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Prospects for a MERS-CoV spike vaccine

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Abstract

Introduction: Six years have passed since Middle East respiratory syndrome (MERS) coronavirus (MERS-CoV), a newly emerging infectious virus, was first reported in 2012. Although MERS-CoV has had a consistently high mortality rate in humans, no vaccines have been approved to prevent MERS-CoV infection in humans. MERS-CoV spike (S) protein is a key target for development of MERS vaccines.

Areas covered: In this review, we illustrate the structure and function of S protein as a vaccine target, describe available animal models for evaluating MERS vaccines, and summarize recent progress on MERS-CoV S-based vaccines, focusing on their ability to elicit antibody and/or cellular immune responses, neutralizing antibodies, and protection against MERS-CoV infection in different models. Prospects for future MERS-CoV S-based vaccines are discussed.

Expert commentary: The majority of MERS vaccines under development are based on MERS-CoV S protein, including full-length S, S1, and receptor-binding domain (RBD). While it is essential to evaluate the safety of full-length S and S1-based MERS vaccines, further improvement of the efficacy of RBD-based vaccines using novel strategies would be necessary. Overall, this review provides informative guidance for designing and developing safe and effective MERS vaccines based on viral S protein.

Keywords

MERS; MERS-CoV; Spike protein; Receptor-binding domain; Vaccine; Immune responses; Neutralizing antibodies; Protection

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1. Introduction

Middle East respiratory syndrome (MERS) coronavirus (MERS-CoV), a newly emerging infectious virus, was first identified in Saudi Arabia in June 2012 [1]. Since then, MERS-CoV has infected 2,229 humans, including 791 deaths (mortality rate ~35%), in at least 27 countries, including Saudi Arabia, South Korea, Iran, Jordan, Qatar, United Kingdom, United States, China, and Thailand [2]. Of note, Saudi Arabia has the largest number of MERS cases, covering 80% of all cases, while in 2015, South Korea reported the largest outbreak of MERS-CoV infection in humans outside Saudi Arabia [3,4]. MERS-CoV has been added to the WHO Blueprint list of priority viruses that cause public health emergency but still lack efficacious drugs and vaccines [5].

MERS-CoV is a zoonotic viral pathogen. It likely uses bats as its natural host, but MERS-CoV replicates in bats without showing clinical signs of disease [6,7]. Dromedary camels are evidenced as the intermediate host of MERS-CoV. The virus is able to transmit from camels to camels, and dromedary camels demonstrate high seropositivity to MERS-CoV [8-10]. Transmission of MERS-CoV from camels to humans occurs, and a number of risk factors, such as direct contact with MERS-CoV-infected dromedary camels, have been identified in camel workers [11–13]. Although MERS-CoV is less able to cause human-tohuman transmission compared to severe acute respiratory syndrome coronavirus (SARS-CoV), another coronavirus which led to the 2003 outbreak in humans [14] and is in the same β -genus as MERS-CoV, it does lead to human infections among healthcare workers and patients through healthcare or household-acquired infection [12,15,16]. MERS-CoV continues to cause cases and outbreaks in the Middle East, constituting an ongoing risk to global health security [13,17]. Therefore, effective medical countermeasures, such as vaccines and therapeutic agents, are urgently needed to block camel-to-camel and camel-tohuman transmission of MERS-CoV and to prevent and treat MERS-CoV infection in humans.

The MERS-CoV RNA genome encodes four structural proteins, including spike (S), membrane (M), nucleocapsid (N), and envelope (E) (Fig. 1A), as well as accessory open reading frame (ORF) proteins, such as non-structural proteins (nsp1–16) and ORF3, 4a, 4b, 5, and 8b [18,19]. Although several MERS-CoV proteins still have unknown functions in the process of MERS-CoV life cycle, infection, or pathogenesis, progress has been made on the structures and/or functions of MERS-CoV accessory proteins, some of which are directly or indirectly related to MERS-CoV pathogenesis [20,21]. For example, MERS-CoV accessory ORF3, 4a, 4b, and 5, as well as nsp16, proteins are necessary for viral infection and pathogenesis. The deletion or attenuation of ORF3–5 proteins dysregulates host responses and augments inflammation, while mutation of nsp16 significantly attenuates MERS-CoV infection in cell cultures and *in vivo* [22,23].

2. MERS-CoV S protein

Located at the MERS-CoV surface, S protein engages with viral cellular receptor dipeptidyl peptidase 4 (DPP4, e.g., CD26), mediating viral attachment to host cells and subsequent virus-cell membrane fusion [24–26]. MERS-CoV S protein consists of S1 and S2 subunits.

