ORIGINAL RESEARCH

High throughput screening against pantothenate synthetase identifes amide inhibitors against *Mycobacterium tuberculosis* **and** *Staphylococcus aureus*

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Abstract

Pantothenate is a crucial enzyme for the synthesis of coenzyme A and acyl carrier protein in *Mycobacterium tuberculosis* and *Staphylococcus aureus*. It is indispensable for the growth and survival of these bacteria. Amides analogs are designed and have been used as inhibitors of pantothenate synthetase. Molecular docking approach has been used to design and predict the drug activity of molecule to the specifc disease. In this work, more than hundred amides have been screened by Discovery Studio molecular docking programme to search best suitable molecule for the treatment of *Mycobacterium tuberculosis*. Pharmacophore generation has been done to recognize the binding modes of inhibitors in the receptor active site. To observe the stability and fexibility of inhibitors molecular dynamics (MD) simulation has been done; Lipinski's rule of fve protocols is followed to screen drug likeness and ADMET (absorption, distribution, metabolism, excretion and toxicity) fltration is also used to value toxicity. DFT computation of optimized geometry and derivation of MOs has been used to correlate the drug likeness. The small diference in energy between HOMO and LUMO may help to activate the drug in the protein environment quickly. 2-Hydroxy-5-[(E)-2-{4-[(prop-2-enamido)sulfonyl]phenyl}diazen-1-yl]benzoic acid (M1) shows best theoretical efficiency against *Mycobacterium tuberculosis* (MTB) pantothenate synthetase and so does 2-hydroxy-5-[(E)-2-{4-[(2-phenylacetamido)sulfonyl]phenyl}diazen-1-yl]benzoic acid (M2) against *Staphylococcus aureus* pantothenate synthetase. These compounds also bind to Adenine–Thymine region of tuberculosis DNA.

Graphical abstract

Extended author information available on the last page of the article

Keywords Pantothenate synthetase inhibitors · Heterocyclic amide compounds · Structure based drug design · Molecular docking · ADMET · MD simulation

Introduction

Virtually one-third of the human population of the world is sufering from *Mycobacterium tuberculosis* (MTB) infection (Onyango [2011\)](#page-22-0). Despite the existence of approved drug against TB, it continues to claim approximately 1.5 million lives every year due to drug-resistant TB problem (multidrug resistant tuberculosis, MDR-TB and extensively drug resistant tuberculosis, XDR-TB). So, there is an urgent need to develop new anti-TB drugs (Onyango [2011\)](#page-22-0). In the search for MTB vaccine, growth and virulence of pantothenate synthetase (panC) auxotrophs has been severely collaborated, sustaining the theory of the necessity of this enzyme and its prettiness as an antimicrobial target (White et al. [2007](#page-22-1)).

Pantothenate synthetase (PS) plays critical roles in many cellular processes like, fatty acid metabolism. It catalyzes the adenosine triphosphate (ATP)-dependent condensation of pantoate and β-alanine to form pantothenate (vitamin B5), and key precursor for the synthesis of coenzyme A (CoA) and acyl carrier protein (ACP) (von Delft et al. [2001](#page-22-2)). Upon action of fatty acid synthases on Acetyl-CoA and NADPH fatty acids are generated which combine with glycerol followed by phosphorylation could form phospholipid (Leonardi and Jackowski [2007](#page-22-3)). The bulk of the lipid bilayers those make up cell wall and surround the organelles within the cells have been synthesized from phospholipids (Berg et al. [2002](#page-21-0); Chafey et al. [2003](#page-21-1)). Lipid-rich cell wall of MTB is an essential element of intracellular survival and pathogenicity, and also thought to contribute to the difficulty of efectively delivering antimicrobial agents into the cell. The signifcance of this lipid-rich cell wall is underscored by the large number of genes (approximately 250) encoding enzymes in fatty acid metabolism present in the MTB genome, making this pathway a promising target for new antibacterial drug discovery (Cole et al. [1998](#page-21-2)). Indeed, several anti-tubercular agents are known to inhibit cell wall biosynthesis.

PanC is absent in mammals. It searches pantothenate from their diet using pantothenate permease, of which, there is no homolog in MTB (Table [1\)](#page-2-0) (Grassl [1992;](#page-22-4) Vallari and Rock [1985](#page-22-5)). Literature search shows that MTB mutant defective in the de novo biosynthesis of pantothenate is highly attenuated in both immune compromised and immune competent mice. It points out that functionality of pantothenate in biosynthetic pathway is crucial (Fig. [1\)](#page-3-0) for virulence of MTB (Wang and Eisenberg [2003\)](#page-22-6). Different industries invest over 50 billion dollars on research and development each year to identify potential new drug targets. Pantothenate Synthetase may explore an opportunity to design the drug resistant TB drugs (Overington et al. [2006](#page-22-7)).

Amides are known to play a crucial role in supramolecular anion sensing technology (McMurry et al. [2017\)](#page-22-8). They may also be used as antibacterial agents against Gram positive and Gram negative bacteria in the future design of drugs (Stefańska et al. [2015](#page-22-9)). Amide derivatives (Table [2\)](#page-4-0) show numerous types of biological features as anthelmintic, antihistaminic, antifungal, and antibacterial including *Staphylococcus aureus* and *Mycobacterium tuberculosis* (Yildiz et al. [2008;](#page-22-10) Jagessar and Rampersaud [2007;](#page-22-11) The Sulfa Derivatives in the Treatment of Tuberculosis [1944](#page-22-12)).

