

Article

C₁₉-Norditerpenoid Alkaloids from *Aconitum szechenyianum* and Their Effects on LPS-Activated NO Production

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Abstract: Three new C₁₉-norditerpenoid alkaloids (1–3), along with two known C₁₉-norditerpenoid alkaloids (4–5) have been isolated from *Aconitum szechenyianum*. Their structures were established by extensive spectroscopic techniques and chemical methods as *szechenyianine* A (1), *szechenyianine* B (2), *szechenyianine* C (3), *N*-deethyl-3-acetylaconitine (4), and *N*-deethyldeoxyaconitine (5). Additionally, compounds 1–5 were tested for the inhibition of NO production on LPS-activated RAW264.7 cells with IC₅₀ values of 36.62 ± 6.86, 3.30 ± 0.11, 7.46 ± 0.89, 8.09 ± 1.31, and 11.73 ± 1.94 μM, respectively, while the positive control drug dexamethasone showed inhibitory activity with IC₅₀ value of 8.32 ± 1.45 μM. The structure-activity relationship of aconitine alkaloids were discussed.

Keywords: *Aconitum szechenyianum*; C₁₉-norditerpenoid alkaloids; anti-inflammatory activity; NO production; structure-activity relationship

1. Introduction

The plant *Aconitum szechenyianum* Gay., a species in the *Aconitum* genus of Ranunculaceae, is widely distributed in the west of China and used as a folk medicine in Shaanxi province, known as “Tie-Bang-Chui” [1]. Phytochemical studies revealed that *A. szechenyianum* contained mainly C₁₉ and C₂₀ diterpenoid alkaloids [2–5], possessing aconitine-type, 7,17-secoaconitine-type, and napeline-type skeletons. Aconitine-type have no oxygen-containing functionality at C-7, and secoaconitine-type skeleton contains N, C-17, and C-7, C-8 double bonds. Pharmacological studies revealed that these C₁₉ and C₂₀ diterpenoid alkaloids had demonstrated various activities as anti-inflammatory, analgesic, anticancer, anti-epileptiform, antiparasite, and cardiovascular action [6,7]. As part of our research project to explore more bioactive lead compounds from the medicinal herbs in the Qinba mountains of China [8–16], the chemical constituents and pharmacological studies of *A. szechenyianum* were studied, and three new C₁₉-norditerpenoid alkaloids, *szechenyianine* A (1), *szechenyianine* B (2), and *szechenyianine* C (3), along with two known ones, *N*-deethyl-3-acetylaconitine (4) and *N*-deethyldeoxyaconitine (5) were isolated (Figure 1). Since the roots of *A. szechenyianum* were commonly used to treat rheumatism and fracture [17], the isolated compounds were evaluated for their effects on the inhibition of NO production on LPS-activated RAW264.7 cells (Table 2 and Figure 5), and the structure-activity relationship of these compounds were discussed.

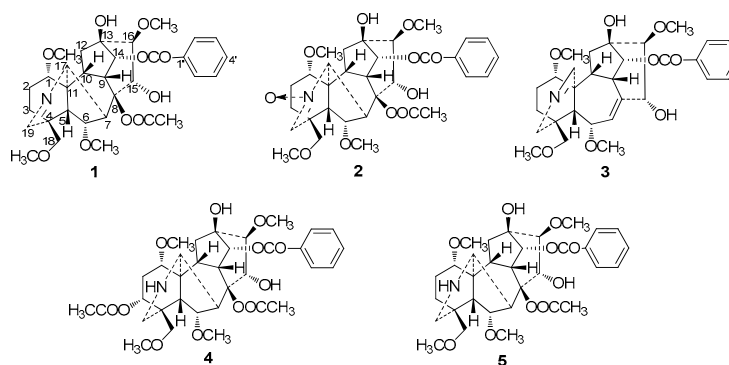


Figure 1. Structures of compounds 1–5.

