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Growth Hormone Releasing Hormone Excess and Blockade in X-LAG Syndrome

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Abstract

X-linked acroigantism (X-LAG) syndrome is a newly-described form of inheritable pituitary gigantism that begins in early childhood and is usually associated with markedly elevated growth hormone (GH) and prolactin secretion by mixed pituitary adenomas/hyperplasia.

Microduplications on chromosome Xq26.3 including the *GPR101* gene cause X-LAG syndrome. In individual cases random GH-releasing hormone (GHRH) levels have been elevated. We performed a series of hormonal profiles in a young female sporadic X-LAG syndrome patient and subsequently undertook *in vitro* studies of primary pituitary tumor culture following neurosurgical resection. The patient demonstrated consistently elevated circulating GHRH levels throughout preoperative testing, which was accompanied by marked GH and prolactin hypersecretion; GH demonstrated a paradoxical increase following TRH administration. *In vitro*, the pituitary cells showed baseline GH and prolactin release that was further stimulated by GHRH administration. Co-incubation with GHRH and the GHRH receptor antagonist, acetyl-(D-Arg²)-GHRH (1–29) amide, blocked the GHRH-induced GH stimulation; the GHRH receptor antagonist alone significantly reduced GH release. Pasireotide, but not octreotide, inhibited GH secretion. A ghrelin receptor agonist and an inverse agonist led to modest, statistically significant increases and decreases in GH secretion, respectively. GHRH hypersecretion can accompany the pituitary

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abnormalities seen in X-LAG syndrome. These data suggest that the pathology of X-LAG syndrome may include hypothalamic dysregulation of GHRH secretion, which is in keeping with localization of GPR101 in the hypothalamus. Therapeutic blockade of GHRH secretion could represent a way to target the marked hormonal hypersecretion and overgrowth that characterizes X-LAG syndrome.

Introduction

Pituitary gigantism is a very recognizable but rare disorder (Rostomyan, et al. 2015). Patients with pituitary gigantism develop growth hormone (GH) excess from a pituitary tumor before closure of epiphyseal growth plates. The etiology of pediatric or adolescent pituitary tumors in general, and pituitary gigantism in particular, remains relatively obscure, with about 50% of cases of the latter having no known genetic cause (Rostomyan et al. 2015). The most frequent known genetic cause is aryl hydrocarbon receptor interacting protein gene (*AIP*) mutations (Daly and Beckers 2015). Recently we described a new condition, termed X-linked acrogigantism, (X-LAG) syndrome, which is characterized by gigantism occurring during infancy (Trivellin, et al. 2014). X-LAG syndrome patients develop pituitary adenomas/hyperplasia and greatly elevated growth hormone (GH) and insulin-like growth factor 1 (IGF-1) levels that are usually accompanied by hyperprolactinemia. Unlike other forms of pituitary gigantism due to single gene mutations/deletions, the underlying abnormality in X-LAG syndrome is a microduplication in chromosome Xq26.3 that includes the orphan G-protein coupled receptor (GPCR) gene, *GPR101* (Daly, et al. 2015; Trivellin et al. 2014).

The clinical features of patients with X-LAG syndrome have been described in detail recently (Beckers, et al. 2015). Patients with X-LAG syndrome usually develop pituitary macroadenomas, while some have hyperplasia alone or in conjunction with adenoma. The elevated GH/IGF-1 levels are poorly responsive to first-generation somatostatin analogs despite the presence of ample somatostatin subtype 2 receptors (Beckers et al. 2015; Trivellin et al. 2014). Control of growth and hormonal hypersecretion can be achieved by radical resection of the anterior pituitary. Even small amounts of residual tumor are capable of maintaining IGF-1 levels in the acromegalic range for many years, thereby necessitating multimodal therapy (Beckers et al. 2015). The GH receptor antagonist, pegvisomant has been a useful option for IGF-1 and growth control in a subgroup of cases (Beckers et al. 2015). Even so, the use of multiple treatment modalities does not guarantee control of growth, making X-LAG syndrome a challenging form of pituitary disease (Naves, et al. 2015). As clinical experience with X-LAG syndrome during long-term follow-up remains limited, the contribution of modalities like radiotherapy remain uncertain. The underlying pathological mechanism driving hyperplasia, adenoma formation and secretory behavior in X-LAG syndrome remains obscure. To address this issue, we performed a series of hormonal studies and *in vitro* experiments using primary pituitary culture of an adenoma resected from a young female patient with X-LAG syndrome. The results of these studies strongly suggest an important role for growth hormone releasing hormone (GHRH) dysregulation in X-LAG syndrome.

Methods

Case Report

The patient was a two-year-old female with no family history of growth disorders. Brief details of the patient were reported as part of the original X-LAG syndrome cohort studies (Beckers et al. 2015; Trivellin et al. 2014). She was born at 35.5 weeks gestation and had a birth weight of 2306 g and a birth length of 44.5 cm, which were both below the 3rd percentile. Beginning from two months of age she began to grow excessively in terms of length and weight, exceeding the 97th percentile of her growth charts by the ages of 12 and 14 months, respectively. She was diagnosed with marked GH, IGF-1 and prolactin excess and a pituitary MRI demonstrated a large, uniform sellar lesion (17×8×8 mm) with suprasellar extension. A random level of GHRH was raised, although no potential ectopic sources were identified despite exhaustive imaging, including radionuclide studies (Octreoscan). Treatment with octreotide (150 mcg sc t.i.d.) and cabergoline (0.5 mg/week) was initiated, but IGF-1 levels remained consistently elevated. She was diagnosed with X-LAG syndrome based on an array CGH study that showed a microduplication on chromosome Xq26.3 that was confirmed by fluorescent in situ hybridization studies on peripheral blood leukocytes. As chronic treatment with somatostatin analogs and dopamine agonists for 12 months had no effect on the patient's excessive growth and her pituitary lesion began impinging on the optic chiasm, she was referred for neurosurgical treatment.

