

REVIEW



New modalities of allergen immunotherapy

Manish Ramesh and Merhunisa Karagic

Montefiore Medical Center, Bronx, NY, USA

ABSTRACT

Allergen immunotherapy is a rapidly evolving field. Although subcutaneous immunotherapy has been practiced for over a hundred years, improved understanding of the underlying immunological mechanisms has led to the development of new, efficacious and better tolerated allergen-derivatives, adjuvants and encapsulated allergens. Diverse routes of allergen immunotherapy – oral, sublingual, epicutaneous and intralymphatic – are enabling immunotherapy for anaphylactic food allergies and pollen-food allergy syndrome, while improving the tolerability and effectiveness of aeroallergen immunotherapy. The addition of Anti-IgE therapy decreases adverse effects of subcutaneous and oral immunotherapy.

ARTICLE HISTORY

Received 11 January 2018
Revised 27 June 2018
Accepted 13 July 2018

KEYWORDS

Allergen Immunotherapy;
Oral Immunotherapy;
Subcutaneous
Immunotherapy; Sublingual
Immunotherapy;
Epicutaneous
Immunotherapy; adjuvants;
Anti-IgE

Introduction

In this review we will address non-registered forms of allergen immunotherapy for various kinds of IgE mediated hypersensitivity, including aeroallergies, food allergy and pollen food allergy syndrome. Allergen Immunotherapy has been in use for over a hundred years. At its core, subcutaneous immunotherapy (SCIT) for aeroallergies and stinging insect hypersensitivity has changed very little since inception. However, the field has moved forward with exciting new developments in immunotherapy including novel routes of delivery, modified allergens, allergen derivatives and combination therapy with biologics.

Our understanding of the immunological basis of allergy and the changes associated with allergen immunotherapy has vastly improved over the last three to four decades. In parallel, advances in molecular characterization have led to a better understanding of allergens at the protein and sequence level.

The process of preparing extracts from various naturally occurring source materials and periodic subcutaneous injection has remained the same. The evidence supporting subcutaneous immunotherapy for aeroallergies is plentiful but patchy.^{1,2} In the United States, standardization of extracts across manufacturers and allergens is limited to a few allergens. Despite these limitations, the overwhelming consensus and the practice parameters back the use of subcutaneous immunotherapy (SCIT) for inhalant allergies and *Hymenoptera* venom allergies.^{3–7} However, there is a risk of local and systemic reactions and poor adherence with SCIT.⁸ Several approaches have been adopted to remedy this including alternate routes and rapid build-up schedules. SCIT has been abandoned for food allergies due to a high rate of adverse reactions.^{9,10}

In this review, we will evaluate newer modalities of immunotherapy. Conventional SCIT for aeroallergens and

venom anaphylaxis have been reviewed extensively elsewhere. We will not discuss FDA approved products such as newly approved SLIT products that are outside the purview of this review article. We will present data based on some products that are in development (such as Epicutaneous immunotherapy), as they best illustrate a new technology. Here we will briefly summarize the evidence. We will discuss updates to our understanding of the immunological mechanism underlying immunotherapy. We evaluate the evidence supporting the treatment of food allergies using oral immunotherapy (OIT), sublingual immunotherapy (SLIT), and epicutaneous immunotherapy (EPIT). We will assess the evidence backing the use of anti-IgE in combination with other forms of immunotherapy. Finally, we will examine novel methods of antigen preparation and delivery such as intralymphatic immunotherapy.

Immunological mechanism of allergen immunotherapy

Despite a century of successful immunotherapy, the molecular and cellular mechanisms of immunotherapy remain poorly understood. There is some understanding of the immunological changes associated with SCIT. These include desensitization of mast cells and basophils, changes in immunoglobulin production, and the generation of regulatory T cells (Treg). We refer you to excellent reviews on the topic by Shamji and Durham,¹¹ Soyer OU et al.¹² and, Berin and Shreffler.¹³ Here we briefly summarize some of the theories and critical findings that impact the modalities of immunotherapy discussed below.

Aeroallergen SCIT, SLIT and OIT are associated with changes in mast cell activation. While the phenomenon of the decreased mast cell and basophil responsiveness is well

established,¹⁴ the mechanism is debatable and not directly observable. Partial degranulation, endocytosis of surface IgE¹⁵ and actin remodeling,¹⁶ have all been proposed based on in vitro and animal studies

Treg suppressor activity mediated by TGF β and IL10 are responsible for late effects on mast cell number and function. Allergen-specific activation of peripheral basophil activity is diminished particularly in OIT and SLIT where it has the potential for use as a marker of successful desensitization.^{17–20} This, however, is not universal. Peanut epicutaneous immunotherapy did not show significant changes in basophil activation.^{14,21,22}

Dendritic cell mediated induction of regulatory T and B cells of various kinds – natural Tregs, inducible Tregs, T follicular regulatory cells, B regs have all been described in various forms of immunotherapy. However, the role of dendritic cells is highly complex and varies tremendously by the site. Allergen (is) first encountered by different populations of mature and immature dendritic and Langerhans cells depending on the route of allergen immunotherapy. While epicutaneous immunotherapy relies on dermal resident dendritic cell populations, intralymphatic immunotherapy entirely bypasses them. With mucosal exposure as in SLIT mucosal Langerhans-like dendritic cells play a critical role by capturing the allergen within the oral mucosa. In response, Th2 cytokine production is decreased with concurrent increase in Th1 and suppressive cytokines (TGF β and IL10) and various T reg subpopulations – natural Treg, inducible T reg and T follicular regulatory cells. Natural and inducible T regs suppress the development of allergic diseases via (a) induction of suppressive cytokines, such as interleukin-10 (IL-10) and transforming growth factor β (TGF- β); (b) suppression of allergen-specific IgE and induction of IgG4 and IgA; (c) suppression of mast cells, basophils, eosinophils, and inflammatory dendritic cells and (d) by suppression of effector TH1, Th2 and Th17 cells.^{12,23} Multiple studies have shown that aeroallergen immunotherapy leads to an increase in local FOXP3⁺ CD25⁺ T cells in the nasal mucosa in allergic rhinitis and airways of asthmatics.^{24–27} Similarly, OIT is also thought to increase the migratory potential of Tregs cells towards intestinal epithelial cell and induce epigenetic changes enhancing their function via FoxP3 + locus.²⁸ It increases CD4⁺ CD25⁺ FoxP3⁺ Tregs and, Foxp3 + mRNA and protein expression, in CD4 cells from mesenteric lymph nodes in the jejunum with OIT.²⁹ Besides Tregs, a population of newly described IgG4 producing IL10 secreting, regulatory B-cells (Bregs) have been observed in subjects on venom immunotherapy and may have a role in other forms of immunotherapy.³⁰

Subcutaneous immunotherapy produces humoral changes. Initially, there is an early increase in specific IgE, but blunting of the seasonal rise in specific IgE, and is followed by a gradual decline in serum specific IgE over months to years of immunotherapy.³¹ Immunotherapy results in a 10–100-fold increase in allergen-specific antibodies in IgG1 and IgG4 subclasses, particularly IgG4.

A vital issue in food allergen immunotherapy is the development of sustained unresponsiveness, a way station for the development of true immunological tolerance. A limited

number of participants in OIT, SLIT and EPIT trials develop sustained unresponsiveness.^{14,17,32} Individuals who haven't developed this state, have a rapid loss of desensitization with the return of basophil reactivity whereas those who develop sustained unresponsiveness do not react for prolonged periods and continue to have suppressed basophil reactivity.^{32,33} The real locus of sustained unresponsiveness is not known. Subjects who fail to develop sustained unresponsiveness have higher starting specific IgE levels, suggesting a humoral component.³³ The use of anti-IgE therapy can decrease basophil reactivity but does not improve rates of sustained unresponsiveness.³⁴

The diversity of allergen immunotherapy options

Immunotherapy has evolved beyond SCIT to a panoply of routes and allergen preparations including OIT, SLIT, EPIT, and peptide-based immunotherapy. (See Figure 1) While the bulk of immunotherapy is based on native extracts, allergoids³⁵ and recombinant allergens³⁶ have been trialed. Modified allergens can be associated with reduced risk of reactions.³⁵ Acetone precipitated dog extracts have increased the concentration of major allergens compared to native allergen extracts and consequently achieved better desensitization.³⁷ Peptide Immunotherapy bypasses IgE mediated mast cell activation and induces Tregs and IL-10 mediated immunological tolerance.^{38,39}

Summary of evidence supporting subcutaneous immunotherapy for inhalant allergies

Translating research involving Immunotherapy into clinical practice is challenging. Subcutaneous immunotherapy as is practiced in the United States often comprises multiple allergens in allergen mixes reflecting polysensitization commonly seen in clinical practice. Studies, however, are confined to single allergens. Aggregated evidence from meta-analyses^{40,41} and expert opinion form the basis of guideline documents for subcutaneous immunotherapy.³ However, the quality of the included studies limits meta-analyses. Differences in outcome measures, duration of therapy, allergens and treatment regimens limit comparisons. The Cochrane review, suggests that aero-allergen SCIT lowers symptom and medication scores.⁴² Unlike other meta-analyses and systematic reviews, it calculates statistics based on studies meeting their inclusion criteria but also cite data from studies that did not meet inclusion criteria but had significant results. Their principal conclusions are supported by the statistical analysis and the majority of rejected trials as well. The AAAAI/ACAAI Practice Parameters now endorse SLIT Immunotherapy for the treatment of aeroallergies.⁴³ Several commercial products are available for co-seasonal and year-round use.

Food allergy – oral and sublingual immunotherapy

Investigations into the treatment of food allergies have lagged the treatment of aeroallergies. However, the explosion in the incidence and awareness of food allergies has led to a push for immunotherapy. Initial SCIT studies had high rates of reactions.^{9,10} Many things set food allergy immunotherapy

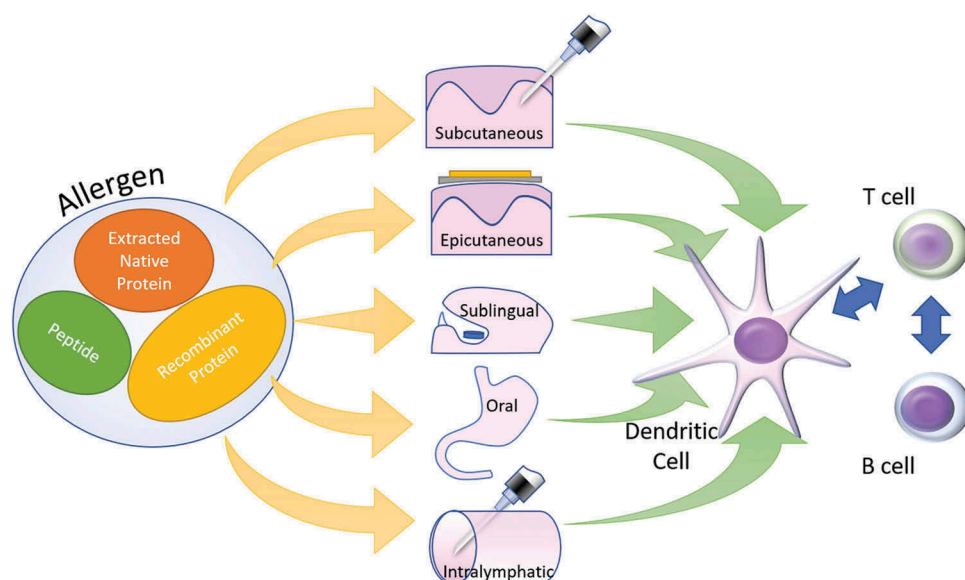


Figure 1. Different allergen forms can be administered through a variety of routes eventually leading to similar immunological changes. Subcutaneous – Subcutaneous Immunotherapy, Sublingual – Sublingual Immunotherapy, Oral – Oral Immunotherapy, Epicutaneous – Epicutaneous Immunotherapy and Intralymphatic – Intralymphatic immunotherapy.

apart from respiratory allergies or venom allergies. The route of exposure is different (oral vs, inhaled or parenteral). The dose per exposure, for food, is several orders of magnitude larger. Partial desensitization or tolerance (i.e., Mild food allergic reactions) is not an acceptable outcome. Strict avoidance is possible but has nutritional, social and financial consequences. Though there are few head-to-head comparisons, OIT is the most efficacious therapy for food allergy. OIT has the highest rates of treatment-associated adverse reactions. Most studies address, peanut,^{28,32,33,44–49} milk^{50–53} and egg allergies.^{51,54–58} Limited studies have been carried out with wheat,^{59,60} multi-food immunotherapy⁶¹ and as discussed elsewhere pollen-food allergy syndrome (PFAS). OIT and SLIT comparison studies have demonstrated higher rates of desensitization and adverse effects with OIT.³² In designing oral immunotherapy for food allergies it is important to define the objective – the ability to consume conventional portions of the food or prevention of allergic reactions due to small or accidental exposures. There hasn't been a systematic effort to qualify the objectives and outcomes in these terms. Most milk and egg trials target the ability of the patient to consume conventional portions of the food as the goal of treatment. All oral immunotherapy trials for all foods published to date cumulatively include approximately 1500 subjects. Several major food allergens are yet to be in clinical trials. This lack of data despite enormous efforts, across a large number of sites and at great cost is the single greatest limitation to our understanding of oral immunotherapy.

