

Review



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The pervasiveness of macropinocytosis in oncological malignancies

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In tumour cells, macropinocytosis functions as an amino acid supply route and supports cancer cell survival and proliferation. Initially demonstrated in oncogenic *KRAS*-driven models of pancreatic cancer, macropinocytosis triggers the internalization of extracellular proteins via discrete endocytic vesicles called macropinosomes. The incoming protein cargo is targeted for lysosome-dependent degradation, causing the intracellular release of amino acids. These protein-derived amino acids support metabolic fitness by contributing to the intracellular amino acid pools, as well as to the biosynthesis of central carbon metabolites. In this way, macropinocytosis represents a novel amino acid supply route that tumour cells use to survive the nutrient-poor conditions of the tumour microenvironment. Macropinocytosis has also emerged as an entry mechanism for a variety of nanomedicines, suggesting that macropinocytosis regulation in the tumour setting can be harnessed for the delivery of anti-cancer therapeutics. A slew of recent studies point to the possibility that macropinocytosis is a pervasive feature of many different tumour types. In this review, we focus on the role of this important uptake mechanism in a variety of cancers and highlight the main molecular drivers of macropinocytosis in these malignancies.

This article is part of the Theo Murphy meeting issue 'Macropinocytosis'.

1. Introduction

A possible link between macropinocytosis and cancer became apparent with the discoveries that proto-oncogenes, such as *RAS*, *SRC* and *EGFR*, have the ability to stimulate this uptake mechanism [1–4]. Although it was clear that activating mutations in these proto-oncogenes leads to macropinocytic induction, a physiological role for this boost in uptake capacity remained uncertain for many years. Taking a clue from the role that macropinocytosis plays in axenic nutrient acquisition in the soil amoeba *Dictyostelium* [5], and inspired by the renaissance of the study of tumour metabolism, we established that macropinocytosis functions as a nutrient delivery pathway in cancer cells [6]. By internalizing extracellular proteins and targeting them for lysosomal degradation, macropinocytosis operates in tumours as an amino acid supply route, providing an endocytic source of protein-derived amino acids that support cancer cell bioenergetics and biosynthesis.

Since our initial studies, which focused on *KRAS*-mutated pancreatic cancer, it has become apparent that cancer-associated macropinocytosis may be more prevalent than we initially surmised (table 1). Recently, studies employing different human *in vitro* systems and *in vivo* animal models have demonstrated that macropinocytosis is a metabolic feature of other solid tumours, including those found in the lung, prostate, bladder and colon. Since macropinocytosis functions as an endocytic gateway for nanomedicines, including nanoparticles, liposomes, nanotubes, DNA nanostructures and protein–drug nanoconjugates [39–41], there are many *in vitro* examples of macropinocytosis in cancer cells that originate from a variety of tissues. The function of macropinocytosis in

Table 1. Examples of macroinocytosis in different tumour types.

| type of cancer | molecular driver | cancer cell lines | <i>in vivo</i> models | refs ^a |
|--|---|---|--|------------------------|
| bladder urothelial carcinoma | <i>HRAS</i> , <i>KRAS</i> , <i>PTEN</i> | T24, J82, UM-UC-3 | | [6,7] |
| brain and nervous system | glioma/glioblastoma neuroblastoma medulloblastoma | U251, U87MG, C6, U87 SH-SY5Y Daoy | | [8–11] [12] [13] |
| breast | ? | MDA-MB-231, MCF-7, HTB-20, MDA-MB-157 | | [14–17] |
| colorectal adenocarcinoma | <i>KRAS</i> , <i>P13K?</i> , <i>APC</i> | DLD-1, HCT116, HT-29, CT26 | colonic epithelium in R26-rtTA Tg-Apc.3374 (shAPC) genetically engineered mouse model | [17–21] |
| haematologic malignancies | ? | X63, J558 L, F8, Molt-4 | | [22–24] |
| hepatocellular carcinoma | <i>NRAS?</i> | HepG2 | | [25] |
| human papillomavirus-related cervical adenocarcinoma | ? | HeLa, SiHa | | [26,27] |
| lung adenocarcinoma | <i>KRAS</i> , integrin $\alpha v \beta 3$, galectin-3 | H1792, A549, SKLU1, H1299 | subcutaneous patient-derived (PDX) xenograft tumours | [28–30] |
| osteosarcoma | ? | HOS, 6647 | | [31,32] |
| ovarian adenocarcinoma | ? | OVCAR-3, SKOV-3, OV2008 | | [33,34] |
| pancreatic ductal adenocarcinoma | <i>KRAS</i> | MIA PaCa-2, PL45, Panc-1 | 1. subcutaneous xenograft tumours 2. autochthonous tumours in <i>P48-cre; Isl-Kras^{G12D}</i> ; 3. human tumours from whipple patients subcutaneous allograft tumours | [6,35–37] |
| prostate | <i>PTEN</i> | PC3, LNCaP, DU145, mPCE | | [38] |

^aFor each cancer type, the earliest or most extensive studies were selected as representative references.

drug delivery is becoming an important field of research in cancer biology and has been the topic of many other excellent reviews [41–44]. Intriguingly, in precursor cell malignancies affecting the brain and nervous system, such as glioblastoma, neuroblastoma and medulloblastoma, macropinocytosis needs to be finely balanced as amplifying the macropinocytic capacity in these contexts has the propensity to lead to a form of nonapoptotic cell death known as methuosis [45]. Like many examples of macropinocytosis in the oncological context, methuosis is linked to the elevated expression of oncogenic forms of *RAS*. The mechanistic underpinnings of oncogenic Ras-induced macropinocytosis have been reviewed elsewhere, but they entail the induction of downstream effector pathways that regulate actin cytoskeleton remodelling and membrane ruffling behaviour [46].

In this review, we aim to provide a semi-systematic analysis of the pervasiveness of macropinocytosis in cancer by focusing on the role of this important uptake mechanism in different tumour types. We highlight the key molecular drivers of macropinocytosis in these malignancies and provide an overview of similarities between the various pathological settings.

