



Article

Combination of Berberine with Resveratrol Improves the Lipid-Lowering Efficacy

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Abstract: The natural compound berberine has been reported to exhibit anti-diabetic activity and to improve disordered lipid metabolism. In our previous study, we found that such compounds upregulate expression of sirtuin 1—a key molecule in caloric restriction, it is, therefore, of great interest to examine the lipid-lowering activity of berberine in combination with a sirtuin 1 activator resveratrol. Our results showed that combination of berberine with resveratrol had enhanced hypolipidemic effects in high fat diet-induced mice and was able to decrease the lipid accumulation in adipocytes to a level significantly lower than that in monotherapies. In the high fat diet-induced hyperlipidemic mice, combination of berberine (30 mg/kg/day, oral) with resveratrol (20 mg/kg/day, oral) reduced serum total cholesterol by 27.4% \pm 2.2%, and low-density lipoprotein-cholesterol by $31.6\% \pm 3.2\%$, which was more effective than that of the resveratrol (8.4% \pm 2.3%, 6.6% \pm 2.1%) or berberine (10.5% \pm 1.95%, 9.8% \pm 2.58%) monotherapy (p < 0.05 for both). In 3T3-L1 adipocytes, the treatment of 12 μmol/L or 20 μmol/L berberine combined with 25 μmol/L resveratrol showed a more significant inhibition of lipid accumulation observed by Oil red O stain compared with individual compounds. Moreover, resveratrol could increase the amount of intracellular berberine in hepatic L02 cells. In addition, the combination of berberine with resveratrol significantly increases the low-density-lipoprotein receptor expression in HepG2 cells to a level about one-fold higher in comparison to individual compound. These results implied that the enhanced effect of the combination of berberine with resveratrol on lipid-lowering may be associated with upregulation of low-density-lipoprotein receptor, and could be an effective therapy for hyperlipidemia in some obese-associated disease, such as type II diabetes and metabolic syndrome.

Keywords: berberine; resveratrol; sirtuin 1; high fat diet; low-density-lipoprotein receptor

1. Introduction

Obesity is recognized as a major risk factor for clinically obesity-associated cardiometabolic complications, including metabolic syndrome components, type 2 diabetes, and cardiovascular diseases [1,2]. Excessive adiposity, especially visceral fat, as an endocrine organ, is associated with overmuch plasma free fatty acid-induced lipotoxicity and a chronic proinflammatory state by releasing

excessive inflammatory cytokines, such as tumor necrosis factor- α (TNF- α), resulting in insulin resistance, hyperglycemia, dyslipidemia, hypertension, and a prothrombotic state [3–5]. Therefore, ameliorating dyslipidemia and fat deposition may contribute to reverse the chronic proinflammatory state, and then to delay or prevent the morbidity of these obese-associated diseases.

Berberine (Ber), an active nature component of traditional Chinese herb Coptidis Rhizoma, attracted many researchers' attention as a new hypolipidemic drug, which showed potent effect against hyper-cholesteremia and type II diabetes. Moreover, Ber reduced adipocytes differentiation and lipid accumulation by inhibiting the expression of lipogenic genes. However, transient gastrointestinal adverse effects, including diarrhea, flatulence, constipation, could be observed in clinical trials because of a high dose of Ber alone (500 mg/2 or 3 times a day) [5–8]. For example, 10.3% diarrhea, 19.0% flatulence, and 6.9% constipation were occurred in patients after administration of Ber alone in a clinical trial. In addition, all of these adverse effects disappeared in one week after reduction in berberine dosage [9]. In our previous study, it was found that Ber could resist oxidative stress induced apoptosis or cellular senescence by upregulation of sirtuin 1 (SIRT1) expression at a low dosage [10,11]. SIRT1 was regarded as a master metabolic sensor by mimicking the beneficial effect of caloric restriction in mammals [12]. Resveratrol (Res), a chemical activator of SIRT1, also exhibited improvement of lipid metabolism in pathologic state, such as obesity, type II diabetes and atherosclerosis [13,14]. Therefore, in this study, we investigated the effect of Ber in combination with Res on lipid metabolism in hyperlipidemia animal and 3T3-L1 adipocytes. The possible enhanced mechanism of Ber in combination with Res was also studied.

