

## Basic Study

# Interplay between post-translational cyclooxygenase-2 modifications and the metabolic and proteomic profile in a colorectal cancer cohort

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## Abstract

### BACKGROUND

Colorectal cancer (CRC) is the second most common cause of cancer death worldwide. It is broadly described that cyclooxygenase-2 (COX-2) is mainly overexpressed in CRC but less is known regarding post-translational modifications of this enzyme that may regulate its activity, intracellular localization and stability. Since metabolic and proteomic profile analysis is essential for cancer prognosis and diagnosis, our hypothesis is that the analysis of correlations between these specific parameters and COX-2 state in tumors of a

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#### Institutional review board

**statement:** Colorectal cancer biopsies were obtained from 45 patients under informed consent, and were used in accordance with the procedures approved by Clinical Investigation ethics committees of the Germans Trias i Pujol Hospital (Badalona, Spain) and Bellvitge Hospital (Barcelona, Spain).

#### Conflict-of-interest statement:

MAP is cofounder and equity holder of Aniling, a biotech company with no interests in this work. MAP lab has received research funding from Celgene. The rest of the authors declare no conflict of interest.

**Data sharing statement:** No additional data is available.

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high number of CRC patients could be useful for the understanding of the basis of this cancer in humans.

#### AIM

To analyze COX-2 regulation in colorectal cancer and to perform a detailed analysis of their metabolic and proteomic profile.

#### METHODS

Biopsies from both healthy and pathological colorectal tissues were taken under informed consent from patients during standard colonoscopy procedure in the University Hospital of Bellvitge (Barcelona, Spain) and Germans Trias i Pujol University Hospital (Campus Can Ruti) (Barcelona, Spain). Western blot analysis was used to determine COX-2 levels. Deglycosylation assays were performed in both cells and tumor samples incubating each sample with peptide N-glycosidase F (PNGase F). Prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) levels were determined using a specific ELISA. <sup>1</sup>H high resolution magic angle spinning (HRMAS) analysis was performed using a Bruker AVIII 500 MHz spectrometer and proteomic analysis was performed in a nano-liquid chromatography-tandem mass spectrometer (nano LC-MS/MS) using a QExactive HF orbitrap MS.

#### RESULTS

Our data show that COX-2 has a differential expression profile in tumor tissue of CRC patients *vs* the adjacent non-tumor area, which correspond to a glycosylated and less active state of the protein. This fact was associated to a lesser PGE<sub>2</sub> production in tumors. These results were corroborated *in vitro* performing deglycosylation assays in HT29 cell line where COX-2 protein profile was modified after PNGase F incubation, showing higher PGE<sub>2</sub> levels. Moreover, HRMAS analysis indicated that tumor tissue has altered metabolic features *vs* non-tumor counterparts, presenting increased levels of certain metabolites such as taurine and phosphocholine and lower levels of lactate. In proteomic experiments, we detected an enlarged number of proteins in tumors that are mainly implicated in basic biological functions like mitochondrial activity, DNA/RNA processing, vesicular trafficking, metabolism, cytoskeleton and splicing.

#### CONCLUSION

In our colorectal cancer cohort, tumor tissue presents a differential COX-2 expression pattern with lower enzymatic activity that can be related to an altered metabolic and proteomic profile.

**Key words:** Colon; Carcinoma; Cyclooxygenase; Prostaglandin; Proteomics; High resolution magic angle spinning

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**Core tip:** Although increased levels of cyclooxygenase-2 (COX-2) have been broadly related to colorectal cancer (CRC), high variability exists among patients. In our cohort, the electrophoretic band profile of COX-2 was altered in tumor tissue due to COX-2 glycosylation. Furthermore, prostaglandin E<sub>2</sub> levels in tumor tissue were significantly lower than in normal colonic mucosa. In order to further characterize these samples, we have also detected an altered metabolic and proteomic profile in the tumor samples compared to controls that can be associated with the presence of glycosylated COX-2. These findings contribute to delineate the post-translational complexity and heterogeneity of molecular signaling regulation in CRC progression.

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## INTRODUCTION

Colorectal cancer (CRC) is the second most common cause of cancer death worldwide. There were over 18.1 million new cases and 9.6 million deaths in 2018 according to the World Health Organization. Patient prognosis is based on several parameters like the clinical stage of the disease, histological type, tumor grade and the analysis of active oncogenes<sup>[1]</sup>. One of the targets for CRC therapy is cyclooxygenase-2 (COX-2), an inducible enzyme usually absent but transcriptionally upregulated upon neoplastic and pro-inflammatory insults<sup>[2,3]</sup>. In CRC, whereas cyclooxygenase 1 (COX-1) expression remains unaltered when comparing CRC cells and normal colonic mucosa, COX-2 is overexpressed in 40%-50% of benign polyps and in 80%-90% of adenocarcinomas<sup>[4,5]</sup>. Both enzymes catalyze a key step in the conversion of arachidonate (AA) to prostaglandin H<sub>2</sub> (PGH<sub>2</sub>), the immediate substrate for a series of cell-specific prostaglandin (PG) and thromboxane synthases. In particular, PGE<sub>2</sub> is partially pro-inflammatory (depending on the tissue and accumulation) and it has been related to CRC progression<sup>[6,7]</sup>. The potential tumor suppressor enzyme 15-prostaglandin dehydrogenase (15-PGDH), which catalyzes the degradation of PGE<sub>2</sub>, is usually down-regulated in colorectal adenoma and carcinoma cells<sup>[8,9]</sup>. In addition to this, experimental data have demonstrated that, not only the expression levels of COX-2 are important: post-translational modifications of this enzyme may also regulate its function and degradation<sup>[10]</sup>. COX-2 sequence contains five potential N-glycosylation sites, three of which are always glycosylated, one Asn580 in human and mouse that is glycosylated ≤ 50% of the times, and one that is never glycosylated<sup>[11]</sup>. The variability of glycosylation at Asn580 leads to the production of two distinct glycoforms of 72 and 74 kDa, respectively. In this context, it has been described that glycosylation of COX-2 can target the protein for proteolysis through the endoplasmic reticulum pathway and has been associated with alterations in the activity of the enzyme<sup>[12]</sup>. While the above-mentioned studies reported various COX-2 post-translational regulatory mechanisms, the biological and physiological significance of the post-translational regulation of COX-2 remains to be investigated.