On the other hand, sulfa drugs show potent anti-microbial activity and literature has shown that they are active against *Mycobacterium tuberculosis* and *Staphylococcus aureus* (Bartzatt et al. [2010](#page-21-3); Holloway et al. [2016](#page-22-13)). In this work, a series of amide functionalized sulfa drugs have been designed by modifying sulfa drugs amides by structure based drug design. Amides have been docked to examine their action against *Mycobacterium tuberculosis* (MTB) and *Staphylococcus aureus* (SA).

According to some scientists at Lilly Research Laboratories, Eli Lilly & Company, USA two compounds with 4-cyano-1-methyl-3-(4-phenylphenyl)pyrrole-2-carboxylic acid core structure, inhibited MTB pantothenate synthetase (Kumar et al. 2013). The MIC₅₀ values of the two compounds were high (55 and 118 μ M. This suggested the growth inhibitory properties were due to PanC-mediated inhibition[a].The IUPAC name of the compounds are 3‐{[1,1′‐biphenyl]‐4‐ yl}‐4‐cyano‐5‐(ethylsulfanyl)‐1‐methyl‐1H‐pyrrole‐2‐carboxylic acid (compound 1) and $3 - \{[1,1'-biphenyl]-4-yl\}-4$ cyano‐5‐ethyl‐1‐methyl‐1H‐pyrrole‐2‐carboxylic acid (compound 2). In the present work it has been showed that 2-hydroxy-5-[(E)-2-{4-[(prop-2-enamido)sulfonyl]phenyl} diazen-1-yl]benzoic acid and 2-hydroxy-5- $[(E)$ -2- $\{4-[2]$ phenylacetamido)sulfonyl]phenyl}diazen-1-yl]benzoic acid showed better docking score than compound 1 (C1) and compound 2 (C2). Sulfonamide, sulfamoyl groups as cocrystalized structure in the crystal structure of pantothenate synthetase proves their efficiency in binding and inhibition.

Out of 154 diferent amides of sulfa drugs 93 derivatives show higher docking score (binding affinity) than C1 (Table [3](#page-10-0)). For SA also some compounds exhibit docking score higher than C2 (Table [4\)](#page-9-0) which are used to treat bacterial infection by susceptible microorganisms (Kumar et al. [2013;](#page-22-14) Wu et al. [2003](#page-22-15)). Molecular dynamics (MD) simulations have been done to observe the effect of explicit solvent molecules on protein (Pantothenate Synthetase) ligand (amides) complex structure and stability to achieve

time-averaged attributes of the bimolecular system, along with diverse thermodynamic parameters. Drug likeness has been examined following Lipinski's rule of fve flter (Lipin ski et al. [2001\)](#page-22-16). The molecular orbitals are used to calculate the electronic confguration, molecular reactivity and stabil ity of the compounds by density functional theory (DFT) computational process (Rozhenko [2014](#page-22-17)). Pharmacophore map generation has also done to observe steric and elec tronic features of best docked amides that ensured the opti mal interactions with a receptor (Pantothenate Synthetase) and to block its biological response (Wermuth et al. [1998](#page-22-18)). ADMET (absorption, distribution, metabolism, excretion and toxicity) fltration has been applied to check toxicity of the compounds (Hou and Wang [2008\)](#page-22-19). Two compounds out of 154 amides, 2 -hydroxy-5- $[(E)$ - 2 - $\{4$ - $[(prop-2-enamide)]$ sulfonyl]phenyl} diazen-1-yl]benzoic acid (**M1**, Table [2\)](#page-4-0) and 2-hydroxy-5-[(E)-2-{4-[(2-phenylacetamido)sulfonyl] phenyl}diazen-1-yl]benzoic acid (**M2**, Table [2\)](#page-4-0) exhibit bet ter theoretical drug potency than approved and have crossed ADMET and Lipinski's rule of fve flter (Table [2](#page-4-0)). These two compounds also bind to Adenine–Thymine region of tuberculosis DNA (Sirajuddin et al. [2013](#page-22-20)).

Methods and materials

Sequence alignment analysis

Functional similarity between two or more sequences of amino acids has been carried out by using *in silico* multiplesequence alignment (MSA) and is important for fnding the functionality of proteins and evolution history of the species (Nguyen and Pan [2013](#page-22-21)).

High throughput virtual screening (HTVS)

Structure-based drug discovery is a vital process for fast and cost-effective lead discovery and proven to be more efficient than the conventional wet lab drug discovery (Lionta et al. [2014;](#page-22-22) Pradhan and Sinha [2017\)](#page-22-23). High-Throughput Screening (HTS) is an approach to drug discovery that has become a standard tool for structure-based drug discovery. It is basi cally a process of screening of large number of drug like molecules against selected and specifc drug targets. HTS are used for screening of various types of libraries, includ ing combinatorial chemistry, genomics, protein, and peptide libraries (Szymański et al. [2012](#page-22-24)).

Drug‑like amide molecule preparation

We have collected 154 small Lipinski's filter passed amide molecule compounds from the PubChem database which inhibit pantothenate synthetase enzyme (Database Resources

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of the National Center for Biotechnology Information [2013](#page-21-4)). The compounds are drawn by compiling sulfa drugs with amides attached to the amino group of sulfa drugs by Accelrys Draw v4. The 3D coordinates and change of ionization are done by Discovery Studio 4 software's ligand preparation wizard. Compounds are prepared using the "Prepare Ligand" protocol in Discovery Studio v4; default parameters towards performing the CDOCKER simulation with sdf fles (The Sulfa Derivatives in the Treatment of Tuberculosis [1944\)](#page-22-12).

a. Docking preparation of pantothenate synthetase

Pantothenate synthetase of MTB has been retrieved from RCSB PDB database with fve bound-inhibitors. The protein data bank (pdb) fle (pdb id: 4efk) showed a dimer of two chains. However, in the present study, only the monomeric unit (A-chain) has been used in the docking studies because it has two N, N-dimethylthiophene-3-sulfonamide bound inhibitors. The pantothenate synthetase of SA was retrieved from RCSB PDB with three bound-inhibitors (pdb id: 3ag6). The pdb fle was a dimer of two chains, only the monomeric unit (A- chain) was used in the docking studies.