2. Results and Discussion

Szechenyanine A (**1**) was isolated as a white amorphous powder and showed a positive reaction with Dragendorff's reagent. Its molecular formula $C_{32}H_{41}NO_{10}$ was derived from a protonated molecular ion peak at m/z 600.2842 $[M + H]^+$ (calcd. 600.2809) of the HR-ESI-MS spectrum. Comparison of the NMR data of **1** and **5**, indicated almost similar NMR spectroscopic features, except for the number of C-4, C-17, C-19, this deduction was also confirmed by the chemical shift (Table 1) of C-4 (δ_C 39.0), C-19 (δ_C 49.0), and C-17 (δ_C 56.7) to upfield in ^{13}C -NMR spectra of **5** compared with C-4 (δ_C 46.8), C-19 (δ_C 165.9) and C-17 (δ_C 60.6) of **1**, we predicted the existence of $N=CH$ group in compound **1**. The 1H -NMR spectrum (Table 1) of **1** showed the presence of five aromatic proton signals due to a monosubstituted benzene at δ_H 8.02 (2H, d, $J = 7.6$ Hz), 7.55 (1H, t, $J = 7.6$ Hz), and 7.43 (2H, t, $J = 7.6$ Hz); a methine proton of an $N = CH$ group at δ_H 7.31 (1H, s), four OMe protons at δ_H 3.75 (3H, s), 3.29 (3H, s), 3.18 (3H, s), and 3.03 (3H, s); and a strongly shielded proton of an acetoxy group at δ_H 1.32 (3H, s). The ^{13}C -NMR spectrum (Table 1) displayed 32 carbon resonances. Among them, resonances at δ_C 166.2, 133.6, 130.0, 129.8 ($C \times 2$), and 128.9 ($C \times 2$) were attributed to a benzoyloxy group; δ_C 61.3, 59.3, 57.4 and 56.3 were attributed to four OMe groups, δ_C 172.6 and 21.5 were attributed to an acetoxy group, and the NMR features of the remained 19 resonances were characteristic to an aconitine-type alkaloid, in which δ_C 165.9 was attributed to a $N=CH$ group and δ_C 74.3 and 78.9 were attributed to two oxygenated carbons associated with hydroxyl groups. The assignments of the NMR signals associated with **1** were derived from HSQC, HMBC, and ROESY experiments. In the HMBC spectrum (Figure 2), correlations of H-5 (δ_H 2.23) and H-17 (δ_H 3.97) to C-19 (δ_C 165.9) suggested that C-19 was involved in the $N=CH$ group; correlation of H-14 (δ_H 4.90) to the carbonyl carbon signal of benzoyl group (δ_C 166.2) suggested that the benzoyl group was located at C-14; correlation of the proton signal of the acetoxy group (δ_H 1.32) to C-8 (δ_C 90.6) suggested the acetoxy group was located at C-8; correlations of OCH_3 (δ_H 3.18) to C-1 (δ_C 82.3), OCH_3 (δ_H 3.03) to C-6 (δ_C 84.1), OCH_3 (δ_H 3.75) to C-16 (δ_C 89.9), and OCH_3 (δ_H 3.29) to C-18 (δ_C 78.2) suggested four methoxyl groups were linked at C-1, C-6, C-16, and C-18, respectively; correlations of H-12 (δ_H 2.20, 2.21), H-14 (δ_H 4.90), and H-16 (δ_H 3.42) to C-13 (δ_C 74.3), H-9 (δ_H 2.70) and H-16 (δ_H 3.42) to C-15 (δ_C 78.9), suggested two hydroxyl groups were linked at C-13 and C-15, respectively. Thus, the planar structure of **1** was deduced as 14-benzoyloxy-8-acetoxy-13,15-dihydroxy-1,6,16,18-tetramethoxy-19-en-aconitane. Meanwhile, in the ROESY spectrum (Figure 2) of **1**, the NOE correlations of H-1/H-10, H-10/H-14, H-14/H-9, and H-9/H-6 indicated β -orientation of H-1, H-6, H-9, H-10, and H-14, and α -axial configurations of 1- OCH_3 , 6- OCH_3 and 14-benzoyloxy; NOE correlations of H-6/H-5 and H-5/H-18 revealed β -orientation of H-18 and 18- OCH_3 , and α -axial of H-19; NOE correlations of H-17/H-7, H-16 and 15-OH, revealed α -axial of H-16, H-17 and 15-OH, and β -orientation of 16- OCH_3 , 13-OH and 8-acetoxy. Moreover, the NOE correlations of H-1/H-3 and H-5 while no correlation between H-2 and H-5 indicated **1** had ring A (C-1, C-2, C-3, C-4, C-5, and C-11) in the

chair conformation. Thus, according to the literature [18], compound **1** was assigned the name as (*A-c*)-14 α -benzoyloxy-8 β -acetoxyl-13 β ,15 α -dihydroxy-1 α ,6 α ,16 β ,18 β -tetramethoxy-19-en-aconitane.

