



# Amperometric Glucose Sensing at Nanomolar Level Using MOF-Encapsulated TiO<sub>2</sub> Platform

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**S** [Supporting Information](#page-5-0)

ABSTRACT: A new synthetic approach is established where both  $TiO<sub>2</sub>$ nanoparticles and glucose oxidase  $(\widehat{GO}_x)$  are together encapsulated into the cavity of ZIF-8 metal−organic framework (MOF) to fabricate a mediator-free glucose sensor in aqueous media. ZIF-8 possesses high stability both physically and chemically. Moreover, its large surface area and tunable cavity size are supportive to encapsulate both nanoparticles (TiO<sub>2</sub>) and enzymes (GO<sub>x</sub>). The as-synthesized nanocomposite is methodically characterized by various advanced analytical techniques, which suggests that  $TiO<sub>2</sub>$  is uniformly distributed within the cavity of ZIF-8 MOF. High surface area and double-layer capacitance of nanostructured  $TiO<sub>2</sub>$ jointly enhance the catalytic biosensor activity. The as-synthesized nanocomposite exhibits commendable stability and is able to detect low-level concentration (80 nM) of glucose in aqueous media by utilizing very low concentration of  $GO_x$  (62  $\mu$ g in 1 mL).



# 1. INTRODUCTION

Nowadays, diabetes mellitus is known to be the most prevalent chronic disease when compared to other harmful diseases that affect human body. There are few reasons given by the World Diabetes Foundation: the disease occurs mainly due to obesity and lack of physical activity. Moreover, it is occurring frequently due to rapid urbanization and modern lifestyle acquired by people. The disease has a direct effect as it destabilizes the level of blood glucose in our body, which ultimately causes grave health problems including cardiac arrest, kidney function failure, and even deterioration of neurons.<sup>1</sup> Generally, doctors recommend patients suffering from this disease to check their level of blood glucose periodically and to take invasive shots of insulin sporadically to manage the level of blood glucose for a long time.<sup>[2](#page-5-0)</sup> Because the available routine methods are associated with invasive methods such as injection, which is painful, glucose monitoring in a noninvasive way is highly necessary.<sup>3 $\leq$ [5](#page-6-0)</sup> It is been a long time since the first proposal from Clark and Lyon in 1962 regarding the concept of glucose sensing, and still new approaches are being invited by the scientific fraternity. In earlier versions, atmospheric oxygen is used as mediator for glucose sensor response. As technology evolves, new-generation glucose sensor had been introduced according to the demand. As per the current requirement, sophisticated techniques are in high demand, requiring less amount of enzyme and operating noninvasively. Saliva/sweat can be taken as an excellent medium of choice for noninvasive point-of-care glucose monitoring. The state of the art in this area is reviewed by various authors. $6-8$  $6-8$  $6-8$  One of the major challenges to date to fabricate sophisticated glucose sensor of such type is the long-

term stability of the enzyme. Generally, enzymes, including also glucose oxidase (GOD), are very much sensitive toward pH and temperature. New concepts were introduced to freeze the activity of  $GO<sub>x</sub>$  by encapsulating into some rigid cavity. Mediator-free direct electron transfer between the electrode and the active center of a redox protein is one of the most important perceptions for the development of next-generation biosensor devices.<sup>[9](#page-6-0)</sup> Flavin adenine dinucleotide (FAD)/ FADH2 present at the prosthetic group of glucose oxidase  $(GO<sub>x</sub>)$  is known to be the active redox species, but it is deeply enclosed inside the nonconducting layer of rigid protein  $(GO_x)$ structure.<sup>[9,10](#page-6-0)</sup>