The bounded inhibitors from the pdb structure are removed before docking. Prepare Protein protocol has been executed such as inserting missing atoms in incomplete residues, modeling missing loop regions, and removing waters from protein. The default parameter values are mostly kept the invariant in the Prepare Protein protocol of Discovery Studio 4.

b. Molecular docking

Molecular docking of 154 selected pantothenate synthetase inhibitors to the receptor enzyme has been carried out in the present study by using CDOCKER with Discovery Studio v4. To perform fexible docking, for the small-molecules all torsion angles are set to be free. CDOCKER is a powerful CHARMm-based docking method that has been used to generate highly accurate docked poses. In this refnement application, the ligands were conceded to tilt around the rigid receptor (Wu et al. [2003](#page-22-15)). In the CDOCKER simulation the following parameters are used: top hits-10, random conformations-10, orientations to refne-10, and force feld-CHARMm. For the assurance of potential relationships between pantothenate synthetase and amides, predicted CDOCKER energy values of the best docked conformations of small-molecule inhibitors (amides and approved drugs) are selected as preliminary binding conformations and saved for observing interactions between pantothenate synthetase and amides.

Drug likeness

Lipinski's rule of fve (RO5) describes four simple physicochemical factors ranges (MWT <500, log $P < 5$, H-bond donors < 5 , H bond acceptors < 10) related with 90% of orally active drugs that have accomplished phase II clinical status (Lipinski et al. [2001\)](#page-22-16). These physicochemical factors are related with aqueous solubility, intestinal permeability

Fig. 1 Activity of pantothenate synthetase

Table 2 Selected amides (IUPAC names are given) showing docking score (CDOCKER energy) against Mycobacterial and *Staphylococcus* p.s. They have been passed the Lipinski's rule and ADMET flter

	IUPAC name of amides	MTB	SA	Lipinski's rule ADMET	
A	Sulfamethoxazole	a.u.	a.u.		
1	(2S)-2-amino-4-({4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}carbamoyl)butanoic acid	-44.63	-46.94	$^{+}$	$^{+}$
$\sqrt{2}$	(2S)-2-amino-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-3-phenylpropana- mide	-46.53	-55.40	$^{+}$	$\boldsymbol{+}$
3	2-Amino-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}benzamide	-45.32	-47.30		$\boldsymbol{+}$
$\overline{4}$	2-Hydroxy-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}benzamide	-56.43	-53.08		$^{+}$
5	4-({4-[(5-Methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}amino)benzene-1-sulfonamide	-48.41	-48.83	$^{+}$	$^{+}$
6	4-({4-[(5-Methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}amino)benzene-1-sulfonamide	-50.02	-50.64	$\ddot{}$	$\pmb{+}$
7	4-Methyl-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}benzene-1-sulfonamide	-40.11	-47.32	$\overline{+}$	$\overline{+}$
8	5-({4-[(5-Methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}amino)naphthalene-1-sulfonamide	-35.012	-52.06	$^{+}$	$^{+}$
9	6-({4-[(5-Methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}amino)pyridine-3-carboxamide	-50.57	-48.74	$\overline{+}$	$\boldsymbol{+}$
10	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl} acetamide	-37.12	-36.75	$\overline{+}$	$\mathrm{+}$
11	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}benzamide	-45.62	-46.02	$\mathrm{+}$	$\boldsymbol{+}$
12	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}hexanamide	-41.38	-49.40	$\overline{+}$	$\boldsymbol{+}$
13	N-(5-methyl-1,2-oxazol-3-yl)-4-(N-methylhydrazido)benzene-1-sulfonamide	${\rm ND}$	-55.37		$\mathrm{+}$
14	4-(N-butylbenzenesulfonamido)-N-(5-methyl-1,2-oxazol-3-yl)benzene-1-sulfonamide	-38.70	-38.31		\ddag
15	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}prop-2-enamide	-38.21	-35.20	$^{+}$	$^+$
16	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}pyrazine-2-carboxamide	-45.32	-46.67	$\overline{+}$	$^+$
17	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}pyridine-3-carboxamide	$\rm ND$	-47.12	$\overline{+}$	\ddag
18	(2R)-4-carbamoyl-2-({4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}amino)butanoic acid	-53.47	-57.71	$^{+}$	\ddag
19	(2R,3S)-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-3-[(4E,7E)-nona-4,7- dienoyl]oxirane-2-carboxamide	-44.86	-62.80	$\overline{+}$	$\, +$
20	(2S)-2,6-diamino-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}hexanamide	-50.59	-52.40	$^{+}$	$\mathrm{+}$
21	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-1,3-benzothiazole-2-sulfonamide	-40.30	-49.45	$^{+}$	$^+$
22	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-1,3-thiazole-2-sulfonamide	-41.62	-45.85	$^{+}$	$\,{}^+$
23	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-2,3-dihydro-1H-indene-5-sulfon- amide	41.73	-47.61	$^{+}$	$\, +$
24	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-2-phenoxyacetamide	-42.48	-49.78	$^{+}$	$\, +$
25	3-Amino-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}benzamide	-48.74	-49.06	$^{+}$	$\, +$
26	N-(4-ethoxyphenyl)-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl} acetamide	-44.95	-50.17	$^{+}$	$\, +$
27	N-methyl-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl} acetamide	-38.90	-39.89	$^{+}$	$\, +$
28	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}thiophene-2-sulfonamide	-38.80	-42.56	$\ddot{}$	$\,^+$
29	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-1,2,3,4-tetrahydroisoquinoline- 7-sulfonamide		$-45.4053 - 49.9116 +$		$^{+}$
30	1-Methyl-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-3-oxo-1,3-dihydro- 2,1-benzothiazole-5-sulfonamide	-42.03	-56.77	$^{+}$	$\overline{+}$
31	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-2-propylpentanamide	-42.87	-48.82	$^{+}$	\pm
32	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-3-oxo-N-[(3S)-2-oxooxolan-3-yl] octanamide	ND	-63.91	$^{+}$	$\mathrm{+}$
33	N-[2-({4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}amino)ethyl]isoquinoline- 5-sulfonamide	ND	-37.33	$\mathrm{+}$	$\mathrm{+}$
34	N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-2-oxopropanamide	-42.18	-40.96	$^{+}$	$\,{}^+$
35	4-(4-Chlorophenyl)-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}piperazine- 1-carboximidamide	${\rm ND}$	ND	$^{+}$	$\hspace{0.1mm} +$
36	4-Amino-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}-1H-imidazole-5-carbox- amide	-43.80	-41.97	$^{+}$	$\overline{+}$
37	5-Hydroxy-N-{4-[(5-methyl-1,2-oxazol-3-yl)sulfamoyl]phenyl}naphthalene-1-sulfon- amide	-39.17	-51.45	$\ddot{}$	$\mathrm{+}$
38	4-Benzenesulfonamido-N-(5-methyl-1,2-oxazol-3-yl)benzene-1-sulfonamide	-43.03	-46.33	$\begin{array}{c} + \end{array}$	$^{+}$

