



Published in final edited form as:

EMS Cancer Sci J. 2018 ; 1(1): .

Therapeutic Considerations for Ron Receptor Expression in Prostate Cancer

Nicholas E. Brown¹, Camille Sullivan¹, and Susan E. Waltz^{1,2,*}

¹Department of Cancer Biology, University of Cincinnati College of Medicine, Cincinnati, OH 45267, USA

²Research Service, Cincinnati Veterans Affairs Medical Center, Cincinnati, OH 45267, USA

Abstract

Introduction: The Ron receptor tyrosine kinase was initially discovered as a protein which played a critical role in regulating inflammatory responses. This effect was primarily determined through studies in various macrophage populations. Since its initial discovery, a role has emerged for Ron as a driver of cancer within epithelial cells. After numerous publications have detailed a role for Ron in promoting tumor initiation, growth, and metastasis, Ron has been designated as an emerging therapeutic option in a variety of cancers.

Areas Covered: This review discusses the current literature regarding the role of Ron in prostate cancer and places special emphasis on the role of Ron in both epithelial cells and macrophages. Whole body loss of Ron signaling initially exposed a variety of prostate cancer growth mechanisms regulated by Ron. With the knowledge that Ron plays an integral part in regulating the function of epithelial cells and macrophages, studies commenced to discern the cell type specific functions for Ron in prostate cancer. A novel role for Ron in promoting Castration Resistant Prostate Cancer has recently been uncovered, and the results of these studies are summarized herein. Furthermore, this review gives a summary of several currently available compounds which show promise at targeting Ron in both epithelial and macrophage populations.

Outlook: Sufficient evidence has been provided for the initiation of clinical trials focused on targeting Ron in both macrophage and epithelial compartments for the treatment of prostate cancer. A number of therapeutic avenues for targeting Ron in prostate cancer are currently available; however, special consideration will need to take place knowing that Ron signaling impacts multiple cell types. Further understanding of the cell type specific functions of Ron in prostate cancer will help inform and shape future clinical research and therapeutic strategies.

Keywords

receptor tyrosine kinase; RON receptor; prostate cancer; hepatocyte growth factor-like protein

*Address correspondence to: Susan E. Waltz, PhD, Department of Cancer Biology, Vontz Center for Molecular Studies, University of Cincinnati College of Medicine, 3125 Eden Ave, Cincinnati, OH 45267-0521, Tel: 513.558.8675, susan.waltz@uc.edu.

Disclosure of potential conflicts of interest: The authors declare no conflicts of interest.

1. Introduction

Receptor tyrosine kinases are becoming increasingly prevalent medicinal targets as new information is uncovered revealing their importance in multiple diseases. Specifically, in cancer, receptor tyrosine kinases have been implicated as drivers of disease and there has been success targeting these proteins, such as the use of the monoclonal antibody trastuzumab to target HER2 in breast cancer (1). One such receptor that is receiving increased attention recently due to seminal findings regarding its crucial role in multiple cancers, such as pancreatic, breast, and prostate, is the Ron receptor/MST1R. The Ron receptor is a member of the Ron and c-Met family of cell surface receptor tyrosine kinases and is primarily expressed on epithelial cells and macrophages, although low levels of expression have been detected in other cell types (2–4). Ron and c-Met are the only two members of this family and the two receptors share some similarities in structure and function. Despite these similarities, a number of specific roles have emerged between the two receptors.

The only known ligand for Ron is Hepatocyte Growth Factor Like protein (HGFL), named due to the structural similarities to the ligand for c-Met, HGF, and the two are believed to have evolved from a common ancestor (5). HGFL is produced primarily by hepatocytes and secreted in the blood in a pro-form. Following cleavage, HGFL forms a heterodimer capable of binding to Ron (6–8). Both the Ron receptor and HGFL have been highly associated with multiple cancers (9–12). Specifically, in prostate cancer Ron was highly expressed in over 85% of primary tumors and in 100% of prostate cancer metastasis (13). The exceptionally high correlation with disease progression is one reason why Ron is the focus of numerous studies for the treatment of prostate cancer. With approximately 30,000 deaths annually in the United States from prostate cancer, the identification of novel targets to treat this disease is a crucial task that needs to be completed, and the Ron receptor is an up and coming therapeutic option (14).

1.1 Ron Structure and Function

The Ron receptor is located on chromosome 3p21.31 in humans and has homologs in several other organisms, such as rat (15), chicken (16), feline (17), mouse (15, 18), xenopus (19) and zebrafish (20). Structurally, the Ron receptor originates as an 185kDa precursor protein, which is cleaved into a 35 kDa extracellular alpha chain that is disulfide linked to a 150 kDa transmembrane beta chain. The extracellular portion of Ron contains a Sema-PSI domain required for ligand binding, while the intracellular portion possesses the kinase domain responsible for signal transduction (21). Ron activation results in receptor dimerization leading to autophosphorylation of kinase domain residues Y1238 and Y1239 and subsequent phosphorylation of Y1353 and Y1360, which induces the activation of multiple downstream signaling cascades (22). Recently, it has been discovered that the intracellular portion of Ron contains an acidic region of the juxtamembrane domain responsible for auto-inhibition; however, phosphorylation of Y1198 in the kinase domain relieves this inhibition and facilitates activation (23). A number of splicing and truncation variants have been identified for Ron, which produce differing effects on function/activation of the receptor (24). One isoform of Ron, known as short form Ron (sf-Ron/ RON 55), is heavily prevalent in

pancreatic cancer and is constitutively phosphorylated, has transforming activity, and is resistant to many therapies targeting the extracellular portion of Ron (25). Structural variants, such as sf-Ron, will need to be taken into consideration during the development of therapeutics targeting Ron signaling.

Initially the ligand for Ron, HGFL, was identified as a protein which induced changes in macrophage shape and spreading (26). Further work indicated that not only did HGFL treatment impact mechanical characteristics of the cell, but it also limited inflammatory responses. HGFL treatment of macrophages reduced nitric oxide production following treatment with a variety of stimuli (27). Shortly thereafter, crosslinking studies were performed to determine that HGFL was binding to Ron at the cell surface (28). Ron was further implicated in mediating inflammatory response as it was determined that Ron signaling deficient mice (TK^{-/-}, lacking Ron tyrosine kinase domain) have a defect in the ability to regulate nitric oxide levels and incur greater tissue damage following inflammatory responses (29). The regulation of inflammatory responses through Ron signaling is a critical aspect of effective wound healing.

Ron has been shown to have both HGFL-dependent and HGFL-independent functions. Overexpression of Ron specifically in the mammary epithelium was sufficient to drive breast cancer, although, overexpression of Ron in the mammary epithelium of HGFL knockout mice produced tumors with a significant delay in mammary tumor formation (30). In this context, HGFL loss in the tumors altered cell signaling patterns, with decreased NF- κ B activation and reduced β -catenin expression. It is interesting to note that genetic knockout of HGFL did not completely prevent tumor formation, indicating that there are HGFL independent functions of Ron that remain oncogenic when Ron is overexpressed. In another breast cancer study, HGFL independent functions of Ron were reported that enhanced cell spreading and survival (31). These reports suggest that Ron activation may function in the absence of HGFL possibly through a yet to be discovered ligand or through cross talk with other receptors. The ideal candidate as an alternative ligand for Ron would be HGF, however, despite having similar structural domains as HGFL, HGF does not appear to activate Ron (32). Ron activation through potential alternative ligand(s) is a current area of investigation. Receptor crosstalk has been reported between Ron and a number of receptors, such as c-Met (22), PDGFR- β (33), IGF1-R (34), Plexins (35) and EGFR (36), making receptor cross-talk another viable option to explain HGFL independent Ron oncogenic function.

