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Evidence for a Causal Role of the *SH2B3*- β_2M Axis in Blood Pressure Regulation: the Framingham Heart Study

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Abstract

Genetic variants at *SH2B3* are associated with blood pressure and circulating beta-2 microglobulin (β_2M), a well-characterized kidney filtration biomarker. We hypothesize that circulating β_2M is an independent risk predictor of hypertension and may causally contribute to its development.

The study sample consisted of 7065 Framingham Heart Study participants with measurements of plasma β_2M . Generalized estimating equations were used to test the association of β_2M with prevalent and new-onset hypertension. There were 2145 (30%) cases of prevalent hypertension at baseline and 886 (21%) cases of incident hypertension during six years of follow-up. A 1-standard deviation increase in baseline plasma β_2M was associated with a greater risk of prevalent (odds ratio [OR] 1.14, 95% confidence interval [CI] 1.05–1.24) and new-onset (OR 1.18, 95% CI 1.07–1.32) hypertension. Individuals within the top β_2M quartile had a greater risk than the bottom quartile for prevalent (OR 1.29, 95% CI .05–1.57) and new-onset (OR 1.59, 95% CI 1.20–2.11) hypertension. These associations remained essentially unchanged in analyses restricted to participants free of albuminuria and chronic kidney disease (CKD). Mendelian randomization demonstrated that lower *SH2B3* expression is causal for increased circulating β_2M levels, and in a hypertensive mouse model, knockout of *Sh2b3* increased β_2M gene expression.

In a community-based study of healthy individuals, higher plasma β_2M levels are associated with increased risk of prevalent and incident hypertension independent of CKD status. Overlapping

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Disclosures

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genetic signals for hypertension and $\beta_2\text{M}$, in conjunction with mouse knockout experiments, suggest that the *SH2B3*- $\beta_2\text{M}$ axis plays a causal role in hypertension.

Summary

Based on overlapping genetic associations, we explored the association between $\beta_2\text{M}$ and hypertension, and demonstrate $\beta_2\text{M}$ to be associated with hypertension both cross-sectionally and longitudinally in Framingham Heart Study participants with and without impaired kidney function. Furthermore, using Mendelian randomization and knockout of *Sh2b3* in a mouse model of hypertension, we provide evidence that *trans*-regulation of $\beta_2\text{M}$ by *SH2B3* may play a causal role in blood pressure regulation.

Keywords

Hypertension; $\beta_2\text{M}$; biomarkers; gene expression; genome-wide association study; epidemiology

Introduction

Beta-2-microglobulin ($\beta_2\text{M}$), an essential component of the class I major histocompatibility complex (MHC),^{1,2} is freely filtered in the glomerulus and reabsorbed in the proximal tubule, with circulating levels tightly maintained in healthy individuals.³ A prior genome-wide association study (GWAS) of circulating $\beta_2\text{M}$ identified genetic variants at the *MHC* and *SH2B3* genetic loci⁴, with one of the top single nucleotide polymorphisms (SNP) for circulating $\beta_2\text{M}$ at the *SH2B3* locus, rs3184504, also being a genome-wide significant SNP for blood pressure (BP)⁵. Therefore, given the concurrence of genetic variants for BP and circulating $\beta_2\text{M}$, we hypothesized that $\beta_2\text{M}$ may be a causal factor for hypertension.

Hypertension is preventable through lifestyle interventions and drug treatment,^{6,7} but it still remains the most widespread non-communicable disease worldwide.^{8,9} Previous population studies of $\beta_2\text{M}$ and hypertension are either limited to cross-sectional analyses¹⁰ or did not consider kidney disease status¹¹. To address these limitations, we sought to determine whether plasma $\beta_2\text{M}$ was associated with hypertension cross-sectionally and longitudinally and to assess its potential clinical utility as a prognostic indicator of hypertension risk. We also performed secondary analyses excluding individuals with albuminuria and low estimated glomerular filtration rate (eGFR) at baseline because elevated urinary $\beta_2\text{M}$ levels have been reported to be associated with impaired kidney function¹⁰. Lastly, we provide results of a functional study in *Sh2b3* knockout mice to explore a causal role of the *SH2B3*- $\beta_2\text{M}$ axis in hypertension.

Materials and Methods

All source data and materials have been made publicly available at dbGaP (accession number phs000363.v17.p11).

Study population:

The cross-sectional study sample consisted of Framingham Heart Study (FHS) Offspring and Third Generation cohort participants.^{12,13,14} A total of 7242 participants had measurements of plasma β_2M ¹⁵ at the baseline examination (Offspring cohort examination 7 [1999–2003, n=3267] and Third Generation cohort examination 1 [2002–2005, n=3975]). Participants with the following conditions were excluded: myocardial infarction or heart failure prior to baseline; missing information regarding use of anti-hypertensive treatment, lipid lowering treatment, and diabetes medication use; and missing information on cigarette smoking, body-mass index, fasting glucose, total- or high-density lipoprotein cholesterol, triglycerides, or serum creatinine – leaving a final cross-sectional sample size of 7065.

Participants who attended the follow-up examination (Offspring cohort examination 8 [2005–2008] and Third Generation cohort examination 2 [2008–2011]) approximately six years after baseline (median 6.0, 25th and 75th percentiles 6.0, 7.0 years after baseline) and were normotensive at baseline, made up the longitudinal study sample (n=4189). Secondary longitudinal analyses were performed after excluding those with albuminuria (defined as urinary albumin-creatinine ratio [UACR] ≥ 30 mg/g) or low eGFR (eGFR <60 mL/min/1.73 m²) at baseline.

Quantification of plasma β_2M protein:

Plasma β_2M concentration was measured at baseline as part of the Systems Approach to Biomarker Research in Cardiovascular Disease (SABRe CVD) Initiative¹⁶. A modified sandwich enzyme-linked immunosorbent assay, multiplexed on a Luminex xMAP platform (Sigma-Aldrich)¹⁷ was used, as described previously¹⁸. All targets were first developed as single assays, after which compatible targets, with appropriate plasma analyte concentration ranges, were combined to form multiplex panels to avoid cross-reactivity. For the β_2M assay, the detection antibody was product #200–406-141s (Rockland Antibodies, Limerick, PA), the capture antibody was GTX20759 (Genetex, Irvine, CA), and the reference protein was #126–11 (Lee BioSolutions, Maryland Heights, MO). The mean intra-assay coefficient of variation for β_2M was 3.7%¹⁷.