In the table 'ND' denotes there were no docking pose for the molecules, 'NA' denotes the molecules hadn't passed the Lipinski rule. '+' sign denotes that the amides have passed Lipinski's rule and ADMET filter, '–' denotes they haven't passe Lipinski's rule and ADMET filter

and oral bioavailability. Because all parameters can be easily computed, the RO5 (or its variants) has become the most widely useful flter in virtual library design (Lipinski [2004\)](#page-22-25).

Quantum chemistry calculation

DFT (density functional theory) calculations have been performed by Gaussian 09W (Frisch et al. [2009](#page-21-5)). Gaussian calculation setup has been done in Gaussian 09 software using Becke's three-parameter exchange potential and Lee–Yang–Parr correlation functional (B3LYP) theory with basis set 6–31G (Becke [1993](#page-21-6); Gill et al. [1992](#page-21-7); Devlin et al. [1995](#page-21-8)). The surfaces (molecular orbital, density, potential) and electrostatic potential charges (EPS) have calculated the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) (Fukui et al. [1952](#page-21-9)). Chemical reactivity, intermolecular interactions and kinetic stability of molecules are characterized by the energy difference of HOMO–LUMO functions (Rauk [1994;](#page-22-26) Fleming [2011](#page-21-10); Strom and Wilson [2013;](#page-22-27) Brownell et al. [2013\)](#page-21-11). Interaction between the HOMO of the drug (amides) and the LUMO of the receptor (pantothenate synthetase) is the key factor of drug activity. Tight binding of drugs with the receptor can be achieved by increasing HOMO energy and decreasing LUMO energy in the drug molecule (El-Henawy et al. [2013](#page-21-12)).

Pharmacophore generation

A pharmacophore model is an ensemble of steric and electronic features that is necessary to ensure the optimal supramolecular interactions with a specifc biological target and to trigger (or block) its biological response (Wermuth et al. [1998](#page-22-18)). The generated pharmacophore models based on receptor-ligand interactions have confrmed all substantial interactions in the compound-receptor interaction modes (Meduru et al. [2016\)](#page-22-28). In a structure-based pharmacophore model, possible interaction area between the drug target (receptor) and ligands are examined (Böhm [1992](#page-21-13)).

The structure-based pharmacophore (SBP) method employed in Discovery Studio is a typical example of a macromolecule-based approach. SBP converts LUDI interaction maps within the protein-binding site into catalyst pharmacophoric features: H-bond acceptor, H-bond donor and hydrophobe. The computer program LUDI is a new method for the de novo design of enzyme inhibitors (Böhm [1992](#page-21-13); Yang [2010\)](#page-22-29). Its interaction maps comprise of a large number of catalyst features (Yang [2010](#page-22-29)).

In the present work, pharmacophore generation executed by receptor-ligand pharmacophore generation with Discovery Studio 4 for study the interactions between protein and ligand. The receptor-ligand pharmacophore generation produces few pharmacophore models from a receptor-ligand complex. The model is generated from the features that are related to the receptor-ligand docking interactions. The following ligand features types are considered: hydrogen bond acceptor, hydrophobic feature, ionizable feature and aromatic ring.

Drug–DNA interaction

Drugs (amides) can interact with different coordinates (groove) of DNA, creating different binding patterns (Chaires [1998](#page-21-14)). Each pattern has its own signifcance (Chen et al. [1993](#page-21-15)). Study and recognition of these patterns lead to fruitful estimate of binding modes and site selectivity which will be contributory for developments in the understanding of new drug molecules as potent and selective gene-regulatory drugs (Chaires [1998](#page-21-14); Chen et al. [1993](#page-21-15)).