Metal−organic framework (MOF) shows promising diversity and fascinated scientific community for its exclusive framework design, large surface-to-volume ratio, and tunable pore sizes[.11](#page-6-0) Zinc-imidazole framework 8 (ZIF-8) is a subclass of zeolitic MOF. ZIF-8 is a well-known microstructure due to its unique cavity and tunable pore size to accommodate small to large biomolecules and protect them from major external stimuli[.12](#page-6-0)<sup>−</sup>[20](#page-6-0) On the other hand, metal nanoparticles possess exceptional physical and chemical properties and can be used as a component of composite as it possesses high surface area, which assists in reducing the electron tunneling distance.<sup>[21](#page-6-0)-[26](#page-6-0)</sup> Tandem catalysis is one of the interesting properties of a catalyst. If a single catalyst can govern two concurrent reactions, it is called a tandem catalyst.  $GO_x$  encapsulated into MOF moiety can act as tandem catalyst for glucose

Received: August 10, 2018 Accepted: October 24, 2018 Published: November 1, 2018 oxidation and  $H_2O_2$  reduction.<sup>[27](#page-6-0)</sup> TiO<sub>2</sub> nanoparticle is known to have high charge-storage ability due to high double-layer capacitance. Using the same, there are reports where  $TiO<sub>2</sub>$  had been used as an active catalyst for nonenzymatic biosensor. The double-layer capacitance of  $TiO<sub>2</sub>$  can be used as an effective source for charge-transfer agent at electrode− electrolyte interface.<sup>[9,28](#page-6-0),[29](#page-6-0)</sup>

In this work, we have proposed an innovative protocol to encapsulate both  $TiO<sub>2</sub>$  and  $GO<sub>x</sub>$  jointly inside the cavity of ZIF-8 to fabricate the as-synthesized nanocomposite  $GO<sub>x</sub>@$  $ZIF-8(TiO<sub>2</sub>)$  and successfully utilize the same for the tandem glucose sensor application. The as-synthesized material is dropcast on a glassy carbon electrode (GCE), and amperometry was performed to obtain the sensor response. The composite shows a detection limit as low as 80 nM of glucose, which accredits the monitoring of glucose from sweat. Commercially available  $TiO<sub>2</sub>$  nanopowder has been used after probe sonication. Moreover, the catalyst shows tandem catalytic behavior to both glucose oxidation and peroxide reduction simultaneously. The oxidation of glucose to gluconate was also confirmed by standard colorimetric experiment by assaying glucose with glucose oxidase−peroxidase (GOD−POD) moiety. The redox adjustment of the couple  $FAD/FADH<sub>2</sub>$ makes the catalyst different toward sophisticated glucometer fabrication. This is the very first time that both a semiconductor nanoparticle and an enzyme are encapsulated into an MOF cavity. The presented work also envisaged low-level detection of glucose (80 nM), which is significantly low compared to the previously reported  $TiO<sub>2</sub>$ -based glucose sensor.

## 2. RESULTS AND DISCUSSION

2.1. Characterization of Materials. The as-synthesized  $GO_x@ZIF-8(TiO_2)$  material shows characteristic features due to the presence of crystalline  $TiO<sub>2</sub>$ . Powder X-ray diffraction (PXRD) was performed to determine the phases of the synthesized probe after the compound was vacuum-dried for 24 h. The PXRD spectra of the as-synthesized  $GO_x@ZIF$ - $8(TiO<sub>2</sub>)$  are depicted in red in Figure 1. The PXRD spectra of



Figure 1. PXRD patterns of synthesized  $GO_x@ZIF-8(TiO_2)$  along with simulated ZIF-8 (JCPDS:  $00-062-1030$ ) GO<sub>x</sub> for comparison.

the as-synthesized composite are compared to the simulated PXRD data of the standard ZIF-8, as depicted in blue of the same figure. It is clearly visible from the PXRD comparison data that there is a >95% PXRD peak matching of the synthesized probe compared to the standard peaks of ZIF-8, suggesting that the native structural property of ZIF-8 remains intact after the encapsulation of  $TiO<sub>2</sub>$  and  $GO<sub>x</sub>$ . Two distinct peaks at 25.36 and 48.17° confirm the presence of anatase  $TiO<sub>2</sub>$  (JCPDS: 21-1272).<sup>[30](#page-6-0)</sup> The standard basal plane of TiO<sub>2</sub> (200) at 48.17° estimated from the XRD data is correlated perfectly with the TEM result.

