

Dual Biocontrol Potential of the Entomopathogenic Fungus, *Isaria javanica*, for Both Aphids and Plant Fungal Pathogens

Beom Ryong Kang^{a*}, Ji Hee Han^{b*}, Jeong Jun Kim^b and Young Cheol Kim^a 

^aDepartment of Applied Biology, College of Agriculture and Life Sciences, Chonnam National University, Gwangju, Republic of Korea; ^bAgricultural Microbiology Division, National Institute of Agricultural Sciences, RDA, Wanju, Republic of Korea

ABSTRACT

Dual biocontrol of both insects and plant pathogens has been reported for certain fungal entomopathogens, including *Beauveria bassiana* and *Lecanicillium* spp. In this study, we demonstrate, for the first time, the dual biocontrol potential of two fungal isolates identified by morphological and phylogenetic analyses as *Isaria javanica*. Both these isolates caused mortality in the greater wax moth, and hence can be considered entomopathogens. Spores of the isolates were also pathogenic to nymphs of the green peach aphid (*Myzus persicae*), with an LC₅₀ value of 10⁷ spores/mL 4 days after inoculation and an LT₅₀ of 4.2 days with a dose of 10⁸ spores/mL. *In vitro* antifungal assays also demonstrated a strong inhibitory effect on the growth of two fungi that are pathogenic to peppers, *Colletotrichum gloeosporioides* and *Phytophthora capsici*. These results indicate that *I. javanica* isolates could be used as novel biocontrol agents for the simultaneous control of aphids and fungal diseases, such as anthracnose and Phytophthora blight, in an integrated pest management framework for red pepper.

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The rapid adoption of organic agricultural practices during the past decade has markedly increased the world-wide demand for plant biopesticides that can be used to control weeds, insect pests, and microbial/viral pathogens. In order to meet this need, the biopesticide industry is eagerly searching for new and effective active agents. Many plant-associated microbes act as plant probiotics, in part because they produce biopesticides [1]. Biocontrol microbes are selected because of their antagonistic activity against a primary target, such as plant pathogens or harmful insects; however, many isolates have been found to have multiple beneficial effects on plant health. For example, certain bacterial biocontrol species, including species of *Pseudomonas* and *Bacillus*, enhance plant growth and induce systemic resistance, which can augment tolerance to plant diseases and abiotic stress [2–6].

Many entomopathogenic fungal isolates have been commercialized world-wide in various formulations as mycoinsecticides for the control of phytophagous insects [7]. However, certain entomopathogenic fungi are also known to directly inhibit plant microbial pathogens, induce systemic resistance, and enhance plant growth [4]. These useful traits are associated with the production of

antimicrobial and/or insecticidal products [8,9]. For example, the peptide isaferin, extracted from the mycelia of *Isaria felina*, is used as an insecticide for control of the oriental armyworm *Leucania separata*, and also functions as a mycocide in control the plant pathogenic fungi *Rhizoctonia solani* and *Sclerotinia sclerotiorum* [10]. Furthermore, certain strains of *Beauveria bassiana* produce a broad-spectrum antimicrobial compound, beauvericin [11,12], which is also produced by other entomopathogens, including isolates of *Paecilomyces*, *Isaria*, and *Fusarium* [13]. The production of beauvericin may also contribute to the suppression of damping-off caused by *R. solani* or *Pythium myriotylum* in tomato and cotton [12]. Isolates of *I. fumosoroseus* and *I. farinosus* control powdery mildew through mycoparasitism of this obligate plant pathogen [14,15]. Furthermore, extracellular chitinase has been proposed as a key virulence factor in insect mortality caused by *I. fumosorosea* [16]. Thus, secreted metabolites and enzymes may be involved in the control of insects and fungal pathogens.

Interestingly, some entomopathogenic fungi with the potential to control plant diseases, including species of *Lecanicillium*, *Beauveria*, and *Metarhizium*,

CONTACT Young Cheol Kim  yckimyc@jnu.ac.kr

*These authors contributed equally to this paper.


 Supplemental data for this article can be accessed [here](#).

Table 1. Comparison of the size and shape of conidia from *Isaria javanica* isolates pf185 and pf212.