The receptor-binding domain (RBD), a functional domain in S1, is divided into a core subdomain and receptor-binding motif (RBM), while the latter contains the motif and key residues to bind DPP4 receptor on the host cell membrane [27,28]. After binding to cellular receptor, MERS-CoV S protein undergoes significant conformational changes, exposing heptad repeat 1 (HR1) and 2 (HR2) regions in S2 subunit, further forming a 6-helix bundle fusion core, leading to membrane fusion [29]. Recent structural studies on MERS-CoV S-trimer also suggest a potential mechanism for initiating the fusion process through receptor-induced triggering which involves sequential activation of protomers [30]. The schematic structure of MERS-CoV S protein and its functional regions are described in Figure 1B.

Theoretically, MERS-CoV S1 and S2 subunits, as well as their functional regions, such as RBD, play different roles in MERS-CoV infection and pathogenesis. Therefore, S1, RBD, and/or full-length S protein all serve as good targets to develop MERS-CoV vaccines and anti-MERS-CoV therapeutic antibodies, while S2, particularly its HR1 region, is an ideal target to develop anti-MERS-CoV fusion inhibitors [18,19]. Indeed, MERS-CoV S protein is able to induce strong antibody responses and/or cellular immune responses in immunized animals, in which S-specific neutralizing antibodies play a key role in preventing MERS-CoV infection [31–33]. It has been shown that an anti-MERS-CoV polyclonal antibody produced from S protein nanoparticle-immunized transchromosomic (Tc) cattle inhibits MERS-CoV infection in mice and demonstrates safety and tolerability in human Phase I trials [34,35]. Also, monoclonal antibodies or nanobodies targeting MERS-CoV S protein significantly protect against MERS-CoV infection in different animal models, including transgenic mice, rabbits, and non-human primates (NHPs) [18,36–38]. These studies highlight the importance of S protein-based vaccines and therapeutics in preventing and treating MERS-CoV infection. Previous reviews have summarized anti-MERS-CoV therapeutics targeting S protein [18]. This review will focus on the MERS-CoV S proteinbased vaccines. The key vaccine targets in MERS-CoV S protein are illustrated in Figure 1C.

3. Animal models available for evaluating the efficacy of MERS vaccines

MERS vaccines, including those based on the S protein, must be evaluated in suitable animal models to prove their efficacy before moving to large-scale production and/or human clinical trials. Other than dromedary camels, MERS-CoV has shown susceptibility to llamas, pigs, and rabbits, but not to goats, sheep and horses [36,39–41]. MERS-CoV may infect NHPs, including rhesus macaques and marmosets [42–44], although the two appear to have varying degrees of histopathological changes and pneumonia in the lungs after MERS-CoV infection [45]. Thus, NHPs can be used as effective animal models to evaluate protective immunity of MERS candidate vaccines.

MERS-CoV does not naturally infect small animals, including mice, ferrets, and hamsters. Mice expressing human DPP4 (hDPP4), including adenovirus 5 (Ad5)/hDPP4-transduced mice, hDPP4-Tg mice (transgenic for expressing hDPP4), and humanized (HuDPP4) mice (replacing mouse DPP4 with hDPP4), have been developed accordingly and are susceptible to wild-type MERS-CoV strains [46–50]. In addition, a mouse model was generated by modifying the mouse genome encoding two mouse amino acids into corresponding human

sequences in the DPP4 receptor, resulting in susceptibility to a mouse-adapted MERS-CoV and viral replication with lung symptoms and diseases [51]. Other mouse models, such as hDPP4-knock-in mice, were also developed, and they are susceptible to a mouse-adapted MERS-CoV clone, showing lung diseases [52]. Compared with large animal models, these small mouse models can be used as convenient and economical vehicles to evaluate the efficacy of MERS vaccines.

4. Advances in the development of MERS-CoV S protein-based vaccines

Like SARS and other viral vaccines, a good MERS vaccine should induce strong immune responses, elicit potent neutralizing antibodies, and demonstrate high protection in vaccinated animals and humans [14,19]. No MERS vaccine is approved for use in humans. Most currently developed vaccines against MERS-CoV are in preclinical stages, the majority of which are based on viral S protein [31,53–55]. These MERS-CoV S-targeting vaccines are summarized in Tables 1 and 2, and they are described later as vaccines based on the full-length S/S-trimer, S1, RBD, and other regions of MERS-CoV S protein, focusing on their ability to induce humoral and/or cellular immune responses and neutralizing antibodies, as well as protection against MERS-CoV infection, in different animal models.

4.1. MERS-CoV full-length S or S-trimer protein-based vaccines

A number of vaccines have been developed using full-length S protein of MERS-CoV as the target (Table 1). These vaccines are categorized as viral vector-, DNA-, nanoparticle-, virus-like particle (VLP)-, and S-trimer protein-based subunit vaccines [30,31,56–60]. Many of the reported full-length S-based vaccines have been tested in suitable animal models, and they demonstrated efficacy against MERS-CoV infection [31,53,57,61,62]. Two of these vaccines, e.g., a full-length S-based simian adenovirus vector vaccine (ChAdOx1) and a DNA vaccine (GLS-5300), are scheduled for clinical trials (Phase I) to test their safety and immunogenicity [63,64].