2. Pancreatic cancer

The vast majority of pancreatic ductal adenocarcinoma (PDAC) tumours exhibit oncogenic mutations in *KRAS* [47]. The initial discoveries that macropinocytosis serves as an amino acid supply route in cancer cells were made in cellular and animal models of *KRAS*-driven PDAC [6,35]. These studies demonstrated that PDAC cells that harbour oncogenic *KRAS* mutations display robust levels of macropinocytosis that are dependent on expression of the oncogene. By contrast, PDAC cells that express wild-type *KRAS* have low levels of macropinocytic induction. Importantly, these observations were recapitulated *in vivo* using tumour xenograft models. Additionally, macropinocytic induction was observed in an autochthonous model of PDAC. In this model, mice of the genotype *P48-cre; lsl-Kras^{G12D}; Trp53^{-/+}* (KPC) develop early pancreatic intraepithelial neoplasia (PanIN) lesions within four weeks of birth and progress to invasive PDAC between 9 and 13 weeks [48,49]. Using this model, macropinosomes were detected in pancreatic acinoductal cells found in mid- to late-stage PanIN lesions but not in pancreata from wild-type mice [6]. The clinical relevance of macropinocytosis in PDAC was established by scrutinizing human specimens. Although some intratumoral variability was observed, stimulated macropinocytosis was evident in all of the PDAC tumour samples analysed [35]. Altogether, these studies showed that macropinocytosis is an attribute of PDAC tumours and suggested that PDAC cells could take up fluids and their solubilized components from their extracellular environment.

Extracellular physiological fluid is mainly composed of proteins, with serum albumin being the most abundant. Therefore, it was a reasonable assertion that PDAC tumour cells were engaging in macropinocytosis to facilitate the uptake of extracellular serum albumin from the tumour interstitium. This was initially examined *in vitro* with colocalization assays that revealed the macropinocytic uptake of fluorescently labelled bovine serum albumin (BSA) [6]. To serve as a nutrient source, this macropinocytosed BSA

would need to be proteolytically degraded into its constituent amino acids. Indeed, using a self-quenched form of BODIPY-dye-conjugated BSA (DQ-BSA) that only emits a bright fluorescent signal upon digestion, it was shown that the incoming BSA is degraded within macropinosomes in a lysosome-dependent manner. The protein-derived amino acids that are produced by macropinocytosis in *KRAS*-mutant cells feed into a multitude of metabolic pathways, including the TCA cycle, glutamine anaplerosis/oxidation, acetyl-coenzyme A metabolism, reductive carboxylation and serine/glycine cycling. Importantly, these protein-derived amino acids have the ability to suppress the deleterious effects of nutrient starvation, leading to albumin-dependent proliferation and growth. Interestingly, catabolism of macropinocytosed serum albumin contributes substantially to PDAC intracellular amino acid pools, even in the absence of amino acid deprivation [35]. An important avenue of further study is determining whether amino acid deprivation can enhance macropinocytic uptake and to what extent macropinocytosis contributes to cellular fitness under nutrient-replete conditions.

The evaluation of the role of macropinocytosis in PDAC tumour growth took advantage of the fact that macropinocytosis is unique in relation to other endocytic pathways, because it is sensitive to 5-(*N*-ethyl-*N*-isopropyl) amiloride (EIPA) and other amiloride analogues that inhibit Na⁺/H⁺ exchange [50]. Miniaturized plasma exchange experiments demonstrated *in situ* that live tumours actively engage in macropinocytosis and break down albumin [51]. This albumin catabolism was suppressed by EIPA and resulted in the modulation of amino acid levels within the tumour. Importantly, EIPA administration in animals bearing *KRAS*-mutant xenograft tumours resulted in an attenuation of tumour growth, and in some cases regression, relative to vehicle-only controls [6]. The observed effects of EIPA on PDAC tumours might be specific to highly macropinocytic *KRAS*-mutant tumours because EIPA treatment had no bearing on the growth rate of tumours derived from *KRAS*-wild-type PDAC cells. Altogether, these exciting results implicated macropinocytosis as a potential therapeutic target in PDAC. Further research is required to assess whether EIPA or any other macropinocytosis inhibitors have therapeutic benefit in human PDAC patients.

3. Bladder cancer

While only a subset of bladder carcinomas harbour oncogenic *RAS* mutations, the majority of these genetic perturbations occur in *HRAS* [52]. Similar to PDAC cells, *HRAS*-mutant bladder cancer cells display robust levels of macropinocytic induction that they employ to augment their metabolism through the lysosome-mediated degradation of the extracellular protein [6]. Much knowledge about macropinocytosis in bladder cancer has been gained by studying the internalization of Bacille Calmette-Guerin (BCG), an attenuated strain of *Mycobacterium bovis* that is widely used as an effective intravesical therapy in bladder carcinoma. The mechanisms underlying the anti-tumour effects of BCG are unclear, but likely involve immune processes and/or direct cytotoxic effects on the tumour cells. BCG entry into bladder cancer cells is mediated by macropinocytosis, although not all bladder cancer cells are susceptible to treatment [7]. Sensitivity to BCG was identified to be associated with *PTEN* deletion,

which is a negative regulator of P13K signalling, or oncogenic mutations in *HRAS* or *KRAS*. These BCG-sensitive cells permitted the internalization of BCG via macropinocytosis, which was dependent upon Rho-family GTPases, such as Cdc42 and Rac1, as well as the kinase Pak1. In BCG-resistant bladder cancer cells, either oncogenic *HRAS* or *KRAS* ectopic expression or *PTEN* knockdown was sufficient to enhance BCG uptake. These important findings implicate macropinocytosis not only in supporting bladder tumours metabolically, but also in possibly representing a mode of BCG delivery to bladder tumours that can be dialed-up to optimize effectiveness. Indeed, a recent live-attenuated *Mycobacterium tuberculosis* vaccine that enters bladder cancer cells more readily through macropinocytosis than BCG, has been shown to be more effective in terms of its anti-tumour effects in an orthotopic murine model of bladder cancer [53]. Considering this information, it would be beneficial to explore ways to enhance macropinocytosis in order to improve microbial-based therapies.