For cancer prognosis and diagnosis, it also gains special relevance the analysis of alterations in both the metabolic and protein profile in tumor tissues of patients in contrast to the non-tumor adjacent regions. Nuclear magnetic resonance techniques, such as high resolution magic angle spinning (HRMAS) provide an in-depth analysis for the identification of distinctive metabolites in cancer and can be used to correlate changes in metabolic profiles with the severity of the disease and probable prognosis<sup>[13-15]</sup>. Although several recent studies have been carried out in CRC patients with nuclear magnetic resonance (NMR) techniques, they have been mainly focused on the improvement of screening (in serum, urinary tissue and fecal extracts) and early diagnosis of the disease<sup>[16-18]</sup>. Additionally, proteomic analyses have been effective in the identification of peptides present in different pathological contexts, including cancer<sup>[19-21]</sup>. Thus, the analysis of correlations between metabolic and molecular parameters in tumors of a high number of CRC patients could be useful for the understanding of the basis of this cancer in humans.

In this study, we have analyzed in samples from CRC patients the post-translational regulation of COX-2 together with a detailed analysis of both the metabolic and the proteomic profile of the tumor tissue compared to the non-tumor adjacent area to elucidate whether distinctive COX-2 post-translational modifications can be associated to a specific, altered metabolic profile in these CRC samples.

## MATERIALS AND METHODS

### *Human colon biopsies*

Biopsies from both healthy and pathological colorectal tissues were taken under informed consent from patients during standard colonoscopy procedure in the University Hospital of Bellvitge (Barcelona, Spain) and Germans Trias i Pujol University Hospital (Campus Can Ruti) (Barcelona, Spain). The mean age of participants was 71 ± 15 years old. All procedures were approved by the Ethical Committee for Clinical Investigation of University Hospital of Bellvitge and University Hospital of Germans Trias i Pujol (Barcelona, Spain).

### *Cell culture*

The colorectal adenocarcinoma cell line HT-29 was maintained in DMEM medium (Gibco) containing 100 U/mL penicillin, 100 µg/mL streptomycin and 10% heat-inactivated fetal bovine serum (FBS).

**Deglycosylation assay in vivo for HT29 cells**

Three hundred thousand cells were seeded per well on 6-well plates. When confluence was reached, cells were starved in serum-free media for 18 h. Then, deglycosylation was performed by treating for 24 h with 5 mmol/L glucosamine-hydrochloride (Sigma #G4875). To also evaluate PGE<sub>2</sub> production, cells were then treated with 30 μmol/L arachidonic acid (Sigma #10931) for 30 min. Two hundred microliter of media was collected before and after treatment with arachidonic acid and plates were stored at -80 °C until processing.

**Deglycosylation assay in vitro for tumor samples**

Tumor samples were denaturalized by mixing 50 μg of total protein extract with 50 μL of 50 mmol/L NaH<sub>2</sub>PO<sub>4</sub> buffer (pH 7.5) and 5 μL of “denaturing solution” containing 0.2% SDS and 1% 2-mercaptoethanol. After heating at 100 °C for 10 min and recovery of room temperature, deglycosylation was performed incubating each sample with 500 U/mL of PNGase F (Sigma #P7367) and 5 μL of 15% Triton X-100 for 18 h at 37 °C. Reaction was then stopped by boiling samples at 100 °C for 5 min.

**Preparation of total protein cell extracts**

Cells were homogenized in a medium containing 10 mmol/L Tris-HCl, pH 7.5; 1 mmol/L MgCl<sub>2</sub>, 1 mmol/L EGTA, 10% glycerol, 0.5% CHAPS and proteases and phosphatases inhibitors cocktails (Sigma). The extracts were vortexed for 30 min at 4 °C and after centrifuging for 15 min at 13000 rpm, the supernatants were stored at -20 °C. Protein levels were determined with Bradford reagent (Bio-Rad).

**Preparation of total protein extracts from human tissue**

Several milligrams of tissue were pulverized and then were homogenized in 10 mmol/L Tris-HCl, pH 7.5; 1 mmol/L MgCl<sub>2</sub>, 1 mmol/L EGTA, 10% glycerol, 0.5% CHAPS and proteases and phosphatases inhibitors cocktails (Sigma) at 4 °C using an Ultra-turrax dispersing instrument (Ika). The extracts were vortexed for 30 min at 4 °C and after centrifuging for 15 min at 13000 rpm, the supernatants were stored at -80 °C. Protein levels were determined with Bradford reagent (Bio-Rad).

**Western blot analysis**

Equal amounts of protein (20-40 μg) from each fraction obtained were loaded into a 10%-12% SDS-PAGE. Proteins were size fractionated, transferred to a PVDF membrane (Bio-Rad) and, after blocking with 5% non-fat dry milk, incubated with the corresponding antibody for COX-1 (sc-1752) or COX-2 (sc-1747) from Santa Cruz Biotechnology. Blots were normalized by the measurement of the amount for GAPDH (ThermoFisher Scientific #AM4300) and developed by ECL protocol and different exposition times were performed for each blot to ensure the linearity of the band intensities. Values of densitometry were determined using Image J software.

**PGE<sub>2</sub> measurement**

Levels of PGE<sub>2</sub> were determined in tumor and normal colonic mucosa samples and in the culture medium of HT29 cells using a specific Enzyme Immunoassay Kit (Arbor Assays, #K051-H1) following the manufacturer's instructions.

**<sup>1</sup>H HRMAS analysis**

All <sup>1</sup>H HRMAS experiments were carried out at 277.15 K on a Bruker AVANCE III 500 MHz spectrometer (Bruker Biospin, Germany) using a <sup>1</sup>H probe with a sample spin rate of 4000 Hz. Each tissue sample was individually placed in D<sub>2</sub>O saline (99.2% <sup>2</sup>H, Apollo Scientific Limited, Stockport, United Kingdom) and inserted into a zirconium oxide rotor (50 μL) for all NMR acquisitions. In order to attenuate NMR signals of macromolecules, a Carr-Purcell-Meiboom-Gil (CPMG) spin-echo spectrum was collected for each sample with 2 pulses and TE = 36 ms and TR = 144 ms. An integration routine using Mnova software (Mestrelab Research, Santiago de Compostela, Spain) was used to determine the areas under the peaks (parts per million; ppm) after adjusting phase and baseline.

