


A new era for stroke therapy: Integrating neurovascular protection with optimal reperfusion

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Abstract

Recent advances in stroke reperfusion therapies have led to remarkable improvement in clinical outcomes, but many patients remain severely disabled, due in part to the lack of effective neuroprotective strategies. In this review, we show that 95% of published preclinical studies on “neuroprotectants” (1990–2018) reported positive outcomes in animal models of ischemic stroke, while none translated to successful Phase III trials. There are many complex reasons for this failure in translational research, including that the majority of clinical trials did not test early delivery of neuroprotectants in combination with successful reperfusion. In contrast to the clinical trials, >80% of recent preclinical studies examined the neuroprotectant in animal models of transient ischemia with complete reperfusion. Furthermore, only a small fraction of preclinical studies included long-term functional assessments, aged animals of both genders, and models with stroke comorbidities. Recent clinical trials demonstrate that 70%–80% of patients treated with endovascular thrombectomy achieve successful reperfusion. These successes revive the opportunity to retest previously failed approaches, including cocktail drugs that target multiple injury phases and different cell types. It is our hope that neurovascular protectants can be retested in future stroke research studies with specific criteria outlined in this review to increase translational successes.

Keywords

Stroke, ischemia, reperfusion, neuroprotection, translational research

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Introduction

Acute ischemic stroke remains a leading cause of disability and mortality worldwide.¹ Following initially modest progress after the introduction of intravenous thrombolysis in 1995,^{2–5} dramatic gains in reperfusion therapies have recently been achieved with endovascular thrombectomy for the most severe strokes with large vessel occlusion (LVO).⁶ On the other hand, many stroke patients remain severely disabled. The rates of functional independence achieved even after highly effective reperfusion treatments are just under 50%,^{7–9} underscoring an unmet clinical need for adjunctive neuroprotective treatments. There has been a long history of failed clinical trials of neuroprotection in stroke.¹⁰ However, most clinical trials did not test the neuroprotective candidate in combination with successful reperfusion. Timely and successful restoration of blood flow

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may be essential to deliver therapeutic concentrations of neuroprotectants to the ischemic brain. Recognition of this key distinction unveils a new path for more fruitful partnerships between preclinical and clinical scientists. The primary goal of this review is to discuss the evidence-based rationale for retesting neuroprotectants in conjunction with reperfusion therapy in ischemic stroke patients. In this review, we hope to accomplish the following: (1) Highlight recent exciting advances in stroke treatments, (2) identify the remaining areas of paramount clinical need for neuroprotection in stroke, (3) synopsise our analysis of the preclinical literature on neuroprotective strategies, (4) identify mismatches between preclinical and clinical research that may have contributed to the failures of translation to date, (5) discuss the primary goals of stroke treatments and the biological mechanisms they must target, and (6) recommend future approaches for preclinical and clinical research, including combinatorial approaches to decelerate ischemic injury and extend the temporal window during which injured tissue can be salvaged.

Recent progress in stroke reperfusion therapies

Recent progress in stroke reperfusion therapies is largely attributed to the successful translation of key concepts about the pathophysiology of cerebral artery occlusion and ensuing tissue ischemia. Early laboratory and clinical studies demonstrated that acute LVO leads to an ischemic “core” consisting of irreversibly damaged, infarcted tissue, surrounded by a “penumbra” of functionally impaired, but potentially salvageable tissue.^{11–13} Following acute LVO, the ischemic core may expand into the penumbral area over time, but this process may be aborted if tissue reperfusion is achieved sufficiently early.^{14–16} The growth of the ischemic core progresses at varying rates across individuals, depending on collateral blood flow capacity and parenchymal ischemic tolerance.^{17–19} Thus, some individuals with poor collateral flow and low ischemic tolerance after LVO display a larger ischemic core soon after symptom onset. These individuals are known as “fast progressors,” who tend to suffer worse clinical outcomes, with the most severe symptoms, larger infarcts, and increased risk of malignant cerebral edema and hemorrhagic transformation.¹⁹ Further studies are needed to better understand the molecular and clinical predictors of this devastating stroke phenotype. Conversely, patients with superior collateral flow and ischemic tolerance tend to exhibit a smaller ischemic core up to several days after LVO stroke onset. These may be the “slow progressors,” who typically experience superior clinical outcomes in response to reperfusion therapy.^{13,19,20}

Seven randomized clinical trials^{8,9,21,25} of endovascular therapy have now established that LVO reperfusion significantly improved clinical outcomes in patients with severe symptoms and brain tissue that is salvageable within 24 h after stroke onset (Table 1). A clear trend across these clinical trials is that faster and more complete arterial reperfusion correlates with superior treatment efficacy. Fast progressors benefit the most from the earliest possible reperfusion, typically within less than 6 h of stroke onset. Slow progressors may also derive significant benefits from delayed reperfusion, as demonstrated in the recent DAWN and DEFUSE-3 trials.^{8,9} Importantly, the latter trials revealed that slow progressors who do not achieve reperfusion will ultimately experience poor functional outcomes.

Why does reperfusion therapy not improve stroke outcome in all patients?

Despite the successful modernization of reperfusion therapies as outlined in Table 1 and recognition of the importance of selecting patients with salvageable brain tissue rather than solely by fixed time-windows, there remains a large need to improve acute stroke treatments in multiple areas. Although overall rates of partial or complete arterial recanalization after early administration of intravenous tissue plasminogen activator (iv tPA) reach 33%,^{26,27} complete recanalization rates with iv tPA are much lower for occlusions of the intracranial internal carotid artery (ICA; 4%), proximal middle cerebral artery (MCA; 21%), and basilar artery (4%).²⁷ Moreover, a significant number of LVO patients remain severely disabled despite achieving successful reperfusion with the gold standard intervention of intravenous tPA combined with endovascular thrombectomy. In a 2015 meta-analysis of the first five successful trials of stroke endovascular therapy, 18.5 to 32.5% of those patients who achieved successful reperfusion within 3 to 8 h after symptom onset still experienced severe disability or death at 90 days.²⁸ In the DAWN and DEFUSE-3 trials of delayed reperfusion, 84% and 76% of respective endovascular patients achieved successful reperfusion, but only 49% and 45% of these patients experienced long-term functional independence. These data indicate that some patients are less responsive to initially successful reperfusion. In some cases, early re-stenosis or re-occlusion of the recanalized vessel may account for unfavorable long-term outcomes. In the DAWN and DEFUSE-3 trials, the rates of artery recanalization at 24 h were 77% and 78%, respectively, which were only slightly lower or similar to the rates of initial angiographic recanalization. Therefore, in the majority of patients, failure to achieve good outcomes despite successful reperfusion is

Table 1. Pivotal clinical trials in the development of modern stroke reperfusion therapies.

Study name	Study type	Target population	Size	Therapy	Time to intervention	Reperfusion (≥mTICI 2 b)	Clinical outcome (intervention)	Clinical Outcome (Control)	Statistical Significance
NINDS (rt-PA Stroke Study Group ²)	Double-blinded, multi-center RCT ^a	All stroke subtypes, age < 3 h onset	624 (1:1) ^a	IV tPA vs. placebo	Not recorded	Not recorded	MRS 0-1: 39% ^a sICH: 7% Mortality: 13%	MRS 0-1: 26% sICH: 1% Mortality: 16%	OR 1.7 (1.1-2.6) P < 0.001 ns
ECASS III (Hacke et al. ³)	Double-blinded, multi-center RCT	All stroke subtypes, age < 80, 3 to 4.5 h onset	821 (1:1)	IV tPA vs. placebo	Not recorded	Not recorded	MRS 0-1: 52.4% sICH: 2.4% Mortality: 7.7%	MRS 0-1: 45.2% sICH: 0.2% Mortality: 8.4%	OR 1.34 (1.02-1.76) P = 0.008 ns
PROACT II (Furlan et al. ⁴)	Multi-center PROBE design	MCA occlusion, < 6 onset	180 (2:1)	IA r-proUK + IV heparin vs. IV heparin	4.7 h (median, onset to randomization)	66% (IAT) ^b	MRS 0-2: 40% sICH: 10% Mortality: 25%	MRS 0-2: 25% sICH: 2% Mortality: 27%	OR 2.13 (1.02-4.42) P = 0.06 ns
IMS-3 (Broderick et al. ⁵)	Multi-center PROBE design	NIHSS ≥ 10 or LVO, < 6 h onset	656 (2:1)	EVT ^c + IV tPA vs. IV tPA	4.2 h (mean, onset to IAT)	38% ICA, 44% MCA	MRS 0-2: 41% sICH: 6.2% Mortality: 19%	MRS 0-2: 39% sICH: 5.9% Mortality: 21%	RD 1.5% (-6.1 - 9.1) ns ns
MR CLEAN (Berkhemer et al. ¹⁵)	Multi-center PROBE design	ICA, MCA, ACA occlusion, NIHSS > 2, < 6 h onset	500 (1:1)	EVT ± IV tPA vs. standard care ± IV tPA	4.3 h (median, onset to groin puncture)	58.7%	MRS 0-2: 33% sICH: 7.7% Mortality: 19%	MRS 0-2: 19% sICH: 6.4% Mortality: 18%	OR 2.16 (1.39-3.38) ns ns
ESCAPE (Goyal et al. ¹⁸)	Multi-center PROBE design	ICA or MCA occlusion, ASPECTS ≥ 6, < 12 h onset	315 (1:1)	EVT ± IV tPA vs. standard care ± IV tPA	4 h (median, onset to reperfusion)	72.4%	MRS 0-2: 53% sICH: 3.6% Mortality: 10%	MRS 0-2: 29% sICH: 2.7% Mortality: 19%	RR 1.8 (1.4-2.4) ns P = 0.04
SWIFT PRIME (Saver et al. ¹⁹)	Multi-center PROBE design	ICA or MCA occlusion, NIHSS > 8, ASPECTS ≥ 6, < 6 h onset	198 (1:1)	EVT w/ stent-retriever + IV tPA vs. IV tPA	4.1 h (median, onset to 1st deployment)	88%	MRS 0-2: 60% sICH: 0% Mortality: 9%	MRS 0-2: 35% sICH: 3% Mortality: 12%	RR 1.7 (1.23 - 2.3) ns ns
EXTEND IA (Campbell et al. ²⁰)	Multi-center PROBE design	ICA or MCA occlusion, CTP core < 70 ml, < 6 h onset	70 (1:1)	EVT w/ stent-retriever + IV tPA vs. IV tPA	4.1 h (median, onset to reperfusion)	86%	MRS 0-2: 71% sICH: 0% Mortality: 9%	MRS 0-2: 40% sICH: 6% Mortality: 20%	OR 4.2 (1.4-12) ns ns
REVASCAT (Jovin et al. ²¹)	Multi-center PROBE design	ICA or MCA occlusion, ASPECTS ≥ 7 and < 8 h onset	206 (1:1)	EVT w/ stent-retriever ± IV tPA vs. standard care ± IV tPA	5.9 h (median, onset to reperfusion)	66%	MRS 0-2: 44% sICH: 4.9% Mortality: 18%	MRS 0-2: 28% sICH: 1.9% Mortality: 15%	OR 2.1 (1.1-4) ns ns
DAWN (Nogueira et al. ⁴)	Multi-center PROBE adaptive-enrichment design	ICA or MCA occlusion, NIHSS ≥ 10, MRI or CTP core < 5 l cc, 6 to 24 h onset	206 (1:1)	EVT w/ stent-retriever vs. standard care	12.2 h (median, onset to randomization)	84%	MRS 0-2: 49% sICH: 6% Mortality: 19%	MRS 02: 13% sICH: 3% Mortality: 18%	RD 36% (24-47) ns ns

(continued)

Table 1. Continued

Study name	Study type	Target population	Size	Therapy	Time to intervention	Reperfusion (\geq mTICI 2b)	Clinical outcome (intervention)	Clinical Outcome (Control)	Statistical Significance
DEFUSE 3 (Albers et al. ⁵)	Multi-center PROBE design	ICA or MCA occlusion, NIHSS \geq 6, MRI or CTP core < 70 cc w/ perfusion mismatch, 6 to 16 h onset	182 (1:1)	EVT vs. standard care	10.9 h (median, onset to randomization)	76%	MRS 0-2: 45% sICH: 7% Mortality: 14%	MRS 0-2: 17% sICH: 4% Mortality: 26%	RR 2.7 (1.6 - 4.5) ns P = 0.05

mTICI: modified treatment in cerebral ischemia scale, grade 2b relates to greater than 50% reperfusion of the vascular territory; MRS: Modified Rankin scale is a measure of functional status ranging from 0-2 (no or mild symptoms) to 5 (severe disability) and 6 (death), reported at three months after stroke; sICH: symptomatic intracranial hemorrhage within 24-30 h; PROBE: prospective randomized open-label with blinded outcome evaluation; ASPECTS: Alberta stroke program early CT score is a 10-point scale that quantifies early ischemic core size in the MCA vascular territory (8-10, 6-7 and < 5 represent small, moderate and large core sizes, respectively).