2. Results

2.1. The Anti-Hyperlipidemic Effect of Combined Therapy with Berberine and Resveratrol in Hyperlipidemic Mice and 3T3-L1 Adipocytes

The level of plasma total cholesterol (TC), triglyceride (TG) and low-density lipoprotein-cholesterol (LDL-c) in high fat diet (HFD) mice after administration of Ber and/or Res was shown in Figure 1. Compared with control low fat diet (LFD) group, the level of plasma TC, TG and LDL-c in HFD group were significantly increased (p < 0.01). Compared with HFD group administrated with equal volume of 0.9% saline, the level of TC, TG and LDL-c were decrease by 27.4% \pm 2.2%, 17.2% \pm 2.42% and 31.6% \pm 3.2%, respectively after administration of 30 mg/kg/day Ber in combination with 20 mg/kg/day Res. However, the monotherapy of Res or Ber showed an 8.4% \pm 2.3% or 10.5% \pm 1.95% reduction in TC, 10.3% \pm 3.42% or 6.9% \pm 2.6% reduction in TG and 6.6% \pm 2.1% or 9.8% \pm 2.58% reduction in LDL-c, respectively. The combination therapy demonstrated a superior hypolipidemic efficacy in level of plasma TC and LDL-c to monotherapy (p < 0.05), but there was no differentiation in plasma TG.

The effects of Ber, Res and their combination on intracellular lipid accumulation were also tested in the established 3T3 L1 adipocytes. As shown in Figure 2, the treatment of Res or Ber alone reduced intracellular lipid accumulation in a dose-dependent manner. However, the lipid accumulation decreased by 40% and 48%, respectively, after treatment with the combination of Res (25 μ mol/L) and Ber (12 or 20 μ mol/L). The results revealed a more significant lipid-accumulation-reducing effect by the combination therapy than that of any monotherapy (p < 0.05).

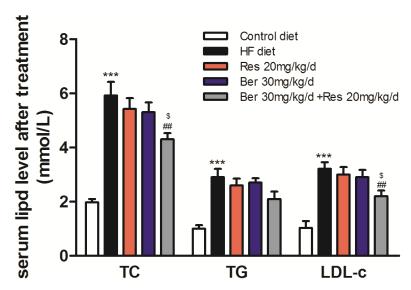


Figure 1. The lipid-lowering effect of Ber, Res and their combination in hyperlipidemic C57BL/6J mice. As Materials and methods mentioned, serums were isolated from mice after different drug treatments. The total cholesterol (TC), Triglyceride (TG) and LDL-cholesterol (LDL-c) levels were measured. Values are mean \pm SD of all of mice in each group (n = 8). *** p < 0.001 vs. that of the control low diet group; ** p < 0.01 vs. that of the HFD group; \$ p < 0.05 vs. that of Ber or Res monotherapy. The result is a representative for three separate experiments.

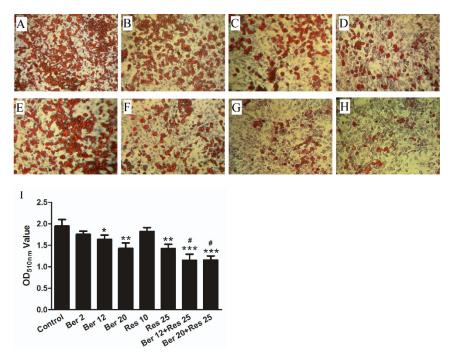


Figure 2. Effect of Ber, Res and their combination on the lipid accumulation in adipocytes. Differentiated 3T3-L1 adipocytes were established by inducing for 10 days (details indicated in Material and Methods) and then treated with different concentration of Ber (**B–D**: 2, 12, 20 μ mol/L), Res (**E–F**: 10, 25 μ mol/L) and their combination (**G–H**: 12 μ mol/L Ber + 25 μ mol/L Res, 20 μ mol/L Ber + 25 μ mol/L Res) for 3 days. Control adipocytes were treated with the vehicle (DMSO < 0.1%) (**A**). Lipid droplets in adipocytes were detected by oil red O staining. The magnification of Microscope for observation was 10 (**I**) the absorbance density of Oil red dye extracted from differentiated adipocytes after Ber and Res treatment. *** p < 0.001, ** p < 0.01, * p < 0.05 vs. control, # p < 0.05 vs. single drug. Data were presented as mean \pm SD from three separate experiments.