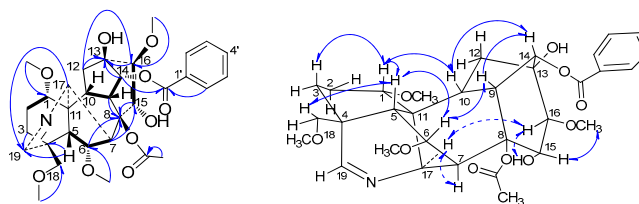


Figure 2. Key ^1H - ^1H COSY (H \leftrightarrow H), HMBC (H \rightarrow C) and ROESY (H \leftrightarrow H) correlations of compound **1**.

Table 1. ^1H -NMR and ^{13}C -NMR spectral data of compounds **1**–**5**.

NO.	1		2		3		4	5
	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{C}
1	82.3	3.20 (d, 4.1)	80.5	3.32 (m)	89.6	2.99 (dd, 4.4, 11.3)	81.0	82.3
2	22.9	1.66 (m, H-2a)	22.0	1.45 (m, H-2a)	24.7	1.10 (m)	31.8	23.5
		1.57 (m, H-2b)		1.81 (m, H-2b)		1.86 (m)		
3	28.2	1.63 (m, H-3a)	29.5	1.70 (m, H-3a)	37.4	1.55 (m)	72.3	29.0
		1.64 (m, H-3b)		1.79 (m, H-3b)		1.69 (m)		
4	46.8		42.1		39.7		43.2	39.0
5	45.8	2.23 (d, 7.1)	44.7	2.35 (d, 7.0)	46.2	2.32 (d, 8.9)	51.2	48.7
6	84.1	3.92 (d, 7.1)	82.7	4.00 (d, 7.0)	80.1	4.45 (m)	83.8	83.2
7	49.6	2.87 (s)	49.7	3.30 (s)	132.1	5.62 (d, 5.5)	45.2	43.6
8	90.6		89.3		137.5		91.7	91.5
9	42.6	2.70 (t, 6.1)	41.9	2.74 (m)	43.0	3.18 (s)	43.9	43.2
10	40.6	2.17 (m)	39.1	2.26 (m)	41.7	2.43 (s)	40.9	40.3
11	51.4		51.9		48.5		49.4	49.7
12	36.4	2.20 (m, H-12a)	36.5	2.27 (m, H-12a)	38.9	2.45 (m)	35.2	35.4
		2.21 (m, H-12b)		1.98 (m, H-12b)				
13	74.3		74.1		75.6		74.4	74.0
14	79.3	4.90 (d, 4.9)	78.8	4.89 (d, 4.8)	79.4	5.08 (d, 4.2)	79.1	78.9
15	78.9	4.48 (dd, 2.9, 5.3)	78.7	4.48 (dd, 3.0, 4.9)	74.1	4.80 (dd, 3.0, 5.8)	79.2	79.0
16	89.9	3.42 (d, 5.3)	89.6	3.45 (d, 5.0)	92.2	3.30 (d, 6.0)	90.0	89.5
17	60.6	3.97 (s)	72.9	4.02 (s)	166.4	7.86 (br s)	55.8	56.7
		3.78 (d, 8.5, H-18a)		3.79 (d, 8.5, H-18a)		3.16 (d, 8.4)		
18	78.2	3.42 (d, 8.5, H-18b)	77.9	3.33 (d, 8.5, H-18b)	80.6	3.86 (d, 8.4)	73.8	79.8
19	165.9	7.31 (s)	138.9	6.70 (d, 1.2)	58.3	3.53 (m)	41.6	49.0
8-OAc	172.6		172.1			3.45 (m)	172.3	172.0
	21.5	1.32 (s)	21.4	1.32 (s)			21.5	21.3
1-OCH ₃	56.3	3.18 (s)	56.6	3.21 (s)	58.2	3.20 (s)	56.0	55.4
6-OCH ₃	57.4	3.03 (s)	57.3	3.05 (s)	56.9	3.19 (s)	58.3	57.9
16-OCH ₃	61.3	3.75 (s)	61.4	3.77 (s)	61.8	3.75 (s)	61.4	61.1
18-OCH ₃	59.3	3.29 (s)	59.3	3.27 (s)	59.1	3.27 (s)	59.1	59.1
ArC=O	166.2		166.2		166.4		166.2	165.9
ArC-1'	130.0		129.9		130.0		130.0	130.7
3', 5'	128.9	7.43 (t, 7.6)	129.0	7.44 (t, 7.3)	128.7	7.42 (t, 7.5)	128.9	128.6
2', 6'	129.8	8.02 (d, 7.6)	129.9	8.01 (d, 7.3)	130.1	8.03 (d, 7.5)	129.8	129.6
4'	133.6	7.55 (t, 7.6)	133.8	7.57 (t, 7.3)	133.5	7.53 (t, 7.5)	133.5	133.3