OR (odds ratio), RR (relative risk) and RD (risk difference) are reported with 95% confidence intervals.

NS denotes no statistical significance.

^aReported OR for independent functional outcome is for part 2 cohort that included 333 patients.

^bReperfusion rates were reported with thrombolysis in myocardial infarction scale (TIMI) grades 2 and 3, which included partial or complete proximal MCA recanalization.

^cOne of four mechanical thrombectomy devices and/or intra-arterial thrombolysis was employed (less than 2% of EVT patients were treated with newer generation stent-retrievers).

more readily attributed to a large pre-intervention infarct and to a lesser extent, the eloquence of the affected brain area.²⁹ Another possible explanation is that some LVO patients experience relatively fast infarct growth due to collaterals failing between the initial time of brain imaging and subsequent vessel recanalization.¹⁹ Other factors, most notably age but also other co-morbidities, in-hospital complications, and rehabilitation quality may also influence the long-term efficacy of reperfusion therapies. Further studies are needed to characterize the mechanisms of ineffective arterial recanalization and suboptimal response to adequate reperfusion.

Among current stroke therapies, another need for improvement relates to hemorrhagic transformation and other deleterious effects of reperfusion after intravenous thrombolysis with or without endovascular thrombectomy.³⁰ Intravenous thrombolysis has been associated with an increased risk of symptomatic intracranial hemorrhage (sICH) ranging between 2 and 7% within the first 24 to 30 h after ischemic stroke onset.^{31,32} The risk of sICH or large parenchymal hematoma ranged from 4 to 5% in both the intervention and control groups in the HERMES meta-analysis, which included more than 1200 patients pooled from the landmark endovascular therapy trials in 2015.⁷ Furthermore, approximately 60 to 80% of LVO patients who developed early ICH after endovascular thrombectomy experienced severe disabilities based on the modified Rankin score (mRS 4 or 5) or mortality at 90 days in a large single-center retrospective study.³³ To maximize the benefits of reperfusion therapies, future neuroprotective therapies must protect the blood-brain barrier, reduce the risk of hemorrhagic transformation, and minimize other types of reperfusion injury after ischemic strokes.

How can stroke reperfusion therapies be further improved?

Although stroke reperfusion therapy is effective under ideal circumstances, its effectiveness in the real world remains limited because the majority of stroke patients lack timely access to intravenous thrombolysis, with or without endovascular therapy. This is due in large part to failure of early recognition, unknown duration of symptoms, or late arrival of stroke patients to the hospital in remote geographical locations. Early epidemiological studies demonstrated that less than 10% of patients diagnosed with acute ischemic stroke in US hospitals are treated with reperfusion therapies, mainly because of delayed presentation.³⁴ This number is likely to increase with the modernization of stroke healthcare systems, and as more patients with LVO are now eligible for endovascular therapy up to

24 h after symptom onset, based on the DAWN and DEFUSE-3 trials.³⁵ On the other hand, no more than 25% of all acute ischemic strokes in the general population are thought to be due to LVO.³⁶ Therefore, the majority of non-LVO patients with distal arterial branch occlusions or small vessel occlusion who could be eligible for intravenous thrombolysis remain excluded from reperfusion, strictly due to delayed clinical presentation beyond 4.5 h of symptom onset. This problem underscores the unmet demand for neuroprotective therapies to preserve penumbral brain tissue (with consequent delay in tissue infarction), prevent hemorrhagic transformation, and extend the therapeutic time window for reperfusion.

Why did earlier clinical trials of neuroprotection in stroke fail despite previous success in preclinical models?

The last two decades have witnessed a remarkable increase in the number, breadth, and depth of preclinical research studies on acute ischemic stroke.³⁷ Recent animal studies have identified a significant number of neuroprotectants that reduce acute ischemic brain injury in preclinical models.³⁸ Between 1990 and 1994, there were ~25 publications per year assessing the effects of neuroprotectants in animal models of acute ischemic stroke. This number increased three-fold (77 per year) between 2000 and 2004 and almost six-fold after 2010 (144 per year). Between 1990 and 1994, approximately 500 clinical trials had commenced on >200 neuroprotective candidates for ischemic stroke, and ~350 of these trials had been completed as of 28 February 2018 (data from www.clinicaltrials.gov). However, among the ~200 neuroprotective candidates tested in patients, only nine successfully transitioned to Phase III trials with the support of promising preclinical studies. An additional candidate, DP-B99, transitioned to Phase III trials without the support of published preclinical work.^{39–41} However, none of these completed Phase III trials demonstrated significantly positive treatment effects to support translation of neuroprotectants into standard clinical practice.^{11,13,19,20}

The neuroprotectant uric acid accounts for as much as two-thirds of total antioxidant capacity in plasma⁴² and is among the top 10 candidates from the above-mentioned Phase III stroke trials. Preclinical studies have demonstrated that uric acid can prevent glutamate-induced cell death *in vitro*⁴³ and improve functional outcomes in animal models of ischemic stroke by neutralizing reactive oxygen species.⁴⁴ Consistent with these preclinical data, the administration of uric acid decreased lipid peroxidation in a Phase II pilot study without eliciting significant adverse reactions.⁴⁵

However, the randomized, double-blinded, placebo-controlled Phase IIb/III multicenter URICO-ICTUS trial demonstrated that co-administration of uric acid with tissue plasminogen activator (tPA) was not superior to reperfusion (intravenous or intra-arterial) alone; the addition of uric acid failed to change primary outcomes, or the percentage of patients with modified Rankin scores of 0–1 during follow-up at 90 days.⁴⁶ However, a post hoc analysis conducted on patients undergoing thrombectomy in URICO-ICTUS revealed statistically significant differences in clinical outcomes favoring the uric acid arm.⁴⁷ A plethora of other preclinically validated neuroprotective candidates, including cerebrolysin,⁴⁸ magnesium sulfate,⁴⁹ citicoline,⁵⁰ human albumin,⁵¹ ginsenoside Rb1,⁵² and recombinant human erythropoietin (rhEPO)⁵³ all failed to provide meaningful clinical benefits, raising important questions about the low rate of translation of drug efficacy from experimental models to clinical trials.⁵⁴ Why has it been so difficult to achieve meaningful neuroprotective effects in clinical trials of stroke? This failure may be attributed to multiple factors, including a series of key mismatches between preclinical and clinical stroke research that are identified below.

Analysis of pre-clinical and clinical research mismatches

In order to identify the discrepancies between basic and clinical stroke research, we analyzed all the original preclinical studies on neuroprotectants in ischemic stroke that were indexed in PubMed and Web of Science databases between January 1, 1990 and February 28, 2018 and published in English. The MeSH search terms entered into PubMed were: “Neuroprotective Agents”[Majr] AND “Brain Ischemia”[Majr]. The search terms for the Web of Science database were: (TS=((((Brain OR cerebral OR intracranial OR cranial OR intracerebral) AND (ischemic OR ischemia OR infarct OR hypoxic OR hypoxia OR anoxic OR anoxia)) AND (neuroprotective OR neuroprotectant OR neuroprotect))) AND LANGUAGE: (English) AND DOCUMENT TYPES: (Article). These search strategies identified a total of 3943 publications in PubMed and 6749 publications in Web of Science. We excluded studies duplicated in both databases (3182 in number) and all records that did not directly report the testing of neuroprotectants (2928 in number). The remaining 4582 studies were screened to ensure compliance with remaining inclusion/exclusion criteria. For example, review articles, clinical studies, *in vitro* studies, studies focused on physiotherapy, hemorrhagic stroke, or global ischemic brain injuries were excluded. In total, 2489 publications were closely examined in the final

analyses (Figure 1). All of the included studies evaluated biological, chemical, and cell-based therapies in preclinical settings, and we identified a number of important inconsistencies between preclinical models and clinical protocols that we believe may have influenced the general failure to translate animal work. Here we list five major characteristics of the clinical population that are not faithfully recapitulated in most preclinical research design:

Mismatch I: Neuroprotectants have been tested in preclinical models of transient ischemia but not tested in combination with adequate reperfusion in stroke clinical trials

Our attention was first drawn to 10 therapies that had entered Phase III clinical trials and were initially

deemed neuroprotective based on preclinical studies. Further scrutiny of the preclinical work on these neuroprotectants revealed that most were tested in models of transient ischemia (Figure 2(a)). As experimental models of transient ischemia typically last for 1 h in mice or 1.5–2 h in rats, the likelihood of successful translation of the preclinical neuroprotectant might be greater in patients who are recanalized with tPa and EVT within these time frames. Preclinical models of permanent cerebral ischemia are much less frequently used, although the vast majority of stroke patients had not experienced significant reperfusion before the recent successes of EVT.⁵⁵ For example, uric acid was only tested and proven efficacious in transient injury models—all four preclinical studies adhering to our inclusion criteria consistently found a reduction in brain damage after transient focal ischemia.^{43,44,56,57} As mentioned earlier, the Phase III clinical trial, URICO-ICTUS, revealed that uric acid combined

