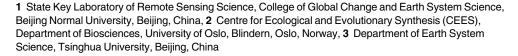


# The ecological dynamics of hantavirus diseases: From environmental variability to disease prevention largely based on data from China

Huaiyu Tian 1\*, Nils Chr. Stenseth 2,3\*



\* tianhuaiyu@gmail.com (HT); n.c.stenseth@ibv.uio.no (NCS)



# OPEN ACCESS

Citation: Tian H, Stenseth NC. (2019) The ecological dynamics of hantavirus diseases: From environmental variability to disease prevention largely based on data from China. PLoS Negl Trop Dis 13(2): e0006901. https://doi.org/10.1371/journal.pntd.0006901

**Editor:** Patricia V. Aguilar, University of Texas Medical Branch, UNITED STATES

Published: February 21, 2019

Copyright: © 2019 Tian, Stenseth. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: Funding for this study was provided by the Beijing Natural Science Foundation (JQ18025) (http://kw.beijing.gov.cn/jjb/), the National Natural Science Foundation of China (81673234,81460520, 31870400,41476161) (http://www.nsfc.gov.cn/), the Young Elite Scientist Sponsorship Program by CAST(YESS) (2018QNRC001) (http://www.cast.org.cn/), the Fundamental Research Funds for the Central Universities (http://www.moe.gov.cn/), the National Key Research and Development Program of China (2016YFA0600104) (http://www.most.gov.cn/), and the health industry's special research funds for public welfare projects (Grant 201502020) (http://www.nhfpc.gov.cn/).

## Abstract

Hantaviruses can cause hantavirus pulmonary syndrome (HPS) in the Americas and hemorrhagic fever with renal syndrome (HFRS) in Eurasia. In recent decades, repeated outbreaks of hantavirus disease have led to public concern and have created a global public health burden. Hantavirus spillover from natural hosts into human populations could be considered an ecological process, in which environmental forces, behavioral determinants of exposure, and dynamics at the human—animal interface affect human susceptibility and the epidemiology of the disease. In this review, we summarize the progress made in understanding hantavirus epidemiology and rodent reservoir population biology. We mainly focus on three species of rodent hosts with longitudinal studies of sufficient scale: the striped field mouse (*Apodemus agrarius*, the main reservoir host for Hantaan virus [HTNV], which causes HFRS) in Asia, the deer mouse (*Peromyscus maniculatus*, the main reservoir host for Sin Nombre virus [SNV], which causes HPS) in North America, and the bank vole (*Myodes glareolus*, the main reservoir host for Puumala virus [PUUV], which causes HFRS) in Europe. Moreover, we discuss the influence of ecological factors on human hantavirus disease outbreaks and provide an overview of research perspectives.

#### Introduction

Hantaviruses are enveloped RNA viruses belonging to the family Hantaviridae, genus *Orthohantavirus* [1]. They can cause serious diseases in humans, with some outbreaks resulting in case fatality rates of 12% (for hemorrhagic fever with renal syndrome [HFRS] in Europe and Asia) and up to 40% (for hantavirus pulmonary syndrome [HPS] in the Americas), depending on the hantavirus type and the resulting clinical syndrome [2,3]. Hantavirus disease came to global attention when two major outbreaks were reported during the last century. The first, an HFRS outbreak, occurred during the Korean War (1950 to 1953), when more than 3,000 United Nations troops fell ill [4]. The second was an outbreak of HPS that occurred in the



The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

Four Corners region of the southwestern United States in 1993 [5]. Hantaviruses remain a global threat to public health; they have been estimated to affect approximately 200,000 humans annually worldwide in recent years [6]. Moreover, the number of countries reporting human cases of hantavirus infection is still on the rise [7].

Human infections with hantaviruses result from contact with infected rodents or exposure to virus-contaminated aerosols; Andes virus (ANDV) is the only hantavirus in which personto-person transmission has been documented so far [8–11]. Outbreaks of hantavirus disease are therefore considered to be associated with the primary rodent host and pathogen dynamics [12,13]. However, the mechanism of zoonotic pathogen dynamics is complex, and the relationships between or among environmental change, host–pathogen dynamics, and human spill-over is far from clear [14,15]. For example, variations in incidence rates are not simply, as expected, a result of changes in rodent demography or virus prevalence [16–18]. Furthermore, although numerous research efforts have been undertaken, no WHO-licensed vaccine against hantavirus infection is available [19] (except Hantavax, which is only licensed for human use in the Republic of Korea). Current efforts to curb hantavirus transmission focus on avoiding contact between humans and host rodents [20,21]. Due to the complexity of these systems, hantaviruses deserve the attention of research scientists in the contexts of both public health and wildlife conservation.

Here, we present a review of the ecology of hantavirus diseases in an attempt to improve our understanding of the mechanisms underlying disease outbreaks. We mainly focus on three species of rodent hosts, on which there have been a wealth of longitudinal studies of population and prevalence dynamics: the striped field mouse (*Apodemus agrarius*, the main reservoir host for Hantaan virus [HTNV] [22–25]) in Asia, the deer mouse (*Peromyscus maniculatus*, the main reservoir host for Sin Nombre virus [SNV] [12,18,26–28]) in North America, and the bank vole (*Myodes glareolus*, the main reservoir host for Puumala virus [PUUV] [29–32]) in Europe. A deeper understanding of the natural ecological dynamics of host–pathogen interactions would be of great value in developing future strategies for disease prevention and control.

#### **Methods**

### Search strategy and selection criteria

We searched the MEDLINE (via PubMed) online database and Google Scholar for articles with the key words "hantavirus," "ecology," or "modelling" in the title, with no date limit, published before 31 July 2017, with restriction to mainly English papers. Key words used in Medical Subject Headings were "hantavirus," "hemorrhagic fever with renal syndrome," "hantavirus pulmonary syndrome," and "rodent reservoir." Inclusion criteria were predefined as research providing information on viral infections (including human incidence, prevalence of hantavirus infections in rodent hosts, and/or host–pathogen interactions), environmental change and rodent reservoir population dynamics, and information on environmental factors that may trigger hantavirus disease outbreaks. Study data extracted included study year, location, hantavirus type, main rodent reservoir, study design, and environmental factors.

#### Results

#### Geographic distribution

Hantaviruses that cause illness in humans have been identified across the globe [3,7,33] (Fig 1). The major hantavirus disease burden in the Old World is HFRS, and in the New World it is HPS. HTNV in Asia, PUUV and Dobrava virus in Europe, and Seoul virus (SEOV) worldwide

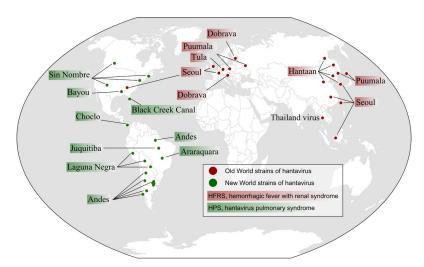


Fig 1. Map of Old World and New World hantavirus genotypes reported to be pathogenic for humans. Hantaviruses that have been shown to cause HFRS are shown in red, and those that cause HPS are shown in green. PUUV, which causes a milder form of HFRS (NE), is found in Europe. The described African hantavirus, Sangassou virus, was found in Guinea in 2016. In recent studies, THAIV is considered to act as an additional causative agent of HFRS. It should be noted that SEOV is harbored by *Rattus norvegicus* (brown rat) worldwide, but only those locations where reports of human infections with SEOV are shown. The map was created specifically for this manuscript and was generated by ArcGIS 9.2 (ESRI, Redlands, CA, USA) based on World Countries (http://www.arcgis.com/home/item.html?id=d974d9c6bc924ae0a2ffea0a46d71e3d). HFRS, hemorrhagic fever with renal syndrome; HPS, hantavirus pulmonary syndrome; NE, nephropathia epidemica; PUUV, Puumala virus; SEOV, Seoul virus; THAIV, Thailand virus.

https://doi.org/10.1371/journal.pntd.0006901.g001

are the causative agents of HFRS. SNV, ANDV, and related viruses have been identified as causative agents of HPS in the Americas [3,12]. Recent studies indicate that the medical problem caused by hantavirus infections may be underestimated in Africa, India, Southeast Asia (where Thailand virus [THAIV] has been isolated), and even Europe [25,31,34–37] and North America [38,39]. It is estimated that hantavirus diseases are heavily underdiagnosed in Europe; only 20% of PUUV infections have been diagnosed, and no human infections data exist in several countries [31].

## Environment variation and hantavirus reservoir population dynamics

A bottom-up trophic cascade hypothesis has been proposed to explain the chain reactions resulting from climatic conditions, primary productivity, and host demography [27,40,41]. A bottom-up trophic cascade suggests that a change in nutrient supply could lead to similar changes in equilibrium abundances at all trophic levels [42,43]. In rodent host-hantavirus systems, climatic conditions are one of the many factors that can affect rodent population dynamics and, consequently, the prevalence of virus infection in rodent reservoirs and risk of virus exposure in humans [44] (see example shown in Fig 2). A review of the available longitudinal studies of rodent communities supports the hypothesis that hantavirus reservoir populations in both the Old World and New World are significantly influenced by climate, either directly (via winter survival) or indirectly (through food limitation).