Viral vector-based vaccines encoding full-length S protein of MERS-CoV have been studied extensively, among which human or simian Ad, modified Vaccinia Ankara (MVA), measles virus (MV), and vesicular stomatitis virus (VSV) can be employed as the vaccine vehicles. For example, MERS-CoV S/RBD-specific systemic, mucosal, and/or cellular immune responses, as well as neutralizing antibodies against pseudotyped and live MERS-CoV, are induced in mice after immunizing them with human Ad5-based and Ad41-based or simian adenovirus vector (ChAdOx1)-based MERS-CoV full-length S-encoding vaccines [56,61,65,66], protecting hDPP4-Tg mice against MERS-CoV infection [61]. Also, MERS-CoV S-specific systemic and mucosal antibody responses and T cell responses, particularly neutralizing antibodies, are elicited in mice and/or camels immunized with MERS-CoV fulllength S-expressing MVA vaccines, protecting Ad5/hDPP4-transduced mice and dromedary camels against MERS-CoV infection [31,62,67]. It is noted that MERS-CoV S-specific immune responses and neutralizing antibodies are significantly improved by the Ad (ChAdOx1)-S vaccine priming and MVA-S vaccine boosting approach [65]. In addition, a full-length S-encoding MV vaccine induces MERS-CoV S-specific antibody and T cell responses, as well as MERS-CoV neutralizing antibodies, protecting Ad5/hDPP4-transduced

mice from MERS-CoV challenge [68]. Furthermore, a VSV-based MERS-CoV full-length S vaccine is shown to elicit MERS-CoV neutralizing antibodies and T cell responses in mice and/or NHPs [69].

Other types of MERS-CoV S-based vaccines, including those based on DNA, S-trimer protein, nanoparticle, and VLP, have shown immunogenicity and/or protective efficacy against MERS-CoV infection in mouse, camel, and NHP models [30,53,57–59]. For instance, an optimized DNA expressing full-length MERS-CoV S protein induces effective neutralizing antibodies and cellular immune responses in mice, camels and NHPs, protecting NHPs from MERS-CoV-caused disease [57]. Immunization of mice and NHPs with full-length S-DNA priming followed by S1 protein boosting elicits sufficient neutralizing antibodies, which protect NHPs against MERS-CoV-induced radiographic pneumonia [58]. A full-length S-based VLP vaccine induces MERS-CoV S-RBD-specific antibody responses and neutralizing antibodies in NHPs in the presence of Alum adjuvant [59]. A baculovirus insect cell-derived full-length S nanoparticle vaccine with Alum or Matrix-M1 adjuvant also induces MERS-CoV neutralizing antibodies in mice and Tc bovines, protecting Ad5/hDPP4-transduced mice from MERS-CoV infection [34,53,60].

Depending on antigen doses, injection doses, or immunization routes, MERS-CoV S/RBDspecific antibody and cellular immune responses and MERS-CoV neutralizing antibody titers induced by viral vectored full-length S vaccines might vary [61,62], while the titer of neutralizing antibodies elicited by other vaccine types, such as nanoparticle vaccines, could be significantly affected by adjuvants, not by antigen doses [53,60]. It is shown that some MERS-CoV full-length S-based DNA ($1:10^3$) or viral vector (MVA) ($1:10^3$) vaccines elicit a relatively higher titer of serum neutralizing antibodies than VLP ($<1:10^2$) or nanoparticle $(1:10^2-10^3)$ vaccines against live MERS-CoV infection [31,57,59,60,62,67]. Here, 50% tissue culture infectious dose (TCID₅₀) or 50% fluorescence-reduction neutralization assay (FRNA₅₀) is utilized to measure neutralizing antibodies (e.g. 50% or 90% neutralization titers). Therefore, the difference of neutralizing antibody titers induced by the aforementioned vaccines might result from variant neutralization assays tested, in addition to different neutralizing immunogenicity of vaccines, MERS-CoV strains, or virus titers. Currently, it appears that there is no international antibody standard to harmonize the data obtained from different neutralization assays. Surely, the breadth and potency of MERS-CoV neutralizing antibodies are improved by a prefusion MERS-CoV S-trimer protein with proline substitutions in the S2-HR1 region [30]. Notably, in addition to inducing MERS-CoV-specific immune responses and neutralizing antibodies, viral vectorbased full-length MERS-CoV S vaccines generally elicit anti-vector immune responses and/or neutralizing antibodies [31,62,65,68], a phenomenon that can be easily eliminated by S-based other vaccine types, such as DNA, S-trimer protein, nanoparticle, and VLP, as described earlier.