<i>I. javanica</i> Isolate ^a	Conidial shape	Conidia (n = 40, μm)		Phialides (n = 70, μm)	
		Length	Width	Length	Width
Pf185	Cylindrical to fusiform	4.92 ± 1.76	2.05 ± 0.50	7.98 ± 2.81	1.57 ± 0.27
Pf212	Cylindrical to fusiform	7.16 ± 1.41	2.33 ± 0.53	7.71 ± 1.79	1.65 ± 0.42
<i>p</i> values		<i>p</i> < 0.0001	<i>p</i> = 0.017	<i>p</i> = 0.491	<i>p</i> = 0.190

^aIsolates were grown on potato dextrose agar for 5 d at 25 °C to facilitate spore formation. Data were analyzed with Student's *t*-test (*p* < 0.05) using SPSS (version 23; SPSS Inc.).

are plant endophytes [4,11,12,17–19]. Given that no additional biocontrol agent applications are required once such isolates have colonized a plant, they are ideal for utilization as formulations in integrated pest management under field conditions [20–22].

In the present study, we initially screened isolated fungi for their entomopathogenic activity, and subsequently examined whether they were inhibitory to the growth of plant pathogenic fungi, thereby establishing whether these isolates have dual biocontrol activity. On the basis of morphological and molecular phylogenetic analyses, we identified two isolates of *I. javanica* with such biocontrol potential. These isolates showed potential for use in the field as part of cost- and labor-effective integrated pest management programs.

To isolate potential entomopathogenic fungi, 10 larvae of the greater wax moth *Galleria mellonella* were placed in sterile Petri dishes, each containing soil collected from various agricultural fields in Korea. The larvae of *G. mellonella* are model insects used to isolate microbial entomopathogens [20]. After incubation for 14 d at 25 °C, infected larvae were suspended in sterile water by vortexing vigorously. To obtain pure fungal isolates, the suspension was serially diluted and plated out on potato dextrose agar (PDA; Difco Inc. Detroit, MI, USA) containing streptomycin (50 μg/mL) to prevent bacterial growth. Single fungal colonies were picked and cultivated on PDA. Segments of agar containing mycelium of the purified isolates grown on PDA plates were transferred to 10% sterile glycerol and stored at –80 °C.

Each isolate causing mortality in the moth larvae was initially examined for its ability to inhibit growth of the fungus *Colletotrichum gloeosporioides* (KACC 40896), obtained from the Korean Agricultural Culture Collection (KACC) National Agrobiodiversity Center, Wanju, South Korea. This fungus is the causal agent of anthracnose, one of the most important limiting factors for pepper production in Korea [21]. The entomopathogenic fungal isolate and the pepper pathogen were simultaneously inoculated onto PDA by placing 7 mm² plugs of the respective mycelia, previously grown on PDA, 5 cm apart from each other. Growth inhibition of the fungal pathogen was determined 7 d after initiating co-incubation. From an initial screen of over 200 different entomopathogenic fungal isolates, two

isolates, pf185 and pf212, were selected for further study because they showed the strongest antifungal activity against *C. gloeosporioides*. These two isolates originated from soil collected in different regions of South Korea: isolate pf185 was isolated from soil collected from a location with GPS coordinates of latitude 36.496975 and longitude 126.361456, and isolate pf212 was isolated from soil collected at a location with GPS coordinates latitude 35.106851 and longitude 121.814057.

Phylogenetic analysis of the isolated entomopathogens was based on sequences derived from two genomic regions, the internal transcribed spacer (ITS) of rRNA genes and a partial sequence of the gene encoding β-tubulin [22,23]. The fungi were grown in potato dextrose broth (PDB) for 5 d at 25 °C, with agitation at 150 rpm. Total genomic DNA was extracted from the fungal mass using a DNeasy Plant Mini Kit (Qiagen Inc., Valencia, CA, USA), according to the manufacturer's instructions. Sequences were amplified by the polymerase chain reaction (PCR) using the primers ITS1 (5'-TCCGTAGGTGAACCTGCGG-3') and ITS4 (5'-TCCGTAGGTGAACCTGCGG-3') for the ITS region [23], and Bt2a (5'-GGTAACCAAATCGGTGCTGCTTTC-3') and Bt2b (5'-ACCCTCAGTGTAGTGACCCTTGGC-3') for the tubulin gene [22]. PCR reactions were performed using a Bioneer premix (Bioneer Inc., Daejeon, Korea), and products of approximately 560 bp and 330 bp were generated from the ITS region and the tubulin gene, respectively. The PCR products were purified using a purification kit (Bioneer) and cloned into a pGEM T-easy vector (Promega, Madison, WI, USA). Nucleotide sequences of the PCR products were generated by Solgent Inc. (Daejeon, Korea). These sequences have been deposited in GenBank (Supplemental Table 1) as LY490787.1 (ITS) and MH340496 (β-tubulin) for pf185, and LY490786.1 (ITS) and MH340497 (β-tubulin) for pf212.

