 β ,28-epoxy-18 α -olean-2-ene by the Prins reaction. *Chem. Nat. Comp.* **2006**, *42*, 740-741.
49. Sejbál, J.; Klinot, J.; Protiva, J.; Vystrčil, A. Triterpenes. Part. LXXIII. Reactions of triterpenoid ketones with sulfur and morpholine under Willgerodt-Kindler reaction conditions. *Collect. Czech. Chem. Commun.* **1986**, *51*, 118-127.
50. Barton, D.H.R.; Head, A.J.; May, P.J. Long-range effects in alicyclic systems. II. Rates of condensation of some triterpenoid ketones with benzaldehyde. *J. Chem. Soc.* **1957**, 935-944.

51. Kim, H.O.; Tolstikov, G.A.; Bazalitskaya, V.S. Triterpenoids. XXI. Condensation of 3-oxotriterpenes with esters and synthesis of C-substituted triterpenoid azoles. *Zh. Obsh. Khim.* **1970**, *40*, 492-497.
52. Klinot, J.; Světlý, J.; Kudláčková, D.; Buděšinský, M.; Vystrčil, A. Triterpenes. 59 Preparation of 2-methyl-3-oxotriterpenoids of 18- α -oleanane series and the conformation of ring A. *Collect. Czech. Chem. Commun.* **1979**, *44*, 211-225.
53. Tolmacheva, I.A.; Anikina, L.V.; Vikharev, Y.B.; Shelepen'kina, L.N.; Grishko, V.V.; Tolstikov, G.A. Synthesis and immunotropic activity of 2-alkylaminomethylene-19 β ,28-epoxyolean-3-ones. *Russ. J. Bioorg. Chem.* **2008**, *34*, 125-129.
54. Biedermann, D.; Sarek, J.; Klinot, J.; Hajduch, M.; Dzubak, P. Fluorination of betulinines and other triterpenoids with DAST. *Synthesis* **2005**, 1157-1163.
55. Klinot, J.; Vystrčil, A. Conformation of epimeric 2-bromoallobetulones. *Chem. Ind.* **1960**, 1360-1361.
56. Hanna, R.; Ourisson, G. Studies of cyclic ketones. VIII. Preparation and properties of polycyclic α -diketones. *Bull. Soc. Chim. Fr.* **1961**, 1945-1951.
57. Green, G.F.H.; Long, A.G. Compounds related to the steroid hormones. II. Action of hydrogen bromide on 2-bromo-3-oxo- Δ 1-5 α -steroids. *J. Chem. Soc.* **1961**, 2532-2543.
58. Lehn, J.M.; Ourisson, G. Cyclic ketones. XIV. Conformation of 2-bromo- and 2,2-dibromo-3-oxotriterpenes. *Bull. Soc. Chim. Fr.* **1963**, 1113-21.
59. Klinot, J.; Vystrčil, A. Triterpenes. 7. Stereochemistry of 2-bromo derivatives of allobetulin and alloheterobetulin. *Collect. Czech. Chem. Commun.* **1966**, *31*, 1079-1091.
60. Hajduch, M.; Sarek, J. Preparation of betulin type triterpenoid derivatives as antitumor agents. *PCT Int. Appl. WO 2001090096*, 2001, [CA 2001, 136, 6180].
61. Klinot, J.; Richt, V.; Vystrčil, A. Triterpenes. XLIII. Conformation of ring A of 2,3-disubstituted triterpenes with chlorine or an alkoxy group in position 2. *Collect. Czech. Chem. Commun.* **1975**, *40*, 1758-1767.
62. Novotny, J.; Podlaha, J.; Klinot, J. Triterpenes. CII. Crystal structure and conformation of ring A of 2 α -bromo-19 β ,28-epoxy-18 α -oleanan-3-one. *Collect. Czech. Chem. Commun.* **1993**, *58*, 2737-2744.
63. Tolstikova, L.F.; Tolstikov, G.A. Triterpenoids. XXIII. Synthesis and reactions of acetylenic derivatives of triterpenes. *Izv. Akad. Nauk Kazakh. SSR, Ser. Khim.* **1971**, *21*, 65-71.
64. Kvasnica, M.; Rudovska, I.; Cisarova, I.; Sarek, J. Reaction of lupane and oleanane triterpenoids with Lawesson's reagent. *Tetrahedron* **2008**, *64*, 3736-3743.
65. Sejbál, J.; Klinot, J.; Vystrčil, A. Triterpenes. Part LXXXIII. Reaction of 3-keto and 2-ketotriterpenoids with 3-chloroperoxybenzoic acid in aliphatic alcohols. A new method of preparation of α -hydroxy ketones. *Collect. Czech. Chem. Commun.* **1987**, *52*, 1052-1061.
