Volatile Compounds in Seasoning Sauce Produced from Soy Sauce Residue by Reaction Flavor Technology

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ABSTRACT: Volatile flavor compounds created in the mixture of enzymatic hydrolysates of soy sauce residue and defatted soybean by reaction flavor technology (RFT) were analyzed and compared using solid phase micro-extraction/gas chromatography/mass selective detector to develop a seasoning sauce. Using response surface methodology analysis, RFT was performed by adding 0.50% fructose, 0.33% glutamic acid, 0.68% arginine, 0.37% methionine, and 0.86% glycine in the base and reaction conditions at 93°C for 120 min. A total of 57 compounds, 43 in RFT and 45 in control, were detected, including 8 aldehydes and ketones, 6 aromatic hydrocarbons, 3 acids, 12 alcohols, 6 esters, 4 furans, 9 nitrogen-containing compounds, 4 sulfur-containing compounds, and 5 miscellaneous. In RFT samples, aldehydes and ketones, aromatic hydrocarbons, alcohols, esters, and S-containing compounds were significantly increased. Sulfur-containing compounds were increased by 687 fold compared to control samples (P<0.05). Among these, the main contributors to the aroma activity of RFT samples were considered to be, with a very low threshold, the newly generated compounds, dimethyl disulfide (roasted onion/garlic-like/meaty odor), dimethyl trisulfide (roasted garlic-like/meaty odor), and methional (roasted potato/potato chip-like odor).

Keywords: soy sauce residue, enzymatic hydrolysate, response surface methodology, reaction flavor

INTRODUCTION

Soy sauce has been flavored as a liquid condiment for a long time in Asia. In Korea, soy sauces can be roughly split into two categories by their ingredients: Korean-style soy sauce (hansik ganjang) and modernized soy sauce (gaeryang ganjang) (1,2). The Korean Ministry of Food and Drug Safety Food Code classifies modernized soy sauce into four categories by their different processing methods: brewed soy sauce, acid-hydrolyzed soy sauce, enzyme-hydrolyzed soy sauce, and blended soy sauce (1,2). Brewed soy sauce is traditionally made from soybean and wheat. Nowadays, defatted soybean is widely used as a cost-saving ingredient because the necessary component for soy sauce production is the protein, not the oil. The rich fermented soybean flavor and the savory soy sauce taste make brewed soy sauce widely accepted by consumers. The consumption of brewed soy sauce has shown rapid growth, and the Korea Agro-Fisheries and Food Trade Corporation has reported that consumption in the domestic market was 64,500 tons in 2014 (3).

Soy sauce residue (SSR) is a by-product of the brewed soy sauce manufacturing process. However, it is usually discarded without suitable application or is partially used as feedstuff. Assuming $13 \sim 15\%$ of total soy sauce is converted to SSR as a by-product, approximately 9,000 tons of SSR are produced every year by the soy sauce industry of Korea. SSR still contains many nutrients such as crude protein (23.09 g/100 g) and amino-N (788.00 mg/100 g) (4).

Chen et al. (5) investigated the hydrolysis efficiency of SSR using ultrasonic probe-assisted enzymolysis technology. Gao et al. (6) researched the isolation, identification and amino acid composition of proteins in SSR. However, most previous studies on SSR have focused on the extraction of high value-added compounds and the hydrolysis condition. There is limited research on SSR protein as an effective resource. If SSR can be used to partly replace defatted soybean in the production of seasoning sauce, and the sensory quality of the product made with SSR can be improved, this should prove advantageous for reducing the cost of developing flavoring agents.