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2.2. Resveratrol Increased the Accumulation of Berberine in Hepatic L02 Cells

Ber can be detected by flow cytometry after uptake and accumulation in living cells because of its fluorescence property [15,16]. Indeed, as shown in Figure 3A, the mean fluorescence intensity (MFI) of Ber was increased significantly in a dose-dependent manner. For example, the MFI value of Ber in hepatic cells after treatment with 50 μ mol/L was almost four-fold that of control and 2.4 times that of treatment with 12.5 μ mol/L Ber. Intriguingly, MFI value in hepatic cells after exposure to 50 μ mol/L Ber and 10 or 25 μ mol/L Res was increased significantly to 1.4 times that of the single Ber treatment, and the results revealed that resveratrol could increase the accumulation of berberine in hepatocytes.

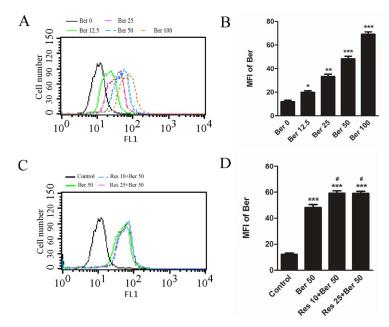


Figure 3. Intracellular accumulation of Berberine enhanced by Res in Hepatic L02 cells. (**A,B**) intracellular fluorescence of Ber in a dose-dependent manner (0–100 μ mol/L), and mean fluorescence intensity (MFI) calculated and compared in different dose of Ber groups. *** p < 0.001, ** p < 0.01, * p < 0.05 vs. untreated control. (**C,D**) Res at 10 or 25 μ mol/L added in advance (details indicated in Material and Methods) enhanced intracellular fluorescence of Ber at 50 μ mol/L, MFI calculated and compared, *** p < 0.001 vs. untreated control, * p < 0.05 vs. single Ber treatment. Data shown are representative of three separate assays.

2.3. The Combined Therapy Can Increase Low-Density-Lipoprotein Receptor Expression in Hepatocytes

It was reported that the cholesterol lowering effect of Ber was closely associated with its regulation of low-density-lipoprotein receptor (LDLR) expression [6]. Therefore, the effect of combination of Ber with Res on LDLR expression was analyzed. Firstly, the LDLR expression in cells cultured with different concentrations of fetal bovine serum (FBS) was examined in order to evaluate the effects of FBS. As shown in Figure 4A, the expression level of LDLR increased gradually in a serum concentration-dependent manner. When the serum concentration exceeds 5%, the expression of the LDLR receptor no longer increases. Therefore, in order to exclude interference of FBS on LDLR expression, culture medium with 1% FBS was used to detect the effects of the combination of Res and Ber on LDLR expression. The expression of mature LDLR in the combined treatment group (Ber at $12 \ \mu mol/L$ and Res at $10 \ \mu mol/L$) significantly increased by 50% compared with the single treatment group as revealed by Figure 4B.

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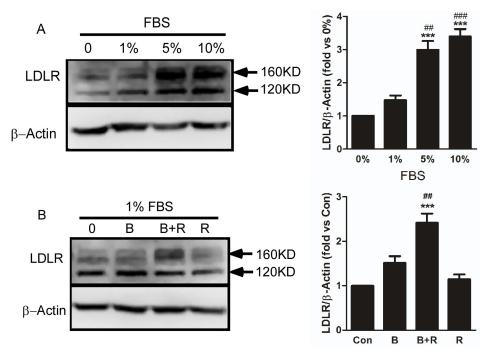


Figure 4. The combination of berberine with resveratrol increased LDLR expression in HepG2 cells. Cells were cultured in 6-well plate for 24 h with a 3×10^5 cell density, and then culture medium were replaced by fresh medium containing different concentration of FBS indicated in A, or 1% FBS and drugs indicated in B for another 24 h followed by extraction of the total proteins from the cells. A: the effect of different concentration FBS on LDLR expression were analyzed by western blot assay. Then the band intensity was quantified by grey scanning analysis, and the intensity ratio of LDLR to β-actin in 0% FBS group was set to 1. *** p < 0.001 vs. 0% FBS group, *## p < 0.001, ## p < 0.01 vs. 1% FBS group. B: effect of the combination 12 μmol/L berberine (B) with 10 μmol/L resveratrol (R) on LDLR expression. The intensity ratio of LDLR to β-actin in untreated control group was set to 1. *** p < 0.001 vs. untreated control, ## p < 0.01 vs. single Ber or Res treatment. The results are representative of three separate experiments.