The BCG-resistant bladder cancer cell lines, which display very low levels of macropinocytosis and do not harbour *RAS* or *PTEN* mutations, served as a platform to screen for signal transduction pathways that have the capacity to activate macropinocytosis when they are suppressed [18]. This whole-genome gain-of-function screen employed a flow cytometry-based read-out to measure uptake of fluorescent BCG and found that negative regulators of the canonical Wnt signalling pathway could activate macropinocytosis when their expression was suppressed by the shRNA-mediated knockdown. These macropinocytosis modulators, which included *DKK2*, *KREMEN1*, *NKDI1*, *SMAD4* and *MAPK9*, might be specific to wild-type Ras cells or to bladder cancer cells because they were not identified as modulators of macropinocytosis in oncogenic *HRAS*-expressing HeLa cells [54]. The activation of the Wnt pathway in various BCG-resistant bladder cancer cell lines was sufficient to support albumin-dependent proliferation under conditions of limiting nutrients, demonstrating that, as was observed in PDAC, macropinocytosis can support metabolic fitness in bladder cancer [18]. Additionally, the Wnt pathway seems to be constitutively activated in BCG-sensitive bladder cancer cells and in other *RAS*-transformed cells, supporting macropinocytosis in these contexts as well. It might be beneficial to establish an animal model of bladder cancer that can be scrutinized for macropinocytic induction *in vivo* and then examine whether Wnt pathway activation can be used to improve BCG therapy.

4. Lung cancer

About a quarter of non-small cell lung cancer (NSCLC) tumours harbour oncogenic *KRAS* mutations [55]. As has been observed in other cancer models, only some of these tumours are dependent on oncogenic *KRAS* expression for their viability—a phenomenon termed oncogene addiction [56]. Recent work exploring the role of macropinocytosis in NSCLC has been in the context of *KRAS* addiction [28]. NSCLC dependence upon oncogenic *KRAS* can be mediated by cell surface receptor integrin $\alpha\beta3$, and its modulator galectin-3. Integrin $\alpha\beta3$ clusters on the surface of tumour cells and plays a role in anchorage-independent growth, controlling tumour progression and metastasis [57]. Clustered integrin $\alpha\beta3$ can potentiate signal transduction, a function

influenced by its binding partner galectin-3, and together this interaction has the ability to induce *KRAS* clustering [58]. Interestingly, NSCLC cells that express integrin $\alpha\beta3$ are uniquely addicted to oncogenic *KRAS*, and these cells employ macropinocytosis to sustain their cellular fitness, while NSCLC cells that are not addicted to oncogenic *KRAS* exhibit low levels of uptake [28]. Macropinosomes produced in integrin $\alpha\beta3$ -positive cells drive the internalization of extracellular protein that is targeted for degradation. This process relies on integrin $\alpha\beta3$ and its modulator galectin-3, as depletion of either suppresses macropinocytic capacity. A clinically active inhibitor of galectin-3, GCS-100, blocks the integrin $\alpha\beta3$ /galectin-3 interaction, and as a consequence blocks macropinocytic uptake. GCS-100 has potent anti-tumour effects in integrin $\alpha\beta3$ -positive mouse models of NSCLC and PDAC; however, the contribution of macropinocytic suppression to this phenotype is unclear because GCS-100 also increases reactive oxygen species levels.

In addition to serum albumin, macropinocytosis may facilitate the internalization of extracellular ATP into lung cancer cells to support viability under conditions of nutrient stress [29]. Uptake of ATP could be physiologically important in the cancer setting because ATP levels can be much higher in tumours than in normal tissues [59]. In an NSCLC xenograft tumour model, the internalization of ATP is mediated by macropinocytosis, but might also involve other endocytic pathways, such as clathrin- and caveolae-mediated endocytosis [30]. Extracellular ATP is known to induce resistance to various chemotherapeutic drugs in cancer cells. Interestingly, drug resistance to sunitinib in NSCLC tumour cells is linked to macropinocytic uptake because it is suppressed by knockdown of *PAK1*, a known regulator of macropinocytosis [60]. The uptake of ATP via macropinocytosis is a prime example that macropinocytic cargo can vary in tumour cells, and depending on the context, identifying novel cargoes could provide clues to the breadth of function for macropinocytosis in cancer [46,61].

5. Prostate cancer

While oncogenic mutations in *RAS* are not associated with prostate cancer, *PTEN*, a negative regulator of P13K signalling, is the most frequently deleted tumour suppressor gene in prostate tumours. It was recently reported that *PTEN*-deficient prostate cancer cells exhibit macropinocytosis and that *PTEN* reconstitution suppresses uptake in these cells [38]. The macropinocytosis in this setting is mediated by AMPK, as a $\beta1$ subunit-specific AMPK activator, A769662, can transiently enhance macropinocytic capacity and inhibition of AMPK or expression of a dominant negative form of AMPK can suppress uptake. AMPK functions to control energy homeostasis, and its role in cancer has been extensively explored in the context of nutrient stress; however, the macropinocytic induction observed in *PTEN*-deficient prostate cancer cells is nutrient-independent. This might be accounted for by the finding that prostate tumours have high basal AMPK activity relative to normal prostate tissue [62]. Intriguingly, serum albumin uptake in *PTEN*-deficient prostate tumour cells seems to be independent of macropinocytosis, possibly relying on other endocytic pathways for its internalization [38]. Instead, necrotic cell debris is the nutritious macropinocytic cargo that is favoured by prostate

cancer cells. Under conditions of limiting amino acids, the uptake of necrotic cell debris significantly impacted the proliferative capacity of various prostate cancer cells, an activity that was suppressed by *PTEN* reconstitution. Not only does necrotic cell debris provide a source of amino acids, but it also serves to supply lipids that are critical to maintaining cellular membranes and intracellular lipid stores. Importantly, inhibiting macropinocytosis via EIPA administration suppressed prostate tumour growth in a prostate allograft model. Important avenues for further study include exploring to what extent AMPK regulates macropinocytosis in other pathological contexts and whether other tumour types also use necrotic cell debris as a nutrient source to fuel growth.