**Proteomic analysis**

Proteins from normal and tumor colon tissue were extracted using tissue homogenization with ceramic beads (MagNa Lyser Green Beads apparatus, Roche, Germany) in extraction buffer (50 mmol/L Tris-HCl pH 6.8; 4% SDS, 10 mmol/L DTT). Proteins were digested using the filter aided sample preparation (FASP) protocol as previously described<sup>[22]</sup> and the resulting peptides were analyzed by nano-liquid chromatography-tandem mass spectrometry (nanoLC-MS/MS) using a QExactive HF orbitrap mass spectrometer (Thermo Scientific). Peptides were injected onto a C18 reversed phase (RP) nano-column (75 μm I.D. and 50 cm, Acclaim

PepMap100, Thermo Scientific) in buffer A [0.1% formic acid (v/v)] and eluted with a 240 min lineal gradient of buffer B [90% acetonitrile, 0.1% formic acid (v/v)]. MS runs consisted of enhanced FT-resolution spectra (140000 resolution) followed by data-dependent MS/MS spectra of the 15 most intense parent ions acquired along the chromatographic run. HCD fragmentation was performed at 27% of normalized collision energy. For peptide identification, the MS/MS spectra were searched with the SEQUEST HT algorithm implemented in Proteome Discoverer 2.1 (Thermo Scientific). The results were analyzed using the probability ratio method<sup>[23]</sup> and the false discovery rate (FDR) of peptide identification was calculated based on the search results against a decoy database using the refined method<sup>[24]</sup>. Peptide and scan counting were performed assuming as positive events those with a FDR equal or lower than 1%. Enrichment analyses were performed by using the DAVID functional annotation database (<https://david.ncifcrf.gov/>).

### Statistical analysis

The values in graphs correspond to the means  $\pm$  SD. Statistical analysis was performed by qualified personnel. Analysis was based on continuous variables, and statistical significance was assessed with the Student-*t* test for paired observations and the two-way ANOVA for multiple comparisons. Correlations between continuous variables were summarized with the Pearson coefficient (indicated in the corresponding figures).

## RESULTS

### Study of post-translational COX-2 modifications

COX-2 seems to play an important role in multiple CRC cellular functions including apoptosis, cell invasiveness, and angiogenesis. We have analyzed both COX isoforms and 15-PGDH levels in 45 CRC patients whose data are detailed in [Table 1](#). We first analyzed the levels of these proteins in tumors and the corresponding adjacent non-tumor tissues. In contrast to COX-1 or 15-PGDH, in which no significant differences were detected ([Figure 1A-C](#)), COX-2 exhibited a change in its protein expression profile in tumors presenting two bands instead of one, at approximately 66 and 72 kDa ([Figure 1D](#)). Regarding all non-tumor/tumor pairs included in this study, whereas non-tumor tissue predominantly exhibits the lower band (79% of cases) ([Figure 1E](#)), only a 27% of tumor samples had this band and 60% had the upper band or both. In fact, tumor samples have significantly more “upper COX-2” band and less “lower COX-2” band than their corresponding non-tumor tissue. To determine whether this modification can be related to the activity of the enzyme, we measured the content of PGE<sub>2</sub> in both groups ([Figure 1F](#)). Our results show that tumor tissue had a significantly lower PGE<sub>2</sub> content compared to their paired non-tumor regions. This situation also prevailed in tumors that only presented the upper band ([Figure 1G](#)).

COX-2 glycosylation has been described as one of the main post-translational modifications that can affect the electrophoretic mobility of the protein. We have performed deglycosylation assays in several tumor samples and observed that in deglycosylated tumors (DT) COX-2 band appears at a lower molecular weight, while the upper initial band was missing ([Figure 2A and B](#)). These experiments were confirmed *in vitro* using the colon cancer cell line HT29 ([Figure 2C](#)). These cells presented only COX-2 upper band under basal conditions, but when treated for deglycosylation, several bands of lower molecular weight appeared along with the upper one that decreased ([Figure 2D](#)). These results indicate that COX-2 appears mainly in a glycosylated state in tumor cells and in tumor samples from CRC patients. Moreover, as it has been described that post-translational modifications can affect COX-2 activity, PGE<sub>2</sub> was measured in cells before and after deglycosylation ([Figure 2E](#)). We obtained higher levels of PGE<sub>2</sub> after deglycosylation indicating that the glycosylated-state of COX-2 appears to be less active than controls in both HT29 and in CRC samples.

### Analysis of the metabolic profile of CRC samples

Another important aspect to consider in CRC is the metabolic profile of the patient, which may reflect the general state of the organism. We have done a comparative study in non-tumor *vs.* tumor colorectal tissues from the same patient by HRMAS (representative obtained profile shown in [Figure 3A](#)), demonstrating that, in tumors, there is an altered metabolic profile with significantly higher presence of taurine (both peaks at  $\delta$  ppm 3.26 and 3.42) ([Figure 3B](#)). We then analyzed whether these differences in taurine for both peaks could be sex- or tumor stage-related. We found a

Table 1 Patient demographic and pathological data

	Age (median)	75 (25-95)
Sex	Male	28
	Female	17
Anatomical location	Distal	15
	Proximal	12
	Unknown	18
T stage	T1	1
	T2	19
	T3	20
	T4	5
N stage	N0	17
	N1	2
	N2	7
	Unknown	19

significant correlation between the levels of both peaks of taurine 3.26 and 3.42 and the tumor stage, being higher in stage 1-2 when compared to stage 3-4 (Figure 3C and D). However, we did not find any significant difference between sexes (Supplementary Figure 1). Moreover, in HRMAS analysis we also detected significantly higher levels of phosphocholine  $[(\text{CH}_3)_3\text{N}]$  in tumors (peak at  $\delta$  ppm 3.22; Figure 3E). Other metabolites, such as lactate (Figure 3F), acetate and glycine did not show significant difference (Supplementary Figure 2A-B). To further investigate if there is a correlation between taurine levels and the presence of COX-2 (both bands) and 15-PGDH, we performed a broad statistical analysis calculating Pearson's correlation coefficient between these variables (Figure 4A-F). Interestingly, only the levels of taurine at 3.26 peak significantly correlates with the levels of the upper COX-2 band (Figure 4A).