To fnd the binding pattern of Amides with MTBDNA (pdb id: 3pvp), docking method is used. Drug-DNA docking is done by autodock vina software on Windows platform with 8 Gb RAM and Intel I 5 processor (Trott and Olson [2010](#page-22-30)).

Table 3 Selected sulfonamides of higher docking score than C1 with MTB pantothenate synthetase and passed the Lipinski's rule and ADMET flter. IUPAC names (Serial No. Table [2\)](#page-4-0) and CDOCKER energy (CDE). The name of the drug group are given below. Added functional groups are in coloured circle

ADMET

Before a drug applies pharmacodynamics effect on the body via interaction with its target, it must transport through the body to reach the site of drug action. Pharmacokinetics denotes to the expedition of the drug from its point of entrance to the site of action. Generally speaking, this process can be defned by the following phases: absorption, distribution, metabolism, excretion and toxicity called ADMET (Roncaglioni et al. [2013\)](#page-22-31). Competence of the drug to reach pharmacologically active concentration at the drug targets without any undesirable effect is taken care by ADMET properties which include the calculation of a series of factors (Lin et al. [2013\)](#page-22-32).

In the present study, ADMET has been carried out by evaluating water solubility, human intestinal absorption, oral bioavailability, blood–brain barrier penetration, transporter, plasma protein binding, volume of distribution, CYP450, **Table 4** Selected sulfonamides of higher docking score than C2 with SA pantothenate synthetase and passed the Lipinski's rule and ADMET flter. IUPAC names (Serial no; Table [2\)](#page-4-0) and CDOCKER energy (CDE).The name of the drug group are given below. Added functional groups are in coloured circle

toxicity etc. by support vector machine (SVM) algorithm (Cheng et al. [2012](#page-21-16)).

Molecular dynamics simulation

Molecular dynamics simulation (MD simulation) is calculated to confrm further the interaction strength and stability of the receptor-ligand complex determined from molecular docking by Discovery Studio's standard dynamics cascade wizard.

The same pdb file which is modified for docking and 2-hydroxy-5-[(E)-2-{4-[(prop-2-enamido) sulfonyl] phenyl} diazen-1-yl] benzoicacid are taken as protein–ligand complexes. For DNA MD simulation, same DNA is chosen. Prior to performing MD simulations, charm force field has been applied to each of the protein–ligand complexes and solvation is set to explicit periodic boundary (Brooks et al. [1983](#page-21-17)). The parameters for MD simulations are set with following conditions: both steepest descent of energy minimization and steps of conjugate gradient minimization are done in order to obtain constant and reasonable conformation of biomolecules (Petrova and Solov'ev [1997](#page-22-33); Hestenes and Stiefel [1952](#page-22-34)). The system was heated from an initial temperature of 50 K to the target temperature of 300 K, and the equilibration steps are done. Moreover, the parameters of electrostatics are chosen as particle mesh Ewald (PME) for long-range electrostatic constrains (Darden et al. [1993\)](#page-21-18). The total production time of 53 ps simulations and are performed with NVT (dynamics without temperature/pressure control) (Beard and Qian [2010\)](#page-21-19). Default setting values are adopted for other parameters.

Results and discussions

Sequence alignment analysis

The sequence alignment analysis has been carried out by Clustal Omega and the results are tabulated in Table [1.](#page-2-0) These results used to observe similarities between diferent pantothenate synthetase (Sievers and Higgins [2014](#page-22-35)). There is no signifcant match between MTB pantothenate synthetase and other pantothenate synthetase. Clustal Omega reveals that *Neisseria gonorrhoeae* pantothenate synthetase has 45.71% similarity with MTB pantothenate synthetase.

Clustal Omega with the support of percent identity matrix (PIM) shows that SA pantothenate synthetase has 45.71% similarity with Thermotoga maritima pantothenate synthetase (Chenna et al. [2003](#page-21-20)). These sequences give an idea of biological species evolution. The sequence similarity would be a motive for drug designers to work with other species containing pantothenate synthetase.

Fig. 2 Docked comprehensive perception of MTB pantothenate synthetase and **M1** after docking. **a** p.s is represented by ribbon and M1 is represented by stick and coloured according to elements. **b** secondary structure of p.s represented by hydrophobic surface and M1 represented is by stick model, **c** interactions of M1 with p.s amino acids. Bonds are in dots. M1 surrounding amino acids are in three letters code, represented in blue

Docking and bond analysis

The specifc binding of a ligand (drug) to a drug target molecule is the key to drug action. Each ligand binds favorably to a specifc site on the surface of the target molecule. Identifcation of the ligand-binding site for each specifc protein molecule is crucially important when trying to fnd a suitable drug molecule for the target, and it is also important to understand the function of the protein (Soga et al. [2007\)](#page-22-36). The docking analysis scores of 93 compounds with MTB and SA pantothenate synthetase have been recorded in Table [2.](#page-4-0) On comparing with CDOCKER energy of C1 and C2, useful tuberculosis drugs A (Table [1](#page-2-0)) and B (Table [2](#page-4-0)) show best score for MTB. The CDOCKER energy of M1 -66.41 a.u. (MTB pantothenate synthetase) and energy of C1 is − 40.58 a.u. M2 shows best score for SA, and CDOCKER energy is − 70.69 a.u. and − 44.31 a.u. for C2.