3. Discussion

Numerous active nature compounds extracted from medicinal plants are used for treatment of dyslipidemia in some diseases, such as morbid obesity or metabolism syndrome, many of which are the sources of currently prescribed synthetic drugs [17]. The present study was to explore the efficacy and possible mechanisms of the combination of nature compounds, Berberine and Resveratrol. Our results demonstrated that the combination of two agents exerted a significant enhanced effect on lipid-lowering in hyperlipidemic mice and intracellular lipid accumulation-decreasing in adipocytes as compared to individual agent alone, which was closely associated with SIRT1 and LDLR.

Berberine is an alkaloid isolated from many plants, such as *Coptidis Rhizoma*, *Hydrastis Canadensis*. It was found that Ber could be not only a new cholesterol-lowering drug by stabilizing LDLR messenger RNA [6], but also ameliorate whole dysregulation of lipid metabolism to decrease development of fatty liver by activating AMP-dependent protein kinase (AMPK) and regulating SIRT1 in animal experiments [18,19]. Resveratrol, a polyphenol from several plants, has showed beneficial effect on obesity, diabetes and cardiovascular disease as a SIRT1 activator. Reports have also showed that Res and cholesterol-lowering drug statin in combination not only enhanced the lipid-lowering efficacy, but also increase myocardial protection in hypercholesterolemia rats [13,14,20]. Owing to their common target, SIRT1, we speculated that a better efficacy of regulating lipid metabolism may be achieved by Ber and Res in combination. Indeed, in vivo results showed that when combined with Res at 20 mg/kg/day, the dose of Ber reduced by 70% still achieved an effective activity of lowering plasma LDL-c, TG and TC in hyperlipidemia mice, which exhibited significant 27.4% decrease in TC

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and 31.6% decrease in LDL-c (p < 0.05), but no significance in TG. However, monotherapy of Ber at 30 mg/kg/day or Res at 20 mg/kg/day had not significant effect on decrease of plasma LDL-c, TC and TG in hyperlipidemia mice. As compared with monotherapy, combination therapy showed significant 18.9% and 24.1% decrease in TC and LDL-c of, respectively (p < 0.05). In animal studies, the dose of hypolipidemic efficacy of Ber was commonly used at 100 mg/kg/day or above [6,21]. However, in our study, the only less one third of 100 mg/kg/day Ber administration in combination with Res could also exhibit a significant lipid-lowering effect. In addition, in differentiated 3T3-L1 adipocytes, our results showed that a decrease of intracellular lipid accumulation was observed in adipocytes treated with Ber or Res alone in a dose dependent manner. However, the lipid accumulation was significantly decreased when adipocytes were treated with Ber at 12 μ mol/L or 24 μ mol/L combined with Res at 25 μ mol/L, which was better than single drug treatment (p < 0.05). These data suggested that combination therapy may be a better way to clinic application of hyperlipidemia than monotherapy of Ber because of efficacy at low dose.

Although there are some adverse effects of Ber in clinic monotherapy for hyperlipidemia and diabetes, some studies revealed that multidrug resistance of Ber may occur in a long-term single administration because of Ber being the substrate for p-glycoprotein, which resulted in a decrease of its efficacy in LDLR regulation [9,22,23]. Moreover, in previous studies, it showed that Res had a reversal effect on multidrug resistance of cancer chemotherapy by inhibition of p-glycoprotein function or downregulation of p-glycoprotein expression [24,25]. Therefore, it was interesting to examine whether the enhanced effect of their combination was partially related with the increased accumulation of Ber in cells caused by Res. As a fluorescent dye for cell, mitochondria and DNA analysis, Ber can be detected through flow cytometry at 550 nm under excitation light [15,16]. As shown in Figure 3, the mean fluorescence intensity (MFI) of hepatocytes after treatment with Ber increased in a concentration-dependent manner, which reflected the accumulation of intracellular Ber. After exposure to the combined agents, the MFI values of cells increased more significantly than that of the single Ber treatment. Meanwhile, in some reports [26] or our study, single Res showed a non-fluorescent property in hepatocytes detected by flow cytometry (Figure S2). Therefore, the increased MFI values were due to the increased accumulation of Ber within the cells. However, the intracellular accumulation of Ber remained unchanged when the dose of resveratrol increased. This data implied that the enhanced effect of the combination may be partially explained by the increased accumulation of Ber in cells that was caused by Res treatment.

