

RESEARCH ARTICLE

Carbonylation accumulation of the *Hypsibius exemplaris* anhydrobiote reveals age-associated marks

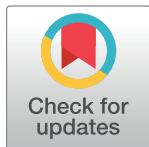
Mira Kuzmic¹*, Myriam Richaud^{1,2}, Pierre Cuq², Sandrine Frelon¹‡, Simon Galas^{1,2}*[†]

1 IRSN/PSE-ENV/SRTE - Laboratoire d'écotoxicologie des radionucléides, St Paul lez Durance Cedex, France, **2** IBMM, University of Montpellier, CNRS, ENSCM, Montpellier, France

* These authors contributed equally to this work.

‡ These authors are last authors on this work.

* simon.galas@umontpellier.fr



Abstract

Together with nematodes and rotifers, tardigrade belong to micrometazoans that can cope with environmental extremes such as UV and solar radiations, dehydration, supercooling or overheating. Tardigrade can resist the harshest conditions by turning to cryptobiosis, an anhydrobiotic state that results from almost complete dehydration and is characterized by an ametabolic status. Although reports have challenged the molecular basis of the mechanisms underlying genomic injury resistance, little is yet known regarding the possible involvement of other tardigrade macromolecules in injury during a stress experience. In this report, we show that the tardigrade *Hypsibius exemplaris* can accumulate molecular damages by means of *in situ* detection of carbonyls. Furthermore, we demonstrate that living tardigrade can accumulate carbonylation. Finally, we reveal that anhydrobiotic tardigrade can be constitutively affected by carbonylation that marks aging in other metazoans.

OPEN ACCESS

Citation: Kuzmic M, Richaud M, Cuq P, Frelon S, Galas S (2018) Carbonylation accumulation of the *Hypsibius exemplaris* anhydrobiote reveals age-associated marks. PLoS ONE 13(12): e0208617. <https://doi.org/10.1371/journal.pone.0208617>

Editor: Michael R Volkert, University of Massachusetts Medical School, UNITED STATES

Received: May 18, 2018

Accepted: November 20, 2018

Published: December 26, 2018

Copyright: © 2018 Kuzmic et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: Part of this work was funded by the grant N° 265880- Giga18 of the AAP—Défi Origines 2018 funding program of the CNRS (<http://www.cnrs.fr/mi/spip.php?article1345>) for SG.

Competing interests: The authors have declared that no competing interests exist.

Introduction

Tardigrade are minute metazoan animals, ranging from 0.1 to 1.2 mm in size, that inhabit diverse habitats such as freshwater, marine or the water films of terrestrial moss and lichens. Approximately 1000 species of tardigrade have been identified to date [1–3], and several have been reported to have the ability to cope with the harshest environments [4]. For example, they can survive up to 10 kJ/m² UV exposure [5], a ten-day space flight exposure to solar radiations at low earth orbit in the space vacuum [6], or exposure of up to 7.5 GPa pressure [7], which equals the pressure at a depth of up to 180 km from the earth's surface. Moreover, tardigrade can withstand treatment with organic solvents [8], extreme temperatures (ranging from -272 to 151 °C) or high radiation doses (kGy) [9].

However, all these observations of tardigrade extreme resistance have been made during a latent stage of life [10–12] that forms by dehydration and allows for the resistance to desiccation. When they lose their surrounding water film, terrestrial tardigrade initiate contractions along their anterior-posterior body axis and retract their limbs to form an anhydrobiotic

organism with a characteristic “tun” shape. Tardigrade can then lose up to 97% of their bound and free body water content during the anhydrobiosis process [13–15]. Together with small organisms, such as rotifera and nematoda, tardigrade can enter anhydrobiosis at any stage of their life cycle [16–22].

Reactive oxygen species (ROS) originate from intracellular sources, mainly mitochondria, and are biproducts of energy metabolism. ROS are highly unstable oxygen derivative molecules that can damage, by oxidation, various intra-cellular molecules such as lipids, proteins, sugars and nucleic acids (DNA, RNA) either directly or indirectly, thereby reducing the normal activity and function of numerous macromolecular components [23]. Thymidine dimer formation is believed to be the main DNA injury caused by UVC [24]. Moreover, gamma radiation and UV light can induce ROS formation [25,26], resulting in damaged cellular macromolecules. For example, up to 70% of the DNA damage can be attributed to ROS formation when water radiolysis occurs upon radiation [24]. The main ROS-target of tardigrade assessed to date concern the DNA molecule [27] but ROS-damaged proteins can also accumulate as carbonylated products which may impact normal biochemical activity and are associated with aging and human degenerative diseases [28,29].

Because of the admittedly high resistance of the tardigrade to several of the harshest treatments, we then assessed if the tardigrade *Hypsibius exemplaris* can accumulate carbonylations during the anhydrobiotic or fully active period. Here, we used a new technique that allows the detection of total carbonylated proteins [30]. Thereby, we showed that the tardigrade *Hypsibius exemplaris* can accumulate carbonylations during both the anhydrobiotic and hydrated periods. Furthermore, we uncovered a gradual accumulation of carbonylation as desiccated tardigrade aged.

Results

Carbonylation in active tardigrade induced by UVC

Living tardigrade exposed to rising doses of UVC revealed a carbonyl content increase (Fig 1A). We measured a high correlation degree (linear regression analysis: $y = 0.195x + 10.75$; $R^2 = 0.998$; $P < 0.05$ and Spearman correlation coefficient $\rho = 0.76$) between the carbonylation detection and the UVC dose of the irradiated group. Moreover, while the carbonyl accumulation with a 60 kJ.m^{-2} UVC irradiation dose did not differ significantly from the control, both UVC dose 120 kJ.m^{-2} and 180 kJ.m^{-2} showed a significant difference (Kruskall-Wallis test, $p < 0.05$). With the fluorescence background staining of the control group, we observed an almost 5-fold increase in the carbonylation staining for the 180 kJ.m^{-2} irradiation condition (Fig 1A). Interestingly, the carbonyl content detected did not plateau between the 120 kJ.m^{-2} and 180 kJ.m^{-2} irradiation doses (Fig 1A). This observation indicates that perhaps the accumulation of UVC-related carbonylation damage may override the highest measured level in the present study.