2‑Hydroxy‑5‑[(E)‑2‑{4‑[(prop‑2‑enamido)sulfonyl]phenyl} diazen‑1‑yl]benzoic acid (M1) and pantothenate synthetase

M1 is docked in the active site of pantothenate synthetase of MTB. It forms 9 hydrogen bonds with amino acids of the pantothenate synthetase in the ranging distance 2.78–3.34 Å (Table [5,](#page-14-0) Fig. [2](#page-11-0)). It also forms 2 electrostatic bonds with amino acids of the ps. The hydrogen bond forming amino acids are asparagine (ARG278), arginine (ARG198), serine (SER197 and SER19), tyrosine (TYR82). The CDOCKER energy is -66.41 a.u. and CDOCKER energy is -39.54 .

C1 and pantothenate synthetase

C1 (Table [1\)](#page-2-0) is docked in the active site of pantothenate synthetase of MTB. It forms three hydrogen bonds with amino acids of the pantothenate synthetase within the ranging distance of 2.23–3.09 Å (Table [5](#page-14-0), Fig. [3](#page-12-0)).The hydrogen bond forming amino acids are Glycine (GLY158), Tyrosine TYR82, Glycine GLN164, Aspartate ASP161. The CDOCKER energy is − 40.58 a.u. and CDOCKER energy is − 5.52 a.u (Fig. [3](#page-12-0)).

5‑[(E)‑2‑{4‑[(4‑aminobenzenesulfonyl) sulfamoyl] phenyl} diazen‑1‑yl]‑2‑hydroxybenzoic acid (M2) and *Staphylococcus aureus* **pantothenate synthetase**

M2 is docked in the active site of *Staphylococcus aureus* pantothenate synthetase. It forms twelve hydrogen bonds ranging between 1.[6](#page-15-0)7 and 3.08 \AA (Table 6, Fig. [4\)](#page-13-0). It also forms an electrostatic interaction at a distance of 3.60 Å. The hydrogen bond forming amino acids are serine (SER186), glutamine (GLN154), lysine (LYS150), arginine (ARG188) histidine (HIS38), methionine (MET31) and threonine (THR30). The CDOCKER energy is − 85.41 a. u. followed by CDOCKER energy is 57.64 a.u. It also forms 1 electrostatic bonds with amino acids of the ps

C2 and pantothenate synthetase

C2 is docked in the active site of SA pantothenate synthetase. It forms three hydrogen bonds with amino acids of the pantothenate synthetase within the ranging distance of 1.9–3.9 Å (Table [6,](#page-15-0) Fig. [5\)](#page-14-1).The hydrogen bond forming amino acids are Arginine (ARG122,ARG188, ARG273), Histidine (HIS38), Lysine(LYS150), Threonine (THR30), Methionine(MET31). It also forms 1 electrostatic bonds with amino acids of the ps. The CDOCKER energy is − 44.31 a.u and CDOCKER energy is − 9.07 a.u.

Table 5 Noncovalent bond distances between 2-hydroxy-5-[(E)-2-{4-[(prop-2-enamido) sulfonyl] phenyl} diazen-1-yl] benzoic acid, C1 and pantothenate synthetase

plots of *M1*. The positive

(c) HOMO, E(HOMO = -0.213889 eV

(d) LUMO, $E(LUMO) = -0.053971$

ADMET property analysis

There are total 26 parameters in ADMET data which were available in literature (Cheng et al. [2012\)](#page-21-16). In the present work a '+' sign is marked when 23 parameters are positive (81%) and '++' is assigned (Table [7\)](#page-18-0) when more than 23 parameters are passed (Cheng et al. [2012\)](#page-21-16). Pharmacokinetic properties and toxicities are predicted by ADMET which can predict permeability for BBB (blood–brain barrier), HIA (human intestinal absorption), P-glycoprotein substrate/ inhibitor, renal organic cation transporter, etc. ADMET result shows 2-hydroxy-5-[(E)-2-{4-[(prop-2-enamido)sulfonyl]phenyl}diazen-1-yl]benzoic acid and 2-hydroxy-5-[(E)- 2-{4-[(2-phenylacetamido)sulfonyl]phenyl}diazen-1-yl] benzoic acid are positive (+) in HIA, BBB permeability. It suggests that the molecules are well absorbed in human

Table 6 Recognizable bonds between *M2*, C2 and *Staphylococcus aureus* Pantothenate synthetase

Noncovalent bond distances Distance (A) bonds between $M2$ (LIG3) and SA pantothenate synthetase		Bonding category	Noncovalent bond distances Distance (A) between C2 (Lig4) and SA ps		Category
A: LYS150:HZ3 - :LIG3: O 1.67		Hydrogen bond; Electro- static	A:ARG188:HH22- : $LIG4:O$	1.9	Hydrogen bond; Elec- trostatic
A: SER186: HG - :LIG3: O	1.91	Hydrogen bond	A:HIS38:HE2 - :LIG4:O	2.11	Hydrogen bond
A: GLN154:HE22 - :LIG3: 1.93 Ω		Hydrogen bond	A:LYS150:HZ3 - :LIG4:N	2.43	Hydrogen bond
A: LYS150:HZ2 - :LIG3: O 2.15		Hydrogen bond; Electro- static	A:LYS150:HZ2 - :LIG4:N	2.63	Hydrogen bond
A:ARG188:HH22 - : $LIG3:N$	2.26	Hydrogen bond	$A:THR30:HA -:LIG4:O$	2.72	Hydrogen bond
A: HIS38:HE2 - :LIG3: O	2.38	Hydrogen bond	A:ARG273:HH11 - :LIG4:O	2.75	Hydrogen bond; Elec- trostatic
A: MET31: HN - :LIG3: O	2.45	Hydrogen bond	A:MET31:HN - :LIG4:O	2.77	Hydrogen bond
A: THR30: HA - :LIG3: O	2.51	Hydrogen bond	A:THR30:HB - :LIG4:O	2.78	Hydrogen bond
A: SER186:HB2 - :LIG3: O	2.68	Hydrogen bond	A:ARG122:NH2 - :LIG4:O	2.83	Electrostatic
A: LYS150:HE1 - :LIG3: O	2.69	Hydrogen bond	A:HIS38:HD2 - :LIG4:O	2.94	Hydrogen bond
A: SER186:HB2 - :LIG3: O	2.81	Hydrogen bond	A:ARG122:NH2 - :LIG4:O	2.95	Electrostatic
A: SER186: HA - :LIG3: O	3.08	Hydrogen bond	A:ARG122:NH2 - :LIG4:O	3.00	Electrostatic
A:ARG188:NH2 - :LIG3	3.60	Electrostatic	A:ARG188:NH2 - :LIG4	3.19	Electrostatic
			A:ARG188:HH11 - : $LIG4:O$	3.20	Hydrogen bond; Elec- trostatic
			$A:ARG188:NH2 -:LIG4:O$	3.90	Electrostatic