Field emission scanning electron microscopy (FE-SEM) was done to investigate the surface morphology of  $GO<sub>x</sub>@ZIF 8(TiO<sub>2</sub>)$  and is depicted in [Figure 2A](#page-2-0), which represents a distinct flowerlike structure having many sheets at low magnification. At high magnification, a hexagonal sheet was observed, as depicted in [Figure 2](#page-2-0)B. The morphology reveals high surface area of the probe due to the presence of  $TiO<sub>2</sub>$ nanopowder.  $GO_x$  is supposed to play a vital role along with  $TiO<sub>2</sub>$  in governing the morphology. It was previously known that  $GO_x$  has an affinity toward imidazole by hydrophobic interaction and hydrogen bonding.<sup>27</sup> Moreover, imidazole is considered to be the building block of ZIF-8, and hence  $GO_x$ controls the morphology of the as-synthesized probe. Further, transmission electron microscopy (TEM) was performed to investigate the spatial structure of the synthesized microstructure and is depicted in [Figure 2C](#page-2-0). Distinctive lattice fringes were observed, probably depicting the presence of TiO<sub>2</sub>. High-resolution transmission electron microscopy (HR-TEM) was performed to obtain the plane of  $TiO<sub>2</sub>$  lattice, and the result is depicted in [Figure 2](#page-2-0)D. The HR-TEM image shows the presence of (200) plane of anatase  $TiO<sub>2</sub>$  with distinct metal fringes and a d-spacing of 0.189 nm.

The result also matches with the PXRD data, which confirms the presence of  $(200)$  plane of TiO<sub>2</sub>. Scanning transmission electron microscope−energy-dispersive X-ray spectroscopy (STEM−EDX) had been done to get further concrete information regarding the encapsulation of  $TiO<sub>2</sub>$  into ZIF-8 moiety, and the result is depicted in [Figure S1](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information). The result depicts that the  $TiO<sub>2</sub>$  nanoparticle is distributed homogeneously with an almost equal spatial distance within the cavity of ZIF-8 microstructure.

ZIF-8 possesses tunable pore size, which can accommodate biomolecules. Fourier transform infrared spectroscopy (FT-IR) had been performed to check whether  $GO_x$  possesses its native structure after encapsulation. The FT-IR spectrum is depicted in [Figure 3.](#page-2-0) Characteristic IR peaks for  $GO_x$  are observed at 1651 and 1420 cm<sup>−</sup><sup>1</sup> . The peak at 1651 cm<sup>−</sup><sup>1</sup> is considered to be the stretching vibration of peptidic  $-C=O$  group of amide-I, whereas the peak at  $1420 \text{ cm}^{-1}$  corroborates the in-plane bending vibration of  $N_5H$  in FAD group.<sup>[31](#page-6-0)</sup> It can be clearly seen from the FT-IR spectrum that 1654 cm<sup>-1</sup> peak of amide-I slightly shifted to 1651 cm<sup>-1</sup>. The resultant shift is due to the effect of amide-I, which is already reported to be highly sensitive toward structural modification.<sup>3</sup>

One of the prime concerns to obtain FT-IR spectra is to validate the presence of  $GO<sub>x</sub>$  in its native state, and the existence of two peaks at 1651 and 1420 cm<sup>−</sup><sup>1</sup> clearly suggests that  $GO_x$  is not degraded by any external stimuli and possesses its native biological structure inside the cavity of ZIF-8 MOF, showing biocatalytic activity.

Thermogravimetric analysis (TGA) had been done to estimate the thermal stability of the as-synthesized probe. The TGA profile is depicted in [Figure S2](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information). It is clearly seen from [Figure S2](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information) that there is a steep decrease of weight loss percentage due to MOF decomposition. The weight loss at 165 °C corresponds to the evaporation of occluded water and organic components. The latter corresponds to framework

<span id="page-2-0"></span>

Figure 2. FE-SEM image of synthesized GO<sub>x</sub>@ZIF-8(TiO2): (A) Multisheet-assembled growth of the probe; (B) magnified image showing a hexagonal single sheet having large surface area; (C) TEM image of TiO<sub>2</sub> growth-assembled nanoparticles; (D) HR-TEM image of TiO<sub>2</sub> lattice fringes consisting of 200  $TiO<sub>2</sub>$  plane (anatase).