δ in CDCl₃, in ppm from TMS; coupling constants (J) in Hz; ^1H -NMR at 400 MHz and ^{13}C -NMR at 100 MHz.

Szechenyanine B (**2**) was isolated as a white amorphous powder. The NMR spectroscopic data indicated that **2** was an analogue of **1** with similar skeleton and substituent groups. However, the molecular formula of **2** was deduced as C₃₂H₄₁NO₁₁ from the protonated molecular ion peak at m/z 616.2783 [M + H]⁺ (calcd. 616.2758), suggesting that an *N*-oxidation group was included in compound **2**. This deduction was also confirmed by the chemical shift (Table 1) of C-4 (δ_{C} 42.1) and C-19 (δ_{C} 138.9) to upfield, and C-17 (δ_{C} 72.9) to downfield in ^{13}C -NMR spectra of **2** compared with C-4 (δ_{C} 46.8), C-19 (δ_{C} 165.9) and C-17 (δ_{C} 60.6) of **1**. Thus, compound **2** was identified by HSQC, ^1H - ^1H COSY, HMBC, and ROESY experiments (Table 1 and Figure 3) as

(A-c)-14 α -benzoyloxy-8 β -acetoxyl-13 β ,15 α -dihydroxy-1 α ,6 α ,16 β ,18 β -tetra-methoxy-19-en-aconitane-N-oxide.

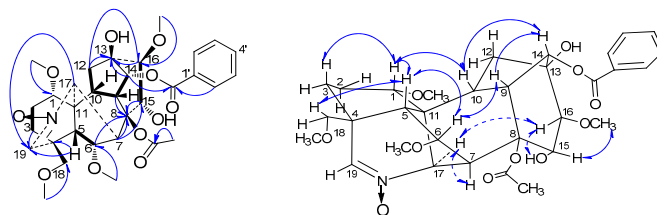


Figure 3. Key ^1H - ^1H COSY (H—H), HMBC (H→C) and ROESY (H↔H) correlations of compound 2.