Reaction flavor technology (RFT) has been widely used in food production for improving sensory quality (7-9). Kim and Kim (7) developed meat flavor extract by using non-enzymatic browning of reaction precursors. Using

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reaction flavor, Kim and Beak (8) developed a baked beef flavor, and Kim et al. (9) developed a boiled-type shrimp flavor. These thermally induced flavor compounds are mainly concerned with development of cooked, nutty, and meat-like odor notes, and mainly consisted of pyrazines, aldehydes, and sulfur-containing compounds (10). Hwang et al. (11) studied the relative reactivities of amino acids in pyrazine formation using glycine, glutamic acid, and arginine, and reported that a large amount of pyrazines are generated by adding glycine. Hou et al. (12) also reported that glycine is the simplest amino acid to generate pyrazines from and is usually used to induce meat-like flavor, and Yu and Ho (13) reported that sulfur-containing compounds can be generated from a thermal reaction of methionine. Therefore, the objective of this study was to develop and compare reaction flavor compounds in seasoning sauce by applying RFT to the base of enzymatic hydrolysates of SSR and defatted soybean (DS).

MATERIALS AND METHODS

Materials

SSR (moisture: 32.91%, crude protein: 23.09%, crude lipid: 7.68%, and crude ash: 8.85%) (4) and DS (moisture: 6.95%, crude protein: 51.58%, crude lipid: 1.35%, and crude ash: 6.12%) were donated by Sungsim Master Food Co. (Changnyeong, Korea). SSR was packaged by 2-layer polyethylene film, stored in a low temperature room $(5 \sim 8^{\circ}C)$, and crushed before use. DS was crushed and boiled in an autoclave (Gaon Science, Bucheon, Korea) at 120°C for 15 min, dried, crushed again, and stored in a low temperature room (5~8°C) until use. The commercial proteases Alcalase[®] 2.4 L, Protamex[®], and Flavourzyme[®] 500 MG were produced by Novozymes (Copenhagen, Demark) and donated by Biosis Co., Ltd. (Busan, Korea). The amino acids arginine (purity: >98.5%, Ajinomoto Co., Inc., Tokyo, Japan), glycine (purity: >98.5%, Ajinomoto Co.), methionine (purity: >99.0%, DL-methionine, Dongeun Co., Pyeongtek, Korea), glutamic acid (purity: >99.0%, Dongeun Co.), and fructose (ADM Decatur, Decatur, IL, USA) used for reaction flavor experiments were purchased from a local food additives shop.

Preparation for the enzymatic hydrolysates of SSR and DS To enzymatically hydrolyze SSR and DS, three commercial enzymes (Alcalase[®] 2.4 L, Protamex[®], and Flavourzyme[®] 500 MG) were selected and used under their respective optimum conditions, as provided by Novozymes (14). Hydrolysis was conducted with a double-jacket reactor made of pyrex (500 mL capacity, custom-production). Optimal hydrolysis conditions for SSR have been described by Cha and Wang (14) and are as fol-

lows: combine 8.79% (w/v) SSR and 100 mL distilled water in a reactor, adjust pH to 7.0 using 0.1 N NaOH, hydrolyze with 0.40% (v/w) Alcalase[®] 2.4 L at 50°C under agitation. After 2 h, 0.43% of mixed enzymes (Flavourzyme[®] 500 MG : Protamex[®]=1:1 w/w) are put into the reactor and hydrolyzed for 4.43 h. Optimal hydrolvsis conditions described for DS by Cha and Wang (15) are as follows: combine 10% (w/w) pretreated DS powder, 0.40% (w/w) Protamex[®], and 100 mL distilled water in a reactor, hydrolyze at 50°C for 4 h under agitation, hydrolyze with 0.40% (w/w) Flavourzyme[®] 500 MG for 5 h. The reactants are then inactivated at 98°C for 10 min and the enzymatic hydrolysates of SSR and DS were obtained after filtration. The mixture of hydrolysates from SSR and DS (1:1, v/v) were used as a base for sensory evaluation (data not shown).

Establishment of optimal reaction flavor conditions by response surface methodology (RSM)

To induce formation of reaction flavor in the base, four amino acids such as glutamic acid, methionine, glycine, and arginine, and the sugar fructose were selected through sensory evaluation (odor) in the preliminary experiments.

