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Sex Bias in Autoimmunity

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Summary

calculated based on prevalence data of a complex interplay of the sex chromosomes, sex hormones, the microbiota, and additional environmental and sociological factors.

Keywords

sex bias; autoimmunity; sex-dependent gene regulation; X chromosome; VGLL3

Introduction

The immune system functions to defend against infection. Responses must be robust and specific enough to ward off or overcome infection without causing undue harm to the organism. Autoimmune disease arises when an exaggerated or misdirected immune response damages native tissues or organs. While individual autoimmune diseases are rare, they are collectively among the most prevalent diseases in Western society (1). Despite intensive investigation, our understanding of the pathogenesis of autoimmune disease is incomplete. A growing body of evidence supports a model wherein environmental and lifestyle factors precipitate development of autoimmunity in genetically susceptible hosts. Cures have been elusive, and lifetime treatment is often required.

Cellular and humoral immunity are generally stronger in women; women have higher levels of circulating antibodies, more circulating CD4 T cells, more robust cytokine production in response to infection, and enhanced rejection of tumors and allografts (2). Many autoimmune diseases show a striking female sex bias (Figure 1) (3). Systemic lupus erythematosus (SLE), Sjogren's syndrome, Grave's disease, and Hashimoto's thyroiditis are seven to ten times more common in women than men; multiple sclerosis (MS), rheumatoid arthritis (RA), and scleroderma are two to three times more common (4). Overall, it is estimated that 78% of people affected with autoimmune diseases are women (5). For many diseases such as SLE, genome-wide association studies and meta-analyses have identified

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Purpose of review

To give an overview of recently published articles addressing the mechanisms underlying sex bias in autoimmune disease.

Conflicts of interest

The authors have no conflicts of interest.

protein UNC93B1 (28). Estrogens also function through ER α to downregulate the autoimmune regulator (AIRE), a critical factor in central tolerance, through promoter methylation(29); AIRE expression is also downregulated by progesterone and upregulated by the androgen dihydrotestosterone (29, 30). The effects of sex hormones on sex-biased DNA methylation and autoimmunity are otherwise as yet unclear (31). Additionally, estrogens regulate miRNAs (32), who in turn regulate estrogen-dependent signaling (33). Estrogens increase neutrophil numbers (34, 35) but overall inhibit their activation and trafficking via multiple mechanisms, some mediated by NF- κ B inhibition (reviewed in (36)). Estrogens enhance NK cell cytotoxicity and IFN- γ production (37) but downregulate NK cell granzyme B secretion and cell surface activation markers (38). Effects of estrogen on monocytes and macrophages vary by dose: production of pro-inflammatory cytokines is enhanced at low concentrations and suppressed at high (39). The response of dendritic cells to estrogens is mixed, inducing production of both anti-inflammatory and Th1-type pro-inflammatory cytokines (40–42). At lower concentrations, estrogens have immunostimulatory effects, promoting a Th1 response through enhancing the secretion of IFN- γ (27, 43, 44); in contrast, at high concentrations, estrogens promote a Th2 response (45–47). In pregnant SLE patients, this Th2 shift and consequent increased production of autoantibodies often exacerbates disease (48). Effects on Th17 cells are less clear (36). Treg cells increase with estrogens (49, 50). Estrogens promote B cell survival, maturation, class switching, and antibody production (51–53) and interfere with peripheral negative selection of autoreactive B cells (54, 55). Oophorectomy has been reported to be protective in lupus-prone mice (56); however, recent data suggest that this protection may not rely solely on estrogen-mediated effects, as complete absence of ER α does not delay lupus in prone mice (57).

Androgens predominantly downregulate the immune response (58), decreasing pro-inflammatory mediators and inhibiting the proliferation and activation of a number of immune cell populations (reviewed in (36)). Accordingly, androgens have been shown to exert a protective effect in multiple autoimmune rodent models (30, 59, 60). This appears to be mediated in part by androgen-induced upregulation of AIRE in the male thymus: androgens increase AIRE levels by binding the androgen receptor (AR) and targeting the *AIRE* promoter, and the protective effects of androgens and male sex are lost in AIRE-deficient mice in a model of experimental autoimmune encephalitis (30). Additionally, ARs are broadly expressed in neutrophil-lineage cells, with no difference in male and female expression patterns (61), and act to increase the number and trafficking of neutrophils (62); however, androgens decrease pro-inflammatory responses of neutrophils (63), natural killer (NK) cells (64), and macrophages (65). ARs are not expressed in peripheral lymphocytes but are found in lymphoid and non-lymphoid thymic and bone marrow cells (66), where they limit the number of immature thymocytes and restrain active cell cycling (67). While androgens decrease the activation of Th1 and Th2 cells (68), Th17 cell responses are enhanced (69). Treatment to reduce testosterone decreases Treg count (70). Androgens have also been found to limit the number of immature type 2 innate lymphoid cells in the lung (71). In epidermal cells, androgens were found to modulate the expression of *PRDMI*, a transcriptional repressor involved in thymic T cell apoptosis and other immune cell

processes that is implicated in female-dependent autoimmune risk (72). B cell number is negatively regulated by androgens (62).

Some studies support a role for psychosocial stress in initiation or exacerbation of autoimmune disease, although causation has been challenging to establish (73). Response to stress occurs through the hypothalamus-pituitary-adrenal (HPA) axis, which also exhibits sexual dimorphism in cortisol response to psychosocial stressors (reviewed in (31)). Stress triggers release of glucocorticoids, which generally inhibit immune responses through decreasing production of pro-inflammatory cytokines and inhibiting activation and proliferation of multiple immune cell types. The HPA axis also produces prolactin, another hormone with immunological effects. The prolactin receptor is widely expressed in immune cells (74), and prolactin signaling is largely immunostimulatory (75). In particular, prolactin may promote autoimmunity by inhibiting negative selection of autoreactive B cells, augmenting autoantibody production (76–78). Positive correlations of prolactin levels and disease activity have been identified in SLE patients, but the causality remains to be determined (79).

Sex chromosomes

The presence of two X chromosomes in the female also contributes to sex bias of autoimmunity. While canonically one of the X chromosomes is inactivated in early development, this process is imperfect, with approximately 15% of genes escaping X chromosome inactivation (XCI) (80). A majority of the genes that escape XCI show female expression bias (81–83), and there is variation among individuals in which genes escape (84). Males with Klinefelter syndrome (karyotype XXY) have an increased risk of SLE commensurate with that of females (85), and one male patient with severe pre-pubertal SLE was found to have an XX karyotype due to an X-Y translocation (86), demonstrating the influence of X dosage. In contrast, females with Turner syndrome, who have complete (XO) or partial X chromosome monosomy, are at increased risk of developing autoimmune disease relative even to XX females, but the excess risk may be greater for male-predominant autoimmune conditions such as ankylosing spondylitis (87, 88). In contrast, females with Turner syndrome (karyotype XO) are at increased risk of developing autoimmune disease, but the risk is strongest for male-predominant conditions (87). In most females, XCI is random, resulting in half of cells expressing the maternal and half the paternal X chromosome; however, some females show non-random silencing leading to an 80% or more predominance of one X chromosome. This skewed inactivation is more common in patients with autoimmune diseases (89, 90), although it may be a consequence of autoimmunity rather than a cause (91).