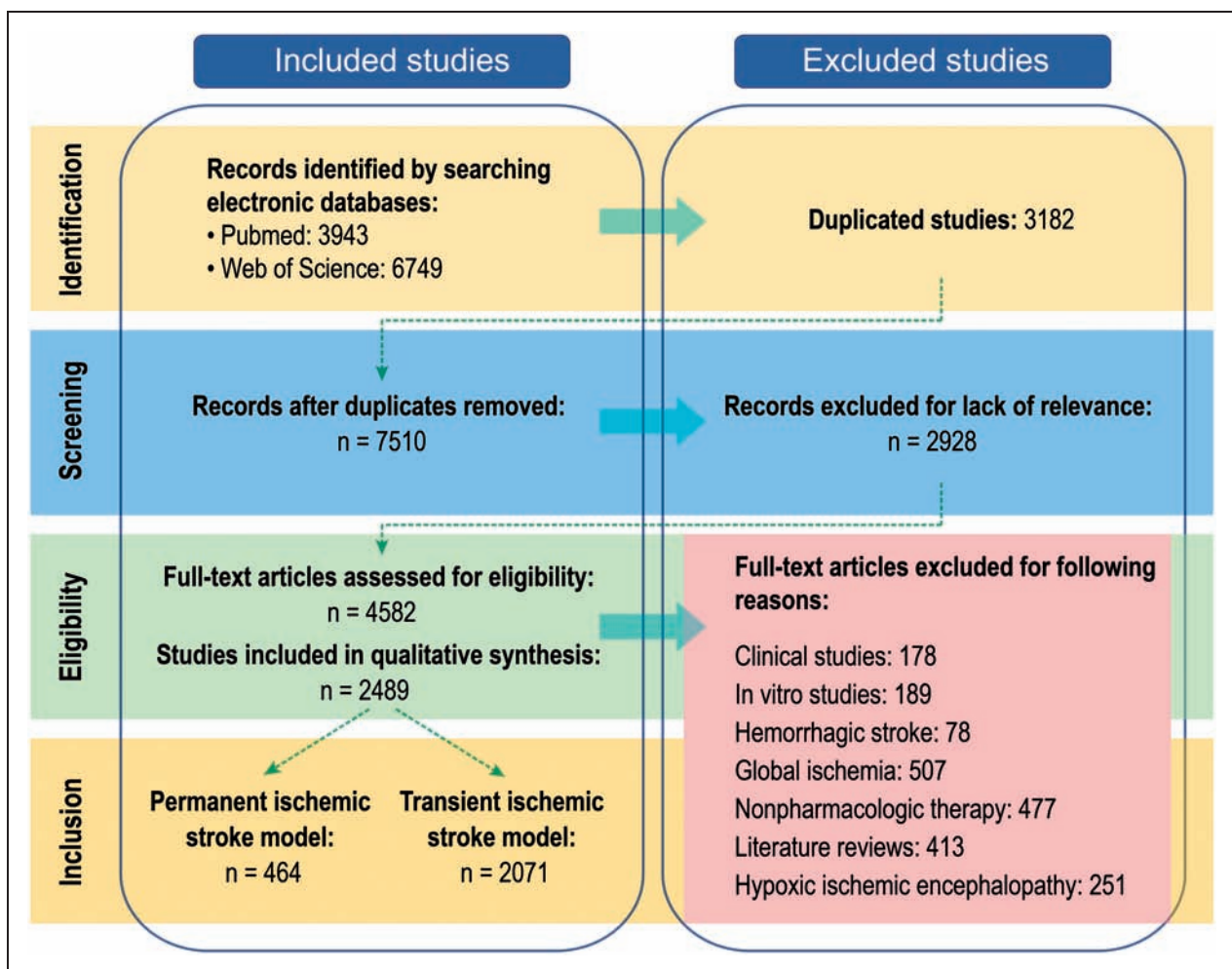


Figure 1. Flowchart of study selection. A total of 7510 studies on Pubmed and Web of Science were screened by evaluators. After exclusion for lack of relevance and various other criteria, 2489 studies were eligible for the qualitative synthesis.

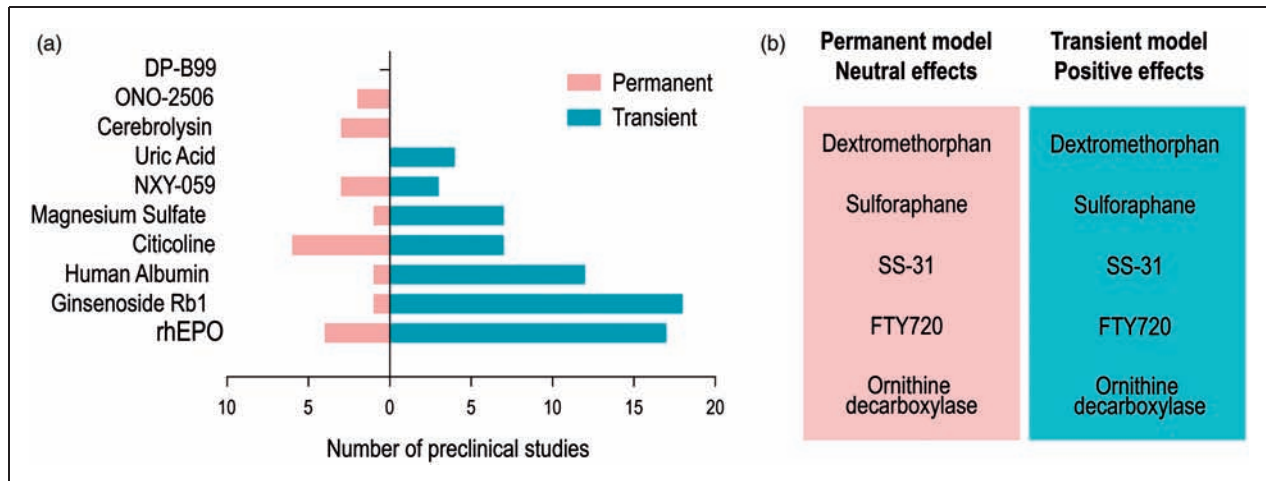


Figure 2. Neuroprotectants in preclinical models of transient and permanent ischemia. (a) Preclinical studies of ten neuroprotectants that entered Phase III clinical trials are divided into those employing permanent (pink) or transient (blue) ischemic insults. (b) Neuroprotectants with positive effects in transient models but neutral or negative effects in permanent models are listed.

with intravenous tPA within 4.5 h of stroke onset had no additional salutary effects on functional recovery at 90 days post-injury compared to tPA alone.⁴⁶ However, further analyses of the URICO-ICTUS trial suggested that uric acid was more effective than the placebo in reducing infarct volume in patients with early onset recanalization ($P=0.015$), but not in patients with delayed recanalization ($P=0.889$) or permanent occlusion ($P=0.233$).⁶⁰ These important observations highlight the importance of accounting for the temporal kinetics of ischemia in both preclinical and clinical work.

In addition to uric acid, magnesium sulfate,^{49,59,60} human albumin,^{51,61,62} and Ginsenoside Rb1^{63–65} were also tested in preclinical models of transient ischemia (Figure 2(a)), and also failed in Phase III trials. Indeed, more than 80% of preclinical studies involved transient ischemic stroke, in contrast to the clinical studies, in which only about one-third of patients exhibited revascularization.²⁷ This mismatch between preclinical experimental models and human subjects offers one likely explanation for the high rate of failure of neuroprotectants in clinical trials.

A series of preclinical rodent studies employing transient middle cerebral artery occlusion (MCAO) reported that human albumin reduced infarct volumes, ameliorated brain edema, and improved neurobehavioral function.^{57,61,66,68} Among numerous potential mechanisms, an improvement in collateral perfusion to the ischemic penumbra and regulation of the microcirculatory environment were identified.^{62,69} These preclinical results led to initiation of a Phase III randomized, double-blinded, placebo-controlled multicenter clinical trial, ALIAS, to examine the potential neuroprotective effects of human albumin on acute

ischemic stroke patients.⁷⁰ The ALIAS clinical trial included 841 patients across 89 sites in four countries and failed to report any significant difference between patients administered high-dose human albumin treatment versus placebo. Although 89% of the patients enrolled in ALIAS received either intravenous thrombolytic therapy with recombinant tissue plasminogen activator or underwent an endovascular procedure, the clinical report did not include recanalization rates. Administration of thrombolytic therapy is no guarantee of sufficient recanalization. Only 33% of patients with proximal LVO receiving intravenous thrombolysis typically undergo partial or complete early recanalization according to a meta-analysis of 26 studies encompassing 2063 stroke patients.²⁷ In other words, the majority of patients enrolled in clinical trials involving neuroprotectants do not undergo recanalization. This conclusion is further substantiated by the SAINT-II trial testing NXY-059 as a potential treatment for acute ischemic stroke, as only 44% of 3306 subjects received tPA in this trial.⁷¹ Given that only 33% of patients with proximal LVO who receive tPA achieve successful recanalization,²⁷ one might estimate that ~85% of subjects in the SAINT-II trial received NXY-059 without significant reperfusion. In contrast, almost all animals undergo complete recanalization once the suture is removed in experimental models of transient ischemia, revealing a mismatch in the degree of reperfusion achieved in preclinical versus clinical studies of neuroprotection in stroke.

Preclinical studies also support the notion that neuroprotection is reperfusion-dependent because the treatment effect size is typically smaller after permanent ischemia than transient ischemia.⁷² For example, dextromethorphan,^{73,74} sulforaphane,^{75,76} SS-31,⁷⁷

FTY720,^{78,79} and ornithine decarboxylase^{80,81} were significantly protective in transient stroke models but failed to elicit any protection against permanent cerebral ischemia (Figure 2(b)). Therefore, recent advances in EVT offer unprecedented opportunities to test neuroprotective therapies that were previously unsuccessful on non-reperfused ischemic tissue.⁵⁵ Restoration of blood flow with EVT is likely to improve the delivery of neuroprotectants into ischemic brain tissue.⁸² In previous clinical work, the lack of reperfusion may have acted as a bottleneck preventing effective delivery of neuroprotectants to cells in the ischemic penumbra that would be the most dynamically responsive to therapeutic intervention. In light of these arguments, we believe that some of the neuroprotectants that are effective in transient MCAO models may be worth reevaluating clinically in combination with modern EVT.

The efficacy of reperfusion after ischemic stroke is determined not only by recanalization of the occluded vessel, but also by the inherent properties of the collateral circulation. The collateral circulation in the vicinity of the necrotic infarct core may be recruited after vessel occlusion to supply missing nutrients and oxygen to the endangered surrounding tissue and exerts a profound influence over the evolution of the penumbral infarct zone.^{83,84} If there is insufficient collateral blood flow in stroke patients, the infarct tends to grow in size.⁸⁴ It is important to bear in mind that specific features of the collateral circulation in humans, such as number of collaterals, arterial diameter, the anatomical structure of the circle of Willis, and the presence of anterior communicating arteries are not always readily modeled in rodents.⁸⁵ Although collateral blood flow may be useful as a prognostic factor, various types of collateral blood flow measurements have been underemployed in preclinical work. A recent comprehensive analysis has demonstrated that 80% of the genetic variation of leptomeningeal collateral anatomy has been linked to a single nucleotide polymorphism of the *Rabep2* gene across numerous mouse strains.⁸⁶ Therefore, researchers should stratify animals according to natural variations in collateral perfusion to control against intragroup and strain variability.⁸³

Mismatch II: Treatment effect sizes are determined by infarct volumes in preclinical models but by long-term functional outcomes in clinical trials

Aside from the mismatch between transient ischemia models and the lack of reperfusion in clinical trials of neuroprotection in stroke, another concern is that 70%

of the preclinical studies relied on “infarct volume” as the single endpoint for the evaluation of therapeutic effects. In contrast, the effectiveness of a clinical treatment is mainly based on the 90-day modified Rankin score for long-term neurological function.^{41,87,88} A significant discrepancy between the severity of the clinical deficits and the infarct volumes is not uncommon in human subjects,⁸⁹ suggesting that overreliance on infarct volumes in preclinical studies should be avoided. The recent DAWN trial on thrombectomy revealed that acute stroke patients with a mismatch between clinical deficits and infarct volumes, or “clinical-core mismatch,” were excellent candidates for successful EVT irrespective of time of presentation within 24h after stroke onset.⁸ These clinical findings highlight the critical necessity of functional evaluations in preclinical studies, at both acute injury and chronic recovery/repair phases. Furthermore, the preclinical measurements of infarct volume are typically made at the histological level on postmortem tissue, which may be difficult to relate to the neuroimaging that is typically performed in clinical subjects.