Szechenyanine C (**3**) was isolated as a white amorphous powder. Its molecular formula $\text{C}_{30}\text{H}_{39}\text{NO}_8$ was derived from a protonated molecular ion peak at m/z 542.2783 $[\text{M} + \text{H}]^+$ (calcd. 542.2754) of the HR-ESI-MS spectrum. The ^1H -NMR spectrum (Table 1) of **3** showed the presence of five aromatic protons signals due to a monosubstituted benzene at δ_{H} 8.03 (2H, d, $J = 7.5$ Hz), 7.53 (1H, t, $J = 7.5$ Hz), and 7.42 (2H, t, $J = 7.5$ Hz); two olefinic protons signals at δ_{H} 7.86 (1H, brs) due to $\text{N}=\text{CH}$ and δ_{H} 5.62 (1H, d, $J = 5.5$ Hz) due to $\text{C}=\text{CH}$, respectively; and four OMe protons at δ_{H} 3.75 (3H, s), 3.27 (3H, s), 3.20 (3H, s) and 3.19 (3H, s). The ^{13}C -NMR spectrum (Table 1) displayed 30 carbon resonances. Among them, resonances at δ_{C} 166.4, 133.5, 130.0, 130.1 ($\text{C} \times 2$) and 128.7 ($\text{C} \times 2$) were attributed to a benzoyl group; δ_{C} 61.8, 59.1, 58.2 and 56.9 were attributed to four OMe groups; and the NMR features of the remained 19 resonances were characteristic to a 7, 17-secoaconitine alkaloid, in which δ_{C} 166.4 was attributed to a $\text{N}=\text{CH}$ group, and δ_{C} 132.1 and 137.5 were attributed to an olefinic bond. In the HMBC spectrum (Figure 4), correlations of H-1 (δ_{H} 2.99), H-5 (δ_{H} 2.32), H-10 (δ_{H} 2.43), and H-19 (δ_{H} 3.53) to C-17 (δ_{C} 166.4) suggested that C-17 was involved in the $\text{N}=\text{CH}$ group, and correlations of H-5 (δ_{H} 2.32), H-6 (δ_{H} 4.45) to C-7 (δ_{C} 132.1), H-6 (δ_{H} 4.45), H-14 (δ_{H} 5.08), and H-15 (δ_{H} 4.80) to C-8 (δ_{C} 137.5) suggested the olefinic bond was located at C-7 and C-8, which supported the presence of skeleton of the 7,17-secoaconitine alkaloid. Moreover, HMBC correlation of H-14 (δ_{H} 5.08) to the carbonyl carbon signal of benzoyl group (δ_{C} 166.4) suggested that the benzoyl group was located at C-14; correlations of OCH_3 (δ_{H} 3.20) to C-1 (δ_{C} 89.6), OCH_3 (δ_{H} 3.19) to C-6 (δ_{C} 80.1), OCH_3 (δ_{H} 3.75) to C-16 (δ_{C} 92.2), and OCH_3 (δ_{H} 3.27) to C-18 (δ_{C} 80.6) suggested four methoxyl groups were linked at C-1, C-6, C-16 and C-18, respectively; correlations of H-10 (δ_{H} 2.43) and H-14 (δ_{H} 5.08) to C-13 (δ_{C} 75.6), H-7 (δ_{H} 5.62) and H-16 (δ_{H} 3.30) to C-15 (δ_{C} 74.1) suggested two hydroxyl group were linked at C-13 and C-15, respectively. Thus, the planar structure of **3** was deduced as 14-benzoyloxy-13,15-dihydroxy-1,6,16,18-tetramethoxy-7(8),17-dien-7,17-secoaconitane. Meanwhile, in the ROSEY spectrum (Figure 4) of **3**, the NOE correlations of H-1/H-10, H-10/H-14 and H-14/H-9 indicated β -orientation of H-1, H-9, H-10 and H-14, and α -axial configurations of 1-OCH₃ and 14-benzoyloxy; the NOE correlations of H-6/H-5 and H-5/H-18 revealed β -orientation of H-18 and 18-OCH₃, and α -axial of 6-OCH₃; NOE correlations of H-17/H-16 and 15-OH, H-15/16-OCH₃ revealed α -axial of H-16 and 15-OH, and β -orientation of 16-OCH₃ and 13-OH. Moreover, the NOE correlations of H-1/H-3 and H-5 while no correlation between H-2 and H-5 indicated **3** had ring A (C-1, C-2, C-3, C-4, C-5, and C-11) in the chair conformation. Thus, according to the literature [18], compound **3** was assigned the name as (A-c)-14 α -benzoyloxy-13 β ,15 α -dihydroxy-1 α ,6 α ,16 β ,18 β -tetramethoxy-7(8),17-dien-7,17-secoaconitane.

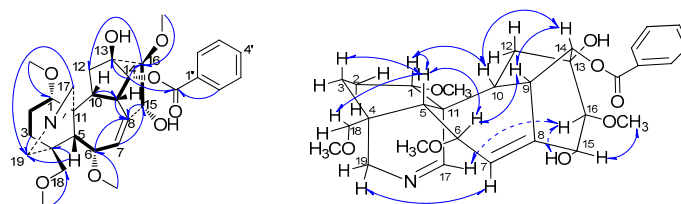


Figure 4. Key ^1H - ^1H COSY (H—H), HMBC (H→C) and ROESY (H↔H) correlations of compound 3.