2. Ron in Prostate Cancer

2.1 Ron-Dependent Signaling Mechanisms in Prostate Cancer

Numerous studies within the past decade have expanded upon the role of the Ron receptor in prostate cancer. Our group was the first to demonstrate that Ron is critical for prostate tumor growth (13, 37). We showed that whole-body genetic ablation of Ron signaling in the Transgenic Adenocarcinoma of Mouse Prostate (TRAMP) mouse model leads to decreased prostate tumor growth (37). Accompanying this research was the discovery of several tumor cell-intrinsic processes regulated by Ron to promote prostate cancer. Prostate tumors isolated from Ron-deficient TRAMP mice exhibited markedly increased tumor apoptosis

and decreased microvessel density compared with controls (37). Similarly, loss of HGFL in TRAMP mice led to increased prostate tumor cell death, and this was at least in part due to down regulation of a Ron-STAT3-Bcl2-dependent pro-survival mechanism (38). Thus, Ron signaling provides a survival advantage to prostate tumor cells. Interestingly, neither loss of Ron nor HGFL in TRAMP mice led to changes in tumor cell proliferation, whereas loss of Ron in the polyoma middle T antigen model of breast cancer led to a significant decrease in cellular proliferation (37–39). These studies suggest that Ron provides different oncogenic functions across cancers from different tissues.

Ron has also been identified as an important regulator of tumor-associated macrophages (30, 38, 40, 41). Whole-body loss of Ron signaling in mouse models of breast and prostate cancer leads to increased localization of F4/80-positive macrophages within tumors coupled with altered expression of macrophage activation markers (30, 38). This suggests Ron regulates macrophage recruitment and activation within the tumor microenvironment to promote cancer. As these *in vivo* studies cannot differentiate the contributions of Ron signaling in individual cell compartments to modulation of tumor-associated macrophages, further investigation into cell type-specific mechanisms is necessary to gain a full understanding of the complex roles of Ron in maintaining a pro-tumorigenic microenvironment.

2.2 Epithelial-Specific Roles for Ron in Prostate Cancer

Many studies have demonstrated the importance of epithelial-specific Ron signaling in supporting tumorigenesis in a variety of epithelial cancers. Loss of Ron signaling diminishes tumorigenic activities of several established and primary thyroid (42), colorectal (43–45), and pancreatic (46) cancer cell lines. Furthermore, epithelial-specific overexpression of Ron in the lung, breast, and pancreas induces adenocarcinomas with metastasis in mammary and pancreatic models (47–49). Recent work has similarly uncovered the functions of prostate cancer epithelial specific Ron signaling. Knockdown of Ron in human prostate cancer cell xenografts in immunodeficient mice revealed that loss of Ron in prostate tumor epithelial cells significantly reduces tumor growth (13). Conversely, overexpression of Ron in prostate epithelial cells is sufficient to induce prostate cancer in mice (50). Taken together, this work demonstrates that epithelial Ron expression promotes murine and human prostate tumor growth *in vivo*. Interestingly, Ron overexpression selectively within prostate epithelial cells was associated with changes in both cell proliferation and cell death (50). This contrast in phenotype between whole-body and cell type-specific modulation of Ron signaling may suggest Ron signaling across multiple cell types communicates within a tumor, however further examination is needed to delineate these mechanisms.

While the role of Ron in cancer cell metastasis has been characterized in several cancers, such as breast (12, 39, 49, 51–53) pancreatic (9, 34, 54), and lung (55), few studies have addressed the importance of Ron in prostate cancer cell migration, invasion, and metastasis. Initial *in vitro* studies have suggested an important role for epithelial Ron in regulating some of these phenotypes. Jiang *et al.* showed that inhibition of Ron activation with a neutralizing antibody reduced PC-3 cell migration while stimulation with recombinant HGFL increased cell migration (56). A second study revealed consistent results with these findings by

demonstrating that knockdown of Ron in PC-3 and DU-145 cells attenuates cell migration and invasion. Moreover, treatment with HGFL was sufficient to induce migration and invasion. In these models, ERK1/2 was shown to mediate HGFL-induced cell migration and invasion, suggesting this pathway plays a key role in Ron-mediated prostate cancer cell metastasis (57). These data suggest that epithelial Ron signaling is an important promoter of prostate cancer cell migration and invasion; however additional studies are needed to test the role of Ron in prostate cancer metastasis *in vivo* and expand upon the mediating mechanisms.

Prostate cancer epithelial-specific Ron expression has also been established as a key regulator of the tumor microenvironment. Ron in human prostate cancer cells positively regulates production of angiogenic chemokines through activation of NF- κ B (13). Furthermore, Ron expression in these cells was deemed necessary for endothelial cell recruitment and prostate tumor vascularization. Studies performed using TRAMP mice support the role of Ron signaling in angiogenesis, as loss of Ron or HGFL leads to decreased prostate tumor microvessel density as measured by CD31 staining (37, 38).

2.3 Macrophage-Specific Roles for Ron in Prostate Cancer

Continuing the investigation into cell type specific functions for Ron has led to multiple discoveries regarding the role of Ron in macrophages to promote prostate cancer. Research has uncovered that Ron is expressed primarily on tissue resident macrophages and terminally differentiated macrophages, but Ron expression is markedly lower in circulatory monocytes (4, 58–61). This observation was also supported in prostate tumor tissue with Ron expression detected in tumor resident macrophages in an orthotopic model of prostate cancer (41). Using a model of breast cancer, Ron expression was connected specifically to a subset of tumor associated macrophages that express Tie2 (62). The significance of macrophage specific Ron expression for prostate tumor growth was first directly examined when mice harboring a myeloid specific deletion of the Ron tyrosine kinase domain (LysMCre TKf/f) were orthotopically injected with syngeneic murine C2RE3 prostate cancer cells. In this model, mice with a myeloid specific Ron loss developed significantly smaller prostate tumors and exhibited increased tumor cell apoptosis compared to transplantation into Ron proficient counterparts (41). Interestingly, prostate tumors in Lys-M-Cre TKf/f mice had an increase in the number of tumor-infiltrated macrophages. This observation was consistent with whole body loss of Ron signaling in both prostate and breast cancer murine models (30, 38), suggesting that macrophage-specific Ron expression is at least partly responsible for regulate macrophage tumor infiltration.