Data collection for covariates:

Each FHS examination consisted of an in-person interview, physician-administered physical examination, anthropometric measurements, BP measurements, and blood/urine sample collection. BMI was calculated as weight (kg) divided by the square of height (m²). Current cigarette smoking was defined as self-reported smoking of at least one cigarette per day, on average, during the year preceding the examination. Information on antihypertensive and lipid lowering medication-use was collected via a self-administered questionnaire and verified using medications that participants brought to the research clinic visit. Diabetes was defined as fasting blood glucose greater than or equal to 126 mg/dL, or the current use of insulin or hypoglycemic medication. Participant spot urine samples were used to measure urinary albumin and creatinine levels, from which the UACR was calculated. Albuminuria was defined as a UACR equal to or greater than 30 mg/g¹⁹. Serum creatinine levels, measured using the Roche Hitachi 911 reader, were quantified using the modified Jaffe method²⁰ with a modified picric acid buffer solution to minimize interference and weighted

by population-specific distributions of age and sex²¹. eGFR was calculated using the CKD-epi equation.^{22, 23} CKD was defined as eGFR < 60mL/min/1.73m².²⁴

BP measurement:

BP was measured at the baseline and follow-up examinations. Systolic and diastolic BP were each measured twice by a physician in the left arm after five minutes of rest in the seated position, and mean values were used. Hypertension was defined as systolic BP ≥ 140 mm Hg, diastolic BP ≥ 90 mm Hg, or current antihypertensive medication use²⁵.

Statistical Analyses:

Generalized estimating equations (GEE) were used to account for familial correlations in testing the association of baseline β_2M concentration with prevalent and new-onset hypertension. In the GEE models, nuclear family members were considered as clusters of measurements. The logit link function was applied for the dichotomous response variable, hypertension. All analyses for prevalent hypertension were adjusted for the following baseline covariates: age, sex, study cohort, BMI, diabetes, cigarette smoking, levels of total and HDL-cholesterol, statin-use, and eGFR. Analyses for new-onset hypertension were adjusted for the same baseline covariates, along with baseline systolic and diastolic BP. Secondary analyses were performed after excluding individuals with prevalent CKD or albuminuria.

Reclassification analyses were conducted by comparing the predictive abilities of regression models with and without plasma β_2M for new-onset hypertension, and used the integrated discrimination improvement (IDI) and net reclassification improvement (NRI) metrics.^{26,27} Bootstrapping with 1000 iterations was used for probability estimates,²⁶ and two-sided *P* values of less than 0.05 were considered statistically significant. All analyses were conducted using SAS software version 9.4.

Mendelian Randomization (MR):

A two-sample inverse-variance weighted MR approach was used to test whether *SH2B3* expression was causal for plasma β_2M levels. Pruned *cis*-eQTL variants (LD $r^2 < 0.1$) for *SH2B3* expression²⁸ were used as instrumental variables (IVs) for *SH2B3* expression with *SH2B3* expression as the exposure and circulating β_2M levels as the outcome. *SH2B3* expression was measured as previously described²⁸. Briefly, fasting whole blood samples were collected from FHS participants in PAXgeneTM tubes (PreAnalytiX, Hombrechtikon, Switzerland) and stored at -80 °C. Total RNA was isolated from frozen PAXgene tubes by Asuragen, Inc, and used to generate cDNA. The cDNA was fragmented and labeled using the FL-OvationTM cDNA Biotin Module, after which the resultant cDNA fragments were allowed to hybridize overnight to the Affymetrix human exon 1.0 ST microarray platform according to the manufacturer's protocol (Affymetrix, Santa Clara, CA, USA). The GeneChipTM 7G GCS3000 scanner (Thermo Fisher Scientific, Waltham, MA) was used to laser scan the microarray following washing and staining using the GeneChipTM Fluidics Station 450. Intensity values for each gene chip were compiled and normalized using the robust multi-chip average method from Affymetrix Power Tools Software version 1.12.0 (Affymetrix) before and after quality control.

All MR analyses were conducted using MRbase (<http://www.mrbase.org/>). Causal effect estimates of *SH2B3* expression on plasma β_2M levels are reported as effect per risk allele on inverse rank-normalized protein level. Because we lacked *cis*-pQTL variants for circulating β_2M levels, we were unable to conduct MR analyses to determine if plasma β_2M is causal for hypertension.

Mouse Models:

Wild-type (WT) C57B1/6J mice were purchased from Jackson Laboratories (Bar Harbor, ME). *Sh2b3*-deficient C57BL/6J mice were generated by deleting exons 3–8 of *Sh2b3*, as previously described^{29,30}. Osmotic mini-pumps (model 2002, Alzet, DURECT Corporation, City, State) were implanted subcutaneously in mice approximately 8–12 weeks of age for angiotensin II (490 ng/kg/min) or vehicle (sodium chloride/acetic acid solution) infusion for 14 days. RNA was extracted from whole blood of WT and *Sh2b3*^{-/-} mice using a RiboPure™ RNA purification Kit (Cat# AM1928, Life Technologies, Carlsbad, CA), and cDNA was constructed using VANTAGE (Vanderbilt Technologies for Advanced Genomics, Vanderbilt University Medical Center, Nashville, TN). RNA sequencing reads first underwent quality control (QC) using the FASTX-Toolkit package (http://hannonlab.cshl.edu/fastx_toolkit/), after which they were mapped to the mouse reference genome (UCSC mm10) using Tophat v2.0³¹. mRNA levels were estimated and normalized using Cufflinks v2.2 and are reported in expected number of fragments per kilobase of transcript sequence per million base pairs sequenced (FPKM)³². Cuffdiff was used to identify differential expression of β_2M between WT and *SH2B3*^{-/-} mice (<http://cole-trapnell-lab.github.io/cufflinks/cuffdiff/>).

Results

Baseline characteristics:

Summary demographic and clinical characteristics of the study sample (mean age 49 years, 54% women) are shown in Table 1 according to β_2M quartiles and in Table S1 for the overall study sample.

Cross-sectional analyses:

Baseline β_2M concentration was positively associated with prevalent hypertension after adjusting for clinical covariates with an odds ratio (OR) of 1.14 (95% confidence interval [CI] 1.05–1.24, $P=0.003$) per standard deviation increase in log-transformed β_2M , and an OR of 1.29 (95% CI 1.05–1.57, $P=0.01$) for the top versus bottom β_2M quartile (Table 2).