4.2. MERS-CoV S1-based vaccines

The MERS-CoV S1 subunit has potential as a vaccine target. Several vaccines have been constructed based on this region, which are categorized as viral vectored (Ad and rabies virus: RABV), DNA, and protein vaccines (Table 2). An Ad vector encoding MERS-CoV S1

extracellular domain (Ad5.MERS-S1) and a RABV vector encoding S1 elicit antibody responses and neutralizing antibodies in mice, while the RABV vectored S1 also protects Ad5/hDPP4-transduced mice from MERS-CoV challenge [56,70]. In addition, DNA-based MERS-CoV S1 vaccines elicit antibody and cellular immune responses and neutralizing antibodies capable of protecting the above mouse model from MERS-CoV infection [55,71]. Particularly, S1 protein boost of a full-length MERS-CoV S-DNA or S1 protein elicits similar levels of MERS-CoV neutralizing antibodies in mice, protecting NHPs from MERS-CoV challenge and pulmonary disease. However, a slightly lower titer of such antibodies is seen in S1/S1-vaccinated, compared to S-DNA/S1-vaccinated, NHPs, resulting in a higher peak volume of pulmonary disease in the S1/S1 group after MERS-CoV challenge [58]. Similar to MERS-CoV full-length S-based viral vectored vaccines, MERS-CoV S1-based viral vectored vaccines also induce a varying degree of anti-vector immune responses and neutralizing antibodies [70].

4.3. MERS-CoV RBD-based vaccines

As a short fragment inside the S1 subunit of MERS-CoV S protein, the RBD is applied as a key target for developing MERS vaccines in the categories of VLP and subunit vaccines, and their immunogenicity and efficacy have been tested in mouse, rabbits, and NHP models (Table 2).

One study indicates that a chimeric, spherical VLP (sVLP) expressing MERS-CoV RBD induces RBD-specific antibody and cellular immune responses in mice, neutralizing pseudotyped MERS-CoV entry into target cells [72]. Almost all other MERS-CoV RBDbased vaccines reported thus far are based on recombinant proteins expressed in mammalian or insect cell expression systems [32,73–77]. It has been demonstrated that a mammalian cell-expressed recombinant RBD protein containing residues 377-662 of MERS-CoV S induces robust humoral systemic and mucosal immune responses and neutralizing antibodies in immunized mice [32,78]. Studies have also shown that residues 358–588 and 367–606 of MERS-CoV RBD expressed in mammalian and insect cells, respectively, elicit RBDspecific antibody or cellular immune responses and neutralizing antibodies in mice and/or rabbits [79,80]. By optimizing and comparing five RBD fragments with different lengths, a RBD fragment containing residues 377–588 of MERS-CoV S protein is identified to induce the highest titer of antibody responses and neutralizing antibodies in immunized mice and rabbits with the capability of protecting Ad5/hDPP4 and hDPP4-Tg mice from MERS-CoV challenge with no evidence of immunological toxicity or eosinophilic immune enhancement [32,33,75,76,81]. In addition to its high immunogenicity and strong protection in mice and rabbits, MERS-CoV RBD is also immunogenic and protective in NHPs. This is evidenced by the induction of robust and sustained immune responses and neutralizing antibodies and protection against MERS-CoV challenge with minimal pathological effects in NHPs immunized with a RBD protein [73]. Importantly, MERS-CoV RBD protein-induced antibodies are shown to potently cross-neutralize multiple pseudotyped MERS-CoVs expressing S proteins of human and camel MERS-CoV strains, live human MERS-CoV strains, and mAb escape MERS-CoV mutants [74].

It should be noted that the immunogenicity of MERS-CoV RBD-based subunit vaccines is not significantly affected by antigen dosage, but by injection doses and vaccination intervals. For example, RBD dosage down to one microgram is sufficiently immunogenic to elicit strong immune responses in animals, and a regimen of two doses of RBD at 4-week intervals elicits the strongest antibody responses and neutralizing antibodies against MERS-CoV infection among one dose and two doses at 1-, 2-, and 3-week intervals tested, protecting hDPP4-Tg mice from MERS-CoV challenge [82,83]. Also, intranasal vaccination of the RBD protein induces much stronger local mucosal immune responses and neutralizing antibodies than subcutaneous immunization [84]. In particular, MERS-CoV RBD-elicited immune responses and neutralizing antibodies are significantly improved in the presence of suitable adjuvants, such as MF59, or fusion with appropriate immunopotentiators, such as Fc tag [32]. Compared with the wild-type RBD, a mutant RBD vaccine with a non-neutralizing epitope masked by a glycan probe provided 100% survival of hDPP4-Tg mice against lethal MERS-CoV infection [75]. These studies suggest that the RBD of MERS-CoV is a critical neutralizing domain and a strong immunogen for developing MERS vaccines.