At transsphenoidal surgery, a gross total resection of the anterior pituitary was performed. Histology revealed a densely cellular tissue specimen comprised of cords of polymorphic cells with abundant cytoplasm (some acidophilic, some chromophobic) and rounded nuclei. Some nuclear atypia was seen, as were some mitotic figures. Small calcifications were identified but no necrotic foci were seen. The pathological findings are shown in Figure 1. Silver staining demonstrated widespread pituitary hyperplasia with widening of the pituitary acini, which was accompanied by focal loss of acinar structure in specific regions indicating adenoma formation. Immunohistochemistry revealed widespread positivity for GH (Biomedica) and prolactin (DakoCytomation) throughout the lesion. Staining for ACTH and TSH (both DakoCytomation) was limited to the periphery of the resected tissue and rare cells disseminated within the lesion (LH staining was very rare and FSH staining was absent (DakoCytomation)). Nuclear positivity for p53 (DO-7 clone, BioCare Medical) was focally intense in some cells within the lesion and the Ki67 index (MIB-1, DakoCytomation) was elevated at 5%. CAM 5.2 (Becton-Dickinson) staining revealed cytoplasmic peri-nuclea, dot-like positivity patterns. The overall pathological result was determined to be a combination of extensive GH/prolactin positive hyperplasia of the anterior pituitary with an atypical mixed GH/prolactin adenoma. Staining for somatostatin receptors subtypes 2 and 5 was present at moderate to high levels.

Postoperatively she had permanent cortisol and thyroid deficiencies and diabetes insipidus, which all were all replaced. During 10 months of follow-up her GH levels were between 0.9 and 1.4 ng/ml (normal: <4 ng/mL), while her IGF-1 levels were near to or above the upper limit of the normal range for her age/sex (160.7–180.4 ng/mL; normal range: 51–170 ng/

mL). Her height (116.5 cm; +3 SDS) and weight (23.5 kg; +2.4 SDS) remain elevated 10 months postoperatively.

Cell culture

A surgical sample of the human pituitary adenoma in Dulbecco's modified Eagle's medium (DMEM) was dissociated mechanically and enzymatically as described previously (Jaquet, et al. 1985). A sample of the medium in which this preparation was performed was retained for hormonal analysis. A total of 50,000 tumor cells per well were plated in 24 well plates coated with extracellular matrix from bovine endothelial corneal cells required for cell adhesion as previously described (Barlier, et al. 1999; Jaquet et al. 1985; Jaquet, et al. 2005; Saveanu, et al. 2002). The total incubation times were short and have been established not to influence cell number/viability in the setting of pituitary somatotrope primary cultures (Barlier et al. 1999; Jaquet et al. 1985; Jaquet et al. 2005; Saveanu et al. 2002); hence hormone levels were not normalized to the number of living cells. The cells were cultured in DMEM depleted of L-valine (D-valine-DMEM) to block fibroblast proliferation, supplemented with 1% fetal calf serum, penicillin (100U/ml), streptomycin (100µg/ml), and glutamine (100U/ml) at 37°C in a water-saturated atmosphere containing 7% CO₂. After 48h, culture medium was changed to D-valine DMEM containing 1% Insulin Transferin Selenium and 1% fetal calf serum for another 48h. Cells were incubated with drugs at a range of concentrations (10⁻¹²-10⁻⁷ mol/L) for 16h (overnight). After these initial studies, the cells were washed three times before undergoing GHRH and GHRH antagonist studies. All experiments were performed in triplicate. The medium was collected and stored frozen for hormonal assays.

Pharmacological compounds

The SSAs, octreotide and pasireotide were kindly provided by Novartis (Basel, Switzerland). Forskolin was obtained from Sigma-Aldrich (St. Quentin Fallavier, France). Cabergoline was obtained from Pfizer (Pharmacia & Upjohn Kalamazoo, USA) and MK-0677 (ghrelin analog) was obtained from Europeptide, Argenteuil France; MSP (ghrelin receptor-GHS-R inverse agonist) came from Polypeptide Laboratories (Strasbourg, France). The GHRH receptor antagonist, acetyl-(D-Arg²)-GHRH (1-29) amide, and GnRH₁₋₅ were obtained from Bachem, Europe. Peptides used for dynamic hormonal stimulation studies in vivo, such as GnRH, were obtained from HRF-Ayerst, Belgium.

Hormonal assays

GH secretion by the primary culture cells was quantified using the AlphaLisa human growth hormone GH kit (Perkin Elmer USA). For that assay, the intra-assay coefficient of variation (CV) is 4.9-5.4% and the inter-assay CV is 6.1-6.9% over the concentration range studied. Prolactin levels were measured in culture medium using a commercial IRMA kit (PRL IRMA Kit-Beckman Coulter Immunotech, Marseille, France). Human GHRH concentrations in plasma were measured using an in-house double-antibody radio-immunoassay as described previously and using a standard solution of Stimu-GH 50 µg (Ferring) and the matrix was nonused culture medium (Girard, et al. 1987).

Statistical analysis

Data are expressed as mean \pm SEM. The statistical tests used were the non-parametric paired Wilcoxon and non-paired Mann-Whitney tests. Significance was set at $p < 0.05$.