Longitudinal analyses:

A total of 886 (21%) participants who were normotensive at baseline developed new-onset hypertension after a mean follow-up of six years. Results of the GEE model revealed a significant association between baseline β_2M and risk of new-onset hypertension with an OR of 1.18 (95% CI 1.07–1.32, $P=0.001$) per standard deviation increase in log-transformed β_2M (Table 3). Results of the quartile analysis showed that the top β_2M quartile had a 1.59-fold risk (95% CI 1.20, 2.11, $P=0.001$) of incident hypertension versus those in the bottom

quartile (Table 3). Excluding individuals with albuminuria and low eGFR at baseline did not materially change the results (Table S2).

Reclassification Analyses:

Reclassification metrics revealed that β_2M improved the prediction of new-onset hypertension over the clinical model, which consisted of the following baseline covariates: age, sex, study cohort, BMI, diabetes, cigarette smoking, levels of total and HDL-cholesterol, statin-use, eGFR, and systolic and diastolic BP. Addition of β_2M to the clinical model improved the discrimination for new onset hypertension in the overall sample (integrated discrimination improvement [IDI] 0.0021, $P=0.03$) and in those free of baseline albuminuria and CKD (IDI=0.0036, $P=0.008$; Table 4). The continuous net reclassification index (NRI) was statistically significant in participants free of baseline albuminuria and low eGFR (NRI 0.030, $P=0.02$) but not in the general sample ($P>0.05$; Table 4).

Mendelian randomization:

As the top genetic variant (rs3184504) associated with plasma β_2M levels is also associated with *SH2B3* expression³³, we postulated that the *trans*-effects (>1 megabase upstream or downstream of the transcription start site) of the rs3184504 polymorphism on β_2M levels may be mediated through its *cis*-effects (within 1 megabase) on *SH2B3* expression. To test this hypothesis, we conducted two-sample inverse-variance weighted MR using all independent *cis* variants (LD $r^2<0.1$) for *SH2B3* expression as instrumental variables, *SH2B3* expression as the exposure, and circulating β_2M levels as the outcome. Decreased *SH2B3* expression was found to be causal for higher β_2M levels ($\beta=-16.8$, $P=8.7E-11$; Fig. 1).

Sh2b3^{-/-} mouse model:

To recapitulate the effects of *cis* genetic variation within *SH2B3*, we tested the effects of *SH2B3* deficiency on β_2M expression at baseline and in response to angiotensin II-induced hypertension. The rs3184504 polymorphism is associated with reduced *SH2B3* function in humans,³⁴ and we previously reported that *Sh2b3*^{-/-} mice had a greater hypertensive response and more severe renal/vascular dysfunction in response to angiotensin II infusion³⁵. Analysis of RNA sequencing performed on whole blood from wild type and *Sh2b3*^{-/-} mice revealed a significant induction of β_2M expression in response to angiotensin II administration only in *Sh2b3*^{-/-} mice ($P=0.037$; Fig. 2) suggesting that *SH2B3* normally constrains β_2M expression in response to hypertensive stimuli and that enhanced β_2M expression may be causally related to the exaggerated hypertensive response and end-organ damage seen in *Sh2b3*^{-/-} animals.

Discussion

Motivated by GWAS variants at the *SH2B3* locus that were reported to be associated both with BP⁵ and circulating β_2M levels⁴, we first explored the association of circulating β_2M with prevalent and incident hypertension in a large community-based cohort study. We found that circulating β_2M was associated with clinically relevant increases in prevalent and new-onset hypertension, replicating association results previously reported by investigators from

the Atherosclerosis Risk in Communities study¹¹. Secondary longitudinal association analyses between β_2M and hypertension conducted in FHS participants free of baseline CKD and albuminuria demonstrated similar effect sizes and consistent directions of effect compared to the overall study sample, thus suggesting β_2M is more than a marker of renal dysfunction. Our MR analyses and mechanistic study in mice suggest that the *SH2B3*- β_2M axis may play a causal role in BP regulation. Reclassification analyses, however, reveal that whereas the addition of β_2M to a clinical model improved hypertension prediction, the magnitude of effect suggests that circulating β_2M level is not likely to be a clinically useful biomarker of hypertension risk.

Hypertension is a complex disorder with a prominent renal component characterized by impaired renal sodium excretion, vascular remodeling, inflammation, and fibrosis.^{36,37} Genes within the *HLA* and *SH2B3* regions have been implicated in inflammation,³⁸ and recent *Sh2b3*-knockout studies in mice have linked mutations or deletions in *SH2B3* with kidney inflammation³⁹, hypertension^{30,40,41}, and atherosclerotic CVD^{34,42,43, 44}. Therefore, we sought to determine if there was a mechanistic link between *SH2B3*, β_2M , and hypertension. To this end, we first conducted MR and found that lower *SH2B3* expression increases circulating β_2M levels (Fig. 1). Due to a lack of *cis* genetic variants for circulating β_2M , we were unable to conduct MR to determine whether circulating β_2M is causal for hypertension. We previously reported that *Sh2b3*^{-/-} mice develop an exaggerated BP response and more severe end-organ damage (renal and vascular inflammation and dysfunction) in response to angiotensin II³⁰. We extend these findings in the present study by reporting that β_2M expression is significantly increased in response to angiotensin II infusion in *Sh2b3*^{-/-} but not wild type mice (Fig. 2). We thus provide evidence for a causal role of the *SH2B3*- β_2M axis in hypertension.

To our knowledge, ours is the first study to demonstrate that *SH2B3* might regulate hypertension not only via production of its coded protein, the lymphocyte adaptor protein (LNK), but also through effects on β_2M expression. Deficiency of LNK increases interferon gamma producing CD8⁺ T-cells in the spleen and kidneys of hypertensive mice, and both interferon gamma and CD8⁺ T cells are mediators of hypertension^{30,45}. β_2M is essential for MHC-I-mediated CD8⁺ T-cell activation⁴⁶. Our finding that loss of LNK increases β_2M expression under hypertensive conditions is consistent with our MR results. We hypothesize that loss of *SH2B3* and a resultant increase in β_2M levels likely leads to increased CD8⁺ T cell activation and interferon gamma production, which promote hypertension and hypertensive end-organ damage (Fig. 1).