Research regarding Ron expression in macrophages has established that Ron is capable of promoting a M2 macrophage phenotype, as Ron expression promotes arginase expression and inhibits inducible nitric oxide synthase (iNOS) expression (30, 41, 62–64). Macrophage activation can be characterized as a continuum between M1 and M2, with M1 traditionally being inhibitory toward tumor growth and M2 being tumor promoting (65). M2 macrophages are anti-inflammatory in nature and are known to promote angiogenesis and matrix remodeling in cancer (65, 66). Thus, loss of Ron in macrophages suppresses the M2

phenotype and produces a macrophage that is capable of infiltrating tumors and suppressing tumor growth.

Macrophages are known to impact the function of other cells within the tumor microenvironment. Given that Ron in macrophages has been shown to suppress inflammatory responses in several injury and infection models, macrophage Ron signaling may play a crucial role in regulating the tumor microenvironment (7, 60, 67–70). Moreover, Ron activation in macrophages suppresses TLR4 signaling, which could limit the activation of neighboring immune cells within the tumor thereby suppressing tumor immune surveillance (71). Indeed, myeloid specific loss of Ron resulted in reduced cytotoxic T-cell function in prostate tumors (41). This result is consistent with what was observed in a murine model of breast cancer, where T-cells isolated from Ron signaling deficient mice had increased proliferation rates, increased expression of T-cell activation markers, and increased *in vitro* cytotoxicity when co-cultured with breast cancer cells. Furthermore, the increased cytotoxic T cell response seen in tumor-bearing Ron deficient mice was correlated with reduced tumor growth and metastasis (30). Taken together, these studies implicate Ron signaling in macrophages as a key regulator of the antitumor immune response.

2.4 Ron in Castration Resistant Prostate Cancer

Ron signaling had been established as a critical player in prostate cancer growth and development, but until recently the role of this signaling pathway had not been evaluated in the most deadly form of prostate cancer, Castration Resistant Prostate Cancer (CRPC). Ron mRNA and protein expression in patients was determined to be elevated in hormone refractory prostate cancer samples relative to hormone naïve samples (72, 73). Further, recent data from our laboratory has shown that Ron is functionally important for the development of castration resistance in several murine allograft and human xenograft mouse models (73). The ability of Ron to promote castration resistance is, at least in part, dependent on activation of β -catenin, NF- κ B, and the androgen receptor. Activation of β -catenin through Ron in prostate cancer had yet to be detected, however, in breast cancer Ron has been shown to activate β -catenin for promotion of growth and in the regulation of cancer stem cells (49, 74, 75). Under androgen deprivation, Ron activation of the androgen receptor appears to be dependent on β -catenin and NF- κ B. Interestingly, Ron has been reported to have differential effects on the androgen receptor depending on the presence or absence of androgens. When androgens are present, the relationship between Ron and the androgen receptor may be inhibitory (72). However, under androgen deprived conditions the relationship appears to be mutually active as Ron overexpression was shown to induce activation of the androgen receptor (72, 73) and re-expression of the androgen receptor in PC-3 cells was shown to induce transcription of Ron (72).

Understanding the differential effects of Ron on the androgen receptor is critical for the treatment of patients with CRPC, because the majority CRPCs have low levels of androgens due to treatment with androgen deprivation therapy. Additionally, several androgen receptor variants have been uncovered which play pivotal roles in prostate cancer, most notably AR-variant 7 (76). Further studies should focus on determining if Ron expression alters androgen receptor variant expression, and if so under what conditions. Moreover, reports

have shown that macrophage androgen receptor expression plays an important role in the development/initiation of prostate cancer (77). With this information, understanding the impact that Ron inhibition may have on macrophage function and on androgen receptor signaling in during the treatment of CRPC patients may prove to be crucial. CRPC is a devastating disease with limited treatment options and these initial studies established the scientific underpinnings for targeting Ron in CRPC.

3 Available Therapeutic Options for RON

3.1 Small Molecules

Possessing an intracellular kinase domain has made targeting Ron with a small molecule inhibitor (SMI) a realistic possibility. As such, several inhibitors have been developed that show efficacy against Ron. Specifically, Foretinib (EXEL-2880) is a SMI with high specificity against Ron, c-Met, and VEGF and was shown to reduce proliferation in cancer cells (78). A clinical trial with Foretinib has yet to be completed for prostate cancer; however, a phase II clinical trial was completed in Triple Negative Breast Cancer showing a clinical benefit rate of 46% (79). Recent pre-clinical work in prostate cancer cell lines indicates that Foretinib treatment may be beneficial in prostate cancer as treatment suppressed metastasis and reversed epithelial to mesenchymal transition (57).

Another intriguing SMI is the compound known as ASLAN002/BMS-777607. ASLAN002 is a dual Ron/c-Met tyrosine kinase inhibitor, but is one of the few compounds available that has preferential action against Ron over c-Met (80). A phase 1 clinical trial of ASLAN002 recently completed in patients with metastatic solid cancers and showed that the inhibitor is well tolerated and suggested that a phase 2 clinical trial begin with the treatment of 300mg twice daily (81). Recently, preclinical work from our laboratory with ASLAN002 illustrated that treatment in combination with castration therapy for castration resistant prostate tumors in a murine model of CRPC inhibits tumor growth (73). Additionally, bone metastases are frequent occurrences in metastatic prostate cancer patients and work by Andrade *et al* showed that treatment with ASLAN002 limits cancer-mediated bone destruction in murine models (82). The numerous reports demonstrating that ASLAN002 is safe and possibly effective at treating prostate cancer warrants further clinical study regarding use of this compound in prostate cancer patients.

A more recently developed inhibitor for Ron/c-Met is Merestinib/LY2801653 (83). Preclinical work with this compound showed its ability to inhibit cancer cell proliferation and cell scattering, and showed potent in vivo antitumor effects in xenograft mouse models (83). Merestinib recently completed a phase 1 clinical trial to determine tolerability in humans and the results have yet to be released (trial I3O-MC-JSBA, NCT01285037). Each of these SMI compounds targeting Ron has the potential to benefit prostate cancer patients. However, knowing that Ron promotes prostate cancer through its expression in both epithelial cells and macrophages, further research into how treatments should be targeted in patients is warranted.

3.2 Antibodies

A number of monoclonal antibodies have been generated toward the Ron receptor, with some making progress in clinical trials. Monoclonal antibodies against Ron can be used to directly target Ron signaling in cancer or they can be fused to a cytotoxic agent and used to guide that agent toward the tumor with Ron overexpression. Narnatumab/IMC-RON8 is a fully humanized monoclonal antibody that binds with high affinity to Ron, subsequently preventing the association of Ron with HGFL. A phase 1 clinical trial of Narnatumab has completed, determining that Narnatumab is well tolerated and provides limited antitumor activity (84). This study produced less than ideal results, however, with only 1 patient reaching the trough concentration at which Narnatumab produced antitumor activity in animal models.

A different approach to target Ron has been used by M.H. Wang's group, where these investigators developed three antibodies that target the Maturation Required Sequence of Ron located on the extracellular domain of this receptor. These antibodies are known as Zt/g4, Zt/f2, and Zt/c9 and rather than preventing ligand binding, this interaction induces receptor internalization and degradation (45, 85, 86). The induction of receptor internalization has been exploited by this group as a means to transport cytotoxic compounds inside the cell. These antibodies have been successfully coupled to doxorubicin, 5-fluorouracil, Gemcitabine, as well as other compounds, and have show preclinical efficacy (45, 85, 87). These proofs of concept studies establish the utility of Ron antibodies in cancer therapy, although none of these current antibodies have been tested in preclinical models for the treatment of prostate cancer. Furthermore, no studies have examined the impact of antibodies targeting Ron in the epithelial versus macrophage compartments.