Fig 2 shows representative images of direct *in situ* carbonyl labeling. We detected a faint carbonylation signal when active tardigrade were irradiated with 60 kJ.m^{-2} UVC (Fig 2B). However, *in situ* carbonylation staining progressively strengthened with 120 (Fig 2C) and 180 kJ.m^{-2} (Fig 2D) UVC irradiation doses, illustrating the correlation between the delivered UVC irradiation dose and carbonyl staining. Moreover, no active tardigrade survived exposure to the UVC radiation doses assessed here.

UVC irradiation can induce carbonylation in anhydrobiotes

Anhydrobiotic tardigrade result from physiological and molecular modifications at a deep level. We then assessed if dehydrated tardigrade can accumulate carbonyl signatures to the

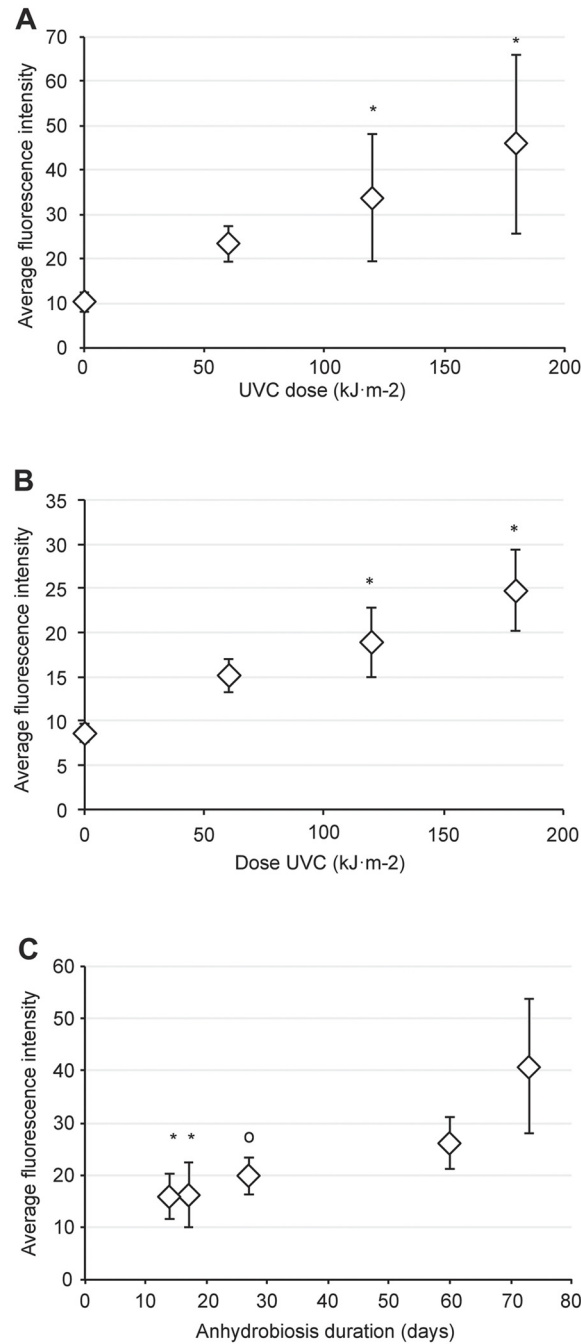


Fig 1. Carbonylation accumulation of the *Hypsibius exemplaris* tardigrade. Average intensity of the *in situ* carbonylation staining of active (A) and anhydrobiote (B) tardigrade in response to rising UVC doses or during anhydrobiote aging (C). Errors bars indicate the standard deviation from at least three repeated experiments. Total animals scored was $n = 34$ for (A), $n = 20$ for (B) and $n = 37$ for (C). The asterisk represents the respective significant difference of the Kruskal-Wallis test at $P < 0.05$. The respective curve correlation degree are $y = 0.195x + 10.75$; $R^2 = 0.998$ for (A), $y = 0.087x + 9.07$; $R^2 = 0.990$ for (B) and $y = 0.36x + 9.92$; $R^2 = 0.88$ for (C). The respective Spearman's correlation coefficient are $P < 0.05$, $\rho = 0.76$ for (A) and $P < 0.05$, $\rho = 0.91$ for (B). The mini-circle marks a less significant difference (Kruskal-Wallis's test, $P < 0.1$) with carbonyl accumulation of the desiccated tardigrade at 73 days (C).

<https://doi.org/10.1371/journal.pone.0208617.g001>

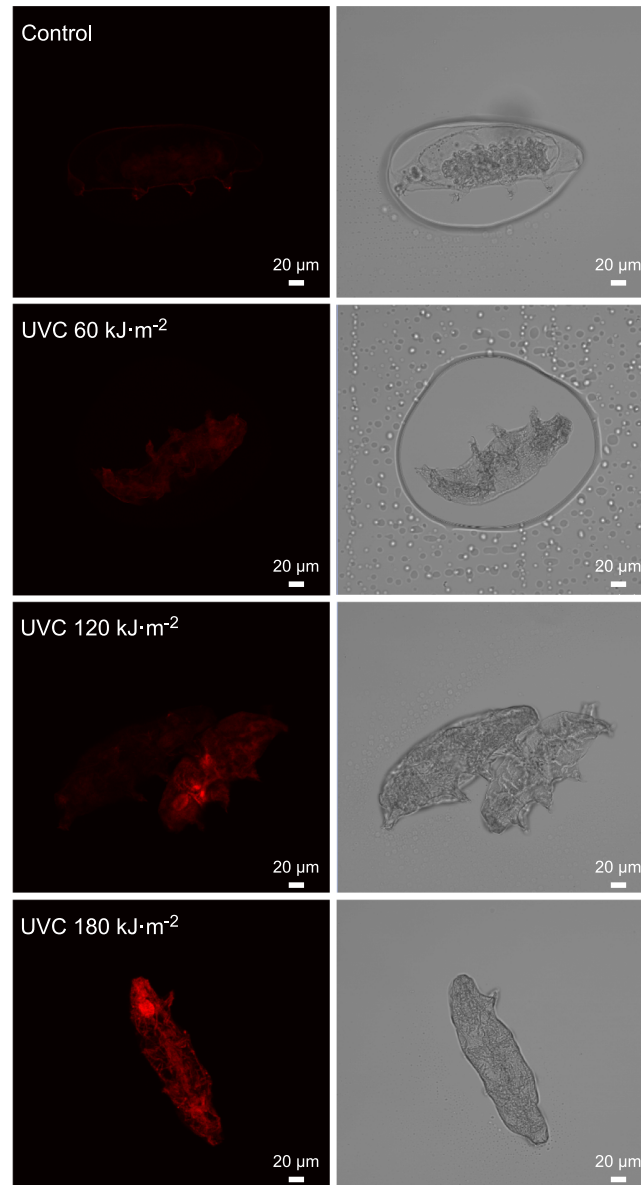


Fig 2. Confocal detection of carbonylation upon exposure of the tardigrade to rising UVC doses during the hydrated state. The pictures are representative of the three independent experiments depicted in Fig 1A. Scale bar = 100 μm .