Results

Dynamic hormone profiles

Pre-operatively a series of dynamic pituitary hormone tests were performed, including a 180 minute test for GH, prolactin and GHRH pulsatility, a GnRH test (25 μ g i.v. GnRH to study prolactin, GH and GHRH (0, 15, 30 and 90 minutes) and a TRH test (125 μ g i.v.) with measurements of prolactin, GH and GHRH (0, 15, 30 and 90 minutes). These were all performed in the immediate preoperative period after sufficient washout of octreotide (>48 hrs.) and cabergoline (4 weeks). The pulsatility study (Figure 2A) demonstrated markedly raised levels of GH and PRL throughout and all GHRH levels were elevated at all study time points. Fluctuations in elevated GHRH levels also occurred, and these were partially but not completely in phase with alterations in GH. While the elevated prolactin levels varied by less than $\pm 7\%$ from baseline throughout the study, changes in GH levels were more marked (-23.6% to $+61.5\%$).

A TRH test (Figure 2B) led to a paradoxical rise in GH that was accompanied by the expected rise in prolactin. Following GnRH administration (Figure 2C), the grossly elevated baseline level of GH (212 ng/mL) decreased to 50 ng/mL. Again, the GHRH level was also elevated at baseline (138 pg/mL) and remained elevated throughout.

Cell culture experiments

In the medium used for cell expansion before pharmacological incubation studies, the GHRH concentration was 0.3ng/50,000 cells. In 1% FCS and ITS, the 24h secretion for 50,000 cells was 40ng/ml for GH and 300ng/ml for prolactin indicating that the tumor cells were actively producing hormone. The GHRH concentration was below the lower limit of the assay (<62 pg/mL) under these same experimental conditions.

As shown in Figure 3A, basal GH secretion was further stimulated by incubation with GHRH reaching a peak at a GHRH concentration of 10^{-10} M ($+41\pm 0.4\%$). In contrast, the putative GPR101 agonist, GnRH1–5, had little effect on GH across the dose range studied ($+15\pm 1.5\%$ at 10^{-10} M). Prolactin secretion was slightly stimulated by GHRH, ($+11\pm 0.4\%$ at 10^{-8} M). GnRH1–5 did not stimulate prolactin release (Figure 3B).

When tumor cells were incubated with the SSTR2-specific somatostatin analog, octreotide, no inhibition in GH secretion was seen, while the prolactin decrease was $-26\pm 0.03\%$ at 10^{-9} M (Figure 3C). In contrast, following incubation with pasireotide, a multi-somatostatin receptor agonist, a decrease in GH secretion was seen, reaching a maximum inhibition at 10^{-8} M ($-18\pm 0.6\%$; pasireotide had no effect on prolactin secretion in these tumor cells, Figure 3D). The D2-receptor agonist, cabergoline, displayed stronger inhibition than either somatostatin analog on GH secretion ($-32\pm 0.8\%$ at 10^{-8} M; Figure 3C), but the inhibition of prolactin was less ($-15\pm 0.04\%$; Figure 3D)).

We then studied the effect of a GHRH receptor antagonist, acetyl-(D-Arg²)-GHRH (1–29) amide on GH and prolactin secretion (Figure 4). There was an initial stimulatory effect of the GHRH-receptor antagonist on GH secretion at 10⁻¹³ M to 10⁻¹¹M (+18±0.8% and +13±1.9% respectively). This was followed by a dose dependent inhibition of GH secretion, reducing GH secretion to approximately -34±1.9% of controls at the highest GHRH-receptor antagonist dose studied (Figure 4A). Interestingly, incubation with the GHRH receptor antagonist also markedly decreased prolactin secretion, -73±0.1%. at a concentration of 10⁻⁸ M (Figure 4B). Co-incubation of GHRH (10⁻¹⁰M) with the GHRH antagonist led to complete abolition of the GH and PRL responses to GHRH antagonist (Figure 4A, B).

Figure 5 shows the effects of incubation with a ghrelin analog, MK-0677, and the inverse agonist of the ghrelin receptor, MSP. MK-0677 had a minor stimulatory effect on GH secretion as compared with control (+15±0.5%), while no effect on prolactin secretion occurred. MSP did, however, decrease GH and PRL secretion by -24%±0.3% and -12±0.08%, respectively, as compared with controls.

Discussion

X-LAG syndrome is a novel condition due to microduplications on chromosome Xq26.3 involving the orphan GPCR, GPR101 (Trivellin et al. 2014). This condition leads to a dramatic phenotype of early childhood onset gigantism due to large mixed GH and prolactin secreting pituitary adenomas and/or hyperplasia (Beckers et al. 2015). A progression from normal tissue through hyperplasia to adenoma has been seen in resected tumor tissue (Trivellin et al. 2014). The pituitary pathology progression suggests that while there may be an underlying abnormality (such as GHRH secretion) causing the hyperplasia, second hits may be involved in leading to the adenoma formation; this remains to be investigated fully in X-LAG syndrome. Usually both GH and prolactin secretion are highly elevated at diagnosis. We also reported previously that while GHRH staining was absent in tumors from X-LAG syndrome patients, abundant GHRH-R was seen both in hyperplastic and adenomatous regions (Beckers et al. 2015; Trivellin et al. 2014). The X-LAG syndrome cohort and literature cases with an identical phenotype include cases with individually high circulating GHRH levels in the absence of a peripheral source like a neuroendocrine tumor. As those cases were largely retrospective, more detailed hormonal and tumoral studies were not possible. The current patient is the youngest case diagnosed in the University of Liège-NIH collaborative cohort, which includes 22 cases at the time of writing. Subsequent to the diagnosis of X-LAG syndrome, a single elevated GHRH was noted during her clinical workup. In the current study during the preoperative period we found increased GHRH levels at all time points, ranging between 100–200 pg/ml. These moderately elevated GHRH levels were seen against a backdrop of markedly increased circulating GH and prolactin concentrations in excess of 200 ng/ml and 500 ng/ml, respectively, at baseline. A pulsatility study showed elevations in GHRH, GH and prolactin throughout the 180 mins of the study. One peak of GHRH secretion was captured and was seemingly accompanied by a concomitant peak in GH but not prolactin secretion. The elevated GHRH was not markedly altered by TRH or GnRH administration. While a longer testing period would have provided more information on GHRH patterns this was not feasible due to the young age of the

patient. No peripheral source of GHRH was found (in keeping with other X-LAG syndrome cases), indicating that GHRH hypersecretion could be an intrinsic part of the etiology of X-LAG syndrome, at least in some patients.