Initial analysis using BLASTN (available at the National Center for Biotechnology Information) using sequences for the ITS region and the partial gene sequences for beta-tubulin of both isolates showed identity with *Isaria* spp. Therefore, the PCR sequences were aligned with the sequences of *Isaria* sp. ex-type strains obtained from GenBank (Supplemental Table 1), using CLUSTAL X, and

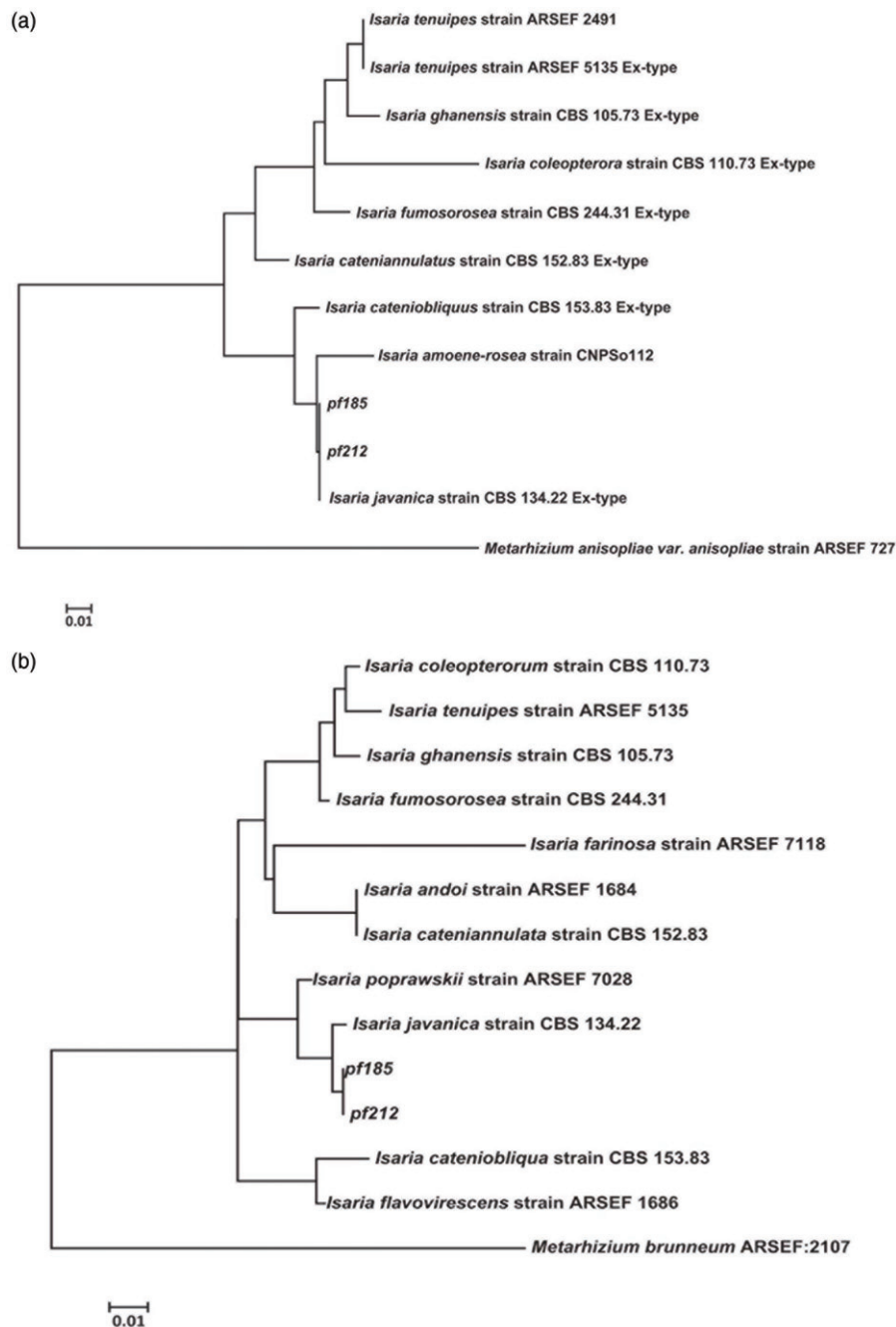


Figure 1. Phylogenetic trees based on the internal transcribed spacer (ITS, A) and partial β -tubulin (TUB2, B) sequences of isolates pf185 and pf212 and ex-type strains of the *Isaria* genus (Supplemental Table 1). The phylogenetic trees were constructed using the neighbor-joining method and Kimura's two-parameter model. The numbers at nodes indicate consensus bootstrap values based on 1,000 replications. Scale bar = 0.01 substitutions per nucleotide position. Sequences from *Metarhizium anisopliae* and *M. brunneum* were used as the outgroups for ITS and β -tubulin sequence analyses, respectively. Abbreviations: ARSEF, Agricultural Research Service Collection of Entomopathogenic Fungal Cultures (Ithaca, NY, USA); CBS, Centraal Bureau voor Schimmelcultures (Utrecht, The Netherlands).