Since the roots of *A. szechenyianum* are commonly used to treat rheumatism and fracture [17], in which inflammation is involved in the pathophysiological process and inhibitors of NO release are considered as potential anti-inflammatory agents for the treatment of these diseases [18–20], the isolated compounds from *A. szechenyianum* were evaluated using the Griess assay [21] for their effects on the inhibition of NO production in LPS-activated RAW264.7 cells. Dexamethasone (DEX) was selected as a positive control. As shown in Table 2 and Figure 5, all compounds with aconitine or 7,17-secoaconitine skeleton exhibited anti-inflammatory activities in a dose-dependent manner. Compared the activity with the substituent groups of 1, 2, 4, and 5, the structure-activity relationship may be due to the chemical environment of *N* atom. The compound 1 could hinder the inhibition of NO production with IC_{50} value of $36.62 \pm 6.86 \mu\text{M}$. The compound 2 exhibited excellent active performance with IC_{50} value of $3.30 \pm 0.11 \mu\text{M}$, indicated that the presence of $N \rightarrow O$ might increase anti-inflammatory activities. Moreover, compound 4 exhibited effective inhibitory activity with IC_{50} value of $8.09 \pm 1.31 \mu\text{M}$ and compound 5 showed inhibitory activity with IC_{50} value of $11.73 \pm 1.94 \mu\text{M}$. In addition, compound 3 as a 7,17-secoaconitine type alkaloid also exhibited potent inhibitory activity on NO production with IC_{50} value of $7.46 \pm 0.89 \mu\text{M}$.