4. The Outlook of Ron in Prostate Cancer

Significant progress has been made detailing the important role of Ron in prostate cancer since Ron was initially discovered. This progress has laid a solid groundwork for future studies to catapult the idea of the treatment of Ron signaling for prostate cancer into mainstream therapeutics. New areas of research will need to focus on the different possibilities for Ron to be used as a biomarker and the mechanism for directly targeting Ron in patients with prostate cancer. As a biomarker, plasma levels of HGFL have already been shown to correlate with prostate cancer progression and Ron expression shown to correlate with Gleason score and response to hormone therapy (72, 88). As a direct target, several compounds outlined previously are already available to begin testing in patients. Additionally, a major problem with treating prostate tumors is that they are known to have low immunogenicity, making many immunotherapies ineffective. Knowing that macrophage loss of Ron impacts macrophage infiltration and alters T-cell function provides a basis for changing that constraint. Although not in prostate cancer, it has recently been shown that Ron inhibition in breast cancer enhances response to anti-CTLA-4 immunotherapy in murine models (89). This suggests that Ron inhibition may be able to suppress antitumor immunity in prostate cancer to increase immunogenicity and sensitize tumors to immunotherapies.

With the number of studies demonstrating that Ron can impact the prostate tumor microenvironment, it is imperative to determine what role the tumor microenvironment plays in regulating castration resistant prostate cancer. Specifically, Ron has been shown to alter endothelial cells, macrophages, and T-cells of the prostate tumor microenvironment making these cell types a primary focus. If Ron produces significant changes to the tumor microenvironment to promote CRPC, then coupling Ron to other therapies, such as immunomodulatory agents, may prove effective in Ron overexpressing CRPC tumors. Lastly, as Ron has been linked to therapeutic resistance in prostate cancer, and Ron has been shown to regulate stemness in breast cancer, studies should be performed which focus on the ability of Ron to regulate stemness in prostate cancer to drive therapeutic resistance. Producing research focused in these areas will enhance our ability to discern what patients will benefit from Ron directed therapy and what method will be best suited for targeting Ron in prostate cancer.

Acknowledgments

Grant Support: This work was funded by the United States Department of Veterans Affairs research grant 1IOBX000803 (SEW); National Institutes of Health Grants T32 CA117846 (SEW, NEB), F31-CA200390 (NEB), and CA125379 (SEW) and Department of Defense Prostate Cancer Research Program Awards (W81XWH-10-2-0056 and W81XWH-10-2-0046) for the Prostate Cancer Biorepository Network (PCBN).

References

1. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *N Engl J Med*. 2001 3 15;344(11):783–92. PubMed PMID: . Epub 2001/03/15. eng. [PubMed: 11248153]
2. Gaudino G, Avantiaggiato V, Follenzi A, Acampora D, Simeone A, Comoglio PM. The proto-oncogene RON is involved in development of epithelial, bone and neuro-endocrine tissues. *Oncogene*. 1995 12 21;11(12):2627–37. PubMed PMID: . Epub 1995/12/21. eng. [PubMed: 8545120]
3. Okino T, Egami H, Ohmachi H, Takai E, Tamori Y, Nakagawa A, et al. Immunohistochemical analysis of distribution of RON receptor tyrosine kinase in human digestive organs. *Dig Dis Sci*. 2001 2;46(2):424–9. PubMed PMID: . Epub 2001/04/03. eng. [PubMed: 11281194]
4. Iwama A, Wang MH, Yamaguchi N, Ohno N, Okano K, Sudo T, et al. Terminal differentiation of murine resident peritoneal macrophages is characterized by expression of the STK protein tyrosine kinase, a receptor for macrophage-stimulating protein. *Blood*. 1995 11 01;86(9):3394–403. PubMed PMID: . Epub 1995/11/01. eng. [PubMed: 7579443]
5. Pegram MD, Lipton A, Hayes DF, Weber BL, Baselga JM, Tripathy D, et al. Phase II study of receptor-enhanced chemosensitivity using recombinant humanized anti-p185HER2/neu monoclonal antibody plus cisplatin in patients with HER2/neu-overexpressing metastatic breast cancer refractory to chemotherapy treatment. *J Clin Oncol*. 1998 8;16(8):2659–71. PubMed PMID: . Epub 1998/08/15. eng. [PubMed: 9704716]
6. Bezerra JA, Witte DP, Aronow BJ, Degen SJ. Hepatocyte-specific expression of the mouse hepatocyte growth factor-like protein. *Hepatology*. 1993 8;18(2):394–9. PubMed PMID: . Epub 1993/08/01. eng. [PubMed: 8340069]
7. Nanney LB, Skeel A, Luan J, Polis S, Richmond A, Wang MH, et al. Proteolytic cleavage and activation of pro-macrophage-stimulating protein and upregulation of its receptor in tissue injury. *J Invest Dermatol*. 1998 10;111(4):573–81. PubMed PMID: . Epub 1998/10/09. eng. [PubMed: 9764835]
8. Wang MH, Yoshimura T, Skeel A, Leonard EJ. Proteolytic conversion of single chain precursor macrophage-stimulating protein to a biologically active heterodimer by contact enzymes of the