Fig. 7 The result of pharmacophore features of *M1* based on receptor-ligand pharmacophore generation. The hydrogen bond acceptor, hydrogen bond donor, positive ionizablefeature, aromatic ring and negative ionizable features are shown as green, orange and blue respectively

Fig. 8 The result of pharmacophore features of *M2* based on receptor-ligand pharmacophore generation. The hydrogen bond acceptor, hydrogen bond donor, positive ionizablefeature, aromatic ring and negative ionizable features are shown as green, orange, and blue respectively

Fig. 9 M1 interaction with Tuberculosis's DNA in minor groove. **a** Drug is represented by stick and double helical DNA structure is represented by ladder and rings, **b** double helical structure of DNA represented by *M1* represented by stick model and are coloured according to elements, **c** interactions of ligand with DNA base pairs (A, T, G, C); the interaction types. Hydrogen bonds are in green. Ligand surrounding base pairs are in three letters code represented in black

body (Table [7](#page-18-0)). Inhibition and initiation of P-glycoprotein have been reported as the causes of drug–drug interactions (Lin and Yamazaki [2003](#page-22-37)). Two best docked molecules (M1 and M2) are P-glycoprotein non-inhibitor. Data in Table [8](#page-19-0) show the best ftted ligand in permissible limit (Lin et al. [2013](#page-22-32)). Organiccation transporters are responsible for drug absorption and disposition in the kidney, liver, and intestine (Zhang et al. [1998](#page-23-0)). ADMET result of two best docked molecules shows that they are non-inhibitor of renal organic cation transporter. The human cytochromes P450 (CYPs), particularly isoforms 1A2, 2C9, 2D6 and 3A4 are responsible for about 90% oxidative metabolic reactions. Inhibition of CYP enzymes will lead to inductive or inhibitory failure of drug metabolism (Uttamsingh et al. [2005\)](#page-22-38).

The Ames test is a widely employed method that uses bacteria to test whether a given chemical can cause cancer. More formally, it is a biological assay to assess the mutagenic potential of chemical compounds (Ames et al. [1972](#page-21-21); Mortelmans and Zeiger [2000\)](#page-22-39).

Human Ether-à-go–go-Related Gene (hERG) is a gene sensitive to drug binding (Sanguinetti and Tristani-Firouzi [2006\)](#page-22-40). ADMET results show that the drugs M1, M2 are weak inhibitor and non-inhibitor of hERG (predictor I and II) (Sanguinetti and Tristani-Firouzi [2006\)](#page-22-40). The aqueous solubility (logS) of a compound considerably affects its absorption and distribution properties. The predicted logS values of the studied compounds are within the acceptable limit (Vyas et al. [2013](#page-22-41)). The solubility (logS) of organic molecules in water is considered in the ADMET, because this parameter generally has important impact on many ADMET concerned properties of drugs, such as uptake, distribution, transport and eventually bioavailability (Hou

Table 7 ADMET properties of M1 and C1

et al. [2004](#page-22-42)). Understanding of ADMET properties together with their measurement and prediction gives an idea about the dose size and dose frequency, drugs solubility, metabolism of drug and its toxicity. Before synthesizing drug like molecules in laboratory, a profound knowledge of drug properties will save time and resources.

Density functional theory analysis

HSFs from the best binding pose have been transferred to Gauss view 5 and the diference in HOMO and LUMO energy, known as band gap, indicates the electronic excitation energy, necessary to compute the molecular reactivity and stability of the compounds (Becke [1993](#page-21-6); Gill et al. [1992](#page-21-7); Devlin et al. [1995](#page-21-8); Fukui et al. [1952](#page-21-9)). For 2-hydroxy-5-[(E)- 2-{4-[(prop-2-enamido)sulfonyl]phenyl} diazen-1-yl] benzoic acid, eigen values of HOMO and LUMO are − 0.182773 and − 0.138828 eV respectively and the HOMO–LUMO gap is − 0.043945. For compound 1, eigen values of HOMO and LUMO are − 0.213889 and − 0.053971 eV and the HOMO–LUMO gap is − 0.159918 eV (Figs. [5](#page-14-1), [6](#page-15-0); Table 6).