The optimal reaction conditions were established by RSM and consist of a central composite design with three independent variables coded at five levels (-2, -1, 0, 1,and 2), as described by Gontard et al. (16). The independent variables were set as the concentration of arginine, methionine, and glycine (Table 1). The dependent variables were expressed as scores of odor and taste from 19 treatments by quantitative description analysis (QDA).

Two reactants, 0.33% (w/v) glutamic acid and 0.50% (w/v) fructose, were selected as the precursors in preliminary experiments, and were put into the base in advance. After adding optimal concentrations of the other 3 amino acids (Arg, Met, and Gly), thermal reaction was carried out at 93° C for 2 h.

QDA

Odor and taste profiles for QDA were assessed by a 10member panel (2 males and 8 females), consisting of undergraduates, graduates and staff, who had trained for

Table 1. Coded level of independent variables in experimental $\ensuremath{\mathsf{design}}^{1j}$

Coded units	Arg	Met	Gly	
-2	0.67	0.17	0.33	
-1	0.83	0.25	0.67	
0	1.00	0.33	1.00	
1	1.17	0.42	1.33	
2	1.33	0.50	1.67	

¹⁾The content (w/v) was g, % of each additive per 100 mL of the mixture of enzymatic hydrolysates of soy sauce residue and defatted soybean (1:1 ratio, v/v).

over 3 months. The panelists first analyzed the samples individually with two commercial seasoning sauces (S and D companies) as references. The panel leader recorded and compiled the results. Three to five open discussions were held to clarify the QDA results, and five expressions of odors and tastes about target samples were established. The scores for odor and taste notes were based on a 9-point scale (1: not detectable, 9: extremely intense). A transparent glass cup (60 mL capacity) was used as the sample vessel and the samples were coded with 3 randomly selected 3-digit numbers. Participants were served a cup of water to rinse after the test. All procedures were approved by the institution review board of Changwon National University (104027-201607-HR-016).

Solid phase microextraction (SPME)/gas chromatography (GC)/mass selective detector (MSD) analysis

A SPME device was used with a SupelcoTM solid phase microextraction fiber holder (Supelco Inc., Bellefonte, PA, USA) and polydimethylsiloxane/divinylbenzene fiber (0.65 µm coating thickness, Supelco Inc.). The sample (6 mL) and 1 µL hexyl acetate (91.11 ng) (Sigma-Aldrich Co., St. Louis, MO, USA) as an internal standard (IS) compound were put into a 20 mL headspace glass vial (Supelco Inc.). Extraction was carried out at 40°C for 25 min using a magnetic stirrer. Further details have been described by Cha et al. (4). Extractions were performed on each sample in triplicate.

The GC/MS system consisted of a Perkin Elmer clarus 600T GC/MSD (Perkin Elmer Inc., Fremont, CA, USA) with splitless mode equipped with a DB-WAXTM capillary column (60 m length×0.25 mm I.D.×0.25 µm film thickness, J&W Scientific, Folsom, CA, USA). The oven temperature was initially programmed at 40°C (held for 5 min) and then increased to 220°C (held for 10 min) at a rate of 4°C/min. The MSD condition was as follows: He carrier gas flow 1.0 cm/s; capillary direct interface temperature 220°C; ion source temperature 204°C; ionization energy 70 eV; mass range 33~350 amu; electron multiplier voltage 1,500 V.

Identification and relative abundance of volatile flavor compounds

Volatile flavor compounds were identified by comparing retention indices (RI) and using NIST (version 2.0 g, The National Institute of Standards and Technology, Gaithersburg, MD, USA) standard MS library data (Perkin Elmer Inc.). The concentration (ng/g) of tentatively identified compounds were calculated as a relative content to the amount of IS put into the samples.

Statistical analysis

Statistical analysis of RSM design data was analyzed

through Statistical Analysis System version 19 (SAS institute Inc., Cary, NC, USA). Statistical analysis of other experimental data was conducted using SPSS (version 22.0, SPSS Inc., Chicago, IL, USA), with a 95% confidence level used to indicate statistical significance.