Figure 3. FT-IR spectra of  $GO_0 \times QZIF-8(TiO_2)$  showing distinctive peaks for the presence of glucose oxidase  $(GO_x)$  in its native form.

degradation. The analysis envisaged the stability profile of the as-synthesized probe, and it can be concluded that the probe is stable up to ∼200 °C.

The nitrogen adsorption−desorption curve was plotted and a hysteresis was observed in the high-pressure region (>0.8 P/  $P_0$ ), proposing the formation of type IV isotherm, which is associated with capillary condensation taking place in mesopores and the limited uptake over a range of high  $P/P<sub>0</sub>$ . The initial part of the type IV isotherm is attributed to monolayer−multilayer adsorption since it follows the same path as the corresponding part of the type II isotherm obtained with the given adsorption on the same surface area of the adsorbent in a nonporous form. The adsorption−desorption Brunauer−Emmett−Teller (BET) isotherm is depicted in Figure 4. The BET surface area was measured as 353.8  $\mathrm{m}^2/\mathrm{g}$ , which is quite superior to that of the previously reported  $TiO<sub>2</sub>$ composite. High surface area clearly envisaged uniform encapsulation of  $GO_x$  and  $TiO_2$  into the ZIF-8 cavity. To



Figure 4. Nitrogen adsorption−desorption isotherm of GO<sub>x</sub>@ZIF- $8(TiO<sub>2</sub>)$  showing hysteresis at >0.8  $P/P<sub>0</sub>$ . The average pore volume and pore diameter are calculated from the BJH plot as depicted in the inset.

obtain the pore size distribution of the synthesized probe, the Barrett−Joyner−Halenda (BJH) plot was plotted and is depicted in the inset of Figure 4. The average pore volume and pore diameter are found to be 0.248853  $\text{cm}^3/\text{g}$  (STP) and 28 Å, respectively. The result is attributed to the ability of ZIF-8 matrix to accommodate  $TiO<sub>2</sub>$  and  $GO<sub>x</sub>$ .

2.2. GOD-POD Test to Assay D-Glucose. It has been already shown that  $GO_x$  can possess its native structure inside ZIF-8 moiety due to the exceptional tunable pore size of ZIF-8, which accommodates  $GO_x$  and protects it from the external chemical stimuli. Furthermore, to check the bioactivity of  $GO_x$ inside the ZIF-8 MOF, glucose oxidase−peroxidase (GOD− POD) assay test was done. The general principle of this test is basically the oxidation of glucose to gluconate in the presence of  $GO_x$ , which acts as an enzyme. The reaction ends up with the formation of hydrogen peroxide  $(H_2O_2, eqs \ 1 \text{ and } 2),$  $(H_2O_2, eqs \ 1 \text{ and } 2),$  $(H_2O_2, eqs \ 1 \text{ and } 2),$  <span id="page-3-0"></span>which can be uptaken by 4-aminophenazone, a chromogenic oxygen acceptor, and in the presence of phenol, a peroxidase (POD) quinone dye is formed, which shows red color upon completion of the reaction<sup>[27](#page-6-0)</sup>

$$
\beta-\text{D-glucose} + \text{O}_2 + \text{H}_2\text{O} \xrightarrow{\text{GO}_2} \text{gluconate} + \text{H}_2\text{O}_2 \tag{1}
$$

 $H_2O_2$  + phenol + (4-AP)  $\stackrel{POD}{\longrightarrow}$  red quinone dye +  $H_2O$ (2)

GOD−POD was performed to examine the activity of the probe with possible interferent analytes like fructose, galactose, lactose, and urea, and the outcome is visible by the naked eye, which is depicted in [Figure S3](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information). It is clearly visible that the synthesized probe is exclusively selective toward glucose. Characteristic UV−vis absorbance peak at 505 nm wavelength was found, which confirms the presence of  $GO_x$  inside the as-synthesized composite probe. The outcome is depicted in [Figure S4](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information). The result suggests that  $GO_x$  is biocatalytically active and definitely encapsulated into ZIF-8 matrix.