In Central China, an environmentally induced cascading effect on the population dynamics of the striped field mouse (*A. agrarius*, the main reservoir host for HTNV) was found by combining 30 years of field surveillance and satellite images. The normalized difference vegetation index (NDVI) value for farmland, and precipitation two months previously, were important in determining striped field mouse survival and recruitment rates, respectively [45]. An extreme

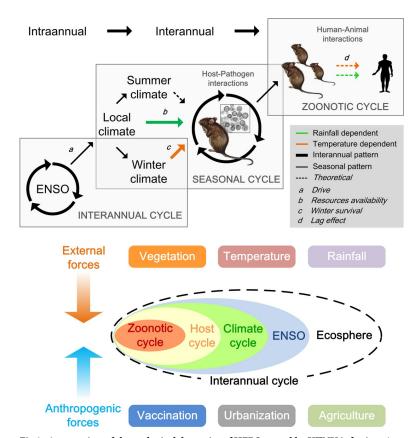


Fig 2. An overview of the ecological dynamics of HFRS caused by HTNV infection. Arrows represent connections affected by environment: the green line represents rainfall, and the orange line represents temperature. The solid line indicates available data, used in models linking the ENSO (Nino3.4 index) with local climate (rainfall and temperature), rodent population density (capture rate), and human HTNV infections. The rectangles delimit the seasonal, interannual, and zoonotic cycles of HTNV. Source: Adapted from [46]. ENSO, El Niño Southern Oscillation; HFRS, hemmorrhagic fever with renal syndrome; HTNV, Hantaan virus.

https://doi.org/10.1371/journal.pntd.0006901.g002

drought-induced food shortage is thought to increase mortality in striped field mouse populations, as they are a species that relies on farm crops [45]. Furthermore, winter temperature was found to exert complex effects on overwinter survival, thereby affecting the population growth rate in the following year [46]. In South China, the population density of the main reservoir host for SEOV, the brown rat (Rattus norvegicus), is correlated with temperature, precipitation, and the NDVI value for farmland [47]. In northeastern China, in Huludao City, an endemic area for HFRS due to SEOV, climate is considered to affect HFRS epidemics mainly through its effect on the population density of the brown rat (the most abundant rodent species present, accounting for more than 80% of the total rodent population) [48]. In Europe, food-related factors (seed production, climate affecting vegetation biomass) have been linked to rodent populations in deciduous forests [30,49]. Outbreaks of nephropathia epidemica (NE; a mild form of HFRS caused by PUUV infection) are hypothesized to have an ecological causal connection with the staple food source for the main carrier of PUUV, the bank vole, in mast years [32,50,51]. Population densities of bank voles were found to be associated with summer temperature and autumn temperature, both of which favor seed development [52,53]. A higher average winter temperature is believed to reduce winter survival of bank voles in Scandinavia because of a shorter period of protective snow cover against predators. On the contrary, increasing winter temperatures are found to provide a survival benefit to increasing populations

of striped field mouse in the spring in Central China; subsequently, a large population of rodents might be accompanied by intraspecific competition due to food or space limitation, resulting in a negative-feedback effect of population density [46]. In North America, the HPS outbreak in 1993 in the Four Corners region of the southwestern US was considered to be driven by increased precipitation induced by an El Niño-Southern Oscillation (ENSO) event. The HPS outbreaks in 1997 in the same region were also preceded by an ENSO event, which brought increased precipitation, favoring deer mouse host populations [26,27]. NDVI is also used to identify locations that can be monitored for the abundance of deer mouse and presence of SNV by examining the seasonal dynamics of vegetative patterns [54]. In central Montana, US, the survival and recruitment rates of the deer mouse were found to be associated with precipitation and temperature, implying a complex relationship between climate and population dynamics [55].

## Rodent population dynamics and risk to humans

Long-term observations on the prevalence of hantavirus infection in rodent hosts are critical to understanding the dynamics of hantavirus diseases in humans. Similarly, analysis of host–pathogen interactions is important to successfully implement disease control strategies [56]. Hantaviruses are known to be directly transmitted from infected to susceptible hosts (horizontal transmission). Therefore, changes in rodent population densities are expected to increase or decrease the transmission and prevalence of hantavirus infection in rodent reservoir populations, resulting in greater or lower levels of spillover to humans [28]. The dynamics of rodent population density and hantavirus infection prevalence are linked by contact rates; the virus may become extinct below a certain host density, corresponding to the minimum number of hosts required for virus maintenance in a population [57]. A critical population density threshold of striped field mice has been observed in Central China, below which HTNV cannot invade and persist in the population [45]. A similar population density threshold has been observed in the SNV reservoir, the deer mouse, in North America [58,59]. Rodent community composition has also been found to affect the risk of human hantavirus infection among different landscapes [60].

Studies have demonstrated positive correlations between relative population density of bank voles and prevalence of PUUV infection among rodent populations [61]. Bank vole density indices were also positively correlated with risk of HFRS in humans in northern Sweden, Finland, and Norway [62-64]. However, the relationship between rodent demography and disease dynamics in China and North America is more complicated. In Central China, fluctuation in HFRS incidence is highly correlated with striped field mouse population density and the prevalence of HFRS infection in Shaanxi Province, an endemic area for HFRS due to HTNV [13]. In northeastern China, HFRS cases are associated with a virus-carrying index, an indicator to describe the combined effects of rodent population density and prevalence of virus infection, with a one-month lag, in the SEOV endemic area of HFRS [48]. However, in South China, the number of HFRS cases is not associated with rodent population density in Chenzhou and Changsha (the main species captured are R. norvegicus, R. flavipectus, and Mus musculus), which are mixed HTNV and SEOV endemic areas [65,66]. In North America, different relationships have been found between deer mouse population density and SNV infection prevalence in the deer mouse population—positive correlations [17,27,67], no linear correlation [59,68,69], and even negative correlations [18,70,71]. (It should also be noted that the different correlational relationship may be due to temporal issues; e.g., some studies take account of time lags when analyzing the data while others do not.) Because hantaviruses are horizontally transmitted, these inconsistent results may be due to two conflicting effects of

population dynamics on prevalence [28]. During breeding seasons, rapid population growth of juveniles not yet infected may increase the proportion of uninfected rodents and potentially decrease hantavirus prevalence [72,73], whereas the resulting higher population size may eventually increase contact between individuals and prevalence. Taken together, these studies point to several issues that could be of importance to understanding the relationship between reservoir population dynamics and pathogen transmission [57]. Future demographic studies should aim to establish causal mechanisms linking behavior, demography, and virus prevalence dynamics.

## Environmental factors that trigger hantavirus disease outbreaks

Environmental factors are major triggers that affect reservoir ecology and virus ecology and thus are likely to affect hantavirus transmission from rodent reservoir to humans (or risk of virus exposure in humans). However, although outbreaks appear to be a result of these factors, it is difficult to predict the exact outcome, chiefly due to the complex and multifactorial mechanisms that drive hantavirus disease outbreaks. Studies have highlighted the importance of the interplay between extrinsic and intrinsic factors in determining hantavirus disease dynamics [45,46,74], e.g., when the size of outbreaks is small due to low population susceptibility levels as the result of vaccination or the rodent reservoir population density is below the threshold level required to maintain the virus due to environmental limitations. In this section, studies addressing environmental variability and human hantavirus infections were included (Table 1), and we mainly review three factors that play an important role in altering hantavirus disease dynamics and have been documented sufficiently—precipitation, temperature, and landscape alteration.

Table 1. Synthesis of the environmental factors, during the same year or before, that may trigger hantavirus disease outbreak.

Trigger factor	Detail	Disease	Area	Rodent host
Temperature	Summer (Y-2) and autumn temperature (Y-1) <sup>+</sup>	NE	Belgium [50,52], Central Europe	M. glareolus
	Winter and spring temperature (Y 0) <sup>+</sup>	NE	Southwestern Germany [53], Central Europe	M. glareolus
	Winter temperature-	HPS	Southern Argentina [91], South America	Oligoryzomys longicaudatus
	Annual mean temperature <sup>+</sup>	HPS	Atlantic forest, Brazil [128], South America	Family Cricetidae
	Maximum temperature (Y 0) <sup>+</sup>	HFRS	Heilongjiang Province, North China [129], East Asia	Unknown
	Mean temperature (Y 0) <sup>+</sup>	HFRS	Inner Mongolia, North China [89]; South Korea [130]; East Asia	Unknown
	Mean temperature (Y 0) <sup>+</sup>	HFRS	Huludao City, North China [48]; Changsha City, South China [20,66]; East Asia	R. norvegicus
	Average temperature (Y 0)-	HFRS	Shandong Province, North China [90], East Asia	Unknown
	Summer temperature (Y 0)-	HFRS	Weihe Plain, Central China [46], East Asia	Apodemus agrarius
Rainfall	Summer rainfall (Y-3) <sup>+</sup>	NE	Belgium [50], Central Europe	M. glareolus
	Rainfall (Y-1) <sup>+</sup>	HPS	the Four Corners region of New Mexico and Arizona, US [27], North America	P. maniculatus
	Winter rainfall <sup>+</sup>	HPS	Southern Brazil [131], South America	Unknown
	Annual rainfall <sup>+</sup>	HPS	Southern Argentina [91], South America	O. longicaudatus
	Monthly rainfall (Y 0) <sup>+</sup>	HPS	Chile [132], South America	Unknown
	Summer and autumn rainfall (Y $0$ ) <sup>+</sup> ; annual rainfall (Y-1) <sup>+</sup>	HFRS	Xi'an City, Central China [46,100], East Asia	A. agrarius
	Monthly rainfall (Y 0) <sup>+</sup>	HFRS	Huludao City, North China [48], East Asia	R. norvegicus
	Monthly rainfall (Y 0) <sup>+</sup>	HFRS	Changsha City, South China [78]; Inner Mongolia, North China [89]; South Korea [130], East Asia	Unknown
	Monthly rainfall (Y 0)-	HFRS	Jiaonan County, North China [133]; Shandong Province, North China [90]; Anhui Province, South China [85], East Asia	Unknown