Our study has several strengths. First, the sample size was large and adequate to assess the relations of β_2M to prevalent and incident hypertension. Second, BP and eGFR were meticulously ascertained, minimizing recall bias. Third, association results for prevalent and incident hypertension largely replicate those of an independent external study¹¹.

We acknowledge several study limitations. First, circulating β_2M was only measured at the baseline examination, preventing analyses of temporal variation in plasma β_2M . In addition, this was a European ancestry study sample; the extent to which our results are indicative of what would be observed in other racial/ethnic groups is unknown. Third, a substantial

proportion of participants did not have measurement of urinary albumin, resulting in a smaller sample size for secondary longitudinal analyses that excluded those with CKD or albuminuria. Fourth, we applied an inclusive definition for hypertension that included elevated BP or antihypertensive medication use. Finally, we were unable to conduct MR of plasma β_2M in relation to hypertension due to a lack of *cis* genetic variants for circulating β_2M .

In summary, we have identified β_2M to be cross-sectionally and longitudinally associated with hypertension risk independent of albuminuria and kidney disease and provide functional evidence of a causal role of the *SH2B3*- β_2M axis in hypertension. Future studies are needed to further understand the mechanistic role of β_2M in hypertension and determine if there is therapeutic efficacy in targeting the *SH2B3*- β_2M axis for hypertension treatment or prevention.

Perspectives

β_2M is cross-sectionally and longitudinally associated with hypertension in Framingham Heart Study participants with and without kidney dysfunction. Mendelian randomization analysis and studies using a murine model of hypertension provide evidence for a causal role of the *SH2B3*- β_2M axis in blood pressure regulation. Future studies are needed to determine whether β_2M is a clinically-useful prognostic indicator of hypertension and whether there is therapeutic utility in targeting the *SH2B3*- β_2M axis for hypertension prevention and treatment.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

1. Ohta Y, Shiina T, Lohr RL, Hosomichi K, Pollin TI, Heist EJ, Suzuki S, Inoko H & Flajnik MF Primordial linkage of beta2-microglobulin to the MHC. *J Immunol* 186, 3563–3571, doi:10.4049/jimmunol.1003933 (2011). [PubMed: 21321107]
2. Li L, Dong M & Wang XG The Implication and Significance of Beta 2 Microglobulin: A Conservative Multifunctional Regulator. *Chin Med J (Engl)* 129, 448–455, doi: 10.4103/0366-6999.176084 (2016). [PubMed: 26879019]
3. Bonventre JV, Vaidya VS, Schmourder R, Feig P & Dieterle F Next-generation biomarkers for detecting kidney toxicity. *Nat Biotechnol* 28, 436–440, doi:10.1038/nbt0510-436 (2010). [PubMed: 20458311]

4. Tin A, Astor BC, Boerwinkle E, Hoogeveen RC, Coresh J & Kao WH Genome-wide association study identified the human leukocyte antigen region as a novel locus for plasma beta-2 microglobulin. *Hum Genet* 132, 619–627, doi:10.1007/s00439-013-1274-7 (2013). [PubMed: 23417110]
5. Levy D, Ehret GB, Rice K, Verwoert GC, Launer LJ, Dehghan A, Glazer NL, Morrison AC, Johnson AD, Aspelund T, Aulchenko Y, Lumley T, Kottgen A, Vasan RS, Rivadeneira F, Eiriksdottir G, Guo X, Arking DE, Mitchell GF, Mattace-Raso FU, Smith AV, Taylor K, Scharpf RB, Hwang SJ, Sijbrands EJ, Bis J, Harris TB, Ganesh SK, O'Donnell CJ, Hofman A, Rotter JJ, Coresh J, Benjamin EJ, Uitterlinden AG, Heiss G, Fox CS, Witteman JC, Boerwinkle E, Wang TJ, Gudnason V, Larson MG, Chakravarti A, Psaty BM & van Duijn CM Genome-wide association study of blood pressure and hypertension. *Nat Genet* 41, 677–687, doi:10.1038/ng.384 (2009). [PubMed: 19430479]
6. Salomon JA, Haagsma JA, Davis A, de Noordhout CM, Polinder S, Havelaar AH, Cassini A, Devleeschauwer B, Kretzschmar M, Speybroeck N, Murray CJ & Vos T Disability weights for the Global Burden of Disease 2013 study. *Lancet Glob Health* 3, e712–723, doi:10.1016/S2214-109X(15)00069-8 (2015). [PubMed: 26475018]
7. Campbell NR, Lackland DT, Niebylski ML, World Hypertension League C & International Society of Hypertension Executive, C. High blood pressure: why prevention and control are urgent and important: a 2014 fact sheet from the World Hypertension League and the International Society of Hypertension. *J Clin Hypertens (Greenwich)* 16, 551–553, doi:10.1111/jch.12372 (2014). [PubMed: 25040331]
8. Whitworth JA & World Health Organization, I. S. o. H. W. G. 2003 World Health Organization (WHO)/International Society of Hypertension (ISH) statement on management of hypertension. *Journal of hypertension* 21, 1983–1992, doi:10.1097/01.hjh.0000084751.37215.d2 (2003). [PubMed: 14597836]
9. Mortality GBD & Causes of Death C Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* 385, 117–171, doi:10.1016/S0140-6736(14)61682-2 (2015). [PubMed: 25530442]
10. Mashima Y, Konta T, Kudo K, Takasaki S, Ichikawa K, Suzuki K, Shibata Y, Watanabe T, Kato T, Kawata S & Kubota I Increases in urinary albumin and beta2-microglobulin are independently associated with blood pressure in the Japanese general population: the Takahata Study. *Hypertens Res* 34, 831–835, doi:10.1038/hr.2011.42 (2011). [PubMed: 21525950]
11. Huang M, Matsushita K, Sang Y, Ballew SH, Astor BC & Coresh J Association of kidney function and albuminuria with prevalent and incident hypertension: the Atherosclerosis Risk in Communities (ARIC) study. *American journal of kidney diseases : the official journal of the National Kidney Foundation* 65, 58–66, doi:10.1053/j.ajkd.2014.06.025 (2015). [PubMed: 25151408]
12. Kannel WB, Dawber TR, KAGAN A, REVOTSKIE N & STOKES J, III. Factors of risk in the development of coronary heart disease—six year follow-up experience. The Framingham Study. *Ann. Intern. Med* 55, 33–50 (1961). [PubMed: 13751193]
13. Feinleib M, Kannel WB, Garrison RJ, McNamara PM & Castelli WP The Framingham Offspring Study. Design and preliminary data. *Prev. Med* 4, 518–525 (1975). [PubMed: 1208363]
14. Splansky GL, Corey D, Yang Q, Atwood LD, Cupples LA, Benjamin EJ, D'Agostino RB, Sr., Fox CS, Larson MG, Murabito JM, O'Donnell CJ, Vasan RS, Wolf PA & Levy D The Third Generation Cohort of the National Heart, Lung, and Blood Institute's Framingham Heart Study: design, recruitment, and initial examination. *Am. J. Epidemiol* 165, 1328–1335, doi:kwm021 [pii]; 10.1093/aje/kwm021 [doi] (2007). [PubMed: 17372189]
15. Yao Chen CG, Song Ci, Mendelson Michael, Tianxiao Huan, Annika Laser, Wu Hongsheng, Ho Jennifer E, Couchresne Paul, Lyass Asya, Larson Martin G, Gieger Christian, Graumann Johannes, Johnson Andrew D, Hwang Shih-Jen, Liu Chunyu, Suhre Karsten, Levy Daniel. Genome-wide Association Study Of Plasma Proteins Identifies Putatively Causal Genes, Proteins, And Pathways For Cardiovascular Disease bioRxiv, doi: 10.1101/136523 (2017).
16. Yin X, Subramanian S, Hwang SJ, O'Donnell CJ, Fox CS, Courchesne P, Muntendam P, Gordon N, Adourian A, Juhasz P, Larson MG & Levy D Protein biomarkers of new-onset cardiovascular