2.3. Biosensor Response of  $GO_x@ZIF-8$  (TiO<sub>2</sub>). High surface coverage of  $TiO<sub>2</sub>$  and the excellent biocatalytic activity of GO<sub>x</sub> demonstrate the as-synthesized ZIF-8 composite to be an admirable candidate for the fabrication of glucose biosensor, accrediting a low-level detection of glucose from sweat/saliva. Cyclic voltammetry (CV) technique is used to check the faradic behavior of electrolyte or species attached with electrode, and for this purpose, CV experiment was performed using  $GO_x \omega ZIF-8(TiO_2)$ -modified GCE at a scan rate of 50 mV/s. Prior to use, GCE was polished in a 0.3  $\mu$  alumina slurry by putting in a high-mesh pad, followed by ultrasonication in 1:1 ethanol−water mixture for 15 min. Finally, the GCE was washed with Milli-Q water several times and dried under room temperature. The electrode was kept covered to avoid any unavoidable dust particles. The as-synthesized composite (5 mg) was taken and homogeneous dispersion was prepared by taking the same 1 mL of 0.1 M pH 7.4 phosphate-buffered saline (PBS), and the resultant solution was ultrasonicated for 10 min. The resulting dispersion was used as the aliquot for further electrochemical experiment. A small amount of the aliquot (10  $\mu$ M) was pipetted out using a micropipette and drop-cast over the clean GCE of 5 mm diameter. The electrode was dried at room temperature for 6 h, covered to avoid external dust, and stored overnight at 4 °C.

The tailored electrode was then immersed into a different concentration of glucose in 0.1 M pH 7.4 PBS. The concentration of glucose was increased from 0 to 10 mM, and CV was taken in the range of −0.6 to +0.6 V. Pt wire was used as the counter electrode, and Ag/AgCl (sat. KCl) was used as the reference electrode. A decrement of cathodic current at around −0.45 V vs Ag/AgCl (sat. KCl) was observed while glucose concentration was increased. The result is depicted in Figure 5. The cathodic potential for the possible glucose oxidation is quite similar to that of the other reported catalysts (enzymatic and nonenzymatic). Cathodic peak current was observed to decrease with increasing concentration and plotted against concentration of glucose. A linear increment of anodic current was found with increasing concentration, and the result is depicted in the inset of Figure 5.

The result shows the excellent catalytic activity of the assynthesized probe to glucose oxidation reaction due to the



Figure 5. Cyclic voltammetry of synthesized probe with different concentration of glucose in pH 7.4 PBS using a three-electrode cell setup. Calibration of cathodic peak current versus glucose concentration is depicted in the inset.

production of  $H_2O_2$ , which results in a decrement of cathodic current/increment of anodic current. Although the nature of the obtained current is nonfaradic (mostly capacitive), the response is well sharp. The possible oxidation reaction occurring at the electrode (GCE)−electrolyte (glucose) interface can be correlated by the following equations (eqs 3 and  $4)$ 

glucose +  $GO_x$  – FAD  $\rightarrow$  gluconate +  $GO_x$  – FADH<sub>2</sub> (3)  $GO_x - FADH_2 + O_2 \rightarrow GO_x - FAD + H_2O_2$  (4)