- coagulation cascade. *J Biol Chem.* 1994 2 4;269(5):3436–40. PubMed PMID: . Epub 1994/02/04. eng. [PubMed: 7508914]
9. Camp ER, Yang A, Gray MJ, Fan F, Hamilton SR, Evans DB, et al. Tyrosine kinase receptor RON in human pancreatic cancer: expression, function, and validation as a target. *Cancer.* 2007 3 15;109(6):1030–9. PubMed PMID: . Epub 2007/02/22. eng. [PubMed: 17311308]
 10. Maggiora P, Marchio S, Stella MC, Giai M, Belfiore A, De Bortoli M, et al. Overexpression of the RON gene in human breast carcinoma. *Oncogene.* 1998 6 4;16(22):2927–33. PubMed PMID: . Epub 1998/07/22. eng. [PubMed: 9671413]
 11. Wagh PK, Peace BE, Waltz SE. Met-related receptor tyrosine kinase Ron in tumor growth and metastasis. *Adv Cancer Res.* 2008;100:1–33. PubMed PMID: . Pubmed Central PMCID: 4102433. Epub 2008/07/16. eng. [PubMed: 18620091]
 12. Welm AL, Sneddon JB, Taylor C, Nuyten DS, van de Vijver MJ, Hasegawa BH, et al. The macrophage-stimulating protein pathway promotes metastasis in a mouse model for breast cancer and predicts poor prognosis in humans. *Proc Natl Acad Sci U S A.* 2007 5 1;104(18):7570–5. PubMed PMID: . Pubmed Central PMCID: 1855278. Epub 2007/04/26. eng. [PubMed: 17456594]
 13. Thobe MN, Gurusamy D, Pathrose P, Waltz SE. The Ron receptor tyrosine kinase positively regulates angiogenic chemokine production in prostate cancer cells. *Oncogene.* 2010 1 14;29(2): 214–26. PubMed PMID: . Pubmed Central PMCID: 2806938. Epub 2009/10/20. eng. [PubMed: 19838218]
 14. Noone AM HN, Krapcho M, Miller D, Brest A, Yu M, Ruhl J, Tatalovich Z, Mariotto A, Lewis DR, Chen HS, Feuer EJ, Cronin KA (eds). *SEER Cancer Statistics Review.* 2018.
 15. Iwama A, Okano K, Sudo T, Matsuda Y, Suda T. Molecular cloning of a novel receptor tyrosine kinase gene, STK, derived from enriched hematopoietic stem cells. *Blood.* 1994 6 1;83(11):3160–9. PubMed PMID: . Epub 1994/06/01. eng. [PubMed: 8193352]
 16. Huff JL, Jelinek MA, Borgman CA, Lansing TJ, Parsons JT. The protooncogene c-sea encodes a transmembrane protein-tyrosine kinase related to the Met/hepatocyte growth factor/scatter factor receptor. *Proc Natl Acad Sci U S A.* 1993 7 1;90(13):6140–4. PubMed PMID: . Pubmed Central PMCID: 46883. Epub 1993/07/01. eng. [PubMed: 8392188]
 17. De Maria R, Maggiora P, Biolatti B, Prat M, Comoglio PM, Castagnaro M, et al. Feline STK gene expression in mammary carcinomas. *Oncogene.* 2002 3 7;21(11):1785–90. PubMed PMID: . Epub 2002/03/16. eng. [PubMed: 11896610]
 18. Wang MH, Iwama A, Skeel A, Suda T, Leonard EJ. The murine stk gene product, a transmembrane protein tyrosine kinase, is a receptor for macrophage-stimulating protein. *Proc Natl Acad Sci U S A.* 1995 4 25;92(9):3933–7. PubMed PMID: . Pubmed Central PMCID: 42076. Epub 1995/04/25. eng. [PubMed: 7732008]
 19. Nakamura T, Aoki S, Takahashi T, Matsumoto K, Kiyohara T. Cloning and expression of Xenopus HGF-like protein (HLP) and Ron/HLP receptor implicate their involvement in early neural development. *Biochem Biophys Res Commun.* 1996 7 16;224(2):564–73. PubMed PMID: . Epub 1996/07/16. eng. [PubMed: 8702427]
 20. Bassett DI. Identification and developmental expression of a macrophage stimulating 1/ hepatocyte growth factor-like 1 orthologue in the zebrafish. *Dev Genes Evol.* 2003 7;213(7):360–2. PubMed PMID: . Epub 2003/05/24. eng. [PubMed: 12764615]
 21. Chao KL, Tsai IW, Chen C, Herzberg O. Crystal structure of the Sema-PSI extracellular domain of human RON receptor tyrosine kinase. *PLoS One.* 2012;7(7):e41912 PubMed PMID: . Pubmed Central PMCID: 3405059. Epub 2012/08/01. eng. [PubMed: 22848655]
 22. Follenzi A, Bakovic S, Gual P, Stella MC, Longati P, Comoglio PM. Cross-talk between the proto-oncogenes Met and Ron. *Oncogene.* 2000 6 22;19(27):3041–9. PubMed PMID: . Epub 2000/06/29. eng. [PubMed: 10871856]
 23. Wang X, Yennawar N, Hankey PA. Autoinhibition of the Ron receptor tyrosine kinase by the juxtamembrane domain. *Cell Commun Signal.* 2014 4 16;12:28 PubMed PMID: . Pubmed Central PMCID: 4021555. Epub 2014/04/18. eng. [PubMed: 24739671]
 24. Lu Y, Yao HP, Wang MH. Multiple variants of the RON receptor tyrosine kinase: biochemical properties, tumorigenic activities, and potential drug targets. *Cancer Lett.* 2007 11 18;257(2):157–64. PubMed PMID: . Epub 2007/09/25. eng. [PubMed: 17889431]

25. Chakedis J, French R, Babicky M, Jaquish D, Mose E, Cheng P, et al. Characterization of RON protein isoforms in pancreatic cancer: implications for biology and therapeutics. *Oncotarget*. 2016 7 19;7(29):45959–75. PubMed PMID: . Pubmed Central PMCID: 5216774. Epub 2016/06/22. eng. [PubMed: 27323855]
26. Leonard EJ, Skeel A. A serum protein that stimulates macrophage movement, chemotaxis and spreading. *Exp Cell Res*. 1976 10 15;102(2):434–8. PubMed PMID: . Epub 1976/10/15. eng. [PubMed: 976357]
27. Wang MH, Cox GW, Yoshimura T, Sheffler LA, Skeel A, Leonard EJ. Macrophage-stimulating protein inhibits induction of nitric oxide production by endotoxin- or cytokine-stimulated mouse macrophages. *J Biol Chem*. 1994 5 13;269(19):14027–31. PubMed PMID: . Epub 1994/05/13. eng. [PubMed: 7514598]
28. Wang MH, Ronsin C, Gesnel MC, Coupey L, Skeel A, Leonard EJ, et al. Identification of the ron gene product as the receptor for the human macrophage stimulating protein. *Science*. 1994 10 7;266(5182):117–9. PubMed PMID: . Epub 1994/10/07. eng. [PubMed: 7939629]
29. Waltz SE, Eaton L, Toney-Earley K, Hess KA, Peace BE, Ihlendorf JR, et al. Ron-mediated cytoplasmic signaling is dispensable for viability but is required to limit inflammatory responses. *J Clin Invest*. 2001 8;108(4):567–76. PubMed PMID: . Pubmed Central PMCID: 209396. Epub 2001/08/24. eng. [PubMed: 11518730]
30. Benight NM, Wagh PK, Zinser GM, Peace BE, Stuart WD, Vasiliaskas J, et al. HGFL supports mammary tumorigenesis by enhancing tumor cell intrinsic survival and influencing macrophage and T-cell responses. *Oncotarget*. 2015 7 10;6(19):17445–61. PubMed PMID: . Pubmed Central PMCID: 4627320. Epub 2015/05/06. eng. [PubMed: 25938541]
31. Feres KJ, Ischenko I, Hayman MJ. The RON receptor tyrosine kinase promotes MSP-independent cell spreading and survival in breast epithelial cells. *Oncogene*. 2009 1 15;28(2):279–88. PubMed PMID: . Pubmed Central PMCID: 2628450. Epub 2008/10/07. eng. [PubMed: 18836480]
32. Gaudino G, Follenzi A, Naldini L, Collesi C, Santoro M, Gallo KA, et al. RON is a heterodimeric tyrosine kinase receptor activated by the HGF homologue MSP. *EMBO J*. 1994 8 1;13(15):3524–32. PubMed PMID: . Pubmed Central PMCID: 395256. Epub 1994/08/01. eng. [PubMed: 8062829]
33. Kobayashi T, Furukawa Y, Kikuchi J, Ito C, Miyata Y, Muto S, et al. Transactivation of RON receptor tyrosine kinase by interaction with PDGF receptor beta during steady-state growth of human mesangial cells. *Kidney Int*. 2009 6;75(11):1173–83. PubMed PMID: . Epub 2009/02/27. eng. [PubMed: 19242504]
34. Jaquish DV, Yu PT, Shields DJ, French RP, Maruyama KP, Niessen S, et al. IGF1-R signals through the RON receptor to mediate pancreatic cancer cell migration. *Carcinogenesis*. 2011 8;32(8):1151–6. PubMed PMID: . Pubmed Central PMCID: 3149203. Epub 2011/05/14. eng. [PubMed: 21565828]
35. Conrotto P, Corso S, Gamberini S, Comoglio PM, Giordano S. Interplay between scatter factor receptors and B plexins controls invasive growth. *Oncogene*. 2004 7 1;23(30):5131–7. PubMed PMID: . Epub 2004/06/09. eng. [PubMed: 15184888]
36. Peace BE, Hill KJ, Degen SJ, Waltz SE. Cross-talk between the receptor tyrosine kinases Ron and epidermal growth factor receptor. *Exp Cell Res*. 2003 10 1;289(2):317–25. PubMed PMID: . Epub 2003/09/23. eng. [PubMed: 14499632]
37. Thobe MN, Gray JK, Gurusamy D, Paluch AM, Wagh PK, Pathrose P, et al. The Ron receptor promotes prostate tumor growth in the TRAMP mouse model. *Oncogene*. 2011 12 15;30(50):4990–8. PubMed PMID: . Pubmed Central PMCID: 3165145. Epub 2011/06/01. eng. [PubMed: 21625214]
38. Vasiliaskas J, Nashu MA, Pathrose P, Starnes SL, Waltz SE. Hepatocyte growth factor-like protein is required for prostate tumor growth in the TRAMP mouse model. *Oncotarget*. 2014 7 30;5(14):5547–58. PubMed PMID: . Pubmed Central PMCID: 4170603. Epub 2014/07/02. eng. [PubMed: 24980820]
39. Peace BE, Toney-Earley K, Collins MH, Waltz SE. Ron receptor signaling augments mammary tumor formation and metastasis in a murine model of breast cancer. *Cancer Res*. 2005 2 15;65(4):1285–93. PubMed PMID: . Epub 2005/03/01. eng. [PubMed: 15735014]