Many genes with established roles in autoimmunity are located on the X chromosome. Several of these have been found to be overexpressed or hypomethylated in female but not male SLE patients (92, 93), and dosage of the X-linked *TLR7* and *TLR8* genes has been shown to influence development of SLE in humans and lupus-prone mice (94–98). Recently, escape of *TLR7* inactivation in a substantial number proportion of immune cells has been described in females and men with Klinefelter syndrome; this biallelic expression of *TLR7* primes for increased class switching in activated B cells and increased *TLR7* reactivity (99).

The X chromosome is also highly enriched in microRNAs (miRNAs) (100). MicroRNAs, including some located on the X chromosome, are essential for maintenance of immune tolerance (reviewed in (101)), and a subset of X-linked miRNAs was found to be overexpressed in females, but not males, with SLE (93). Finally, the X chromosome can become partially reactivated in lymphocytes in women, resulting in overexpression of immunity genes and possibly contributing to sex bias in SLE (102).

The Y chromosome has garnered much less attention as a driver of sex bias in autoimmunity, but evidence is accumulating. In a mouse model of autoimmune disease, Y chromosome polymorphisms, including gene copy number variation, correlate with disease susceptibility and severity (103–105), although the observed effects may also reflect impaired balancer function in mismatched X and Y chromosomes that evolved in different strains (106). However, data from men with MS suggest the influence of the Y chromosome on autoimmunity may extend to humans (105). Further investigation of male mice with specific Y-linked defects in immunity (107, 108) and examples of human Y-linked immune variation (109) is ongoing and may shed additional light.

Gut immunology and the microbiota

The gut microbiota plays a critical role in maturation and modulation of innate and adaptive immunity (110) yet is itself shaped by the immune system. Both the gut immune system and microbiota exhibit sexual dimorphism. Immune tissues in the gut of male and female rodents differ in representation of immune cells, with an overall trend toward enhanced innate immunity and attenuated adaptive immunity in the male gut relative to the female (111, 112), and many immune genes show sex-biased expression in mouse gut (113, 114). Although some studies have observed no differences in the diversity or composition of male and female gut microbiota, sex differences in the human gut microbiota have been extensively documented (115), raising interest in the gut microbiota as a potential driver of sex bias in autoimmune disease. Additionally, microbiome aberrations have been observed in the vast majority of immune-mediated diseases, although demonstrating causation has proven a significant obstacle thus far (116), with many resorting to animal models for further investigation.

Female mice show increased microbiota diversity relative to male, and many bacterial species show sex-biased enrichment that occasionally varies with strain, age, and diet (reviewed in (115)). Sex hormones likely also play a role. In human twin studies, the microbiota of opposite-sex twins becomes more divergent after puberty relative to same-sex twins (117). Similarly, in the nonobese diabetic (NOD) mouse model of spontaneous autoimmune type I diabetes (T1D), the gut microbiota does not differ in prepubertal male and female mice; however, microbial diversity decreases in intact postpubertal males, while this does not occur in females and castrated males (118). Transfer of male microbiota into germ-free female mice and female microbiota into germ-free male mice revealed that some manifestations of immunological sexual dimorphism appear to depend on sex-specific gut microbiota: regardless of the sex of the recipient, $ROR\gamma^+Foxp3^+$ cells are increased in gut immune tissues of mice who received male microbiota, and T cell precursors are increased in mice who received female microbiota (113).

There is mounting evidence of a direct role for the gut microbiota in driving sex-biased autoimmunity. Female NOD mice develop spontaneous autoimmune T1D at twice the rate of male mice. Under germ-free conditions, however, incidence of T1D is equal in both sexes (119), indicating that male microbiota may confer protection. Gavage of female NOD weanlings with male NOD intestinal microbiota results in elevated serum testosterone and protects against development of T1D. This effect is abrogated in recipient female mice treated with androgen receptor antagonist, suggesting protection is conferred by a testosterone-dependent mechanism (119). Female MRL/Mp-Fas^{lpr} mice, who develop lupus at far higher rates than males, show significantly higher gut microbiota diversity but lower abundance of *Lactobacillales* species and increased intestinal permeability (120). In female and castrated male Fas^{lpr} mice, *Lactobacillales* gavage restores gut mucosal barrier function, promoting an anti-inflammatory cytokine environment in which autoantibody production decreases and renal disease improves, with increased renal Treg cells and suppression of renal Th17 cells (120). The same benefits were not observed in intact male Fas^{lpr} mice, suggesting that *Lactobacillus* species in the gut attenuate renal disease in lupus-prone mice in a sex hormone-dependent manner. In aggregate, the animal data suggest that sex and androgens appear to regulate gut microbiota composition and function, which reciprocally influences the immune response and development of autoimmunity.

Other factors

Recently, we identified the transcription factor VGLL3 to be critical in orchestrating sex-biased expression of key autoimmune genes in a sex hormone-independent fashion (83). VGLL3 is required for robust expression of genes implicated in autoimmunity and for mounting a full IFN-I response. In healthy skin, VGLL3 shows nuclear localization that is more prominent in women than in men. In lesional skin of patients with cutaneous lupus, however, VGLL3 shows nuclear localization in both sexes, indicating disease-dependent regulation. This suggests VGLL3 governs an autoimmunity pathway that is constitutively active in women but must be triggered by other means in men (83). VGLL3 is located on chromosome 3 and appears to be epigenetically regulated (unpublished observation), and its exact role in driving autoimmune diseases is being actively explored.

Sex bias is prominent in DNA methylation, affecting chromatin accessibility of immune genes (121). The X and Y chromosome may influence DNA methylation, as was shown for an autosomal locus in human cells (122). Prenatal environmental exposures also show sex-specific epigenetic effects on DNA methylation (reviewed in (123)), although the underlying mechanisms remain unknown. Fetal microchimerism, wherein circulating fetal cells travel to the mother and persist for years after pregnancy (124), may predispose to development of autoimmune disease; however, studies disagree on whether autoimmune diseases are more common in women with prior pregnancies, and a definitive connection has not been established (125). Additionally, men and women differ in exposure to environmental endocrine-disrupting chemicals, with estrogenic and anti-estrogenic properties that may affect genetic and epigenetic regulation of immunity (reviewed in (126, 127)). Finally, sociological differences between the sexes, such as rates of smoking in men versus women, may influence the development and manifestations of autoimmune disease (128).

Conclusion

In treating severe autoimmune disease, physicians often must turn to broad immunosuppressive therapies with high side effect burden and inherent risks of infection and malignancy due to decreased immune surveillance activity. The trend toward stratifying immunological research studies by gender and continuous improvements in high-throughput technologies have enabled increasingly sophisticated characterization of the differences between the male and female autoimmune phenotypes; this is evidenced by work such as the recent description of sex bias in the influence of HLA associations on T cell selection and expansion as revealed by TCR immunosequencing of large cohorts (129). These nuanced descriptions are helping to explain observed sex differences in infection susceptibility and autoimmunity, but we must continue to interrogate the mechanistic underpinnings of sex-biased autoimmunity to pave the way toward development of highly specific therapeutics that spare patients the dangers of broad immunosuppression. Targeting the precise pathways that drive the female autoimmune disease burden above the baseline male prevalence will be an immense boon to human welfare, particularly if accompanied only by the relatively modest increased risk of infection and malignancy native to the male immune system.