The probe  $GO_x \omega ZIF-8(TiO_2)$  not only catalyzes the glucose oxidation reaction but also shows distinctive tandem catalytic property, which makes itself different from other reported amperometric glucose sensors. The probe possesses tandem catalytic activity, which is very unique as it can catalyze the reduction of  $H_2O_2$ . A characteristic increase of cathodic current was observed over different concentrations of  $H_2O_2$  at −0.45 V vs Ag/AgCl (sat. KCl). The cathodic current increases due to the possible oxygen reduction reaction occurring at the electrode surface. The result is depicted in [Figure S5](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information). The possible electrode reaction at the electrode surface is depicted in the following equation (eq 5)

$$
GO_x - FAD + H_2O_2 \rightarrow GO_x - FADH_2 + O_2 \tag{5}
$$

We have also performed control experiment to understand the role of  $TiO<sub>2</sub>$  in this composite, and for this purpose, we synthesized the material without having  $TiO<sub>2</sub>$ . The synthesized  $GO_x@ZIF-8$  was drop-cast over GCE, and CV was taken in an air-saturated 10 mM glucose solution in pH 7.4 buffer and compared to the CV with  $GO_x \omega ZIF-8(TiO_2)$ . The result is depicted in [Figure S6.](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) The result clearly suggests that there is an active role of  $TiO<sub>2</sub>$  as a charge-storage tank as the control experiment shows an almost nonfaradic linear current−voltage response with mostly the migration of ions present in PBS. The as-synthesized probe is not much affected by ion migration and can store charges originated at the electrode surface due to the generation of  $H_2O_2$ , which is uptaken by TiO<sub>2</sub> and transferred to the electrode surface, resulting in the appearance of cathodic peak.

The bioelectrocatalytic activity of the synthesized probe is estimated by calculating electroactive protein density  $(\Gamma, \text{mol})$ cm<sup>2</sup>) using eq 6

$$
Q = nFA\Gamma \tag{6}
$$

$$
(6)
$$

where  $n$ , F, and A carry their usual meaning. To calculate  $\Gamma$ , CV was performed in 0.1 M pH 7.4 PBS at a scan rate of 100 mV/s.  $\Gamma$  is calculated to be 1.56  $\times$  10<sup>-10</sup> mol/cm<sup>2</sup>, which is quite superior to that of other  $TiO_2$ -based amperometric glucose sensors.

To estimate the steady-state response of the probe, the potentiostatic amperometric technique was performed. In this technique, the potential needs to be constant while an increment/decrement of current is observed with respect to time. For amperometric glucose measurement, an increment of current is measured with increasing concentration of glucose added at a constant rate. To proceed with the experiment, 0.1 mM glucose has been added successively into the air-saturated pH 7.4 buffer solution. The potential chosen for chronoamperometry was −0.45 V vs Ag/AgCl (sat. KCl), and the result is depicted in Figure 6. Dynamic stairs of current change were



Figure 6. Amperometric response of synthesized probe at −0.45 V vs Ag/AgCl (sat. KCl) by adding 0.1 mM glucose successively in airsaturated pH 7.4 PBS, resulting in a dynamic stair with quick steady state of current. Calibration plot is depicted in the inset.

observed with constant rise and saturation of current. A quick steady state upon successive addition of glucose (∼5 s) was observed, which depicts the efficacy of the probe toward glucose reduction reaction. % Relative standard deviation (RSD) was calculated to check the analytical performance of the synthesized probe, and 1.2% RSD was found for 20 successive dilutions, depicting the nature and stability of the synthesized probe. The steady-state current, obtained from the amperometric plot, was plotted with the change of concentration, and the result is depicted in the inset of Figure 6. A linear calibration plot was obtained, which depicts the linear proportionality of the obtained current with change of concentration. The first rise of current was designated as lower detection limit of this biosensor, and it was calculated at 80 nM glucose concentration. Dissolved oxygen plays an immense role in this experiment as the current achieved steady state gradually after 18 successive dilution. This is probably due to the increment of dissolved oxygen, which vigorously disturbs the  $FAD/FADH<sub>2</sub>$  equilibrium. This low-level detection of glucose can be strongly correlated with the measurement of blood glucose from sweat/saliva and can be an active component of noninvasive monitor of glucose.