Among the 2489 preclinical studies included in our analyses, only 819 evaluated neurological functional outcomes in stroke animals (Figure 3(a)). Even among those that reported neurological function, fewer than 30% of studies monitored functional recovery over extended periods (defined here as >14 days) (Figure 3(a)). Among those neuroprotectants that improved long-term functional recovery in animal studies, only 20 agents transitioned to Phase II or III trials. These agents include 17 beta-estradiol,⁹⁰ atorvastatin,⁹¹ cilostazol,⁹² erythropoietin,⁹³ ginsenoside Rd,⁹⁴ filgrastim,⁹⁵ hydrogen (Recruiting), melatonin (registered), minocycline,⁹⁶ NXY-059,⁹⁷ human albumin,⁹⁸ cerebrolysin,⁹⁹ citicoline,¹⁰⁰ propofol (Recruiting), JPI-289 (Recruiting), memantine (Recruiting), 3K3A-APC (NCT02222714, completed), FTY720 (Recruiting), YM872 (NCT00044057, NCT00044070, completed), and edaravone.¹⁰¹ Thus, more than 80% of the neuroprotectants that elicit improvements in long-term functional outcomes in preclinical studies have never been tested clinically. For example, preclinical evidence support the neuroprotective role of progesterone following acute CNS injuries, including traumatic brain injury and ischemic stroke.^{102–104} Progesterone significantly reduces infarct volumes and improves long-term functional outcomes in both permanent and transient models of ischemia.^{105,106} Although the therapeutic potential of progesterone following traumatic brain injury has been tested in two multi-center Phase III clinical trials,^{107,108} no clinical trials have tested the efficacy of progesterone as a treatment for ischemic stroke.

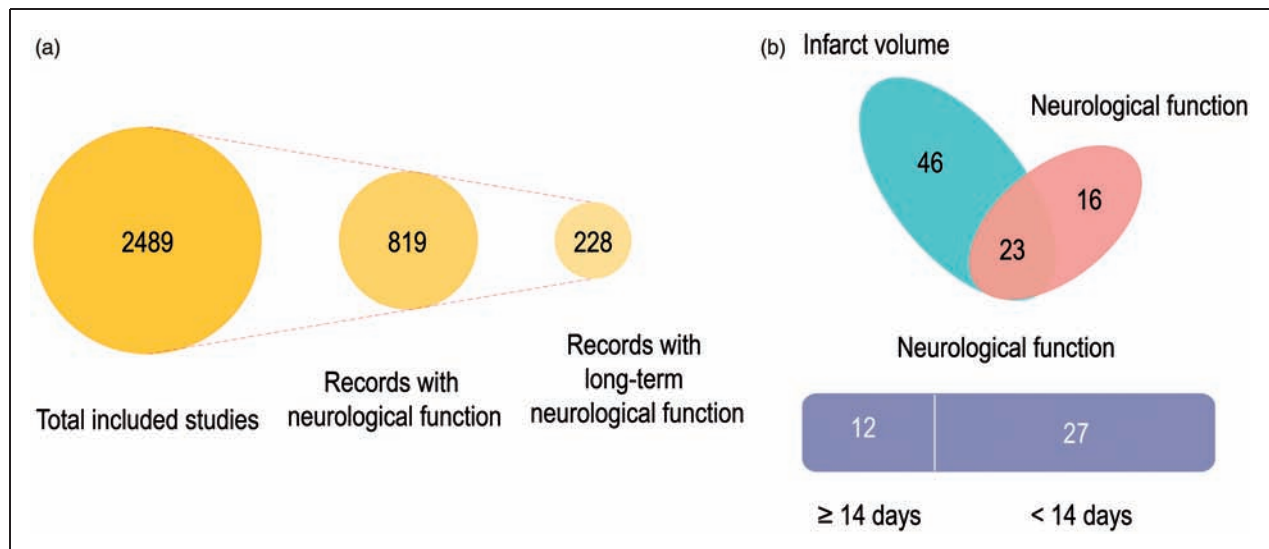


Figure 3. Neurological functional assessments in preclinical studies of neuroprotectants. (a) Only 32% of the eligible preclinical studies included measurements of neurological functional outcomes in stroke animals. Even among those that reported neurological function, fewer than 30% of studies monitored functional recovery over extended periods (defined here as > 14 days). (b) Among the 85 preclinical studies on the ten neuroprotectants that entered Phase III clinical trials, only 39 measured neurological function as an endpoint, and only 12 evaluated neurological functions longer than 14 days after stroke.

Of the 85 preclinical studies on the 10 neuroprotectants that entered Phase III clinical trials, only 39 measured neurological function as an endpoint, and only 12 evaluated neurological functions longer than 14 days after stroke (Figure 3(b)). For example, DP-B99 is a lipophilic chelator with a moderate affinity for zinc and was observed to exert beneficial effects against ischemic stroke in double-blinded placebo-controlled Phase II trials.⁴⁰ However, the Phase III Membrane Activator Chelator Stroke Intervention trial on 446 subjects with acute ischemic stroke reported that only 20.6% patients receiving DP-B99 displayed modified Rankin scores ≤ 1 , which was lower than placebo (28.8%).³⁹ To the best of our knowledge, there are no preclinical studies on the impact of DP-B99 in experimental models of acute ischemic stroke. In addition to DP-B99, the therapeutic candidates ONO-2506, cerebrolysin, and uric acid have also not received sufficient preclinical support. Citicoline has been tested in various ischemic models involving different species, but only one of these studies measured neurological functions longer than 14 days after stroke.¹⁰⁹ Similar concerns apply to some of the other neuroprotectants, including human albumin, magnesium sulfate, and NXY-059. Therefore, it seems worthwhile to focus future clinical tests on those neuroprotectants that achieve significant improvements in long-term functional recovery in preclinical studies conducted by multiple independent laboratories.

Mismatch III: Stroke is most prevalent in elderly men and women, whereas preclinical models mostly test young animals

The third concern raised by our analysis is that the majority of preclinical studies employ young adult animals, although it is well established that the risk for human stroke rises dramatically with age.¹¹⁰ As shown in Figure 4(a), out of more than 2000 treatments, only two (3K3A-activated protein C¹¹¹ and magnesium chloride¹¹²) have been evaluated in an aged rodent model of permanent ischemia, and nine have been assessed in aged rodent models of transient ischemia. The remarkable differences in stroke pathophysiology and outcomes between young and old animals have been known for many years.¹¹³ Aging is associated with a notable decline in the capacity of the brain to restore cellular and biochemical functions after ischemic stroke.^{57,113} Preclinical studies indicate that the mortality rate for animals subjected to transient middle cerebral artery occlusion is 6.3% for young adult rats and 43.5% for aged rats.¹¹⁴ Aging can result in greater oxidative DNA damage and impair neuroprotective responses in the peri-infarct cortex in the early stages after ischemic stroke.¹¹⁵ Our group previously observed that aged mice exhibited worse long-term learning and memory deficits after experimental stroke than young mice, and this age-related exacerbation of functional outcomes was associated with

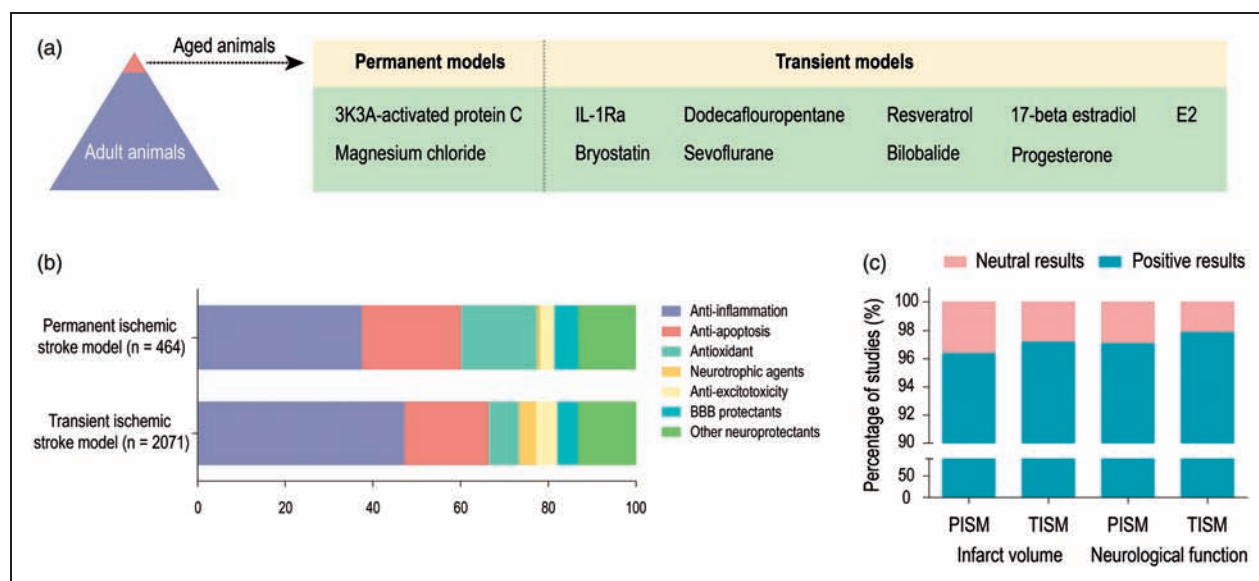


Figure 4. Aging imbalance, therapeutic targets, and publication bias in preclinical studies of ischemic stroke. (a) Out of more than 2,000 treatments, only two have been evaluated in an aged rodent model of permanent ischemia, and nine neuroprotectants have been assessed in aged rodent models of transient ischemia. (b) The potential mechanisms underlying the therapeutic properties of neuroprotectants in both transient and permanent stroke models. More than 80% of the neuroprotectants in the current analyses target inflammation, apoptosis, oxidative stress, neurotrophs, and excitotoxicity. (c) 95% of studies fitting all of our inclusion criteria reported positive outcomes in preclinical models. PISM: permanent ischemic stroke model; TISM: transient ischemic stroke model.

impaired cerebral perfusion, greater infarct sizes, white matter injury, and dysregulation of pro versus anti-inflammatory microglia.¹¹⁶ Similar age-related declines in endogenous defenses are also observed in clinical settings. Almost 72% of stroke occurrences are recorded in patients above the age of 65.¹¹⁷ A recent report from the American Heart Association (2018) indicated that the 30-day mortality rate of patients with ischemic stroke was 9.0% between 65 and 74 years of age, 13.1% between 74 and 84 years of age, and 23.0% in those 85 years of age or older.¹¹⁸ Aged animals can respond quite differently to treatment regimens than young animals.¹¹⁹ For example, melatonin prevents neuronal apoptosis by protecting nuclear and mitochondrial DNA damage when administered chronically to aged rats, but increases neuronal apoptosis when administered to younger rats,¹²⁰ perhaps because melatonin levels drop with age.¹²¹ Thus, future experimental stroke studies need to account for age-related changes in the brain and test the effects of neuroprotective agents in appropriately aged animals. In preclinical studies, researchers may also need to focus on distal branch occlusion for better translation to elderly humans, whereas in preclinical studies, middle cerebral artery occlusion is commonly employed.

The National Institutes of Neurological Disorders and Stroke reported that ~55,000 more women are afflicted annually by stroke than men.¹²² Women have a lower incidence of ischemic stroke than men when

they are younger, but this trend is reversed after 85 years of age.¹²³ In addition, women have higher post-stroke rates of mortality, disability, depression, and dementia compared to men.¹²⁴ Although this has been partially attributed to higher life expectancies in women, this gender disparity remains present even after adjusting for age differences in medical history and presentation.¹²⁵ In preclinical studies, young female rodents exhibit less brain injury than males after ischemic stroke.¹²⁶ This relative resilience can be abolished by ovariectomy or after reproductive senescence,¹²⁷ perhaps because the circulating sex steroid hormones, estrogen and progesterone, are potential neuroprotective factors after ischemic stroke.^{128,129}

In our analysis, only 3% of the preclinical ischemic stroke studies reported the effects of potential neuroprotectants in female animals. Given the gender dimorphisms presented above, it seems necessary to assess neuroprotective strategies in both males and females prior to clinical translation. For example, uric acid failed to achieve any overall improvement in patients with acute ischemic stroke, but it exerted a positive effect on functional recovery in female patients.⁴⁶ In the URICO-ICTUS trial, women displayed significantly lower serum uric acid concentrations than men, but their levels rose by 13% after uric acid therapy.⁴⁶ Therefore, it seems plausible that uric acid may be more valuable as a therapeutic agent against oxidative stress after ischemic stroke in female

patients. Nevertheless, our extensive analysis did not reveal any preclinical studies that assess the effects and potential mechanisms of uric acid in female animals.