The source of the chronically elevated circulating GHRH is likely to be central, based on a number of lines of evidence. As noted above, no peripheral sources were seen following exhaustive imaging studies. Also the GHRH level was moderately elevated and was not consistent with the magnitude of the GHRH elevation that is encountered typically in ectopic GHRH hypersecretion from a neuroendocrine tumor. The pathology of pituitary disease in this and other cases of X-LAG syndrome bears strong resemblance to that seen in other settings of GHRH excess. As noted previously, peripheral GHRH hypersecretion from a neuroendocrine tumor is accompanied by either pituitary somatotrope hyperplasia or mixed GH and prolactin secreting adenomas (Borson-Chazot, et al. 2012). Central GHRH hypersecretion from hypothalamic gangliocytomas leads to pituitary adenoma and acromegaly (Asa, et al. 1984). Hypersecretion of GHRH in mice transgenic for the human GHRH gene leads to the development of early and massive pituitary hyperplasia (somatotrope, mammotrope and somatomammothrope) (Mayo, et al. 1988); similar extensive hyperplasia accompanied adenoma formation in the current case and in other X-LAG syndrome patients (Trivellin et al. 2014). Furthermore, in older transgenic mice, chronic GHRH hypersecretion leads to pituitary adenoma formation, predominantly with GH and prolactin staining (Asa, et al. 1992). Taken together, the sum effect of chronic GHRH hypersecretion on the pituitary is somatotrope, mammotrope and somatomammothrope hyperplasia and/or mixed GH and prolactin secreting adenomas, which is the picture seen in this and other cases of X-LAG syndrome. This strongly suggests that the central GHRH hypersecretion seen in the current case may explain the pituitary histological changes that occur in X-LAG syndrome.

One further area that requires more investigation is the GHRH-receptor activity in this and other pituitary tumors. We found that not only did a GHRH receptor antagonist block GHRH-related stimulation of GH, but the GHRH-antagonist itself was capable of reducing basal GH secretion. This suggests that the cells may have some intrinsic GHRH-receptor related GH secretion at baseline, which requires further study, as tumors in patients with X-LAG syndrome express significant numbers of GHRH receptors (Beckers et al. 2015; Naves et al. 2015; Trivellin et al. 2014). Indeed a whole range of studies into the behavior of the pituitary in X-LAG syndrome may help elucidate some of the specific features of somatotrope behavior in response to GHRH in early life (fetal/neonatal) as compared with the adult (Collins, et al. 1996; Torronteras, et al. 2012; Torronteras, et al. 1997).

Further support for a role for GHRH hypersecretion in X-LAG syndrome comes from the *in vitro* pituitary culture data, which are the first results to be reported in a tumor from a known case of X-LAG syndrome. These results confirm basal GH and prolactin hypersecretion, which was further stimulated by GHRH co-incubation. Crucially, the addition of the GHRH antagonist acetyl-(D-Arg²)-GHRH (1–29) was able to reduce basal and stimulated GH and prolactin secretion in a dose dependent manner. Co-incubation of acetyl-(D-Arg²)-GHRH (1–29) with GHRH abolished the stimulatory effect of GHRH itself. This combination of findings suggests that clinical benefit could be yielded from pharmacological treatment with

GHRH antagonists in X-LAG syndrome. Previous work in normal volunteers and in a patient with ectopic tumoral GHRH hypersecretion shows that such an approach with acetyl-(D-Arg²)-GHRH (1–29) and other GHRH analogs is clinically feasible (Jaffe, et al. 1997; Jaffe, et al. 1993; Kovacs, et al. 1997). Newer GHRH antagonists are being tested clinically in other settings but could also have applications in X-LAG syndrome (Kovacs, et al. 2010; Szalontay, et al. 2014).

Post-operatively the GHRH was tested at a single time-point (lengthy dynamic testing was declined) and found to be in the normal range. We cannot rule out that GHRH remains intermittently elevated at this time. This normal GHRH level also could be interpreted as pointing to the resected tissue being a source of the GHRH secretion. However, immunohistochemical studies of this and other X-LAG syndrome cases are negative for GHRH (Beckers et al. 2015; Trivellin et al. 2014). In addition, RNA studies of tumor from multiple X-LAG syndrome cases show no GHRH expression (Daly, Trivellin, Palmeira Beckers & Stratakis, Unpublished data). These two findings argue strongly against the tumor itself being the GHRH source. Following surgery the patient exhibited deficiency in thyroid and adrenal axes, plus diabetes insipidus, indicating significant pituitary stalk impairment. Despite this, however, the IGF-1 secretion has remained at or above the upper limit of normal. Taken together, this suggests that the patient might have a small adenomatous remnant that is autonomously producing GH. Such small tumor remnants in X-LAG syndrome have led to chronic GH hypersecretion over decades in affected patients, requiring long-term medical treatment (Beckers et al. 2015; Trivellin et al. 2014). We suggest that tumor resection led to pituitary stalk impairment that prevented normal flow of hypothalamic factors-including GHRH- to the pituitary and into the circulation via the venous channels. This could explain the picture of significant GH/IGF-1 secretion in the absence of elevated GHRH as had been present before surgery. Alternatively, the pituitary tissue itself could have contained a factor that led to feedback dysregulation of hypothalamic GHRH and surgical resection could have removed this abnormality. This remains speculative, but cannot be completely discounted in the setting of tumoral GPR101 overexpression, for which no regulators, ligands, or physiological/pharmacological modulators are yet known.