One of the important mechanisms for hypolipidemic function of Ber is the involvement in cholesterol-lowering by upregulation LDLR expression [6]. The formation process of functional LDLR begins from a precursor-like receptor in endoplasmic reticulum to a mature receptor after being O-linked glycosylated in the Golgi apparatus, and migrating to cell membrane [2]. Meanwhile, it was reported that serum factors could upregulate LDLR expression [27]. This was confirmed by our study that normal concentration of serum (10%) in cell culture medium have a direct effect on increasing the level of mature LDLR. To remove the influence of serum, the cells were cultured in medium with low concentration of serum (1%), and then were treated with Ber plus Res. The results revealed that the combination treatment could significantly increase the level of mature LDLR compared with single Ber treatment. These data implied that the reason for enhanced efficacy of combination may be associated with increase of the intracellular accumulation of Ber by Res, which enhanced hypolipidemic effect by Ber-induced LDLR expression. However, in some report, deletion of SIRT1 could decrease the level of LDLR mRNA and a SIRT1-specific activator could enhance LDLR expression [28,29]. In view of the upregulation SIRT1 protein of Ber (Figure S1) and activation SIRT1 enzyme activity of Res, another explanation may be not rule out that SIRT1 targeted by Ber and Res may be partially participating in upregulation of LDLR expression.

In conclusion, our results demonstrated that berberine at low dose in combination with resveratrol may be a better therapy than monotherapy during the lipid metabolism disorder scenario, which was closely associated with enhanced LDLR expression by Res-mediated intracellular accumulation of Ber.

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It might imply that this combination could be an effective trial for clinical obesity-associated disease therapy, such as type II diabetes, metabolic syndrome.

4. Materials and Methods

4.1. Experimental Animals and Study Protocol

Female C57BL/6J mice were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd (Beijing, China) at 7–8 weeks of age, accommodated for one week after receipt, and maintained at a temperature and humidity of 25 °C and 55%, respectively and a 12 h:12 h light:dark cycle. All mice were randomly assigned to either control low fat diet (LFD) (Research Diets number D12450J, 10% kcal from fat) or high fat diet (HFD) (Research Diets number 12,492, 60% kcal from fat) for 12 weeks. Then Mice with HFD were divided randomly into five groups and administrated orally with different dose of drugs for 4 weeks: (1) 0.9% saline at equal volume, (2) Ber at 30 mg/kg/day, (3) Res at 20 mg/kg/day, (4) Ber at 100 mg/kg/day, (5) Ber at 30 mg/kg/day plus Res at 20 mg/kg/day. Ber was administrated twice a day at 8 A.M. and 5 P.M., respectively, whereas Res was given once a day at 5 P.M. Berberine and Resveratrol was purchased from Hangzhou Huadong Medicine Group Kangrun Pharmaceutical Co., Ltd. (Hangzhou, China).

Blood samples were taken after the treatment by intraocular canthus venous plexus after 6-h fast. Serums were isolated by centrifugation. Serum LDL-c, total cholesterol (TC), and triglyceride (TG) levels were assayed by Standard enzymatic methods with commercially available kits purchased from Najing jiancheng Bioengineering institute (Nanjing, China). All animal procedures and experiments were conducted in accordance with Chinese Administration Rule of Laboratory Animal and approved by the Ethics Committees and the Research Committees of Xinxiang Medical University (81373135-2014, 15 March 2014).

4.2. Cell Culture and Adipocytes Differentiation

Human hepatic L02 and HepG2 cells were cultured in RPMI-1640 medium containing 10% fetal bovine serum, 100 IU/mL penicillin, and 100 µg/mL streptomycin. 3T3-L1 preadipocytes were cultured in Dulbecco's Modified Eagle Medium (DMEM) containing 15% fetal bovine serum, 100 IU/mL penicillin, and 100 µg/mL streptomycin. All cells were cultured in an incubator with a humidified atmosphere of 5% CO2, 95% air at 37 °C. The 3T3-L1 differentiation was performed as previously described [30]. Briefly, 3 \times 10⁴ 3T3-L1 cells were plated in 60 mm dishes, grown to confluence and fed every two days during growth. At two days of post-confluence (day 0), the medium supplemented with methyllisobutylxanthine (0.5 mM), dexamethasone (1 µM), and insulin (10 g/mL) was added to dishes for 2 days induction (day 2). In addition, then the medium was replaced with a new medium supplemented only with insulin (10 g/mL) for another 2 days. At day 4, the medium was changed to complete DMEM medium for maintenance. At day 10, differentiated 3T3-L1 adipocytes were treated with Ber, Res and their combination for 3 days. Berberine and Resveratrol for cell assay was purchased from Selleck Chemicals company (Shanghai, China).