<https://doi.org/10.1371/journal.pone.0208617.g002>

same extent as the active tardigrade. We observed a positive link between the rising dose of UVC exposure and the carbonyl content in desiccated tardigrade (Fig 1B). Furthermore, we noticed a high degree of correlation between the UVC radiation dose and the carbonyl signal (linear regression analysis: $y = 0.087x + 9.07$; $R^2 = 0.990$ and Spearman correlation coefficient $P < 0.05$, $\rho = 0.91$). Moreover, similar to the living tardigrade groups observed in the above section, we detected a highly significant difference in the carbonyl content between the control group and the $120 \text{ kJ}\cdot\text{m}^{-2}$ or $180 \text{ kJ}\cdot\text{m}^{-2}$ UVC irradiated groups (Kruskall-Wallis test, $p < 0.05$), which showed a 2- and 2.5-fold increase in carbonyl content, respectively. Conversely, the

tardigrade group that received a 60 kJ.m^{-2} UVC radiation dose did not differ in carbonyl content from the control group.

Fig 3 shows selected images of the direct *in situ* carbonyl staining of anhydrobiotic tardigrade. We observed a UVC dose-dependent increase in carbonyl labeling with a faint carbonylation staining for the 60 kJ.m^{-2} UVC irradiation dose (Fig 3B), followed by stronger carbonyl labeling for the 120 kJ.m^{-2} (Fig 3C) and 180 kJ.m^{-2} (Fig 3D) UVC irradiation doses.

Because of the absence of a plateau in the carbonyl staining curve observed between the 120 kJ.m^{-2} and 180 kJ.m^{-2} UVC irradiation doses, it is plausible that the maximal UVC irradiation dose assessed in this study did not allow for the highest possible carbonylation content of the anhydrobiotic tardigrade to be reached. Nevertheless, we observed up to a two-fold increase in total carbonylation (Fig 1B) in the irradiated groups of desiccated tardigrade compared to that in the control group of desiccated tardigrade.

UVC impacted both the survival and progeny hatching of anhydrobiotic tardigrade

The UVC-dependent carbonyl measured in the anhydrobiotes prompted us to evaluate their possible impact during the tardigrade rehydration process. Fig 4A shows the anhydrobiotic exit of desiccated tardigrade subjected to various UVC irradiation doses. We observed that compared with the rehydrated and active tardigrade of the control group, the rehydrated and active tardigrade of the 60 kJ.m^{-2} UVC dose group diminished by a two-fold factor. Moreover, a comparison between both tardigrade groups showed a significant difference (Paired Z-test, $P < 0.05$, p -value = 0.029).

Likewise, we noticed that the time needed for the tardigrade group subjected to the 60 kJ.m^{-2} UVC irradiation dose to fully rehydrate was postponed by 19 hours compared with that needed for the control group. This means that perhaps UVC irradiation either directly or indirectly impaired the anhydrobiotic exit process.

In contrast, both 120 and 180 kJ.m^{-2} UVC irradiation doses resulted in a total failure of the anhydrobiotes to fully rehydrate, and these groups died. However, we recovered a unique individual from the group of dehydrated tardigrade irradiated with the 120 kJ.m^{-2} UVC dose who started rehydration with a normal appearance but instead retained an uncommon gray color and a failure to regain any motility until 5 days post rehydration.

As we have shown, the anhydrobiotic tardigrade group irradiated with the 60 kJ.m^{-2} UVC dose yielded a survivor fraction during the rehydration process assay (Fig 4A). Based on this observation, we then assessed if the brood size was also affected. As shown in Fig 4B, we observed comparable egg production in both the control and the 60 kJ.m^{-2} UVC irradiated tardigrade groups.

However, even though it was normal in appearance, egg production can be concomitant with an impairment in the egg hatchability. Fig 4C depicts the hatchability of the eggs laid by the control and 60 kJ.m^{-2} UVC irradiated groups. We detected a drastic decrease in the hatchability of the eggs laid by the irradiated tardigrade group compared with those laid by the control group (Fig 4C). However, we observed that the irradiated anhydrobiotes can produce viable embryos even after exposure to a high UVC dose up to 60 kJ.m^{-2} .

Constitutive carbonylation accumulation of desiccated tardigrade

In this report, we have shown that both hydrated and anhydrobiotic tardigrade can show a detectable carbonyl signature in response to the UVC irradiation doses that we assessed. We then investigated if the anhydrobiotic tardigrade may constitutively accumulate carbonyls. To conduct this investigation, we left groups of dehydrated tardigrade for different times (see