Pharmacophore generation

The generated pharmacophore models based on receptorligand interactions by docking have confrmed all major interactions in the drug-receptor interaction modes. The number of features, feature set and selectivity score from

Table 8 ADMET properties of M2 and C2

Table 9 Noncovalent bond distances between M1 and DNA (A and B chain)

Fig. 10 Shows bond energy graph of *M1*, other four molecules of Table [3](#page-10-0) and C1 with pantothenate synthetase of *Mycobacterium tuberculosis*. Bond energy and number of trajectory frames are plotted along X- and Y-axis respectively. Best docked molecule is in blue and C1 is in pink. (Series numbers are maintained as in Table [3](#page-10-0))

Fig. 11 Shows bond energy graph of *M2* with pantothenate synthetase of *S. aurues*. X axis and Y-axis represent the bond energy and number of trajectory frames respectively. Best docked molecule is in pink and C2 is in blue. (Series numbers are maintained as in Table [4](#page-9-0))

Fig. 12 Shows bond energy graph of 2-hydroxy-5-[(E)-2-{4-[(prop-2-enamido) sulfonyl] phenyl} diazen-1-yl] benzoic acid and C1 with DNA of *Mycobacterium tuberculosis*. X-axis and Y-axis represent bond energy and number of trajectory frames respectively. Best docked molecule is in green and C1 is in violet

pharmacophore generation are observed from the two best docked molecules. Pharmacophore generation of 2-Hydroxy-5- $[(E)$ -2- $[4-[(prop-2-enamide) sul$ fonyl]phenyl} diazen-1-yl]benzoic acid (M1) and

Fig. 13 Shows results of molecular dynamics, last conformation (structure) superimposed (red) with frst conformation (blue) of pantothenate synthetase of *Mycobacterium tuberculosis*

2-hydroxy-5-[(E)-2-{4-[(2-phenylacetamido)sulfonyl] phenyl}diazen-1-yl]benzoic acid (M2) are shown in Figs. [7](#page-16-0) and [8](#page-16-1), respectively.

Drug–DNA interaction

The drug (amides)–DNA interactions have been computationally examined by docking techniques used to study interactions between DNA and small ligand molecules those are potentially pharmaceutical interest. Autodock vina software is used for docking (Trott and Olson [2010](#page-22-30)). Drug-DNA interaction gives an idea about binding pattern of drug with DNA. Amides compounds (drug) docked with *M*. *tuberculosis's* DNA is used to understand the drug-DNA interaction. We have observed that amide compounds interact with AT-rich regions (represented by red and turquoise coloured rings respectively) of DNA in the minor groove by forming hydrogen bonding and hydrophobic interactions (Fig. [9](#page-17-0)). The amino group of guanine (represented by green ring) prevents 2-hydroxy-5-[(E)-2-{4-[(prop-2-enamido) sulfonyl] phenyl} diazen-1-yl] benzoic acid from binding to the G·C base pairs by steric hindrance (Table [9\)](#page-19-1), and thus conferring AT-selectivity on the drug molecule. Minor groove binding molecules are usually constructed of a series of heterocyclic or aromatic hydrocarbon rings that possess rotational freedom. This allows these molecules to ft into the minor groove, with displacement of water; these drugs can form hydrogen bonds to bases (Sangeetha Gowda et al. [2014](#page-22-43)).

Molecular dynamics simulation

The MD simulation study of best docked molecules with Mtb and *S. aureus* pantothenate synthetase and Mtb DNA are carried out for 53 ps by 50 thousand steps. MD production run and the trajectory of the various energy profles are created and analyzed.

Potential energy of *M1* after MD simulation was − 14,633 kcal/mol and C1 − 14,607.2 kcal/mol and bond energy was 1526.3 and 1503.13 kcal/mol respectively. Same for S. aureus pantothenate $M2 - 16,245.9$ kcal/mol bond energy was 1642.68 and 1533.34 kcal/mol respectively.

For Mtb DNA and 2-hydroxy-5- $[(E)$ -2- $[4-[(prop-2-])$ enamido) sulfonyl]phenyl} diazen-1-yl] benzoic acid potential energy was − 2394.2 kcal/mol and bond energy was 398.096 kcal/mol. For C1 potential energy was − 2357.67 kcal/mol bond energy 334.856 kcal/mol. Bond energy graphs are shown in Figs. [10](#page-20-0), [11](#page-20-1) and [12.](#page-20-2)

As the graphs revealed that in all situations for the best docked molecules, the bond strength are increased from initial position, ended from the starting point of bond strength and it was higher till then end compared to corresponding drugs. So it can be clearly predicted that molecules formed stable conformation with Mtb and *S*. *aureus* pantothenate synthetase and DNA. Super imposed structures of frst conformation (trajectory frame) and last conformation of Mtb pantothenate synthetase show the deviation of end point of the dynamics from the initial point of dynamics (Fig. [13](#page-20-3)).

Conclusion

Amide functionalized sulfa drugs show potent anti-microbial activity and also active against MTB and *S*. *aureus* in vivo. The best docked compounds have better docking score than approved drugs and also show better ADMET efficiency.

Out of 154 compounds, 2-hydroxy-5- $[(E)-2-(4-[(prop-$ 2-enamido)sulfonyl]phenyl}diazen-1-yl] benzoic acid and 2-hydroxy-5-[(E)-2-{4-[(2-phenylacetamido)sulfonyl]phenyl}diazen-1-yl] benzoic acid exhibit signifcantly higher docking score than approved drugs, C1 and C2. Molecular orbital, pharmacophore, drug likeness and ADMET predicted the idea about electrostatic pharmacological and nontoxic properties. Molecular dynamics simulation enriches the knowledge of stability of drug like molecule. We hope, will open new avenues to amide drug research. This computational prediction about a better tuberculosis drug will encourage and assist experimentalists to a great extent to design and synthesize potential new drugs for removal of this epidemic disease, spreading globally with a rapid span.

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