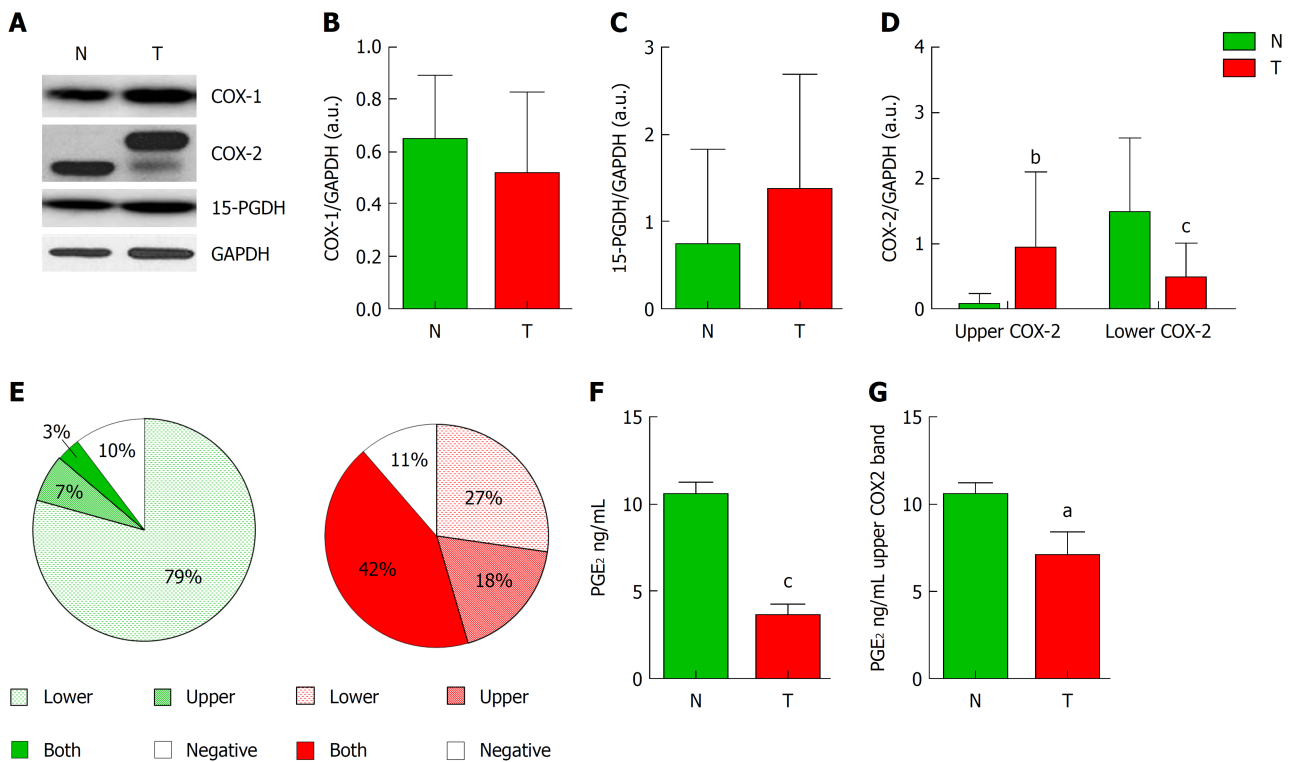
### Proteomic analysis of CRC samples

To complete the molecular analysis of our CRC cohort, we performed a proteomic *shotgun* analysis (Figure 5). Our results shown that tumor tissues have a higher number of identified proteins, detecting more than 4000 proteins in tumors of which 1573 are shared with non-tumor adjacent tissue but 2518 are exclusive (Figure 5A). To evaluate the implications of these proteins, we then selected only the exclusive proteins from tumors and established a threshold for the score of "peptide counting" above 5, resulting in a total of 271 proteins that we have classified according to their biological functions (Figure 5B). We have detected that most of tumor exclusive proteins have mitochondrial activity (23.6%) and the rest were mainly implicated in metabolism (16.2%), DNA/RNA processing (12.2%), and cytoskeleton (8.1%), vesicular trafficking (5.5%) and splicing (5.5%) among others pathways. A simplified list of the main proteins detected in tumor *vs.* normal mucosa, containing those with a score of "peptide counting" over 15, is shown in Figure 5C. All of them are included in the biological functions previously cited.

## DISCUSSION

Cancer includes several complex diseases that can affect different organs in the body. The recent report about global burden of cancer "GLOBOCAN 2018" estimates that it causes more than 9 millions of deaths per year worldwide<sup>[25]</sup>. Many medical and research efforts have been made trying to improve the treatments against this complex disease. Nevertheless, it is still necessary to study in depth the molecular, metabolic and proteomic features of cancer tissue to find the remaining answer of cancer problematic.

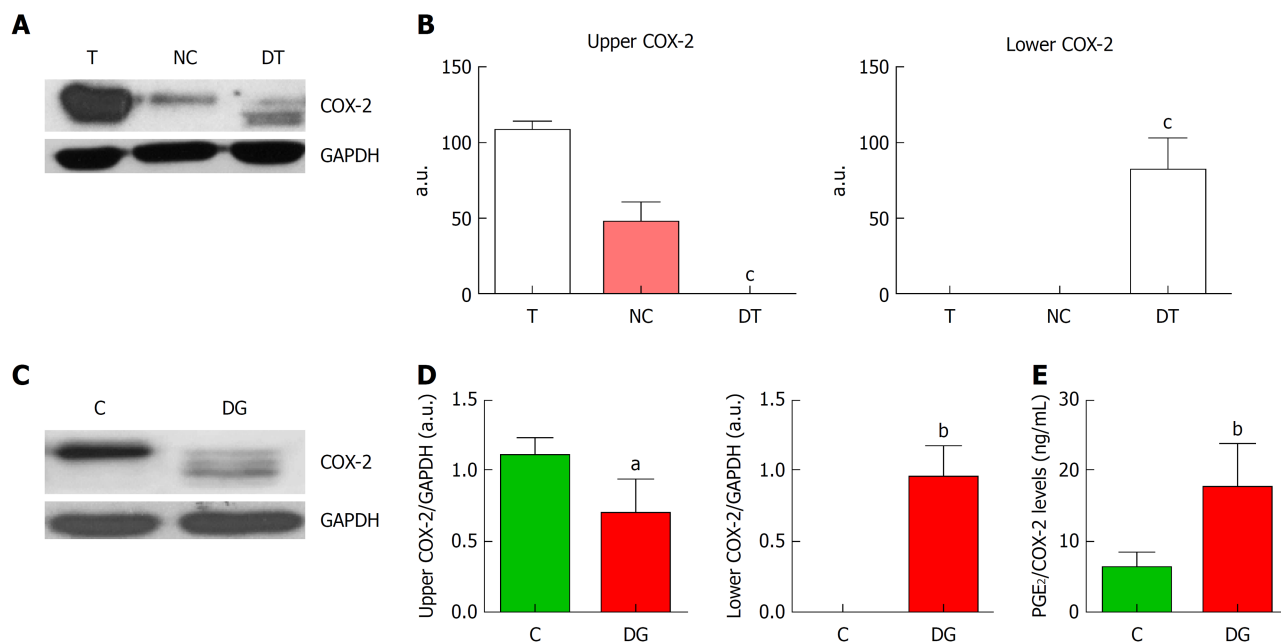
At the molecular level, numerous studies have related increased levels of COX-2 and derived prostanoids (mainly PGE<sub>2</sub>) to cancer development<sup>[26]</sup>. Thus, elevated COX-2 expression has been associated to larger cancer size, greater depth of cancer invasion and reduced patient survival<sup>[27,28]</sup>. Due to these findings, COX-2 was proposed as a potential molecular predictor for CRC progression. However, its predictive relevance is controversial because other studies have been unable to