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Recent findings

Recent studies investigating the origins of sex bias in autoimmune disease have revealed an extensive and interconnected network of genetic, hormonal, microbial, and environmental influences. Investigation of sex hormones has moved beyond profiling the effects of hormones on activity and prevalence of immune cell types to defining the specific immunity-related genes driving these changes. Deeper examination of the genetic content of the X and Y chromosomes and genetic escapees of X chromosome interaction has revealed some key drivers of female-biased autoimmunity. Animal studies are offering further insights into the connections between microbiota, particularly that of the gut, and the immune system.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
1. El-Gabalawy H, Guenther LC, Bernstein CN. Epidemiology of immune-mediated inflammatory diseases: incidence, prevalence, natural history, and comorbidities. *J Rheumatol Suppl.* 2010;85:2–10. [PubMed: 20436161]
 2. Nalbandian G, Kovats S. Understanding sex biases in immunity: effects of estrogen on the differentiation and function of antigen-presenting cells. *Immunol Res.* 2005;31(2):91–106. [PubMed: 15778508]
 3. Whitacre CC. Sex differences in autoimmune disease. *Nat Immunol.* 2001;2(9):777–80. [PubMed: 11526384]

4. Fish EN. The X-files in immunity: sex-based differences predispose immune responses. *Nat Rev Immunol.* 2008;8(9):737–44. [PubMed: 18728636]
5. Jacobson DL, Gange SJ, Rose NR, Graham NM. Epidemiology and estimated population burden of selected autoimmune diseases in the United States. *Clin Immunol Immunopathol.* 1997;84(3):223–43. [PubMed: 9281381]
6. Lim SS, Bayakly AR, Helmick CG, Gordon C, Easley KA, Drenkard C. The incidence and prevalence of systemic lupus erythematosus, 2002–2004: The Georgia Lupus Registry. *Arthritis Rheumatol.* 2014;66(2):357–68. [PubMed: 24504808]
7. Bentham J, Morris DL, Graham DSC, Pinder CL, Tomblinson P, Behrens TW, et al. Genetic association analyses implicate aberrant regulation of innate and adaptive immunity genes in the pathogenesis of systemic lupus erythematosus. *Nat Genet.* 2015;47(12):1457–64. [PubMed: 26502338]
8. Heldring N, Pike A, Andersson S, Matthews J, Cheng G, Hartman J, et al. Estrogen receptors: how do they signal and what are their targets. *Physiol Rev.* 2007;87(3):905–31. [PubMed: 17615392]
9. Hewitt SC, Winuthayanon W, Korach KS. What’s new in estrogen receptor action in the female reproductive tract. *J Mol Endocrinol.* 2016;56(2):R55–71. [PubMed: 26826253]
10. Chang CS, Kokontis J, Liao ST. Molecular cloning of human and rat complementary DNA encoding androgen receptors. *Science.* 1988;240(4850):324–6. [PubMed: 3353726]
11. Hannah MF, Bajic VB, Klein SL. Sex differences in the recognition of and innate antiviral responses to Seoul virus in Norway rats. *Brain Behav Immun.* 2008;22(4):503–16. [PubMed: 18053684]
12. Gubbels Bupp MR, Jorgensen TN. Androgen-Induced Immunosuppression. *Front Immunol.* 2018;9:794. [PubMed: 29755457]
13. Lateef A, Petri M. Hormone replacement and contraceptive therapy in autoimmune diseases. *J Autoimmun.* 2012;38(2–3):J170–6. [PubMed: 22261500]
14. Zarkavelis G, Kollas A, Kampletsas E, Vasiliou V, Kaltsonoudis E, Drosos A, et al. Aromatase inhibitors induced autoimmune disorders in patients with breast cancer: A review. *Journal of advanced research.* 2016;7(5):719–26. [PubMed: 28275510]
15. Lee JH, Lydon JP, Kim CH. Progesterone suppresses the mTOR pathway and promotes generation of induced regulatory T cells with increased stability. *Eur J Immunol.* 2012;42(10):2683–96. [PubMed: 22740122]
16. Klein SL, Flanagan KL. Sex differences in immune responses. *Nat Rev Immunol.* 2016;16(10):626–38. [PubMed: 27546235]
17. Teilmann SC, Clement CA, Thorup J, Byskov AG, Christensen ST. Expression and localization of the progesterone receptor in mouse and human reproductive organs. *J Endocrinol.* 2006;191(3):525–35. [PubMed: 17170211]
18. Druckmann R, Druckmann MA. Progesterone and the immunology of pregnancy. *J Steroid Biochem Mol Biol.* 2005;97(5):389–96. [PubMed: 16198558]
19. Arruvito L, Giulianelli S, Flores AC, Paladino N, Barboza M, Lanari C, et al. NK cells expressing a progesterone receptor are susceptible to progesterone-induced apoptosis. *J Immunol.* 2008;180(8):5746–53. [PubMed: 18390760]
20. Butts CL, Shukair SA, Duncan KM, Bowers E, Horn C, Belyavskaya E, et al. Progesterone inhibits mature rat dendritic cells in a receptor-mediated fashion. *Int Immunol.* 2007;19(3):287–96. [PubMed: 17289656]
21. Jones LA, Kreem S, Shweash M, Paul A, Alexander J, Roberts CW. Differential modulation of TLR3- and TLR4-mediated dendritic cell maturation and function by progesterone. *J Immunol.* 2010;185(8):4525–34. [PubMed: 20844199]
22. Piccinni MP, Giudizi MG, Biagiotti R, Beloni L, Giannarini L, Sampognaro S, et al. Progesterone favors the development of human T helper cells producing Th2-type cytokines and promotes both IL-4 production and membrane CD30 expression in established Th1 cell clones. *J Immunol.* 1995;155(1):128–33. [PubMed: 7541410]
23. Hardy DB, Janowski BA, Corey DR, Mendelson CR. Progesterone receptor plays a major antiinflammatory role in human myometrial cells by antagonism of nuclear factor-kappaB

activation of cyclooxygenase 2 expression. *Mol Endocrinol*. 2006;20(11):2724–33. [PubMed: 16772530]