OIT proceeds through three phases – initial rapid up-dosing (especially with omalizumab as premedication), a slow escalation phase and a maintenance phase. Studies assess desensitization (a food challenge while on daily dosing) and sustained unresponsiveness (food challenge after a period of abstinence after achieving desensitization). The duration maintenance therapy and the duration of abstinence before a sustained unresponsiveness challenge vary amongst studies.

Oral immunotherapy for peanut (Table 1A) serves as a prototype for the study of OIT for anaphylactic food allergies. However, the generalizability of these findings will not be known until a large cohort of foods has been studied. Desensitization rates have varied from 60 to 80%, and this has been the case for most foods. Early studies used a large goal dose for desensitization (about 4000mg) However, data suggests that lower maintenance doses such as 1000mg or 300mg of peanut protein may achieve higher rates of desensitization, with lower rates of adverse effects and similar ability to tolerate more substantial amounts upon challenge. High rates of desensitization and sustained unresponsiveness were associated with the use of lactobacillus in combination with peanut.⁴⁵ However, the trial involved younger subjects (mean age about six years), and sustained unresponsiveness challenges were carried an earlier than other studies (2–5 weeks)

In milk and egg allergies, the introduction of baked milk or egg products in baked milk or baked egg-tolerant subjects respectively has revolutionized care. It is associated with high rates of resolution of allergy to unheated milk and egg.⁶⁴ Paradoxically immunotherapy with baked milk in baked milk-reactive subjects has had dismal results due to high rates of reactions.⁶⁵ Milk powder and fluid milk have been used for milk oral immunotherapy. (Table 1B) Fluid milk doses have varied from 100 to 200 ml. The limited number of trials and small populations preclude systematic analysis of the best strategy. Likewise, egg oral immunotherapy trials (Table 1C) have varied in dose used for desensitization and maintenance. Most clinical trials use dried or lyophilized egg white as agent for desensitization and challenge to one egg. However, egg clinical trials show the greatest variability in how maintenance dosing is conducted; using raw, undercooked or cooked egg; 1/3rd to a whole egg in quantity and daily to twice weekly dosing intervals. As with other foods, there is a paucity head to head comparisons. However, high rates of desensitization (80–94%) have been

Table 1A. Peanut oral immunotherapy.

Study	Food allergen	N	Age Range	Study design	Maintenance Dose	Desensitization Outcome	Long-term results	Adverse events
Hofmann et al. 2009 ⁶² Jones et al. 2009 ⁴⁶	Peanut	39	1–16 years	Open-label, not controlled.	300–1800 mg	71% reached 300 mg daily dose.	69% (27/39) passed OFC at 1800 mg (total 3.9 g).	Escalation: 92% symptomatic, 15% needed epinephrine. 3.7 % of 14,773 doses during up-dosing or maintenance, mostly minor.
Blumchen et al. 2010 ⁴⁷	Peanut	23	3–14 years	Open label, not controlled.	2500 mg	61% (14/23) reached 500 mg daily for 8 weeks.	After 2 weeks off OIT, 14/23 (100% who reached maintenance) tolerated 500 mg in DBPCFC, 17% tolerated 4 g.	Escalation: subjective symptoms with 25/317 (7.9 %) doses. Reactions with 160/6137 (2.6 %) of buildup/maintenance doses.
Varshney et al. 2011 ⁴⁸	Peanut	28 (2:1 OIT: placebo)	1–16 years	RCT, placebo-controlled, double-blind.	Up to 4000 mg daily	80% on OIT tolerated 4000 mg daily for 1 year, 80% on OIT passed 5000 mg DBPCFC after 1 year.	N/A	Escalation: 47% (9/19) of OIT had symptoms, 2 required epinephrine. Reactions with 1.2 % of buildup/maintenance doses.
Vickery et al. 2014 ³³	Peanut	39	1–16 years	Open label, not controlled (enrolled patients from ⁵³ and ²⁴).	300 mg OR 1800 mg (if passed 3900mg OFC), then 4000 mg daily	66% (26/39) reached 4000 mg daily.	After 4 weeks off OIT, 31% (12/39) tolerated 5000 mg OFC and open dose.	15% withdrew for allergic side effects.
Anagnostou et al. 2014 ⁶³	Peanut	99	7–16 years	RCT, placebo-controlled, double-blind. (OIT/placebo), control group crossed over to active OIT.	800 mg	Phase 1: 84% OIT, 0 % placebo passed 1400 mg DBPCFC. Phase 2: placebo patients switched to OIT, 54% passed the same DBPCFC.	Phase 1: 62% OIT, 0 % placebo tolerated 800 mg daily at 26 weeks. Phase 2: placebo patients switched to OIT, 91% tolerated 800 mg daily at 26 weeks. QoL improved in both groups.	Up-dosing: Mild adverse events in 6.3 % of doses. 1 patient received epinephrine for 2 separate reactions.
Narisety et al. 2015 ³²	Peanut	21	7–13 years	RCT, parallel intervention, double blind (1:1 OIT/SLIT).	2000 mg OIT, 3.7 mg SLIT	50% SLIT, 45% OIT passed OFC 10g at 6–12 months.	Sustained unresponsiveness: 50% (2/4) OIT, 20% (1/5) SLIT passed repeat OFC 10 g after 4 weeks peanut avoidance.	Reactions to 43% of OIT, 9 % of SLIT. 5 OIT reactions required epinephrine. 1 OIT patient developed eosinophilic esophagitis and withdrew.
Syed et al. 2014 ²⁸	Peanut	43	5–45 years	RCT, open label (OIT/peanut avoidance).	4000 mg	20/23 OIT, 0 % (0/20) control desensitized (passed DBPCFC 4 g) at 24 months.	Sustained unresponsiveness: after off peanut OIT for 3 months, 30% (7/23) passed DBPCFC at 27 months. After off peanut for 6 months, 13% (3/7 SU, 3/23 ITT) remained tolerant (passed DBPCFC).	N/A
Tang et al. 2015 ⁴⁵	Peanut and lactobacillus	62	1–10 years	RCT, placebo controlled, double blind (Probiotic?+? OIT vs. placebo?+? placebo OIT).	2000 mg	90% probiotic?+? OIT, 7 % placebo desensitized (DBPCFC).	Sustained unresponsiveness: 82% probiotic, 4 % placebo (DBPCFC 2–5 weeks after probiotic discontinued).	45% probiotic, 32% placebo patients had ?1 severe adverse event. 1 patient had anaphylaxis to probiotic.

Table 1B. Milk oral immunotherapy.

Study	Food allergen	N	Age Range	Study design	Maintenance Dose	Desensitization Outcome	Long-term results	Adverse events
Meglio et al. 2008 ⁵⁰ Meglio et al. 2004 ⁶⁶	Fluid milk	21	5–10 years	Open-label, not controlled.	Up to 200 mL milk or milk-containing foods	71% (15/21) tolerated 200 mL, 14% (3/21) reached 40–80 mL. Mean duration 201 days (range 183–234 days).	After 48–51 months, 70% in follow-up (14/20) tolerating some milk. 43% (9/14) taking milk ad lib.	Desensitization: 3/21 (14%) had mild symptoms so discontinued; 3/21 (14%) had symptoms so took a lower daily dose. Follow-up: no reactions requiring epinephrine.
Pajno et al. 2013 ⁶⁷	Fluid milk	32	4–13 years	RCT, open-label, comparison of maintenance regimens.	150–200 mL daily (group A) or twice weekly (group B) and milk ad lib	100% in both groups continued maintenance, a similar frequency of allergic symptoms.	N/A	8/15 in group A and 9/15 in group B had recorded events.
Salmivesi et al. 2013 ⁶⁸ Paassilta et al. 2016 ⁶⁹	Fluid milk	24	6–14 years (from initial study 2013)	RCT, placebo control, double-blind, open-label crossover.	200 mL (6400 mg) daily	89% (16/18) OIT desensitized, not assessed in placebo. 100% (10/10) control patients desensitized in the crossover to open-label OIT. At 12 months, 13/18 (72%) of OIT taking 6400 mg CM daily.	At 3 years, 85% OIT (including original OIT and crossover) tolerating milk daily, 58% at 7 years.	Desensitization: 100% of OIT, 80% crossover, 63% of placebo patients reported symptoms. 6–12 months maintenance: 62% had symptoms, none severe; 50% at 3 years, 19% at 7 years.
Longo et al. 2008 ⁵²	Fluid milk, then foods with milk.	60	5–17 years	RCT, open-label, (OIT/milk-free diet).	150 mL (4800 mg) then milk containing foods	At 1 year, 37% (11/30) OIT fully desensitized to 150 ml daily, 53% (16/30) OIT partially (5–15), 0 % control passed DBPCFC.	N/A	Desensitization: 4 patients required IM epinephrine. Home dosing: 2 needed epinephrine. 20% control had an adverse reaction to accidental milk.
Morrisset et al. 2007 ⁵¹	Fluid milk.	57	1–6 years	RCT, open-label (OIT/milk-free diet).	(OIT/milk-free diet).	After 6 months, 11% OIT and 40% control had positive SBPCFC (< 200 mL).	N/A	Reactions in 3 OIT, 0 control patients.
Martorell et al. 2011 ⁷⁰	Fluid milk.	60	2–3.5 years	RCT, open label. (OIT/milk-free diet).	200 mL (6400 mg) daily	At 1 year, 90% milk OIT tolerant to 200 mL daily, 13% (3/23) of control passed DBPCFC. RR 7.7 for milk tolerance in OIT vs. placebo.	N/A	80% OIT patients had ?1 reaction, all mild-moderate. Reactions with 15% (114/738) doses.
Skpirak et al. 2008 ⁵³ Narisety et al. 2009 ⁷¹ Keet et al. 2013 ⁷²	Milk powder.	20	6–21 years	RCT, placebo controlled, double blind, then open-label crossover. (2:1 OIT/placebo)	500 mg daily for 13 weeks, then 7000 mg (if pass 8g DBPCFC).	23% OIT, 0 % placebo, 67% crossover OIT passed DBPCFC (8000 mg); 92% active OIT, 0% placebo, 83% of cross-over OIT tolerant to ?2540 mg.	At 5 years: 19% of 13 in follow-up tolerating unlimited milk, 31% 1 serving/day, 34% limited amounts, 16% no milk/avoiding completely.	Desensitization: reactions to 45% of 2437 doses in OIT, 11% of 1193 in placebo patients. 4 patients required epinephrine. Follow-up open-label OIT: reactions in 17% of 2465 home doses.
Goldberg et al. 2015 ⁶⁵	Baked milk.	15	6–12 years	Open-label, uncontrolled.	Baked milk products daily as tolerated.	21% tolerated 1.3 g baked milk OFC. Maximum tolerated dose 900 mg unheated milk.	N/A	8/15 did not complete desensitization due to IgE-mediated reactions. 2 had anaphylaxis requiring epinephrine with home doses.

achieved in many trials.^{54,55,77,78,80} Only two clinical trials of wheat OIT (Table 1D), with a total 24 subjects have been published. These serve as little more than proof of concept currently.