We also studied a number of other stimulatory and inhibitory factors. We previously reported poor clinical responses to SSTR2-specific somatostatin analogs in X-LAG syndrome, despite strong immunohistochemical staining for SSTR2 in tumor samples (Beckers et al. 2015; Naves et al. 2015; Trivellin et al. 2014). In keeping with this, octreotide had no inhibitory effect on GH or prolactin in the X-LAG syndrome pituitary cell culture, which supports the poor response to octreotide during chronic treatment in this case. Pasireotide had a small inhibitory effect on GH secretion, which suggests that some clinical benefit might be obtained in X-LAG syndrome via its binding at SSTR5 and SSTR3, as these receptor types are expressed in X-LAG syndrome tumor tissue (Beckers et al. 2015). The GnRH_{1–5} fragment was suggested elsewhere to potentially act as a ligand for GPR101, the orphan GPCR overexpressed due to the Xq26.3 microduplication causing X-LAG syndrome (Cho-Clark, et al. 2014; Cho-Clark, et al. 2015). In our study, GnRH_{1–5} had only a minor stimulatory effect on GH at lower doses and a small inhibitory effect at the highest dose 10⁻⁸M. The lack of a marked effect in this tissue culture might be due to inter-tissue

differences, as much of the previous data on GnRH₁₋₅ was generated in an endometrial cell line. As some patients with X-LAG syndrome exhibit increased appetite in conjunction with their gigantism, we were interested in studying the effects of ghrelin in the tumor tissue. Ghrelin agonism itself had a minor stimulatory effect on GH secretion, while a ghrelin receptor inverse agonism inhibited GH secretion at 10⁻⁸ and 10⁻⁷ M. These results suggest that patients with X-LAG syndrome may have GH dysregulation across various control pathways, although specific studies are needed.

In conclusion, this study provides new evidence that central GHRH dysregulation is a feature of the pituitary pathology and gigantism seen in X-LAG syndrome. It may be that chronic GHRH hypersecretion from the hypothalamus in X-LAG syndrome contributes to the pituitary pathology of mixed GH/prolactin positive hyperplasia and adenoma, which mirrors that seen in other experimental and clinical settings with GHRH excess. Inhibition of GH and prolactin secretion from cell culture in X-LAG syndrome using a GHRH antagonist suggests that the challenging clinical management of X-LAG syndrome could be improved by targeting GHRH inhibition