24. Phiel KL, Henderson RA, Adelman SJ, Elloso MM. Differential estrogen receptor gene expression in human peripheral blood mononuclear cell populations. *Immunol Lett*. 2005;97(1):107–13. [PubMed: 15626482]
25. Kovats S Estrogen receptors regulate innate immune cells and signaling pathways. *Cell Immunol*. 2015;294(2):63–9. [PubMed: 25682174]
26. Shen H, Panchanathan R, Rajavelu P, Duan X, Gould KA, Choubey D. Gender-dependent expression of murine *Irf5* gene: implications for sex bias in autoimmunity. *J Mol Cell Biol*. 2010;2(5):284–90. [PubMed: 20802013]
27. Fox HS, Bond BL, Parslow TG. Estrogen regulates the IFN-gamma promoter. *J Immunol*. 1991;146(12):4362–7. [PubMed: 1904081]
28. Panchanathan R, Liu H, Choubey D. Expression of murine *Unc93b1* is up-regulated by interferon and estrogen signaling: implications for sex bias in the development of autoimmunity. *Int Immunol*. 2013;25(9):521–9. [PubMed: 23728775]
- 29 • Dragin N, Bismuth J, Cizeron-Clairac G, Biferi MG, Berthault C, Serraf A, et al. Estrogen-mediated downregulation of AIRE influences sexual dimorphism in autoimmune diseases. *J Clin Invest*. 2016;126(4):1525–37.
 Estrogen downregulates expression of AIRE in the thymus of female humans and mice by inducing epigenetic changes at the AIRE promoter through ER α receptors. In a mouse model of experimental autoimmune thyroiditis, the protective effects of male sex are abrogated by downregulation of Aire, suggesting that estrogen-mediated regulation of AIRE likely contributes to female-biased autoimmunity.
 [PubMed: 26999605]
- 30 • Zhu ML, Bakhru P, Conley B, Nelson JS, Free M, Martin A, et al. Sex bias in CNS autoimmune disease mediated by androgen control of autoimmune regulator. *Nat Commun*. 2016;7:11350
 Androgens bind AR and target the AIRE promoter to drive higher AIRE expression in the thymus of male humans and mice. In a mouse model of experimental autoimmune encephalitis, androgens and male sex are protective against disease in an Aire-dependent manner, suggesting that androgen-mediated regulation of AIRE likely contributes to female-biased autoimmunity.
 [PubMed: 27072778]
31. Edwards M, Dai R, Ahmed SA. Our Environment Shapes Us: The Importance of Environment and Sex Differences in Regulation of Autoantibody Production. *Front Immunol*. 2018;9:478. [PubMed: 29662485]
32. Klinge CM. Estrogen Regulation of MicroRNA Expression. *Curr Genomics*. 2009;10(3):169–83. [PubMed: 19881910]
33. Khan D, Dai R, Ansar Ahmed S. Sex differences and estrogen regulation of miRNAs in lupus, a prototypical autoimmune disease. *Cell Immunol*. 2015;294(2):70–9. [PubMed: 25619140]
34. Jilma B, Eichler HG, Breiteneder H, Wolzt M, Aringer M, Graninger W, et al. Effects of 17 beta-estradiol on circulating adhesion molecules. *J Clin Endocrinol Metab*. 1994;79(6):1619–24. [PubMed: 7527406]
35. Robinson DP, Hall OJ, Nilles TL, Bream JH, Klein SL. 17beta-estradiol protects females against influenza by recruiting neutrophils and increasing virus-specific CD8 T cell responses in the lungs. *J Virol*. 2014;88(9):4711–20. [PubMed: 24522912]
36. Bereshchenko O, Bruscoli S, Riccardi C. Glucocorticoids, Sex Hormones, and Immunity. *Front Immunol*. 2018;9:1332. [PubMed: 29946321]
37. Nakaya M, Tachibana H, Yamada K. Effect of estrogens on the interferon-gamma producing cell population of mouse splenocytes. *Biosci Biotechnol Biochem*. 2006;70(1):47–53. [PubMed: 16428820]

38. Hao S, Zhao J, Zhou J, Zhao S, Hu Y, Hou Y. Modulation of 17beta-estradiol on the number and cytotoxicity of NK cells in vivo related to MCM and activating receptors. *Int Immunopharmacol.* 2007;7(13):1765–75. [PubMed: 17996687]
39. Karpuzoglu E, Phillips RA, Gogal RM, Jr., Ansar Ahmed S. IFN-gamma-inducing transcription factor, T-bet is upregulated by estrogen in murine splenocytes: role of IL-27 but not IL-12. *Mol Immunol.* 2007;44(7):1808–14. [PubMed: 17046061]
40. Liu HY, Buenafe AC, Matejuk A, Ito A, Zamora A, Dwyer J, et al. Estrogen inhibition of EAE involves effects on dendritic cell function. *J Neurosci Res.* 2002;70(2):238–48. [PubMed: 12271473]
41. Delpy L, Douin-Echinard V, Garidou L, Bruand C, Saoudi A, Guery JC. Estrogen enhances susceptibility to experimental autoimmune myasthenia gravis by promoting type 1-polarized immune responses. *J Immunol.* 2005;175(8):5050–7. [PubMed: 16210608]
42. Subramanian S, Yates M, Vandebark AA, Offner H. Oestrogen-mediated protection of experimental autoimmune encephalomyelitis in the absence of Foxp3+ regulatory T cells implicates compensatory pathways including regulatory B cells. *Immunology.* 2011;132(3):340–7. [PubMed: 21091909]
43. Grasso G, Muscettola M. The influence of beta-estradiol and progesterone on interferon gamma production in vitro. *Int J Neurosci.* 1990;51(3–4):315–7. [PubMed: 2126259]
44. Karpuzoglu-Sahin E, Hissong BD, Ansar Ahmed S. Interferon-gamma levels are upregulated by 17-beta-estradiol and diethylstilbestrol. *J Reprod Immunol.* 2001;52(1–2):113–27. [PubMed: 11600182]
45. Marzi M, Vigano A, Trabattoni D, Villa ML, Salvaggio A, Clerici E, et al. Characterization of type 1 and type 2 cytokine production profile in physiologic and pathologic human pregnancy. *Clin Exp Immunol.* 1996;106(1):127–33. [PubMed: 8870710]
46. Matalka KZ. The effect of estradiol, but not progesterone, on the production of cytokines in stimulated whole blood, is concentration-dependent. *Neuro Endocrinol Lett.* 2003;24(3–4):185–91. [PubMed: 14523355]
47. Sabahi F, Rola-Pleszczynski M, O'Connell S, Frenkel LD. Qualitative and quantitative analysis of T lymphocytes during normal human pregnancy. *Am J Reprod Immunol.* 1995;33(5):381–93. [PubMed: 7576120]
48. Eudy AM, Siega-Riz AM, Engel SM, Franceschini N, Howard AG, Clowse MEB, et al. Effect of pregnancy on disease flares in patients with systemic lupus erythematosus. *Ann Rheum Dis.* 2018;77(6):855–60. [PubMed: 29463519]
49. Polanczyk MJ, Hopke C, Huan J, Vandebark AA, Offner H. Enhanced FoxP3 expression and Treg cell function in pregnant and estrogen-treated mice. *J Neuroimmunol.* 2005;170(1–2):85–92. [PubMed: 16253347]
50. Tai P, Wang J, Jin H, Song X, Yan J, Kang Y, et al. Induction of regulatory T cells by physiological level estrogen. *J Cell Physiol.* 2008;214(2):456–64. [PubMed: 17654501]
51. Asaba J, Bandyopadhyay M, Kindy M, Dasgupta S. Estrogen receptor signal in regulation of B cell activation during diverse immune responses. *Int J Biochem Cell Biol.* 2015;68:42–7. [PubMed: 26299327]
52. Straub RH. The complex role of estrogens in inflammation. *Endocr Rev.* 2007;28(5):521–74. [PubMed: 17640948]
53. Herblot S, Aplan PD, Hoang T. Gradient of E2A activity in B-cell development. *Mol Cell Biol.* 2002;22(3):886–900. [PubMed: 11784864]
54. Bynoe MS, Grimaldi CM, Diamond B. Estrogen up-regulates Bcl-2 and blocks tolerance induction of naive B cells. *Proc Natl Acad Sci U S A.* 2000;97(6):2703–8. [PubMed: 10694576]
55. Grimaldi CM, Michael DJ, Diamond B. Cutting edge: expansion and activation of a population of autoreactive marginal zone B cells in a model of estrogen-induced lupus. *J Immunol.* 2001;167(4):1886–90. [PubMed: 11489967]
56. Cunningham MA, Wirth JR, Scott JL, Eudaly J, Collins EL, Gilkeson GS. Early Ovariectomy Results in Reduced Numbers of CD11c+/CD11b+ Spleen Cells and Impacts Disease Expression in Murine Lupus. *Front Immunol.* 2016;7:31. [PubMed: 26913030]

- 57 • Scott JL, Wirth JR, Eudaly J, Ruiz P, Cunningham MA. Complete knockout of estrogen receptor alpha is not directly protective in murine lupus. *Clin Immunol.* 2017;183:132–41. Lupus prone mice with complete absence of ER α are protected from lupus disease expression only if their ovaries are intact, suggesting another protective factor, such as hypogonadism with resultant androgen elevation, may be involved.