edited using BioEdit 7 [24]. Phylogenetic trees were generated using MEGA 7 software [25] based on the neighbor-joining method, with the Kimura two-parameter model (Kimura, 1980) and bootstrap values based on 1,000 replications. Phylogenetic analysis revealed that the ITS and β -tubulin sequences of both isolates were 99.99% identical to those of *I. javanica* ex-type CBS134.22 (Figure 1). Interestingly, our phylogenetic analysis indicated that *I. javanica* pf185 and pf212 belong to a distinct subgroup containing *I. fumosorosea* ex-type CBS244.31.

However, given that recent reports have indicated the incorrect identification of some *Isaria* isolates based on molecular analyses [26,27], we also examined the morphology of conidia and phialides for comparison with published data on isolates characterized as ex-types of the *Isaria* genus [27,28]. Isolates pf185 and pf212 were grown on PDA for 7 d at 25 °C to facilitate spore formation. Slides were prepared by suspending scrapes from these fungal cultures in sterile water, followed by light microscopy examination at $\times 200$, $\times 400$, and $\times 1,000$

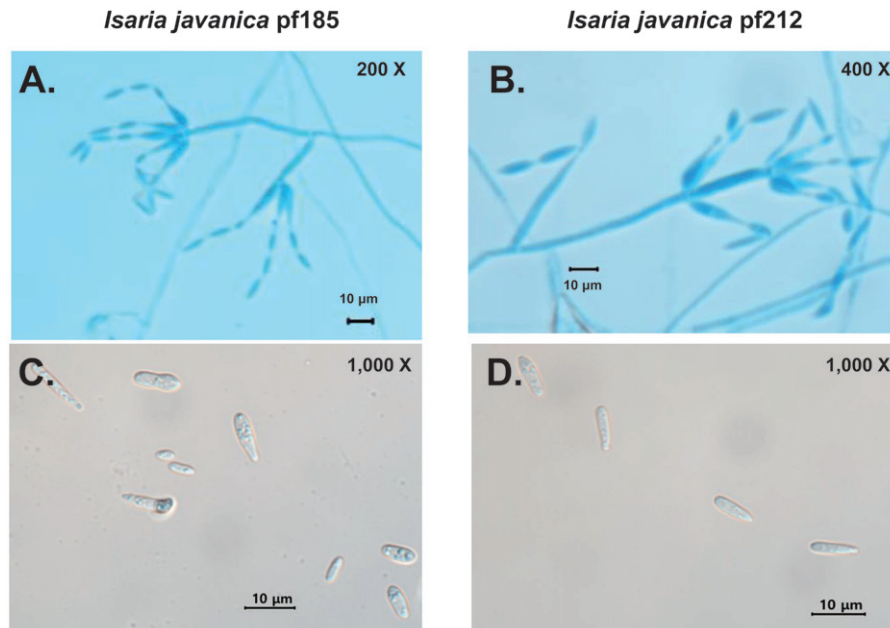


Figure 2. Morphology of the conidia and phialides of *Isaria javanica* isolates pf185 and pf212. Phialides with developing conidial chains of *I. javanica* pf185 (A) and *I. javanica* pf212 (B). Typical shapes of individual conidia of *I. javanica* pf185 (C) and *I. javanica* pf212 (D). Scale bars represent 10 μm . The images are representative of two independent observations with at least 40 conidia and 70 phialides.