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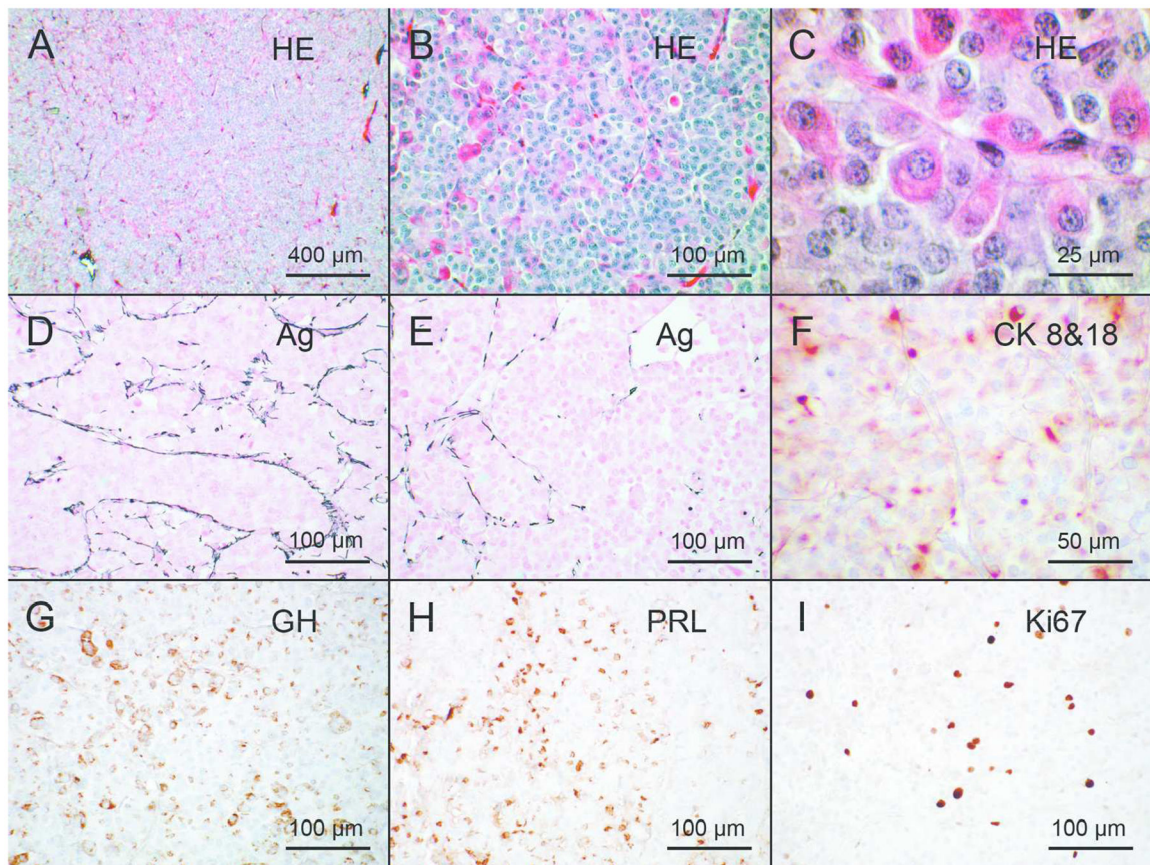
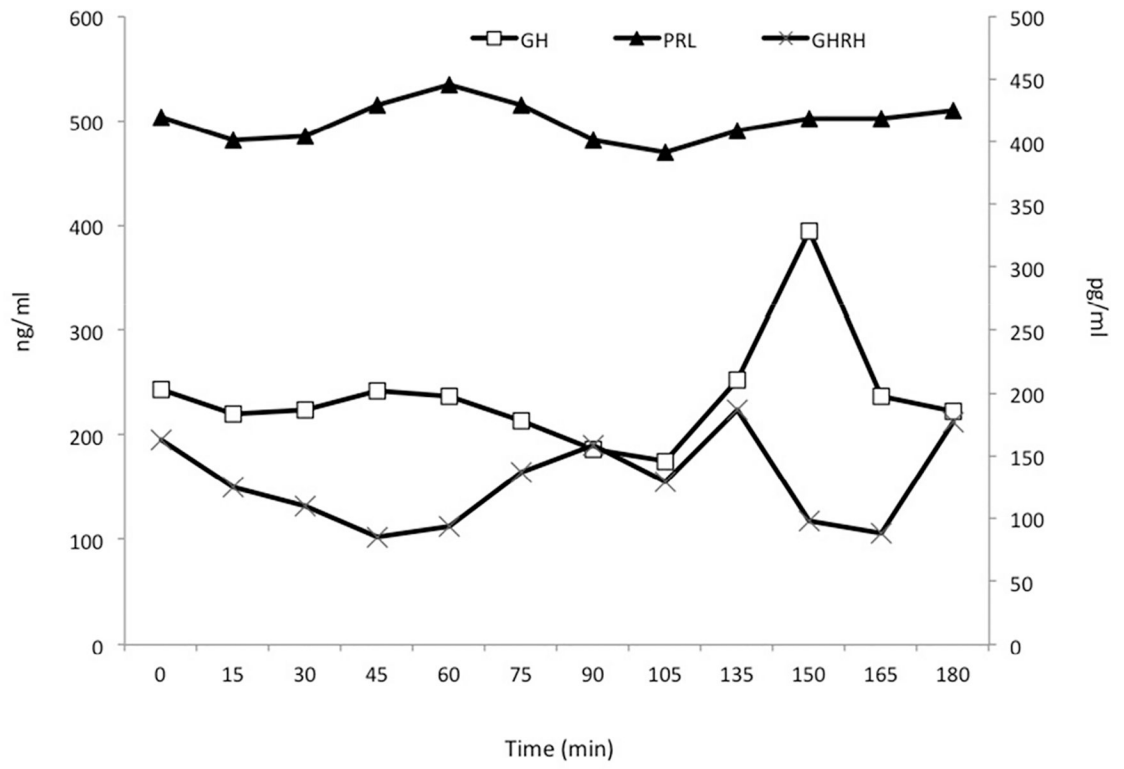
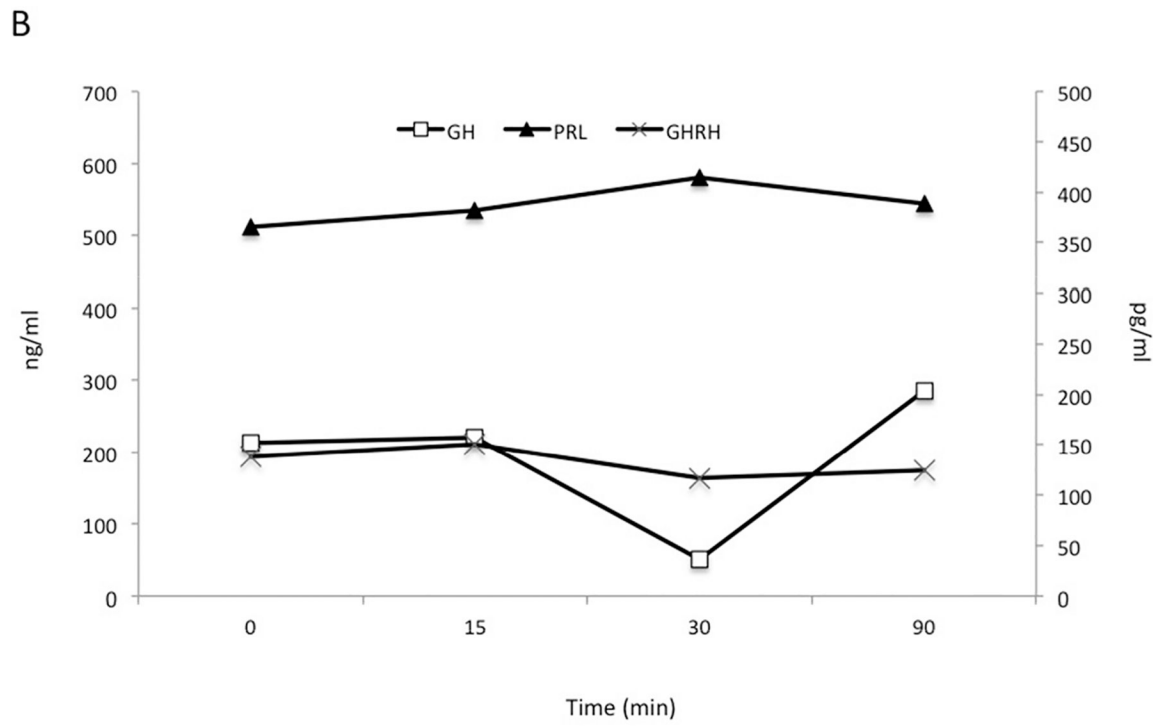
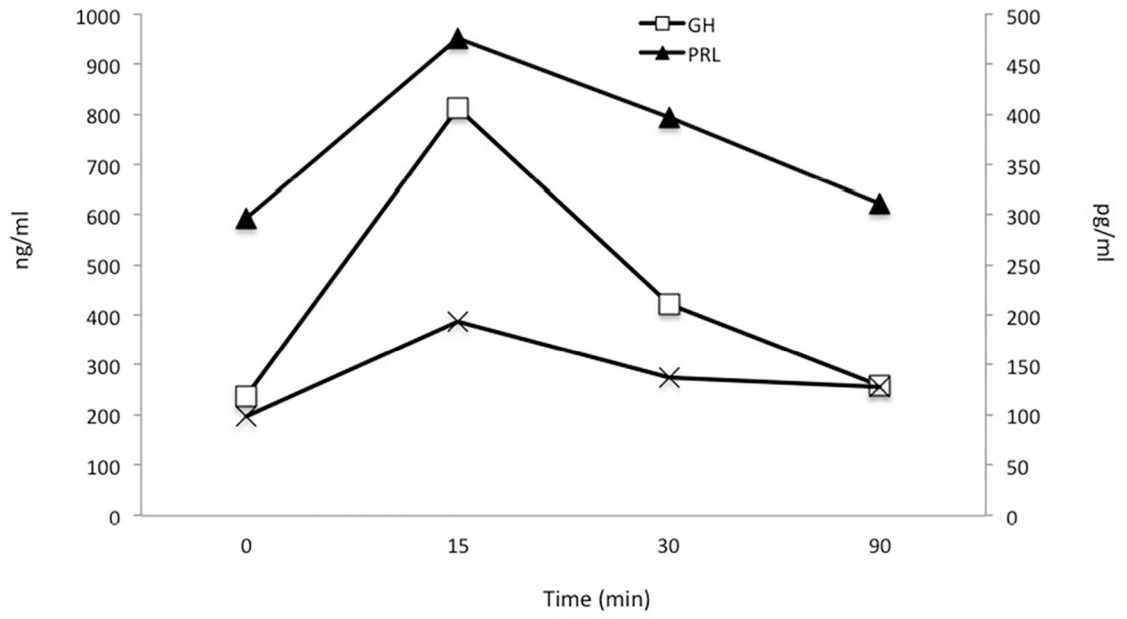