**Figure 1 Analysis of cyclooxygenase-2 protein levels and prostaglandin E<sub>2</sub> measurement.** A: Representative image of western blot analysis of cyclooxygenase-1 (COX-1), cyclooxygenase-2 (COX-2) and 15-hydroxyprostaglandin dehydrogenase (15-PGDH) normalized by glyceraldehyde-3-phosphate dehydrogenase (GAPDH) of tumor (T) or non-tumor (N) samples. B and C: Graphs represent normalized COX-1 (B) or 15-PGDH (C) bands in western blot in T vs N tissue. D: Graph represent normalized COX-2 protein levels corresponding to upper (Up) or lower (Down) band in western blot in T vs N tissue. E: Graphs represent the percentage of samples expressing lower, upper, both or none bands in western blot for COX-2. F: Levels of PGE<sub>2</sub> in Tumor vs Non-tumor samples. G: Relative levels of prostaglandin E<sub>2</sub> only in samples from colorectal cancer patients showing the upper band of COX-2 in western blot. *n* = 45. Graphs show mean  $\pm$  SD. <sup>a</sup>*P*  $\leq$  0.05, <sup>b</sup>*P*  $\leq$  0.01 and <sup>c</sup>*P*  $\leq$  0.001 vs the non-tumor condition. COX-1: Cyclooxygenase-1; COX-2: Cyclooxygenase-2; 15-PGDH: 15-hydroxyprostaglandin dehydrogenase; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase; T: Tumor; N: Non-tumor; PGE<sub>2</sub>: Prostaglandin E<sub>2</sub>.

identify a significant correlation between CRC patient outcome and COX-2 expression mainly due to the high variability in COX-2 levels among CRC patients<sup>[29]</sup>. In our study, when we analyzed by Western blot the COX-2 levels in tumor tissues from CRC patients, we observed significantly higher levels of total protein and also a change in the expression profile compared to non-tumor colonic mucosa, comprising several bands. Although most reports in the literature refer to only one homogenous COX-2 protein band in animal tissue, some groups have reported the presence of several bands corresponding to COX-2 in tumors<sup>[30,31]</sup>. They proposed that this multiplicity of bands can be associated to post-translational modifications of the enzyme, mainly due to COX-2 glycosylation which can be related to several pathologies such as lung adenocarcinoma<sup>[32]</sup>. Interestingly, we detected non-glycosylated COX-2 protein in the majority of non-tumor mucosa samples, while glycosylated COX-2 protein was mainly detected in tumor tissue from CRC patients. These findings were confirmed by deglycosylation assays performed both in tumor tissue from CRC patients and in HT29 cells. Moreover, it is known that PGE<sub>2</sub> is the most abundant prostaglandin in human colon and the levels of PGE<sub>2</sub> are usually increased in colorectal neoplasia compared to normal tissue, which demonstrates its tumor expansion and metastatic functions. Interestingly, in our tumor samples we detected less PGE<sub>2</sub> compared to normal colonic mucosa. These results may be due to the presence of glycosylated COX-2, as previous work pointed out that post-translational modifications such as glycosylation of COX-2 can lead to a decrease in both protein levels and activity. In fact, the levels of PGE<sub>2</sub> were significantly lower in tumors that only present the upper COX-2 band, corresponding to those with a glycosylated state of the protein. Moreover, in spite of the high variability of the samples, we have also detected a tendency towards an increase in 15-PGDH content in tumors, while levels of EP4 receptors remained unaltered (data not shown). Thus, low levels of PGE<sub>2</sub> detected in tumor samples of our CRC cohort can be associated not only to COX-2 protein alterations, but also to an inhibitory scenario of the downstream signaling pathways.