40. Benight NM, Waltz SE. Ron receptor tyrosine kinase signaling as a therapeutic target. *Expert Opin Ther Targets*. 2012 9;16(9):921–31. PubMed PMID: . Pubmed Central PMCID: 4075176. Epub 2012/07/28. eng. [PubMed: 22834780]
41. Gurusamy D, Gray JK, Pathrose P, Kulkarni RM, Finkleman FD, Waltz SE. Myeloid-specific expression of Ron receptor kinase promotes prostate tumor growth. *Cancer Res*. 2013 3 15;73(6):1752–63. PubMed PMID: . Pubmed Central PMCID: 3602275. Epub 2013/01/19. eng. [PubMed: 23328584]
42. Wang MH, Lee W, Luo YL, Weis MT, Yao HP. Altered expression of the RON receptor tyrosine kinase in various epithelial cancers and its contribution to tumorigenic phenotypes in thyroid cancer cells. *J Pathol*. 2007 12;213(4):402–11. PubMed PMID: . Epub 2007/10/24. eng. [PubMed: 17955509]
43. Wang MH, Lao WF, Wang D, Luo YL, Yao HP. Blocking tumorigenic activities of colorectal cancer cells by a splicing RON receptor variant defective in the tyrosine kinase domain. *Cancer Biol Ther*. 2007 7;6(7):1121–9. PubMed PMID: . Epub 2007/07/06. eng. [PubMed: 17611409]
44. Xu XM, Wang D, Shen Q, Chen YQ, Wang MH. RNA-mediated gene silencing of the RON receptor tyrosine kinase alters oncogenic phenotypes of human colorectal carcinoma cells. *Oncogene*. 2004 11 4;23(52):8464–74. PubMed PMID: . Epub 2004/09/21. eng. [PubMed: 15378025]
45. Li Z, Yao H, Guin S, Padhye SS, Zhou YQ, Wang MH. Monoclonal antibody (mAb)-induced down-regulation of RON receptor tyrosine kinase diminishes tumorigenic activities of colon cancer cells. *Int J Oncol*. 2010 8;37(2):473–82. PubMed PMID: . Epub 2010/07/03. eng. [PubMed: 20596675]
46. Logan-Collins J, Thomas RM, Yu P, Jaquish D, Mose E, French R, et al. Silencing of RON receptor signaling promotes apoptosis and gemcitabine sensitivity in pancreatic cancers. *Cancer Res*. 2010 2 1;70(3):1130–40. PubMed PMID: . Pubmed Central PMCID: 2943733. Epub 2010/01/28. eng. [PubMed: 20103639]
47. Michele L Babicky M, Evangeline Mose, BSc, Karly Maruyama, BSc, Meg Harper, BSc, Dawn Jaquish, BSc, Randall French, PhD, Andrew M. Lowy, MD, FACS. RON overexpression accelerates tumorigenesis and induces metastasis in a KRAS mutant mouse model of pancreatic cancer. *Journal of the American College of Surgeons*. 2011;213(3):S131.
48. Chen YQ, Zhou YQ, Fu LH, Wang D, Wang MH. Multiple pulmonary adenomas in the lung of transgenic mice overexpressing the RON receptor tyrosine kinase. *Recepteur d'origine nantais. Carcinogenesis*. 2002 11;23(11):1811–9. PubMed PMID: . Epub 2002/11/07. eng. [PubMed: 12419829]
49. Zinser GM, Leonis MA, Toney K, Pathrose P, Thobe M, Kader SA, et al. Mammary-specific Ron receptor overexpression induces highly metastatic mammary tumors associated with beta-catenin activation. *Cancer Res*. 2006 12 15;66(24):11967–74. PubMed PMID: . Epub 2006/12/21. eng. [PubMed: 17178895]
50. Gray JK, Paluch AM, Stuart WD, Waltz SE. Ron receptor overexpression in the murine prostate induces prostate intraepithelial neoplasia. *Cancer Lett*. 2012 1 1;314(1):92–101. PubMed PMID: . Pubmed Central PMCID: 3225593. Epub 2011/10/19. eng. [PubMed: 22004727]
51. Narasimhan M, Ammanamanchi S. Curcumin blocks RON tyrosine kinase-mediated invasion of breast carcinoma cells. *Cancer Res*. 2008 7 1;68(13):5185–92. PubMed PMID: . Epub 2008/07/03. eng. [PubMed: 18593918]
52. Thangasamy A, Rogge J, Ammanamanchi S. Regulation of RON tyrosine kinase-mediated invasion of breast cancer cells. *J Biol Chem*. 2008 2 29;283(9):5335–43. PubMed PMID: . Epub 2008/01/01. eng. [PubMed: 18165235]
53. Cunha S, Lin YC, Goossen EA, DeVette CI, Albertella MR, Thomson S, et al. The RON receptor tyrosine kinase promotes metastasis by triggering MBD4-dependent DNA methylation reprogramming. *Cell Rep*. 2014 1 16;6(1):141–54. PubMed PMID: . Pubmed Central PMCID: 5312658. Epub 2014/01/07. eng. [PubMed: 24388747]
54. Zhao S, Ammanamanchi S, Brattain M, Cao L, Thangasamy A, Wang J, et al. Smad4-dependent TGF-beta signaling suppresses RON receptor tyrosine kinase-dependent motility and invasion of pancreatic cancer cells. *J Biol Chem*. 2008 4 25;283(17):11293–301. PubMed PMID: . Pubmed Central PMCID: 2431051. Epub 2008/03/04. eng. [PubMed: 18310076]