Figure 1.

Histological and immunohistochemical appearance of the pituitary lesion. Histological sections were stained with hematoxylin and eosin (A, B, C), and with silver to highlight the reticulinic fibres (D, E) or immunostained with the CAM 5.2 monoclonal mouse antibody against cytokeratins 8 and 18 (F), with polyclonal rabbit antibody against growth hormone (G) or against prolactin (H), or with the MIB-1 mouse monoclonal antibody against Ki67 (I). Hematoxylin and eosin-stained sections (A-C) show the abundance of acidophilic somatotroph cells. Argyrophilic staining show that reticulinic fibres are still present in most parts of the lesion (D), indicating its hyperplastic nature, but tend to disappear in areas (E), suggesting development of an adenoma within the hyperplastic tissue. Pituitary cells contain cytokeratin filaments, most often diffusely distributed in the cytoplasm but concentrated in a perinuclear dot in a few cells (F). Many cells contain growth hormone (G) or prolactin (H). About 5% of pituitary cells are proliferating as indicated by Ki67 immunostaining (I). Bars correspond to 400 μm in A, to 100 μm in B, D, E, G, H, I, to 50 μm in F and to 25 μm in C.



A



C

Figure 2.

Dynamic and pulsatility testing. Elevated growth hormone (GH), prolactin (PRL) and GHRH levels were seen throughout an extended testing period of 180 minutes (Panel A). GH levels are seen to peak at 150 min after an earlier GHRH rise between 105min and 135 min. A TRH test induced an immediate and marked increase in GH levels and GHRH remained largely unchanged (Panel B). GnRH administration led to suppression of GH levels at 30 mins, whereas GHRH levels remained unaltered and prolactin rose mildly (Panel C). GH and PRL are measured in ng/mL; GHRH is measured in pg/mL.

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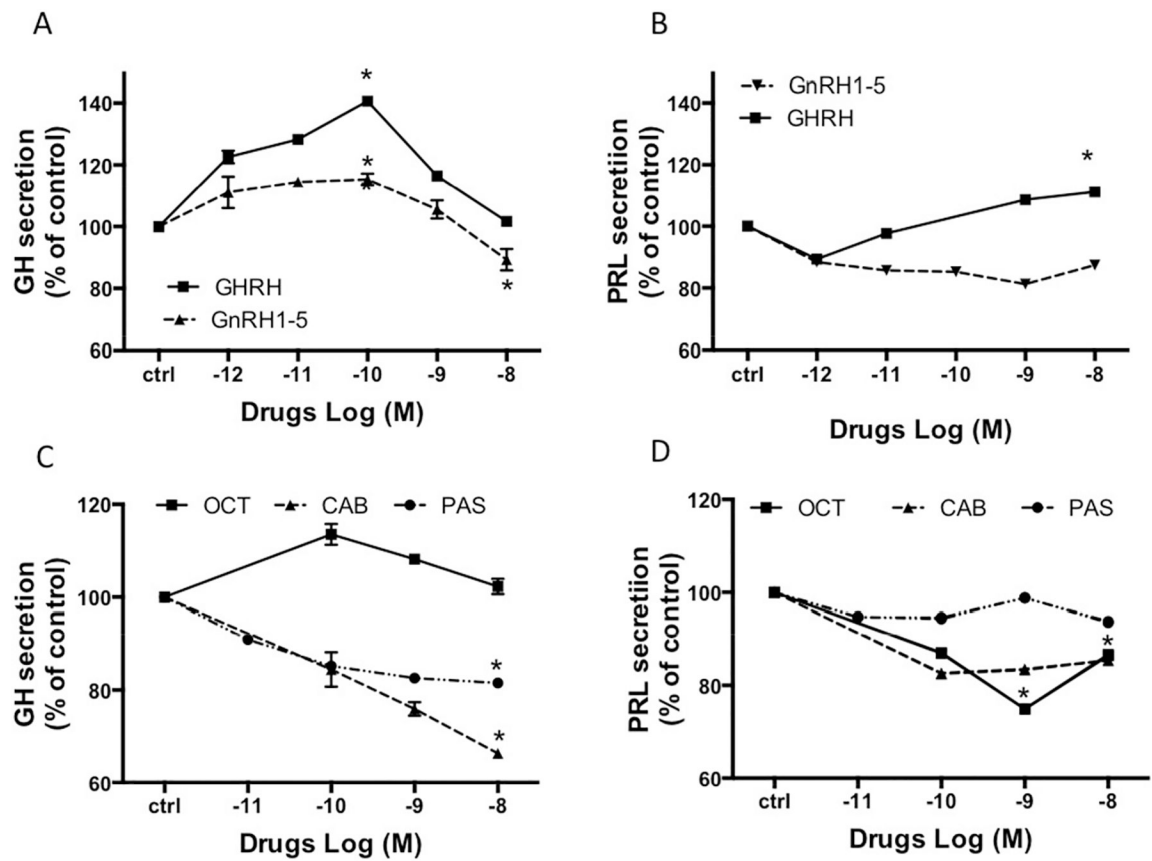