66. Tolmacheva, Shelepen'kina, L.N.; Shashkov, A.S.; Grishko, V.V.; Glushkov, V.A.; Tolstikov, G.A. Reaction of 3-acetoxy-(2,3),(19 β ,28)diepoxyoleanane with cyclic and linear amines. *Chem. Nat. Comp.* **2007**, *43*, 153-158.
67. Sejbál, J.; Klinot, J.; Vystrčil, A. Triterpenes. LXXXV. Photolysis of 19 β , 28-epoxy-18 α -oleanan-2 β -ol nitrites: functionalization of 10 β - and 8 β -methyl groups. *Collect. Czech. Chem. Commun.* **1988**, *53*, 118-131.

68. Sejbal, J.; Klinot, J.; Budesinsky, M. Triterpenes. XCIV. Photolyses and pyrolyses of triterpenoid nitrites. *Collect. Czech. Chem. Commun.* **1991**, *56*, 1732-1743.
69. Sejbal, J.; Klinot, J.; Budesinsky, M. Triterpenes. Part CV. Oleanane triterpenoids functionalized at C-25 and C-26. *Collect. Czech. Chem. Commun.* **1996**, *61*, 1360-1370.
70. Flekhter, O.B.; Medvedeva, N.I.; Karachurina, L.T.; Baltina, L.A.; Zarudi, F.S.; Galin, F.Z.; Kabal'nova, N.N.; Tolstikov, G.A. Synthesis and anti-inflammatory activity of new acylated betulin derivatives. *Pharm. Chem. J.* **2002**, *36*, 488-491.
71. Krasutsky, P.A.; Carlson, R.M. Preparation of triterpene derivatives having fungicidal activity against yeast. *PCT Int. Appl. WO 2002026761*, 2002, [CA 2002, 136, 294955].
72. Krasutsky, P.A.; Avilov, D.V. Preparation of triterpene quaternary salts having antibacterial, antifungal, and surfactant properties. *PCT Int. Appl. WO 2003062260*, 2003, [CA 2003, 139, 133696].
73. Nekrasov, D.D.; Obukhova, A.S. Benzoylacetylation of allobetulin and allobetulone oxime by benzoylketene generated *in situ* during thermolysis of 5-phenyl-2,3-dihydrofuran-2,3-dione. *Chem. Het. Comp.* **2005**, *41*, 1426-1427.
74. Odinokova, L.E.; Denisenko, M.V.; Denisenko, V.A.; Uvarova, N.I. Glycosylation of triterpene alcohols of the lupane series. *Khim. Prirod. Soedin.* **1988**, 212-217.
75. Baltina, L.A.; Flekhter, O.B.; Vasil'eva, E.V.; Tolstikov, G.A. Stereoselective synthesis of 2-deoxy- α -D-arabino-hexopyranosides of triterpenic alcohols. *Izv. Akad. Nauk, Ser. Khim.* **1996**, 2340-2346.
76. Baltina, L.A.; Flekhter, O.B.; Vasiljjeva, E.V. Stereoselective synthesis of triterpene 3-O-2-deoxy- α -glycosides. *Mendeleev Commun.* **1996**, 63-64.
77. Baltina, L.A.; Flekhter, O.B.; Vasil'eva, E.V.; Davydova, V.A.; Ismagilova, A.F.; Zarudii, F.S.; Tolstikov, G.A. The synthesis of triterpene 2-deoxy- α -D-hexopyranosides from glycals and their effect on reparative processes. *Bioorg. Khim.* **1997**, *23*, 512-518.
78. Flekhter, O.B.; Medvedeva, N.I.; Tret'yakova, E.V.; Galin, F.Z.; Tolstikov, G.A. Synthesis of methyl esters of betulinic acid 2-deoxy- α -glycosides and 28-oxo-19,28-epoxyoleanane. *Chem. Nat. Comp.* **2006**, *42*, 706-709.
79. Eleutério, M.I. P.; Schimmel, J.; Ritter, G.; Costa, M.D.; Schmidt, R.R. Synthesis of saponins with allobetulin and glycyrrhetic acid as aglycones. *Eur. J. Org. Chem.* **2006**, 5293-5304.
80. Gauthier, C.; Legault, J.; Piochon, M.; Lavoie, S.; Tremblay, S.; Pichette, A. Synthesis, cytotoxicity and haemolytic activity of chacotrioside lupane-type neosaponins and their germanicane-type rearrangement products. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2310-2314.
81. Pichette, A.; Legault, J.; Gauthier, C. Synthesis of triterpene derivatives as antitumor or anti-inflammatory agents. *Can. Pat. Appl. CA 2586614*, 2008, [CA 2008, 148, 538411].