6. Brain and nervous system cancers

Enhanced activation of the Ras signalling pathway can drive macropinocytotic induction in glioblastoma cells [63]. Indeed, this characteristic has been co-opted to study the uptake of potentially therapeutic nanomedicines, especially those that can cross the blood–brain barrier [8]. Macropinocytosis in glioblastoma cells requires fine-tuned regulation because hyperstimulation of the pathway by either a boost in Ras signalling or by small molecules that drive the process can lead to a necrosis-like form of cell death called methuosis. Methuosis was first described as a cellular response to ectopic expression of oncogenic *HRAS* in glioblastoma cells, which induces extreme cytoplasmic vacuolization as a result of hyperactive macropinocytotic induction [9]. This peculiar trait might represent a unique vulnerability that can be targeted in glioblastoma. Similar cell death effects were observed when glioblastoma cells were treated with Bacoside A, a natural compound mixture [64]. Although the mechanism driving macropinocytosis in response to bacosides is unclear, it might involve calcium release from the smooth endoplasmic reticulum.

In addition to glioblastoma, macropinocytosis is also evident in neuroblastoma and medulloblastoma. In neuroblastoma cells, two distinct forms of macropinocytosis have been observed, both dependent on nonmuscle myosin II [12]. Insulin-like growth factor-1 induces macropinosomes that are predominantly located in the cell bodies and require P13K signalling for their formation, while phorbol 12-myristate 13-acetate induces macropinocytosis in the neurites that is P13K independent. Similar to glioblastoma, hyperstimulation of macropinocytosis in neuroblastoma cells leads to methuosis, suggesting this might be a common trait in nervous system cancers [65,66]. Indeed, in medulloblastoma cells, nerve growth factor activation of the tropomyosin receptor kinase A (TrkA) puts macropinocytosis in overdrive, stimulating vacuolar accumulation and cell death by methuosis [13,67].

7. Haematologic malignancies

Haematologic malignancies include forms of cancer that originate from cells of the immune system or in blood-forming tissue, such as bone marrow. It is not surprising that macropinocytosis is prevalent in these pathological states since, unlike solid tumours, this uptake mechanism is employed as an aspect of normal function in these cells. For example,

dendritic cells are thought to macropinocytose large quantities of exogenous solute as part of their sentinel function [68,69] and macropinocytosis in macrophages is linked to antigen surveillance, as well as foam cell formation through uptake of LDL [70,71]. It is unclear whether the macropinocytosis observed in malignancies such as leukaemia, lymphoma and multiple myeloma supports tumour cell metabolism; however, there are examples of this endocytic pathway being active in these settings, suggesting a possible role in treatment or progression of these diseases. Nanoparticle uptake via macropinocytosis has been demonstrated in an *in vitro* murine model of adult T-cell leukaemia/lymphoma and in myeloma cells [22,23]. Macropinocytosis has also been harnessed to deliver a cell-penetrating peptide with a lymph node-homing motif to leukaemia and lymphoma cells [24]. It is not clear whether macropinocytosis is critical to all haematological malignancies because dendritic cells obtained from chronic myeloid leukaemia patients displayed lower levels of uptake than dendritic cells from healthy individuals, although macropinocytotic capacity can still be used in these cells as a functional test [72,73]. It will be important to determine whether the inhibition of macropinocytosis has the ability to modulate the onset or progression of these diseases.

8. Future perspectives

The studies that have focused on the role of macropinocytosis in Ras-driven cancers have been centred on pancreatic, lung and bladder cancers. An important extension of this research is to explore whether macropinocytosis contributes to tumour cell metabolic fitness in other Ras-mutant tumours. For example, *KRAS* mutations are prevalent in colorectal cancer and these aggressive tumours do not respond to established chemotherapies [74]. An attractive hypothesis is that macropinocytosis in these tumours might represent a novel vulnerability that can be exploited as a treatment. Evidence that macropinocytosis is a feature of these tumours both *in vitro* and *in vivo* underscores this idea (table 1). In addition, oncogenic Ras mutations are prevalent in many other malignancies, including multiple myeloma, melanoma, uterine cancer, thyroid cancer, acute myeloid leukaemia and stomach cancer [75]. Future endeavours could be focused on delineating the possible role that macropinocytosis might have in augmenting metabolism in these cancers and evaluating whether macropinocytotic inhibition can be harnessed as a therapy in these settings.

In addition to *RAS* mutations, studies have linked the induction of macropinocytosis to other oncogenes, as well as tumour suppressors. EGFR activation, which acts upstream of wild-type Ras, is a classic example of a macropinocytotic driver; therefore, tumours with EGFR gain-of-function mutations or EGFR overexpression might present with enhanced macropinocytotic capacity. Overexpression of EGFR is an attribute of both triple-negative breast cancer and inflammatory breast cancer [76]. Studies in breast cancer cells have linked EGFR activation in response to growth factor stimuli to enhanced macropinocytotic uptake [77], but it is uncertain whether targeting macropinocytosis in breast cancer cells would have deleterious effects on viability. Along the same lines, macropinocytosis might be a robust feature of tumours with oncogenic mutations in other receptor tyrosine kinases, such as PDGFR and HER2/ErbB2 [78]. Another classic

example of oncogene-driven macropinocytosis is uptake caused by Src signal transduction. Activating mutations in c-Src, a non-receptor tyrosine kinase, are not very common in cancer, but enhanced pathway activation driven by elevated protein levels is observed in some colon, breast and prostate tumours [79]. Macropinocytosis depends on the activity of Rac1, which orchestrates the actin cytoskeleton dynamics that lead to membrane ruffling and macropinosome formation. Some sun-exposed melanomas harbour the *RAC1*^{P29S} mutation, which leads to Rac1 protein that exhibits enhanced inherent GDP/GTP nucleotide exchange [80]. Hyperactive Rac1^{P29S} leads to an enhanced macropinocytic response that is dependent on Dock1, a Rac-specific guanine nucleotide exchange factor implicated in a variety of cancers [14]. Interestingly, these studies showed that a Dock1-selective inhibitor suppresses macropinocytosis in these *RAC1*-mutant cells and future work will hopefully determine whether this compound has anti-tumour effects in preclinical models of melanoma. Similar to melanoma cells, some breast cancer cell lines harbour the *RAC1*^{P29S} mutation, potentially making these tumours susceptible to Dock1-inhibition [14].