Mismatch IV: Stroke is more devastating in patients with multiple comorbidities not captured in preclinical models

As mentioned above, almost 72% of ischemic strokes occur in patients above the age of 65.¹¹⁷ Serious comorbidities are present in this elderly cohort, including hypertension and diabetes, which are likely to influence the efficacy of neuroprotectants.¹³⁰ The INTERSTROKE study estimated that 90% of ischemic strokes arise due to 10 established risk factors, including hypertension, diabetes, high alcohol intake, current smoking habits, poor dietary choices, obesity, physical inactivity, psychosocial stress, and cardiac and lipid abnormalities.¹³¹ Despite these observations, the vast majority of experimental stroke studies have been performed on young and healthy rodents housed under optimal pathogen-free conditions.¹³² Of course, no single model can simulate all the comorbidities known to influence stroke outcomes in humans. However, several attempts have been made to devise animal models displaying the most frequent comorbidities, including diabetes^{133,134} and hypertension.^{135,136} Approximately 50 preclinical stroke studies have employed hypertensive animals and reported larger infarct volumes compared to normotensive animals.¹³⁷ Fewer preclinical investigations have assessed the effect of diabetes on the response to therapeutic interventions for ischemic stroke.^{138,139} For example, the genetically altered db/db mouse is used as a model of adult onset type-2 diabetes.¹⁴⁰ The db/db mice display larger infarct volumes, increased cerebral edema, and impaired functional recovery following ischemic stroke than db/+ mice.¹⁴⁰ In addition, db/db mice display higher expression of matrix metalloprotease(MMP)-9 than db/+ mice, resulting in increased blood-brain barrier (BBB) permeability, greater infiltration of macrophages and neutrophils into the infarct area, and higher inflammation.¹⁴¹

A number of issues need to be addressed in order to properly evaluate therapeutic interventions in animals with comorbid diseases, including selection of appropriate animal models. For example, there are important post-stroke sequelae not commonly assessed in animals, such as depression and other psychiatric consequences,¹⁴² obstructive sleep apnea,¹⁴³ and immunodepression, which frequently results in fever.¹⁴⁴ Given these complexities, it has been argued by some authors that “experimental models of stroke can cover only individual, specific aspects of this multifaceted

disease.”¹⁴⁵ Therefore, it is recommended that the effects of a therapeutic intervention on stroke injury and recovery should first be examined in adult male control and diseased (diabetic, hypertensive, or hyperlipidemic) animals, followed by studies in female and aged animals.¹⁴⁶ Ensuring that therapies are effective in multiple independent models that encompass specific comorbidities may increase the likelihood of clinical translation.

Mismatch V: Therapeutic targets and publication bias in preclinical versus clinical studies

The primary goal of neuroprotectants is to salvage the ischemic penumbra surrounding the infarcted core (Figure 4(b)). More than 80% of the neuroprotectants in the current analyses aid in neurotrophs and target inflammation, apoptosis, oxidative stress, and excitotoxicity (Figure 4(b)). The actions of these neuroprotective agents range from pharmacological blockade of neurotransmitter receptors to interference with cell death pathways. It is well established that one of the first molecular mechanisms underlying ischemic brain damage involves excitotoxicity.¹⁴⁷ Aside from massive and rapid release of the excitatory amino acid glutamate after ischemic stroke, the reuptake of this transmitter is also inhibited,¹⁴⁸ and the subsequent extracellular accumulation of glutamate activates downstream signaling pathways that flood cells with intracellular calcium, leading to activation of proteases and failure of mitochondrial bioenergetics.¹⁴⁸ Numerous pharmacological inhibitors of glutamate receptors have been examined in models of ischemic stroke and in stroke patients, but all failed to show efficacy in clinical trials, including N-methyl-D-aspartate (NMDA) receptor antagonists that reduce glutamate excitotoxicity, such as Selfotel, Aptiganel, and Gavestinel.^{149,150}

NMDA antagonists have been shown to reduce neurogenesis in the recovery phase of ischemic stroke, apart from inhibiting proteasomal function, which can lead to selective neuronal apoptosis.¹⁵¹ Although NMDA antagonists may alleviate early neuronal injury by suppressing glutamate excitotoxicity in the acute phase, the inhibition of long-term neurogenesis during the chronic recovery/repair phase may counteract the former benefits and contribute to failure of clinical translation. Thus, investigators have begun to target the signaling pathways downstream of glutamate receptor binding. For example, NA-1 is an inhibitor of the postsynaptic density-95 protein (PSD95), which couples NMDA receptor activation to nitric oxide neurotoxicity.¹⁵² NA-1 has been shown to halt glutamate excitotoxicity, reduce infarct volume, and improve long-term functional outcomes in rodents and

non-human primates after ischemic stroke.¹⁵³ Although clinical trials confirmed the safety of NA-1, no significant effects on functional recovery have been achieved.¹⁵⁴ However, the clinical study was geared towards testing neuroprotective effects in patients with stroke following endovascular aneurysmal repair and were therefore unlikely to harbor a salvageable penumbra anyway. Previous studies testing NA-1 in ischemic stroke primarily focused on its ability to protect neurons from excitotoxic damage.¹⁵⁵ Few studies were designed to characterize the influence of NA-1 on neurogenesis. A randomized double-blinded clinical trial, ESCAPE-NA-1 (NCT02930018), is currently ongoing to test the safety and efficacy of NA-1 administration in LVO patients with salvageable penumbral tissue and sufficient collateral circulation who have been treated with EVT.

In addition to the aforementioned issues contributing to the failure to translate preclinically effective treatments into clinical interventions (e.g. the absence of recanalization or functional evaluations, overreliance on young male animals, and a neglect of comorbidities), there have been many other impediments to meaningful progress. For example, neutral or negative results are difficult to publish in major peer-reviewed journals, which may introduce biases into preclinical and clinical research and may therefore influence the conclusions drawn in this report. Surprisingly, 95% of studies fitting all of our inclusion criteria reported positive outcomes in preclinical models (Figure 4(c)). As we do not believe that our search terms or inclusion criteria would have skewed the data dramatically, this degree of bias towards only reporting positive outcomes may be serious. There are too few rewards for publishing neutral or negative data and the resulting publication bias needs to be addressed at the levels of reviewers, editors, publishers, and funding bodies.

Perspectives

The primary function of stroke treatments should be to improve recanalization rates in the early stages after ischemic stroke. Recently, a number of novel thrombolytic drugs (e.g. tenecteplase, desmoteplase) with superior pharmacological profiles compared to alteplase (tPA) have been tested in Phase III trials.¹⁵⁶ Our previous meta-analysis of 819 patients from five clinical trials indicated that desmoteplase administered over an extended time window (3 to 9 h after stroke onset) exerted no significant effects upon functional recovery, but had a favorable safety profile in patients with acute ischemic stroke.¹⁵⁷ Furthermore, greater early-stage (4 to 8 h) reperfusion was observed in the desmoteplase-treated group (47.4%) compared to the placebo-treated group (37.5%). Although tenecteplase was not superior

to alteplase in improving recanalization rate in ischemic stroke patients, it was associated with a reduced risk of bleeding.¹⁵⁸ Recent data indicate that tenecteplase is superior to alteplase as a bridging agent for patients with LVO stroke undergoing intravenous thrombolysis in the 0–4.5 h time window prior to intended thrombectomy.¹⁵⁹

Reperfusion therapies with thrombolysis and mechanical thrombectomy are aimed at achieving tissue reperfusion and not simply recanalization. Clinical trials have shown that 70–80% of patients receiving thrombectomy can be successfully reperfused.⁷ This area of clinical intervention has great potential for growth. Stroke researchers will need to identify more effective and safer thrombolytic agents and improve the mechanical thrombectomy technique to not only achieve LV recanalization in the largest number of ischemic stroke patients, but also obtain maximum small vessel reperfusion in ischemic brains.

Effective neuroprotectants are needed to alleviate ischemia–reperfusion injury in multiple cell types. Ischemic stroke causes a failure in cellular bioenergetics in the ischemic core, including in neurons, astrocytes and endothelial cells of the BBB.¹⁶⁰ Impairments in the function of the ischemic BBB facilitate the entry of water and pro-inflammatory cytokines into the brain, leading to further activation of inflammatory responses and production of excess free radicals after reperfusion, culminating in the phenomenon of “reperfusion injury.”¹⁶⁰ Furthermore, ischemia and reperfusion injury can both induce hemorrhagic transformation.¹⁶⁰ Hence, drugs that protect the BBB and maintain its integrity might alleviate reperfusion injury and prevent hemorrhagic transformation after complete recanalization.

Neuroprotective candidates should be tested in both transient and permanent ischemia stroke models, and in young and aged animals of both genders before the initiation of a clinical trial. Permanent ischemia models may be superior for the testing of neuroprotectants that can maintain salvageable ischemic penumbra until reperfusion (i.e. bridging therapy), whereas transient ischemia models may be most suited to test reperfusion injury. In addition, preclinical studies are essential for establishing a comprehensive toxicity profile prior to any human exposures, and for identifying and then characterizing the precise molecular mechanism of action, which can accelerate the discovery of additional drug targets. Hypertensive and diabetic animals are particularly valuable in revealing the therapeutic potential of the neuroprotective candidate in more realistic settings of age-related diseases. Furthermore, experimental studies need to focus not only on short-term neuroprotection, which may be transient and only delay injury, but also on long-term

repair and recovery processes, including neurogenesis and other structural changes in the brain. Finally, all stroke studies should incorporate long-term assessments of multiple functional outcomes into their study design, particularly those that encompass higher order cognitive functions as well as anxiety and depression. In order to properly compare clinical testing conditions such as Rankin scores to animal research, assessments at 90 days post-injury may also be necessary. Furthermore, live animal imaging may be easier to relate to the functional imaging on live stroke patients rather than relying only on postmortem histological assessments.

Based on our extensive literature analyses, we recommend that combinations of neuroprotectants with thrombolytic drugs or endovascular thrombectomy should be studied as new therapeutic strategies for acute ischemic stroke. Rather than targeting one biological system/pathway or cell type with monotherapies, a multidimensional cocktail approach that includes thrombolytic agents, neuroprotectants, glioprotectants, vasoprotectants, anti-inflammatory drugs, anti-excitatory agents, and antioxidants delivered at both the optimal dose and time after onset of symptoms might improve success rates. The combination drug cocktail approach may also extend the duration of the ischemic penumbra.¹⁶¹ Other important points to consider are whether the temporal window of tPA or other intravenous thrombolytic agents such as tenecteplase can really be extended without increasing the risk for hemorrhagic transformation or blunting thrombolytic efficacy, but these risks may be mitigated by the concurrent use of neuroprotective interventions such as hypothermia, which is thought to mitigate reperfusion injury.^{162,163}

Conclusions

To summarize, the lack of success of preclinically validated neuroprotectants in clinical trials of stroke may be attributed to: (1) Failure to test the candidate molecule in combination with rapid, substantial reperfusion in patients, as in the experimental models, which suffer from overreliance on transient rather than permanent brain ischemia models; (2) paucity of preclinical studies designed to analyze treatment effects on clinically relevant long-term outcomes; (3) use of healthy young adult male animals instead of aged males and females with diabetes and other conditions; and (4) a publication bias toward reporting statistically significant, positive outcomes in preclinical studies versus large clinical trials.