(Continued)



Table 1. (Continued)

Trigger factor	Detail	Disease	Area	Rodent host
Humidity	Absolute humidity (Y 0) <sup>+</sup>	HFRS	Changsha City, South China [78]	Unknown
	Relatvie humidity (Y 0)–	HFRS	Jiaonan County, North China [133]; Shandong Province, North China [90], East Asia	Unknown
	Relative humidity (Y 0) <sup>+</sup>	HFRS	Inner Mongolia, North China [89]; Liaoning Province, North China [134]; South Korea [130], East Asia	Unknown
	Relative humidity (Y 0)+	HFRS	Huludao City, North China [48], East Asia	R. norvegicus
	Temperature vegetationdryness index <sup>+</sup>	HFRS	Changsha City, Zhuzhou City, Xiangtan city, Hengyang City, South China [127], East Asia	R. norvegicus, M. musculus, A. agrarius, R. flavipectus
	Proportion of thin particles (<10 μm) (Y 0) <sup>+</sup>	NE	Northern Belgium [81], Central Europe	M. glareolus
Air pressure	Mean air pressure (Y 0)-	HFRS	Huludao City, North China [48], East Asia	R. norvegicus
Air pollution	PM <sub>10</sub> (Y 0) <sup>+</sup>	HFRS	South Korea [130], East Asia	Unknown
Flood	Water-level difference of Huai River (Y 0)–	HFRS	Anhui Province, South China [86], East Asia	Unknown
ENSO	Southern oscillation index (Y 0)-	HFRS	Heilongjiang Province, North China [129]; Anhui Province, South China [84,135], East Asia	Unknown
	Multivariate ENSO index (Y 0) <sup>+</sup>	HFRS	Changsha City, South China [66,78]; Inner Mongolia, North China [89], East Asia	Unknown
	Multivariate ENSO index (Y-1) <sup>+</sup>	HPS	Four Corners region of New Mexico and Arizona, US [136], North America	P. maniculatus
Land cover change	Forestation <sup>+</sup>	HFRS	Liaoning Province, North China [134], East Asia	Unknown
	Developed land-; Rice paddy <sup>+</sup> ; Orchard <sup>+</sup>	HFRS	Beijing, North China [119,137], East Asia	Unknown
	Area of deciduous forest <sup>+</sup>	NE	Northern Belgium [81,108], Central Europe	M. glareolus
	Cover of beech forest, cover of seed plant <sup>+</sup>	NE	Southwestern Germany [53], Central Europe; Temperate Europe [138]	M. glareolus
	Proportion of land cultivated for sugarcane <sup>+</sup>	HPS	São Paulo [128], South America	Family Cricetidae
	Proportion of forest cover <sup>+</sup>	HPS	Atlantic forest, Brazil [128], South America	Family Cricetidae
Food availability for rodent host	Seed production (Y-1) <sup>+</sup>	NE	Western and central European countries [52,53,139-141]	M. glareolus
	NDVI for trapping site (Y-1)	NE	Northern Belgium [108], Central Europe	M. glareolus
	High photosynthetic mass <sup>+</sup>	HPS	Southern Brazil [131], South America	Unknown
	NDVI (Y 0) <sup>+</sup>	NE	the Franche-Comté region, France [82], Central Europe	Unknown
	NDVI for farmland (Y 0) <sup>+</sup>	HFRS	Inner Mongolia [142], North China, East Asia	Unknown
	NDVI for rice paddy (Y 0) <sup>+</sup>	HFRS	Changsha City, South China [66], East Asia	M. musculus, R. flavipectus, R. norvegicus
	net photosynthesis (Y 0)+	HFRS	Weihe Plain, North China [41], East Asia	A. agrarius
Socioeconomic factors	Gross domestic product and the urbanization rate (Y 0)–	HFRS	Chenzhou City, South China [20], East Asia	R. norvegicus, R. flavipectus
	Human development index-	HPS	São Paulo [128], South America	Family Cricetidae
Selenium deficiency	Selenium content in feed-	HFRS	China [143], East Asia	Unknown

<sup>&</sup>lt;sup>+</sup>Positive correlation with hantavirus infections

**Abbreviations:** ENSO, El Niño Southern Oscillation; HFRS, hemorrhagic fever with renal syndrome; HPS, hantavrius pulmonary syndrome; NDVI, normalized difference vegetation index; NE, nephropathia epidemica; (Y 0), during the same year; Y-n, n years before

https://doi.org/10.1371/journal.pntd.0006901.t001

It is becoming increasingly more apparent that climatic variations have profound impacts on infectious disease dynamics [75,76], especially for climate-sensitive infectious diseases; e.g., human hantavirus diseases are considered climate-sensitive infectious diseases [77].

Negative correlation with hantavirus infections

Understanding the influence of climatic drivers on hantavirus disease ecology can help in fore-casting and prevention, which is even more urgently needed in this era of climate change. Recently, Tian and colleagues [46] investigated the role extrinsic factors (climatic conditions) play in determining hantavirus disease dynamics in Central China. A unique data set from Central China covering half a century showed the existence of a climate-driven transmission mechanism for HTNV from the striped field mouse to humans [21]. HFRS outbreaks were highly correlated with specific environmental conditions—low summer temperature and abundant summer precipitation. Conversely, very few disease outbreaks occurred under conditions of high summer temperatures and drought (Fig 3A) [46]. In South China, HFRS incidence was also found to be positively correlated with annual precipitation and absolute humidity during 1991 to 2010 [78]. HPS outbreaks in the Four Corners region of the US are considered to be driven by ENSO-associated precipitation events as well [26,79].

However, precipitation can also negatively affect or have no effect on the incidence of hantavirus disease, depending on magnitude and region [80]. In Europe, no clear relationship of rainfall with human hantavirus infections was shown [77], except that one study illustrated rainfall in the summer three years before hantavirus disease incidence increased [50]. Other studies also demonstrated no significant association between rainfall and hantavirus disease [52,81,82]. In Cascade and Polson, Montana, US, increases in primary productivity, deer mouse population densities, and human disease risk are less sensitive to changes in the amount of rainfall [83]. In low-lying regions of China, such as Anhui Province, HFRS incidence and rodent population density (dominated by the striped field mouse) are all negatively correlated with the total precipitation [84–86]. Heavy precipitation there may kill rodents by flooding their burrows and nests, thereby reducing host population density and the risk of human exposure to pathogens. Floods could also cause movement of rodent populations (even community-level changes [87]) to new habitats and eventually disease emergence in new sites.

Temperature could influence disease dynamics through its impact on the rodent reservoir population dynamics and pathogen survival in the external environment, subsequently influencing human–animal contact patterns. However, compared to precipitation, the effects of temperature have been less investigated, and contradictory findings make it difficult to draw

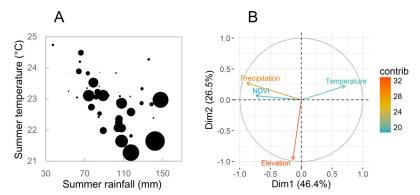


Fig 3. Environmental factors associated with hantavirus disease outbreaks. (A) Relationship between summer temperature, summer rainfall, and HFRS cases in Weihe Plain, North China, 1960 to 2013. Circle size is proportionate to the number of HFRS cases [46]. (B) Contribution of the environmental variables to the explained variance of hantavirus-antibody–positive in rodents using a multivariate principal component analysis in Hunan Province, South China, 2007 to 2010. Dimensions 1 and 2 are the spaces where variables are expressed. The length (angle) of the arrows represents the magnitude (direction) of the correlation coefficient between the variable and the principal components. The contributions of the variables to the hantavirus-antibody–positive in rodents are ranked with colors ranging from green to red, respectively (reproduced from Xiao 2016 with permission of the publisher [127]). Dim, dimension; HFRS, hemorrhagic fever with renal syndrome; NDVI, normalized difference vegetation index.

https://doi.org/10.1371/journal.pntd.0006901.g003

firm conclusions. For example, in Central China, higher summer temperatures were followed by a lower incidence of HFRS in autumn (Fig 3A) [46]—this was presumably due to a reduced frequency of contacts between rodents and between rodents and humans, coupled with an unfavorable environment for virus survival [88]—whereas in Belgium, a higher summer temperature two years before led to higher NE incidence [50,52]. A potential explanation is that the higher temperature two years before might have stimulated bud formation that contributed to heavy masting one year before. Likewise, opposite associations between mean temperature and hantavirus infection incidence were found across China [20,66,89,90]. In South America, HPS caused by ANDV was also negatively associated with winter temperature [91]. Conversely, in North America, a positive relationship between hantavirus disease incidence and temperature was reported [80]. There is a need for more studies to investigate the mechanisms behind these relationships.