[PubMed: 28822833]

58. Foo YZ, Nakagawa S, Rhodes G, Simmons LW. The effects of sex hormones on immune function: a meta-analysis. *Biol Rev Camb Philos Soc.* 2017;92(1):551–71. [PubMed: 26800512]
59. Roubinian JR, Talal N, Greenspan JS, Goodman JR, Siiteri PK. Effect of castration and sex hormone treatment on survival, anti-nucleic acid antibodies, and glomerulonephritis in NZB/NZW F1 mice. *J Exp Med.* 1978;147(6):1568–83. [PubMed: 308087]
60. Ahmed SA, Penhale WJ. The influence of testosterone on the development of autoimmune thyroiditis in thymectomized and irradiated rats. *Clin Exp Immunol.* 1982;48(2):367–74. [PubMed: 7049452]
61. Mantalaris A, Panoskaltis N, Sakai Y, Bourne P, Chang C, Messing EM, et al. Localization of androgen receptor expression in human bone marrow. *J Pathol.* 2001;193(3):361–6. [PubMed: 11241417]
62. Lai JJ, Lai KP, Zeng W, Chuang KH, Altuwaijri S, Chang C. Androgen receptor influences on body defense system via modulation of innate and adaptive immune systems: lessons from conditional AR knockout mice. *Am J Pathol.* 2012;181(5):1504–12. [PubMed: 22959669]
63. Pergola C, Dodt G, Rossi A, Neunhoeffler E, Lawrenz B, Northoff H, et al. ERK-mediated regulation of leukotriene biosynthesis by androgens: a molecular basis for gender differences in inflammation and asthma. *Proc Natl Acad Sci U S A.* 2008;105(50):19881–6. [PubMed: 19064924]
64. Hou J, Zheng WF. Effect of sex hormones on NK and ADCC activity of mice. *Int J Immunopharmacol.* 1988;10(1):15–22. [PubMed: 3366506]
65. D'Agostino P, Milano S, Barbera C, Di Bella G, La Rosa M, Ferlazzo V, et al. Sex hormones modulate inflammatory mediators produced by macrophages. *Ann N Y Acad Sci.* 1999;876:426–9. [PubMed: 10415638]
66. Olsen NJ, Kovacs WJ. Effects of androgens on T and B lymphocyte development. *Immunol Res.* 2001;23(2–3):281–8. [PubMed: 11444393]
67. Olsen NJ, Viselli SM, Shults K, Stelzer G, Kovacs WJ. Induction of immature thymocyte proliferation after castration of normal male mice. *Endocrinology.* 1994;134(1):107–13. [PubMed: 8275924]
68. Liva SM, Voskuhl RR. Testosterone acts directly on CD4+ T lymphocytes to increase IL-10 production. *J Immunol.* 2001;167(4):2060–7. [PubMed: 11489988]
69. Zhang MA, Rego D, Moshkova M, Kebir H, Chruscinski A, Nguyen H, et al. Peroxisome proliferator-activated receptor (PPAR)alpha and -gamma regulate IFN γ and IL-17A production by human T cells in a sex-specific way. *Proc Natl Acad Sci U S A.* 2012;109(24):9505–10. [PubMed: 22647601]
70. Page ST, Plymate SR, Bremner WJ, Matsumoto AM, Hess DL, Lin DW, et al. Effect of medical castration on CD4+ CD25+ T cells, CD8+ T cell IFN-gamma expression, and NK cells: a physiological role for testosterone and/or its metabolites. *Am J Physiol Endocrinol Metab.* 2006;290(5):E856–63. [PubMed: 16352669]
71. Laffont S, Blanquart E, Savignac M, Cenac C, Laverny G, Metzger D, et al. Androgen signaling negatively controls group 2 innate lymphoid cells. *J Exp Med.* 2017;214(6):1581–92. [PubMed: 28484078]
72. Kretzschmar K, Cottle DL, Donati G, Chiang MF, Quist SR, Gollnick HP, et al. BLIMP1 is required for postnatal epidermal homeostasis but does not define a sebaceous gland progenitor under steady-state conditions. *Stem Cell Reports.* 2014;3(4):620–33. [PubMed: 25358790]
73. Porcelli B, Pozza A, Bizzaro N, Fagiolini A, Costantini MC, Terzuoli L, et al. Association between stressful life events and autoimmune diseases: A systematic review and meta-analysis of retrospective case-control studies. *Autoimmun Rev.* 2016;15(4):325–34. [PubMed: 26708168]