Two related limitations in oral immunotherapy have been the need for indefinite daily maintenance dosing and low rates of “sustained unresponsiveness.” It is the maintenance of desensitization despite withholding the food. The rates across foods and studies have been low. Therefore, most subjects need to consume the food daily to maintain desensitization. A small trial with 17 subjects suggests that intermittent twice weekly dosing may be as effective as a daily dosing regimen

after desensitization. The incidence of allergic reactions was the same.⁶⁷ However, this is going to need further validation. Longer duration of therapy is associated with higher rates of sustained unresponsiveness.⁸⁶ This would be particularly significant because many allergic subjects have an aversion to the food. Even in the absence of food aversion, daily dosing is onerous. Sustained unresponsiveness has been assessed after 3–6 weeks of withdrawal. We do not know what would happen if the subjects did not consume the food for more extended periods of time. The only way to assess sustained unresponsiveness is the withdrawal of the food and challenge which carries the risk of anaphylaxis. There is a desperate

need for a surrogate marker. IgG4 levels and BAT responses are potential markers but will need extensive validation for each food.⁸⁶

Determinants of successful desensitization and sustained unresponsiveness are not known. A long-term study suggests that the starting specific IgE level and peak IgE level during desensitization may determine the long-term sustained unresponsiveness.³³ Trials of very young children with peanut and milk allergies has demonstrated that in this age group OIT was associated with higher rates of desensitization and was associated with lower rates of reactions.^{51,87} When coupled with studies of the primary and secondary prevention of peanut allergy by the early introduction of peanut⁸⁸ it suggests that there may be a continuum of immunological plasticity, which applies to peanut allergy and possibly other foods. This degree of plasticity might decrease with age resulting in lower desensitization rates with age. Studies are needed to determine the generalizability to other foods and different forms of immunotherapy.

Pollen food allergy syndrome

Pollen-food allergy syndrome (PFAS) is caused by anti-pollen antibodies cross-reacting with antigenically similar protein in several plant-based foods principally, fruits, vegetables, and nuts. It is unique because both the inciting pollen and the secondary food are targets for immunotherapy. The most commonly studied is the birch-apple allergy caused by cross-reactivity between, the PR10 family proteins, Bet v1 in birch and Mal d 1 in apple. Seven studies have investigated three modalities of therapy, subcutaneous immunotherapy using birch pollen, SLIT using birch pollen and OIT using raw apple. Fundamental limitations in these studies include the lack of uniformity in intervention, the lack of randomization and placebo controls and the frequent use of open challenges with apple. These studies have varied in outcomes. (see Table 2) Unlike prior studies, Hansen et al. were the first to perform a double-blind, double-dummy placebo-controlled study.⁹³ However, this study ultimately suffered from a small number of subjects who had challenge confirmed PFAS in each group and threshold-dose insensitive challenge protocol.⁹³ Likewise, the only study using apple DBPFC for diagnosis did not use a graded challenge but instead relied on a VAS score for symptoms with a fixed dose of apple. This method likely impaired their ability to measure changes in the degree of apple sensitivity as determined by threshold dose for eliciting symptoms.⁹⁴

The lack of improvement in apple PFAS with Birch pollen SLIT is explained by lack change IgE, IgG4 or T-cell responses with the intervention despite concomitant amelioration in Bet v1 specific responses.⁹⁴ However, a recent study by the same group using Mal d1 SLIT demonstrated significant improvement in Mal d1-specific sublingual challenge, IgE, and IgG4 responses.⁹³ OIT with apple and Mal d1 SLIT suggest that though the pathogenesis of birch-apple PFAS involves birch pollen sensitization desensitization with apple rather than Birch may be a superior approach to therapy.

Epicutaneous immunotherapy

Epicutaneous immunotherapy involves transdermal administration of allergen under an occlusive dressing that promotes allergen absorption. Three allergens, grass pollen,⁹⁸ milk⁹⁹ and peanut^{14,100} have been studied using this modality. Skin preparation before application of the patch has also varied – no preparation, abrasion with a foot file and tape stripping. In a head-to-head comparison of the two skin prep methods, the more aggressive abrasion method was associated with a higher risk of systemic allergic reactions.¹⁰¹ Mouse studies have demonstrated the uptake of allergen by dendritic cells after prolonged application and resulting immunological changes.¹⁰² Trials involving milk and peanut without skin preparation, use a different proprietary patch, precluding direct comparison, however, the rates of systemic reactions were low.¹⁰⁰ Comparison of different protein doses in patches shows a dose effect¹⁰⁰ Local reactions, most commonly patch site pruritus, was virtually universal.^{99–101,103} All trials compare efficacy compared to placebo. There are no trials comparing grass pollen EPIT to SCIT or SLIT. Similarly, there no trials comparing milk or peanut EPIT to SLIT or OIT. However, comparison of completed studies shows a clear distinction in the rates of systemic adverse effects favoring EPIT. Peanut EPIT resulted in desensitization in 48% subjects (passing a 1000mg of peanut food challenge or 10-fold increase in maximum tolerated dose). It was more efficacious in 4–11 age group than in older subjects.¹⁰⁰ In comparison, the largest peanut OIT trial which used similar entry criteria resulted in desensitization in 62% of subjects (passing a 1400mg challenge).⁴⁴ Excluding subjects on EPIT who did not pass a 1000mg challenge but did have tenfold increase in threshold further widens the gap between OIT and EPIT, suggesting OIT is likely more efficacious than EPIT.⁴⁴ Like OIT, EPIT outcomes appear to be better in younger subjects.⁸⁷

Intralymphatic immunotherapy

Intralymphatic immunotherapy (ILIT) is a new modality of injectable allergen immunotherapy. It involves the repeated injection of allergen directly into the lymph node. The dose of allergen is lower than in subcutaneous therapy.¹⁰⁴ It involves fewer injections and has fewer side effects.¹⁰⁴ Injections were performed under ultrasound guidance.¹⁰⁴ Tolerance achieved was long lasting and equivalent to SCIT.¹⁰⁴ It has been used for Grass pollen,¹⁰⁴ Birch pollen,^{105,106} Cat,^{103,107,108} Dog,^{107,108} Dust mite.^{107,108} (Key studies are summarized in Table 3) However, unlike SCIT expertise and studies are confined to a limited number of centers. Though the results are encouraging and carry the promise of improved compliance, studies are limited and without replication. Another limitation is that the studies are randomized but open-label. Ideally, SCIT and ILIT comparison studies should be double-blind double-dummy. However, this may not be ethically feasible. The methodology and optimal injection schedule are open to



Table 1C. Egg oral immunotherapy.

Study	Food allergen	N	Age Range	Study design	Maintenance Dose	Desensitization Outcome	Long-term results	Adverse events
Staden et al. 2007 ³	Lyophilized egg white.	45 (11 egg avoidance control, 24 other foods)	0.6–12 years.	RCT, open-label (OIT/egg-free diet).	1600 mg egg daily plus deliberate intake	64% (16/25) fully or partially desensitized (milk and egg OIT assessed jointly), 35% of control later tolerant.	36% of egg OIT had sustained tolerance (passed DBPCFC after stopping OIT for 2 months) vs. 35% of controls.	All OIT patients had some symptoms; 20% (4/20) of control had symptoms with accidental ingestion.
Buchanan et al. 2007 ⁴ Burks et al. 2008 ⁵ Vickery et al. 2010 ⁶	Dried egg white.	21 (initially 7)	1–16 years.	Open-label, no control.	300 mg daily	90% of 21 reached 300 mg daily, 57% of 7 initial patients passed DBPCFC for 8 g egg.	26% of 7 had sustained unresponsiveness (pass DBPCFC after stopping OIT 3–4 months).	Up-dosing: 1/7 initial patients reacted. Maintenance: no reactions in 7/7 patients, 3 did not react to accidental egg exposure.
Burks et al. 2012 ⁷	Dried egg white.	8	3–13 years.	Open-label, no control (same protocol as ^{3,2} different patients).	300 mg–3.6 g daily	75% (6/8) OIT reached maintenance. After 4 months, 62.5% (5/5 tested) passed OFC to 3.9 g egg, 1 not tested. After median 34 months, 75% (6/6 who reached maintenance) passed DBPCFC.	Sustained unresponsiveness: After 1 month off OIT, 75% (6/6) passed 2nd DBPCFC. All incorporated egg into the diet.	Buildup/4/6 who completed had reactions. Maintenance: no reactions. No reactions requiring epinephrine.
Fuentes-Aparicio et al. 2013 ⁵⁷	Dried egg white.	55	5–11 years.	RCT, placebo-controlled, double-blind (OIT/ placebo).	2 g daily (approx 1/3 egg)	At 10 months, 55% on OIT, 0% on placebo desensitized (OFC 5 g). At 22 months, 75% on OIT desensitized (OFC 10 g).	Sustained unresponsiveness: at 24 months, after off egg for 4–6 weeks, 27.5% OIT passed 10 g OFC. Not tested in placebo.	Desensitization: adverse events with 25% of 11,680 OIT doses, 4% 4018 placebo doses. No adverse events in patients with SU consuming egg ad lib.
Caminiti et al. 2015 ⁵⁵	Dried egg white.	72	5–15 years.	RCT, open-label (OIT/egg-free diet).	10 g daily	After 1 month: 92% (37/40) OIT, 22% (7/32) control desensitized to or tolerated egg.	54% (20/37) OIT passed OFC to 10 g raw egg and introduced into the diet.	Desensitization: 21/37 had symptoms. 5 reactions required epinephrine. 1 patient withdrew with eosinophilic esophagitis.
Escudero et al. 2015 ⁵⁴	Dried egg white.	31	4–10 years.	RCT, placebo control, double blind (OIT/ placebo).	1 cooked egg twice weekly	94% (16/17) OIT, 0% control (0/14) desensitized (passed DBPCFC 4 g).	After 6 months of egg diet then 3 months avoidance, 29% (5/17) OIT, 7% (1/14) placebo had sustained tolerance (DBPCFC).	Desensitization: 3 patients had adverse events, 1 discontinued OIT. Egg containing diet phase: 2 patients had symptoms.
Ruiz-García et al. 2012 ⁷	Dried egg white – OVIDES NMI kit.	61	5–17 years.	RCT, open label (OIT/egg avoidance).	1 undercooked egg every 2 days and egg ad lib	93% (28/30) on OIT desensitized. At 4 months, 37% OIT passed DBPCFC 2808 mg (11/25 challenged); 1% control passed DBPCFC (1/31 challenged).	After 1 month avoidance, 37% OIT, 3% control had sustained unresponsiveness (DBPCFC 2808 mg).	145 reactions in OIT, 14% in dose escalation, 54% in buildup, 31% in maintenance. 70% of patients had 71 reaction. 1 required epinephrine.
Tortajada et al. 2012 ⁸	Raw e ⁷⁹ gg.	17	6–38 years.	Open-label, not controlled.	3 eggs per week	82% (14/17) reached maintenance dose and tolerated for 9 months.	N/A	70.5% (12/17) symptomatic during OIT. 3 required epinephrine during up-dosing or at home.
Ojeda et al. 2012 ⁸⁰	Raw egg.	19	3–14 years.	Open-label, prospective, no control.	0.5 egg twice weekly for several months, then 1 egg weekly.	89% (17/19) desensitized to 30 g, no control.	N/A	2/19 had anaphylactic reactions requiring epinephrine, discontinued OIT.
		31	6–15 years.	Open-label, controlled (not randomized).	0.5 raw (pasteurized) egg 3 times weekly.	81% (25/31) in OIT reached maximum dose, 74% passed OFC for 1 unpasteurized egg. 3% partially desensitized, 10% not desensitized but increased reactivity threshold. Control outcomes N/A.	N/A	74% of OIT patients reacted. 180 adverse events, 86% were grades 1–2. 4 required epinephrine at home.

(Continued)

Table 1C. (Continued).