#### Human-animal interface

Human-animal interface constitutes the boundary/barrier for cross species transmission of disease and the environment (including ecological and anthropological factors) within which these species exist [92,93]. In the preceding sections, we have summarized the ecological factors and processes that affect hantavirus diseases dynamics; hereafter, anthropological factors will be reviewed, including agricultural activity, human-driven land-use change, and vaccination. In HPS endemic regions of America, agriculture-associated activities were also most commonly reported as potential risk factors [94–97], and seropositive rodents were found with high frequency in agricultural landscapes [98,99]. In the HTNV-type endemic area of central China, seasonal pattern of HFRS dynamics is found to coincide with the increase in potential contact between rodents and humans in the dry season due to seasonal agricultural activities [45,100]. Besides, the breeding season of striped field mouse, the local rodent host, is closely associated with agricultural activity. Moreover, it could be concluded that agricultural activity may influence the activity and life cycles of local striped field mouse and in turn shape disease dynamics.

Human-driven landscape change could influence rodent host behavior and the composition of reservoir communities in such a way as to impact pathogen transmission [101,102]. The major consequences of landscape alteration are habitat loss and changes in species composition due to the loss of specialist species and the increase in generalist species [74], and these consequences either increase or decrease the risk of disease transmission at the human-animal interface. For example, HFRS epidemics in China peaked in autumn and winter in the area where the dominant hantavirus is the Apodemus-borne HTNV, whereas the area with Rattusborne SEOV saw epidemic peaks in spring [103]. These two distinct rodent species have different breeding sites with special landscape attributes [104], which changes the epidemiological characteristics of hantavirus disease. Human disturbance also affects survival probabilities and reproduction of rodent hosts [105]. Another consequence of landscape alteration is loss of biodiversity, which is considered to affect the transfer of pathogens among species and influence the risk of infection to humans. Biodiversity loss may result in increased hantavirus infection prevalence in host populations [106], affecting the dynamics of SNV in the US, Choclo hantavirus in Panama, and PUUV in Europe [68,107-111]. For some directly transmitted and vector-borne zoonotic diseases, it has been hypothesized that increased species diversity would result in a lower pathogen prevalence in competent hosts and therefore lower risk of infection to humans by a mechanism called the "dilution effect" [112], although there is still a debate about the (positive or negative) relationship between biodiversity and zoonotic disease transmission, e.g., SNV and deer mice system in America [113]. To our knowledge, few attempts

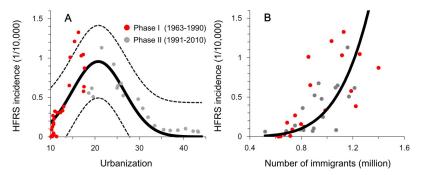


Fig 4. Urbanization, immigration, and hantavirus disease epidemics in an endemic area of south China. (A) Urbanization and HFRS incidence in Hunan Province. A biphasic inverted U-shaped relationship was found between hantavirus disease epidemics and urbanization. (B) The number of immigrants and HFRS incidence. This result indicates that the effect of urbanization on HFRS epidemics changed, whereas the effect of immigration remained constant. Source: Adapted from [114]. HFRS, hemorrhagic fever with renal syndrome.

https://doi.org/10.1371/journal.pntd.0006901.g004

have been made to evaluate this effect with regards to HTNV/striped field mouse system in China. Further testing of this theory is therefore still required. Another human-driven land-scape change is induced by urbanization (i.e., urban expansion). However, the relationship between the diffusion of zoonotic pathogens and urbanization is complex because of the contrasting effects (Fig 4): cities with faster economic growth may attract more immigrants and reach their endemic turning points later (endemic turning point defined as point at which incidence changes from increasing to decreasing), whereas economic growth that contributes towards improvements in the living conditions may decrease contact between rats and humans [114].

Vaccination programs against HFRS have been in practice for more than 20 years in China, where the incidence of HFRS has decreased with occasional small fluctuations. It has been reported that the age distribution of HFRS infections has clearly changed in recent years, ever since the Expanded Program of Immunization for regions with high HFRS incidence was implemented in 2008 [115]. Simulations conducted on data from a county in Central China have shown that vaccination will alter the dynamics of HFRS outbreaks [45]. The vaccination-induced reduction in susceptible human population gradually led to the reduction of human hantavirus infections and successfully averted further epidemics. The findings in China therefore highlight the necessity for a vaccination strategy and provide important insights for other countries.

### **Discussion**

The dynamics of hantavirus epidemics involve multiple phases, including environmental drivers that influence infectious diseases, transmission in the animal reservoir, and spillover transmission to humans. The complexity of disease dynamics has been highlighted in recent decades by contradicting trends; the incidence of HFRS has decreased in China [22,116] and increased in Europe [31]. Although pioneering research efforts to study both New World and Old World strains of hantaviruses in rodents have been undertaken, as well as related prevention strategies, it is evident that much work remains to be done. By further studying the transmission dynamics of hantaviruses, better prediction and prevention measures can be implemented to protect human health, and critical insight can be obtained into the ecology of hantaviruses and their rodent hosts.

The geographic distributions of hantaviruses reflect the distributions of their reservoir hosts [3], and our knowledge of the host associations of hantaviruses is expanding. For

example, the hantavirus isolated in Africa, Sangassou virus, was recently found in the African wood mouse (*Hylomyscus simus*) [117]. Although, in general, each hantavirus has been considered to be associated with a specific reservoir host, there have been cases that suggest rodent host expansion (e.g., pathogens that can infect multiple host species) for both HTNV and SEOV in China [22,118–120]. This challenges the strict rodent–virus coevolution theory and demonstrates that at least some hantaviruses can infect other susceptible rodents in addition to primary hosts, expanding the number of potential animal hosts [121]. These cases raise the question as to what role undetected or potential hosts play in hantavirus ecology and further influence the risk of human infections—a question largely ignored in the past and an area of further research. Further research is also needed to clarify the relationships between diversity and prevalence across types of ecosystems and host species, in particular, as the dilution effect on prevalence dynamics has broad potential applicability in predicting virus prevalence among rodent hosts.

To the best of our knowledge, this is the first systematic review to compare hantavirus disease dynamics, from environmental variability to rodent reservoir and to public health, among the main endemic areas across the globe. This review is timely because of the increasing public awareness of hantavirus diseases in Europe, in particular, over the last years. Limitations of this systematic review were that individual studies had differences—surveillance strategies for hantavirus infections, rodent sampling methods, rodent community compositions, socioeconomic factors, and environmental conditions. Therefore, studies were not all directly comparable, especially among China, European countries, and the US. Additionally, it should be noted that unusual human exposure (like war-induced exposure) to otherwise "normal" environment is an often forgotten risk factor for limited outbreaks [122]. Besides, there are more recent studies on ANDV-like viruses in South America that explore the environment conditions relevant to the occurrence of the host and the circulation of the virus, scrutinize the evidence for climate sensitivity of related disease risk, and recognize those areas of high risk for humans [91,123–126]. However, in this review, more attention was paid to longitudinal studies of sufficient scale. Finally, our focus on China in the review reflects the fact that for this country highly comprehensive data exist.

Establishing a mechanistically determined predictive framework for rodent-borne hantavirus disease prediction and prevention is urgently required to proactively protect the public from the increasing threat of hantaviruses. Such a framework would also provide insight into climate change, landscape alteration, rodent community composition, and pathogen spillover.

## **Key Learning Points**

With this review we have demonstrated the following:

- 1. The complex seasonality and interannual cycles of hantavirus disease dynamics are a direct result of the (nonlinear) interaction between the population dynamics of the rodent host, environmental forcing, and human–wildlife contact patterns.
- 2. Specific environmental conditions can trigger hantavirus disease outbreaks, but the outcomes may differ among strains or areas mediated by the underlying mechanisms of hantavirus transmission. For example, in Weihe Plain of Central China, higher summer temperatures were followed by a lower incidence of HFRS in autumn; conversely, in North America, a positive relationship between hantavirus disease incidence and temperature was reported, indicating the complexity across different systems.

3. Current efforts to curb hantavirus transmission focus on avoiding contact between humans and host rodents as there is no WHO-licensed vaccine against hantavirus infections available (except Hantavax, which is licensed for human use only in the Republic of Korea). Hence, a broader appreciation for the epidemiological links among human beings, animals, and environment can result in more effective control of disease outbreak.

# **Top Five Papers**

- 1. Yates TL, Mills JN, Parmenter CA, Ksiazek TG, Parmenter RR, et al. (2002) The ecology and evolutionary history of an emergent disease: hantavirus pulmonary syndrome. Bioscience 52: 989–998.
- 2. Glass GE, Yates TL, Fine JB, Shields TM, Kendall JB, et al. (2002) Satellite imagery characterizes local animal reservoir populations of Sin Nombre virus in the southwestern United States. Proc Natl Acad Sci USA 99: 16817–16822.
- 3. Clement J, Vercauteren J, Verstraeten W, Ducoffre G, Barrios J, et al. (2009) Relating increasing hantavirus incidences to the changing climate: the mast connection. International Journal of Health Geographics 8: 1.
- 4. Jonsson CB, Figueiredo LTM, Vapalahti O (2010) A global perspective on hantavirus ecology, epidemiology, and disease. Clinical Microbiology Reviews 23: 412–441.
- 5. Yan L, Fang LQ, Huang HG, Zhang LQ, Feng D, et al. (2007) Landscape elements and Hantaan virus-related hemorrhagic fever with renal syndrome, People's Republic of China. Emerging Infectious Diseases 13(9): 1301–1306.