4.4. Vaccines based on other regions of MERS-CoV S protein

As indicated earlier, most MERS-CoV vaccines under development focus on the full-length S protein, S1, or RBD. However, a few MERS vaccines are reported based on other regions of MERS-CoV S protein, such as truncated S, S-NTD, and S1/S2 regions (Table 2). For example, a MV-vectored MERS-S lacking the transmembrane domain (truncated S) is able to elicit MERS-CoV neutralizing antibodies, but the titer is slightly lower than that elicited by a full-length MERS-CoV S vaccine in the same viral vector [68]. Also, a MERS-CoV S-NTD-based protein vaccine is shown to induce favorable antibody and cellular responses, as well as neutralizing antibodies, reducing lung abnormalities in Ad5/hDPP4-transduced mice after MERS-CoV challenge [54]. In addition, a peptide spanning the S1/S2 regions, i.e., residues 736–761, has neutralizing activities by inhibiting pseudotyped MERS-CoV entry and membrane fusion, pointing out the possibility of developing vaccines covering this neutralizing epitope [85]. Nevertheless, the protective efficacy of such vaccines needs to be further confirmed in a lethal challenge model. At present, no vaccines are reported solely based on the MERS-CoV S2 subunit, potentially because of its low immunogenicity and inability to induce strong neutralizing antibodies.

5. Summary and Conclusions

This review introduces the structure and function of MERS-CoV S protein, summarizes current animal models available for evaluating the efficacy of MERS vaccines, and describes recent advances in MERS vaccines based on viral S protein, including those on the full-length S/S-trimer, S1, RBD, and other S regions. These S-based vaccines are further categorized as different types, including viral vector, DNA, VLP, nanoparticle, and protein-based vaccines, and their ability to elicit antibody and/or cellular immune responses, neutralizing antibodies, and protective efficacy against MERS-CoV infection was compared in different models. Such data will provide useful information and important guidance for designing and developing safe and effective MERS-CoV S protein-based vaccines.

6. Expert commentary

Although full-length S-based vaccines show protection against MERS-CoV infection and demonstrate promise for further development, no safety profile has been established. More specifically, some non-neutralizing immunodominant epitopes on the full-length S protein may compete with the neutralizing epitopes to attenuate neutralizing activity or even enhance MERS-CoV infection, potentially causing immunopathological effects or other harmful immune responses, as seen in the case of full-length S-based SARS vaccines [86–88]. In addition, MERS-CoV S1 subunit, which is much longer than the RBD, also contains some non-neutralizing immunodominant epitopes. Such immunodominant epitope in SARS-CoV S1 is shown to elicit epitope sequence-dependent enhancement of viral infection [89]. Therefore, evaluation of the safety and potential immunopathological consequences is essential for full-length S and S1-based MERS vaccines before moving them to large-scale development and beginning clinical trials.

Compared with vaccines based on other targets of MERS-CoV S, such as full-length S protein, RBD-based vaccines have a strong safety profile. The RBD does not contain nonneutralizing epitopes that may cause harmful immune responses, and RBD-based vaccines do not show immunological toxicity and immunopathological effects in the animals tested [81]. Similar to the SARS-CoV RBD, which covers multiple conformation-dependent neutralizing epitopes [90], the RBD of MERS-CoV also contains a critical neutralizing fragment with major neutralizing epitopes capable of inducing highly potent neutralizing antibodies [76]. like SARS-CoV RBD-based vaccines that induce high titers of crossneutralizing antibodies against divergent strains of human and animal SARS-CoV [91], MERS-CoV RBD-based vaccines also elicit broad-spectrum neutralizing antibodies and cross-protective immunity against infections of divergent MERS-CoV strains from humans, camels, and antibody escape mutants [74]. Future strategies can be applied to improve the potency and breadth of MERS-CoV RBD candidate vaccines by using structure-based rational design to mask unfavorable non-neutralizing immunodominant epitopes and expose neutralizing epitopes within the RBD [75], or incorporating RBD with favorable neutralizing epitopes in the S2 or non-RBD S1 regions. Such vaccination approaches are also expected to improve the efficiency of vaccines against escape antibody mutant virus strains [92].

In some cases, viral vectored vaccines encoding MERS-CoV S1 or S protein, such as RABV-MERS-S1 or MVA-MERS-S vaccines, induce anti-vector responses helpful in preventing host-derived RABV and camelpox virus in immunized mice or camels [31,70]. However, viral vectored vaccines, such as those based on Ad, against other viruses, including HIV, may induce a rapid memory immune response against the vector, enhance virus infection, or elicit limited efficacy in immunized hosts, resulting in early halting of clinical trials [93–95]. Thus, in addition to investigating immunogenicity and protection of viral-vectored MERS-CoV S candidate vaccines against MERS-CoV infection, careful design and selection of suitable viral vectors, comprehensive investigation of the possibility of anti-vector immunity in preventing MERS-CoV-specific immune responses, as well as extensive evaluation of their safety and potential toxicity, are needed before moving such vaccines forward for trials in humans.

Generally speaking, neutralizing antibodies are indispensable for preventing MERS-CoV infection, and the titers of neutralizing antibodies are correlated with protection [33,83]. Therefore, improvement of a vaccine's neutralizing immunogenicity to induce highly potent neutralizing antibodies would be the key for further development of next-generation MERS-CoV S-based vaccines. In addition to humoral immune responses, cellular immune responses or various cell populations may play different roles in fighting against MERS-CoV infection or regulating pathogenesis. For instance, airway memory CD4⁺ T cells mediate protective immunity against MERS-CoV, while CD8⁺ T cells and macrophages regulate MERS-CoV-induced pathology and clinical symptoms of the disease [96,97]. In this regard, cellular immunity of MERS-CoV S vaccines may well complement their humoral immunity to increase the vaccine's potency in preventing MERS-CoV infection.