Study	Food allergen	N	Age Range	Study design	Maintenance Dose	Desensitization Outcome	Long-term results	Adverse events
Lertran et al. 2012 ⁷⁹	Raw egg.	17	4–14 years.	Open label, not controlled.	1 egg omelet (or fraction tolerated)	35% (6–17) fully desensitized.	N/A	Up-dosing: 4 reactions reported. Maintenance: 47% (8/17) had mild symptoms, 1 patient required emergency room visit.
Meglio et al. 2013 ⁸¹	Raw egg.	20	4–14 years.	RCT, open label (OIT/egg-free diet)	Raw egg or food with eggs 3 times/week	80% (8/10) OIT, 0% control tolerated 25 mL (13.6 g) daily; 10% OIT had partial tolerance.	N/A	7/10 OIT had some symptoms during desensitization, 5 resolved, 1 tolerated limited amounts, 1 discontinued.
Dello Iacono et al. 2012 ⁸²	Raw egg.	20	5–11 years.	RCT, open label (OIT/egg free diet)	10–40 mL egg daily.	After 6 months, 0% (0/10) in egg OIT desensitized to 1 raw egg (40 mL); 90% partially desensitized (tolerated 10–40 mL). 0% control passed DBPCFC.	N/A	100% OIT patients had 53 total adverse events during desensitization. 24% were grades 1–2. None required epinephrine. 30% control patients had 5 total adverse reactions from accidental egg ingestion.
Vazquez-Ortiz et al. 2014 ⁵⁶	Raw egg.	82	5–18 years	Non-randomized, controlled, open label. (Parallel OIT/avoidance).	1 egg twice weekly	Desensitization: 80% (40/50) OIT reached 2 eggs/week.	At 12 months of maintenance, 56% (28/50) OIT passed OFC for 1 raw egg, 8% (4/50) of OIT partially desensitized (tolerated 0.8–1.7 g). 16% (5/32) of control spontaneously developed tolerance at 12 months (passed DBPCFC).	Dose-related reactions in 45/50 (90%) and 7.6% (1024/13,551) of egg-OIT doses. 61% were mild (grades 1–2), 18 required epinephrine (in 13 patients).
Garcia-Rodriguez et al. 2011 ⁸³	Raw and cooked egg.	23	5–17 years.	Open-label, no control.	1 cooked egg daily for 3 months, then every 2–3 days for 12 months.	87% (20/23) OIT desensitized (tolerated 1 cooked egg), 61% (14/23) within 5 days.	2 patients (9%) who failed initial desensitization switched to slow protocol, desensitized in 60–80 days.	78% (18/23) had an allergic reaction during desensitization. Total reactions 55, 35 mild and 20 moderate. 2 patients had reactions during daily cooked egg maintenance.
Morrisset et al. 2007 ⁵¹	Hard-boiled egg.	84	1–8 years.	RCT (OIT/egg-free diet). (Both egg and milk in the same study).	Daily intake including creamy desserts and flan	69% OIT (32/49), 51% (18/35) control patients desensitized and passed SBPCFC (7 g raw egg white).	N/A	N/A
Itoh et al. 2010 ⁸⁴	Scrambled egg.	6	7–12 years.	Open-label, no control.	1 heated whole egg twice weekly	100% desensitized to 1 scrambled egg (60 g), meantime 12 days, 50% also passed OFC 1 g powdered egg after 9–12 months maintenance.	16–21 months after rush desensitization, all continuing to eat 1 heated egg twice weekly.	100% had allergic symptoms during desensitization. None required epinephrine.
Leonard et al. 2012 ⁸⁵	Baked egg.	70	0.5–25 years.	Prospective, historical control.	1–3 servings of baked egg product daily	53% (42/70) tolerated partially cooked egg (OFC 6.5 g). 36% (28/70) did not but continued baked egg.	Retrospective: 28% (13/47) tolerant to partially cooked egg, 13% (6/47) to baked egg, 59% (28/47) avoidant.	No reactions to baked egg consumption, 1 patient reacted to accidental exposure to unbaked egg.

Table 1D. Wheat oral immunotherapy.

Study	Food allergen	N	Age Range	Study design	Maintenance Dose	Desensitization Outcome	Long-term results	Adverse events
Sato et al. 2015 ⁶⁰	Wheat noodles.	18	5–13 years.	Open-label, historical controls.	5.2 g daily	89% reached maintenance and tolerating 5.2 g daily.	After 2 years off OIT, 61% passed OFC, 9.1% in historical control.	Rush/dose escalation: 26% (42/143) doses resulted in symptoms; none required epinephrine. Maintenance: 6.8% (486/5778) doses with symptoms, 1 required epinephrine.
Rodriguez del Rio et al. 2014 ⁵⁹	Wheat porridge or pasta.	6	5–11 years	Open-label, no control.	100 g daily	83% (5/6) completed maintenance, 80% also tolerant of oat.	N/A	Up-dosing: 6 adverse reactions in 2 patients (6.25%, 6/96 doses). Maintenance: 1 patient had exercise-induced symptoms.

Table 2. Therapy for pollen food syndrome.

Author/Year of publication	Food	Pollen	Route	Intervention	Age (years)	N	Outcome	Limitations
Moller C 1989 ⁸⁹	Apple	Birch	SCIT	SCIT (Birch Pollen) vs OIT (Birch pollen)	21–47	15	neither intervention improved food sensitivity significantly	
Herrmann D et al. 1995 ⁹⁰	Apple	Birch	SCIT	Observational study	Adults	20	improvement in 56% of patients	not a controlled study
Asero et al. 1998 ⁹¹	Apple	Birch	SCIT	SCIT (aluminum hydroxide-adsorbed birch pollen extracts) vs. no treatment	Mean age 34.4 years	SCIT: 49 Controls 22	treatment group 45% complete resolution, 39% partial reduction, 16% unchanged control group 0% unchanged	No information about food challenge methodology, age or gender of subject allocation
Bucher et al. 2004 ⁹²	Apple/Hazelnut	Birch	SCIT	SCIT (birch-hazel-alder± ash pollen extract) vs no treatment		SCIT: 15 Controls: 12	87% of the treatment group and 8% of control group improved tolerance after treatment	no sham group; no randomization
Hansen et al. 2004 ⁹³	Apple	Birch	SCIT & SLIT	Double-blind, double-dummy placebo-controlled Birch pollen SCIT and Birch pollen SLIT	mean 32	M: 42, F: 32. Actual number of challenge confirmed PFAS subjects SCIT 10, SLIT 4, Placebo 10	No significant change in number of subjects who passed a food challenge	a small number of subjects for SLIT, Two-step challenge 10g of apple and whole apple not sensitive for detecting changes in tolerated dose.
Kinaciyani et al. 2007 ⁹⁴	Apple	Birch	SLIT	Birch pollen SLIT	21–47 mean 33.2	M: 5 F:15	No significant change in DBPFC (VAS) score	DBPFC used VAS to fixed dose not threshold dose of reactivity
Mauro et al. 2011 ⁹⁵	Apple	Birch	SCIT & SLIT	Birch pollen SCIT and SLIT	18–60 mean 37.8	SLIT (M: 11, F: 9) SCIT (M: 10, F: 10)	25% of SCIT and 14.2% of SLIT complete tolerance, 37.5% of SCIT and 28.6% of SLIT developed increase in the provocative dose	No Placebo group
Kopac et al. 2012 ⁹⁶	Apple	Birch	OIT	Apple OIT	18–61	OIT (M: 9, F:18); Control (M: 3, F: 10)	17/27 subjects in treatment and 0/13 subjects in control achieved desensitization	Open challenges
Kinaciyani et al. 2017 ⁹⁷	Apple	Birch	SLIT	Bet v1 SLIT, Mal d1 SLIT or Placebo	18–65	Bet v1 SLIT: 20, Mal d1 SLIT: 20 and Placebo: 20	Mal d1 SLIT performed significantly better than Bet v1 SLIT or placebo in Mal d 1 sublingual challenge	No Apple challenge

SCIT – Subcutaneous Immunotherapy, SLIT – Sublingual Immunotherapy, OIT – Oral Immunotherapy, Bet v1 SLIT – SLIT using the recombinant birch pollen protein Bet v1, Mal d1 SLIT – SLIT using the recombinant apple protein Mal d1

debate.¹⁰⁹ At this time four clinical trials of ILIT are recruiting or active. (ClinicalTrials.gov)

Combination immunotherapy with biologics

Anti-IgE therapy has found application in settings where there are high rates of reactions to allergen immunotherapy – food OIT, aeroallergen rush immunotherapy, and venom immunotherapy. Compared to OIT alone, the addition of Omalizumab significantly reduces the risk of mild and severe reactions.^{49,110} (See Table 4) Anti-IgE monotherapy with a different biologic (TNX-901) suggests that it increases the threshold of reactivity to the food allergen.¹¹⁶ The

combination with OIT appears to confirm this benefit, facilitating faster desensitization using rush protocols with low rates of adverse reactions.^{49,111,112,114} Carefully designed placebo-controlled studies suggest that it does not improve the overall rate of desensitization over OIT alone. A case series suggested that it enables desensitization in subjects who have previously failed immunotherapy with conventional protocols.¹¹⁵ However, another small case series opposes this. Here patients refractory to OIT were successfully desensitized on Omalizumab but reverted to allergy after discontinuing anti-IgE therapy.¹¹³ Mechanistic studies confirm a transient decrease in basophil reactivity but no change in the number of Tregs.³⁴ Such findings are mirrored in aeroallergen SCIT where Omalizumab has been shown to decrease

Table 3. Table intralymphatic immunotherapy.

Author/Year	Disease	Allergen	Intervention	Age (years)	Number of Patients	Outcome
Hylander et al. 2016 ¹⁰⁶	Allergic Rhinitis	Birch or grass pollen	Double Blind Randomized Placebo Control	20–54	21 (8 females) (15 controls)	Overall met the primary endpoint of symptom reduction by VAS.
Senti et al. 2008 ¹⁰⁴	Allergic Rhinitis	Grass pollen	ILT vs. SCIT	32 ± 8.7 years	58 ILT (20 females) 54 SCIT	ILT was safe, effective, induced tolerance faster and tolerance was durable
Senti et al. 2012 ¹⁰³	Allergic Rhinitis	Cat (MAT-Fel d 1)	Randomized Placebo controlled	34.6 ± 11.9	12 (8 Females) 8 Placebo	Met the primary end point of increased nasal tolerance
Lee et al. 2015 ¹⁰⁴	Allergic Rhinitis	Dust mite cat, dog	Dust mite, cat and dog ILT	Not reported	10	ILT can rapidly improve rhinitis symptoms, however severe systemic reactions (2) and severe local reaction (1) occurred.

the adverse effect and anaphylaxis related to ragweed rush immunotherapy;¹¹⁷ have a synergistic effect in decreasing seasonal allergic rhinitis symptoms,¹¹⁸ and improve the safety and efficacy of SCIT in allergic asthma.^{119–121} Newer biologics such as Dupilumab (Anti-IL4 Receptor) that have a more direct effect on atopy might have a greater effect on T-cells improving rates of desensitization and are the subjects of ongoing studies.

Encapsulated allergen

Encapsulation of allergen is a new concept in the allergy world but has been studied and used extensively in pharmacology. Nanoparticles or microparticles can be used to enclose allergens. These can then be injected or ingested like other forms of AIT. The chief advantage of encapsulating allergen is that it can help potentially shield the allergen from mast cells. Other benefits may include reduced allergen dose, co-encapsulation of adjuvants, targeting and improved uptake. Encapsulating agents include Liposomes, Virus-Like particles, natural polymers (Chitosan, Dextran)¹²² and synthetic polymers (poly (lactic acid), poly (lactic-co-glycolic acid) (PLGA))

A randomized placebo-controlled trial of Liposome encapsulated dust mite extract was noted to be safe and effective in 55 asthmatics.¹²³ Over 12 months of therapy, medication scores were reduced and healthy days increased. The study documented reduced allergen-specific responses in skin tests and bronchial challenges. The study did not include extensive mechanistic investigations. It did not compare responses (or adverse effects) against conventional subcutaneous immunotherapy.¹²³

In an older study, which aggregated four small studies assessing the cutaneous tolerance of the allergens, the safety of empty and allergen filled liposomes and lastly the safety and efficacy of liposomal therapy. The study concluded that systemic safety was poor.¹²⁴ However, their sample sizes were limited (12 patients in 4 treatment arms). The same trial noted reduced local reaction upon subcutaneous injection. Interestingly, alpha-tocopherol used as an antioxidant in the production of these liposomes caused contact dermatitis in two patients. Despite the promise of lower rates of allergic reactions and extensive development of liposome-based therapies in other fields such as oncology there have been no new trials in this field. It is not clear if this is due to unpublished negative data, lack of funding or lack of interest.