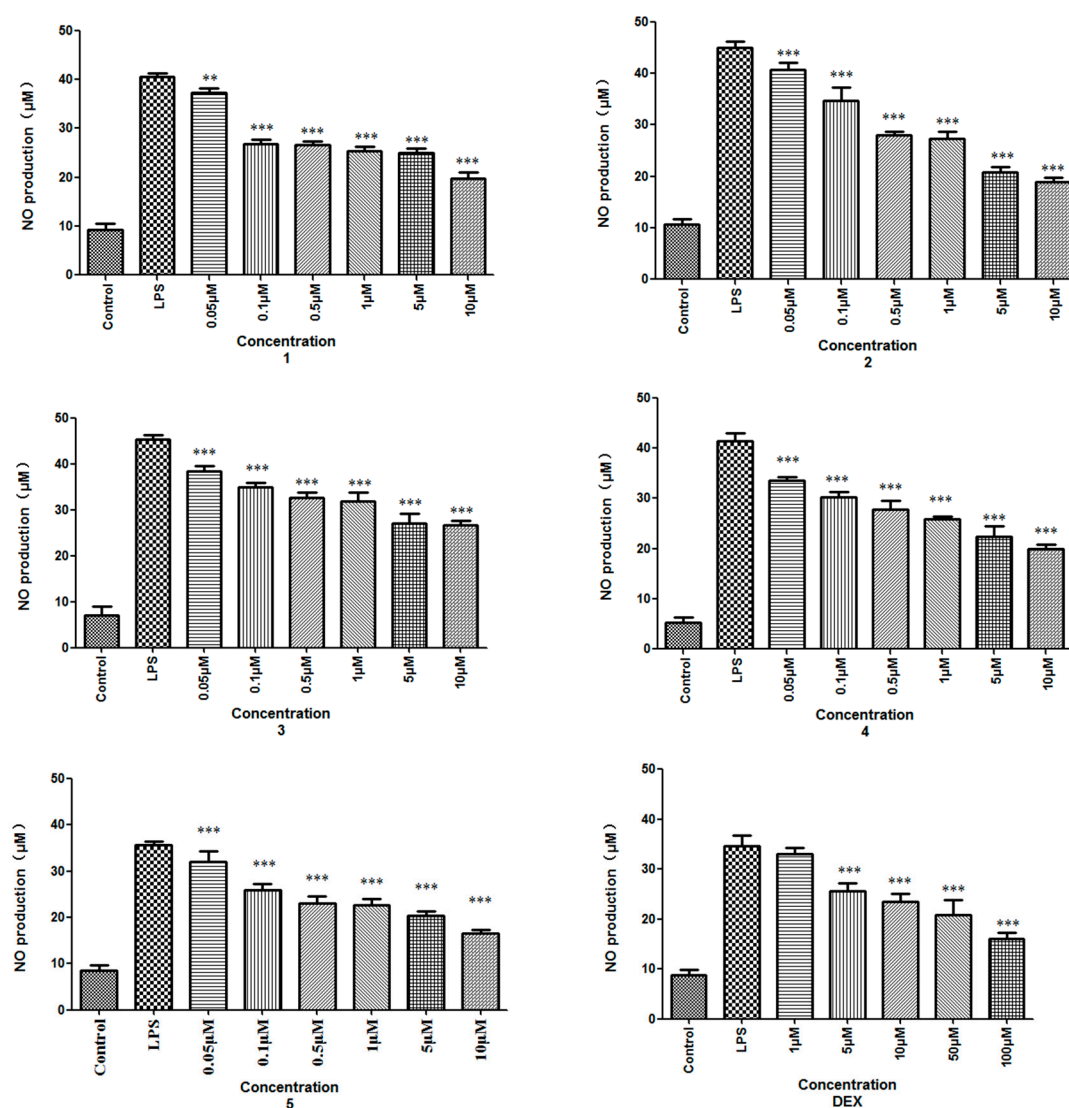


Figure 5. NO inhibitory effects of compounds from *A. szechenyianum* on LPS-activated RAW264.7 cells. Results represent the mean \pm SD of three independent experiments; results differ significantly from the LPS-treated, ** $p < 0.01$, *** $p < 0.001$; dexamethasone (DEX) was used as a positive control.

Table 2. IC₅₀ values of the compounds from *A. szechenyianum* on NO production in LPS-activated RAW264.7 cells.

Compound	1	2	3	4	5	Dexamethasone
IC ₅₀ (μM)	36.62 ± 6.86	3.30 ± 0.11	7.46 ± 0.89	8.09 ± 1.31	11.73 ± 1.94	8.32 ± 1.45

Results are expressed as IC₅₀ values in μM and the values are means ± SD; *n* = 3; dexamethasone was used as a positive control.

3. Experimental Section

3.1. General Information

ESI-MS was performed on a Quattro Premier instrument (Waters, Milford, MA, USA). The HR-ESI-MS spectra were recorded on an Agilent Technologies 6550 Q-TOF (Santa Clara, CA, USA). 1D and 2D-NMR spectra were recorded on Bruker-AVANCE 400 instrument (Bruker, Rheinstetten, Germany) with TMS as an internal standard. The analytical HPLC was performed on a Waters e2695 Separations Module coupled with a 2998 Photodiode Array Detector and a Accurasil C-18 column (4.6 mm × 250 mm, 5 μm particles, Ameritech, Chicago, IL, USA). Semipreparative HPLC was performed on a system comprising an LC-6AD pump equipped with an SPD-20A UV detector (Shimadzu, Kyoto, Japan) and an Ultimate XB-C18 (10 mm × 250 mm, 5 μm particles) or YMS-Pack-ODS-A (10 mm × 250 mm, 5 μm particles). Silica gel was purchased Qingdao Haiyang Chemical Group Corporation (Qingdao, China).

3.2. Plant Material

The roots of *Aconitum szechenyianum* Gay. were collected from the Xi Mountains of Gansu Province of China in July 2014, and identified by senior experimentalist Jitao Wang. A voucher specimen (herbarium No. 20140728) has been deposited in the Medicinal Plants Herbarium (MPH), Shaanxi University of Chinese Medicine, Xianyang, China.

3.3. Extraction and Isolation

The air-dried and powdered underground parts of *A. szechenyianum* (5.0 kg) were extracted with 80% EtOH at 80 °C for three times (each time 40 L for 1.5 h). After removal of EtOH solvent under reduced pressure, the extract (2 L) was dispersed in water (1.5 L), adjusted with 9% HCl solution to pH 0.8, and extracted with petroleum ether (PE). The acidic water solution was alkalinized to pH 10.26 with 25% ammonia solution, extracted with CHCl₃ three times, and evaporated under pressure to give crude alkaloids (50 g). The crude alkaloids (47 g) were chromatographed on silica gel column, eluting with gradient solvent system (PE/acetone/diethylamine, 50:1:0.1–1:1:0.1) to give 12 fractions (Fr.1–Fr.12). Fr.6 (3.2 g) was purified by HPLC (YMC-Pack-ODS-A, 10 mm × 250 mm, 5 μm particles, flow rate: 1.0 mL·min⁻¹) with CH₃OH/H₂O (70:30) as mobile phase to obtain Fr.6-1 (30 mg; *t*_R = 5 min), Fr.6-2 (120 mg; *t*_R = 30 min), Fr.6-3 (130 mg; *t*_R = 42 min), Fr.6-4 (120 mg; *t*_R = 63 min), Fr.6-5 (140 mg; *t*_R = 77 min), and Fr.6-6 (200 mg; *t*_R = 63 min). Fr.6-3 (130mg) was purified by HPLC with CH₃OH/H₂O (60:40) as mobile phase to afford **1** (13 mg; *t*_R = 50 min) and **2** (12 mg; *t*_R = 65 min). Fr.6-4 (120 mg) was purified by HPLC with CH₃OH/H₂O (65:35) as mobile phase to afford **3** (10 mg; *t*_R = 45 min), **4** (20 mg; *t*_R = 65 min), and **5** (25 mg; *t*_R = 75 min). See more detailed spectrums in the supplementary materials.