55. Sato S, Hanibuchi M, Kuramoto T, Yamamori N, Goto H, Ogawa H, et al. Macrophage stimulating protein promotes liver metastases of small cell lung cancer cells by affecting the organ microenvironment. *Clin Exp Metastasis*. 2013 3;30(3):333–44. PubMed PMID: . Epub 2012/09/27. eng. [PubMed: 23011677]
56. Jiang WG, Ye L, Ablin RJ, Kynaston HG, Mason MD. The prostate transglutaminase, TGase-4, coordinates with the HGFL/MSP-RON system in stimulating the migration of prostate cancer cells. *Int J Oncol*. 2010 8;37(2):413–8. PubMed PMID: . Epub 2010/07/03. eng. [PubMed: 20596668]
57. Yin B, Liu Z, Wang Y, Wang X, Liu W, Yu P, et al. RON and c-Met facilitate metastasis through the ERK signaling pathway in prostate cancer cells. *Oncol Rep*. 2017 6;37(6):3209–18. PubMed PMID: . Pubmed Central PMCID: 5442400. Epub 2017/04/26. eng. [PubMed: 28440432]
58. Brunelleschi S, Penengo L, Lavagno L, Santoro C, Colangelo D, Viano I, et al. Macrophage stimulating protein (MSP) evokes superoxide anion production by human macrophages of different origin. *Br J Pharmacol*. 2001 11;134(6):1285–95. PubMed PMID: . Pubmed Central PMCID: 1573047. Epub 2001/11/13. eng. [PubMed: 11704649]
59. Kurihara N, Iwama A, Tatsumi J, Ikeda K, Suda T. Macrophage-stimulating protein activates STK receptor tyrosine kinase on osteoclasts and facilitates bone resorption by osteoclast-like cells. *Blood*. 1996 5 1;87(9):3704–10. PubMed PMID: . Epub 1996/05/01. eng. [PubMed: 8611695]
60. Stuart WD, Kulkarni RM, Gray JK, Vasiliauskas J, Leonis MA, Waltz SE. Ron receptor regulates Kupffer cell-dependent cytokine production and hepatocyte survival following endotoxin exposure in mice. *Hepatology*. 2011 5;53(5):1618–28. PubMed PMID: . Pubmed Central PMCID: 3082400. Epub 2011/04/27. eng. [PubMed: 21520175]
61. Suzuki Y, Funakoshi H, Machide M, Matsumoto K, Nakamura T. Regulation of cell migration and cytokine production by HGF-like protein (HLP) / macrophage stimulating protein (MSP) in primary microglia. *Biomed Res*. 2008 4;29(2):77–84. PubMed PMID: . Epub 2008/05/16. eng. [PubMed: 18480548]
62. Sharda DR, Yu S, Ray M, Squadrito ML, De Palma M, Wynn TA, et al. Regulation of macrophage arginase expression and tumor growth by the Ron receptor tyrosine kinase. *J Immunol*. 2011 9 1;187(5):2181–92. PubMed PMID: . Pubmed Central PMCID: 4042865. Epub 2011/08/04. eng. [PubMed: 21810604]
63. Chen YQ, Fisher JH, Wang MH. Activation of the RON receptor tyrosine kinase inhibits inducible nitric oxide synthase (iNOS) expression by murine peritoneal exudate macrophages: phosphatidylinositol-3 kinase is required for RON-mediated inhibition of iNOS expression. *J Immunol*. 1998 11 1;161(9):4950–9. PubMed PMID: . Epub 1998/10/30. eng. [PubMed: 9794431]
64. Morrison AC, Correll PH. Activation of the stem cell-derived tyrosine kinase/RON receptor tyrosine kinase by macrophage-stimulating protein results in the induction of arginase activity in murine peritoneal macrophages. *J Immunol*. 2002 1 15;168(2):853–60. PubMed PMID: . Epub 2002/01/05. eng. [PubMed: 11777982]
65. Aras S, Zaidi MR. TAMEless traitors: macrophages in cancer progression and metastasis. *Br J Cancer*. 2017 11 21;117(11):1583–91. PubMed PMID: . Pubmed Central PMCID: 5729447. Epub 2017/10/25. eng. [PubMed: 29065107]
66. Jetten N, Verbruggen S, Gijbels MJ, Post MJ, De Winther MP, Donners MM. Anti-inflammatory M2, but not pro-inflammatory M1 macrophages promote angiogenesis in vivo. *Angiogenesis*. 2014 1;17(1):109–18. PubMed PMID: . Epub 2013/09/10. eng. [PubMed: 24013945]
67. Nikolaidis NM, Gray JK, Gurusamy D, Fox W, Stuart WD, Huber N, et al. Ron receptor tyrosine kinase negatively regulates TNF α production in alveolar macrophages by inhibiting NF- κ B activity and Adam17 production. *Shock*. 2010 2;33(2):197–204. PubMed PMID: . Pubmed Central PMCID: 4082961. Epub 2009/06/03. eng. [PubMed: 19487969]
68. Caldwell CC, Martignoni A, Leonis MA, Ondiveeran HK, Fox-Robichaud AE, Waltz SE. Ron receptor tyrosine kinase-dependent hepatic neutrophil recruitment and survival benefit in a murine model of bacterial peritonitis. *Crit Care Med*. 2008 5;36(5):1585–93. PubMed PMID: . Pubmed Central PMCID: 4105018. Epub 2008/04/25. eng. [PubMed: 18434891]
69. McDowell SA, Mallakin A, Bachurski CJ, Toney-Earley K, Prows DR, Bruno T, et al. The role of the receptor tyrosine kinase Ron in nickel-induced acute lung injury. *Am J Respir Cell Mol Biol*. 2002 1;26(1):99–104. PubMed PMID: . Epub 2001/12/26. eng. [PubMed: 11751209]