The use of TLR ligands

Besides liposomes, studies have used virus-like particles for allergic rhinitis.^{1,125,126} VLP with CpG, a TLR9 ligand has been used with¹²⁵ and without allergen immunotherapy. In a dose-ranging study of 299 subjects, subjects treated with the highest dose a VLP containing CpG, *without an allergen*, had a significantly reduced combined symptom and medication score.¹²⁶ In a subsequent study, 20 patients underwent open-label therapy with the CpG containing VLP and conventional house dust mite extract in increasing doses over ten weeks. Conjunctival provocation eliciting dose and protective changes IgE and IgG levels improved.¹²⁵ In this study House dust mite was adsorbed on alum which is a confounder. A 2006 study that utilized ragweed conjugated to CpG (without Alum) showed similar promise with significant improvement in symptom scores over two seasons following a single treatment. However, a follow-up phase 2 study for Amb a 1 CpG conjugate was prematurely withdrawn due to no meaningful effect in the first ragweed season (ClinicalTrials.gov NCT00387738). The same company has since launched a clinical trial using a proprietary TLR9 agonist for eosinophilic asthma without a conjugated allergen (ClinicalTrials.gov NCT02898662)

Several animal studies have explored various biodegradable, and non-biodegradable substrates have been studied in animal models. These studies are summarized by Pohlit et al.¹²⁷

Conclusion

Immunotherapy of allergic disorders is entering a new phase. New therapies are available, particularly for food allergies. There is an improved understanding of the immunological changes that occur during immunotherapy. This has led to the development of new formulations and methods of administering immunotherapy. While local effects on the immune milieu occur with sublingual, oral and epicutaneous immunotherapy, the core immunological changes in mast cells, antibody production, and T-cell changes follow a similar pattern. Our understanding of immunology has led to the investigation of adjuvants such as CpG to improve efficacy, anti-IgE therapy to reduce adverse effects, and encapsulating-agents that do both.

The route of allergen administration influences efficacy and adverse effects. As illustrated in the treatment of food allergies there are tradeoffs between efficacy and adverse effects. Further studies would be needed to determine if

Table 4. Oral immunotherapy combined with Anti-IgE therapy.

Study	Food	N	Age	Study Design	Duration of Omalizumab	Target Maintenance Dose	Adverse effects	Desensitization
Nadeau et al. 2011 ¹¹¹	Milk	11	7–17 years	Open-label Single arm, Rush OIT, Total duration of OIT 24 weeks	16 wk (Pre OIT 8 wks)	2000 mg of Milk protein	Total 1.8% (41/2301 doses) Severe 0.1% Epi 3/2301	Desensitization: 82% (9/11) achieved the primary objective of desensitization to a daily dose of 2000 mg milk
Wood et al. 2016 ^{110**}	Milk	57 (O 28, P 29)	7–32 years	Randomized double-blind placebo controlled for Omalizumab. Unblinded milk administration, dose escalation 22–40 weeks, Duration of OIT 28 months	28 months (Pre-OIT 4 months)	3800 mg of Milk Protein	Total during escalation O: 8.5% (442/5226doses) P: 26.1% (1634/6252 doses) Maintenance O: 0.7% (110/15418 doses) P: 14.4% (1983/13745 doses) Epi O: 2 subjects (2 doses) P: 9 subjects (18 doses)	No significant difference in desensitization Omalizumab (24/27; 88.9%) and Placebo (20/28; 71.4%) (P = .18)
Schneider et al. 2013 ¹¹²	Peanut	13	7–15 years	Open-label Single arm, Rush escalation, Duration of OIT: from 12 weeks to 30–32 week	20 weeks (Pre-OIT 12 wks)	4000 mg of peanut protein	Total 2% (72/3502) Epi 3 subjects 5 doses	Desensitization: 92% (12/13) tolerated oral food challenge with 8000 mg peanut flour (about 20 peanuts)
MacGinnitie et al. 2017 ⁴⁹	Peanut	37 (O 29, P 8)	6–19 years	Randomized double-blind placebo-controlled for Omalizumab. Unblinded peanut administration, Open-Label Omalizumab (2 active, 6 control) who failed to reach 250mg of peanut by week 8	19 weeks (Pre-OIT 12 wks)	2000 mg of peanut protein	Epi – 8 subjects (14 reactions) P: 2 subjects (3 reactions), O: 3 (4 reactions), Open-label 7 doses. EoE 8% (3/37) (2 in the active group and 1 in control)	Desensitization to 2000 mg of peanut protein 6 wk after withdrawal of omalizumab. Omalizumab 79% (23/29) Placebo 12.5% (1/8) p < 0.01
Lafuente et al. 2014 ¹¹³	Egg	3	9–10 years	Case series in OIT failures	4–7 months (Pre-OIT 8–12 wks)	1 egg 3 times a week	3/3 patients had recurrence of symptoms 3–4 mo after omalizumab as discontinued	Desensitization to 50 ml egg white: 100% (3/3)
Begin et al. 2014 ¹¹⁴	Multiple	25	4–15 years	Open-label single-arm Phase 1. Up to 5 allergens; rush escalation	16 wk (Pre OIT 8 wks)	4000 mg of protein for each food	rush phase 52% of subjects; escalation 5.7% (13/227 doses) Maintenance 5.3% (401/7530 doses) Epi 1 dose	76% (19/25) tolerated all 6 steps of the initial escalation day (up to 1250 mg of combined food proteins), requiring minimal or no rescue therapy. All subjects achieved 4000mg/food by 9 months
Martorell-Calatayud et al. 2016 ¹¹⁵	Milk, Egg	N = 14 (Egg 9, Milk 5)	3–13 years	Case series Open-label single arm OIT in patients who failed conventional OIT	Variable till 2 months after maintenance dose is achieved (Pre-OIT 9 wks)	Milk: 200ml (6600 mg Milk protein), Egg white 17 ml (1800mg egg protein)	Rush phase: 28% (4/14) Late maintenance (2.5–4 months after discontinuation of omalizumab): 42% (6/14)	Desensitization: 100% after end of induction phase (egg and milk)

Key: O – Omalizumab, P – Placebo, OIT – Oral Immunotherapy, Epi – Epinephrine administration, wk – weeks.

**Key study.

therapy could be started with a low adverse effect modality such as EPIT, and then transitioned to OIT.

Looking ahead, one of the significant challenges confronting us is the patchwork of studies, without uniform methodologies, limited comparability and a shortage of well-

designed head-to-head comparisons of different treatment modalities. The multitude of allergens further exacerbates this problem. As an example, all the studies on pollen-food allergy syndrome use one of two pollen-food combinations, birch-apple or birch-hazelnut. The question remains – are the

results of birch-apple generalizable to all pollen-food combinations, or even to all birch related foods?

The other major challenge is the long duration of therapy. SCIT, SLIT, OIT, and EPIT are all associated with treatments that last months to years, and for food allergies may even be life-long. Intralymphatic therapy promises to shorten the duration of therapy, but the relevant studies are limited to a handful of aeroallergens.

Disclosure of potential conflicts of interest

No potential conflicts of interest were disclosed.

References

- Dhami S, Nurmatov U, Arasi S, Khan T, Asaria M, Zaman H, Agarwal A, Netuveli G, Roberts G, Pfaar O, et al. Allergen immunotherapy for allergic rhinoconjunctivitis: a systematic review and meta-analysis. *Allergy*. 2017;72(11):1597–1631. doi:10.1111/all.13201.
- Turkalj M, Banic I, Anzic SA. A review of clinical efficacy, safety, new developments and adherence to allergen-specific immunotherapy in patients with allergic rhinitis caused by allergy to ragweed pollen (*Ambrosia artemisiifolia*). *Patient Prefer Adherence*. 2017;11:247–257. doi:10.2147/PPA.S70411.
- Cox L, Nelson H, Lockey R, Calabria C, Chacko T, Finegold I, Nelson M, Weber R, Bernstein DI, Blessing-Moore J, et al. Allergen immunotherapy: a practice parameter third update. *J Allergy Clin Immunol*. 2011;127(1, Supplement):S1–S55. doi:10.1016/j.jaci.2010.09.034.
- Fass PT. American academy of otolaryngic allergy endorses the allergen immunotherapy practice parameter. *J Allergy Clin Immunol*. 2008;121(1):269–70; author reply 270. doi:10.1016/j.jaci.2007.10.030.
- Golden DB, Demain J, Freeman T, Graft D, Tankersley M, Tracy J, Blessing-Moore J, Bernstein D, Dinakar C, Greenhawt M, et al. Stinging insect hypersensitivity: A practice parameter update 2016. *Ann Allergy Asthma Immunol*. 2017;118(1):28–54. doi:10.1016/j.anai.2016.10.031.
- Golden DB, Moffitt J, Nicklas RA, Freeman T, Graft DF, Reisman RE, Tracy JM, Bernstein D, Blessing-Moore J, Cox L, et al. Stinging insect hypersensitivity: a practice parameter update 2011. *J Allergy Clin Immunol*. 2011;127(4):852–4 e1-23. doi:10.1016/j.jaci.2011.01.025.
- Joint Task Force on Practice, P., et al. Allergen immunotherapy: a practice parameter second update. *J Allergy Clin Immunol*. 2007;120(3 Suppl):S25–85. doi:10.1016/j.jaci.2007.06.019.
- Cox LS, Hankin C, Lockey R. Allergy immunotherapy adherence and delivery route: location does not matter. *J Allergy Clin Immunol Pract*. 2014;2(2):156–160. doi:10.1016/j.jaip.2014.01.010.
- Oppenheimer JJ, Nelson HS, Bock SA, Christensen F, Leung DY. Treatment of peanut allergy with rush immunotherapy. *J Allergy Clin Immunol*. 1992;90(2):256–262.
- Nelson HS, Lahr J, Rule R, Bock A, Leung D. Treatment of anaphylactic sensitivity to peanuts by immunotherapy with injections of aqueous peanut extract. *J Allergy Clin Immunol*. 1997;99(6 Pt 1):744–751.
- Shamji MH, Durham SR. Mechanisms of allergen immunotherapy for inhaled allergens and predictive biomarkers. *J Allergy Clin Immunol*. 2017;140(6):1485–1498. doi:10.1016/j.jaci.2017.10.010.
- Soyer OU, Akdis M, Akdis CA. Mechanisms of subcutaneous allergen immunotherapy. *Immunol Allergy Clin North Am*. 2011;31(2):175–90, vii–viii. doi:10.1016/j.iac.2011.02.006.
- Berin MC, Shreffler WG. Mechanisms underlying induction of tolerance to foods. *Immunol Allergy Clin North Am*. 2016;36(1):87–102. doi:10.1016/j.iac.2015.08.002.
- Jones SM, Sicherer SH, Burks AW, Leung DYM, Lindblad RW, Dawson P, Henning AK, Berin MC, Chiang D, Vickery BP, et al. Epicutaneous immunotherapy for the treatment of peanut allergy in children and young adults. *J Allergy Clin Immunol*. 2017;139(4):1242–1252 e9. doi:10.1016/j.jaci.2016.08.017.
- Oka T, Rios EJ, Tsai M, Kalesnikoff J, Galli SJ. Rapid desensitization induces internalization of antigen-specific IgE on mouse mast cells. *J Allergy Clin Immunol*. 2013;132(4):922–932.e16. doi:10.1016/j.jaci.2013.05.004.
- Ang WXG, Church AM, Kulis M, Choi HW, Burks AW, Abraham SN. Mast cell desensitization inhibits calcium flux and aberrantly remodels actin. *J Clin Invest*. 2016;126(11):4103–4118. doi:10.1172/JCI87492.
- Burks AW, Jones SM, Wood RA, Fleischer DM, Sicherer SH, Lindblad RW, Stablein D, Henning AK, Vickery BP, Liu AH, et al. Oral immunotherapy for treatment of egg allergy in children. *N Engl J Med*. 2012;367(3):233–243. doi:10.1056/NEJMoa1200435.
- Burks AW, Wood RA, Jones SM, Sicherer SH, Fleischer DM, Scurlock AM, Vickery BP, Liu AH, Henning AK, Lindblad R, et al. Sublingual immunotherapy for peanut allergy: long-term follow-up of a randomized multicenter trial. *J Allergy Clin Immunol*. 2015;135(5):1240–8 e1-3. doi:10.1016/j.jaci.2014.12.1917.
- Swamy RS, Reshamwala N, Hunter T, Vissamsetti S, Santos CB, Baroody FM, Hwang PH, Hoyte EG, Garcia MA, Nadeau KC. Epigenetic modifications and improved regulatory T-cell function in subjects undergoing dual sublingual immunotherapy. *J Allergy Clin Immunol*. 2012;130(1):215–24 e7. doi:10.1016/j.jaci.2012.04.021.
- Caruso M, Cibella F, Emma R, Campagna D, Tringali G, Amaradio MD, Polosa R. Basophil biomarkers as useful predictors for sublingual immunotherapy in allergic rhinitis. *Int Immunopharmacol*. 2018;60:50–58. doi:10.1016/j.intimp.2018.04.034.
- Horak F, Zieglmayer P, Zieglmayer R, Lemell P, Devillier P, Montagut A, Méléac M, Galvain S, Jean-Alphonse S, Van Overtvelt L, et al. Early onset of action of a 5-grass-pollen 300-IR sublingual immunotherapy tablet evaluated in an allergen challenge chamber. *J Allergy Clin Immunol*. 2009;124(3):471–7, 477 e1. doi:10.1016/j.jaci.2009.06.006.
- Patil SU, Shreffler WG. Immunology in the Clinic Review Series; focus on allergies: basophils as biomarkers for assessing immune modulation. *Clin Exp Immunol*. 2012;167(1):59–66. doi:10.1111/j.1365-2249.2011.04503.x.
- Akdis CA, Akdis M. Mechanisms and treatment of allergic disease in the big picture of regulatory T cells. *J Allergy Clin Immunol*. 2009;123(4):735–46; quiz 747–8. doi:10.1016/j.jaci.2009.02.030.
- Aslam A, Chan H, Warrell DA, Misbah S, Ogg GS, Unutmaz D. Tracking antigen-specific T-cells during clinical tolerance induction in humans. *PLoS One*. 2010;5(6):e11028. doi:10.1371/journal.pone.0011028.
- Radulovic S, Jacobson MR, Durham SR, Nouri-Aria KT. Grass pollen immunotherapy induces Foxp3-expressing CD4+ CD25+ cells in the nasal mucosa. *J Allergy Clin Immunol*. 2008;121(6):1467–72, 1472 e1. doi:10.1016/j.jaci.2008.03.013.
- Tsai YG, Chiou Y-L, Chien J-W, Wu H-P, Lin C-Y. Induction of IL-10+ CD4+ CD25+ regulatory T cells with decreased NF-kappaB expression during immunotherapy. *Pediatr Allergy Immunol*. 2010;21(1 Pt 2):e166–73. doi:10.1111/j.1399-3038.2009.00870.x.
- Tsai YG, Yang KD, Niu D-M, Chien J-W, Lin C-Y. TLR2 agonists enhance CD8+Foxp3+ regulatory T cells and suppress Th2 immune responses during allergen immunotherapy. *J Immunol*. 2010;184(12):7229–7237. doi:10.4049/jimmunol.1000083.
- Syed A, Garcia MA, Lyu S-C, Bucayu R, Kohli A, Ishida S, Berglund JP, Tsai M, Maecker H, O’Riordan G, et al. Peanut oral immunotherapy results in increased antigen-induced regulatory T-cell function and hypomethylation of forkhead box protein 3 (FOXP3). *J Allergy Clin Immunol*. 2014;133(2):500–510. doi:10.1016/j.jaci.2013.12.1037.
- Wang M, Yang IV, Davidson EJ, Joetham A, Takeda K, O’Connor BP, Gelfand EW. Forkhead box protein 3 demethylation is associated with tolerance induction in peanut-induced intestinal allergy. *J Allergy Clin Immunol*. 2017;141(2):659–670.e2. doi:10.1016/j.jaci.2017.04.020. Epub 2017 May 4. PubMed PMID: 28479331"