magnification (Leica M165 FC; Leica Microsystems, Wetzlar, Germany). For observation of phialides, samples were stained with lactophenol cotton blue with iodine glycerol reagent, as previously described [29]. Images were photographed digitally using a Leica DFC 450 C camera and analyzed using an Image analyzer (IMT i-Solution Inc., Scarborough, Canada). At least 40–70 independent measurements of the lengths and widths of conidia ($n=40$) and phialides ($n=70$) were made for each isolate. Observations were performed twice and data were analyzed with Student's t -test ($p < 0.05$) using SPSS (version 23; SPSS Inc., Chicago, IL, USA).

Morphological features of the pf185 and pf212 isolates and their comparison with the ex-type strain are summarized in Table 1 and illustrated in Figure 1. For pf185, spores were $4.9 \pm 1.8 \mu\text{m}$ in length and $2.1 \pm 0.5 \mu\text{m}$ in width (Table 1), whereas those for pf212 were longer at $7.2 \pm 1.4 \mu\text{m}$ ($t = -5.711$, $p < 0.0001$) but similar in width at $2.3 \pm 0.5 \mu\text{m}$ ($t = -2.436$, $p = 0.017$). Growth of other *I. javanica* isolates for 5 d on a different medium has been shown to result in conidial lengths of between 2 and 5 μm and widths of 1 to 2 μm [27,28]. However, all conidia were cylindrical to fusiform in shape, although unlike the conidia of other *I. javanica* isolates, the conidia from pf185 and pf212, were hyaline and not pink in color (Figure 2).

The phialides of the pf185 and pf212 isolates displayed a wide globose basal portion with a long distal neck, and overall lengths $8.0 \pm 2.8 \mu\text{m}$ and $7.7 \pm 1.8 \mu\text{m}$, and widths of $1.6 \pm 0.3 \mu\text{m}$ and

$1.6 \pm 0.4 \mu\text{m}$, respectively (Table 1 and Figure 2(A, B)). Although these measurements were not significantly different, ($t = 0.691$, $p = 0.491$ for length and $t = 318$, $p = 0.190$ for width) (Table 1), they tended to be longer and less wide than those of other *I. javanica* isolates, which showed lengths of 4–6 μm and widths of 2–3 μm [27,28]. Differences in growth conditions could account for the variability reported here. All phialides were smooth- and colorless, with few or no synnemata [27,28]. Consequently, combining information obtained from the phylogenetic and morphological analyses, the pf185 and pf212 isolates were classified as isolates of *I. javanica* and have been deposited in the KACC with designated numbers KACC93241P and KACC93240P, respectively.

To extend our characterization of the antifungal activity of isolates pf185 and pf212, *in vitro* coinoculation assays on PDA plates were conducted against additional phytopathogenic fungi, namely, *Fusarium oxysporum* (KACC 44452), a wilt pathogen of tomato, and *Rhizoctonia solani* (KACC 40111), the causative agent of damping-off in many crops. We also examined activity against the pepper phytophthora blight pathogen *Phytophthora capsici* (KACC 44716). Assays of growth inhibition caused by *I. javanica* isolates pf185 and pf212 on PDA were performed *in vitro*. Fungal growth was generally measured after a 7-d co-incubation, although for the fast-growing fungus *R. solani*, growth was recorded at 3 d. Growth of each fungal pathogen was measured as the average diameter of the colony size in four different directions. Percentage fungal growth inhibition was calculated as $[(A - B)/$