82. Klinot, J.; Waisser, K.; Streinz, L.; Vystrčil, A. Triterpenes. XX. Participation of dimethylformamide in the addition of halogens to the Δ^2 -double bond-a route for the preparation of β -epoxides. *Collect. Czech. Chem. Commun.* **1970**, *35*, 3610-3617.
83. Klinot, J.; Krumpolc, M.; Vystrčil, A. Triterpenes. IX. Reaction of isomeric 2,3-epoxides with a Grignard reagent. *Collect. Czech. Chem. Commun.* **1966**, *31*, 3174-3181.
84. Tolstikov, G.A.; Kim, H.-O.; Goryaev, M.I. Synthesis of triterpenoid indoles. *Zh. Obsh. Khim.* **1967**, *37*, 1690.

85. Kim, H.-O.; Tolstikov, G.A.; Shumov, I.P.; Nasonova, A.M. Triterpenoids. XX. Synthesis of triterpenoid indoles. *Zh. Org. Khim.* **1969**, *5*, 1987-1991.
86. Urban, M.; Sarek, J.; Kvasnica, M.; Tislerova, I.; Hajduch, M. Triterpenoid Pyrazines and Benzopyrazines with Cytotoxic Activity. *J. Nat. Prod.* **2007**, *70*, 526-532.
87. Kim, H.-O.; Tolstikov, G.A.; Goryaev, M.I. Triterpenoids. XXII. Heterocyclic derivatives of glycyrrhetic acid. *Izv. Akad. Nauk Kazakh. SSR Ser. Khim.* **1969**, *19*, 46-48.
88. Kim, H.-O.; Tolstikov, G.A.; Goryaev, M.I. Triterpenoids. XXIV. Isomerism of triterpene azoles. *Izv. Akad. Nauk Kazakh. SSR Ser. Khim.* **1970**, *20*, 49-54.
89. Pettit, G.R.; Green, B.W.; Bowyer, W.J. Steroids and related natural products. VI. The structure of α -apoallobetulin. *J. Org. Chem.* **1961**, *26*, 2879-2883.
90. Dischendorfer, O.; Crillmayer, H. Phytochemistry. III. Betulin. 3. *Monatsh. Chem.* **1926**, *47*, 419-425.
91. Bradbury, B.J.; Huang, M. Substituted taraxastanes useful for treating viral infections. *U.S. Pat. Appl. US 20070197646*, 2007, [CA 2007, 147, 269186].
92. Vystrčil, A.; Klinot, J. Structure of alloheterobetulin. *Collect. Czech. Chem. Commun.* **1959**, *24*, 3273-3278
93. Kazakova, O.B.; Giniyatullina, G.V.; Yamansarov, E.Y.; Tolstikov, G.A. Betulin and ursolic acid synthetic derivatives as inhibitors of Papilloma virus. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 4088-4090.
94. Hase, T. Exhaustive Baeyer-Villiger oxidation of the allobetulone triterpenoid. *J. Chem. Soc. Chem. Commun.* **1972**, 755-756.
95. Sejbál, J.; Klinot, J.; Hrnčířová, D.; Vystrčil, A. Triterpenes. Part LXXI. Oxidation of 19 β , 28-epoxy-18 α -oleanan-3-one and -1-one with peracids. *Collect. Czech. Chem. Commun.* **1985**, *50*, 2753-2759.
96. Klinot, J.; Vystrčil, A. Beckmann rearrangement of triterpene 3-ketoximes. *Collect. Czech. Chem. Commun.* **1962**, *27*, 377-386.
97. Hase, T. Dehydration of triterpenoid ring A ϵ -lactams. *Acta Chem. Scand.* **1970**, *24*, 364-365.
98. Rao, K.L.; Ramraj, S.K.; Sundararamaiah, T. Aza triterpenes. II. A-Aza triterpenes of the lactones of methyl oleanonate and methyl betulonate. *J. Ind. Chem. Soc.* **1980**, *57*, 833-834.
99. Tolmacheva, I.A.; Galaiko, N.V.; Grishko, V.V. Synthesis of acylhydrazones from lupane and 19 β ,28-epoxy-18 α -oleanane 2,3-seco-aldehydonitriles. *Chem. Nat. Comp.* **2010**, *46*, 39-43.
100. Tolmacheva, I.A.; Igosheva, E.V.; Grishko, V.V.; Zhukova, O.S.; Gerasimova, G.K. The synthesis of triterpenic amides on the basis of 2,3-seco-1-cyano-19 β ,28-epoxy-18 α -olean-3-oic acid. *Russ. J. Bioorg. Chem.* **2010**, *36*, 377-382.