The best-characterized tumour suppressor that controls macropinocytosis is *PTEN*. *PTEN* is a negative regulator of P13K signalling, which has a well-established role in controlling macropinocytosis. *PTEN* is one of the most frequently disrupted tumour suppressors in cancer and macropinocytosis induction driven by the loss of *PTEN* has been demonstrated in both bladder and prostate cancer (table 1). As *PTEN* is a phosphatase that suppresses P13K signalling, a loss in *PTEN* expression unleashes P13K-driven cascades that control actin reorganization, as well as macropinosome closure [81,82]. From this perspective, we would predict that tumours harbouring gain-of-function mutations that enhance

P13K signal potentiation are robustly macropinocytic. Oncogenic mutations in *PIK3CA*, which encodes the catalytic subunit of P13K, are quite common in cancer [83]. It might be conceivable that treatment strategies tailored for P13K aberrations are effective, at least in part, owing to their inhibitory properties affecting macropinocytosis induction.

Hyperstimulation of macropinocytosis might not be advantageous in terms of supporting tumour cell viability as it leads to methuosis [63]. This phenomenon has been mainly observed in blastomas that affect immature undifferentiated precursor cells in the brain and nervous system. Cell death is likely attributed to vacuolar expansion that is beyond the trafficking tolerances of these cells and as a result, disrupts endocytic homeostasis. It is not clear whether methuosis is uniquely a feature of blastoma cells; however, evidence that synthetic lethality underlies the mutual exclusivity of oncogenic *KRAS* and *EGFR* mutations in lung cancer suggests that such cell death might occur in other settings [84]. Indeed, forced expression of both oncogenes led to loss of viability that was associated with vacuolarization and increased macropinocytosis [84]. Altogether, these studies highlight that macropinocytosis is highly regulated in tumours and underscore important considerations when designing therapeutic strategies centred on macropinocytosis, especially from the perspective of metabolic interventions and nanomedicines.

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References

- Brunk U, Schellens J, Westermarck B. 1976 Influence of epidermal growth factor (EGF) on ruffling activity, pinocytosis and proliferation of cultivated human glia cells. *Exp. Cell Res.* **103**, 295–302. (doi:10.1016/0014-4827(76)90266-4)
- Haigler HT, McKanna JA, Cohen S. 1979 Rapid stimulation of pinocytosis in human carcinoma cells A-431 by epidermal growth factor. *J. Cell Biol.* **83**, 82–90. (doi:10.1083/jcb.83.1.82)
- Bar-Sagi D, Feramisco JR. 1986 Induction of membrane ruffling and fluid-phase pinocytosis in quiescent fibroblasts by ras proteins. *Science* **233**, 1061–1068. (doi:10.1126/science.3090687)
- Veithen A, Cupers P, Baudhuin P, Courtoy PJ. 1996 v-Src induces constitutive macropinocytosis in rat fibroblasts. *J. Cell Sci.* **109**, 2005–2012.
- Clarke M, Kayman SC. 1987 The axenic mutations and endocytosis in *Dictyostelium*. *Methods Cell Biol.* **28**, 157–176. (doi:10.1016/S0091-679X(08)61642-8)
- Commisso C *et al.* 2013 Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature* **497**, 633–637. (doi:10.1038/nature12138)
- Redelman-Sidi G, Iyer G, Solit DB, Glickman MS. 2013 Oncogenic activation of Pak1-dependent pathway of macropinocytosis determines BCG entry into bladder cancer cells. *Cancer Res.* **73**, 1156–1167. (doi:10.1158/0008-5472.CAN-12-1882)
- Huang JL *et al.* 2017 Lipoprotein-biomimetic nanostructure enables efficient targeting delivery of siRNA to Ras-activated glioblastoma cells via macropinocytosis. *Nat. Commun.* **8**, 15144. (doi:10.1038/ncomms15144)
- Overmeyer JH, Kaul A, Johnson EE, Maltese WA. 2008 Active Ras triggers death in glioblastoma cells through hyperstimulation of macropinocytosis. *Mol. Cancer Res.* **6**, 965–977. (doi:10.1158/1541-7786.MCR-07-2036)
- Mo L, Hou L, Guo D, Xiao X, Mao P, Yang X. 2012 Preparation and characterization of teniposide PLGA nanoparticles and their uptake in human glioblastoma U87MG cells. *Int. J. Pharm.* **436**, 815–824. (doi:10.1016/j.ijpharm.2012.07.050)
- Ruan S *et al.* 2015 Noninvasive *in vivo* diagnosis of brain glioma using RGD-decorated fluorescent carbonaceous nanospheres. *J. Biomed. Nanotechnol.* **11**, 2148–2157. (doi:10.1166/jbn.2015.2105)
- Jiang J, Kolpak AL, Bao ZZ. 2010 Myosin IIB isoform plays an essential role in the formation of two distinct types of macropinosomes. *Cytoskeleton Hoboken* **67**, 32–42. (doi:10.1002/cm.20419)
- Li C, Macdonald JI, Hryciw T, Meakin SO. 2010 Nerve growth factor activation of the TrkA receptor induces cell death, by macropinocytosis, in medulloblastoma Daoy cells. *J. Neurochem.* **112**, 882–899. (doi:10.1111/j.1471-4159.2009.06507.x)
- Tomino T *et al.* 2018 DOCK1 inhibition suppresses cancer cell invasion and macropinocytosis induced by self-activating Rac1^{P29S} mutation. *Biochem. Biophys. Res. Commun.* **497**, 298–304. (doi:10.1016/j.bbrc.2018.02.073)
- Kapoor M, Burgess DJ. 2013 Cellular uptake mechanisms of novel anionic siRNA lipoplexes. *Pharm. Res.* **30**, 1161–1175. (doi:10.1007/s11095-012-0952-9)
- Corsi F *et al.* 2009 Towards ideal magnetofluorescent nanoparticles for bimodal detection of breast-cancer cells. *Small* **5**, 2555–2564. (doi:10.1002/smll.200900881)
- Delpout S, Sisson G, Black KM, Richardson CD. 2017 Measles virus enters breast and colon cancer cell lines through a PVRL4-mediated macropinocytosis pathway. *J. Virol.* **91**, 10. (doi:10.1128/JVI.02191-16)