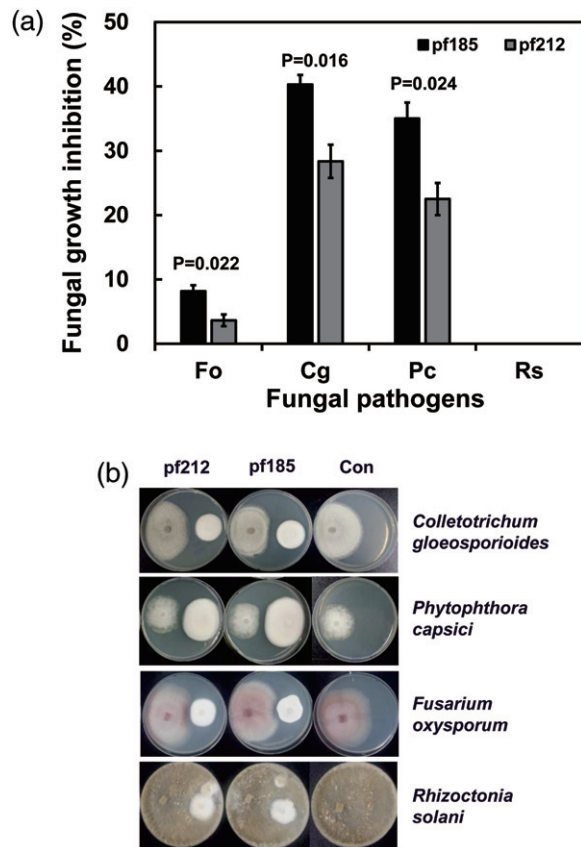


Figure 3. Growth inhibition of four plant pathogens caused by *Isaria javanica* pf185 and pf212 when grown in dual cultures on potato dextrose agar. Fo, *Fusarium oxysporum*; Cg, *Colletotrichum gloeosporioides*; Pc, *Phytophthora capsici*; and Rs, *Rhizoctonia solani*. (A) Error bars represent the mean standard error of 10 total replicates per treatment in two independent experiments. *P*-values are provided for the significant differences in growth under different treatments (Student's *t*-test, $\alpha < 0.05$). (B) The images were obtained at 7 d after co-cultivation. The fungal pathogen was inoculated on the left-hand side of the plate, and *I. javanica* pf185 or pf212 on the right-hand side. The images are representative of two independent experiments with five plates for each experiment.

$A] \times 100$, where *A* is the radial growth of the fungi in the absence of *I. javanica* and *B* is the radial growth in plates inoculated with both test organism and *I. javanica* isolate. The experiment was performed twice, with each treatment being replicated five times. Data were analyzed with Student's *t*-test ($p < 0.05$) using SPSS (version 23; SPSS Inc.).

In the dual growth assays, *I. javanica* pf185 was marginally more antagonistic than isolate pf212. Although inhibition was strongest against *C. gloeosporioides*, it was still evident against *P. capsici*. Substantially lower inhibition was observed against *F. oxysporum*, whereas no inhibition was observed against *R. solani* (Figure 3), which is in contrast to previous reports for an isolate of *I. felina* [10]. Thus, it will be useful to explore the extent of differences in the biocontrol potential of different *Isaria* genus isolates.

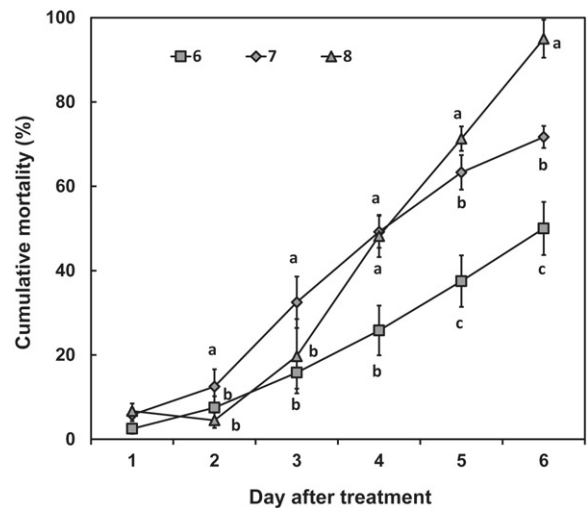


Figure 4. The percentage cumulative mortality of second-instar green peach aphid nymphs, following treatment with spore suspensions of *I. javanica* pf185. The nymphs were treated with spores at concentrations of 10^6 (6), 10^7 (7), and 10^8 (8) spores/mL (8), as described in the text. Error bars represent the mean standard error of three replicates per treatment in two independent experiments. Data were analyzed using ANOVA ($p < 0.05$), and if the *F* test was significant, differences were further elucidated using Duncan's multiple range test. Different letters indicate significant differences at $p < 0.05$ at each time point.