74. Orbach H, Zandman-Goddard G, Amital H, Barak V, Szekanez Z, Szucs G, et al. Novel biomarkers in autoimmune diseases: prolactin, ferritin, vitamin D, and TPA levels in autoimmune diseases. *Ann N Y Acad Sci.* 2007;1109:385–400. [PubMed: 17785327]
75. Borba VV, Zandman-Goddard G, Shoenfeld Y. Prolactin and Autoimmunity. *Front Immunol.* 2018;9:73. [PubMed: 29483903]
76. Buckley AR. Prolactin, a lymphocyte growth and survival factor. *Lupus.* 2001;10(10):684–90. [PubMed: 11721694]
77. Kochendoerfer SK, Krishnan N, Buckley DJ, Buckley AR. Prolactin regulation of Bcl-2 family members: increased expression of bcl-xL but not mcl-1 or bad in Nb2-T cells. *J Endocrinol.* 2003;178(2):265–73. [PubMed: 12904174]
78. Saha S, Gonzalez J, Rosenfeld G, Keiser H, Peeva E. Prolactin alters the mechanisms of B cell tolerance induction. *Arthritis Rheum.* 2009;60(6):1743–52. [PubMed: 19479826]
79. Song GG, Lee YH. Circulating prolactin level in systemic lupus erythematosus and its correlation with disease activity: a meta-analysis. *Lupus.* 2017;26(12):1260–8. [PubMed: 28420051]
80. Carrel L, Willard HF. X-inactivation profile reveals extensive variability in X-linked gene expression in females. *Nature.* 2005;434(7031):400–4. [PubMed: 15772666]
81. Cotton AM, Ge B, Light N, Adoue V, Pastinen T, Brown CJ. Analysis of expressed SNPs identifies variable extents of expression from the human inactive X chromosome. *Genome Biol.* 2013;14(11):R122. [PubMed: 24176135]
82. Shen JJ, Wang TY, Yang W. Regulatory and evolutionary signatures of sex-biased genes on both the X chromosome and the autosomes. *Biol Sex Differ.* 2017;8(1):35. [PubMed: 29096703]
- 83 • Liang Y, Tsoi LC, Xing X, Beamer MA, Swindell WR, Sarkar MK, et al. A gene network regulated by the transcription factor VGLL3 as a promoter of sex-biased autoimmune diseases. *Nat Immunol.* 2017;18(2):152–60.
- The transcription factor VGLL3 is upregulated in women and functions independently of sex hormones to regulate a novel inflammatory pathway promoting female-biased autoimmunity. In male skin affected by lupus, this pathway appears to be activated, suggesting that VGLL3 contributes to female sex bias in autoimmunity but is also relevant to autoimmune disease in men.
- [PubMed: 27992404]
84. Zhang Y, Castillo-Morales A, Jiang M, Zhu Y, Hu L, Urrutia AO, et al. Genes that escape X-inactivation in humans have high intraspecific variability in expression, are associated with mental impairment but are not slow evolving. *Mol Biol Evol.* 2013;30(12):2588–601. [PubMed: 24023392]
85. Scofield RH, Bruner GR, Namjou B, Kimberly RP, Ramsey-Goldman R, Petri M, et al. Klinefelter's syndrome (47,XXY) in male systemic lupus erythematosus patients: support for the notion of a gene-dose effect from the X chromosome. *Arthritis Rheum.* 2008;58(8):2511–7. [PubMed: 18668569]
86. Chagnon P, Schneider R, Hebert J, Fortin PR, Provost S, Belisle C, et al. Identification and characterization of an Xp22.33;Yp11.2 translocation causing a triplication of several genes of the pseudoautosomal region 1 in an XX male patient with severe systemic lupus erythematosus. *Arthritis Rheum.* 2006;54(4):1270–8. [PubMed: 16575839]
87. Jorgensen KT, Rostgaard K, Bache I, Biggar RJ, Nielsen NM, Tommerup N, et al. Autoimmune diseases in women with Turner's syndrome. *Arthritis Rheum.* 2010;62(3):658–66. [PubMed: 20187158]
88. Goldacre MJ, Seminog OO. Turner syndrome and autoimmune diseases: record-linkage study. *Arch Dis Child.* 2014;99(1):71–3. [PubMed: 24064113]
89. Uz E, Mustafa C, Topaloglu R, Bilginer Y, Dursun A, Kasapcopur O, et al. Increased frequency of extremely skewed X chromosome inactivation in juvenile idiopathic arthritis. *Arthritis Rheum.* 2009;60(11):3410–2. [PubMed: 19877028]
90. Rubtsova K, Marrack P, Rubtsov AV. Sexual dimorphism in autoimmunity. *J Clin Invest.* 2015;125(6):2187–93. [PubMed: 25915581]

91. Kanaan SB, Onat OE, Balandraud N, Martin GV, Nelson JL, Azzouz DF, et al. Evaluation of X Chromosome Inactivation with Respect to HLA Genetic Susceptibility in Rheumatoid Arthritis and Systemic Sclerosis. *PLoS One*. 2016;11(6):e0158550. [PubMed: 27355582]
92. Lu Q, Wu A, Tesmer L, Ray D, Yousif N, Richardson B. Demethylation of CD40LG on the inactive X in T cells from women with lupus. *J Immunol*. 2007;179(9):6352–8. [PubMed: 17947713]
93. Hewagama A, Gorelik G, Patel D, Liyanarachchi P, McCune WJ, Somers E, et al. Overexpression of X-linked genes in T cells from women with lupus. *J Autoimmun*. 2013;41:60–71. [PubMed: 23434382]
94. Umiker BR, Andersson S, Fernandez L, Korgaokar P, Larbi A, Pilichowska M, et al. Dosage of X-linked Toll-like receptor 8 determines gender differences in the development of systemic lupus erythematosus. *Eur J Immunol*. 2014;44(5):1503–16. [PubMed: 24500834]
95. Garcia-Ortiz H, Velazquez-Cruz R, Espinosa-Rosales F, Jimenez-Morales S, Baca V, Orozco L. Association of TLR7 copy number variation with susceptibility to childhood-onset systemic lupus erythematosus in Mexican population. *Ann Rheum Dis*. 2010;69(10):1861–5. [PubMed: 20525845]
- 96 •. Shen N, Fu Q, Deng Y, Qian X, Zhao J, Kaufman KM, et al. Sex-specific association of X-linked Toll-like receptor 7 (TLR7) with male systemic lupus erythematosus. *Proc Natl Acad Sci U S A*. 2010;107(36):15838–43.
- TLR7 escapes X inactivation in some B cells and myeloid cells in females and men with Klinefelter syndrome. These biallelic B cells show enhanced TLR7-driven responses and greater propensity to class switch toward immunoglobulin G that may contribute to increased risk of SLE in women and men with Klinefelter syndrome.
- [PubMed: 20733074]
97. Subramanian S, Tus K, Li QZ, Wang A, Tian XH, Zhou J, et al. A Tlr7 translocation accelerates systemic autoimmunity in murine lupus. *Proc Natl Acad Sci U S A*. 2006;103(26):9970–5. [PubMed: 16777955]
98. McDonald G, Cabal N, Vannier A, Umiker B, Yin RH, Orjalo AV, Jr., et al. Female Bias in Systemic Lupus Erythematosus is Associated with the Differential Expression of X-Linked Toll-Like Receptor 8. *Front Immunol*. 2015;6:457. [PubMed: 26441962]
99. Souyris M, Cenac C, Azar P, Daviaud D, Canivet A, Grunewald S, et al. TLR7 escapes X chromosome inactivation in immune cells. *Sci Immunol*. 2018;3(19).
100. Pinheiro I, Dejager L, Libert C. X-chromosome-located microRNAs in immunity: might they explain male/female differences? The X chromosome-genomic context may affect X-located miRNAs and downstream signaling, thereby contributing to the enhanced immune response of females. *Bioessays*. 2011;33(11):791–802. [PubMed: 21953569]
101. Lam IKY, Chow JX, Lau CS, Chan VSF. MicroRNA-mediated immune regulation in rheumatic diseases. *Cancer Lett*. 2018;431:201–12. [PubMed: 29859876]
102. Wang J, Syrett CM, Kramer MC, Basu A, Atchison ML, Anguera MC. Unusual maintenance of X chromosome inactivation predisposes female lymphocytes for increased expression from the inactive X. *Proc Natl Acad Sci U S A*. 2016;113(14):E2029–38. [PubMed: 27001848]
103. Teuscher C, Noubade R, Spach K, McElvany B, Bunn JY, Fillmore PD, et al. Evidence that the Y chromosome influences autoimmune disease in male and female mice. *Proc Natl Acad Sci U S A*. 2006;103(21):8024–9. [PubMed: 16702550]
104. Spach KM, Blake M, Bunn JY, McElvany B, Noubade R, Blankenhorn EP, et al. Cutting edge: the Y chromosome controls the age-dependent experimental allergic encephalomyelitis sexual dimorphism in SJL/J mice. *J Immunol*. 2009;182(4):1789–93. [PubMed: 19201829]
105. Case LK, Wall EH, Dragon JA, Saligrama N, Krementsov DN, Moussawi M, et al. The Y chromosome as a regulatory element shaping immune cell transcriptomes and susceptibility to autoimmune disease. *Genome Res*. 2013;23(9):1474–85. [PubMed: 23800453]
106. Arnold AP. Y chromosome's roles in sex differences in disease. *Proc Natl Acad Sci U S A*. 2017;114(15):3787–9. [PubMed: 28360199]