RESULTS AND DISCUSSION

Establishment of optimal reaction flavor conditions by RSM Scores of odor and taste from the 19 treatments are shown in Table 2. In the QDA analysis, five kinds of odor (savory, smoke, nutty, salty, and sweet) and five kinds of taste (savory, dried shrimp-like, nutty, smoke, and acidic) were described during the sensory evaluation (data not shown). During SAS analysis, savory for odor and dried shrimp-like for taste were considered suitable for RSM and were therefore chosen for analysis.

Odor and taste values analyzed using SAS and statistical results are shown in Table 3. Considering that the purpose of this study was to generate reaction flavor from the mixture of enzymatic hydrolysates from SSR and DS (1:1 ratio, v/v), we focused on odor over taste with RFT, since adjusting taste is possible using food additives in later process whereas flavor is hard to change. The odor results are mainly discussed here for establishing the op-

Table 2. Response surface methodology (RSM) of dependentvariables for making seasoning sauce from the mixture of en-zymatic hydrolysates of soy sauce residue (SSR) and defattedsoybean (DS)

Design Indepen		dent variables ¹⁾		Dependent variables ²⁾		
point	Arg	Met	Gly	Odor	Taste	
1	-1	-1	-1	4.97±0.21	4.93±0.35	
2	1	-1	-1	4.37±0.15	5.10±0.72	
3	-1	1	-1	5.33±0.42	4.73±0.25	
4	1	1	-1	4.77±0.12	5.10±0.17	
5	-1	-1	1	5.20±0.46	5.20±0.26	
6	1	-1	1	5.07±0.25	5.47±0.42	
7	-1	1	1	5.23±0.32	4.87±0.15	
8	1	1	1	5.00±0.61	5.23±0.15	
9	2	0	0	4.50±0.52	5.23±0.31	
10	-2	0	0	5.67±0.32	4.33±0.12	
11	0	2	0	4.97±0.47	5.03±0.45	
12	0	-2	0	5.13±0.06	4.83±0.31	
13	0	0	2	5.10±0.10	5.03±0.65	
14	0	0	-2	4.63±0.59	5.00±0.17	
15	0	0	0	4.50±0.30	4.90±0.44	
16	0	0	0	4.50±0.26	4.70±0.26	
17	0	0	0	4.47±0.25	4.77±0.15	
18	0	0	0	4.50±0.15	4.80±0.52	
19	0	0	0	4.70±0.35	4.60±0.81	

¹⁾The coded levels of independent variables are the same as represented in Table 1.

²⁾Odor, savory odor value; Taste, dried shrimp-like taste value (mean value, n=3).

Table 3. Model coefficients estimated by multiple linear regression for dependent variables for making seasoning sauce from the mixture of enzymatic hydrolysates of soy sauce residue and defatted soybean

Factor	Odor	Taste	
Constant	4.558**	4.799**	
Linear			
[Arg]	-0.241**	0.186*	
[Met]	0.025	-0.023	
[Gly]	0.125**	0.061	
Quadratic			
[Arg] ²	0.147**	0.023	
[Met] ²	0.138*	0.061	
[Gly] ²	0.092*	0.082	
Crossproduct			
[Arg]×[Met]	-0.008	0.036	
[Arg]×[Gly]	0.100	0.011	
[Met]×[Gly]	-0.100	-0.046	
Model			
Linear	<0.001	0.030	
Quadratic	0.002	0.275	
Crossproduct	0.134	0.882	
R-Square	0.915	0.682	
Total model	0.001	0.136	
Lack of fit	0.113	0.062	

*P<0.05 and **P<0.01.

timal conditions of reaction flavors.