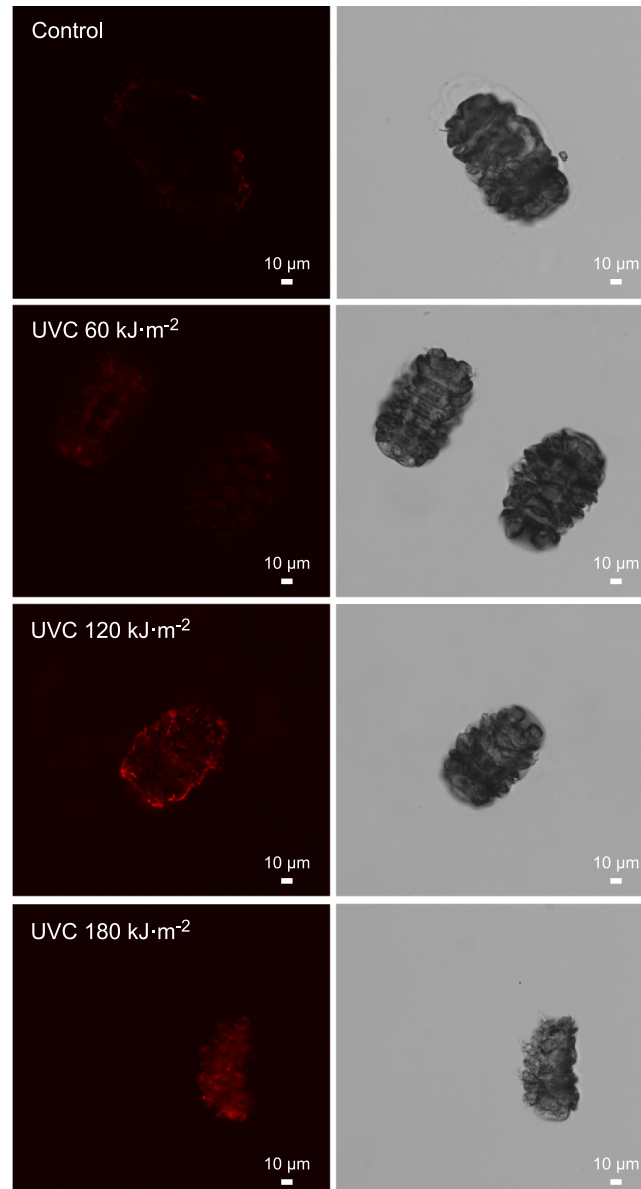


Fig 3. Confocal detection of carbonylation in the tardigrade anhydrobiotes in response to UVC. Representative images corresponding to the three independent experiments plotted in Fig 1B. Scale bar = 100 μm .

<https://doi.org/10.1371/journal.pone.0208617.g003>

Materials and methods section) and then subjected the groups to the *in situ* carbonyl detection labeling. As shown in Fig 1C, we observed that tardigrade anhydrobiotes progressively accumulated a carbonyl signature. Moreover, we determined that carbonyl labeling levels are well correlated with the time spent in the anhydrobiotic state. Thus, we noted that carbonyl labeling remained at approximately the same intensity level after both 14 (mean value 15.9) and 17 (mean value 16.3) days of anhydrobiosis (Fig 1C). Next, the carbonyl staining increased over 27 days (mean value 19.9) at 60 days (mean value 26.1) of anhydrobiosis, the staining reached almost twice the value observed at 14 days (mean value 15.9). Finally, at 73 days (mean value 40.8) of anhydrobiosis, we observed a value twice that of the one observed at 27 days (mean

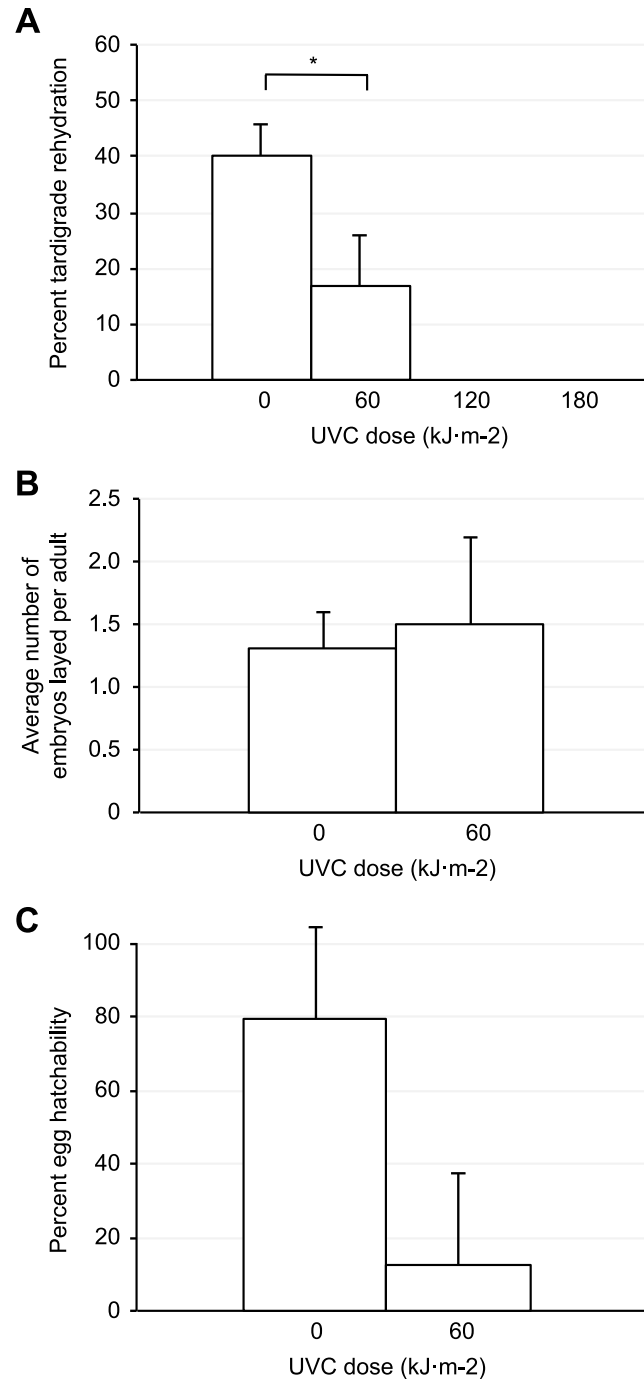


Fig 4. Survival and progeny hatching of *Hypsibius exemplaris* tardigrade irradiated with UVC during anhydrobiosis. (A) Mean percentage of survival after UVC irradiation and rehydration. Error bars represent standard deviation (n = 200). The asterisk indicates respective significant difference with control (Pairwise Z-test, $P < 0.05$, p -value = 0.029). (B) Average number of embryos laid per rehydrated tardigrade. The error bars represent standard deviation (n = 37). (C) Mean percentage of the embryos hatchability. Error bars represent standard deviation (n = 37).

<https://doi.org/10.1371/journal.pone.0208617.g004>

value 19.9) and almost three times the mean value observed at 14 days of anhydrobiosis (mean value 15.9).

We observed a significant difference (Kruskal-Wallis test $P < 0.05$) between the carbonyl labeling intensity in both the 14 and 17 day anhydrobiotic groups in compared with that in the older 73 day anhydrobiotic tardigrade group. Furthermore and to a lesser extent, we observed a significant difference (Kruskal-Wallis test $P < 0.1$) of the carbonyl labeling between the anhydrobiotic group aged 27 days and the older group aged 73 days. Altogether, the data plotted in Fig 1C showed the following linear regression analysis: $y = 0.36x + 9.92$; $R^2 = 0.88$.

In summary, we have uncovered an unexpected constitutive carbonyl accumulation in tardigrade anhydrobiotes.