- disease: prospective study from the systems approach to biomarker research in cardiovascular disease initiative. *Arterioscler Thromb Vasc Biol* 34, 939–945, doi:10.1161/ATVBAHA.113.302918 (2014). [PubMed: 24526693]
17. Ho JE, Lyass A, Courchesne P, Chen G, Liu C, Yin X, Hwang SJ, Massaro JM, Larson MG & Levy D Protein Biomarkers of Cardiovascular Disease and Mortality in the Community. *J Am Heart Assoc* 7, doi:10.1161/JAHA.117.008108 (2018).
 18. dupont NC, Wang K, Wadhwa PD, Culhane JF & Nelson EL Validation and comparison of luminex multiplex cytokine analysis kits with ELISA: determinations of a panel of nine cytokines in clinical sample culture supernatants. *J Reprod Immunol* 66, 175–191, doi:10.1016/j.jri.2005.03.005 (2005). [PubMed: 16029895]
 19. McMahan GM, Hwang SJ, Tanner RM, Jacques PF, Selhub J, Muntner P & Fox CS The association between vitamin B12, albuminuria and reduced kidney function: an observational cohort study. *BMC Nephrol* 16, 7, doi:10.1186/1471-2369-16-7 (2015). [PubMed: 25644490]
 20. Moore JF & Sharer JD Methods for Quantitative Creatinine Determination. *Curr Protoc Hum Genet* 93, A 30 1–A 30 7, doi:10.1002/cphg.38 (2017).
 21. Fox CS, Larson MG, Leip EP, Culeton B, Wilson PW & Levy D Predictors of new-onset kidney disease in a community-based population. *JAMA* 291, 844–850, doi:10.1001/jama.291.7.844 (2004). [PubMed: 14970063]
 22. Levey AS, Inker LA & Coresh J GFR estimation: from physiology to public health. *Am J Kidney Dis* 63, 820–834, doi:10.1053/j.ajkd.2013.12.006 (2014). [PubMed: 24485147]
 23. Matsushita K, Selvin E, Bash LD, Astor BC & Coresh J Risk implications of the new CKD Epidemiology Collaboration (CKD-EPI) equation compared with the MDRD Study equation for estimated GFR: the Atherosclerosis Risk in Communities (ARIC) Study. *Am J Kidney Dis* 55, 648–659, doi:10.1053/j.ajkd.2009.12.016 (2010). [PubMed: 20189275]
 24. Stevens PE, Levin A & Kidney Disease: Improving Global Outcomes Chronic Kidney Disease Guideline Development Work Group, M. Evaluation and management of chronic kidney disease: synopsis of the kidney disease: improving global outcomes 2012 clinical practice guideline. *Ann Intern Med* 158, 825–830, doi:10.7326/0003-4819-158-11-201306040-00007 (2013). [PubMed: 23732715]
 25. Staessen JA, Wang J, Bianchi G & Birkenhager WH Essential hypertension. *Lancet* 361, 1629–1641, doi:10.1016/S0140-6736(03)13302-8 (2003). [PubMed: 12747893]
 26. Pencina MJ, D’Agostino RB, Sr. & Demler OV Novel metrics for evaluating improvement in discrimination: net reclassification and integrated discrimination improvement for normal variables and nested models. *Stat Med* 31, 101–113, doi:10.1002/sim.4348 (2012). [PubMed: 22147389]
 27. Shao F, Li J, Fine J, Wong WK & Pencina M Inference for reclassification statistics under nested and non-nested models for biomarker evaluation. *Biomarkers* 20, 240–252, doi:10.3109/1354750X.2015.1068854 (2015). [PubMed: 26301882]
 28. Joehanes R, Zhang X, Huan T, Yao C, Ying SX, Nguyen QT, Demirkale CY, Feolo ML, Sharopova NR, Sturcke A, Schaffer AA, Heard-Costa N, Chen H, Liu PC, Wang R, Woodhouse KA, Tanriverdi K, Freedman JE, Raghavachari N, Dupuis J, Johnson AD, O’Donnell CJ, Levy D & Munson PJ Integrated genome-wide analysis of expression quantitative trait loci aids interpretation of genomic association studies. *Genome Biol* 18, 16, doi:10.1186/s13059-016-1142-6 (2017). [PubMed: 28122634]
 29. Takaki S, Sauer K, Iritani BM, Chien S, Ebihara Y, Tsuji K, Takatsu K & Perlmutter RM Control of B cell production by the adaptor protein I κ N. Definition Of a conserved family of signal-modulating proteins. *Immunity* 13, 599–609 (2000). [PubMed: 11114373]
 30. Saleh MA, McMaster WG, Wu J, Norlander AE, Funt SA, Thabet SR, Kirabo A, Xiao L, Chen W, Itani HA, Michell D, Huan T, Zhang Y, Takaki S, Titze J, Levy D, Harrison DG & Madhur MS Lymphocyte adaptor protein LNK deficiency exacerbates hypertension and end-organ inflammation. *J Clin Invest* 125, 1189–1202, doi:10.1172/JCI76327 (2015). [PubMed: 25664851]
 31. Trapnell C, Pachter L & Salzberg SL TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* 25, 1105–1111, doi:10.1093/bioinformatics/btp120 (2009). [PubMed: 19289445]
 32. Trapnell C, Williams BA, Pertea G, Mortazavi A, Kwan G, van Baren MJ, Salzberg SL, Wold BJ & Pachter L Transcript assembly and quantification by RNA-Seq reveals unannotated transcripts