The model coefficients are shown in Table 3. In linear, [Arg] and [Gly] were significant at a 99% confidence level; in quadratic, $\left[Arg\right]^2$ was significant at a 99% confidence level, and $\left[Met\right]^2$ and $\left[Gly\right]^2$ were significant at a 95% confidence level; neither were considered significant in crossproduct. This response model was determined as an adequate model because R^2 was 0.915, near to 1. Lack of fit (0.113) showed P>0.05 and the total model (0.001) showed P<0.05. Therefore, the model equation obtained from the SAS result was: odor score =4.558-0.241[Arg]+0.025[Met]+0.125[Gly]+0.147 $[Arg]^{2}+0.138[Met]^{2}+0.092[Gly]^{2}-0.008[Arg][Met]+$ 0.100[Arg][Gly]-0.100[Met][Gly]. The predicted odor score was 4.27 during the stationary point. From the ridge analysis for accepting high scores, the optimal flavoring conditions were set as 0.68% (w/v) arginine, 0.37% (w/v) methionine and 0.86% (w/v) glycine of the base, and a high odor score (5.66) was obtained.

In conclusion, the optimal reaction flavor conditions were as follows: the optimum condition of the precursors was 0.33% (w/v) glutamic acid, 0.50% (w/v) fructose, 0.68% (w/v) arginine, 0.37% (w/v) methionine, and 0.86% (w/v) glycine. The reactions were then carried out in a shaking water bath for 2 h at 93° C.

Volatile flavor compounds of seasonings produced with and without RFT

Volatile compounds in the seasonings produced with and



Fig. 1. Mean amounts of group compounds detected in the mixture of enzymatic hydrolysates of soy sauce residue and defatted soybean with reaction flavor technology (RFT) and without (Control). Mean values within the same group with asterisks are significantly different (P<0.05).

without RFT were analyzed by SPME and GC/MSD. A total of 57 compounds, 43 in RFT and 45 in control, were detected in samples, including 8 aldehydes and ketones, 6 aromatic hydrocarbons, 3 acids, 12 alcohols, 6 esters, 4 furans, 9 nitrogen-containing compounds, 4 sulfur-containing compounds and 5 miscellaneous compounds (Table 4). Comparison of the mean amounts of each compound group detected with and without RFT are shown in Fig. 1.

Aldehydes: Four aldehydes (2-methylbutanal, 3-methylbutanal, hexanal, and benzaldehyde) were identified in RFT. Among these, the most abundant compound was benzaldehyde (176.00 ng/g), which has a characteristic almond-like odor, and was increased 21.54 fold greater than the control (8.17 ng/g) (Table 4). Benzaldehyde has been reported to have a high odor activity value in Japanese soy sauce (17). The second most abundant compound was hexanal (45.44 ng/g), which has a fruity odor. Hexanal has been reported to be generated by RFT via lipid oxidation (18), and has been identified in a koji-fermented soy sauce (18,19). Two compounds, 2-methylbutanal (roasted cocoa-like odor) and 3-methylbutanal (banana-like odor), regularly found in soy sauces (18-21) were detected, and were increased 3.87 and 4.77 fold by RFT, respectively. The formation of 2-methylbutanal and 3-methylbutanal can be explained by degradation of the amino acids isoleucine and leucine, respectively (17,18), and each consist of one less carbon than the original amino acids. Degradation of isoleucine and leucine has been reported as a pathway of the Maillard reaction (22,23).

Ketones and aromatic hydrocarbons: Four ketones (2-heptanone, 3-octanone, 2-octanone, and 3-hydroxybutanone) were identified in both the RFT and control samples. The abundance of 2-octanone (63.98 ng/g) was increased 10.65 fold in RFT samples compared with controls (6.01

 Table 4. Volatile flavor compounds of the mixture of enzymatic hydrolysates of soy sauce residue and defatted soybean with and without reaction flavor technology¹⁾
 (unit: ng/g)