Discussion

Previous reports of total carbonyl analysis used methods that included sample extraction protocols from whole organisms [31]. However, these protocols have been challenged because of underestimation of carbonylation due to a failure to extract an aggregated protein fraction insolubilized by a UV-dependent mechanism [32]. Moreover, other reports used protocols with chemicals such as 2,4-Dinitrophenylhydrazine (DNPH) or paraformaldehyde (PFA), which are believed to induce a carbonylation overestimation [33,34].

To increase the sensitivity and the specificity of the carbonyl labeling, we adapted an improved *in situ* carbonyl staining technique [30]. This technical procedure can bypass any extraction protocol requirements and reduce spontaneously occurring carbonylation reactions prior to analysis. In a first set of experiments, we performed carbonyl labeling of hydrated tardigrade in response to rising UVC irradiation doses. We were then able to detect up to a five-fold increase in carbonyl staining in the irradiated groups with the maximal UVC dose assessed (180 kJ.m^{-2}). Moreover, all the UVC irradiation doses that we assessed here were higher than the doses used by Horikawa et al. [35] and caused instant death of hydrated tardigrade.

By contrast to other tardigrade species that can withstand direct exposure to relatively low humidity environments, *Hypsibius exemplaris* needs preconditioning pre-exposure to a high humidity environment to fully develop the characteristics of a stress-resistant anhydrobiotic organism [36–39]. Nonetheless, to date, studies [40] have provided new insights into the stress resistance of *Hypsibius exemplaris* when hydrated but not as an anhydrobiote. Unexpectedly, we observed that anhydrobiotes of *Hypsibius exemplaris* could survive a high UVC dose (60 kJ.m^{-2}). Moreover, we noticed that the egg production of the rehydrated animals, previously irradiated as anhydrobiotes with a 60 kJ.m^{-2} UVC dose, was not impaired.

In the next experiment, we assessed if the hatchability of the eggs produced by the anhydrobiotes irradiated with high UVC doses was affected. We observed that the hatchability of the eggs laid by the *Hypsibius exemplaris* groups irradiated with UVC (60 kJ.m^{-2}) appeared drastically reduced (11%). This result is significantly different from the reported hatchability (80%) of *Ramazzottius varieornatus* eggs when irradiated with 20 kJ.m^{-2} of UVC [35]. This difference result perhaps from a higher sensitivity of the *Hypsibius exemplaris* embryo precursors to UVC radiation while inside the anhydrobiotes. Therefore, because the germ cell or embryo status during the *Hypsibius exemplaris* anhydrobiotic stage remains unknown, we cannot yet explain the possible impact of UVC on both fertility and hatchability. Previous studies on tardigrade irradiation were performed on isolated embryos [14,24,40,41–43] but we described here the effect of UVC on the hatching of embryos that were not directly irradiated but instead exposed while inside the anhydrobiotic tardigrade.

In the final experimental section, we were surprised to observe gradual, proportional increases in the carbonyl content of the *Hypsibius exemplaris* anhydrobiotes of different ages. Moreover, we were able to detect up to a 4-fold increase in carbonylation at 73 days of anhydrobiosis compared with that in controls. The degree of carbonylation observed is almost of the same order of magnitude as the maximum carbonylation level that we observed when living tardigrade were irradiated with 180 kJ.m^{-2} of UVC. Moreover, this increase of up to 4-fold of spontaneous carbonylation is well above the 2.5-fold carbonylation increase observed for anhydrobiotes exposed to the 180 kJ.m^{-2} UVC dose. This observation may indicate that tardigrade can accumulate higher carbonylation levels than those observed in this work.

Proteins are the main targets of oxidation by ROS, which can result in protein carbonylation accumulation. In this work, we showed that *Hypsibius exemplaris* can accumulate carbonylations. Based on the report describing the labeling technique also described here [30], it is likely that after UVC exposure, the carbonylation labeling observed is linked to proteins. A protein carbonylation increase has been reported with age in various species such as yeast [44], flies [45], worms [46] and mammals [47].

The carbonylation of proteins is a process recognized as the universal marker of oxidative stress that has been associated with several human disorders and diseases of aging [23]. Because an increase in protein carbonylation marks the *Hypsibius exemplaris* anhydrobiote as it ages, we propose that this species must experience aging. However, whether other tardigrade species also accumulate carbonylation as they age still needs to be assessed. The tardigrade *Hypsibius exemplaris* is not as resistant to harsh conditions as other tardigrade species. However, innovative molecular techniques [48] and recently available genome data [49–53] make *Hypsibius exemplaris* a good integrative model to decipher how tardigrade species can regulate carbonyl clearance and proteostasis.

Materials and methods

Tardigrade culture and desiccation/rehydration protocols

Hypsibius exemplaris tardigrade [54] were fed with unicellular algae *Chlorococcum* sp.; both were purchased from the Sciento company (Manchester, UK). Specimens were maintained in cultures at 15°C on Chalkley's medium as previously described by Gabriel et al. [48].

The desiccation protocol for randomly selected adult tardigrade was adapted from Hengherr et al. [55] and conducted as follows: coverslips were placed at 25°C in a sealed preconditioning box with 85% relative humidity (RH) for 16 hours with a saturated solution of KCl. The coverslips were then placed into a sealed box at 33% relative humidity (RH) for 48 hours with an MgCl_2 saturated solution. Full dehydration of the specimens was monitored by direct observation under a stereomicroscope. The rehydration of desiccated tardigrade on coverslips was initiated by a drop of Chalkley's medium and the motility evidence of rehydrated animals was monitored by direct observation under stereomicroscope. The anhydrobiotes were stored at a relative humidity (RH) of 24% at room temperature until analysis. The brood size of the rehydrated specimens, either UVC irradiated or not, was monitored over a two-week period.

UVC treatment and labeling of carbonyls in anhydrobiotic tardigrade

UVC radiation and carbonyl labeling were performed according to a previous protocol [30] but with an adaptation for tardigrade. UVC treatment of dehydrated tardigrade was performed *in situ* on slides for 10, 20 and 30 minutes with a dose rate of $99.99 \text{ J.m}^{-2}.\text{s}^{-1}$. To keep tardigrade dehydrated and fixed to their slide, we irradiated tardigrade *in situ* without adding any additional liquid, and the carbonyl labeling process, *i.e.*, chemoporation, the labeling of carbonyls and washing, were performed in 100% (iPrOH). To keep the liquid on dehydrated

tardigrade during the protocol and to enable the labeling of carbonyls, we cut a 1 ml pipette tip and placed it around tardigrade on slides. Chemoporation lasted for 3 minutes on ice (150 μ L 100% isopropanol) and labeling of the carbonyls lasted 45 minutes with 4 μ g.ml⁻¹ Cy5 Hz (Cy5 Mono Hydrazide, GE Healthcare Life Sciences, Buckinghamshire, UK) in 100% iPrOH. Washing was performed in 4 cycles with 150 μ l 100% iPrOH. Slides were prepared by pipetting a minimal volume of mounting medium (Eukitt Quick-hardening mounting medium, SIGMA-ALDRICH Co., St. Louis, Missouri, USA) to cover tardigrade and then placement of a cover slip.