As indicated earlier, MERS-CoV S protein serves as a key vaccine target. In addition to Sbased vaccines, MERS vaccines can be engineered by targeting other viral structural or nonstructural proteins, such as E, ORFs and nsp16, to generate mutant live virus vaccine platforms that lack E, delete ORF3–5, or mutate nsp16 [22,23]. These non-S-based vaccines could be applied as an alternative or complement of S-based vaccines to prevent MERS-CoV infection. Overall, in order to further improve immunogenicity and efficacy, S-based MERS vaccines can be used alone or combined with other types of S or non-S vaccines with complementary effects by priming-boosting vaccination approaches, or conjugating with different adjuvants, and optimizing for doses, routes, or intervals. Such vaccines should be tested in different animal models, including large animal models, such as NHPs and/or camels, to confirm immunogenicity, efficacy, toxicity, and immunopathology before processing to human clinical trials.

7. Five-year view

Six years have passed since MERS-CoV was first reported in 2012. In addition to two candidate vaccines that are scheduled for clinical trials, most MERS-CoV vaccines, including those based on viral S protein, are still in preclinical development or not even processed for testing in large animal models, such as NHPs. Lack of sufficient funds to support such studies is potentially one of the main reasons, in addition to regulations and other issues. A number of MERS-CoV S-based vaccine candidates, such as those based on the RBD, with strong safety profile, immunogenicity, and protective efficacy, but without inducing immunopathology in animals, are expected to also induce potent immune responses and efficacy, as well as maintain good safety profile, in humans. Therefore, it is anticipated that sufficient funds will be invested from governments, private entities, or big pharmaceutical companies and that an increasing number of MERS vaccines, particularly those based on the S and/or RBD proteins, can be tested in large animals, moved to clinical trials, and/or approved to prevent MERS-CoV infection in humans in the next five years.

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* of interest

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8. Key issues

- MERS-CoV, a newly emerging infectious coronavirus, continues to infect human populations, particularly in Saudi Arabia, with about 35% mortality rate. As a zoonotic virus, MERS-CoV transmits efficiently among camels and occasionally in people who are in close contact with infected camels and persons, constituting a continual threat to global public health and thus requiring effective medical countermeasures, such as vaccines, to prevent MERS-CoV infection.
- MERS-CoV genome encodes four structural proteins, including S, E, M and N, and a number of accessory proteins. The structures and/or functions of some of these proteins, such as S, ORF3–5, and nsp16, have been extensively studied.
- MERS-CoV S protein consists of S1 and S2 subunits. RBD in S1 is responsible for cellular receptor binding, while HR1 and HR2 regions in S2 mediate virus fusion and entry into the target cell. Receptor binding to the RBD may trigger sequential activation of protomers.
- MERS-CoV can infect dromedary camels, but it does not naturally infect other large animals (e.g., sheep, horses) and small animals (e.g, mice, ferrets and hamsters). Large animal models, including non-human primate models, and small animal models, such as adenovirus/hDPP4-transduced mice, hDPP4-transgenic mice, and hDPP4-knock-in mice, have been developed for testing protective efficacy of MERS vaccines.
- S protein is the primary target for the humoral immune response during infection and thus a key target for developing MERS-CoV vaccines. S-RBD contains a critical neutralizing domain capable of inducing highly potent neutralizing antibodies. S- and RBD-specific neutralizing antibodies play an essential role in preventing MERS-CoV infection, and neutralizing antibody titers are correlated with protection.
- No MERS vaccines have been approved for use in humans. Two MERS-CoV S-based vaccines are scheduled for clinical trials, and all other MERS vaccines are in preclinical development.
- Most MERS vaccines under development are based on viral S protein, including the full-length S/S-trimer, S1, RBD, and other S regions. These vaccines induce MERS-CoV-specific antibody and/or cellular immune responses, in addition to neutralizing antibodies at varying titers, a number of which have been tested in mouse, camel, or NHP models with protective efficacy.
- Each vaccine category has advantages and potential limitations. By comparison, MERS-CoV RBD-based subunit vaccines elicit relatively higher titers of neutralizing antibodies against multiple virus strains, protecting transgenic mice from MERS-CoV challenge without causing

immunopathological effects, demonstrating strong safety profile. Clinical trials are warranted if sufficient funds are available.



Figure 1.

MERS-CoV and S protein structures and vaccine targets in MERS-CoV S protein. (A) Schematic structure of MERS-CoV and its key structural proteins, including S, N, M, and E proteins. (B) Schematic structure of MERS-CoV S protein and its functional regions. S protein is composed of S1 and S2 subunits. NTD, N-terminal domain. RBD, receptorbinding domain. RBM, receptor-binding motif. FP, fusion peptide. HR1 and HR2, heptad

repeat 1 and 2 regions. TM, transmembrane. CP, cytoplasmic tail. (C) Key vaccine targets in MERS-CoV S protein and its fragments.