107. Kremontsov DN, Case LK, Dienz O, Raza A, Fang Q, Ather JL, et al. Genetic variation in chromosome Y regulates susceptibility to influenza A virus infection. *Proc Natl Acad Sci U S A*. 2017;114(13):3491–6. [PubMed: 28242695]
108. Case LK, Teuscher C. Y genetic variation and phenotypic diversity in health and disease. *Biol Sex Differ*. 2015;6:6. [PubMed: 25866616]
109. Maan AA, Eales J, Akbarov A, Rowland J, Xu X, Jobling MA, et al. The Y chromosome: a blueprint for men's health? *Eur J Hum Genet*. 2017;25(11):1181–8. [PubMed: 28853720]
110. Pickard JM, Zeng MY, Caruso R, Nunez G. Gut microbiota: Role in pathogen colonization, immune responses, and inflammatory disease. *Immunol Rev*. 2017;279(1):70–89. [PubMed: 28856738]
111. Elderman M, van Beek A, Brandsma E, de Haan B, Savelkoul H, de Vos P, et al. Sex impacts Th1 cells, Tregs, and DCs in both intestinal and systemic immunity in a mouse strain and location-dependent manner. *Biol Sex Differ*. 2016;7:21. [PubMed: 27051505]
112. Shastri P, McCarville J, Kalmokoff M, Brooks SP, Green-Johnson JM. Sex differences in gut fermentation and immune parameters in rats fed an oligofructose-supplemented diet. *Biol Sex Differ*. 2015;6:13. [PubMed: 26251695]
113. Franssen F, van Beek AA, Borghuis T, Meijer B, Hugenholtz F, van der Gaast-de Jongh C, et al. The Impact of Gut Microbiota on Gender-Specific Differences in Immunity. *Front Immunol*. 2017;8:754.
114. Steegenga WT, Mischke M, Lute C, Boekschoten MV, Pruis MG, Lendvai A, et al. Sexually dimorphic characteristics of the small intestine and colon of prepubescent C57BL/6 mice. *Biol Sex Differ*. 2014;5:11. [PubMed: 25243059]
115. Elderman M, de Vos P, Faas M. Role of Microbiota in Sexually Dimorphic Immunity. *Front Immunol*. 2018;9:1018. [PubMed: 29910797]
116. Round JL, Palm NW. Causal effects of the microbiota on immune-mediated diseases. *Sci Immunol*. 2018;3(20).
117. Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, et al. Human gut microbiome viewed across age and geography. *Nature*. 2012;486(7402):222–7. [PubMed: 22699611]
118. Yurkovetskiy L, Burrows M, Khan AA, Graham L, Volchkov P, Becker L, et al. Gender bias in autoimmunity is influenced by microbiota. *Immunity*. 2013;39(2):400–12. [PubMed: 23973225]
119. Markle JG, Frank DN, Mortin-Toth S, Robertson CE, Feazel LM, Rolle-Kampczyk U, et al. Sex differences in the gut microbiome drive hormone-dependent regulation of autoimmunity. *Science*. 2013;339(6123):1084–8. [PubMed: 23328391]
120. Mu Q, Zhang H, Liao X, Lin K, Liu H, Edwards MR, et al. Control of lupus nephritis by changes of gut microbiota. *Microbiome*. 2017;5(1):73. [PubMed: 28697806]
121. Qu K, Zaba LC, Giresi PG, Li R, Longmire M, Kim YH, et al. Individuality and variation of personal regulomes in primary human T cells. *Cell Syst*. 2015;1(1):51–61. [PubMed: 26251845]
122. Ho B, Greenlaw K, Al Tuwaijri A, Moussette S, Martinez F, Giorgio E, et al. X chromosome dosage and presence of SRY shape sex-specific differences in DNA methylation at an autosomal region in human cells. *Biol Sex Differ*. 2018;9(1):10. [PubMed: 29463315]
123. Chiaroni-Clarke RC, Munro JE, Ellis JA. Sex bias in paediatric autoimmune disease - Not just about sex hormones? *J Autoimmun*. 2016;69:12–23. [PubMed: 26970680]
124. Bianchi DW, Zickwolf GK, Weil GJ, Sylvester S, DeMaria MA. Male fetal progenitor cells persist in maternal blood for as long as 27 years postpartum. *Proc Natl Acad Sci U S A*. 1996;93(2):705–8. [PubMed: 8570620]
125. Ngo ST, Steyn FJ, McCombe PA. Gender differences in autoimmune disease. *Front Neuroendocrinol*. 2014;35(3):347–69. [PubMed: 24793874]
126. Pellegrini M, Bulzomi P, Lecis M, Leone S, Campesi I, Franconi F, et al. Endocrine disruptors differently influence estrogen receptor beta and androgen receptor in male and female rat VSMC. *J Cell Physiol*. 2014;229(8):1061–8. [PubMed: 24347325]
127. Rogers JA, Metz L, Yong VW. Review: Endocrine disrupting chemicals and immune responses: a focus on bisphenol-A and its potential mechanisms. *Mol Immunol*. 2013;53(4):421–30. [PubMed: 23123408]

128. Borchers AT, Gershwin ME. Sociological differences between women and men: implications for autoimmunity. *Autoimmun Rev.* 2012;11(6–7):A413–21. [PubMed: 22155202]
129. Schneider-Hohendorf T, Gorlich D, Savola P, Kelkka T, Mustjoki S, Gross CC, et al. Sex bias in MHC I-associated shaping of the adaptive immune system. *Proc Natl Acad Sci U S A.* 2018;115(9):2168–73. [PubMed: 29440397]

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Key points

- The mechanisms underlying sex bias in autoimmunity remain incompletely understood.
- The effects of sex hormones on autoimmune disease are mediated in part by direct regulation of key immunity factors such as AIRE, IRF5, and IFN- γ and the intracellular TLR trafficking protein UNC93B1.
- Newly discovered non-hormonally-regulated immune modulators such as VGLL3 may contribute to female-biased autoimmunity.
- The gut microbiota influences the immune response and development of autoimmunity and is itself shaped by androgens.
- These and future investigations may yield targets for more selective and therefore less toxic therapies for autoimmune disease.