UVC treatment and labeling of carbonyls in living tardigrade

Radiation was performed in 1 ml M9 buffer (M9) in a 3 cm glass petri dish for 10, 20 and 30 minutes with a dose rate of 99.99 J.m⁻².s⁻¹. Tardigrade were transferred with a 10 μ l pipette to a new empty tube in a minimal volume of buffer. Chemoporation was performed with 150 μ l 100% iPrOH for 3 minutes on ice followed by tardigrade transfer. Labeling of carbonyls was performed with 150 μ l 4 μ g.ml⁻¹ Cy5 Hz in 100% iPrOH for 45 minutes followed by tardigrade transfer. Washing was performed with 150 μ l in 4 cycles with 40% iPrOH. Then, for each test condition, all tardigrade were pipetted with a P10 onto a slide, one drop of mounting medium was added, and then a coverslip was placed.

Confocal microscopy imaging and fluorescence quantification

Images were obtained with an LSM 780 confocal microscope (Carl Zeiss, France) using a 10 \times dry (N.A. 0.45) and 20 \times dry (N.A. 0.80) or 63 \times oil (N.A. 1.4) oil immersion objective, with a pinhole setting of 1.31 A.U. and 8- or 12-bit images. Cy5 was excited with the red HeNe laser (633 nm), and emitted light was collected between 643 and 696 nm. Fluorescence quantification was also performed with a confocal microscope according to a previously described protocol [30].

Acknowledgments

We thank the referees for their advice in evaluating the manuscript and their help in its improvement.

Author Contributions

Conceptualization: Myriam Richaud, Simon Galas.

Data curation: Mira Kuzmic, Myriam Richaud, Sandrine Frelon, Simon Galas.

Formal analysis: Myriam Richaud, Simon Galas.

Funding acquisition: Simon Galas.

Investigation: Myriam Richaud, Sandrine Frelon, Simon Galas.

Methodology: Mira Kuzmic, Myriam Richaud, Sandrine Frelon, Simon Galas.

Project administration: Simon Galas.

Resources: Myriam Richaud, Simon Galas.

Software: Mira Kuzmic, Myriam Richaud, Sandrine Frelon, Simon Galas.

Supervision: Mira Kuzmic, Sandrine Frelon, Simon Galas.

Validation: Mira Kuzmic, Simon Galas.

Visualization: Myriam Richaud, Pierre Cuq, Simon Galas.

Writing – original draft: Myriam Richaud, Simon Galas.

Writing – review & editing: Mira Kuzmic, Myriam Richaud, Sandrine Frelon, Simon Galas.

References

1. Guidetti RG, Bertolani RB. Tardigrade taxonomy: an updated check list of the taxa and a list of characters for their identification. *Zootaxa*. 2005; 845: 1–46. <https://doi.org/10.11646/zootaxa.845.1.1>
2. Degma P, Guidetti R. Notes to the current checklist of Tardigrada. *Zootaxa*. 2007; 1579: 41–53.
3. Degma P, Bertolani R, Guidetti R. Actual checklist of Tardigrada species. 2016. <http://www.tardigrada.modena.unimo.it/miscellanea/Actual%20checklist%20of%20Tardigrada.pdf>. Cited 3/7/2016.
4. Kinchin I. The biology of tardigrade. *J Mar Biol Assoc UK*. 1995; 75:262–262.
5. Altiero T, Guidetti R, Caselli V, Cesari M, Rebecchi L. Ultraviolet radiation tolerance in hydrated and desiccated eutardigrade. *J Zool Syst Evol Res*. 2011; 49;Suppl 1: 104–110. <https://doi.org/10.1111/j.1439-0469.2010.00607.x>
6. Jönsson KI, Rabbow E, Schill RO, Harms-Ringdahl M, Rettberg P. Tardigrades survive exposure to space in low Earth orbit. *Curr Biol*. 2008; 18: R729–RR731. <https://doi.org/10.1016/j.cub.2008.06.048> PMID: 18786368
7. Ono F, Mori Y, Sougawa M, Takarabe K, Hada Y, Nishihira N, et al. Effect of very high pressure on life of plants and animals. *J Phys: Conf Ser*. 2012; 377: 012053–01206p. <https://doi.org/10.1088/1742-6596/377/1/012053>
8. Ramløv H, Westh P. Cryptobiosis in the Eutardigrade *Adorybiotus* (*Richtersius*) coronifer: tolerance to alcohols, temperature and de novo protein synthesis. *Zoologischer Anzeiger—A Journal of Comparative Zoology*. 2001; 240: 517–523. <https://doi.org/10.1078/0044-5231-00062>
9. Jönsson KI, Harms-Ringdahl M, Torudd J. Radiation tolerance in the eutardigrade *Richtersius* coronifer. *Int J Radiat Biol*. 2005; 81: 649–656. <https://doi.org/10.1080/09553000500368453> PMID: 16368643
10. Baumann H. Die Anabiose der Tardigraden. *Zoologische Jahrbuån Cher*. 1992: 501–556.
11. Marcus E. Tardigrada. In: Bronn HG, editor, *Klassen und Ordnungen des Tierreichs*, Bd. 5 Abtlig. IV, Buch 3. Leipzig: Akademische Verlagsgesellschaft; 1929.
12. Keilin D. The problem of anabiosis or latent life: history and current concept. *Proc R Soc Lond B Biol Sci*. 1959; 150: 149–191. <https://doi.org/10.1098/rspb.1959.0013> PMID: 13633975
13. Westh P, Ramløv H. Trehalose accumulation in the tardigrade *Adorybiotus* coronifer during anhydrobiosis. *J Exp Zool*. 1991; 258: 303–311. <https://doi.org/10.1002/jez.1402580305>
14. Horikawa DD, Sakashita T, Katagiri C, Watanabe M, Kikawada T, Nakahara Y, et al. Radiation tolerance in the tardigrade *Milnesium tardigradum*. *Int J Radiat Biol*. 2006; 82: 843–848. <https://doi.org/10.1080/09553000600972956> PMID: 17178624
15. Horikawa DD, Kunieda T, Abe W, Watanabe M, Nakahara Y, Yukuhiro F, et al. Establishment of a rearing system of the extremotolerant tardigrade *Ramazzottius varieornatus*: a new model animal for astrobiology. *Astrobiology*. 2008; 8: 549–556. <https://doi.org/10.1089/ast.2007.0139> PMID: 18554084
16. Doyère PLN. Mémoires sur les Tardigrades. Sur le facilité que possèdent les Tardigrades, les rotifères, les anguillules des toits et quelques autres animalcules, de revenir à la vie après été complètement deséchées. *Ann Sci Nat Zool Biol Anim*. 1842; 2e: 5–35.
17. Crowe JH. The physiology of cryptobiosis in tardigrades. *Mem. Ist Ital Idrobiol*. 1975; 32;Suppl: 37–59.
18. Womersley CZ. Biochemical and physiological aspects of anhydrobiosis. *Comp. Biochem. Physiol. B*. 1981; 70: 669–678. [https://doi.org/10.1016/0305-0491\(81\)90001-8](https://doi.org/10.1016/0305-0491(81)90001-8)
19. Westh P, Ramløv H. Cryptobiosis in Arctic tardigrades with special attention to the appearance of trehalose. In: Höpner Petersen G, editor, *Grönland Exkursion 2*. Kiel: Institut für Polarökologie (Kiel). 1988; pp. 227–245.
20. Tunnacliffe A, Lapinski J. Resurrecting Van Leeuwenhoek’s rotifers: a reappraisal of the role of disaccharides in anhydrobiosis. *Philos Trans R Soc Lond B Biol Sci*. 2003; 358: 1755–1771. <https://doi.org/10.1098/rstb.2002.1214> PMID: 14561331
21. Horikawa DD, Higashi S. Desiccation tolerance of the tardigrade *Milnesium tardigradum* collected in Sapporo, Japan, and Bogor, Indonesia. *Zoolog Sci*. 2004; 21: 813–816. <https://doi.org/10.2108/zsj.21.813> PMID: 15333992
22. Schill RO, Fritz GB. Desiccation tolerance in embryonic stages of the tardigrade. *J Zool*. 2008; 276: 103–107. <https://doi.org/10.1111/j.1469-7998.2008.00474.x>