Previous studies comparing the metabolic content of healthy and cancerous colon

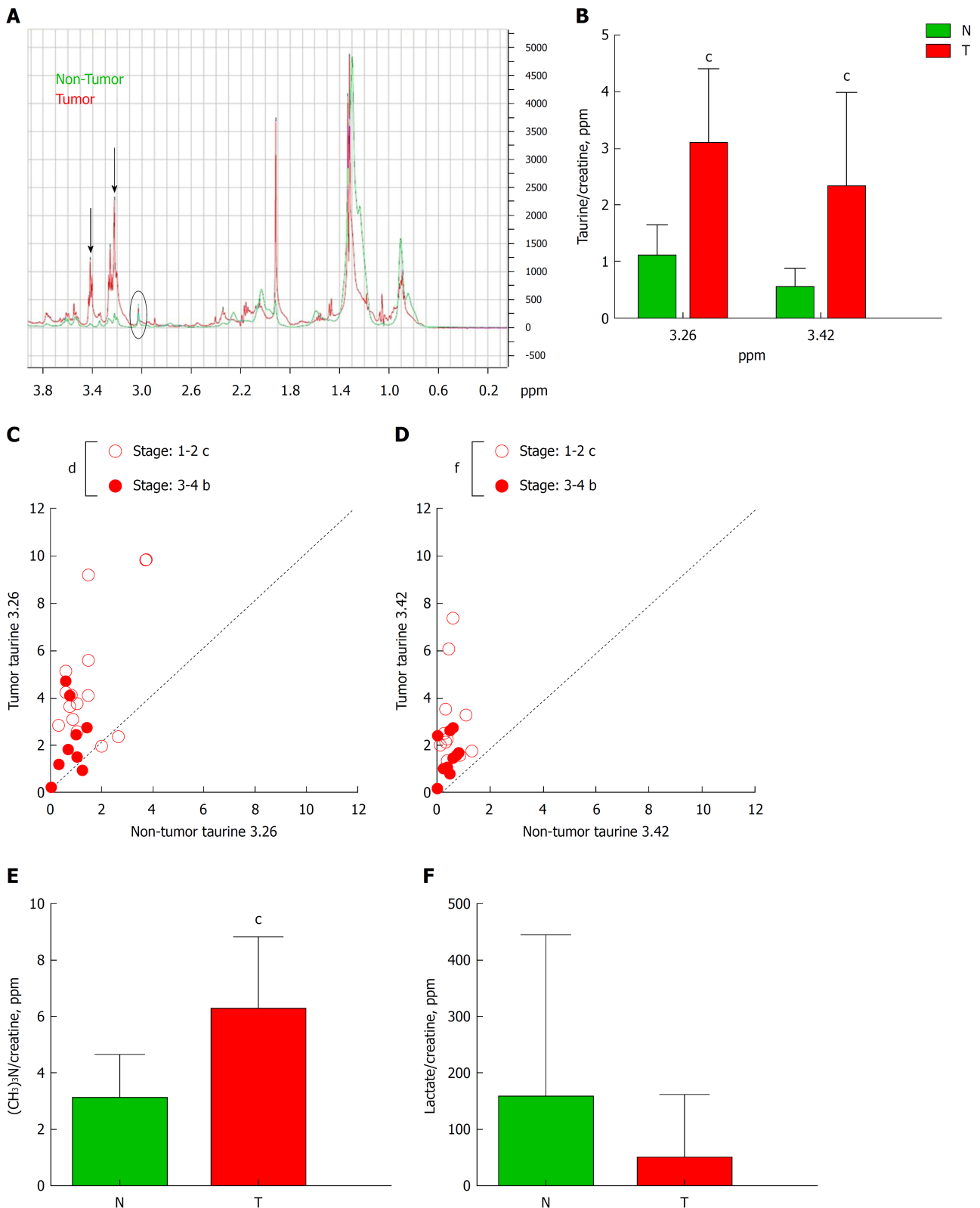


**Figure 2 Deglycosylation assays of cyclooxygenase-2.** A: Representative images of western blot of cyclooxygenase-2 (COX-2) protein levels corresponding to tumor tissue (T), negative control (NC) and deglycosylated tumor tissue (DT). B: Graphs correspond to upper or lower COX-2 bands quantifications.  $n = 4$ . C: Representative images of western blot of COX-2 protein levels in HT29 cells under basal conditions (C) or deglycosylated (DG). D: Graphs correspond to upper or lower COX-2 bands quantifications.  $n = 8$ . E: Relative levels of prostaglandin  $E_2$  (PGE $_2$ ) normalized by total COX-2 band obtained in the corresponding western blot analysis.  $n = 11$ . Graphs show mean  $\pm$  SD. <sup>a</sup> $P \leq 0.05$ , <sup>b</sup> $P \leq 0.01$  and <sup>c</sup> $P \leq 0.001$  vs the glycosylated condition. COX-2: Cyclooxygenase-2; T: Tumor; NC: Negative control; DT: Deglycosylated tumor; C: Cells under basal conditions; DG: Deglycosylated; PGE $_2$ : Prostaglandin  $E_2$ .

biopsies by HRMAS obtained from the same patient have demonstrated that adenocarcinomas are characterized by higher levels of taurine, glutamate, aspartate and lactate among others<sup>[18,33]</sup>. Accordingly, we have detected higher levels of taurine (both peaks) and phosphocholine in tumor samples whose peaks were missing in normal colonic mucosa. In contrast to the broadly described increase in the literature<sup>[34,35]</sup>, lactate levels in our cohort showed a decreasing trend in tumors that did not reach statistical significance due to the high variability between patients. Although some authors have proposed lactate as a non-invasive biomarker to determine malignancy for some kind of tumors<sup>[36]</sup>, further analyses in larger populations will be needed to establish the value of serum lactate to determine the response to therapy or early recurrence in CRC.