Compounds	RI	RFT	Control	t-value
Aldehydes and Ketones (8)		351.72±35.63	56.73±65.73	13.05*
2-Methylbutanal	846	17.03±10.53	4.40±2.87	2.05
3-Methylbutanal	858	35.66±28.76	7.48±6.62	1.65
Hexanal	1,085	45.44±28.47	2)	2.77
2-Heptanone	1,189	10.78±12.76	16.12±14.11	-0.49
3-Octanone	1,259	2,39±0,51	11,93±1,32	-11.67*
2-Octanone	1,290	63,98±9,49	6.01±0.83	10.54*
3-Hydroxybutanone	1,296	0.44±0.34	2.62±0.74	-4.92*
Benzaldehyde	1,542	176.00±4.13	8.17±3.79	51.87*
Aromatic hydrocarbons (6)	,	355,57±56,12	117,68±44,80	5.74*
Methylbenzene	1,045	_	2.65±1.44	-3.18*
2-Methoxyphenol	1,880	30.71±4.11	19.09±6.83	2.52
2-Phenylethanol	1,929	73.41±15.25	33.60±12.17	3.54*
2-Phenylbutanol	2,002	0.12±0.04	2.89±1.21	-3.97*
Phenol	2,024	5.01±0.38	4.11±1.99	0.77
4-Ethyl-2-methoxyphenol	2,051	246.32±48.65	55.34±22.76	6.16*
Acids (3)		63.96±6.13	124.23±53.43	-1.94
Acetic acid	1,462	36.25±3.66	43.55±18.65	-0.67
2-Methylpropanoic acid	1,576	_	18.85±6.15	-5.31*
3-Methylbutanoic acid	1,676	27.71±2.57	61.83±28.76	-2.05
Alcohols (12)		757.63±128.90	198.15±45.07	6.82*
Cyclopropyl methanol	1,101	66.52±41.59	_	1.59
3-Methyl-1-butanol	1,211	626.85±129.66	79.68±15.80	7.26*
Pentanol	1,253	_	1.30±0.94	-2.39
2-Heptanol	1,320	5.32±1.00	2.46±0.38	4.64*
3,4-Dimethyl-1-pentanol	1,355	23.53±0.99	16.38±4.26	2.83*
3-Octanol	1,395	_	74.99±20.63	-6.30*
1-Octen-3-ol	1,452	3.08±0.14	8.61±2.64	-3.62*
Heptanol	1,457	8.29±4.40	_	3.26*
2-Ethyl-1-hexanol	1,491	16.17±1.27	5.37±1.88	8.25*
Octanol	1,560	2.19±0.55	6.33±3.75	-1.89
(Z)-2-Octen-1-ol	1,620	1.51±0.02	3.03±1.11	-2.38
Undecanol	1,899	4.17±4.20	-	1.72*
Esters (6)		50.29±10.18	18.65±6.40	4.56*
Ethyl 2-hydroxypropanoate	1,351	_	2.14±0.60	-6.11*
Diisopropyl carbonate	1,585	2.87±0.46	-	10.85*
Cyclopentyl 4-ethylbenzoate	1,756	12.72±7.28	-	3.02*
Ethylbezene acetate	1,803	20.13±3.48	4.22±1.85	7.00*
Methylbenzene propanoate	1,835	-	2.07±0.67	-5.40^{*}
Tetrahydro-2-furanmethanol acetate	1,887	14.57±0.86	10.22±3.62	2.03
Furans (4)		6.34±0.85	10.42±2.77	-2.76
Furfural	1,479	_	3.87±1.32	-5.10*
2-[(Methylthio)methyl]furan	1,519	0.29±0.04	-	12.56*
2-Furanmethanol	1,670	1.28±0.70	3.48±1.08	-2.96*
3-Phenylfuran	1,876	4.77±0.24	3.07±1.19	2.43
Nitrogen-containing compounds (9)		24.53±2.89	71.70±43.37	-1.88
1,3-Propanediamine	1,079	-	12.90±4.60	-4.86*
2,5-Dimethylpyrazine	1,331	0.15±0.06	8.01±1.86	-7.33*
2,6-Dimethylpyrazine	1,337	2.48±0.39	-	10.94*
2-Acetyl-1-pyrroline	1,338	-	1.32±0.30	-7.73*
2-Ethyl-6-methylpyrazine	1,401	0.44±0.22	1.11±0.51	-2.09
Trimethylpyrazine	1,413	8.78±1.08	7.88±2.53	0.56
2-Ethyl-5,6-dimethylpyrazine	1,471	_	1.90±0.61	-5.39*
Tetramethylpyrazine	1,482	12.52±2.00	14.65±5.98	-0.58
3,5-Diethyl-2-methylpyrazine	1,520	0.16±0.09	23.93±35.61	-1.16
Sulfur-containing compounds (4)		2,980.09±415.99	4.34±1.11	12.34*
Dimethyl disulfide	1,078	1,405.91±258.22	_	9.34*
Dimethyl trisulfide	1,396	1,558.53±154.84	-	17.43*