To investigate the tandem catalytic activity of the probe, the same steady-state response for the reduction of  $H_2O_2$  was also investigated, and for this purpose, 0.1 mM  $H_2O_2$  was added successively to the air-saturated pH 7.4 buffer solution. The potential for the chronopotentiometry is also kept fixed, and it was chosen to be the same as we added for glucose monitoring: −0.45 V vs Ag/AgCl (sat. KCl). The current versus time plot was obtained and is depicted in [Figure S7](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf) (Supporting Information). The result shows a decrement of cathodic current at −0.45 V vs Ag/AgCl (sat. KCl), which shows the excellent tandem catalytic activity of the probe toward H<sub>2</sub>O<sub>2</sub>, having a quick steady state of ~7 s that was achieved successfully.

## 3. CONCLUSIONS

Herein, we report a protocol for successful encapsulation of  $TiO<sub>2</sub>$  and  $GO<sub>x</sub>$  inside ZIF-8 MOF to synthesize a new probe  $GO_x@ZIF-8(TiO_2)$ , applicable for mediator-free amperometric detection of glucose.  $GO_x$  and  $TiO_2$  are jointly encapsulated into ZIF-8 matrix in aqueous media to form the composite  $GO_x \omega ZIF-8(TiO_2)$ . The protocol supports gram-scale synthesis and shows commendable stability and good tandem catalytic activity. The as-synthesized composite was thoroughly characterized by various physicochemical methods. Thermal analysis depicts excellent stability of the probe, whereas X-ray diffraction analysis depicts the crystalline structure of the synthesized probe with  $TiO<sub>2</sub>$  nanoparticles in the anatase phase. The presence of  $TiO<sub>2</sub>$  nanoparticles increases not only the effective surface area but also the charge storage due to the presence of electrical double layer, which ultimately influences charge transfer at the electrode−electrolyte interface, resulting in excellent tandem catalytic activity toward both glucose oxidation and peroxide reduction reaction. Using this probe, a low-level detection of glucose can be achieved (80 nM) in aqueous media by using very low concentration of GO<sub>x</sub> (62  $\mu$ g in 1 mL), which can be attributed to the monitoring of blood sugar by noninvasive way, mainly from sweat. Overall, the synthetic probe can be demonstrated as competent, robust, and aqueous media-tolerable nanocomposite applicable as tandem catalyst for both glucose oxidation and peroxide reduction reaction.

#### 4. EXPERIMENTAL SECTION

4.1. Materials and Methods. Analytical grades of titanium oxide nanopowder  $(TiO<sub>2</sub>)$ , glucose oxidase from Aspergillus niger  $(GO_x)$ , and 2-methylimidazole were purchased from Sigma-Aldrich and used without further purification. Analytical-grade  $Zn(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O$  purchased from SRL Chemicals was also used without further purification. The synthesis of  $GO_x \omega ZIF-8(TiO_2)$  nanocomposite was done in aqueous media at room temperature.

The conventional glassy carbon electrode (GCE) of 5 mm diameter had been used as a working electrode for all electrochemical experiments performed. Prior to the experiment, alumina slurry of size 0.3  $\mu$  was put in a cloth of dense mesh and the electrode was polished gently, followed by sonication for 15 min in 1:1 ethanol−water mixture to get a clean GCE surface for drop-casting of the as-synthesized composite. The electrode was washed several times with ultrapure Milli-Q water and dried at room temperature prior to use for experiment. A conventional electrolytic cell setup consisting of three electrodes was used in all of the electrochemical experiments. Glassy carbon was utilized as the working electrode, Pt wire (0.1 mm diameter) was used as the counter electrode, and Ag/AgCl (sat. KCl) was used as the reference electrode. The  $GO_x$  slurry of  $GO_x \times ZIF-8(TiO_2)$ composite was prepared for electroanalysis. Material (1 mg) is <span id="page-5-0"></span>Scheme 1. Schematic Representation for Fabrication of  $GO<sub>x</sub>(@ZIF-8(TiO<sub>2</sub>))$ 



dispersed in 200  $\mu$ L of pH 7.4 buffer to make the slurry, followed by ultrasonication for 15 min. Furthermore, 5  $\mu$ L of 0.5% nafion is mixed with the slurry, which acts as a binder to make the material strongly attached to the GCE surface. All of the electrochemical experiments were performed in Metrohm Autolab 203 potentiostat/galvanostat.