18. Redelman-Sidi G *et al.* 2018 The canonical Wnt pathway drives macropinocytosis in cancer. *Cancer Res.* **78**, 4658–4670. (doi:10.1158/0008-5472.CAN-17-3199)
19. Zhu BY *et al.* 2017 A new HDAC inhibitor cinnamoylphenazine shows antitumor activity in association with intensive macropinocytosis. *Oncotarget* **8**, 14 748–14 758. (doi:10.18632/oncotarget.14714)
20. Sung S, Choi J, Cheong H. 2015 Catabolic pathways regulated by mTORC1 are pivotal for survival and growth of cancer cells expressing mutant Ras. *Oncotarget* **6**, 40 405–40 417. (doi:10.18632/oncotarget.6334)
21. Dubey RD *et al.* 2017 Novel hyaluronic acid conjugates for dual nuclear imaging and therapy in CD44-expressing tumors in mice *in vivo*. *Nanotheranostics* **1**, 59–79. (doi:10.7150/ntno.17896)
22. Watanabe K *et al.* 2011 The use of cationic nanogels to deliver proteins to myeloma cells and primary T lymphocytes that poorly express heparan sulfate. *Biomaterials* **32**, 5900–5905. (doi:10.1016/j.biomaterials.2011.04.058)
23. Hou KK, Pan H, Ratner L, Schlesinger PH, Wickline SA. 2013 Mechanisms of nanoparticle-mediated siRNA transfection by melittin-derived peptides. *ACS Nano* **7**, 8605–8615. (doi:10.1021/nn403311c)
24. Nishimura S *et al.* 2008 Combinatorial targeting of the macropinocytotic pathway in leukemia and lymphoma cells. *J. Biol. Chem.* **283**, 11 752–11 762. (doi:10.1074/jbc.M708849200)
25. Shao D, Li J, Guan F, Pan Y, Xiao X, Zhang M, Chen L. 2014 Selective inhibition of liver cancer growth realized by the intrinsic toxicity of a quantum dot-lipid complex. *Int. J. Nanomedicine.* **9**, 5753–5769. (doi:10.2147/IJN.S73185)
26. Hattori Y, Nakamura T, Ohno H, Fujii N, Maitani Y. 2013 siRNA delivery into tumor cells by lipid-based nanoparticles composed of hydroxyethylated cholesteryl triamine. *Int. J. Pharm.* **443**, 221–229. (doi:10.1016/j.ijpharm.2012.12.017)
27. Beaudet D, Badilescu S, Kuruvishetti K, Sahrabi Kashani A, Jaunky D, Ouellette S, Piekny A, Packirisamy M. 2017 Comparative study on cellular entry of incinerated ancient gold particles (Swarna Bhasma) and chemically synthesized gold particles. *Sci. Rep.* **7**, 10678. (doi:10.1038/s41598-017-10872-3)
28. Seguin L *et al.* 2017 Galectin-3, a druggable vulnerability for KRAS-addicted cancers. *Cancer Discov.* **7**, 1464–1479. (doi:10.1158/2159-8290.CD-17-0539)
29. Qian Y *et al.* 2014 Extracellular ATP is internalized by macropinocytosis and induces intracellular ATP increase and drug resistance in cancer cells. *Cancer Lett.* **351**, 242–251. (doi:10.1016/j.canlet.2014.06.008)
30. Qian Y, Wang X, Li Y, Cao Y, Chen X. 2016 Extracellular ATP a new player in cancer metabolism: NSCLC cells internalize ATP *in vitro* and *in vivo* using multiple endocytic mechanisms. *Mol. Cancer Res.* **14**, 1087–1096. (doi:10.1158/1541-7786.MCR-16-0118)
31. Patel D, Rorbach J, Downes K, Szukszto MJ, Pekalski ML, Minczuk M. 2017 Macropinocytic entry of isolated mitochondria in epidermal growth factor-activated human osteosarcoma cells. *Sci. Rep.* **7**, 12886. (doi:10.1038/s41598-017-13227-0)
32. Manara MC *et al.* 2016 CD99 triggering induces methuosis of Ewing sarcoma cells through IGF-1R/RAS/Rac1 signaling. *Oncotarget* **7**, 79 925–79 942. (doi:10.18632/oncotarget.13160)
33. van Bracht E *et al.* 2014 Specific targeting of tumor cells by lyophilisomes functionalized with antibodies. *Eur. J. Pharm. Biopharm.* **87**, 80–89. (doi:10.1016/j.ejpb.2014.01.005)
34. Holzer AK, Howell SB. 2006 The internalization and degradation of human copper transporter 1 following cisplatin exposure. *Cancer Res.* **66**, 10 944–10 952. (doi:10.1158/0008-5472.CAN-06-1710)
35. Kamphorst JJ *et al.* 2015 Human pancreatic cancer tumors are nutrient poor and tumor cells actively scavenge extracellular protein. *Cancer Res.* **75**, 544–553. (doi:10.1158/0008-5472.CAN-14-2211)
36. Baek G *et al.* 2014 MCT4 defines a glycolytic subtype of pancreatic cancer with poor prognosis and unique metabolic dependencies. *Cell Reports* **9**, 2233–2249. (doi:10.1016/j.celrep.2014.11.025)
37. Kamerkar S *et al.* 2017 Exosomes facilitate therapeutic targeting of oncogenic KRAS in pancreatic cancer. *Nature* **546**, 498–503. (doi:10.1038/nature22341)