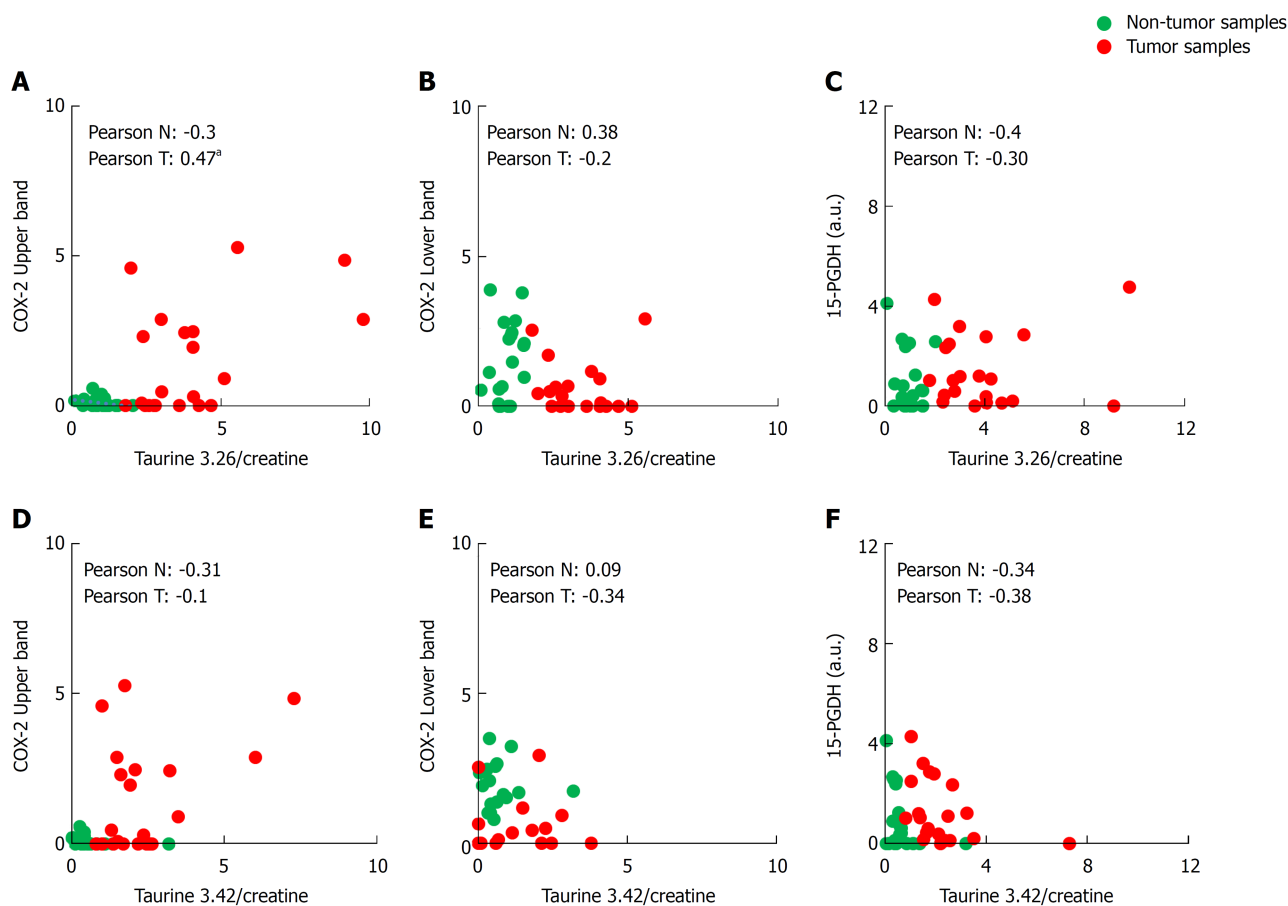
Furthermore, in the last years several proteomic studies of tissue samples obtained from CRC patients has allowed to compare the protein profile between tumor and the adjacent healthy mucosa<sup>[19,37]</sup>. It has been particularly effective in the discovery of prognostic tumor markers. However, these proteomic studies in tumors have also some disadvantages, mainly due to the limited availability of sample and the cellular heterogeneity of tumors. Our proteomic study revealed that tumor tissue has significantly more protein load and higher heterogeneity than its corresponding non-tumor pair. This fact has been recently described in several cancer cell lines, where it has been related to genomic aberrations on protein networks underlying physiological cellular activities<sup>[38]</sup>. Among these proteins, our study highlights some implicated in basic biological functions such as mitochondrial activity, processing of DNA and RNA, vesicular trafficking, cytoskeleton, metabolism and splicing. These findings are in agreement with previously published data in CRC and in other tumors<sup>[19,39,40]</sup>. Particularly, in our cohort of CRC patients, four proteins are missing in healthy mucosa but are highly expressed in tumor with a peptide counting more than 20. Interestingly, two of them are implicated in splicing, another specific hit is importin-5, which is a nuclear transport protein and the other is the apoptotic factor AIF-1. Recent studies have revealed multiple ways by which splicing is pathologically altered to promote the initiation and/or maintenance of cancer<sup>[41,42]</sup>. Furthermore, the importins may also play an important role in cancer by transporting key mediators of oncogenesis across the nuclear membrane in cancer cells. In fact, these proteins are usually overexpressed in multiple tumors including melanoma, pancreatic, breast, colon, gastric, prostate, esophageal, lung cancer and lymphomas<sup>[43]</sup>. Apoptotic pathways dysregulation has a direct relationship with cancer development<sup>[44]</sup>. In particular, it has been described that the pro-apoptotic factor AIF-1 shows higher expression in colorectal tumors<sup>[45]</sup>. In addition to this, it should be noted that among





**Figure 3 High resolution magic angle spinning analysis of tumor tissue.** A: Schematic diagram of high resolution magic angle spinning spectra of tumor tissue (red line) and non-tumor tissue (blue line). Black arrows indicate two peaks corresponding to taurine  $\delta$  ppm 3.26 and 3.42. B: Graph represents the quantification of both taurines normalized by creatine of tumor (T) and non-tumor (N) tissue. C and D: Correlation graphs of taurine 3.26 (C) and taurine 3.42 (D) in T vs N subdivided depending on the stage of the Tumor: 1-2 vs 3-4. E and F: Graphs represent the quantification of phosphocholine (E) or lactate (F) in T vs N.  $n = 29$ . Graphs show mean  $\pm$  SD. <sup>b</sup> $P \leq 0.01$ , <sup>c</sup> $P \leq 0.001$  of T vs N and <sup>d</sup> $P \leq 0.05$ , <sup>f</sup> $P \leq 0.001$  between stages in tumors. T: Tumor; N: Non-tumor.

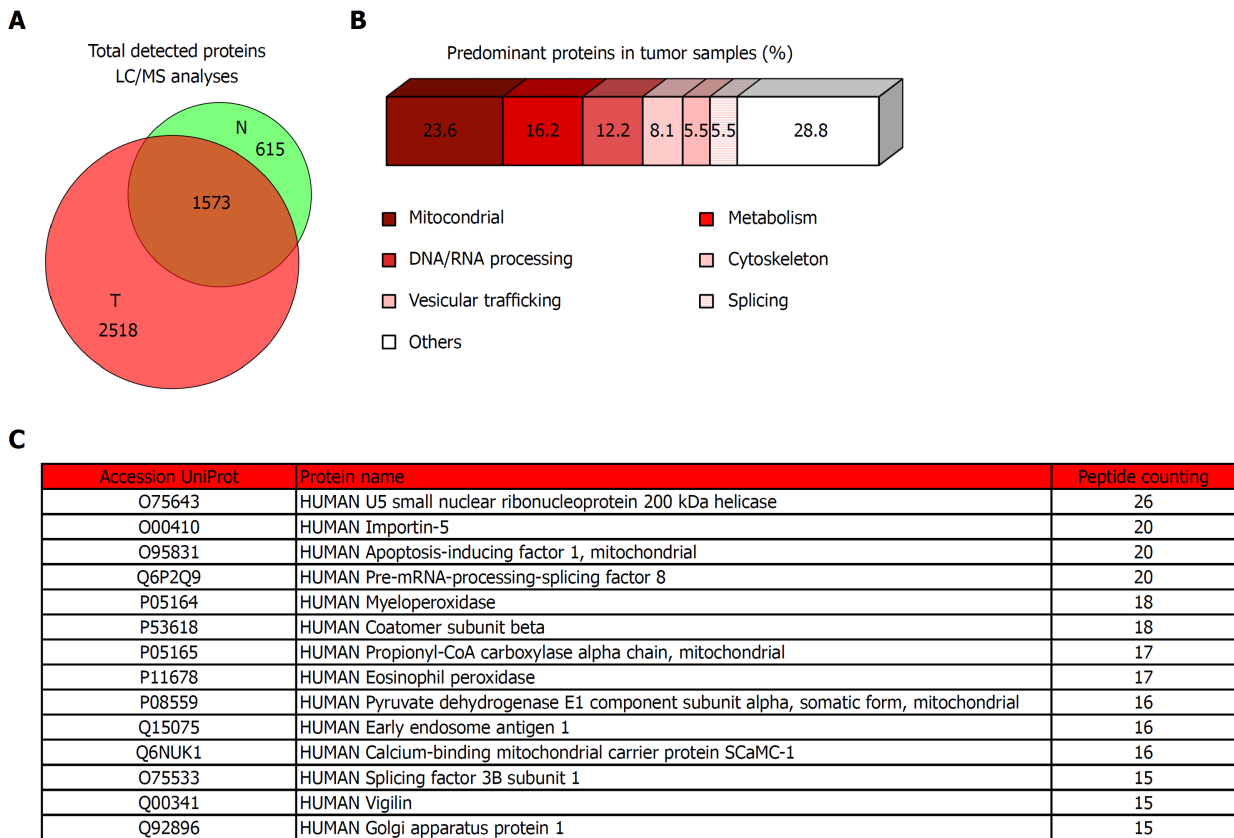
the more significantly upregulated proteins we found neutrophil-associated proteins such as myeloperoxidase and eosinophil peroxidase, which are involved in inflammation. This result is consistent with what we expected since cancer has a strong inflammatory component, which is known to play a significant role in cancer