Figure 3.

The *in vitro* dose-effect on GH (A, C) and PRL (B, D) secretion in somatotroph cells after overnight incubation with (A,B) GHRH, GnRH1–5, (C,D) octreotide (OCT) cabergoline (CAB) or pasireotide (PAS). The results are expressed as the mean percentage of PRL or GH change compared to the values of control wells (ctrl). * : $p < 0.05$.

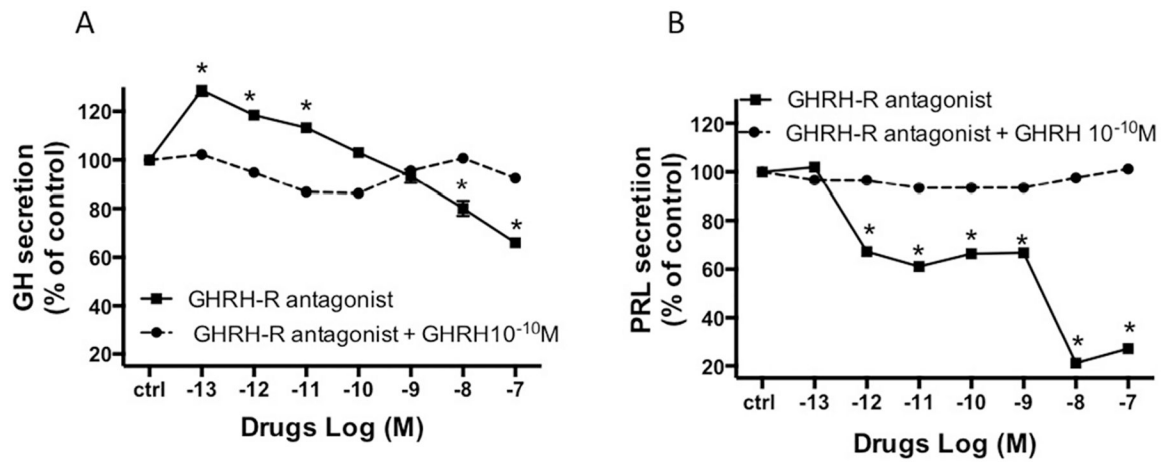


Figure 4.

The *in vitro* dose-effect on GH (A) and PRL (B) secretion in somatotroph cells after overnight incubation with GHSR agonist (MK 0677) or GHSR-inverse agonist (MSP). The results are expressed as the mean percentage of PRL or GH change compared to the values of control wells (ctrl). * : $p < 0.05$.

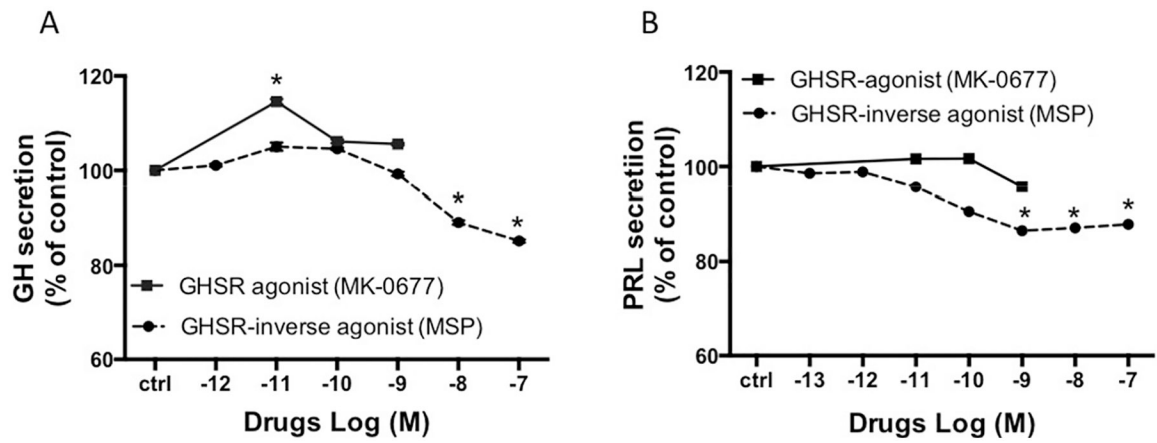


Figure 5.

The *in vitro* dose-effect on GH (A) and PRL (B) secretion in somatotroph cells after overnight incubation with GHRH-R antagonist, with or without GHRH at 10^{-10} M. The results are expressed as the mean percentage of PRL or GH change compared to the values of control wells (ctrl). * : $p < 0.05$.