- and isoform switching during cell differentiation. *Nat Biotechnol* 28, 511–515, doi:10.1038/nbt.1621 (2010). [PubMed: 20436464]
33. Consortium, G. T. The Genotype-Tissue Expression (GTEx) project. *Nat Genet* 45, 580–585, doi: 10.1038/ng.2653 (2013). [PubMed: 23715323]
 34. Wang W, Tang Y, Wang Y, Tascau L, Balcerak J, Tong W, Levine RL, Welch C, Tall AR & Wang N LNK/SH2B3 Loss of Function Promotes Atherosclerosis and Thrombosis. *Circ Res* 119, e91–e103, doi:10.1161/CIRCRESAHA.116.308955 (2016). [PubMed: 27430239]
 35. Huan T, Meng Q, Saleh MA, Norlander AE, Joehanes R, Zhu J, Chen BH, Zhang B, Johnson AD, Ying S, Courchesne P, Raghavachari N, Wang R, Liu P, International Consortium for Blood Pressure, G., O'Donnell CJ, Vasan R, Munson PJ, Madhur MS, Harrison DG, Yang X & Levy D Integrative network analysis reveals molecular mechanisms of blood pressure regulation. *Mol Syst Biol* 11, 799, doi:10.15252/msb.20145399 (2015). [PubMed: 25882670]
 36. Harrison DG The immune system in hypertension. *Trans Am Clin Climatol Assoc* 125, 130–138; discussion 138–140 (2014). [PubMed: 25125726]
 37. Singh M, Singh AK, Pandey P, Chandra S, Singh KA & Gambhir IS Molecular genetics of essential hypertension. *Clin Exp Hypertens* 38, 268–277, doi:10.3109/10641963.2015.1116543 (2016). [PubMed: 27028574]
 38. Astor BC, Shafi T, Hoogeveen RC, Matsushita K, Ballantyne CM, Inker LA & Coresh J Novel markers of kidney function as predictors of ESRD, cardiovascular disease, and mortality in the general population. *American journal of kidney diseases : the official journal of the National Kidney Foundation* 59, 653–662, doi:10.1053/j.ajkd.2011.11.042 (2012). [PubMed: 22305758]
 39. Blass G, Mattson DL & Staruschenko A The function of SH2B3 (LNK) in the kidney. *Am J Physiol Renal Physiol* 311, F682–F685, doi:10.1152/ajprenal.00373.2016 (2016). [PubMed: 27440780]
 40. Dale BL & Madhur MS Linking inflammation and hypertension via LNK/SH2B3. *Curr Opin Nephrol Hypertens* 25, 87–93, doi:10.1097/MNH.000000000000196 (2016). [PubMed: 26717315]
 41. Rudemiller NP, Lund H, Priestley JR, Endres BT, Prokop JW, Jacob HJ, Geurts AM, Cohen EP & Mattson DL Mutation of SH2B3 (LNK), a genome-wide association study candidate for hypertension, attenuates Dahl salt-sensitive hypertension via inflammatory modulation. *Hypertension* 65, 1111–1117, doi:10.1161/HYPERTENSIONAHA.114.04736 (2015). [PubMed: 25776069]
 42. Huan T, Meng Q, Saleh MA, Norlander AE, Joehanes R, Zhu J, Chen BH, Zhang B, Johnson AD, Ying S, Courchesne P, Raghavachari N, Wang R, Liu P, O'Donnell CJ, Vasan R, Munson PJ, Madhur MS, Harrison DG, Yang X & Levy D Integrative network analysis reveals molecular mechanisms of blood pressure regulation. *Molecular systems biology* 11, 799 (2015). [PubMed: 25882670]
 43. Devalliere J & Charreau B The adaptor Lnk (SH2B3): an emerging regulator in vascular cells and a link between immune and inflammatory signaling. *Biochem Pharmacol* 82, 1391–1402, doi:S0006–2952(11)00406–0 [pii]10.1016/j.bcp.2011.06.023 (2011). [PubMed: 21723852]
 44. Swirski FK & Nahrendorf M Bone Marrow Takes Center Stage in Cardiovascular Disease. *Circ Res* 119, 701–703, doi:10.1161/CIRCRESAHA.116.309584 (2016). [PubMed: 27587406]
 45. Trott DW, Thabet SR, Kirabo A, Saleh MA, Itani H, Norlander AE, Wu J, Goldstein A, Arendshorst WJ, Madhur MS, Chen W, Li CI, Shyr Y & Harrison DG Oligoclonal CD8+ T Cells Play a Critical Role in the Development of Hypertension. *Hypertension* 64, 1108–1115, doi: 10.1161/HYPERTENSIONAHA.114.04147 (2014). [PubMed: 25259750]
 46. Argyropoulos CP, Chen SS, Ng YH, Roumelioti ME, Shaffi K, Singh PP & Tzamaloukas AH Rediscovering Beta-2 Microglobulin As a Biomarker across the Spectrum of Kidney Diseases. *Front Med (Lausanne)* 4, 73, doi:10.3389/fmed.2017.00073 (2017). [PubMed: 28664159]

Novelty and Significance**What is New?**

To our knowledge, no study to-date has analyzed in detail the role of the *SH2B3*- β_2 M axis in blood pressure regulation. Moreover, population studies of associations between β_2 M and hypertension are either limited to cross-sectional analyses or do not consider kidney disease status.

What is Relevant?

We provide novel evidence of a causal role of the *SH2B3*- β_2 M axis in blood pressure regulation, which can be further explored as a potential therapeutic target in hypertension.