**Figure 4** Biostatistical analysis of the correlation between the presence of cyclooxygenase-2 bands in western blot and both taurine peaks. A-F: Statistical analysis was performed searching the correlation coefficient between the presence of upper or lower cyclooxygenase-2 band or 15-hydroxyprostaglandin dehydrogenase in western blot and the taurine peaks (3.26 and 3.42  $\delta$  ppm) levels of 29 Tumor/Non-tumor pairs of samples. The Pearson's correlation coefficient is shown in each figure for non-tumor or tumor sample. <sup>a</sup> $P \leq 0.05$ . COX-2: Cyclooxygenase-2; 15-PGDH: 15-hydroxyprostaglandin dehydrogenase.

initiation, promotion and progression.

Our results relate by the first time post-translational COX-2 modifications with a metabolic and proteomic profile and can be useful in building new multivariate classifiers looking for a robust cross-talk among molecular, metabolic and proteomics data to improve the knowledge on CRC, thus contributing to establish new protocols for the diagnosis, prognosis and therapeutics for this cancer.



**Figure 5 Proteomic analysis of tumor tissue of colorectal cancer patients and the corresponding non-tumor adjacent tissue.** A: Venn diagram with specific and overlapped proteins. 2188 and 4091 proteins were identified (FDR < 1%) in non-tumor and tumor tissue, respectively. Among them, 1573 were identified in both tissues, and 615 and 2518 were unique in non-tumor and tumor tissue, respectively. B: Enrichment analysis of specific proteins identified in tumor tissue by mass spectrometry performed with proteins that were identified with 5 or more peptides (271 most abundant proteins). Most of these proteins had mitochondrial and metabolic function, or were involved in processes related to nucleic acids processing, cytoskeleton, vesicular trafficking, or splicing. C: Specific tumor tissue proteins identified with 15 or more peptides in liquid chromatography-mass spectrometry analyses. T: Tumor; N: Non-tumor; LC/MS: Liquid chromatography-mass spectrometry.

## ARTICLE HIGHLIGHTS

### Research background

Colorectal cancer (CRC) is the second most common cause of cancer death worldwide. It is broadly described that cyclooxygenase-2 (COX-2) is mainly overexpressed in CRC but very low is known about the nature and effects of the post-translational modifications of this enzyme within the tumor environment. Using metabolic and proteomic profile analyses which are essential for cancer prognosis and diagnosis, our hypothesis is that integrating these parameters with the study of COX-2 post-translational modifications in a high number of CRC patients may provide new insights on the implications of post-translational modifications for CRC prognosis and therapeutics.

### Research motivation

Many medical and research efforts have been made trying to better evaluate CRC progression. Nevertheless, it is still necessary to study in depth the molecular, metabolic and proteomic features to unravel the mechanisms leading to CRC progression and to provide a rationale for a personalized therapeutic approach.

### Research objectives

The aim of this study was to analyze the regulation of COX-2 in samples from patients with colorectal cancer and to perform a detailed analysis of both the metabolic profile and the proteomic content of the tumor tissue compared to the non-tumor adjacent area.

### Research methods

Both normal and tumor tissue obtained from CRC patients were processed and the protein levels of COX-2 were determined. Deglycosylation assays were performed in both cells and tumor samples before measuring prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) levels. Moreover, metabolic and proteomic profile in both samples types were carried out to complete the study.

### Research results

Tumor tissue of colorectal cancer patients of our cohort presents an altered COX-2 protein expression profile, which correspond to a glycosylated state of the protein. This was associated to a lesser PGE<sub>2</sub> production in tumors. Moreover, high resolution magic angle spinning (HRMAS) analysis indicated that tumor tissue exhibits increased levels of certain metabolites as taurine and phosphocholine and lower levels of lactate. Moreover, we detected an enlarged number of proteins in tumors that are mainly implicated in basic biological functions. Due to the high variability between patients, it will be necessary to analyze a large number of samples in order to achieve sufficient statistical power to find a biomarker that will be applicable in the future as a tool to improve both the diagnosis and prognosis of CRC patients.

### Research conclusions

In our colorectal cancer cohort, tumor tissue presents a differential COX-2 expression pattern compared to non-tumor and a lower activity of this enzyme. Moreover, this tissue showed an altered metabolic and proteomics profile that can be correlated to post-translational COX-2 modifications. The analysis of correlations between metabolic and molecular parameters in tumors of a high number of CRC patients could be useful for the understanding of the basis of this cancer in humans.

### Research perspectives

Our results relate by the first time post-translational COX-2 modifications with a metabolic and proteomic profile and can be useful in building new multivariate classifiers looking for a robust cross-talk among molecular, metabolic and proteomics data to improve the knowledge on CRC contributing to establish new protocols for the diagnosis, prognosis and therapeutics for this cancer. However, more samples should be analyzed in the future in order to discriminate a robust biomarker that can be useful in CRC diagnosis and prognosis.

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