# Acknowledgments

We thank Jan Clement and James N. Mills for their valuable comments, which have been very helpful in improving the manuscript.

#### References

- Maes P, Alkhovsky SV, Bào Y, Beer M, Birkhead M, et al. (2018) Taxonomy of the family Arenaviridae and the order Bunyavirales: update 2018. Arch Virol: <a href="https://doi.org/10.1007/s00705-00018-03843-00705">https://doi.org/10.1007/s00705-00018-03843-00705</a>
- Vaheri A, Strandin T, Hepojoki J, Sironen T, Henttonen H, et al. (2013) Uncovering the mysteries of hantavirus infections. Nat Rev Microbiol 11: 539–550. PMID: 24020072
- Jonsson CB, Figueiredo LTM, Vapalahti O (2010) A global perspective on hantavirus ecology, epidemiology, and disease. Clin Microbiol Rev 23: 412–441. https://doi.org/10.1128/CMR.00062-09 PMID: 20375360
- Hart C, Bennett M (1999) Hantavirus infections: epidemiology and pathogenesis. Microbes Infect 1: 1229–1237. PMID: 10580279



- Nichol ST, Spiropoulou CF, Morzunov S, Rollin PE, Ksiazek TG, et al. (1993) Genetic identification of a hantavirus associated with an outbreak of acute respiratory illness. Science 262: 914–917. PMID: 8235615
- Bi Z, Formenty PB, Roth CE (2008) Hantavirus infection: a review and global update. J Infect Dev Ctries 2: 3–23. PMID: 19736383
- Watson DC, Sargianou M, Papa A, Chra P, Starakis I, et al. (2014) Epidemiology of Hantavirus infections in humans: a comprehensive, global overview. Crit Rev Microbiol 40: 261–272. <a href="https://doi.org/10.3109/1040841X.2013.783555">https://doi.org/10.3109/1040841X.2013.783555</a> PMID: 23607444
- Ferrés M, Vial P, Marco C, Yanez L, Godoy P, et al. (2007) Prospective evaluation of household contacts of persons with hantavirus cardiopulmonary syndrome in Chile. J Infect Dis 195: 1563–1571. https://doi.org/10.1086/516786 PMID: 17471425
- Martinez VP, Bellomo C, San Juan J, Pinna D, Forlenza R, et al. (2005) Person-to-person transmission of Andes virus. Emerg Infect Dis 11: 1848–1853. https://doi.org/10.3201/eid1112.050501 PMID: 16485469
- Padula P, Edelstein A, Miguel S, Lopez N, Rossi C, et al. (1998) Hantavirus pulmonary syndrome outbreak in Argentina: molecular evidence for person-to-person transmission of Andes virus. Virology 241: 323–330. https://doi.org/10.1006/viro.1997.8976 PMID: 9499807
- Martinez-Valdebenito C, Calvo M, Vial C, Mansilla R, Marco C, et al. (2014) Person-to-person house-hold and nosocomial transmission of Andes hantavirus, Southern Chile, 2011. Emerg Infect Dis 20: 1629–1636. https://doi.org/10.3201/eid2010.140353 PMID: 25272189
- Hjelle B, Torres-Pérez F (2010) Hantaviruses in the Americas and their role as emerging pathogens.
  Viruses 2: 2559–2586. https://doi.org/10.3390/v2122559 PMID: 21994631
- Yu P, Tian H, Ma C, Ma C, Wei J, et al. (2015) Hantavirus infection in rodents and haemorrhagic fever with renal syndrome in Shaanxi province, China, 1984–2012. Epidemiol Infect 143: 405–411. https://doi.org/10.1017/S0950268814001009 PMID: 24787374
- Johnson PT, de Roode JC, Fenton A (2015) Why infectious disease research needs community ecology. Science 349: 1259504. https://doi.org/10.1126/science.1259504 PMID: 26339035
- Smith MJ, Telfer S, Kallio ER, Burthe S, Cook AR, et al. (2009) Host–pathogen time series data in wildlife support a transmission function between density and frequency dependence. Proc Natl Acad Sci USA 106: 7905–7909. https://doi.org/10.1073/pnas.0809145106 PMID: 19416827
- Mills JN, Ksiazek TG, Ellis BA, Rollin PE, Nichol ST, et al. (1997) Patterns of association with host and habitat: antibody reactive with Sin Nombre virus in small mammals in the major biotic communities of the southwestern United States. Am J Trop Med Hyg 56: 273–284. PMID: 9129529
- Calisher CH, Sweeney W, Mills JN, Beaty BJ (1999) Natural history of Sin Nombre virus in western Colorado. Emerg Infect Dis 5: 126–134. https://doi.org/10.3201/eid0501.990115 PMID: 10081680
- Douglass RJ, Wilson T, Semmens WJ, Zanto SN, Bond CW, et al. (2001) Longitudinal studies of Sin Nombre virus in deer mouse-dominated ecosystems of Montana. Am J Trop Med Hyg 65: 33–41. PMID: 11504405
- Jiang H, Zheng X, Wang L, Du H, Wang P, et al. (2017) Hantavirus infection: a global zoonotic challenge. Virol Sin 32: 32–43. https://doi.org/10.1007/s12250-016-3899-x PMID: 28120221
- Xiao H, Tian HY, Gao LD, Liu HN, Duan LS, et al. (2014) Animal reservoir, natural and socioeconomic variations and the transmission of hemorrhagic fever with renal syndrome in Chenzhou, China, 2006–2010. PLoS Negl Trop Dis 8: e2615. https://doi.org/10.1371/journal.pntd.0002615 PMID: 24421910
- Forbes KM, Sironen T, Plyusnin A (2018) Hantavirus maintenance and transmission in reservoir host populations. Curr Opin Virol 28: 1–6. https://doi.org/10.1016/j.coviro.2017.09.003 PMID: 29024905
- 22. Zhang YZ, Zou Y, Fu ZF, Plyusnin A (2010) Hantavirus infections in humans and animals, China. Emerg Infect Dis 16: 1195–1203. https://doi.org/10.3201/eid1608.090470 PMID: 20678311
- Lee H, French G, Lee P, Baek L, Tsuchiya K, et al. (1981) Observations on natural and laboratory infection of rodents with the etiologic agent of Korean hemorrhagic fever. Am J Trop Med Hyg 30: 477–482. PMID: 6786119
- Lee HW, Lee PW, Johnson KM (1978) Isolation of the etiologic agent of Korean hemorrhagic fever. J Infect Dis 137: 298–308. PMID: 24670
- Kariwa H, Yoshimatsu K, Arikawa J (2007) Hantavirus infection in East Asia. Comp Immunol Microbiol Infect Dis 30: 341–356. https://doi.org/10.1016/j.cimid.2007.05.011 PMID: 17655929
- Glass GE, Cheek JE, Patz JA, Shields TM, Doyle TJ, et al. (2000) Using remotely sensed data to identify areas at risk for hantavirus pulmonary syndrome. Emerg Infect Dis 6: 238–247. <a href="https://doi.org/10.3201/eid0603.000303">https://doi.org/10.3201/eid0603.000303</a> PMID: 10827113