Table 1.

Summary of vaccines based on MERS-CoV full-length S or S-trimer proteins^a

Vaccines	Categories	Antibody responses and neutralizing antibodies	Cellular immune responses	Protection	Ref.
Viral vector-based	Ad (human Ad5/ Ad41; simian Ad: ChAdOx1)	Induces MERS-CoV S/ RBD-specific systemic and mucosal antibody responses and neutralizing antibodies in mice; neutralizes pseudotyped and live MERS-CoV (EMC2012, most <1:10 ³) and vector Ad	Elicits MERS-CoV S-specific T cell responses in mice	Protects hDPP4- Tg mice from MERS-CoV (EMC2012) challenge; Phase I trial (ChAdOx1)	[56,61,64-66]
	MVA	Induces MERS-CoV S- and/or vector MVA- specific systemic and/or mucosal antibody responses in mice and camels; induces neutralizing antibodies against live MERS-CoV (EMC2012, 1:10 ³), MVA, and camelpox virus	Elicits MERS-CoV S and vector MVA-specific T cell responses in mice	Protects Ad5/ hDPP4-transuced mice and camels from MERS-CoV (EMC2012) challenge; protection correlates with serum neutralizing antibodies	[31,62,65,67]
	MV	Induces MERS-CoV S- and vector MV-specific antibody responses and neutralizing antibodies in mice; neutralizes live MERS-CoV (EMC2012, <1:10 ³)	Elicits MERS-CoV S- and vector MV-specific T cell responses in mice	Protects Ad/ hDPP4-transduced mice from MERS- CoV (EMC2012) challenge	[68]
	VSV	Induces MERS-CoV S- specific antibody responses and neutralizing antibodies in mice and NHPs; neutralizes pseudotyped MERS- CoV	Elicits MERS-CoV S-specific T cell responses in NHPs	Not reported	[69]
DNA-based	DNA	Induces MERS-CoV S- specific antibody responses and neutralizing antibodies in mice, camels, and NHPs; neutralizes divergent pseudotyped and/or live MERS-CoV (human/1390, camel/31, camel/39, EMC2012, 1:10 ² –10 ³)	Elicits MERS-CoV S-specific T cell responses in mice and NHPs	Protects NHPs against MERS- CoV (EMC2012) challenge, without demonstrating clinical or radiographic signs of pneumonia; Phase I trial	[55,57,63]
DNA/protein-based	DNA prime/ protein (S1) boost	Induces persistent MERS-CoV S- targeting antibody responses and neutralizing antibodies in mice and/or NHPs; neutralizes divergent pseudotyped and live MERS-CoV (Jordan- N3)	Not reported	Protects NHPs from MERS-CoV (Jordan-N3) challenge and virus-induced radiographic pneumonia	[58]

Vaccines	Categories	Antibody responses and neutralizing antibodies	Cellular immune responses	Protection	Ref.
VLP-based	VLP	Induces MERS-CoV S- RBD-specific antibody responses and neutralizing antibodies (1:40) in NHPs	Elicits MERS-CoV S-RBD- specific T cell responses in NHPs	Not reported	[59]
Nanoparticle-based	Nanoparticle	Induces MERS-CoV S- specific antibody response and neutralizing antibodies (1:10 ² -10 ³) in mice and Tc bovines; neutralizes live MERS- CoV (Jordan)	Not reported	Protects Ad5/ hDPP4-transduced mice from MERS- CoV (Jordan or EMC2012) challenge	[34,53,60]
S-trimer protein-based	Subunit	Induces neutralizing antibodies in mice; neutralizes variant pseudotyped MERS- CoV	Not reported	Not reported	[30]

^{*a*}Ad, adenovirus; ChAdOx1, simian adenovirus vector; hDPP4, human dipeptidyl peptidase 4; hDPP4-Tg: hDPP4-transgenic; MERS-CoV, Middle East respiratory syndrome coronavirus; MV, measles virus; MVA, Modified Vaccinia Ankara; NHPs, non-human primates; RBD, receptor-binding domain; S, spike; Tc, transchromosomic; VLP, virus-like particle; VSV, vesicular stomatitis virus. MERS-CoV strains used for neutralization and challenge studies, as well as vaccine-induced neutralizing antibody titers, are shown in parentheses.

Table 2.