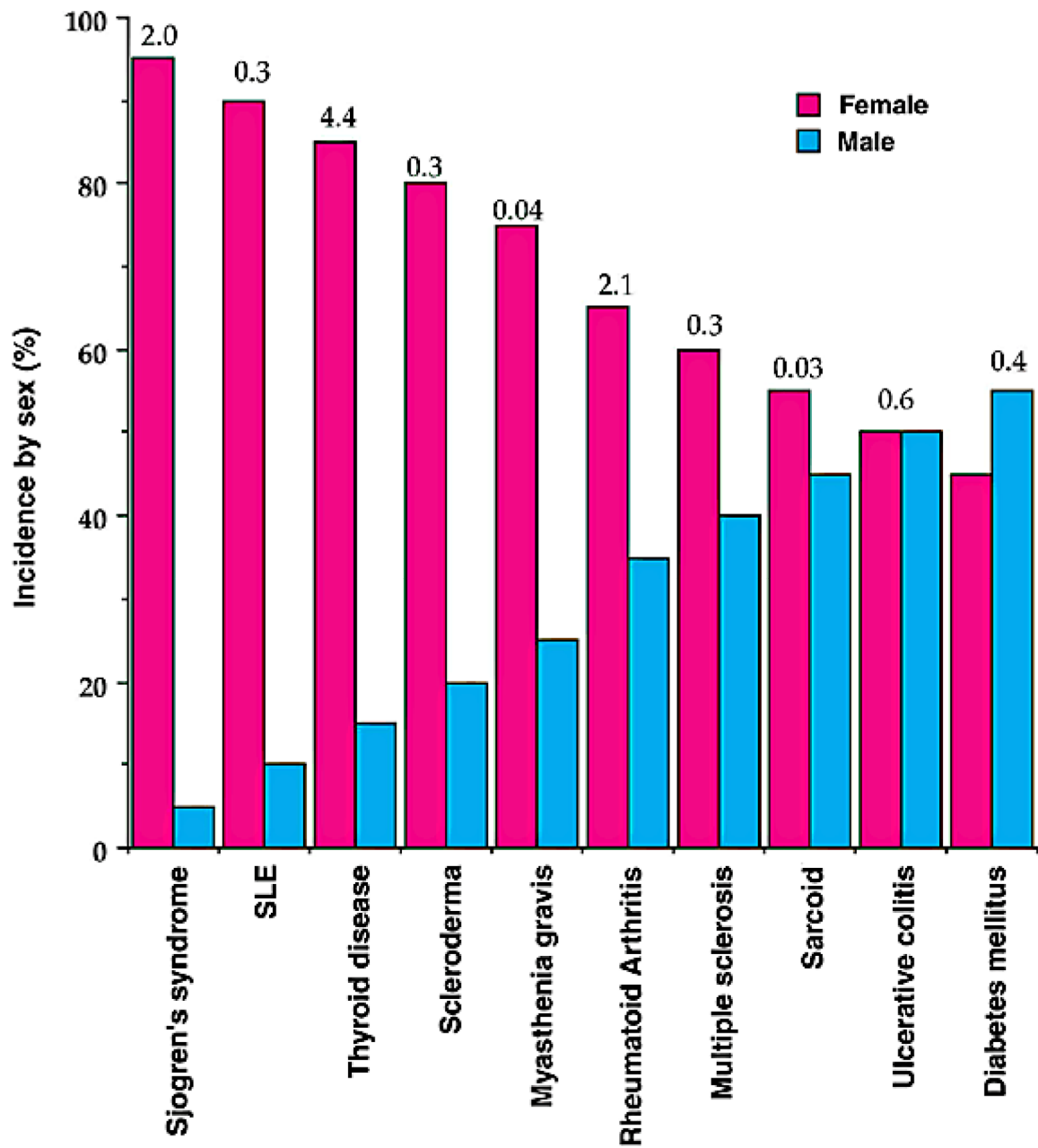


Figure 1. The sex distribution of the major autoimmune diseases.
 The numbers above the bars refer to the total number of disease cases ($\times 1,000,000$) in the USA (3). SLE: systemic lupus erythematosus.

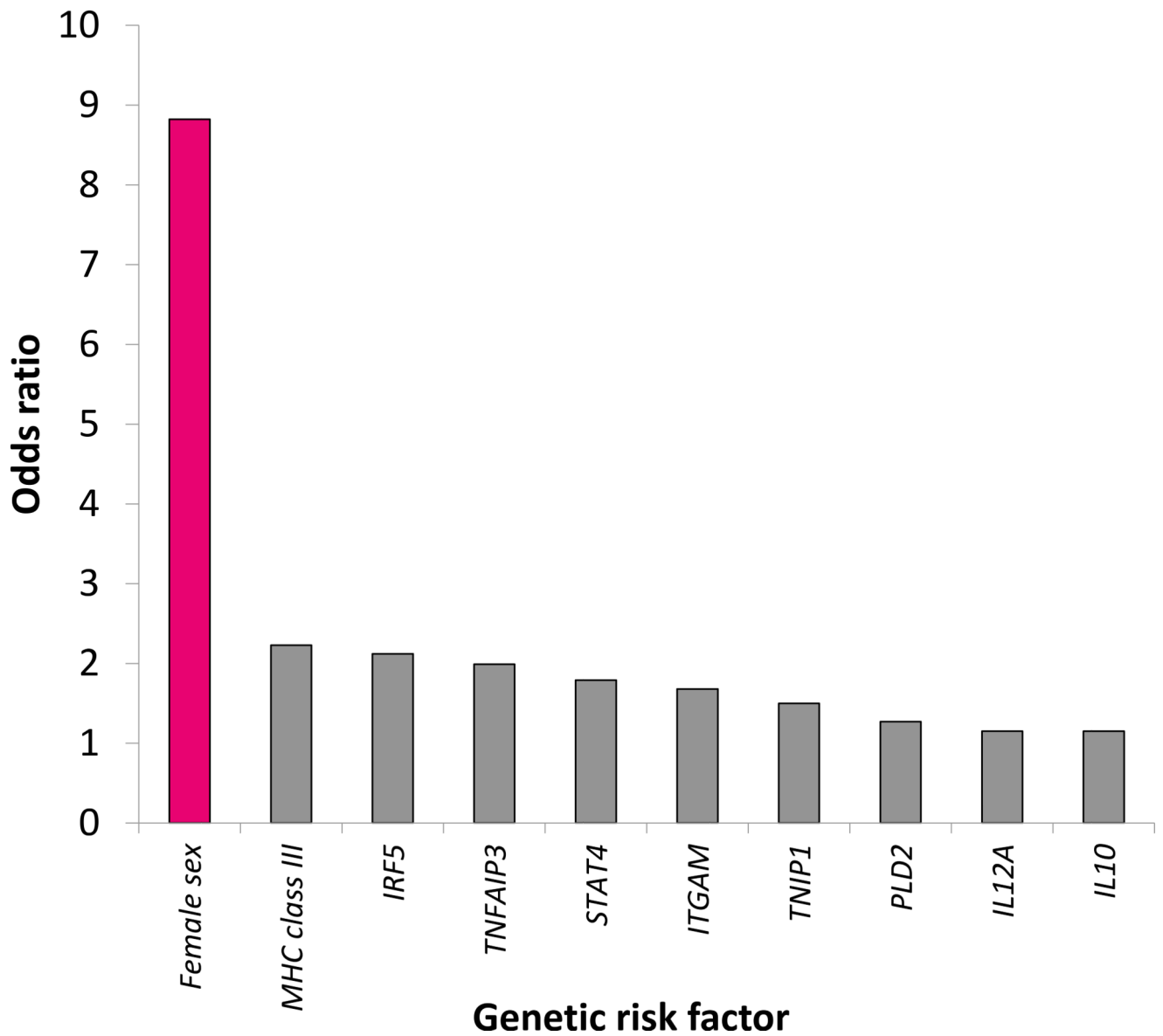


Figure 2. Female sex alone carries a risk of autoimmunity four times greater than any other known genetic risk variant for SLE.

Odds ratio (OR) for female sex was calculated based on prevalence data from the Georgia Lupus Registry from 2002 (6). ORs are shown for allelic associations at SLE susceptibility loci from a genome-wide association replication study (7).