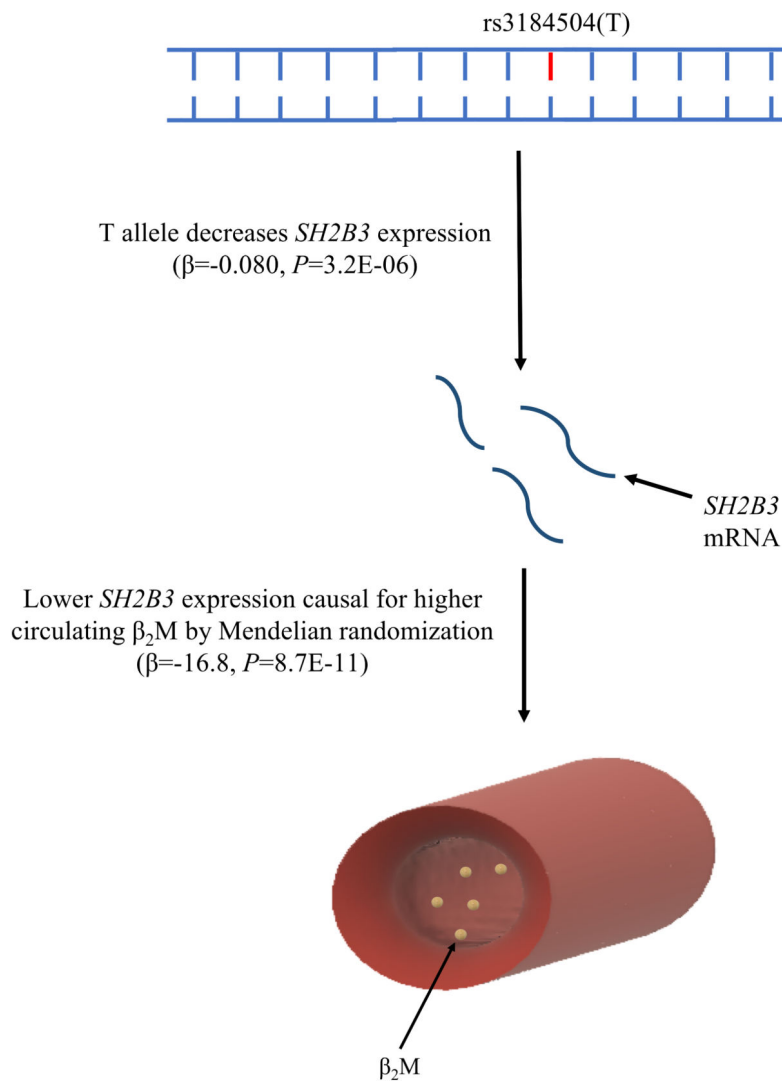


Figure 1. Mendelian randomization demonstrating *SH2B3* expression to be causal for plasma β_2M levels and how both increase hypertension risk

Mendelian randomization (MR) analyses were conducted using rs3184504 to instrument *SH2B3* expression to determine the causal effect of *SH2B3* expression on circulating β_2M levels. rs3184504(T) is a *cis*-eQTL variant for *SH2B3* expression ($\beta=-0.08$, $P=3.2E-06$), *trans*-pQTL variant for circulating β_2M ($\beta=0.12$, $P=9.7E-11$), and GWAS SNP associated with systolic ($\beta=0.58$, $P=5E-09$) and diastolic ($\beta=0.48$, $P=3E-14$) blood pressure³⁵. The MR Wald ratio demonstrated that *SH2B3* expression was causal for circulating β_2M ($\beta=-16.8$, $P=8.7E-11$).

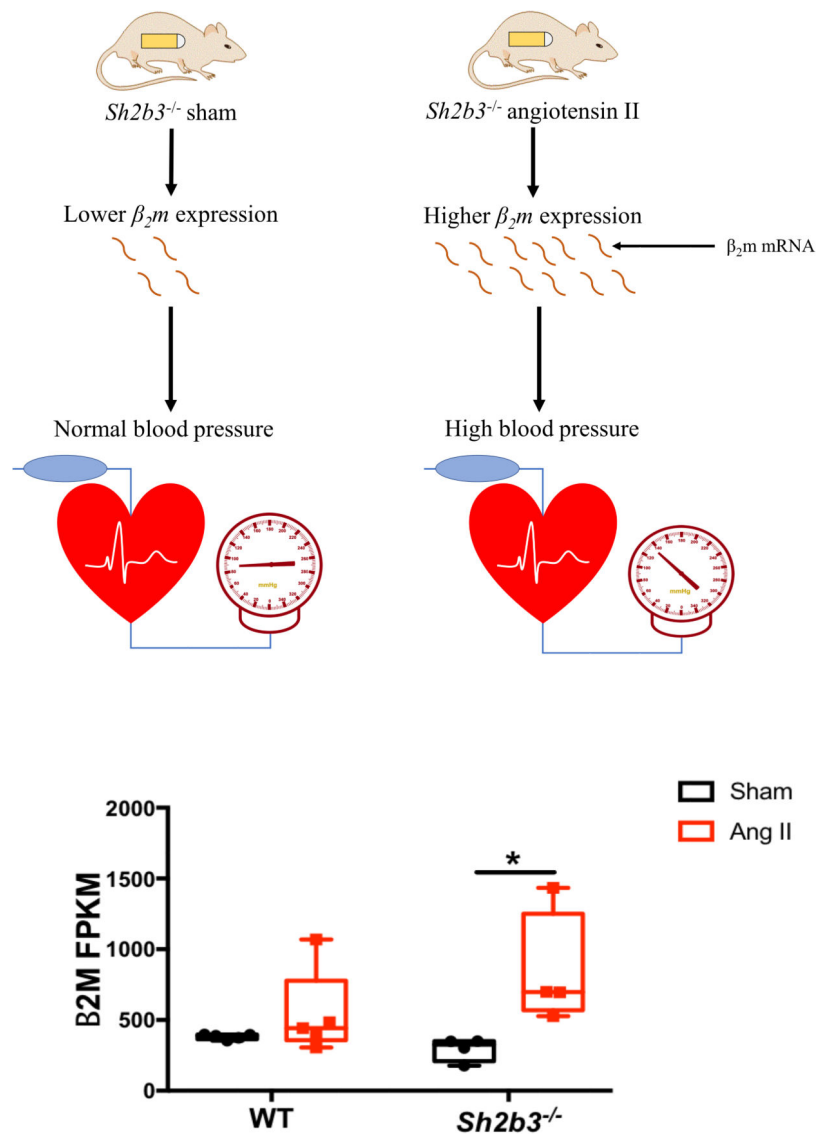


Figure 2. Angiotensin II infusion significantly increases β_2M expression in $Sh2b3^{-/-}$ mice WT and $Sh2b3^{-/-}$ mice⁴⁷ were infused with vehicle (sham) or angiotensin II (Ang II) for 2 weeks. We previously showed that $Sh2b3^{-/-}$ mice develop exaggerated hypertension in response to Ang II infusion. RNA sequencing analysis demonstrated that whole blood β_2M transcripts are increased in $Sh2b3^{-/-}$ mice, but not WT mice, in response to Ang II-induced hypertension ($P=0.037$ by 2-way ANOVA).