The structure of the as-synthesized composite was characterized by various physicochemical techniques. PXRD (PAN Analytical Empyrean Series 2 X-ray diffraction system) was done to investigate the crystal structure of the synthesized probe. The morphology of the synthesized microstructured material was investigated by FE-SEM (JEOL JSM-7100F) and HR-TEM (JEOL JEM-2100). UV−vis spectra were recorded with Varian spectrophotometer. FT-IR experiment was performed using PerkinElmer G-FT-IR. TGA experiment was performed in Mettler Toledo instrument. BET N<sub>2</sub> adsorption− desorption isotherm was measured in Micromeritics, ASAP 2010. The detailed synthetic procedure for the fabrication of  $GO_x@ZIF-8(TiO_2)$  nanocomposite is described below. Metrohm Autolab 203 potentiostat/galvanostat is used for all electrochemical measurements.

4.2. Synthesis of  $GO_x@ZIF-8(TiO_2)$ . ZIF-8 was prepared by a previously reported method.<sup>[13](#page-6-0)</sup> Two distinct synthetic mixtures of 22.70 g of 2-methlyimidazole (108 mM) were dissolved in 400 mL of ultrapure Milli-Q water, and 50 mg of  $GO_x$  (8170 units in 50 mg) was added to the same solution, labeled (A). The resulting solution turned pale yellow. In another synthetic mixture, 1.17 g of  $\text{Zn}(\text{NO}_3)$ <sub>2</sub>·6H<sub>2</sub>O (1.53) mM) was dissolved in another 390 mL of ultrapure Milli-Q water separately and labeled  $(B)$ . TiO<sub>2</sub> nanopowder (5 mg) was taken in 1 mL of Milli-Q water separately (C) and sonicated for 30 min until a homogeneous dispersion formed. The as-prepared (B) and (C) solutions were added directly to the as-prepared mixture (A) under constant stirring of 700 rpm at room temperature, and the total reaction mixture was stirred for 3 h. It was observed that the solution turned hazy from transparent pale yellow, which is the indication for the encapsulation of both  $GO_x$  and  $TiO_2$  jointly inside ZIF-8 moiety. The probable haziness may have appeared due to precipitation of the as-synthesized probe. Then, the mixture was allowed to stand overnight in a 100 mL beaker to settle down the desired precipitate. The settled solid precipitate was slowly removed from the beaker using a micropipette and further centrifuged in 10 000 rpm for 10 min. The precipitate was collected and washed thoroughly with 50 mL of Milli-Q water three times to remove excess  $GO_x$  and  $TiO_2$ , which may have stuck to the surface of the synthesized MOF. Finally, the probe was put in a dessicator and dried under vacuum. An offwhite solid composite was recovered, and this is the first initial confirmation for the formation of  $GO_x \omega ZIF-8(TiO_2)$  nano-

composite. The synthetic route is schematically depicted in Scheme 1. The as-synthesized probe was further characterized by various physicochemical techniques like FT-IR, PXRD, FE-SEM, HR-TEM, and UV−vis spectroscopies to confirm the formation of the desired material.

## ■ ASSOCIATED CONTENT

## **6** Supporting Information

The Supporting Information is available free of charge on the [ACS Publications website](http://pubs.acs.org) at DOI: [10.1021/acsome](http://pubs.acs.org/doi/abs/10.1021/acsomega.8b01968)[ga.8b01968](http://pubs.acs.org/doi/abs/10.1021/acsomega.8b01968).

> STEM-EDX mapping of TiO<sub>2</sub> (red color building block) over ZIF-8 layers (green color building block) showing equal spatial distribution of  $TiO<sub>2</sub>$  (Figure S1); thermogravimetric analysis of  $GO_x \omega ZIF-8(TiO_2)$  showing distinct degradation region at different temperature (Figure S2) [\(PDF](http://pubs.acs.org/doi/suppl/10.1021/acsomega.8b01968/suppl_file/ao8b01968_si_001.pdf))

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#### Notes

The authors declare no competing financial interest.

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