Previous efforts in facilitating the success of clinical translation have prompted STAIR, STEPS, and RIGOR research guidelines, which have also proposed the use of females, aged animals, long-term functional

outcomes, and increased rigor with blinding, power analyses, and appropriate statistical tests.¹⁶⁴ Based on our literature analyses, we also encourage technical development of superior preclinical models that recapitulate permanent ischemia or removal of a thrombus followed by reperfusion. We recognize that these recommendations might involve completing fewer studies due to higher costs and greater animal numbers, but these higher demands are likely to reward biomedical researchers in the long run by accelerating the translation of their benchwork to the bedside and clinic.

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Declaration of conflicting interests

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References

1. Kuklina EV, Tong X, George MG, et al. Epidemiology and prevention of stroke: a worldwide perspective. *Expert Rev Neurother* 2012; 12: 199–208.
2. National Institute of Neurological Disorders, Stroke rt PA Study Group. Tissue plasminogen activator for acute ischemic stroke. *N Engl J Med* 1995; 333: 1581–1587.
3. Hacke W, Kaste M, Bluhmki E, et al. Thrombolysis with alteplase 3 to 4.5 hours after acute ischemic stroke. *N Engl J Med* 2008; 359: 1317–1329.
4. Furlan A, Higashida R, Wechsler L, et al. Intra-arterial prourokinase for acute ischemic stroke. The proact ii study: A randomized controlled trial. Prolyse in acute cerebral thromboembolism. *Jama* 1999; 282: 2003–2011.

5. Broderick JP, Palesch YY, Demchuk AM, et al. Endovascular therapy after intravenous t-PA versus t-PA alone for stroke. *N Engl J Med* 2013; 368: 893–903.
6. Campbell BCV, Donnan GA, Lees KR, et al. Endovascular stent thrombectomy: the new standard of care for large vessel ischaemic stroke. *Lancet Neurol* 2015; 14: 846–854.
7. Goyal M, Menon BK, van Zwam WH, et al. Endovascular thrombectomy after large-vessel ischaemic stroke: a meta-analysis of individual patient data from five randomised trials. *Lancet* 2016; 387: 1723–1731.
8. Nogueira RG, Jadhav AP, Haussen DC, et al. Thrombectomy 6 to 24 hours after stroke with a mismatch between deficit and infarct. *N Engl J Med* 2018; 378: 11–21.
9. Albers GW, Marks MP, Kemp S, et al. Thrombectomy for stroke at 6 to 16 hours with selection by perfusion imaging. *N Engl J Med* 2018; 378: 708–718.
10. Xu SY and Pan SY. The failure of animal models of neuroprotection in acute ischemic stroke to translate to clinical efficacy. *Med Sci Monit Basic Res* 2013; 19: 37–45.
11. Astrup J, Siesjo BK and Symon L. Thresholds in cerebral ischemia – the ischemic penumbra. *Stroke* 1981; 12: 723–725.
12. Olsen TS, Larsen B, Herning M, et al. Blood flow and vascular reactivity in collaterally perfused brain tissue. Evidence of an ischemic penumbra in patients with acute stroke. *Stroke* 1983; 14: 332–341.
13. Marchal G, Beaudouin V, Rioux P, et al. Prolonged persistence of substantial volumes of potentially viable brain tissue after stroke: a correlative PET-CT study with voxel-based data analysis. *Stroke* 1996; 27: 599–606.
14. Jones TH, Morawetz RB, Crowell RM, et al. Thresholds of focal cerebral ischemia in awake monkeys. *J Neurosurg* 1981; 54: 773–782.
15. Darby DG, Barber PA, Gerraty RP, et al. Pathophysiological topography of acute ischemia by combined diffusion-weighted and perfusion MRI. *Stroke* 1999; 30: 2043–2052.
16. Bardutzky J, Shen Q, Henninger N, et al. Characterizing tissue fate after transient cerebral ischemia of varying duration using quantitative diffusion and perfusion imaging. *Stroke* 2007; 38: 1336–1344.
17. Wheeler HM, Mlynash M, Inoue M, et al. The growth rate of early DWI lesions is highly variable and associated with penumbral salvage and clinical outcomes following endovascular reperfusion. *Int J Stroke* 2015; 10: 723–729.
18. Hakimelahi R, Vachha BA, Copen WA, et al. Time and diffusion lesion size in major anterior circulation ischemic strokes. *Stroke* 2014; 45: 2936–2941.
19. Rocha M and Jovin TG. Fast Versus slow progressors of infarct growth in large vessel occlusion stroke: clinical and research implications. *Stroke* 2017; 48: 2621–2627.
20. Copen WA, Rezai Gharai L, Barak ER, et al. Existence of the diffusion-perfusion mismatch within 24 hours after onset of acute stroke: dependence on proximal arterial occlusion. *Radiology* 2009; 250: 878–886.
21. Berkhemer OA, Fransen PS, Beumer D, et al. A randomized trial of intraarterial treatment for acute ischemic stroke. *N Engl J Med* 2015; 372: 11–20.
22. Goyal M, Demchuk AM, Menon BK, et al. Randomized assessment of rapid endovascular treatment of ischemic stroke. *N Engl J Med* 2015; 372: 1019–1030.
23. Saver JL, Goyal M, Bonafe A, et al. Stent-retriever thrombectomy after intravenous t-PA vs. t-PA alone in stroke. *N Engl J Med* 2015; 372: 2285–2295.
24. Campbell BC, Mitchell PJ, Kleinig TJ, et al. Endovascular therapy for ischemic stroke with perfusion-imaging selection. *N Engl J Med* 2015; 372: 1009–1018.
25. Jovin TG, Chamorro A, Cobo E, et al. Thrombectomy within 8 hours after symptom onset in ischemic stroke. *N Engl J Med* 2015; 372: 2296–2306.
26. Bhatia R, Hill MD, Shobha N, et al. Low rates of acute recanalization with intravenous recombinant tissue plasminogen activator in ischemic stroke: real-world experience and a call for action. *Stroke* 2010; 41: 2254–2258.
27. Seners P, Turc G, Maier B, et al. Incidence and predictors of early recanalization after intravenous thrombolysis: a systematic review and meta-analysis. *Stroke* 2016; 47: 2409–2412.
28. Saver JL, Goyal M, van der Lugt A, et al. Time to treatment with endovascular thrombectomy and outcomes from ischemic stroke: a meta-analysis. *JAMA* 2016; 316: 1279–1288.
29. Rangaraju S, Streib C, Aghaebrahim A, et al. Relationship between lesion topology and clinical outcome in anterior circulation large vessel occlusions. *Stroke* 2015; 46: 1787–1792.
30. Mizuma A, You JS and Yenari MA. Targeting reperfusion injury in the age of mechanical thrombectomy. *Stroke* 2018; 49: 1796–1802.
31. Seet RC and Rabinstein AA. Symptomatic intracranial hemorrhage following intravenous thrombolysis for acute ischemic stroke: a critical review of case definitions. *Cerebrovasc Dis* 2012; 34: 106–114.
32. Yaghi S, Willey JZ, Cucchiara B, et al. Treatment and outcome of hemorrhagic transformation after intravenous alteplase in acute ischemic stroke: a scientific statement for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2017; 48: e343–e361.
33. Neuberger U, Mohlenbruch MA, Herweh C, et al. Classification of bleeding events: comparison of ECASS III (European Cooperative Acute Stroke Study) and the New Heidelberg bleeding classification. *Stroke* 2017; 48: 1983–1985.
34. Adeoye O, Hornung R, Khatri P, et al. Recombinant tissue-type plasminogen activator use for ischemic stroke in the United States: a doubling of treatment rates over the course of 5 years. *Stroke* 2011; 42: 1952–1955.
35. Jadhav AP, Desai SM, Kenmuir CL, et al. Eligibility for endovascular trial enrollment in the 6- to 24-hour time window: analysis of a single comprehensive stroke center. *Stroke* 2018; 49: 1015–1017.

36. Dozois A, Hampton L, Kingston CW, et al. PLUMBER study (prevalence of large vessel occlusion strokes in Mecklenburg county emergency response). *Stroke* 2017; 48: 3397–3399.
37. Ginsberg MD. Expanding the concept of neuroprotection for acute ischemic stroke: the pivotal roles of reperfusion and the collateral circulation. *Prog Neurobiol* 2016; 145–146: 46–77.
38. Neuhaus AA, Couch Y, Hadley G, et al. Neuroprotection in stroke: the importance of collaboration and reproducibility. *Brain* 2017; 140: 2079–2092.
39. Lees KR, Bornstein N, Diener HC, et al. Results of membrane-activated chelator stroke intervention randomized trial of DP-b99 in acute ischemic stroke. *Stroke* 2013; 44: 580–584.
40. Diener HC, Schneider D, Lampl Y, et al. DP-b99, a membrane-activated metal ion chelator, as neuroprotective therapy in ischemic stroke. *Stroke* 2008; 39: 1774–1778.
41. Rosenberg G, Bornstein N, Diener HC, et al. The membrane-activated chelator stroke intervention (MACSI) Trial of DP-b99 in acute ischemic stroke: a randomized, double-blind, placebo-controlled, multinational pivotal phase III study. *Int J Stroke* 2011; 6: 362–367.
42. Becker BF. Towards the physiological function of uric acid. *Free Radic Biol Med* 1993; 14: 615–631.
43. Yu ZF, Bruce-Keller AJ, Goodman Y, et al. Uric acid protects neurons against excitotoxic and metabolic insults in cell culture, and against focal ischemic brain injury in vivo. *J Neurosci Res* 1998; 53: 613–625.
44. Romanos E, Planas AM, Amaro S, et al. Uric acid reduces brain damage and improves the benefits of rt-PA in a rat model of thromboembolic stroke. *J Cereb Blood Flow Metab* 2007; 27: 14–20.
45. Amaro S, Soy D, Obach V, et al. A pilot study of dual treatment with recombinant tissue plasminogen activator and uric acid in acute ischemic stroke. *Stroke* 2007; 38: 2173–2175.
46. Chamorro A, Amaro S, Castellanos M, et al. Safety and efficacy of uric acid in patients with acute stroke (URICO-ICTUS): a randomised, double-blind phase 2b/3 trial. *Lancet Neurol* 2014; 13: 453–460.
47. Chamorro A, Amaro S, Castellanos M, et al. Uric acid therapy improves the outcomes of stroke patients treated with intravenous tissue plasminogen activator and mechanical thrombectomy. *Int J Stroke* 2017; 12: 377–382.
48. Zhang C, Chopp M, Cui Y, et al. Cerebrolysin enhances neurogenesis in the ischemic brain and improves functional outcome after stroke. *J Neurosci Res* 2010; 88: 3275–3281.
49. Wang LC, Huang CY, Wang HK, et al. Magnesium sulfate and nimesulide have synergistic effects on rescuing brain damage after transient focal ischemia. *J Neurotrauma* 2012; 29: 1518–1529.
50. Xu F, Hongbin H, Yan J, et al. Greatly improved neuroprotective efficiency of citicoline by stereotactic delivery in treatment of ischemic injury. *Drug Deliv* 2011; 18: 461–467.
51. Park H, Hong M, Jhon GJ, et al. Repeated oral administration of human serum albumin protects from the cerebral ischemia in rat brain following MCAO. *Exp Neurobiol* 2017; 26: 151–157.
52. Yang Y, Li X, Zhang L, et al. Ginsenoside Rg1 suppressed inflammation and neuron apoptosis by activating PPARgamma/HO-1 in hippocampus in rat model of cerebral ischemia-reperfusion injury. *Int J Clin Exp Pathol* 2015; 8: 2484–2494.
53. Wang R, Wu X, Liang J, et al. Intra-artery infusion of recombinant human erythropoietin reduces blood-brain barrier disruption in rats following cerebral ischemia and reperfusion. *Int J Neurosci* 2015; 125: 693–702.
54. Moretti A, Ferrari F and Villa RF. Neuroprotection for ischaemic stroke: current status and challenges. *Pharmacol Ther* 2015; 146: 23–34.
55. Tymianski M. Combining neuroprotection with endovascular treatment of acute stroke: is there hope? *Stroke* 2017; 48: 1700–1705.
56. Onetti Y, Dantas AP, Perez B, et al. Middle cerebral artery remodeling following transient brain ischemia is linked to early postischemic hyperemia: a target of uric acid treatment. *Am J Physiol Heart Circ Physiol* 2015; 308: H862–H874.
57. Zhang B, Yang N, Lin SP, et al. Suitable concentrations of uric acid can reduce cell death in models of OGD and cerebral ischemia-reperfusion injury. *Cell Mol Neurobiol* 2017; 37: 931–939.
58. Amaro S, Llull L, Renu A, et al. Uric acid improves glucose-driven oxidative stress in human ischemic stroke. *Ann Neurol* 2015; 77: 775–783.
59. Song W, Wu YM, Ji Z, et al. Intra-carotid cold magnesium sulfate infusion induces selective cerebral hypothermia and neuroprotection in rats with transient middle cerebral artery occlusion. *Neurol Sci* 2013; 34: 479–486.
60. Izumi Y, Roussel S, Pinard E, et al. Reduction of infarct volume by magnesium after middle cerebral artery occlusion in rats. *J Cereb Blood Flow Metab* 1991; 11: 1025–1030.
61. Wang M, Wang Y, He J, et al. Albumin induces neuroprotection against ischemic stroke by altering Toll-like receptor 4 and regulatory T cells in mice. *CNS Neurol Disord Drug Targets* 2013; 12: 220–227.
62. Defazio RA, Zhao W, Deng X, et al. Albumin therapy enhances collateral perfusion after laser-induced middle cerebral artery branch occlusion: a laser speckle contrast flow study. *J Cereb Blood Flow Metab* 2012; 32: 2012–2022.
63. Li Y, Guan Y, Wang Y, et al. Neuroprotective effect of the ginsenoside Rg1 on cerebral ischemic injury in vivo and in vitro is mediated by PPARgamma-regulated anti-oxidative and anti-inflammatory pathways. *Evid Based Complement Alternat Med* 2017; 2017: 7842082.
64. Yang LX, Zhang X and Zhao G. Ginsenoside Rd attenuates DNA damage by increasing expression of DNA glycosylase endonuclease VIII-like proteins after focal cerebral ischemia. *Chin Med J* 2016; 129: 1955–1962.
65. Zhang B, Hata R, Zhu P, et al. Prevention of ischemic neuronal death by intravenous infusion of a ginseng saponin, ginsenoside Rb(1), that upregulates Bcl-x(L) expression. *J Cereb Blood Flow Metab* 2006; 26: 708–721.