- 27. Yates TL, Mills JN, Parmenter CA, Ksiazek TG, Parmenter RR, et al. (2002) The ecology and evolutionary history of an emergent disease: hantavirus pulmonary syndrome. Bioscience 52: 989–998.
- 28. Mills JN, Yates TL, Ksiazek TG, Peters C, Childs JE (1999) Long-term studies of hantavirus reservoir populations in the southwestern United States: rationale, potential, and methods. Emerg Infect Dis 5: 95–101. https://doi.org/10.3201/eid0501.990111 PMID: 10081676
- **29.** Clement J, Colson P, McKenna P (1994) Hantavirus pulmonary syndrome in New England and Europe. N Engl J Med 331: 545–546.
- Olsson GE, Leirs H, Henttonen H (2010) Hantaviruses and their hosts in Europe: reservoirs here and there, but not everywhere? Vector Borne Zoonotic Dis 10: 549–561. <a href="https://doi.org/10.1089/vbz.2009.0138">https://doi.org/10.1089/vbz.2009.0138</a> PMID: 20795916
- Vaheri A, Henttonen H, Voutilainen L, Mustonen J, Sironen T, et al. (2013) Hantavirus infections in Europe and their impact on public health. Rev Med Virol 23: 35–49. <a href="https://doi.org/10.1002/rmv.1722">https://doi.org/10.1002/rmv.1722</a> PMID: 22761056
- Vapalahti O, Mustonen J, Lundkvist Å, Henttonen H, Plyusnin A, et al. (2003) Hantavirus infections in Europe. Lancet Infect Dis 3: 653–661. PMID: 14522264
- 33. Clement J, Maes P, Lagrou K, Van Ranst M, Lameire N (2012) A unifying hypothesis and a single name for a complex globally emerging infection: hantavirus disease. Eur J Clin Microbiol Infect Dis 31: 1–5. https://doi.org/10.1007/s10096-011-1456-y PMID: 22068273
- 34. Witkowski PT, Klempa B, Ithete NL, Auste B, Mfune JK, et al. (2014) Hantaviruses in Africa. Virus Res 187: 34–42. https://doi.org/10.1016/j.virusres.2013.12.039 PMID: 24406800
- Kruger DH, Figueiredo LTM, Song J-W, Klempa B (2015) Hantaviruses—globally emerging pathogens. J Clin Virol 64: 128–136. https://doi.org/10.1016/j.jcv.2014.08.033 PMID: 25453325
- Reynes JM, Carli D, Bour JB, Boudjeltia S, Dewilde A, et al. (2017) Seoul Virus Infection in Humans, France, 2014–2016. Emerg Infect Dis 23: 973–977. https://doi.org/10.3201/eid2306.160927 PMID: 28368241
- McElhinney LM, Marston DA, Pounder KC, Goharriz H, Wise EL, et al. (2017) High prevalence of Seoul hantavirus in a breeding colony of pet rats. Epidemiol Infect 145: 3115–3124. <a href="https://doi.org/10.1017/S0950268817001819">https://doi.org/10.1017/S0950268817001819</a> PMID: 28965516
- Fill MA, Mullins H, May AS, Henderson H, Brown SM, et al. (2017) Notes from the Field: Multiple Cases of Seoul Virus Infection in a Household with Infected Pet Rats—Tennessee, December 2016— April 2017. MMWR Morb Mortal Wkly Rep 66: 1081–1082. <a href="https://doi.org/10.15585/mmwr.mm6640a4">https://doi.org/10.15585/mmwr.mm6640a4</a> PMID: 29023435
- Kerins JL, Koske SE, Kazmierczak J, Austin C, Gowdy K, et al. (2018) Outbreak of Seoul virus among rats and rat owners—United States and Canada, 2017. MMWR Morb Mortal Wkly Rep 67: 131–134. https://doi.org/10.15585/mmwr.mm6704a5 PMID: 29393924
- 40. Parmenter RR, Brunt JW, Moore DI, Ernest S (1993) The hantavirus epidemic in the Southwest: rodent population dynamics and the implications for transmission of hantavirus-associated adult respiratory distress syndrome (HARDS) in the Four Corners region. Publication No. 31. Sevilleta Long-TermEcological Research Program (LTER), 1993.
- **41.** Yu P, Li Y, Xu B, Wei J, Li S, et al. (2017) Using satellite data for the characterization of local animal reservoir populations of Hantaan virus on the Weihe Plain, China. Remote Sensing 9: 1076.
- **42.** Kagata H, Ohgushi T (2006) Bottom-up trophic cascades and material transfer in terrestrial food webs. Ecol Res 21: 26–34.
- Heath MR, Speirs DC, Steele JH (2014) Understanding patterns and processes in models of trophic cascades. Ecol Lett 17: 101–114. https://doi.org/10.1111/ele.12200 PMID: 24165353
- 44. Hansen A, Cameron S, Liu Q, Sun Y, Weinstein P, et al. (2015) Transmission of haemorrhagic fever with renal syndrome in China and the role of climate factors: a review. Int J Infect Dis 33: 212–218. https://doi.org/10.1016/j.ijid.2015.02.010 PMID: 25704595
- **45.** Tian H, Yu P, Bjørnstad ON, Cazelles B, Yang J, et al. (2017) Anthropogenically driven environmental changes shift the ecological dynamics of hemorrhagic fever with renal syndrome. PLoS Pathog 13: e1006198. https://doi.org/10.1371/journal.ppat.1006198 PMID: 28141833
- 46. Tian H, Yu P, Cazelles B, Xu L, Tan H, et al. (2017) Interannual cycles of Hantaan virus outbreaks at the human–animal interface in Central China are controlled by temperature and rainfall. Proc Natl Acad Sci USA 114: 8041–8046. https://doi.org/10.1073/pnas.1701777114 PMID: 28696305
- 47. Xiao H, Liu HN, Gao LD, Huang CR, Li Z, et al. (2013) Investigating the effects of food available and climatic variables on the animal host density of hemorrhagic fever with renal syndrome in Changsha, China. PLoS ONE 8: e61536. https://doi.org/10.1371/journal.pone.0061536 PMID: 23637849
- **48.** Guan P, Huang D, He M, Shen T, Guo J, et al. (2009) Investigating the effects of climatic variables and reservoir on the incidence of hemorrhagic fever with renal syndrome in Huludao City, China: a 17-year

- data analysis based on structure equation model. BMC Infect Dis 9: 109. https://doi.org/10.1186/1471-2334-9-109 PMID: 19583875
- **49.** Pucek Z, Jedrzejewski W, Jedrzejewska B, Pucek M (1993) Rodent population dynamics in a primeval deciduous forest (Bialowieza National Park) in relation to weather, seed crop, and predation. Acta Theriol 38: 199–232.
- Clement J, Vercauteren J, Verstraeten W, Ducoffre G, Barrios J, et al. (2009) Relating increasing hantavirus incidences to the changing climate: the mast connection. Int J Health Geogr 8: 1. <a href="https://doi.org/10.1186/1476-072X-8-1">https://doi.org/10.1186/1476-072X-8-1</a> PMID: 19149870
- Haredasht SA, Barrios JM, Maes P, Verstraeten WW, Clement J, et al. (2011) A dynamic data-based model describing nephropathia epidemica in Belgium. Biosyst Eng 109: 77–89.
- 52. Tersago K, Verhagen R, Servais A, al e (2009) Hantavirus disease (nephropathia epidemica) in Belgium: effects of tree seed production and climate. Epidemiol Infect 137: 250–256. <a href="https://doi.org/10.1017/S0950268808000940">https://doi.org/10.1017/S0950268808000940</a> PMID: 18606026
- Schwarz AC, Ranft U, Piechotowski I, Childs JE, Brockmann SO (2009) Risk factors for human infection with Puumala virus, southwestern Germany. Emerg Infect Dis 15: 1032–1039. <a href="https://doi.org/10.3201/eid1507.081413">https://doi.org/10.3201/eid1507.081413</a> PMID: 19624917
- 54. Glass GE, Shields T, Cai B, Yates TL, Parmenter R (2007) Persistently highest risk areas for hantavirus pulmonary syndrome: potential sites for refugia. Ecol Appl 17: 129–139. PMID: 17479840
- 55. Luis AD, Douglass RJ, Mills JN, Bjørnstad ON (2010) The effect of seasonality, density and climate on the population dynamics of Montana deer mice, important reservoir hosts for Sin Nombre hantavirus. J Anim Ecol 79: 462–470. https://doi.org/10.1111/j.1365-2656.2009.01646.x PMID: 20015212
- 56. Smith KF, Dobson AP, McKenzie FE, Real LA, Smith DL, et al. (2005) Ecological theory to enhance infectious disease control and public health policy. Front Ecol Environ 3: 29–37. https://doi.org/10.1890/1540-9295(2005)003[0029:ETTEID]2.0.CO;2 PMID: 19838319
- 57. Khalil H, Hörnfeldt B, Evander M, Magnusson M, Olsson G, et al. (2014) Dynamics and drivers of hantavirus prevalence in rodent populations. Vector Borne Zoonotic Dis 14: 537–551. <a href="https://doi.org/10.1089/vbz.2013.1562">https://doi.org/10.1089/vbz.2013.1562</a> PMID: 25072983
- **58.** Luis AD, Douglass RJ, Mills JN, Bjørnstad ON (2015) Environmental fluctuations lead to predictability in Sin Nombre hantavirus outbreaks. Ecology 96: 1691–1701.
- 59. Boone JD, Otteson EW, McGwire KC, Villard P, Rowe JE, et al. (1998) Ecology and demographics of hantavirus infections in rodent populations in the Walker River Basin of Nevada and California. Am J Trop Med Hyg 59: 445–451. PMID: 9749642
- 60. Xiao H, Tong X, Huang R, Gao L, Hu S, et al. (2018) Landscape and rodent community composition are associated with risk of hemorrhagic fever with renal syndrome in two cities in China, 2006–2013. BMC Infect Dis 18: 37. https://doi.org/10.1186/s12879-017-2827-5 PMID: 29329512
- Olsson GE, White N, Ahlm C, Elgh F, Verlemyr A-C, et al. (2002) Demographic factors associated with hantavirus infection in bank voles (Clethrionomys glareolus). Emerg Infect Dis 8: 924–929. <a href="https://doi.org/10.3201/eid0809.020037">https://doi.org/10.3201/eid0809.020037</a> PMID: 12194768
- Olsson GE, Hjertqvist M, Lundkvist Å, Hörnfeldt B (2009) Predicting high risk for human hantavirus infections, Sweden. Emerg Infect Dis 15: 104–106. <a href="https://doi.org/10.3201/eid1501.080502">https://doi.org/10.3201/eid1501.080502</a> PMID: 19116065
- Kallio ER, Begon M, Henttonen H, Koskela E, Mappes T, et al. (2009) Cyclic hantavirus epidemics in humans—predicted by rodent host dynamics. Epidemics 1: 101–107. https://doi.org/10.1016/j. epidem.2009.03.002 PMID: 21352757
- 64. Milhano N, Korslund L, Evander M, Ahlm C, Vainio K, et al. (2017) Circulation and diagnostics of Puumala virus in Norway: nephropatia epidemica incidence and rodent population dynamics. Apmis 125: 732–742. https://doi.org/10.1111/apm.12712 PMID: 28585306
- 65. Xiao H, Tian HY, Gao LD, Liu HN, Duan LS, et al. (2014) Animal reservoir, natural and socioeconomic variations and the transmission of hemorrhagic fever with renal syndrome in Chenzhou, China, 2006–2010. PLoS Negl Trop Dis 8: e2615. https://doi.org/10.1371/journal.pntd.0002615 PMID: 24421910
- 66. Xiao H, Gao L, Li X, Lin X, Dai X, et al. (2013) Environmental variability and the transmission of haemorrhagic fever with renal syndrome in Changsha, People's Republic of China. Epidemiol Infect 141: 1867–1875. https://doi.org/10.1017/S0950268812002555 PMID: 23158456
- Calisher CH, Wagoner KD, Amman BR, Root JJ, Douglass RJ, et al. (2007) Demographic factors associated with prevalence of antibody to Sin Nombre virus in deer mice in the western United States. J Wildl Dis 43: 1–11. https://doi.org/10.7589/0090-3558-43.1.1 PMID: 17347388
- 68. Clay CA, Lehmer EM, Jeor SS, Dearing MD (2009) Sin Nombre virus and rodent species diversity: a test of the dilution and amplification hypotheses. PLoS ONE 4: e6467. <a href="https://doi.org/10.1371/journal.pone.0006467">https://doi.org/10.1371/journal.pone.0006467</a> PMID: 19649283