Results obtained for the aphicidal activities of isolates pf185 and pf212 against the green peach aphid (*M. persicae*) were essentially similar, and therefore here we only present the data for pf185. An aphid culture, obtained from Dr. Han of the Agricultural Microbiology Division, National Academy of Agricultural Science, was propagated on Chinese cabbage. Chinese cabbage seedlings were planted into sterile soil-less medium (peat moss:vermiculite:perlite, 7:3:3, vol/vol/vol) with one seedling per pot, which was watered every 2 d. Healthy apterous adult aphids were transferred onto young cabbage leaves, with approximately 20 aphids per leaf, and allowed to reproduce for 2 d to produce the 1–2-d-old second-instar nymphs used for bioassays. Sections of Chinese cabbage leaves (3.5×3.5 cm) were placed in an insect breeding dish (10×40 cm; SPL Science, Daejeon, Korea) containing sterile cotton, soaked in distilled water, and inoculated with 20 second-instar nymphs by transfer with a soft brush. Suspensions of *I. javanica* pf185 conidia diluted to contain 10^{6-8} spores/mL were applied directly in 20- μ L aliquots onto the nymphs using a micropipette. The conidia were obtained from cultures grown for 7 d at 25 °C on PDA by flooding the plates with sterile water followed by filtration through two layers of sterile cheese cloth to remove hyphal debris. Spore numbers were determined using a hemocytometer. After application of spore suspensions, or sterile water as a control, the insect breeding dishes were incubated at 25 °C and aphid mortality was monitored daily

Table 2. Probit analysis of larval mortality for estimation of the median lethal concentrations of *Isaria javanica* pf185.

Days after treatment	Lethal concentration (Log ₁₀ value spore/mL) (95% confidence limit) ^a	Regression analysis and coefficient ^b	
4	LC ₅₀ = 7.0 (6.8–7.3) LC ₉₀ = 9.5 (8.8–10.9)	$y = -8.21 + 9.71 * x$	$R^2 = 1.000$
5	LC ₅₀ = 6.5 (6.2–6.7) LC ₉₀ = 8.6 (8.1–9.6)	$y = -8.27 + 10.21 * x$	$R^2 = 0.995$

^aLC₅₀ and LC₉₀, median lethal concentration of spore values, 95% confidence limit, the overall parameter is 95% in this range.

^bLinear regressions, where the concentration of *I. javanica* pf185 (X) is the independent variable and aphicidal activity (Y) is the dependent variable, were obtained at 4 or 5 d after exposure. We calculated the Spearman correlation coefficient (R-value) and its corresponding p-value.

Table 3. Probit analysis of detached-leaf bioassays for estimation of changes in the lethal times of *Isaria javanica* pf185 at different spore concentrations for second-instar nymphs of the peach aphid.

Spore concentration (log ₁₀ value spores/mL)	Lethal time (days) (95% confidence limits) ^a	Regression line and coefficient ^b	
8	LT ₅₀ = 4.2 (3.2–4.7) LT ₉₀ = 6.4 (6.0–7.3)	$y = -4.61 + 7.32 * x$	$R^2 = 0.959$
7	LT ₅₀ = 5.0 (3.8–5.5) LT ₉₀ = 10.3 (8.3–24.0)	$y = -2.86 + 4.08 * x$	$R^2 = 0.995$
6	LT ₅₀ = 7.0 (6.5–8.8) LT ₉₀ = 13.6 (10.1–38.7)	$y = -3.76 + 4.44 * x$	$R^2 = 0.999$

^aLT₅₀ and LT₉₀, median lethal time values; 95% confidence limits, the overall parameter is 95% in this range.

^bLinear regressions, where the lethal day (X) is the independent variable and aphicidal activity of *I. javanica* pf185 (Y) is the dependent variable, were obtained for different spore applications. We calculated the Spearman correlation coefficient (R-value) and its corresponding p-value.

for 7 d. Nymphs were considered dead when they failed to move after multiple prods with a fine paint brush. Dead aphids were removed from the colonies and examined for overt mycosis after overnight incubation in a moist chamber at 25 °C. The total mortality for each treatment was corrected according to Abbott's formula based on mortality observed in the control [30]. Corrected mortality (%) = (1 - number in fungal treatment/number in control treatment) × 100. All experiments were performed with two independent treatments, each of which was replicated three times, using 20 second-instar nymphs per treatment. Data were analyzed with ANOVA ($p < 0.05$) using SPSS (version 23; SPSS Inc.), and if the *F* test produced a significant result, differences in each measurement were further examined using Duncan's multiple range test ($p < 0.05$). The lethal concentration (LC) and lethal time (LT) values were grouped using Tukey's post-hoc test, if variance analysis results were significant at the 95% confidence level using SPSS. LC and LT values were also examined with Probit analysis [31] using SPSS. Regression analysis between the aphicidal activity and the LCs or LTs of *I. javanica* pf212 were determined, and Spearman correlation coefficients (R-values) and corresponding p-values were determined using SPSS.