23. Fedorova M, Bollineni RC, Hoffmann R. Protein carbonylation as a major hallmark of oxidative damage: update of analytical strategies. *Mass Spectrom Rev.* 2014; 33: 79–97. <https://doi.org/10.1002/mas.21381> PMID: 23832618
24. Beltrán-Pardo E, Jönsson KI, Harms-Ringdahl M, Haghdoost S, Wojcik A. Tolerance to gamma Radiation in the Tardigrade *Hypsibius dujardini* from Embryo to Adult Correlate Inversely with cellular Proliferation. *PLOS ONE.* 2015; 10: e0133658. <https://doi.org/10.1371/journal.pone.0133658> PMID: 26208275
25. Widel M, Krzywon A, Gajda K, Skonieczna M, Rzeszowska-Wolny J. Induction of bystander effects by UVA, UVB, and UVC radiation in human fibroblasts and the implication of reactive oxygen species. *Free Radic Biol Med.* 2014; 68: 278–287. <https://doi.org/10.1016/j.freeradbiomed.2013.12.021> PMID: 24373962
26. Sinha RP, Häder DP. UV-induced DNA damage and repair: a review. *Photochem Photobiol Sci.* 2002; 1: 225–236. <https://doi.org/10.1039/b201230h> PMID: 12661961
27. Neumann S, Reuner A, Brümmer F, Schill RO. DNA damage in storage cells of anhydrobiotic tardigrades. *Comp Biochem Physiol A Mol Integr Physiol.* 2009; 153: 425–429. <https://doi.org/10.1016/j.cbpa.2009.04.611> PMID: 19361569
28. Dalle-Donne I, Giustarini D, Colombo R, Rossi R, Milzani A. Protein carbonylation in human diseases. *Trends Mol Med.* 2003; 9: 169–176. [https://doi.org/10.1016/S1471-4914\(03\)00031-5](https://doi.org/10.1016/S1471-4914(03)00031-5) PMID: 12727143
29. Baraibar MA, Liu L, Ahmed EK, Friguet B. Protein oxidative damage at the crossroads of cellular senescence, aging, and age-related diseases. *Oxid Med Cell Longev.* 2012; 2012: Article ID 919832. <https://doi.org/10.1155/2012/919832> PMID: 23125894
30. Kuzmic M, Javot H, Bonzom JM, Lecomte-Pradines C, Radman M, Garnier-Laplace J, et al. In situ visualization of carbonylation and its co-localization with proteins, lipids, DNA and RNA in *Caenorhabditis elegans*. *Free Radic Biol Med.* 2016; 101: 465–474. <https://doi.org/10.1016/j.freeradbiomed.2016.11.004> PMID: 27840319
31. Bosshard F, Riedel K, Schneider T, Geiser C, Bucheli M, Egli T. Protein oxidation and aggregation in UVA-irradiated *Escherichia coli* cells as signs of accelerated cellular senescence. *Environ Microbiol.* 2010; 12: 2931–2945. <https://doi.org/10.1111/j.1462-2920.2010.02268.x> PMID: 20545749
32. Maisonneuve E, Fraysse L, Lignon S, Capron L, Dukan S. Carbonylated proteins are detectable only in a degradation-resistant aggregate state in *Escherichia coli*. *J Bacteriol.* 2008; 190: 6609–6614. <https://doi.org/10.1128/JB.00588-08> PMID: 18689474
33. Goudeau J, Aguilaniu H. Carbonylated proteins are eliminated during reproduction in *C. elegans*. *Aging Cell.* 2010; 9: 991–1003. <https://doi.org/10.1111/j.1474-9726.2010.00625.x> PMID: 21040398
34. Possanzini M, Ciccio P, Di Palo V, Draisci R. Determination of low boiling aldehydes in air and exhaust gases by using annular denuders combined with HPLC techniques. *Chromatographia.* 1987; 23: 829–834. <https://doi.org/10.1007/BF02311408>
35. Horikawa DD, Cumbers J, Sakakibara I, Rogoff D, Leuko S, Harnoto R, et al. Analysis of DNA repair and protection in the Tardigrade *Ramazzottius varieornatus* and *Hypsibius dujardini* after exposure to UVC radiation. *PLoS One.* 2013; 8: e64793. <https://doi.org/10.1371/journal.pone.0064793> PMID: 23762256
36. Wright JC. Desiccation tolerance and water-retentive mechanisms in tardigrades. *J Exp Biol.* 1989; 142: 267–292.
37. Welnicz W, Grohme MA, Kaczmarek L, Schill RO, Frohme M. Anhydrobiosis in tardigrades—the last decade. *J Insect Physiol.* 2011; 57: 577–583. <https://doi.org/10.1016/j.jinsphys.2011.03.019> PMID: 21440551
38. Guidetti R, Altiero T, Bertolani R, Grazioso P, Rebecchi L. Survival of freezing by hydrated tardigrades inhabiting terrestrial and freshwater habitats. *Zoology.* 2011; 114: 123–128. <https://doi.org/10.1016/j.zool.2010.11.005> PMID: 21429723
39. Boothby TC, Tapia H, Brozena AH, Piszkiwicz S, Smith AE, Giovannini I, et al. Tardigrades use intrinsically disordered proteins to survive desiccation. *Mol Cell.* 2017; 65: 975–984. e5. <https://doi.org/10.1016/j.molcel.2017.02.018> PMID: 28306513
40. Beltrán-Pardo E, Jönsson KI, Wojcik A, Haghdoost S, Harms-Ringdahl M, Bermúdez-Cruz RM, et al. Effects of Ionizing Radiation on Embryos of the Tardigrade *Milnesium cf. tardigradum* at Different Stages of Development. *PLOS ONE.* 2013; 8: e72098. <https://doi.org/10.1371/journal.pone.0072098> PMID: 24039737
41. Horikawa DD, Yamaguchi A, Sakashita T, Tanaka D, Hamada N, Yukuhiro F, et al. Tolerance of anhydrobiotic eggs of the Tardigrade *Ramazzottius varieornatus* to extreme environments. *Astrobiology.* 2012; 12: 283–289. <https://doi.org/10.1089/ast.2011.0669> PMID: 22490117