- 69. Pearce-Duvet JM, St. Jeor SC, Boone JD, Dearing MD(2006) Changes in Sin Nombre virus antibody prevalence in deer mice across seasons: the interaction between habitat, sex, and infection in deer mice. J Wildl Dis 42: 819–824. https://doi.org/10.7589/0090-3558-42.4.819 PMID: 17255449
- Calisher CH, Root JJ, Mills JN, Rowe JE, Reeder SA, et al. (2005) Epizootiology of Sin Nombre and El Moro Canyon hantaviruses, southeastern Colorado, 1995–2000. J Wildl Dis 41: 1–11. <a href="https://doi.org/10.7589/0090-3558-41.1.1">https://doi.org/10.7589/0090-3558-41.1.1</a> PMID: 15827206
- Biggs JR, Bennett KD, Mullen MA, Haarmann TK, Salisbury M, et al. (2000) Relationship of ecological variables to Sin Nombre virus antibody seroprevalence in populations of deer mice. J Mammal 81: 676–682
- Mills JN, Ksiazek TG, Peters C, Childs JE (1999) Long-term studies of hantavirus reservoir populations in the southwestern United States: a synthesis. Emerg Infect Dis 5: 135–142. <a href="https://doi.org/10.3201/eid0501.990116">https://doi.org/10.3201/eid0501.990116</a> PMID: 10081681
- Davis S, Calvet E, Leirs H (2005) Fluctuating rodent populations and risk to humans from rodentborne zoonoses. Vector Borne Zoonotic Dis 5: 305–314. https://doi.org/10.1089/vbz.2005.5.305 PMID: 16417426
- 74. Prist PR, Metzger JP Landscape, climate and hantavirus cardiopulmonary syndrome outbreaks. Eco-Health 14: 614–629. https://doi.org/10.1007/s10393-017-1255-8 PMID: 28620680
- 75. Patz JA, Campbell-Lendrum D, Holloway T, Foley JA (2005) Impact of regional climate change on human health. Nature 438: 310–317. https://doi.org/10.1038/nature04188 PMID: 16292302
- 76. Altizer S, Ostfeld RS, Johnson PT, Kutz S, Harvell CD (2013) Climate change and infectious diseases: from evidence to a predictive framework. Science 341: 514–519. <a href="https://doi.org/10.1126/science.1239401">https://doi.org/10.1126/science.1239401</a> PMID: 23908230
- Roda Gracia J, Schumann B, Seidler A (2015) Climate variability and the occurrence of human puumala hantavirus infections in Europe: a systematic review. Zoonoses Public Health 62: 465–478. https://doi.org/10.1111/zph.12175 PMID: 25557350
- 78. Xiao H, Tian HY, Cazelles B, Li XJ, Tong SL, et al. (2013) Atmospheric moisture variability and transmission of hemorrhagic fever with renal syndrome in Changsha City, Mainland China, 1991–2010. PLoS Negl Trop Dis 7: e2260. https://doi.org/10.1371/journal.pntd.0002260 PMID: 23755316
- 79. Glass GE, Yates TL, Fine JB, Shields TM, Kendall JB, et al. (2002) Satellite imagery characterizes local animal reservoir populations of Sin Nombre virus in the southwestern United States. Proc Natl Acad Sci USA 99: 16817–16822. https://doi.org/10.1073/pnas.252617999 PMID: 12473747
- **80.** Carver S, Mills JN, Parmenter CA, Parmenter RR, Richardson KS, et al. (2015) Toward a mechanistic understanding of environmentally forced zoonotic disease emergence: Sin Nombre Hantavirus. Bio-Science 65: 651–666. https://doi.org/10.1093/biosci/biv047 PMID: 26955081
- Linard C, Tersago K, Leirs H, Lambin EF (2007) Environmental conditions and Puumala virus transmission in Belgium. Int J Health Geogr 6: 55. <a href="https://doi.org/10.1186/1476-072X-6-55">https://doi.org/10.1186/1476-072X-6-55</a> PMID: 18078526
- 82. Viel JF, Lefebvre A, Marianneau P, Joly D, Giraudoux P, et al. (2011) Environmental risk factors for haemorrhagic fever with renal syndrome in a French new epidemic area. Epidemiol Infect 139: 867–874. https://doi.org/10.1017/S0950268810002062 PMID: 20822577
- 83. Loehman RA, Elias J, Douglass RJ, Kuenzi AJ, Mills JN, et al. (2012) Prediction of Peromyscus maniculatus (deer mouse) population dynamics in Montana, USA, using satellite-driven vegetation productivity and weather data. J Wildl Dis 48: 348–360. https://doi.org/10.7589/0090-3558-48.2.348 PMID: 22493110
- 84. Bi P, Parton KA (2003) El Niño and incidence of hemorrhagic fever with renal syndrome in China. JAMA 289: 176–177.
- Bi P, Tong S, Donald K, Parton K, Ni J (2002) Climatic, reservoir and occupational variables and the transmission of haemorrhagic fever with renal syndrome in China. Int J Epidemiol 31: 189–193. PMID: 11914320
- 86. Bi P, Wu X, Zhang F, Parton KA, Tong S (1998) Seasonal rainfall variability, the incidence of hemorrhagic fever with renal syndrome, and prediction of the disease in low-lying areas of China. Am J Epidemiol 148: 276–281. PMID: 9690365
- Thibault KM, Brown JH (2008) Impact of an extreme climatic event on community assembly. Proc Natl Acad Sci USA 105: 3410–3415. https://doi.org/10.1073/pnas.0712282105 PMID: 18303115
- **88.** Kallio ER, Klingström J, Gustafsson E, Manni T, Vaheri A, et al. (2006) Prolonged survival of Puumala hantavirus outside the host: evidence for indirect transmission via the environment. J Gen Virol 87: 2127–2134. https://doi.org/10.1099/vir.0.81643-0 PMID: 16847107