70. Nikolaidis NM, Kulkarni RM, Gray JK, Collins MH, Waltz SE. Ron receptor deficient alveolar myeloid cells exacerbate LPS-induced acute lung injury in the murine lung. *Innate Immun.* 2011 12;17(6):499–507. PubMed PMID: . Pubmed Central PMCID: 4102430. Epub 2010/11/23. eng. [PubMed: 21088048]
71. Ray M, Yu S, Sharda DR, Wilson CB, Liu Q, Kaushal N, et al. Inhibition of TLR4-induced I κ B kinase activity by the RON receptor tyrosine kinase and its ligand, macrophage-stimulating protein. *J Immunol.* 2010 12 15;185(12):7309–16. PubMed PMID: . Pubmed Central PMCID: 4815273. Epub 2010/11/17. eng. [PubMed: 21078906]
72. Batth I, Yun H, Hussain S, Meng P, Osmulski P, Huang TH, et al. Crosstalk between RON and androgen receptor signaling in the development of castration resistant prostate cancer. *Oncotarget.* 2016 3 22;7(12):14048–63. PubMed PMID: . Pubmed Central PMCID: 4924697. Epub 2016/02/13. eng. [PubMed: 26872377]
73. Brown NE PAM, Nashu M.A., Komurov K., Waltz S.E. Tumor Cell Autonomous RON Receptor Expression Promotes Prostate Cancer Growth Under Conditions of Androgen Deprivation. *Neoplasia.* 2018.
74. Ruiz-Torres SJ, Benight NM, Karns RA, Lower EE, Guan JL, Waltz SE. HGFL-mediated RON signaling supports breast cancer stem cell phenotypes via activation of non-canonical beta-catenin signaling. *Oncotarget.* 2017 8 29;8(35):58918–33. PubMed PMID: . Pubmed Central PMCID: 5601703. Epub 2017/09/25. eng. [PubMed: 28938607]
75. Wagh PK, Gray JK, Zinser GM, Vasiliauskas J, James L, Monga SP, et al. beta-Catenin is required for Ron receptor-induced mammary tumorigenesis. *Oncogene.* 2011 8 25;30(34):3694–704. PubMed PMID: . Pubmed Central PMCID: 3134560. Epub 2011/03/23. eng. [PubMed: 21423209]
76. Antonarakis ES, Lu C, Wang H, Lubner B, Nakazawa M, Roeser JC, et al. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. *N Engl J Med.* 2014 9 11;371(11):1028–38. PubMed PMID: . Pubmed Central PMCID: 4201502. Epub 2014/09/04. eng. [PubMed: 25184630]
77. Fang LY, Izumi K, Lai KP, Liang L, Li L, Miyamoto H, et al. Infiltrating macrophages promote prostate tumorigenesis via modulating androgen receptor-mediated CCL4-STAT3 signaling. *Cancer Res.* 2013 9 15;73(18):5633–46. PubMed PMID: . Pubmed Central PMCID: 3833080. Epub 2013/07/24. eng. [PubMed: 23878190]
78. Qian F, Engst S, Yamaguchi K, Yu P, Won KA, Mock L, et al. Inhibition of tumor cell growth, invasion, and metastasis by EXEL-2880 (XL880, GSK1363089), a novel inhibitor of HGF and VEGF receptor tyrosine kinases. *Cancer Res.* 2009 10 15;69(20):8009–16. PubMed PMID: . Epub 2009/10/08. eng. [PubMed: 19808973]
79. Rayson D, Lupichuk S, Potvin K, Dent S, Shenkier T, Dhesy-Thind S, et al. Canadian Cancer Trials Group IND197: a phase II study of foretinib in patients with estrogen receptor, progesterone receptor, and human epidermal growth factor receptor 2-negative recurrent or metastatic breast cancer. *Breast Cancer Res Treat.* 2016 5;157(1):109–16. PubMed PMID: . Epub 2016/04/27. eng. [PubMed: 27116183]
80. Schroeder GM, An Y, Cai ZW, Chen XT, Clark C, Cornelius LA, et al. Discovery of N-(4-(2-amino-3-chloropyridin-4-yloxy)-3-fluorophenyl)-4-ethoxy-1-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carboxamide (BMS-777607), a selective and orally efficacious inhibitor of the Met kinase superfamily. *J Med Chem.* 2009 3 12;52(5):1251–4. PubMed PMID: . Epub 2009/03/06. eng. [PubMed: 19260711]
81. Roohullah A, Cooper A, Lomax AJ, Aung J, Barge A, Chow L, et al. A phase I trial to determine safety and pharmacokinetics of ASLAN002, an oral MET superfamily kinase inhibitor, in patients with advanced or metastatic solid cancers. *Invest New Drugs.* 2018 5 16 PubMed PMID: . Epub 2018/05/17. eng. [PubMed: 29766337]
82. Andrade K, Fornetti J, Zhao L, Miller SC, Randall RL, Anderson N, et al. RON kinase: A target for treatment of cancer-induced bone destruction and osteoporosis. *Sci Transl Med.* 2017 1 25;9(374). PubMed PMID: . Pubmed Central PMCID: 5771677. Epub 2017/01/27. eng. [PubMed: 28123075]
83. Yan SB, Peek VL, Ajamie R, Buchanan SG, Graff JR, Heidler SA, et al. LY2801653 is an orally bioavailable multi-kinase inhibitor with potent activity against MET, MST1R, and other oncoproteins, and displays anti-tumor activities in mouse xenograft models. *Invest New Drugs.*

- 2013 8;31(4):833–44. PubMed PMID: . Pubmed Central PMCID: 3717159. Epub 2013/01/01. eng. [PubMed: 23275061]
84. LoRusso PM, Gounder M, Jalal SI, Andre V, Kambhampati SRP, Loizos N, et al. Phase 1 study of narnatumab, an anti-RON receptor monoclonal antibody, in patients with advanced solid tumors. *Invest New Drugs*. 2017 8;35(4):442–50. PubMed PMID: . Pubmed Central PMCID: 5502198. Epub 2017/02/06. eng. [PubMed: 28161886]
85. Guin S, Yao HP, Wang MH. RON receptor tyrosine kinase as a target for delivery of chemodrugs by antibody directed pathway for cancer cell cytotoxicity. *Mol Pharm*. 2010 4 5;7(2):386–97. PubMed PMID: . Epub 2009/12/31. eng. [PubMed: 20039696]
86. Padhye SS, Guin S, Yao HP, Zhou YQ, Zhang R, Wang MH. Sustained expression of the RON receptor tyrosine kinase by pancreatic cancer stem cells as a potential targeting moiety for antibody-directed chemotherapeutics. *Mol Pharm*. 2011 12 5;8(6):2310–9. PubMed PMID: . Epub 2011/10/22. eng. [PubMed: 22014215]
87. Yao HP, Zhou YQ, Ma Q, Guin S, Padhye SS, Zhang RW, et al. The monoclonal antibody Zt/f2 targeting RON receptor tyrosine kinase as potential therapeutics against tumor growth-mediated by colon cancer cells. *Mol Cancer*. 2011 7 12;10:82 PubMed PMID: . Pubmed Central PMCID: 3142532. Epub 2011/07/14. eng. [PubMed: 21749705]
88. Sugie S, Mukai S, Yamasaki K, Kamibeppu T, Tsukino H, Kamoto T. Plasma macrophage-stimulating protein and hepatocyte growth factor levels are associated with prostate cancer progression. *Hum Cell*. 2016 1;29(1):22–9. PubMed PMID: . Epub 2015/08/08. eng. [PubMed: 26250899]
89. Ekiz HA, Lai S-CA, Gundlapalli H, Haroun F, Williams MA, Welm AL. Inhibition of RON kinase potentiates anti-CTLA-4 immunotherapy to shrink breast tumors and prevent metastatic outgrowth. *OncoImmunology*. 2018:1–16.