30. Van De Veen W, Stanic B, Yaman G, Wawrzyniak M, Söllner S, Akdis DG, Rückert B, Akdis CA, Akdis M. IgG4 production is confined to human IL-10-producing regulatory B cells that suppress antigen-specific immune responses. *J Allergy Clin Immunol.* 2013;131(4):1204–1212. doi:10.1016/j.jaci.2013.01.014.
31. Norman PS, Lichtenstein LM, Marsh DG. Studies on allergoids from naturally occurring allergens IV. Efficacy and safety of long-term allergoid treatment of ragweed hay fever. *J Allergy Clin Immunol.* 1981;68(6):460–470.
32. Narisety SD, Frischmeyer-Guerrero PA, Keet CA, Gorelik M, Schroeder J, Hamilton RG, Wood RA. A randomized, double-blind, placebo-controlled pilot study of sublingual versus oral immunotherapy for the treatment of peanut allergy. *J Allergy Clin Immunol.* 2015;135(5):1275–82.e1–6. doi:10.1016/j.jaci.2014.11.005.
33. Vickery BP, Scurlock AM, Kulis M, Steele PH, Kamilaris J, Berglund JP, Burk C, Hiegel A, Carlisle S, Christie L, et al. Sustained unresponsiveness to peanut in subjects who have completed peanut oral immunotherapy. *J Allergy Clin Immunol.* 2014;133(2):468–475. doi:10.1016/j.jaci.2013.11.007.
34. Frischmeyer-Guerrero PA, Masilamani M, Gu W, Brittain E, Wood R, Kim J, Nadeau K, Jarvinen KM, Grishin A, Lindblad R, et al. Mechanistic correlates of clinical responses to omalizumab in the setting of oral immunotherapy for milk allergy. *J Allergy Clin Immunol.* 2017;140(4):1043. doi:10.1016/j.jaci.2017.03.028.
35. Morales M, Gallego M, Iraola V, Taulés M, De Oliveira E, Moya R, Carnés J. In vitro evidence of efficacy and safety of a polymerized cat dander extract for allergen immunotherapy. *BMC Immunol.* 2017;18(1):10. doi:10.1186/s12865-017-0193-0.
36. Wallner M, Pichler U, Ferreira F. Recombinant allergens for pollen immunotherapy. *Immunotherapy.* 2013;5(12):1323–1338. doi:10.2217/imt.13.114.
37. Lent AM, Harbeck R, Strand M, Sills M, Schmidt K, Efaw B, Lebo T, Nelson HS. Immunologic response to administration of standardized dog allergen extract at differing doses. *J Allergy Clin Immunol.* 2006;118(6):1249–1256. doi:10.1016/j.jaci.2006.07.055.
38. Verhoef A, Alexander C, Kay AB, Larché M, Platts-Mills T. T cell epitope immunotherapy induces a CD4+ T cell population with regulatory activity. *Plos Medicine.* 2005;2(3):253–261. doi:10.1371/journal.pmed.0020078.
39. Campbell JD, Buckland KF, McMillan SJ, Kearley J, Oldfield WLG, Stern LJ, Grönlund H, Van Hage M, Reynolds CJ, Boyton RJ, et al. Peptide immunotherapy in allergic asthma generates IL-10-dependent immunological tolerance associated with linked epitope suppression. *J Exp Med.* 2009;206(7):1535–1547. doi:10.1084/jem.20082901.
40. Cingi C, Muluk NB, Hanci D, Ulusoy S, Sahin F. Updating the role played by immunotherapy for allergic rhinitis: meta-analysis. *Int Forum Allergy Rhinol.* 2015;5(2):132–142. doi:10.1002/alr.2015.5.issue-2.
41. Abramson M, Puy R, Weiner J. Immunotherapy in asthma: an updated systematic review. *Allergy.* 1999;54(10):1022–1041.
42. Calderon MA, Alves B, Jacobson M, Hurwitz B, Sheikh A, Durham S. Allergen injection immunotherapy for seasonal allergic rhinitis. *Evid Based Child Health.* 2010;5(3):1279–1379. PubMed PMID: 17253469.
43. Greenhawt M, Oppenheimer J, Nelson M, Nelson H, Lockey R, Lieberman P, Nowak-Wegrzyn A, Peters A, Collins C, Bernstein DI, et al. Sublingual immunotherapy: A focused allergen immunotherapy practice parameter update. *Ann Allergy Asthma Immunol.* 2017;118(3):276–282 e2. doi:10.1016/j.anai.2016.12.009.
44. Anagnostou K, Islam S, King Y, Foley L, Pasea L, Bond S, Palmer C, Deighton J, Ewan P, Clark A. Assessing the efficacy of oral immunotherapy for the desensitisation of peanut allergy in children (STOP II): a phase 2 randomised controlled trial. *Lancet.* 2014;383(9925):1297–1304. doi:10.1016/S0140-6736(13)62301-6.
45. Tang ML, Ponsonby A-L, Orsini F, Tey D, Robinson M, Su EL, Licciardi P, Burks W, Donath S. Administration of a probiotic with peanut oral immunotherapy: A randomized trial. *J Allergy Clin Immunol.* 2015;135(3):737–44.e8. doi:10.1016/j.jaci.2014.11.034.
46. Jones SM, Pons L, Roberts JL, Scurlock AM, Perry TT, Kulis M, Shreffler WG, Steele P, Henry KA, Adair M, et al. Clinical efficacy and immune regulation with peanut oral immunotherapy. *J Allergy Clin Immunol.* 2009;124(2):292–300, 300 e1–97. doi:10.1016/j.jaci.2009.05.022.
47. Blumchen K, Ulbricht H, Staden U, Dobberstein K, Beschoner J, De Oliveira LCL, Shreffler WG, Sampson HA, Niggemann B, Wahn U, et al. Oral peanut immunotherapy in children with peanut anaphylaxis. *J Allergy Clin Immunol.* 2010;126(1):83–91 e1. doi:10.1016/j.jaci.2010.04.030.
48. Varshney P, Jones SM, Scurlock AM, Perry TT, Kemper A, Steele P, Hiegel A, Kamilaris J, Carlisle S, Yue X, et al. A randomized controlled study of peanut oral immunotherapy: clinical desensitization and modulation of the allergic response. *J Allergy Clin Immunol.* 2011;127(3):654–660. doi:10.1016/j.jaci.2010.12.1111.
49. MacGinnitie AJ, Rachid R, Gragg H, Little SV, Lakin P, Cianferoni A, Heimall J, Makhija M, Robison R, Chinthrajah RS, et al. Omalizumab facilitates rapid oral desensitization for peanut allergy. *J Allergy Clin Immunol.* 2017;139(3):873–881.e8. doi:10.1016/j.jaci.2016.08.010.
50. Meglio P, Giampietro PG, Gianni S, Galli E. Oral desensitization in children with immunoglobulin E-mediated cow's milk allergy—follow-up at 4 yr and 8 months. *Pediatr Allergy Immunol.* 2008;19(5):412–419. doi:10.1111/j.1399-3038.2007.00670.x.
51. Morisset M, Moneret-Vautrin DA, Guenard L, Cuny JM, Frentz P, Hatahet R, Hanss C, Beaudouin E, Petit N, Kanny G. Oral desensitization in children with milk and egg allergies obtains recovery in a significant proportion of cases. A randomized study in 60 children with cow's milk allergy and 90 children with egg allergy. *Eur Ann Allergy Clin Immunol.* 2007;39(1):12–19.
52. Longo G, Barbi E, Berti I, Meneghetti R, Pittalis A, Ronfani L, Ventura A. Specific oral tolerance induction in children with very severe cow's milk-induced reactions. *J Allergy Clin Immunol.* 2008;121(2):343–347. doi:10.1016/j.jaci.2007.10.029.
53. Skripak JM, Nash SD, Rowley H, Brereton NH, Oh S, Hamilton RG, Matsui EC, Burks AW, Wood RA. A randomized, double-blind, placebo-controlled study of milk oral immunotherapy for cow's milk allergy. *J Allergy Clin Immunol.* 2008;122(6):1154–1160. doi:10.1016/j.jaci.2008.09.030.
54. Escudero C, Rodríguez Del Río P, Sánchez-García S, Pérez-Rangel I, Pérez-Farínós N, García-Fernández C, Ibáñez MD. Early sustained unresponsiveness after short-course egg oral immunotherapy: a randomized controlled study in egg-allergic children. *Clin Exp Allergy.* 2015;45(12):1833–1843. doi:10.1111/cea.12604.
55. Caminiti L, Pajno GB, Crisafulli G, Chiera F, Collura M, Panasci G, Ruggeri P, Guglielmo F, Passalacqua G. Oral Immunotherapy for Egg Allergy: A Double-Blind Placebo-Controlled Study, with Postdesensitization Follow-Up. *J Allergy Clin Immunol Pract.* 2015;3(4):532–539. doi:10.1016/j.jaip.2015.01.017.
56. Vazquez-Ortiz M, Alvaro M, Piquer M, Dominguez O, Machinena A, Martín-Mateos MA, Plaza AM. Baseline specific IgE levels are useful to predict safety of oral immunotherapy in egg-allergic children. *Clin Exp Allergy.* 2014;44(1):130–141. doi:10.1111/cea.12233.
57. Fuentes-Aparicio V, Alvarez-Perea A, Infante S, Zapatero L, D'Oleo A, Alonso-Lebrero E. Specific oral tolerance induction in paediatric patients with persistent egg allergy. *Allergol Immunopathol (Madr).* 2013;41(3):143–150. doi:10.1016/j.aller.2012.02.007.
58. Dello Iacono I, Tripodi S, Calvani M, Panetta V, Verga MC, Miceli Sopo S. Specific oral tolerance induction with raw hen's egg in children with very severe egg allergy: a randomized controlled trial. *Pediatr Allergy Immunol.* 2013;24(1):66–74. doi:10.1111/j.1399-3038.2012.01349.x.
59. Rodriguez Del Rio P, Díaz-Perales A, Sanchez-García S, Escudero C, Do Santos P, Catarino M, Ibañez MD. Oral immunotherapy in children with IgE-mediated wheat allergy: outcome and molecular changes. *J Investig Allergol Clin Immunol.* 2014;24(4):240–248.
60. Sato S, Utsunomiya T, Imai T, Yanagida N, Asaumi T, Ogura K, Koike Y, Hayashi N, Okada Y, Shukuya A, et al. Wheat oral