4.3. Oil Red O Staining Assay

The intracellular lipid accumulation of differentiated cells was detected by oil red O staining. Cells were washed with phosphate buffered saline (PBS) twice, fixed in 10% formalin for 15 min at room temperature. After washed with PBS, cells were stained with oil red O for 20 min, then rinsed with PBS twice and 60% isopropanol once. Cell nuclei were stained with hematoxylin. After dyed, cells were washed and overlaid with PBS, photographed using a phase contrast microscopy. For semi-quantitative analysis of oil red O staining, the dye in intracellular lipid droplets was extracted by incubating cells stained with 60% isopropanol containing 4% Sodium dodecyl sulfate (SDS) at room temperature for 20 min. The optical density of oil red O were monitored spectrophotometrically at 510 nm wavelength.

4.4. Western Blot Assay

After treatment, cells pellets collected immediately were lysed in RIPA cell lysis buffer (10 mM Tris-HCl pH 7.4, 1% Triton X-100, 1% sodium deoxycholate, 0.1% SDS, 160 mM NaCl, 5 mM EDTA, 50 mM NaF, 10% Glycerol, 1 mM Na₃VO₄, and 5 mM sodium pyrophoshate). The total proteins extracted from cells were quantified by bicinchoninic acid (BCA) protein assay. Equal amounts of protein samples were subjected to 8–10% sodium dodecyl sulfate–polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto Polyvinylidene fluoride (PVDF) membrane. The membranes were incubated with various antibodies at 1:1000 dilutions. Then they were incubated with horseradish peroxidase-conjugated secondary antibodies at 1:2000 dilutions. Visualization was detected with ECL Plus Western Blotting Detection System according to the manufacturer's recommendation. Goat antibodies directed against LDLR (C-7, sc-18823), HRP-labeled anti-goat IgG (sc-2354) and anti-β-actin antibody (sc-8432) were from Santa Cruz Biotechnology (Heidelberg, German). SIRT1 Antibody (#2493) were from Cell Signal Technology (Shanghai, China).

4.5. Flow Cytometry Assay

L02 cells were seeded into six-well plates at a density of 4×10^5 /well for 24 h culture. For the intracellular fluorescence of Ber assay, a range of Ber concentrations (12.5–100 µmol/L) was added to dishes of L02 cells for 2 h, and then cells were measured with a FACScan flow cytometer (Shanghai, China) using the FL1 filter set after trypsinization, washing and resuspension in PBS with 5% fetal bovine serum. For combination studies, L02 cells were pretreated with 10 or 25 µmol/L Res for 2 h, and then co-treated with 50 µmol/L Ber for 2 h. The intracellular fluorescence of Ber was detected by flow cytometry.

4.6. Statistical Analysis.

Data analysis was performed using GraphPad Prism 5 (GraphPad software Inc., San Diego, CA, USA) and expressed as mean \pm SD. The significance of differences between two groups was determined with student's-t-test. For multiple comparison, One-way analysis of variance (ANOVA) was used with Turkey's test. A *p*-value less than 0.05 was considered to be significant.

Supplementary Materials: Supplementary materials can be found at http://www.mdpi.com/1422-0067/19/12/3903/s1.