Table 4.	Continued
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Compounds	RI	RFT	Control	t-value
Methional	1,470	8.29±4.40	_	3.26*
2-Acetylthiazole	1,666	7.36±0.50	4.34±1.11	4.28*
Miscellaneous compounds (5)		3.17±0.45	39.06±27.44	-2.27
Limonene	1,200	-	25.18±22.54	-1.94
Trimethyl oxazole	1,202	3.17±0.45	-	12.34*
γ-Terpinene	1,249	-	6.63±5.82	-1.97
3-Methyl-6-(1-methylethylidene)cyclohexene	1,287	-	4.09±0.37	-19.20*
β-Bisabolene	1,736	_	3.16±0.93	-5.91

Data presented as mean \pm SD. (n=3).

RI, retention index on Supelcowax 10[™] column; RFT, reaction flavor technology.

**P*<0.05 by *t*-test.

¹⁾Concentration (ng/g) of each compound was calculated as a relative content to hexyl acetate (91.12 ng/g) put in sample (factor=1). ²⁾Not detected.

ng/g). Ketones are one kind of decomposed product of fat acidification and have herb and flower-like odors (24).

A total of 5 aromatic hydrocarbons were identified in the RFT samples and 6 in the controls. Methylbenzene, which was found in the control but not the RFT samples, may be catalytically oxidized to benzaldehyde. The most abundant aromatic hydrocarbon in the RFT samples was 4-ethyl-2-methoxyphenol (246.32 ng/g), and was 4.45 fold greater than in controls. 4-Ethyl-2-methoxyphenol has been reported to be a significant contributor to the flavor of Japanese soy sauce because of its very strong soy sauce-like odor (17), and can be produced from both lignin pyrolysis and 2-methoxyphenol (25). Four aromatic hydrocarbons, 4-ethyl-2-methoxyphenol, 2-phenylethanol (73.41 ng/g, floral odor), 2-methoxyphenol (30.71 ng/g, smoky odor) and phenol (5.01 ng/g, green odor), have been reported in other soy sauces (19,20).

Acids: Two acids were found in the RFT samples, compared to three in the controls. These can be attributed to the acidic sensory of soy sauce. Acetic acid, a product of microbiology fermentation (26), was found to be the major acid in soy sauce. A similar results has been observed for Korean soy sauce (17). 3-Methylbutanoic acid (sweat /strong cheese-like odor) was the most abundant acid in control samples (61.83 ng/g), and was decreased to 27.17 ng/g following RFT. 2-Methylpropanoic acid, attributable to a rotten butter-like and sweat odor, was found at 18.85 ng/g in control samples, but was not present following RFT. Compounds such as 2-methylpropanoic acid and 3-methylpropanoic acid are reported to be mainly produced from branched-chain amino acids via the Ehrlich pathway by the action of various fungal enzymes during fermentation (27).

Alcohols: Twelve alcohols were overall identified in this study (10 in the RFT samples and 9 in the control samples). In both the RFT and control samples, 3-methyl-1-butanol (isoamyl alcohol/fruity/kiwi-like odor) showed the highest abundance (626.85 ng/g and 79.68 ng/g, respectively), increased 7.87 fold by reaction flavor. 1-Oc-

ten-3-ol (mushroom-like odor) has regularly been reported as one of the major constituents in soy sauce has a favorable impact on soy sauce products (20,21). 3-Octanol (cheese/mint-like odor), which is the dominant constituent in the soy sauce koji, and pentanol (wine-like odor) were not present following RFT. However, cyclopropyl methanol (66.52 ng/g), heptanol (8.29 ng/g), and undecanol (4.17 ng/g) were generated through RFT.