- immunotherapy for wheat-induced anaphylaxis. *J Allergy Clin Immunol.* 2015;136(4):1131–3.e7. doi:10.1016/j.jaci.2015.07.019.
61. Andorf S, Purington N, Block WM, Long AJ, Tupa D, Brittain E, Rudman Spergel A, Desai M, Galli SJ, Nadeau KC, Chinthrajah RS. Anti-IgE treatment with oral immunotherapy in multifoed allergic participants: a double-blind, randomised, controlled trial. *Lancet Gastroenterol Hepatol.* 2018. Feb;3(2):85–94. doi: 10.1016/S2468-1253(17)30392-8. Epub 2017 Dec 12. PubMed PMID: 29242014.
 62. Hofmann AM, Scurllock AM, Jones SM, Palmer KP, Lokhnygina Y, Steele PH, Kamilaris J, Burks AW. Safety of a peanut oral immunotherapy protocol in children with peanut allergy. *J Allergy Clin Immunol.* 2009;124(2):286–91, 291 e1–6. doi:10.1016/j.jaci.2009.03.045.
 63. Anagnostou K, Clark A. Peanut immunotherapy. *Clin Transl Allergy.* 2014;4:30. doi:10.1186/2045-7022-4-30.
 64. Leonard SA. Baked egg and milk exposure as immunotherapy in food allergy. *Curr Allergy Asthma Rep.* 2016;16(4):32. doi:10.1007/s11882-016-0604-y.
 65. Goldberg MR, Nachshon L, Appel MY, Elizur A, Levy MB, Eisenberg E, Sampson HA, Katz Y. Efficacy of baked milk oral immunotherapy in baked milk-reactive allergic patients. *J Allergy Clin Immunol.* 2015;136(6):1601–1606. doi:10.1016/j.jaci.2015.05.040.
 66. Meglio P, Bartone E, Plantamura M, Arabito E, Giampietro PG. A protocol for oral desensitization in children with IgE-mediated cow's milk allergy. *Allergy.* 2004;59(9):980–987. doi:10.1111/j.1398-9995.2004.00542.x.
 67. Pajno GB, Caminiti L, Salzano G, Crisafulli G, Aversa T, Messina MF, Wasniewska M, Passalacqua G. Comparison between two maintenance feeding regimens after successful cow's milk oral desensitization. *Pediatr Allergy Immunol.* 2013;24(4):376–381. doi:10.1111/pai.12077.
 68. Salmivesi S, Korppi M, Mäkelä MJ, Paasilta M. Milk oral immunotherapy is effective in school-aged children. *Acta Paediatr.* 2013;102(2):172–176. doi:10.1111/j.1651-2227.2012.02815.x.
 69. Paasilta M, Salmivesi S, Mäki T, Helminen M, Korppi M. Children who were treated with oral immunotherapy for cows' milk allergy showed long-term desensitisation seven years later. *Acta Paediatr.* 2016;105(2):215–219. doi:10.1111/apa.13251.
 70. Martorell A, De La Hoz B, Ibáñez MD, Bone J, Terrados MS, Michavila A, Plaza AM, Alonso E, Garde J, Nevot S, et al. Oral desensitization as a useful treatment in 2-year-old children with cow's milk allergy. *Clin Exp Allergy.* 2011;41(9):1297–1304. doi:10.1111/j.1365-2222.2011.03749.x.
 71. Narisety SD, Skripak JM, Steele P, Hamilton RG, Matsui EC, Burks AW, Wood RA. Open-label maintenance after milk oral immunotherapy for IgE-mediated cow's milk allergy. *J Allergy Clin Immunol.* 2009;124(3):610–612. doi:10.1016/j.jaci.2009.06.025.
 72. Keet CA, Seopaul S, Knorr S, Narisety S, Skripak J, Wood RA. Long-term follow-up of oral immunotherapy for cow's milk allergy. *J Allergy Clin Immunol.* 2013;132(3):737–739 e6. doi:10.1016/j.jaci.2013.05.006.
 73. Staden U, Rolinck-Werninghaus C, Brewe F, Wahn U, Niggemann B, Beyer K. Specific oral tolerance induction in food allergy in children: efficacy and clinical patterns of reaction. *Allergy.* 2007;62(11):1261–1269. doi:10.1111/j.1398-9995.2007.01501.x.
 74. Buchanan AD, Green TD, Jones SM, Scurllock AM, Christie L, Althage KA, Steele PH, Pons L, Helm RM, Lee LA, et al. Egg oral immunotherapy in nonanaphylactic children with egg allergy. *J Allergy Clin Immunol.* 2007;119(1):199–205. doi:10.1016/j.jaci.2006.09.016.
 75. Burks AW, Jones SM. Egg oral immunotherapy in non-anaphylactic children with egg allergy: follow-up. *J Allergy Clin Immunol.* 2008;121(1):270–271. doi:10.1016/j.jaci.2007.07.066.
 76. Vickery BP, Pons L, Kulis M, Steele P, Jones SM, Burks AW. Individualized IgE-based dosing of egg oral immunotherapy and the development of tolerance. *Ann Allergy Asthma Immunol.* 2010;105(6):444–450. doi:10.1016/j.ana.2010.09.030.
 77. Ruiz Garcia M, Haroun E, Landivar ME, Torres Hernandez JA, Sastre J. Commercial dehydrated egg white for specific oral tolerance induction (SOTI): an easier treatment for egg allergy. *J Investig Allergol Clin Immunol.* 2012;22(7):529–531.
 78. Tortajada-Girbes M, Porcar-Almela M, Martorell-Giménez L, Tallón-Guerola M, Gracia-Antequera M, Codoñer-Franch P. Specific oral tolerance induction (SOTI) to egg: our experience with 19 children. *J Investig Allergol Clin Immunol.* 2012;22(1):75–77.
 79. Letran A, Espinazo M, López MC, Caro FJ, Gómez L, Lobatón P, Dafonte J, Moreno F. Threshold doses in specific oral tolerance induction in children with egg allergy. *J Investig Allergol Clin Immunol.* 2012;22(2):147–149.
 80. Ojeda P, Ojeda I, Rubio G, Pineda F. Home-based oral immunotherapy protocol with pasteurized egg for children allergic to hen's egg. *Isr Med Assoc J.* 2012;14(1):34–39.
 81. Meglio P, Giampietro PG, Carello R, Gabriele I, Avitabile S, Galli E. Oral food desensitization in children with IgE-mediated hen's egg allergy: a new protocol with raw hen's egg. *Pediatr Allergy Immunol.* 2013;24(1):75–83. doi:10.1111/j.1399-3038.2012.01341.x.
 82. Dello Iacono I, Verga MC, Tripodi S. Oral immunotherapy for egg allergy in children. *N Engl J Med.* 2012;367(15):1471; author reply 1472–3.
 83. García Rodríguez R, Urrea JM, Feo-Brito F, Galindo PA, Borja J, Gómez E, Lara P, Guerra F. Oral rush desensitization to egg: efficacy and safety. *Clin Exp Allergy.* 2011;41(9):1289–1296. doi:10.1111/j.1365-2222.2011.03722.x.
 84. Itoh N, Itagaki Y, Kurihara K. Rush specific oral tolerance induction in school-age children with severe egg allergy: one year follow up. *Allergol Int.* 2010;59(1):43–51. doi:10.2332/allergolint.09-OA-0107.
 85. Leonard SA, Sampson HA, Sicherer SH, Noone S, Moshier EL, Godbold J, Nowak-Węgrzyn A. Dietary baked egg accelerates resolution of egg allergy in children. *J Allergy Clin Immunol.* 2012;130(2):473–80 e1. doi:10.1016/j.jaci.2012.06.006.
 86. Jones SM, Burks AW, Keet C, Vickery BP, Scurllock AM, Wood RA, Liu AH, Sicherer SH, Henning AK, Lindblad RW, et al. Long-term treatment with egg oral immunotherapy enhances sustained unresponsiveness that persists after cessation of therapy. *J Allergy Clin Immunol.* 2016;137(4):1117–1127.e10. doi:10.1016/j.jaci.2015.12.1316.
 87. Vickery BP, Berglund JP, Burk CM, Fine JP, Kim EH, Kim JI, Keet CA, Kulis M, Orgel KG, Guo R, et al. Early oral immunotherapy in peanut-allergic preschool children is safe and highly effective. *J Allergy Clin Immunol.* 2017;139(1):173–181.e8. doi:10.1016/j.jaci.2016.05.027.
 88. Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, Brough HA, Phippard D, Basting M, Feeney M, et al. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med.* 2015;372(9):803–813. doi:10.1056/NEJMoa1414850.
 89. Moller C. Effect of pollen immunotherapy on food hypersensitivity in children with birch pollinosis. *Ann Allergy.* 1989;62(4):343–345.
 90. Herrmann D, Henzgen M, Frank E, Rudeschko O, Jäger L. Effect of hyposensitization for tree pollinosis on associated apple allergy. *J Investig Allergol Clin Immunol.* 1995;5(5):259–267.
 91. Asero R. Effects of birch pollen-specific immunotherapy on apple allergy in birch pollen-hypersensitive patients. *Clin Exp Allergy.* 1998;28(11):1368–1373.
 92. Bucher X, Pichler WJ, Dahinden CA, Helbling A. Effect of tree pollen specific, subcutaneous immunotherapy on the oral allergy syndrome to apple and hazelnut. *Allergy.* 2004;59(12):1272–1276. doi:10.1111/j.1398-9995.2004.00626.x.
 93. Hansen KS, Khinchi MS, Skov PS, Bindslev-Jensen C, Poulsen LK, Malling H-J. Food allergy to apple and specific immunotherapy with birch pollen. *Mol Nutr Food Res.* 2004;48(6):441–448. doi:10.1002/mnfr.200400037.
 94. Kinaciyan T, Jahn-Schmid B, Radakovics A, Zwölfer B, Schreiber C, Francis JN, Ebner C, Bohle B. Successful sublingual immunotherapy with birch pollen has limited effects on concomitant food allergy to apple and the immune response to the Bet v 1