66. Belayev L, Busto R, Zhao W, et al. Effect of delayed albumin hemodilution on infarction volume and brain edema after transient middle cerebral artery occlusion in rats. *J Neurosurg* 1997; 87: 595–601.
67. Yao X, Miao W, Li M, et al. Protective effect of albumin on VEGF and brain edema in acute ischemia in rats. *Neurosci Lett* 2010; 472: 179–183.
68. Tang J, Li YJ, Mu J, et al. Albumin ameliorates tissue plasminogen activator-mediated blood-brain barrier permeability and ischemic brain injury in rats. *Neurol Res* 2009; 31: 189–194.
69. Belayev L, Pinard E, Nallet H, et al. Albumin therapy of transient focal cerebral ischemia: in vivo analysis of dynamic microvascular responses. *Stroke* 2002; 33: 1077–1084.
70. Ginsberg MD, Palesch YY, Hill MD, et al. High-dose albumin treatment for acute ischaemic stroke (ALIAS) Part 2: a randomised, double-blind, phase 3, placebo-controlled trial. *Lancet Neurol* 2013; 12: 1049–1058.
71. Shuaib A, Lees KR, Lyden P, et al. NXY-059 for the treatment of acute ischemic stroke. *N Engl J Med* 2007; 357: 562–571.
72. Weng YC and Kriz J. Differential neuroprotective effects of a minocycline-based drug cocktail in transient and permanent focal cerebral ischemia. *Exp Neurol* 2007; 204: 433–442.
73. Britton P, Lu XC, Laskosky MS, et al. Dextromethorphan protects against cerebral injury following transient, but not permanent, focal ischemia in rats. *Life Sci* 1997; 60: 1729–1740.
74. Lo EH and Steinberg GK. Effects of dextromethorphan on regional cerebral blood flow in focal cerebral ischemia. *J Cereb Blood Flow Metab* 1991; 11: 803–809.
75. Porritt MJ, Andersson HC, Hou L, et al. Photothrombosis-induced infarction of the mouse cerebral cortex is not affected by the Nrf2-activator sulforaphane. *PLoS One* 2012; 7: e41090.
76. Alfieri A, Srivastava S, Siow RC, et al. Sulforaphane preconditioning of the Nrf2/HO-1 defense pathway protects the cerebral vasculature against blood-brain barrier disruption and neurological deficits in stroke. *Free Radic Biol Med* 2013; 65: 1012–1022.
77. Kim EH, Tolhurst AT, Szeto HH, et al. Targeting CD36-mediated inflammation reduces acute brain injury in transient, but not permanent, ischemic stroke. *CNS Neurosci Ther* 2015; 21: 385–391.
78. Liesz A, Sun L, Zhou W, et al. FTY720 reduces post-ischemic brain lymphocyte influx but does not improve outcome in permanent murine cerebral ischemia. *PLoS One* 2011; 6: e21312.
79. Kraft P, Gob E, Schuhmann MK, et al. FTY720 ameliorates acute ischemic stroke in mice by reducing thrombo-inflammation but not by direct neuroprotection. *Stroke* 2013; 44: 3202–3210.
80. Lukkarinen JA, Kauppinen RA, Grohn OH, et al. Neuroprotective role of ornithine decarboxylase activation in transient focal cerebral ischaemia: a study using ornithine decarboxylase-overexpressing transgenic rats. *Eur J Neurosci* 1998; 10: 2046–2055.
81. Raghavendra Rao VL, Dogan A, Bowen KK, et al. Ornithine decarboxylase knockdown exacerbates transient focal cerebral ischemia-induced neuronal damage in rat brain. *J Cereb Blood Flow Metab* 2001; 21: 945–954.
82. Fraser JF, Maniskas M, Trout A, et al. Intra-arterial verapamil post-thrombectomy is feasible, safe, and neuroprotective in stroke. *J Cereb Blood Flow Metab* 2017; 37: 3531–3543.
83. Cuccione E, Padovano G, Versace A, et al. Cerebral collateral circulation in experimental ischemic stroke. *Exp Transl Stroke Med* 2016; 8: 2.
84. Campbell BC, Christensen S, Tress BM, et al. Failure of collateral blood flow is associated with infarct growth in ischemic stroke. *J Cereb Blood Flow Metab* 2013; 33: 1168–1172.
85. Howells DW, Porritt MJ, Rewell SS, et al. Different strokes for different folks: the rich diversity of animal models of focal cerebral ischemia. *J Cereb Blood Flow Metab* 2010; 30: 1412–1431.
86. Lucitti JL, Sealock R, Buckley BK, et al. Variants of rab GTPase-effector binding protein-2 cause variation in the collateral circulation and severity of stroke. *Stroke* 2016; 47: 3022–3031.
87. Adams HP Jr, Effron MB, Torner J, et al. Emergency administration of abciximab for treatment of patients with acute ischemic stroke: results of an international phase III trial: abciximab in emergency treatment of stroke trial (AbESTT-II). *Stroke* 2008; 39: 87–99.
88. Lees KR, Zivin JA, Ashwood T, et al. NXY-059 for acute ischemic stroke. *N Engl J Med* 2006; 354: 588–600.
89. Davalos A, Blanco M, Pedraza S, et al. The clinical-DWI mismatch: a new diagnostic approach to the brain tissue at risk of infarction. *Neurology* 2004; 62: 2187–2192.
90. Viscoli CM, Brass LM, Kernan WN, et al. A clinical trial of estrogen-replacement therapy after ischemic stroke. *N Engl J Med* 2001; 345: 1243–1249.
91. Amarenco P, Bogousslavsky J, Callahan A, et al. High-dose atorvastatin after stroke or transient ischemic attack. *N Engl J Med* 2018; 355: 549–559.
92. Shinohara Y, Katayama Y, Uchiyama S, et al. Cilostazol for prevention of secondary stroke (CSPS 2): an aspirin-controlled, double-blind, randomised non-inferiority trial. *Lancet Neurol* 2010; 9: 959–968.
93. Ehrenreich H, Weissenborn K, Prange H, et al. Recombinant human erythropoietin in the treatment of acute ischemic stroke. *Stroke* 2009; 40: e647–656.
94. Liu X, Wang L, Wen A, et al. Ginsenoside-Rd improves outcome of acute ischaemic stroke – a randomized, double-blind, placebo-controlled, multicenter trial. *Eur J Neurol* 2012; 19: 855–863.
95. Alasheev AM, Belkin AA, Leiderman IN, et al. Granulocyte-colony-stimulating factor for acute ischemic stroke: a randomized controlled trial (STEMTHER). *Transl Stroke Res* 2011; 2: 358–365.
96. Padma Srivastava MV, Bhasin A, Bhatia R, et al. Efficacy of minocycline in acute ischemic stroke: a single-blinded, placebo-controlled trial. *Neurol India* 2012; 60: 23–28.
97. Feuerstein GZ, Zaleska MM, Krams M, et al. Missing steps in the STAIR case: a translational medicine