(*A-c*)-14α-Benzoyloxy-8β-acetoxyl-13β,15α-dihydroxy-1α,6α,16β,18β-tetramethoxy-19-en-aconitane (*szechenyianine* A): A white amorphous powder, $[\alpha]_D^{20} +60.2$ (*c* 0.38, MeOH), IR (KBr) ν_{\max} : 3508, 2936, 1718, 1637 and 714 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) and ¹³C-NMR (100 MHz, CDCl₃) spectral data, see Table 1; *m/z* 600.2842 [M + H]⁺ (calcd. for C₃₂H₄₁NO₁₀, 600.2809).

(A-c)-14 α -Benzyloxy-8 β -acetoxyl-13 β ,15 α -dihydroxy-1 α ,6 α ,16 β ,18 β -tetramethoxy-19-en-aconitane-N-oxide (szechenyanine B): A white amorphous powder, $[\alpha]_D^{20} +10.5$ (c 0.44, MeOH), IR (KBr) ν_{\max} : 3510, 2938, 1719, 1603 and 716 cm^{-1} ; $^1\text{H-NMR}$ (400 MHz, CDCl_3) and $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) spectral data, see Table 1; m/z 616.2783 $[\text{M} + \text{H}]^+$ (calcd. for $\text{C}_{32}\text{H}_{41}\text{NO}_{11}$, 616.2758).

(A-c)-14 α -Benzyloxy-13 β ,15 α -dihydroxy-1 α ,6 α ,16 β ,18 β -tetramethoxy-7(8),17-dien-7,17-secoaconitane (szechenyanine C): A white amorphous powder, $[\alpha]_D^{20} +21.6$ (c 0.64, MeOH), IR (KBr) ν_{\max} : 3513, 2930, 2824, 1716, 1645, 1453, 1367, 1275, 1102 and 715 cm^{-1} ; $^1\text{H-NMR}$ (400 MHz, CDCl_3) and $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) spectral data, see Table 1; m/z 542.2783 $[\text{M} + \text{H}]^+$ (calcd. for $\text{C}_{30}\text{H}_{39}\text{NO}_8$, 542.2754).

3.4. Inhibitory Assay of NO Production

Assays for NO production were carried out according to the Griess reaction, using dexamethasone as positive control. Briefly, RAW264.7 cells were seeded into 96-well microplates at a density of $2 \times 10^5 \text{ mL}^{-1}$ and allowed to adhere for 4 h. RPMI1640 (100 μL) containing test samples (final concentration of 10, 5, 1, 0.5, 0.1, and 0.05 μM) dissolved in DMSO (final concentration less than 0.2%) and LPS (final concentration of 1 $\mu\text{g}\cdot\text{mL}^{-1}$) were added. After incubation at 37 $^\circ\text{C}$ for 18 h, 50 μL of cell-free supernatant was mixed with 50 μL of Griess Reagent I and 50 μL of Griess Reagent II to determine NO production. Absorbance was measured at 550 nm against a calibration curve with NaNO_2 standard. The NO productions of the isolated compounds were tested (Figure 5), the inhibitory rate on NO production induced by LPS was calculated by the NO_2^- levels as follows: Inhibitory rate (%) = $100 \times ([\text{NO}_2^-]_{\text{LPS}} - [\text{NO}_2^-]_{\text{LPS+sample}}) / ([\text{NO}_2^-]_{\text{LPS}} - [\text{NO}_2^-]_{\text{untreated}})$, the IC_{50} values were calculated (Table 2). Values are mean \pm SD, $n = 3$, ** $p < 0.01$, *** $p < 0.001$ vs. LPS treated.

Supplementary Materials: IR, HR-ESI-MS, $^1\text{H-NMR}$, $^{13}\text{C-NMR}$ and 2D NMR spectra for compounds 1–3 can be found, in the online version, at <http://www.mdpi.com/1420-3049/21/9/1175/s1>.

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Sample Availability: Samples of the compounds 4–5 are available from the authors.



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