38. Kim SM *et al.* 2018 PTEN deficiency and AMPK activation promote nutrient scavenging and anabolism in prostate cancer cells. *Cancer Discov.* **8**, 866–883. (doi:10.1158/2159-8290.CD-17-1215)
39. Sahay G, Alakhova DY, Kabanov AV. 2010 Endocytosis of nanomedicines. *J. Control Release.* **145**, 182–195. (doi:10.1016/j.jconrel.2010.01.036)
40. Akinc A, Battaglia G. 2013 Exploiting endocytosis for nanomedicines. *Cold Spring Harb. Perspect. Biol.* **5**, a016980. (doi:10.1101/cshperspect.a016980)
41. Garnacho C. 2016 Intracellular drug delivery: mechanisms for cell entry. *Curr. Pharm. Des.* **22**, 1210–1226. (doi:10.2174/1381612822666151216151021)
42. Jones AT. 2007 Macropinocytosis: searching for an endocytic identity and role in the uptake of cell penetrating peptides. *J. Cell. Mol. Med.* **11**, 670–684. (doi:10.1111/j.1582-4934.2007.00062.x)
43. Jones AT. 2008 Gateways and tools for drug delivery: endocytic pathways and the cellular dynamics of cell penetrating peptides. *Int. J. Pharm.* **354**, 34–38. (doi:10.1016/j.ijpharm.2007.10.046)
44. Hillaireau H, Couvreur P. 2009 Nanocarriers' entry into the cell: relevance to drug delivery. *Cell. Mol. Life Sci.* **66**, 2873–2896. (doi:10.1007/s00018-009-0053-z)
45. Overmeyer JH, Young AM, Bhanot H, Maltese WA. 2011 A chalcone-related small molecule that induces methuosis, a novel form of non-apoptotic cell death, in glioblastoma cells. *Mol. Cancer.* **10**, 69. (doi:10.1186/1476-4598-10-69)
46. Recouvreur MV, Commisso C. 2017 Macropinocytosis: a metabolic adaptation to nutrient stress in cancer. *Front. Endocrinol. (Lausanne)* **8**, 261. (doi:10.3389/fendo.2017.00261)
47. Smit VT, Boot AJ, Smits AM, Fleuren GJ, Cornelisse CJ, Bos JL. 1988 KRAS codon 12 mutations occur very frequently in pancreatic adenocarcinomas. *Nucleic Acids Res.* **16**, 7773–7782. (doi:10.1093/nar/16.16.7773)
48. Hingorani SR, Wang L, Multani AS, Combs C, Deramandt TB, Hruban RH, Rustgi AK, Chang S, Tuveson DA. 2005 Trp53R172H and KrasG12D cooperate to promote chromosomal instability and widely metastatic pancreatic ductal adenocarcinoma in mice. *Cancer Cell.* **7**, 469–483. (doi:10.1016/j.ccr.2005.04.023)
49. Nolan-Stevaux O, Lau J, Truitt ML, Chu GC, Hebrok M, Fernandez-Zapico ME, Hanahan D. 2009 *GLI1* is regulated through Smoothed-independent mechanisms in neoplastic pancreatic ducts and mediates PDAC cell survival and transformation. *Genes Dev.* **23**, 24–36. (doi:10.1101/gad.1753809)
50. Ivanov AI. 2008 Pharmacological inhibition of endocytic pathways: is it specific enough to be useful? *Methods Mol. Biol.* **440**, 15–33. (doi:10.1007/978-1-59745-178-9_2)
51. Davidson SM *et al.* 2017 Direct evidence for cancer-cell-autonomous extracellular protein catabolism in pancreatic tumors. *Nat. Med.* **23**, 235–241. (doi:10.1038/nm.4256)
52. Visvanathan KV, Pockock RD, Summerhayes IC. 1988 Preferential and novel activation of H-ras in human bladder carcinomas. *Oncogene Res.* **3**, 77–86.
53. Alvarez-Arguedas S *et al.* 2018 Therapeutic efficacy of the live-attenuated *Mycobacterium tuberculosis* vaccine, MTBVAC, in a preclinical model of bladder cancer. *Transl. Res.* **197**, 32–42. (doi:10.1016/j.trsl.2018.03.004)
54. Fennell M, Commisso C, Ramirez C, Garripa R, Barsagi D. 2015 High-content, full genome siRNA screen for regulators of oncogenic *HRAS*-driven macropinocytosis. *Assay Drug Dev. Technol.* **13**, 347–355. (doi:10.1089/adt.2015.660)
55. Siegfried JM *et al.* 1997 Prognostic value of specific KRAS mutations in lung adenocarcinomas. *Cancer Epidemiol. Biomarkers Prev.* **6**, 841–847.
56. Singh A *et al.* 2009 A gene expression signature associated with 'K-Ras addiction' reveals regulators of EMT and tumor cell survival. *Cancer Cell.* **15**, 489–500. (doi:10.1016/j.ccr.2009.03.022)
57. Desgrosellier JS, Cheresh DA. 2010 Integrins in cancer: biological implications and therapeutic opportunities. *Nat. Rev. Cancer* **10**, 9–22. (doi:10.1038/nrc2748)
58. Seguin L *et al.* 2014 An integrin β_3 -KRAS-RalB complex drives tumour stemness and resistance to EGFR inhibition. *Nat. Cell Biol.* **16**, 457–468. (doi:10.1038/ncb2953)
59. Pellegatti P, Raffaghello L, Bianchi G, Piccardi F, Pistoia V, Di Virgilio F. 2008 Increased level of extracellular ATP at tumor sites: *in vivo* imaging with plasma membrane luciferase. *PLoS ONE* **3**, e2599. (doi:10.1371/journal.pone.0002599)