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Abbreviations

Ber Berberine
Res Resveratrol
SIRT1 Sirtuin 1

TC Total cholesterol

LDLR Low Density Lipoprotein Receptor

HFD High fat diet TG triglyceride

LDL-c Low Density Lipoprotein cholesterol

MFI mean fluorescence intensity

FBS Fetal Bovine Serum

DMSO Dimethyl Sulphoxide

PBS phosphate buffered saline

SDS-PAGE Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis

PVDF Polyvinylidene fluoride

References

1. Kopelman, P.G. Obesity as a medical problem. Nature 2000, 404, 635–643. [CrossRef] [PubMed]

- 2. Eckel, R.H.; Alberti, K.G.; Grundy, S.M.; Zimmet, P.Z. The metabolic syndrome. *Lancet* **2010**, *375*, 181–183. [CrossRef]
- 3. McKeigue, P.M.; Shah, B.; Marmot, M.G. Relation of central obesity and insulin resistance with high diabetes prevalence and cardiovascular risk in South Asians. *Lancet* **1991**, *337*, 382–386. [CrossRef]
- 4. Ye, J. Mechanisms of insulin resistance in obesity. Front. Med. 2013, 7, 14–24. [CrossRef] [PubMed]
- 5. Yazıcı, D.; Sezer, H. Insulin Resistance, Obesity and Lipotoxicity. *Adv. Exp. Med. Biol.* **2017**, *960*, 277–304. [CrossRef] [PubMed]
- 6. Kong, W.; Wei, J.; Abidi, P.; Lin, M.; Inaba, S.; Li, C.; Wang, Y.; Wang, Z.; Si, S.; Pan, H.; et al. Berberine is a novel cholesterol-lowering drug working through a unique mechanism distinct from statins. *Nat. Med.* **2004**, 10, 1344–1351. [CrossRef]
- 7. Lee, Y.S.; Kim, W.S.; Kim, K.H.; Yoon, M.J.; Cho, H.J.; Shen, Y.; Ye, J.M.; Lee, C.H.; Oh, W.K.; Kim, C.T.; et al. Berberine, a natural plant product, activates AMP-activated protein kinase with beneficial metabolic effects in diabetic and insulin-resistant states. *Diabetes* **2006**, *55*, 2256–2264. [CrossRef]
- 8. Huang, C.; Zhang, Y.; Gong, Z.; Sheng, X.; Li, Z.; Zhang, W.; Qin, Y. Berberine inhibits 3T3-L1 adipocyte differentiation through the PPAR-gamma pathway. *Biochem. Biophys. Res. Commun.* **2006**, 348, 571–578. [CrossRef]
- 9. Yin, J.; Xing, H.; Ye, J. Efficacy of berberine in patients with type 2 diabetes mellitus. *Metabolism* **2008**, *57*, 712–717. [CrossRef]
- 10. Zhu, X.; Guo, X.; Mao, G.; Gao, Z.; Wang, H.; He, Q.; Li, D. Hepatoprotection of berberine against hydrogen peroxide-induced apoptosis by upregulation of Sirtuin 1. *Phytother. Res.* **2013**, 27, 417–421. [CrossRef]
- 11. Zhu, X.; Yue, H.; Guo, X.; Yang, J.; Liu, J.; Liu, J.; Wang, R.; Zhu, W. The Preconditioning of Berberine Suppresses Hydrogen Peroxide-Induced Premature Senescence via Regulation of Sirtuin 1. *Oxid. Med. Cell Longev.* 2017, 2017, 2391820. [CrossRef] [PubMed]
- 12. Bordone, L.; Cohen, D.; Robinson, A.; Motta, M.C.; van Veen, E.; Czopik, A.; Steele, A.D.; Crowe, H.; Marmor, S.; Luo, J.; et al. SIRT1 transgenic mice show phenotypes resembling calorie restriction. *Aging Cell* **2007**, *6*, 759–767. [CrossRef] [PubMed]
- 13. Lagouge, M.; Argmann, C.; Gerhart-Hines, Z.; Meziane, H.; Lerin, C.; Daussin, F.; Messadeq, N.; Milne, J.; Lambert, P.; Elliott, P.; et al. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1α. *Cell* **2006**, 27, 1109–1122. [CrossRef] [PubMed]
- 14. Baur, J.A.; Pearson, K.J.; Price, N.L.; Jamieson, H.A.; Lerin, C.; Kalra, A.; Prabhu, V.V.; Allard, J.S.; Lopez-Lluch, G.; Lewis, K.; et al. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 2006, 444, 337–342. [CrossRef] [PubMed]
- 15. Talwalkar, S.S.; Vaidya, A.B.; Godse, C.; Vaidya, A.; Vaidya, R. Plasmodium DNA Fluoresces with berberine: A novel approach for diagnosis of malarial parasites. *Am. J. Clin. Pathol.* **2005**, 124, 408–412. [CrossRef] [PubMed]
- 16. Serafim, T.L.; Oliveira, P.J.; Sardao, V.A.; Perkins, E.; Parke, D.; Holy, J. Different concentrations of berberine result in distinct cellular localization patterns and cell cycle effects in a melanoma cell line. *Cancer Chemother. Pharmacol.* **2008**, *61*, 1007–1018. [CrossRef] [PubMed]
- 17. Balandrin, M.F.; Klocke, J.A.; Wurtele, E.S.; Bollinger, W.H. Natural plant chemicals: Sources of industrial and medicinal materials. *Science* **1985**, 228, 1154–1160. [CrossRef] [PubMed]

18. Kim, W.S.; Lee, Y.S.; Cha, S.H.; Jeong, H.W.; Choe, S.S.; Lee, M.R.; Oh, G.T.; Park, H.S.; Lee, K.U.; Lane, M.D.; et al. Berberine improves lipid dysregulation in obesity by controlling central and peripheral AMPK activity. *Am. J. Physiol. Endocrinol. Metab.* **2009**, 296, E812–E819. [CrossRef] [PubMed]