- homolog Mal d 1. *J Allergy Clin Immunol.* 2007;119(4):937–943. doi:10.1016/j.jaci.2006.11.010.
95. Mauro M, Russello M, Incorvaia C, Gazzola G, Frati F, Moingeon P, Passalacqua G. Birch-apple syndrome treated with birch pollen immunotherapy. *Int Arch Allergy Immunol.* 2011;156(4):416–422. doi:10.1159/000323909.
 96. Kopac P, Rudin M, Gentinetta T, Gerber R, Pichler C, Hausmann O, Schnyder B, Pichler WJ. Continuous apple consumption induces oral tolerance in birch-pollen-associated apple allergy. *Allergy.* 2012;67(2):280–285. doi:10.1111/j.1398-9995.2011.02744.x.
 97. Kinaciyani T, Nagl B, Faustmann S, Frommlet F, Kopp S, Wolkersdorfer M, Wöhrl S, Bastl K, Huber H, Berger U, et al. Efficacy and safety of 4 months of sublingual immunotherapy with recombinant Mal d 1 and Bet v 1 in patients with birch pollen-related apple allergy. *J Allergy Clin Immunol.* 2018 Mar;141(3):1002–1008. doi: 10.1016/j.jaci.2017.07.036. Epub 2017 Sep 1. PubMed PMID: 28870463.
 98. Senti G, Von Moos S, Tay F, Graf N, Sonderegger T, Johansen P, Kündig TM. Epicutaneous allergen-specific immunotherapy ameliorates grass pollen-induced rhinoconjunctivitis: A double-blind, placebo-controlled dose escalation study. *J Allergy Clin Immunol.* 2012;129(1):128–135. doi:10.1016/j.jaci.2011.08.036.
 99. Dupont C, Kalach N, Soulaïnes P, Legoué-Morillon S, Piloquet H, Benhamou P-H. Cow's milk epicutaneous immunotherapy in children: a pilot trial of safety, acceptability, and impact on allergic reactivity. *J Allergy Clin Immunol.* 2010;125(5):1165–1167. doi:10.1016/j.jaci.2010.02.029.
 100. Sampson HA, Shreffler WG, Yang WH, Sussman GL, Brown-Whitehorn TF, Nadeau KC, Cheema AS, Leonard SA, Pongracic JA, Sauvage-Delebarre C, et al. Effect of varying doses of epicutaneous immunotherapy vs placebo on reaction to peanut protein exposure among patients with peanut sensitivity: a randomized clinical trial. *JAMA.* 2017;318(18):1798–1809. doi:10.1001/jama.2017.16591.
 101. Von Moos S, Johansen P, Tay F, Graf N, Kündig TM, Senti G. Comparing safety of abrasion and tape-stripping as skin preparation in allergen-specific epicutaneous immunotherapy. *J Allergy Clin Immunol.* 2014;134(4):965–7.e4. doi:10.1016/j.jaci.2014.07.037.
 102. Dioszeghy V, Mondoulet L, Dhelft V, Ligouis M, Puteaux E, Benhamou P-H, Dupont C. Epicutaneous immunotherapy results in rapid allergen uptake by dendritic cells through intact skin and downregulates the allergen-specific response in sensitized mice. *J Immunol.* 2011;186(10):5629–5637. doi:10.4049/jimmunol.1003134.
 103. Senti G, Cramer R, Kuster D, Johansen P, Martinez-Gomez JM, Graf N, Steiner M, Hothorn LA, Grönlund H, Tivig C, et al. Intralymphatic immunotherapy for cat allergy induces tolerance after only 3 injections. *J Allergy Clin Immunol.* 2012;129(5):1290–1296. doi:10.1016/j.jaci.2012.02.026.
 104. Senti G, Prinz Vavricka BM, Erdmann I, Diaz MI, Markus R, McCormack SJ, Simard JJ, Wüthrich B, Cramer R, Graf N, et al. Intralymphatic allergen administration renders specific immunotherapy faster and safer: a randomized controlled trial. *Proc Natl Acad Sci U S A.* 2008;105(46):17908–17912. doi:10.1073/pnas.0803725105.
 105. Hjalmarsson E, Bierrenbach AL, Calderon MA, Sheikh A, Simons FER, Demoly P. Intralymphatic immunotherapy (ilit) with both grass and birch allergen- a randomized double-blind placebo controlled trial. *Allergy.* 2017;72:120. doi:10.1111/all.13006.
 106. Hylander T, Larsson O, Petersson-Westin U, Eriksson M, Kumlien Georén S, Winqvist O, Cardell L-O. Intralymphatic immunotherapy of pollen-induced rhinoconjunctivitis: a double-blind placebo-controlled trial. *Respir Res.* 2016;17:10. doi:10.1186/s12931-016-0324-9.
 107. Lee SM, Jung JH, Choi SJ, Joe E, Kang SM, Kim YJ, Kyung SY, Park J-W, Jeong SH, Lee SP. The evaluation of efficacy and adverse effect in intralymphatic allergen-specific immunotherapy against house dust mite, cat, and dog allergens in allergic rhinitis. *J Allergy Clin Immunol.* 2015;135(2):AB159–AB159. doi:10.1016/j.jaci.2014.12.1460.
 108. Lee SP, Choi SJ, Joe E, Lee SM, Lee MW, Shim JW, Kim YJ, Kyung SY, Park JW, Jeong SH, et al. A pilot study of intralymphatic immunotherapy for house dust mite, cat, and dog allergies. *Allergy Asthma Immunol Res.* 2017;9(3):272–277. doi:10.4168/aa.2017.9.3.272.
 109. Kundig TM, Johansen P, Bachmann MF, Cardell LO, Senti G. Intralymphatic immunotherapy: time interval between injections is essential. *J Allergy Clin Immunol.* 2014;133(3):930–931. doi:10.1016/j.jaci.2013.11.036.
 110. Wood RA, Kim JS, Lindblad R, Nadeau K, Henning AK, Dawson P, Plaut M, Sampson HA. A randomized, double-blind, placebo-controlled study of omalizumab combined with oral immunotherapy for the treatment of cow's milk allergy. *J Allergy Clin Immunol.* 2016;137(4):1103–1110.e11. doi:10.1016/j.jaci.2015.10.005.
 111. Nadeau KC, Schneider LC, Hoyte L, Borrás I, Umetsu DT. Rapid oral desensitization in combination with omalizumab therapy in patients with cow's milk allergy. *J Allergy Clin Immunol.* 2011;127(6):1622–1624. doi:10.1016/j.jaci.2011.04.009.
 112. Schneider LC, Rachid R, LeBovidge J, Blood E, Mittal M, Umetsu DT. A pilot study of omalizumab to facilitate rapid oral desensitization in high-risk peanut-allergic patients. *J Allergy Clin Immunol.* 2013;132(6):1368–1374. doi:10.1016/j.jaci.2013.09.046.
 113. Lafuente I, Mazon A, Nieto M, Uixera S, Pina R, Nieto A. Possible recurrence of symptoms after discontinuation of omalizumab in anti-IgE-assisted desensitization to egg. *Pediatr Allergy Immunol.* 2014;25(7):717–719. doi:10.1111/pai.12259.
 114. Begin P, Shams-Ghahfarokhi M, Postigo I, Razzaghi-Abyaneh M, Eslamifard A, Gutiérrez A, Suñén E, Martínez J. Phase 1 results of safety and tolerability in a rush oral immunotherapy protocol to multiple foods using Omalizumab. *Allergy Asthma Clin Immunol.* 2014;10(1):7. doi:10.1186/1710-1492-10-41.
 115. Martorell-Calatayud C, Michavila-Gómez A, Martorell-Aragonés A, Molini-Menchón N, Cerdá-Mir JC, Félix-Toledo R, De Las Marinas-Álvarez MD. Anti-IgE-assisted desensitization to egg and cow's milk in patients refractory to conventional oral immunotherapy. *Pediatr Allergy Immunol.* 2016;27(5):544–546. doi:10.1111/pai.12567.
 116. Leung DY, Sampson HA, Yunginger JW, Burks AW, Schneider LC, Wortel CH, Davis FM, Hyun JD, Shanahan WR. Effect of anti-IgE therapy in patients with peanut allergy. *N Engl J Med.* 2003;348(11):986–993. doi:10.1056/NEJMoa022613.
 117. Casale TB, Busse WW, Kline JN, Ballas ZK, Moss MH, Townley RG, Mokhtarani M, Seyfert-Margolis V, Asare A, Bateman K, et al. Omalizumab pretreatment decreases acute reactions after rush immunotherapy for ragweed-induced seasonal allergic rhinitis. *J Allergy Clin Immunol.* 2006;117(1):134–140. doi:10.1016/j.jaci.2005.09.036.
 118. Kuehr J, Brauburger J, Zielen S, Schauer U, Kamin W, Von Berg A, Leupold W, Bergmann K-C, Rolinck-Werninghaus C, Gräve M, et al. Efficacy of combination treatment with anti-IgE plus specific immunotherapy in polysensitized children and adolescents with seasonal allergic rhinitis. *J Allergy Clin Immunol.* 2002;109(2):274–280.
 119. Lambert N, Guiddir T, Amat F, Just J. Pre-treatment by omalizumab allows allergen immunotherapy in children and young adults with severe allergic asthma. *Pediatr Allergy Immunol.* 2014;25(8):829–832. doi:10.1111/pai.12306.
 120. Massanari M, Nelson H, Casale T, Busse W, Kianifard F, Geba GP, Zeldin RK. Effect of pretreatment with omalizumab on the tolerability of specific immunotherapy in allergic asthma. *J Allergy Clin Immunol.* 2010;125(2):383–389. doi:10.1016/j.jaci.2009.11.022.
 121. Kopp MV, Hamelmann E, Zielen S, Kamin W, Bergmann K-C, Sieder C, Stenglein S, Seyfried S, Wahn U. Combination of omalizumab and specific immunotherapy is superior to immunotherapy in patients with seasonal allergic rhinoconjunctivitis and co-morbid seasonal allergic asthma. *Clin Exp Allergy.* 2009;39(2):271–279. doi:10.1111/j.1365-2222.2008.03121.x.
 122. Roy K, Mao HQ, Huang SK, Leong KW. Oral gene delivery with chitosan-DNA nanoparticles generates immunologic protection in a murine model of peanut allergy. *Nat Med.* 1999;5(4):387–391. doi:10.1038/7385.

123. Basomba A, Tabar AI, De Rojas DHF, García BE, Alamar R, Olaguibel JM, Del Prado JM, Martín S, Rico P. Allergen vaccination with a liposome-encapsulated extract of *Dermatophagoides pteronyssinus*: a randomized, double-blind, placebo-controlled trial in asthmatic patients. *J Allergy Clin Immunol*. 2002;109(6):943–948.
124. Galvain S, André C, Vatrinet C, Villet B. Safety and efficacy studies of liposomes in specific immunotherapy. *Curr Ther Res-Clin Exp*. 1999;60(5):278–294. doi:10.1016/S0011-393X(99)80004-6.
125. Senti G, Johansen P, Haug S, Bull C, Gottschaller C, Müller P, Pfister T, Maurer P, Bachmann MF, Graf N, et al. Use of A-type CpG oligodeoxynucleotides as an adjuvant in allergen-specific immunotherapy in humans: a phase I/IIa clinical trial. *Clin Exp Allergy*. 2009;39(4):562–570. doi: 10.1111/j.1365-2222.2008.03191.x. Epub 2009 Feb 16. PubMed PMID: 19226280.
126. Klimek L, Willers J, Hammann-Haenni A, Pfaar O, Stocker H, Mueller P, Renner WA, Bachmann MF. Assessment of clinical efficacy of CYT003-QbG10 in patients with allergic rhinoconjunctivitis: a phase IIb study. *Clin Exp Allergy*. 2011;41(9):1305–1312. doi:10.1111/j.1365-2222.2011.03783.x.
127. Pohlit H, Bellinghausen I, Frey H, Saloga J. Recent advances in the use of nanoparticles for allergen-specific immunotherapy. *Allergy*. 2017;72(10):1461–1474. doi:10.1111/all.13199.