Summary of vaccines based on MERS-CoV S fragments^a

Vaccines	Categories	Antibody responses and neutralizing antibodies	Cellular immune responses	Protection	Ref.	
Vaccines based on MERS-CoV S1				•		
Viral vector-based	Ad (Ad5)	Induces MERS-CoV S- specific antibody responses and neutralizing antibodies (>1:10 ³) in mice; neutralizes live MERS-CoV (EMC2012)	Not reported	Not reported	[56]	
	RABV	Induces MERS-CoV S- and vector RABV-specific antibody responses and neutralizing antibodies (>1:10 ³) in mice; neutralizes live MERS-CoV (Jordan-N3)	Not reported	Protects Ad5/ hDPP4- transuced mice from MERS-CoV (Jordan-N3) challenge	[70]	
DNA-based	DNA	Induces MERS-CoV S/ RBD-specific antibody responses and neutralizing antibodies (1:10 ² -10 ⁴) in mice; neutralizes live MERS-CoV (EMC2012, human/1390, camel/31, and camel/39)	Elicits MERS-CoV S-RBD- specific T cell responses in mice	Protects Ad5/ hDPP4- transuced mice from MERS-CoV (EMC2012) challenge	[55,71]	
Protein-based	Subunit	Induces MERS-CoV S- targeting antibody responses and neutralizing antibodies in mice and NHPs; neutralizes divergent pseudotyped and live MERS-CoV (Jordan-N3)	Not reported	Protects NHPs from MERS-CoV (Jordan-N3) challenge	[58]	
Vaccines based on MERS-CoV RBD						
VLP-based	VLP	Induces MERS-CoV S- RBD-specific	Elicits S-RBD-specific T cell responses in mice	Not reported	[72]	

Vaccines	Categories	Antibody responses and neutralizing antibodies	Cellular immune responses	Protection	Ref.			
Vaccines based on MERS-CoV S1	Vaccines based on MERS-CoV S1							
		antibody responses and neutralizing antibodies in mice; neutralizes pseudotyped MERS-CoV						
Protein-based (residues 358–588)	Subunit	Induces MERS-CoV- specific antibody responses and neutralizing antibodies (>1:10 ³) in rabbits	Not reported	Not reported	[79]			
Protein-based (residues 377–588)	Subunit	Induces MERS-CoV S/ RBD-specific antibody responses and neutralizing antibodies in mice and rabbits; neutralizes divergent strains of pseudotyped and live MERS-CoV (EMC2012, London1– 2012, >1:10 ³)	Elicits MERS-CoV S-RBD- specific T cell responses in mice	Protects Ad5/ hDPP4- transduced mice and hDPP4-Tg mice from MERS-CoV (EMC2012) challenge without causing immuno- pathological effects	[32,33,74–76,81–83]			
Protein-based (residues 377–662)	Subunit	Induces MERS-CoV S/ RBD-specific systemic and mucosal antibody responses and neutralizing antibodies in mice; neutralizes pseudotyped and live MERS-CoV (EMC2012, 1:10 ² -10 ³)	Induces MERS-CoV S- specific T cell responses in mice	Not reported	[77,78,84]			
Protein-based (residues 367–606)	Subunit	Induces MERS-CoV S- RBD-specific antibody responses and neutralizing antibodies in mice and/or NHPs; neutralizes pseudotyped and live MERS-CoV	Elicits MERS-CoV S-RBD- specific T cell responses in mice	Partially protects NHPs from MERS-CoV (EMC2012) challenge	[73,80]			

Vaccines	Categories	Antibody responses and neutralizing antibodies	Cellular immune responses	Protection	Ref.
Vaccines based on MERS-CoV S1	-				
		(EMC2012, <1:10 ³)			
Vaccines based on other S regions	of MERS-Co	V, including trunca	ated S, S-NTD, and S1/S2		
Viral vector-based (truncated S)	MV vector	Induces MERS-CoV S- and vector MV- specific antibody responses and neutralizing antibodies in mice; neutralizes live MERS-CoV (EMC2012, <1:10 ³) and MV	Elicits MERS-CoV S- and vector MV-specific T cell responses in mice	Protects Ad/ hDPP4- transduced mice from MERS-CoV (EMC2012) challenge	[68]
Protein-based (S-NTD)	Subunit	Induces MERS-CoV S- NTD-specific antibody responses and neutralizing antibodies (1:40–10 ²) in mice and rabbits; neutralizes live MERS-CoV (EMC2012)	Elicits MERS-CoV S-NTD- specific T cell responses in mice	Protects Ad5/ hDPP4- transduced mice from MERS-CoV (EMC2012) challenge	[54,79]
Protein-based (S1/S2: residues 736–761)	Subunit	Induces low- titer neutralizing antibodies in rabbits; inhibits pseudotyped MERS-CoV entry and membrane fusion	Not reported	Not reported	[85]

^aAd, adenovirus; hDPP4, human dipeptidyl peptidase 4; hDPP4-Tg: hDPP4-transgenic; MERS-CoV, Middle East respiratory syndrome coronavirus; MV, measles virus; NHPs, non-human primates; RABV, rabies virus; RBD, receptor-binding domain; S, spike; S-NTD, spike protein N-terminal domain; VLP, virus-like particle. MERS-CoV strains used for neutralization and challenge studies, as well as vaccine-induced neutralizing antibody titers, are shown in parentheses.