60. Wang X *et al.* 2017 Extracellular ATP, as an energy and phosphorylating molecule, induces different types of drug resistances in cancer cells through ATP internalization and intracellular ATP level increase. *Oncotarget* **8**, 87 860–87 877. (doi:10.18632/oncotarget.21231)
61. Comisso C, Debnath J. 2018 Macropinocytosis fuels prostate cancer. *Cancer Discov.* **8**, 800–802. (doi:10.1158/2159-8290.CD-18-0513)
62. Khan AS, Frigo DE. 2017 A spatiotemporal hypothesis for the regulation, role, and targeting of AMPK in prostate cancer. *Nat. Rev. Urol.* **14**, 164–180. (doi:10.1038/nrurol.2016.272)
63. Barth RF, Kaur B. 2009 Rat brain tumor models in experimental neuro-oncology: the C6, 9L, T9, RG2, F98, BT4C, RT-2 and CNS-1 gliomas. *J. Neurooncol.* **94**, 299–312. (doi:10.1007/s11060-009-9875-7)
64. John S, Sivakumar KC, Mishra R. 2017 Bacoside A induces tumor cell death in human glioblastoma cell lines through catastrophic macropinocytosis. *Front. Mol. Neurosci.* **10**, 171. (doi:10.3389/fnmol.2017.00171)
65. Nara A, Aki T, Funakoshi T, Uemura K. 2010 Methamphetamine induces macropinocytosis in differentiated SH-SY5Y human neuroblastoma cells. *Brain Res.* **1352**, 1–10. (doi:10.1016/j.brainres.2010.07.043)
66. Nara A, Aki T, Funakoshi T, Uema K, Uemura K. 2012 Hyperstimulation of macropinocytosis leads to lysosomal dysfunction during exposure to methamphetamine in SH-SY5Y cells. *Brain Res.* **1466**, 1–14. (doi:10.1016/j.brainres.2012.05.017)
67. Li C *et al.* 2016 Unravelling the mechanism of TrkA-induced cell death by macropinocytosis in medulloblastoma Daoy cells. *Mol. Cell. Biol.* **36**, 2596–2611. (doi:10.1128/MCB.00255-16)
68. Norbury CC. 2006 Drinking a lot is good for dendritic cells. *Immunology* **117**, 443–451. (doi:10.1111/j.1365-2567.2006.02335.x)
69. West MA, Prescott AR, Eskelinen EL, Ridley AJ, Watts C. 2000 Rac is required for constitutive macropinocytosis by dendritic cells. *Curr. Biol.* **10**, 839–848. (doi:10.1016/S0960-9822(00)00595-9)
70. Michael DR *et al.* 2013 Differential regulation of macropinocytosis in macrophages by cytokines: implications for foam cell formation and atherosclerosis. *Cytokine* **64**, 357–361. (doi:10.1016/j.cyto.2013.05.016)
71. Kruth HS, Jones NL, Huang W, Zhao B, Ishii I, Chang J, Combs CA, Malide D, Zhang W-Y. 2005 Macropinocytosis is the endocytic pathway that mediates macrophage foam cell formation with native low density lipoprotein. *J. Biol. Chem.* **280**, 2352–2360. (doi:10.1074/jbc.M407167200)
72. Dong R *et al.* 2003 Dendritic cells from CML patients have altered actin organization, reduced antigen processing, and impaired migration. *Blood* **101**, 3560–3567. (doi:10.1182/blood-2002-06-1841)
73. Heinzinger M, Waller CF, von den Berg A, Rosenstiel A, Lange W. 1999 Generation of dendritic cells from patients with chronic myelogenous leukemia. *Ann. Hematol.* **78**, 181–186. (doi:10.1007/s002770050497)
74. Tan C, Du X. 2012 KRAS mutation testing in metastatic colorectal cancer. *World J. Gastroenterol.* **18**, 5171–5180. (doi:10.3748/wjg.v18.i37.5171)
75. Hobbs GA, Der CJ, Rossman KL. 2016 RAS isoforms and mutations in cancer at a glance. *J. Cell Sci.* **129**, 1287–1292. (doi:10.1242/jcs.182873)
76. Masuda H, Zhang D, Bartholomeusz C, Doihara H, Hortobagyi GN, Ueno NT. 2012 Role of epidermal growth factor receptor in breast cancer. *Breast Cancer Res. Treat.* **136**, 331–345. (doi:10.1007/s10549-012-2289-9)
77. Bryant DM *et al.* 2007 EGF induces macropinocytosis and SNX1-modulated recycling of E-cadherin. *J. Cell Sci.* **120**, 1818–1828. (doi:10.1242/jcs.000653)
78. Anton IM *et al.* 2003 WIP participates in actin reorganization and ruffle formation induced by PDGF. *J. Cell Sci.* **116**, 2443–2451. (doi:10.1242/jcs.00433)
79. Ishizawa R. 2004 Parsons SJ. c-Src and cooperating partners in human cancer. *Cancer Cell.* **6**, 209–214. (doi:10.1016/j.ccr.2004.09.001)
80. Davis MJ, Ha BH, Holman EC, Halaban R, Schlessinger J, Boggon TJ. 2013 RAC1^{P29S} is a spontaneously activating cancer-associated GTPase. *Proc. Natl Acad. Sci. USA* **110**, 912–917. (doi:10.1073/pnas.1220895110)
81. Hoeller O, Bolourani P, Clark J, Stephens LR, Hawkins PT, Weiner OD, Weeks G, Kay RR. 2013 Two distinct functions for PI3-kinases in macropinocytosis. *J. Cell Sci.* **126**, 4296–4307. (doi:10.1242/jcs.134015)
82. Araki N, Johnson MT, Swanson JA. 1996 A role for phosphoinositide 3-kinase in the completion of macropinocytosis and phagocytosis by macrophages. *J. Cell Biol.* **135**, 1249–1260. (doi:10.1083/jcb.135.5.1249)
83. Samuels Y, Waldman T. 2010 Oncogenic mutations of PIK3CA in human cancers. *Curr. Top. Microbiol. Immunol.* **347**, 21–41. (doi:10.1007/82_2010_68)
84. Unni AM, Lockwood WW, Zejnullahu K, Lee-Lin SQ, Varmus H. 2015 Evidence that synthetic lethality underlies the mutual exclusivity of oncogenic KRAS and EGFR mutations in lung adenocarcinoma. *Elife* **4**, e06907. (doi:10.7554/elife.06907)