101. Klinot, J.; Rozen, J.; Klinotova, E.; Vystrčil, A. Triterpenes. Part LXXXII. A-nor-derivatives of 19 β ,28-epoxy-18 α -oleanane: preparation and stereochemistry. *Collect. Czech. Chem. Commun.* **1987**, *52*, 493-500.
102. Dischendorfer, O.; Polak, O. Phytochemistry. V. Allobetulin. *Monatsh. Chem.* **1929**, *51*, 43-58.
103. Ruzicka, L.; Isler, O. Polyterpenes, and polyterpenoids. CVI. Oxidation of dihydrobetulinol and dihydrobetulonic acid with nitric acid. *Helv. Chim. Acta* **1936**, *19*, 506-519.

104. Shernyukov, A.V.; Mainagashev, I.Y.; Korchagina, D.V.; Gatilov, Y.V.; Salakhutdinov, N.F.; Tolstikov, G.A. Spirocyclization of 2,3-seco-19 β ,28-epoxy-28-oxo-18 α -olean-2,3-dicarboxylic anhydride with benzylamines. *Dokl. Chem.* **2009**, *429*, 286-289.
105. Kazakova, O.B.; Khusnutdinova, E.F.; Kukovinets, O.S.; Zvereva, T.I.; Tolstikov, G.A. Effective synthesis of 2,3-seco-2,3-dicarboxyplatanic acid. *Chem. Nat. Comp.* **2010**, *46*, 393-396.
106. Ruzicka, L.; Govaert, F.; Goldberg, M.W.; Lambertson, A.H. Polyterpenes and polyterpenoids. CXXIII. Degradation of allobetulin and hydroxymethyleneallobetulinone with chromium trioxide. *Helv. Chim. Acta* **1938**, *21*, 73-83.
107. Sejbal, J.; Homolova, M.; Tislerova, I.; Krecek, V. Preparation and conformational analysis of 1,2-seco derivatives of 19 β ,28-epoxy-18 α -oleanane. *Collect. Czech. Chem. Commun.* **2000**, *65*, 1339-1356.
108. Thibeault, D.; Legault, J.; Bouchard, J.; Pichette, A. Useful approach to access germanicanes from betulin. *Tetrahedron Lett.* **2007**, *48*, 8416-8419.
109. Flekhter, O.B.; Medvedeva, N.I.; Tolstikov, G.A.; Galin, F.Z.; Yunusov, M.S.; Mai, H.N.T.; Tien, L.V.; Savinova, I.V.; Boreko, E.I.; Titov, L.P.; Glukhov, I.V. Synthesis of olean-18-(19)-ene derivatives from betulin. *Russ. J. Bioorg. Chem.* **2009**, *35*, 233-239.
110. Kazakova, O.B.; Tolstikov, G.A.; Suponitskii, K.Y. A one-step approach to the synthesis of germanicane triterpenoids from allobetulin. *Russ. J. Bioorg. Chem.* **2010**, *36*, 133-135.
111. Platanov, V.G.; Zorina, A.D.; Gordon, M.A.; Chizhov, N.P.; Balykina, L.V.; Mikhailov, Y.D.; Ivanen, D.R.; Kvi, T.K.; Shavva, A.G. Triterpenoid antiviral activity against influenza A and B viruses. *Pharm. Chem. J.* **1995**, *29*, 42-46.
112. Flekhter, O.B.; Nigmatullina, L.R.; Baltina, L.A.; Karachurina, L.T.; Galin, F.Z.; Zarudii, F.S.; Tolstikov, G.A.; Boreko, E.I.; Pavlova, N.I.; Nikolaeva, S.N.; Savinova, O.V. Synthesis of betulinic acid from betulin extract and study of the antiviral and antiulcer activity of some related terpenoids. *Pharm. Chem. J.* **2002**, *36*, 484-487.
113. Anikina, L.V.; Tolmacheva, I.A.; Vikharev, Y.B.; Grisko, V.V. Effects of lupane and oleanane β -enamino ketones on the number and morphology of white blood cells. *Pharm. Chem. J.* **2009**, *43*, 378-380.
114. Salin, O.; Alakurtti, S.; Pohjala, L.; Siiskonen, A.; Maass, V.; Maass, M.; Yli-Kauhaluoma, J.; Vuorela, P. Inhibitory effect of the natural product betulin and its derivatives against the intracellular bacterium *Chlamydia pneumoniae*. *Biochem. Pharmacol.* **2010**, *80*, 1141-1151.
115. Flekhter, O.B.; Medvedeva, N.I.; Karachurina, L.T.; Baltina, L.A.; Galin, F.Z.; Zarudii, F.S.; Tolstikov, G.A. Synthesis and Pharmacological Activity of Betulin, Betulinic Acid, and Allobetulin Esters. *Pharm. Chem. J.* **2005**, *39*, 401-404.

Sample Availability: Not available.