- perspective on the development of NXY-059 for treatment of acute ischemic stroke. *J Cereb Blood Flow Metab* 2008; 28: 217–219.
98. Palesch YY, Hill MD, Ryckborst KJ, et al. The ALIAS Pilot Trial: a dose-escalation and safety study of albumin therapy for acute ischemic stroke – II: neurologic outcome and efficacy analysis. *Stroke* 2006; 37: 2107–2114.
 99. Ladurner G, Kalvach P, Moessler H, et al. Neuroprotective treatment with cerebrolysin in patients with acute stroke: a randomised controlled trial. *J Neural Transm* 2005; 112: 415–428.
 100. Davalos A, Alvarez-Sabin J, Castillo J, et al. Citicoline in the treatment of acute ischaemic stroke: an international, randomised, multicentre, placebo-controlled study (ICTUS trial). *Lancet* 2012; 380: 349–357.
 101. Edaravone Acute Infarction Study G. Effect of a novel free radical scavenger, edaravone (MCI-186), on acute brain infarction. Randomized, placebo-controlled, double-blind study at multicenters. *Cerebrovasc Dis* 2003; 15: 222–229.
 102. Ishrat T, Sayeed I, Atif F, et al. Effects of progesterone administration on infarct volume and functional deficits following permanent focal cerebral ischemia in rats. *Brain Res* 2009; 1257: 94–101.
 103. Gagnard P, Frechou M, Schumacher M, et al. Progesterone reduces brain mitochondrial dysfunction after transient focal ischemia in male and female mice. *J Cereb Blood Flow Metab* 2016; 36: 562–568.
 104. Gibson CL and Murphy SP. Progesterone enhances functional recovery after middle cerebral artery occlusion in male mice. *J Cereb Blood Flow Metab* 2004; 24: 805–813.
 105. Jiang C, Zuo F, Wang Y, et al. Progesterone changes VEGF and BDNF expression and promotes neurogenesis after ischemic stroke. *Mol Neurobiol* 2016; 54: 571–581.
 106. Yousuf S, Sayeed I, Atif F, et al. Delayed progesterone treatment reduces brain infarction and improves functional outcomes after ischemic stroke: a time-window study in middle-aged rats. *J Cereb Blood Flow Metab* 2014; 34: 297–306.
 107. Wright DW, Yeatts SD, Silbergleit R, et al. Very early administration of progesterone for acute traumatic brain injury. *N Engl J Med* 2014; 371: 2457–2466.
 108. Skolnick BE, Maas AI, Narayan RK, et al. A clinical trial of progesterone for severe traumatic brain injury. *N Engl J Med* 2014; 371: 2467–2476.
 109. Diederich K, Frauenknecht K, Minnerup J, et al. Citicoline enhances neuroregenerative processes after experimental stroke in rats. *Stroke* 2012; 43: 1931–1940.
 110. Powers WJ, Derdeyn CP, Biller J, et al. 2015 American Heart Association/American Stroke Association focused update of the 2013 guidelines for the early management of patients with acute ischemic stroke regarding endovascular treatment: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2015; 46: 3020–3035.
 111. Wang Y, Zhao Z, Chow N, et al. Activated protein C analog protects from ischemic stroke and extends the therapeutic window of tissue-type plasminogen activator in aged female mice and hypertensive rats. *Stroke* 2013; 44: 3529–3536.
 112. Davis M, Perry RH and Mendelow AD. The effect of non-competitive N-methyl-D-aspartate receptor antagonism on cerebral oedema and cerebral infarct size in the aging ischaemic brain. *Acta Neurochir Suppl* 1997; 70: 30–33.
 113. Shapira S, Sapir M, Wengier A, et al. Aging has a complex effect on a rat model of ischemic stroke. *Brain Res* 2002; 925: 148–158.
 114. Wang RY, Wang PS and Yang YR. Effect of age in rats following middle cerebral artery occlusion. *Gerontology* 2003; 49: 27–32.
 115. Li S, Zheng J and Carmichael ST. Increased oxidative protein and DNA damage but decreased stress response in the aged brain following experimental stroke. *Neurobiol Dis* 2005; 18: 432–440.
 116. Suenaga J, Hu X, Pu H, et al. White matter injury and microglia/macrophage polarization are strongly linked with age-related long-term deficits in neurological function after stroke. *Exp Neurol* 2015; 272: 109–119.
 117. Writing Group M, Mozaffarian D, Benjamin EJ, et al. Heart disease and stroke statistics – 2016 update: a report from the American Heart Association. *Circulation* 2016; 133: e38–360.
 118. Benjamin EJ, Virani SS, Callaway CW, et al. Heart disease and stroke statistics – 2018 update: a report from the American Heart Association. *Circulation* 2018; 137: e67–e492.
 119. Dickstein DL, Kabaso D, Rocher AB, et al. Changes in the structural complexity of the aged brain. *Aging Cell* 2007; 6: 275–284.
 120. Krestinina OV, Odinkova IV, Baburina YL, et al. Age-related effect of melatonin on permeability transition pore opening in rat brain mitochondria. *Biochem Suppl Ser A Membr Cell Biol* 2013; 7: 286–293.
 121. Karasek M. Melatonin, human aging, and age-related diseases. *Exp Gerontol* 2004; 39: 1723–1729.
 122. Demel SL, Kittner S, Ley SH, et al. Stroke risk factors unique to women. *Stroke* 2018; 49: 518–523.
 123. Petrea RE, Beiser AS, Seshadri S, et al. Gender differences in stroke incidence and poststroke disability in the Framingham heart study. *Stroke* 2009; 40: 1032–1037.
 124. Kapral MK, Fang J, Hill MD, et al. Sex differences in stroke care and outcomes: results from the Registry of the Canadian Stroke Network. *Stroke* 2005; 36: 809–814.
 125. Park SJ, Shin SD, Ro YS, et al. Gender differences in emergency stroke care and hospital outcome in acute ischemic stroke: a multicenter observational study. *Am J Emerg Med* 2013; 31: 178–184.
 126. Cheng J and Hurn PD. Sex shapes experimental ischemic brain injury. *Steroids* 2010; 75: 754–759.
 127. Alkayed NJ, Murphy SJ, Traystman RJ, et al. Neuroprotective effects of female gonadal steroids in reproductively senescent female rats. *Stroke* 2000; 31: 161–168.
 128. Gibson CL, Gray LJ, Murphy SP, et al. Estrogens and experimental ischemic stroke: a systematic review. *J Cereb Blood Flow Metab* 2006; 26: 1103–1113.

129. Gibson CL, Coomber B and Rathbone J. Is progesterone a candidate neuroprotective factor for treatment following ischemic stroke? *Neuroscientist* 2009; 15: 324–332.
130. Fischer U, Arnold M, Nedeltchev K, et al. Impact of comorbidity on ischemic stroke outcome. *Acta Neurol Scand* 2006; 113: 108–113.
131. O'Donnell MJ, Xavier D, Liu L, et al. Risk factors for ischaemic and intracerebral haemorrhagic stroke in 22 countries (the INTERSTROKE study): a case-control study. *Lancet* 2010; 376: 112–123.
132. Dirnagl U. Thomas Willis lecture: is translational stroke research broken, and if so, how can we fix it? *Stroke* 2016; 47: 2148–2153.
133. Ferris JK, Peters S, Brown KE, et al. Type-2 diabetes mellitus reduces cortical thickness and decreases oxidative metabolism in sensorimotor regions after stroke. *J Cereb Blood Flow Metab* 2018; 38: 823–834.
134. Bevers MB, Vaishnav NH, Pham L, et al. Hyperglycemia is associated with more severe cytotoxic injury after stroke. *J Cereb Blood Flow Metab* 2017; 37: 2577–2583.
135. Chan SL and Cipolla MJ. Treatment with low dose fasudil for acute ischemic stroke in chronic hypertension. *J Cereb Blood Flow Metab* 2017; 37: 3262–3270.
136. Cipolla MJ, Sweet JG and Chan SL. Effect of hypertension and peroxynitrite decomposition with FeTMPyP on CBF and stroke outcome. *J Cereb Blood Flow Metab* 2017; 37: 1276–1285.
137. O'Collins VE, Donnan GA, Macleod MR, et al. Hypertension and experimental stroke therapies. *J Cereb Blood Flow Metab* 2013; 33: 1141–1147.
138. Kelly-Cobbs AI, Prakash R, Li W, et al. Targets of vascular protection in acute ischemic stroke differ in type 2 diabetes. *Am J Physiol Heart Circ Physiol* 2013; 304: H806–815.
139. Yuen NY, Chechneva OV, Chen YJ, et al. Exacerbated brain edema in a rat streptozotocin model of hyperglycemic ischemic stroke: evidence for involvement of blood-brain barrier Na-K-Cl cotransport and Na/H exchange. *J Cereb Blood Flow Metab*. Epub ahead of print 9 May 2018. DOI: 10.1177/0271678X18770844.
140. Hummel KP, Dickie MM and Coleman DL. Diabetes, a new mutation in the mouse. *Science* 1966; 153: 1127–1128.
141. Kumari R, Willing LB, Patel SD, et al. Increased cerebral matrix metalloproteinase-9 activity is associated with compromised recovery in the diabetic db/db mouse following a stroke. *J Neurochem* 2011; 119: 1029–1040.
142. Robinson RG and Jorge RE. Post-stroke depression: a review. *Am J Psychiatry* 2016; 173: 221–231.
143. Swartz RH, Bayley M, Lanctot KL, et al. Post-stroke depression, obstructive sleep apnea, and cognitive impairment: rationale for, and barriers to, routine screening. *Int J Stroke* 2016; 11: 509–518.
144. Haeusler KG, Schmidt WU, Fohring F, et al. Cellular immunodepression preceding infectious complications after acute ischemic stroke in humans. *Cerebrovasc Dis* 2008; 25: 50–58.
145. Sommer CJ. Ischemic stroke: experimental models and reality. *Acta Neuropathol* 2017; 133: 245–261.
146. Ergul A, Hafez S, Fouda A, et al. Impact of comorbidities on acute injury and recovery in preclinical stroke research: focus on hypertension and diabetes. *Transl Stroke Res* 2016; 7: 248–260.
147. Rothman SM and Olney JW. Glutamate and the pathophysiology of hypoxic-ischemic brain damage. *Ann Neurol* 1986; 19: 105–111.
148. Chamorro A, Dirnagl U, Urra X, et al. Neuroprotection in acute stroke: targeting excitotoxicity, oxidative and nitrosative stress, and inflammation. *Lancet Neurol* 2016; 15: 869–881.
149. Lai TW, Zhang S and Wang YT. Excitotoxicity and stroke: identifying novel targets for neuroprotection. *Prog Neurobiol* 2014; 115: 157–188.
150. Hoyte L, Barber PA, Buchan AM, et al. The rise and fall of NMDA antagonists for ischemic stroke. *Curr Mol Med* 2004; 4: 131–136.
151. Arvidsson A, Kokaia Z and Lindvall O. N-methyl-D-aspartate receptor-mediated increase of neurogenesis in adult rat dentate gyrus following stroke. *Eur J Neurosci* 2001; 14: 10–18.
152. Sattler R, Xiong Z, Lu WY, et al. Specific coupling of NMDA receptor activation to nitric oxide neurotoxicity by PSD-95 protein. *Science* 1999; 284: 1845–1848.
153. Cook DJ, Teves L and Tymianski M. Treatment of stroke with a PSD-95 inhibitor in the gyrencephalic primate brain. *Nature* 2012; 483: 213–217.
154. Hill MD, Martin RH, Mikulis D, et al. Safety and efficacy of NA-1 in patients with iatrogenic stroke after endovascular aneurysm repair (ENACT): a phase 2, randomised, double-blind, placebo-controlled trial. *Lancet Neurol* 2012; 11: 942–950.
155. Ballarin B and Tymianski M. Discovery and development of NA-1 for the treatment of acute ischemic stroke. *Acta Pharmacol Sin* 2018; 39: 661–668.
156. Powers WJ, Rabinstein AA, Ackerson T, et al. 2018 Guidelines for the early management of patients with acute ischemic stroke: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2018; 49: e46–e110.
157. Shi L, Liang F, Li Y, et al. Desmoteplase for acute ischemic stroke within 3 to 9 hours after symptom onset: evidence from randomized controlled trials. *Sci Rep* 2016; 6: 33989.
158. Guillermin A, Yan DJ, Perrier A, et al. Safety and efficacy of tenecteplase versus alteplase in acute coronary syndrome: a systematic review and meta-analysis of randomized trials. *Arch Med Sci* 2016; 12: 1181–1187.
159. Campbell BC, Mitchell PJ, Churilov L, et al. Tenecteplase versus alteplase before endovascular thrombectomy (EXTEND-IA TNK): a multicenter, randomized, controlled study. *Int J Stroke* 2018; 13: 328–334.
160. Khatri R, McKinney AM, Swenson B, et al. Blood-brain barrier, reperfusion injury, and hemorrhagic transformation in acute ischemic stroke. *Neurology* 2012; 79: S52–S57.

161. Carmichael ST, Archibeque I, Luke L, et al. Growth-associated gene expression after stroke: evidence for a growth-promoting region in peri-infarct cortex. *Exp Neurol* 2005; 193: 291–311.
162. Yenari MA and Hemmen TM. Therapeutic hypothermia for brain ischemia: where have we come and where do we go? *Stroke* 2010; 41: S72–74.
163. Tang XN, Liu L and Yenari MA. Combination therapy with hypothermia for treatment of cerebral ischemia. *J Neurotrauma* 2009; 26: 325–331.
164. Lapchak PA, Zhang JH and Noble-Haeusslein LJ. RIGOR guidelines: escalating STAIR and STEPS for effective translational research. *Transl Stroke Res* 2013; 4: 279–285.