- 19. Gomes, A.P.; Duarte, F.V.; Nunes, P.; Hubbard, B.P.; Teodoro, J.S.; Varela, A.T.; Jones, J.G.; Sinclair, D.A.; Palmeira, C.M.; Rolo, A.P. Berberine protects against high fat diet-induced dysfunction in muscle mitochondria by inducing SIRT1-dependent mitochondrial biogenesis. *Biochim. Biophys. Acta* 2012, 1822, 185–195. [CrossRef]
- 20. Penumathsa, S.V.; Thirunavukkarasu, M.; Koneru, S.; Juhasz, B.; Zhan, L.; Pant, R.; Menon, V.P.; Otani, H.; Maulik, N. Statin and resveratrol in combination induces cardioprotection against myocardial infarction in hypercholesterolemic rat. *J. Mol. Cell. Cardiol.* 2007, 42, 508–516. [CrossRef]
- 21. Guo, T.; Woo, S.L.; Guo, X.; Li, H.; Zheng, J.; Botchlett, R.; Liu, M.; Pei, Y.; Xu, H.; Cai, Y.; et al. Berberine Ameliorates Hepatic Steatosis and Suppresses Liver and Adipose Tissue Inflammation in Mice with Diet-induced Obesity. *Sci. Rep.* **2016**, *6*, 22612. [CrossRef] [PubMed]
- 22. Kong, W.J.; Zhang, H.; Song, D.Q.; Xue, R.; Zhao, W.; Wei, J.; Wang, Y.M.; Shan, N.; Zhou, Z.X.; Yang, P.; et al. Berberine reduce insulin-resistance through protein kinase C-dependent up-regulation of insulin receptor expression. *Metabolism* **2009**, *58*, 109–119. [CrossRef] [PubMed]
- 23. Shitan, N.; Tanaka, M.; Terai, K.; Ueda, K.; Yazaki, K. Human MDR1 and MRP1 recognize berberine as their transport substrate. *Biosci. Biotechnol. Biochem.* **2007**, *71*, 242–245. [CrossRef] [PubMed]
- 24. Quan, F.; Pan, C.; Ma, Q.; Zhang, S.; Yan, L. Reversal effect of resveratrol on multidrug resistance in KBv200 cell line. *Biomed. Pharmacother.* **2008**, *62*, *622*–*629*. [CrossRef] [PubMed]
- 25. Nabekura, T.; Kamiyama, S.; Kitagawa, S. Effects of chemopreventive citrus phytochemicals on human P-glycoprotein and multidrug resistance protein 1. *Eur. J. Pharmacol.* **2008**, *600*, 45–49. [CrossRef] [PubMed]
- Yang, I.; Kim, E.; Kang, J.; Han, H.; Sul, S.; Park, S.B.; Kim, S.K. Photochemical generation of a new, highly fluorescent compound from non-fluorescent resveratrol. *Chem. Commun.* 2012, 48, 3839–3841.
 [CrossRef] [PubMed]
- 27. Ellsworth, J.L.; Chandrasekaran, C.; Cooper, A.D. Evidence for sterol-independent regulation of low-density lipoprotein receptor activity in Hep-G2 cells. *Biochem. J.* **1991**, 279, 175–187. [CrossRef] [PubMed]
- 28. Purushotham, A.; Schug, T.T.; Xu, Q.; Surapureddi, S.; Guo, X.; Li, X. Hepatocyte-specific deletion of SIRT1 alters fatty acid metabolism and results in hepatic steatosis and inflammation. *Cell. Metab.* **2009**, *9*, 327–338. [CrossRef]
- 29. Miranda, M.X.; van Tits, L.J.; Lohmann, C.; Arsiwala, T.; Winnik, S.; Tailleux, A.; Stein, S.; Gomes, A.P.; Suri, V.; Ellis, J.L.; et al. The Sirt1 activator SRT3025 provides atheroprotection in Apoe-/- mice by reducing hepatic Pcsk9 secretion and enhancing Ldlr expression. *Eur. Heart J.* 2015, *36*, 51–59. [CrossRef]
- 30. Student, A.K.; Hsu, R.Y.; Lane, M.D. Induction of fatty acid synthesis in differentiating 3T3-L1 preadipocytes. *J. Biol. Chem.* **1980**, 255, 4745–4750.



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