Table 1. Demographic and clinical characteristics of all FHS study participants by β_2 M quartile (N=7065)

β_2 M Quartile	1		2		3		4		Trend (P)
	Mean or number	SD or percent	Mean or number	SD or percent	Mean or number	SD or percent	Mean or number	SD or percent	
N	1794		1751		1754		1766		
β_2 M (min,max, mg/L)	1.03	1.19	1.3	1.4	1.49	1.62	1.81	2.2	
Age (year)	42.3	10.4	46.1	12.0	50.8	13.0	58.7	13.8	0.0066
BMI (kg/m ²)	26.0	4.8	27.2	5.2	27.9	5.5	28.8	6.0	0.0002
Systolic BP (mmHg)	116.5	14.8	119.3	15.5	122.5	16.9	127.1	19.3	0.73
Diastolic BP (mmHg)	74.3	9.8	75.4	9.3	75.6	9.6	73.9	10.0	0.0001
Total cholesterol (mg/dL)	190.7	35.1	195.1	36.0	196.2	35.5	194.0	37.4	0.0002
HDL cholesterol (mg/dL)	58.1	16.8	55.0	16.3	52.9	16.0	50.5	16.1	0.014
Fasting blood sugar (mg/dL)	94.8	16.9	96.9	18.7	100.2	23.2	104.6	28.3	0.61
eGFR (mL/min/1.73 m ²)	103.5	13.1	98.5	14.4	92.2	15.2	81.1	19.4	0.40
Women (n, %)	1049	58.5	897	51.2	899	51.3	948	53.7	0.012
Hypertension treatment (n, %)	165	9.2	265	15.1	381	21.7	661	37.4	0.005
Hypertension (n, %)	306	17.1	418	23.9	557	31.8	864	48.9	0.0014
Lipid-lowering treatment (n, %)	167	9.3	177	10.1	276	15.7	367	20.8	0.12
Diabetes treatment (n, %)	24	1.3	41	2.3	70	4.0	140	7.9	0.057
Diabetes (n, %)	43	2.4	75	4.3	134	7.6	206	11.7	0.25
Cigarette smoker (n, %)	294	16.4	245	14.0	249	14.2	217	12.3	0.45
CKD (n, %)	4	0.22	9	0.51	29	1.7	235	13.3	<.0001
Albuminuria (n, %)	51	3.04	48	3.06	66	4.33	156	10.8	0.0004
UACR (median, Q1/Q3,mg/g)	4.41	2.76/8.82	4.06	2.56/7.77	4.22	2.64/8.09	5.25	2.92/ 11.96	0.41

Linear P trend values are adjusted for baseline age, sex, and study cohort. All P values less than 0.05 are shown in **bold**.

Abbreviations: BMI=body mass index; BP=blood pressure; HDL=high density lipoprotein; eGFR=estimated glomerular filtration rate; CKD=chronic kidney disease (defined as eGFR<60 mL/min/1.73m²); UACR=urinary albumin-creatinine ratio (albuminuria defined as UACR 30 mg/g)

Table 2.Odds ratios for prevalent hypertension by β_2 M levels

β_2 M levels	Cases/non-cases	Odds ratio	95% CI		P value
Continuous Model					
Log β_2 M (per 1 SD)	2145/4920	1.14	1.05	1.24	0.0026
Quartile Model					
β_2 M quartile 1	306/1488	Ref			
β_2 M quartile 2	418/1333	1.03	0.86	1.23	0.75
β_2 M quartile 3	557/1197	1.04	0.87	1.24	0.67
β_2 M quartile 4	864/902	1.29	1.05	1.57	0.013

P values less than 0.05 are shown in **bold**.

Abbreviations: CI=confidence interval; SD= standard deviation

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Table 3.Odds ratios for new-onset hypertension by β_2 M levels

β_2 M levels	Cases/non-cases	Odds ratio	95% CI		P value
Continuous Model					
Log (β_2 M(mg/L))	886/3303	1.18	1.07	1.32	0.0011
Quartile Model					
β_2 M quartile 1	153/1103	Ref			
β_2 M quartile 2	194/953	0.95	0.71	1.28	0.76
β_2 M quartile 3	268/761	1.3	0.99	1.7	0.057
β_2 M quartile 4	271/486	1.59	1.20	2.11	0.0011

P values less than 0.05 are shown in **bold**.

Abbreviations: CI=confidence interval; SD= standard deviation

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Table 4.

Reclassification analyses for new-onset hypertension

Integrated Discrimination Improvement (IDI)				
Study population (N-cases/N-total)	IDI	SE	P value	95% CI
Total sample (886/4189)	0.0021	0.001	0.029	(0.0002,0.0041)
Individuals free of baseline albuminuria and CKD (663/3638)	0.0036	0.0014	0.0084	(0.0009,0.0063)
Net Reclassification Index (NRI)				
Study population (N-cases/N-total)	NRI	SE	P value	95% CI
Total sample (886/4189)	0.0073	0.0088	0.41	(-0.01,0.025)
Individuals free of baseline albuminuria and CKD (663/3638)	0.030	0.012	0.017	(0.0055,0.0535)

P values less than 0.05 are shown in **bold**.

The clinical model for hypertension consisted of the following baseline covariates: age, sex, study cohort, BMI, diabetes, cigarette smoking, levels of total and HDL-cholesterol, statin-use, eGFR, and systolic and diastolic blood pressure. For the NRI, the case validation, or additional percentage of events correctly reclassified over clinical model, was 1% in the general sample and 3% in those free of baseline albuminuria and CKoD; the non-case validation, or additional percentage of non-events correctly reclassified over the clinical model, was 0% for the general sample and those free of baseline albuminuria and CKD.

Abbreviations: CI=confidence interval; CKD=chronic kidney disease; IDI=integrated discrimination improvement; NRI=net reclassification index; SE=standard error