Esters: Six esters were identified in both the RFT and control samples. Control samples consisted of 18.66 ng/g esters, significantly increased by RFT to 50.28 ng/g. Tetrahydro-2-furanmethanol acetate was the most abundant ester in control samples (10.22 ng/g), followed by ethylbenzene acetate (4.22 ng/g), which has a wine-like odor. In RFT samples, ethylbezene acetate was the most abundant ester (20.13 ng/g) followed by tetrahydro-2-furanmethanol acetate (14.57 ng/g). Most of the esters are formed via esterification of fatty acids and alcohols during fermentation (28,29), and can contribute to flavor by minimizing the sharpness and bitterness imparted by amines and fatty acids (30,31).

Furans: Four furans were detected in both the RFT (6.34 ng/g) and control (10.42 ng/g) samples, however no significant differences in their abundances were recorded. The heterocyclics in soy sauce, which include furans and pyrazines, are mainly formed via a Maillard reaction (20). The presence of several furans in soy sauce could result from both glucose pyrolysis and Maillard reactions (17). Generally, furans are formed from decomposition of Amadori and Heyns products in a Maillard reaction (32). Nitrogen-containing compounds: Almost all the nitrogen-containing compounds detected in RFT and control samples consisted of pyrazines. Pyrazines are an important group of nitrogen-containing flavor compounds, which are generally formed by the reaction of amino acids with a carbonyl group through Maillard reactions (33,34). Most pyrazines are important contributors to the flavors of roasted, toasted, and heated foods (35). A larger number of nitrogen-containing compounds were detected in the

control samples than following RFT. These data were thought to be a result of the precursor compounds applied to the reaction flavor system. 2,6-Dimethylpyrazine was detected only in RFT samples, and 1,3-propanediamine and 2-acetyl-1-pyrroline were only detected in control samples. Two pyrazines, 2,5-dimethylpyrazine (meaty /nutty odor) and 2,6-dimethylpyrazine (meaty/nutty/ coffee-like odor), are formed from cysteines by thermal treatment (36,37), and are reported to be the main pyrazines in several soy sauces, including Thai soy sauces (26), Korean acid hydrolyzed soy sauces (17), and Japanese soy sauces (38). Trimethylpyrazine (peanut and hazelnut-like odor) and tetramethylpyrazine (coffee/chocolate-like odor) also accounted for a large proportion of the nitrogen-containing compounds in both RFT (21.30 ng/g) and control (22.53 ng/g) samples.

Sulfur-containing compounds: Four sulfur-containing compounds dimethyl disulfide (roasted onion/garlic-like/ meaty odor), dimethyl trisulfide (roasted garlic-like/ meaty odor), methional (roasted potato/potato chip-like odor) and 2-acetylthiazole were predominantly detected following RFT; of these compounds, only 2-acetylthiazole was found in controls. The amount of sulfur-containing compounds was 687 fold higher following RFT samples compared with control samples (P < 0.05) (Fig. 1). Methional is a Strecker aldehyde of methionine, and is detected in cooked potatoes (32). Schutte and Teranishi (39) reported that methional is produced by decomposition of methionine, which was added as the precursor of reaction flavor in this study. Methional was subsequently decomposed into dimethyl sulfide and trimethyl sulfide. These three newly generated compounds (12 ng/g dimethyl disulfide, 0.01 ng/g dimethyl trisulfide, and 0.2 ng/g methional) (40) were, with a very low threshold, considered to be the main contributors to the aroma activity of RFT.

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AUTHOR DISCLOSURE STATEMENT

The authors declare no conflict of interest.

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