The cumulative mortalities caused by application of spores of the pf185 isolate were dependent upon both spore concentration and days after treatment. Mortality gradually increased with time after treatment, and after 4 d an effect of spore concentration was observed (Figure 4). More than 50% mortality was observed at 4 d with spores applied at concentrations of between 10⁷ and 10⁸ spores/mL (Figure 4). Aphids treated with 10⁶ spores/mL showed less than 50% mortality even after

exposure for 6 d. For second-instar nymphs sprayed with 0.02% Tween 80 as a control, the average mortality after 6 d was 10%. In contrast, more than 90% mortality was observed after 6 d among aphids treated with 10⁹ spores/mL (Figure 4).

Calculations indicate that the LC₅₀ for *I. javanica* was log₁₀ = 7 at 4 d, and log₁₀ = 6.5 at 5 d. For LC₉₀, the titer was log₁₀ = 9.5 (with 95% confidence limits of 8.8–10.9) at 4 d, and log₁₀ = 8.6 (with 95% confidence limits of 8.1–9.6) at 5 d. The LT₅₀ increased in treatments with lower spore concentrations (Table 3). The LT₉₀ was 6.4 d (with 95% confidence limits of 6.0–7.3) for 10⁸ spores/mL, 10.3 d (with 95% confidence limits of 8.3–24.0) for 10⁷ spores/mL, and 13.6 d (with 95% confidence limits of 10.1–38.7) for 10⁶ spores/mL (Tables 2 and 3). On the basis of these values, we estimated that the effective concentration (LC₉₀) and time (LT₉₀) for successful control of green peach aphids are 10⁸ spores/mL and 6 d, respectively.

The pattern of aphid mortality caused by isolates pf185 and pf212 is consistent with that previously observed for other *Isaria* isolates: over 70% mortality after 9–15 d exposure in brown citrus aphids treated with an isolate of *I. fumosorosea*, with 100% mortality occurring by day 22 [32]. Our previous studies using *I. javanica* isolate pf04, showed high virulence against the sweet potato whitefly [33,34], with LT₅₀ and LT₉₀ values of 3.8 and 5 d following application of 10⁹ spores/mL, respectively [33]. These findings from the whitefly and aphid studies indicate that prolonged exposure and high spore densities are required for effective insect control. Thus, the medium for growth must be optimized. Previously, we found that *I. javanica* pf04, had optimal conidia formation when grown under solid state

fermentation on a barley substrate supplemented with silkworm powder and moth larvae [33]. The demonstration of antifungal activities in dual culture assays on PDA indicates that extracellular products are involved, and dibutyl succinate from pf185 was identified as a key metabolite for antifungal and aphicidal activity [35]. These findings accordingly support the possibility of the production of effective cross-kingdom metabolites by other entomopathogens [8,9]. Consequently, because maximizing the dual biocontrol potential of strains is important, we are examining the effect of various growth conditions, including nutrition, temperature, and length of culture, utilizing *in vitro* assays for fungal growth antagonism.

In summary, we found that two isolates of the entomopathogenic fungus *I. javanica* showed dual biocontrol activities against different plant pathogenic fungi and the larvae and nymphs of moths and aphids, respectively. Isolate identification was based on a comparison of base sequences from two conserved fungal loci and the morphology of their spores and phialides with those of ex-type strains of the *Isaria* genus. Studies are currently in progress with the aim of optimizing growth conditions for the production of both high conidial concentrations and extracellular metabolites with antifungal activity. The selected *I. javanica* isolates could be used as novel biocontrol agents for dual biocontrol of insects and fungal diseases in red pepper.

Disclosure statement

No potential conflict of interest was reported by the authors.

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ORCID

Young Cheol Kim  <http://orcid.org/0000-0002-7661-7600>

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