42. Jönsson I, Beltran-Pardo E, Haghdoust S, Wojcik A, Bermúdez-Cruz RM, Bernal Villegas JE, et al. Tolerance to gamma-irradiation in eggs of the tardigrade *Richtersius coronifer* depends on stage of development. *J Limnol.* 2013; 72: 73–79. <https://doi.org/10.4081/jlimnol.2013.s1.e9>
43. Jönsson KI, Schill RO. Induction of Hsp70 by desiccation, ionising radiation and heat-shock in the eutardigrade *Richtersius coronifer*. *Comp Biochem Physiol B Biochem Mol Biol.* 2007; 146: 456–460. <https://doi.org/10.1016/j.cbpb.2006.10.111> PMID: 17261378
44. Aguilaniu H, Gustafsson L, Rigoulet M, Nyström T. Asymmetric inheritance of oxidatively damaged proteins during cytokinesis. *Science.* 2003; 299: 1751–1753. <https://doi.org/10.1126/science.1080418> PMID: 12610228
45. Orr WC, Sohal RS. Extension of life-span by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster*. *Science.* 1994; 263: 1128–1130. <https://doi.org/10.1126/science.8108730> PMID: 8108730
46. Adachi H, Fujiwara Y, Ishii N. Effects of oxygen on protein carbonyl and aging in *Caenorhabditis elegans* mutants with long (age-1) and short (mev-1) life spans. *J Gerontol A Biol Sci Med Sci.* 1998; 53: B240–BB244. <https://doi.org/10.1093/gerona/53A.4.B240> PMID: 18314552
47. Stadtman ER. Protein oxidation and aging. *Science.* 1992; 257: 1220–1224. <https://doi.org/10.1126/science.1355616> PMID: 1355616
48. Gabriel WN, McNuff R, Patel SK, Gregory TR, Jeck WR, Jones CD, et al. The tardigrade *Hypsibius dujardini*, a new model for studying the evolution of development. *Dev Biol.* 2007; 312: 545–559. <https://doi.org/10.1016/j.ydbio.2007.09.055> PMID: 17996863
49. Boothby TC, Tenlen JR, Smith FW, Wang JR, Patanella KA, Nishimura EO, et al. Evidence for extensive horizontal gene transfer from the draft genome of a tardigrade. *Proc Natl Acad Sci USA.* 2015; 112: 15976–15981. <https://doi.org/10.1073/pnas.1510461112> PMID: 26598659
50. Boothby TC, Tenlen JR, Smith FW, Wang JR, Patanella KA, Nishimura EO, et al. Correction for Boothby et al. Evidence for extensive horizontal gene transfer from the draft genome of a tardigrade. *Proc Natl Acad Sci USA.* 2016; 113: E5364. <https://doi.org/10.1073/pnas.1613046113> PMID: 27551073
51. Koutsovoulos G, Kumar S, Laetsch DR, Stevens L, Daub J, Conlon C, et al. No evidence for extensive horizontal gene transfer in the genome of the tardigrade *Hypsibius dujardini*. *Proc Natl Acad Sci USA.* 2016; 113: 5053–5058. <https://doi.org/10.1073/pnas.1600338113> PMID: 27035985
52. Arakawa K, Yoshida Y, Tomita M. Genome sequencing of a single tardigrade *Hypsibius dujardini* individual. *Sci Data.* 2016; 3: 160063. <https://doi.org/10.1038/sdata.2016.63> PMID: 27529330
53. Yoshida Y, Koutsovoulos G, Laetsch DR, Stevens L, Kumar S, Horikawa DD, et al. Comparative genomics of the tardigrades *Hypsibius dujardini* and *Ramazzottius varieornatus*. *PLOS Biol.* 2017; 15: e2002266. <https://doi.org/10.1371/journal.pbio.2002266> PMID: 28749982
54. Gašiorek P, Stec D, Morek W, Michalczyk Ł. An integrative redescription of *Hypsibius dujardini* (Doyère, 1840), the nominal taxon for *Hypsibioidea* (Tardigrada: Eutardigrada). *Zootaxa.* 2018; 4415 (1): 45–75. <https://doi.org/10.11646/zootaxa.4415.1.2> PMID: 30313631
55. Hengherr S, Heyer AG, Köhler HR, Schill RO. Trehalose and anhydrobiosis in tardigrades—evidence for divergence in responses to dehydration. *FEBS J.* 2008; 275: 281–288. <https://doi.org/10.1111/j.1742-4658.2007.06198.x> PMID: 18070104