- 89. Zhang WY, Guo WD, Fang LQ, Li CP, Bi P, et al. (2010) Climate variability and hemorrhagic fever with renal syndrome transmission in Northeastern China. Environ Health Perspect 118: 915–920. https:// doi.org/10.1289/ehp.0901504 PMID: 20142167
- 90. Fang L, Wang X, Liang S, Li Y, Song S, et al. (2010) Spatiotemporal trends and climatic factors of hemorrhagic fever with renal syndrome epidemic in Shandong Province, China. PLoS Negl Trop Dis 4: e789. https://doi.org/10.1371/journal.pntd.0000789 PMID: 20706629
- 91. Andreo V, Neteler M, Rocchini D, Provensal C, Levis S, et al. (2014) Estimating Hantavirus risk in southern Argentina: a GIS-based approach combining human cases and host distribution. Viruses 6: 201–222. https://doi.org/10.3390/v6010201 PMID: 24424500
- 92. Hassell JM, Begon M, Ward MJ, Fèvre EM (2017) Urbanization and disease emergence: Dynamics at the wildlife–livestock–human interface. Trends Ecol Evol 32: 55–67. https://doi.org/10.1016/j.tree. 2016.09.012 PMID: 28029378
- Reperant LA, Cornaglia G, Osterhaus AD (2012) The importance of understanding the human–animal interface. Curr Top Microbiol Immunol 365: 49–81.
- Martinez VP, Bellomo CM, Cacace ML, Suárez P, Bogni L, et al. (2010) Hantavirus pulmonary syndrome in Argentina, 1995–2008. Emerg Infect Dis 16: 1853–1860. https://doi.org/10.3201/eid1612.091170 PMID: 21122213
- 95. Mendes WS, Da Silva AA, Aragão LF, Aragão NJ, Raposo MdL, et al. (2004) Hantavirus infection in Anajatuba, Maranhao, Brazil. Emerg Infect Dis 10: 1496–1498. <a href="https://doi.org/10.3201/eid1008.040002">https://doi.org/10.3201/eid1008.040002</a> PMID: 15496259
- 96. Matheus S, Djossou F, Moua D, Bourbigot AM, Hommel D, et al. (2010) Hantavirus pulmonary syndrome, French Guiana. Emerg Infect Dis 16: 739–741. https://doi.org/10.3201/eid1604.090831 PMID: 20350412
- 97. Maurice AdS, Ervin E, Schumacher M, Yaglom H, VinHatton E, et al. (2017) Exposure characteristics of hantavirus pulmonary syndrome patients, United States, 1993–2015. Emerg Infect Dis 23: 733–739. https://doi.org/10.3201/eid2305.161770 PMID: 28418312
- **98.** Goodin DG, Koch DE, Owen RD, Chu YK, Hutchinson JS, et al. (2006) Land cover associated with hantavirus presence in Paraguay. Glob Ecol Biogeogr 15: 519–527.
- 99. Armién B, Ortiz PL, Gonzalez P, Cumbrera A, Rivero A, et al. (2016) Spatial-Temporal Distribution of Hantavirus Rodent-Borne Infection by Oligoryzomys fulvescens in the Agua Buena Region-Panama. PLoS Negl Trop Dis 10: e0004460. https://doi.org/10.1371/journal.pntd.0004460 PMID: 26894436
- 100. Tian H, Yu P, Luis AD, Bi P, Cazelles B, et al. (2015) Changes in rodent abundance and weather conditions potentially drive hemorrhagic fever with renal syndrome outbreaks in Xi'an, China, 2005–2012. PLoS Negl Trop Dis 9: e0003530. https://doi.org/10.1371/journal.pntd.0003530 PMID: 25822936
- 101. Patz JA, Daszak P, Tabor GM, Aguirre AA, Pearl M, et al. (2004) Unhealthy landscapes: Policy recommendations on land use change and infectious disease emergence. Environ Health Perspect 112: 1092–1098. https://doi.org/10.1289/ehp.6877 PMID: 15238283
- 102. Lambin EF, Tran A, Vanwambeke SO, Linard C, Soti V (2010) Pathogenic landscapes: interactions between land, people, disease vectors, and their animal hosts. Int J Health Geogr 9: 54. https://doi.org/10.1186/1476-072X-9-54 PMID: 20979609
- 103. Li LM (2007) Epidemiology (Sixth Edition). Beijing: People's Medical Publishing House.
- 104. Yan L, Fang LQ, Huang HG, Zhang LQ, Feng D, et al. (2007) Landscape elements and Hantaan virus-related hemorrhagic fever with renal syndrome, People's Republic of China. Emerg Infect Dis 13: 1301–1306. https://doi.org/10.3201/eid1309.061481 PMID: 18252099
- 105. Previtali MA, Lehmer EM, Pearce-Duvet J, Jones JD, Clay CA, et al. (2010) Roles of human disturbance, precipitation, and a pathogen on the survival and reproductive probabilities of deer mice. Ecology 91: 582–592. PMID: 20392022
- 106. Keesing F, Belden LK, Daszak P, Dobson A, Harvell CD, et al. (2010) Impacts of biodiversity on the emergence and transmission of infectious diseases. Nature 468: 647–652. https://doi.org/10.1038/nature09575 PMID: 21124449
- **107.** Mills JN (2006) Biodiversity loss and emerging infectious disease: an example from the rodent-borne hemorrhagic fevers. Biodiversity 7: 9–17.
- 108. Tersago K, Schreurs A, Linard C, Verhagen R, Van Dongen S, et al. (2008) Population, environmental, and community effects on local bank vole (Myodes glareolus) Puumala virus infection in an area with low human incidence. Vector Borne Zoonotic Dis 8: 235–244. https://doi.org/10.1089/vbz.2007.0160 PMID: 18370592
- 109. Dizney LJ, Ruedas LA (2009) Increased host species diversity and decreased prevalence of Sin Nombre virus. Emerg Infect Dis 15: 1012–1018. https://doi.org/10.3201/eid1507.081621 PMID: 19624913



- 110. Suzán G, Marcé E, Giermakowski JT, Mills JN, Ceballos G, et al. (2009) Experimental evidence for reduced rodent diversity causing increased hantavirus prevalence. PLoS ONE 4: e5461. https://doi.org/10.1371/journal.pone.0005461 PMID: 19421313
- 111. Clay CA, Lehmer EM, Jeor SS, Dearing MD (2009) Testing mechanisms of the dilution effect: deer mice encounter rates, Sin Nombre virus prevalence and species diversity. EcoHealth 6: 250–259. https://doi.org/10.1007/s10393-009-0240-2 PMID: 19495881
- 112. Mills JN (2005) Regulation of rodent-borne viruses in the natural host: implications for human disease. Arch Virol Suppl 19: 45–57.
- 113. Luis AD, Kuenzi AJ, Mills JN (2018) Species diversity concurrently dilutes and amplifies transmission in a zoonotic host–pathogen system through competing mechanisms. Proc Natl Acad Sci USA 115: 7979–7984. https://doi.org/10.1073/pnas.1807106115 PMID: 30012590
- 114. Tian HY, Hu SX, Cazelles B, Chowell G, Gao LD, et al. (2018) Urbanization prolongs hantavirus epidemics in cities. Proc Natl Acad Sci USA 115: 4707–4712. https://doi.org/10.1073/pnas.1712767115 PMID: 29666240
- 115. He X, Wang S, Huang X, Wang X (2013) Changes in age distribution of hemorrhagic fever with renal syndrome: an implication of China's expanded program of immunization. BMC Public Health 13: 394. https://doi.org/10.1186/1471-2458-13-394 PMID: 23622420
- 116. Fang L, Yan L, Liang S, de Vlas S, Feng D, et al. (2006) Spatial analysis of hemorrhagic fever with renal syndrome in China. BMC Infect Dis 6: 77. https://doi.org/10.1186/1471-2334-6-77 PMID: 16638156
- 117. Klempa B, Fichet-Calvet E, Lecompte E, Auste B, Aniskin V, et al. (2006) Hantavirus in African wood mouse, Guinea. Emerg Infect Dis 12: 838–840. <a href="https://doi.org/10.3201/eid1205.051487">https://doi.org/10.3201/eid1205.051487</a> PMID: 16704849
- 118. Fang L, Zhao L, Wen H, Zhang Z, Liu J, et al. (2015) Reservoir host expansion of hantavirus, China. Emerg Infect Dis 21: 170–171. https://doi.org/10.3201/eid2101.140960 PMID: 25531113
- **119.** Zhang WY, Fang LQ, Jiang JF, Hui FM, Glass GE, et al. (2009) Predicting the risk of hantavirus infection in Beijing, People's Republic of China. Am J Trop Med Hyg 80: 678–683. PMID: 19346399
- **120.** Ma C, Yu P, Nawaz M, Zuo S, Jin T, et al. (2012) Hantaviruses in rodents and humans, Xi'an, PR China. J Gen Virol 93: 2227–2236. https://doi.org/10.1099/vir.0.043364-0 PMID: 22837422
- 121. Clement J, Mc Kenna P, Leirs H, Verhagen R, Lefevre A, et al. (1994) Virus infections of Rodents and lagomorphs, 5th Volume in a series (Osterhaus, ed). In: Horzinek M, editor. Virus infections in Vertebrates. Amsterdam: Elsevier Science BV. pp. 295–316.
- **122.** Clement J, Underwood P, Ward D, Pilaski J, LeDuc J (1996) Hantavirus outbreak during military manoeuvres in Germany. Lancet 347: 336.
- 123. Andreo V, Glass G, Shields T, Provensal C, Polop J (2011) Modeling potential distribution of oligoryzomys longicaudatus, the andes virus (Genus: Hantavirus) reservoir, in Argentina. Ecohealth 8: 332–348. https://doi.org/10.1007/s10393-011-0719-5 PMID: 22130568
- 124. Piudo L, Monteverde MJ, Walker RS, Douglass RJ (2011) Rodent community structure and Andes virus infection in sylvan and peridomestic habitats in northwestern Patagonia, Argentina. Vector Borne Zoonotic Dis 11: 315–324. https://doi.org/10.1089/vbz.2009.0242 PMID: 21332352
- 125. Medina RA, Torres-Perez F, Galeno H, Navarrete M, Vial PA, et al. (2009) Ecology, genetic diversity, and phylogeographic structure of Andes virus in humans and rodents in Chile. J Virol 83: 2446–2459. https://doi.org/10.1128/JVI.01057-08 PMID: 19116256
- 126. Astorga F, Escobar LE, Poo-Muñoz D, Escobar-Dodero J, Rojas-Hucks S, et al. (2018) Distributional ecology of Andes hantavirus: a macroecological approach. Int J Health Geogr 17: 22. <a href="https://doi.org/10.1186/s12942-018-0142-z">https://doi.org/10.1186/s12942-018-0142-z</a> PMID: 29929522
- 127. Xiao H, Huang R, Gao L, Huang C, Lin X, et al. (2016) Effects of humidity variation on the hantavirus infection and hemorrhagic fever with renal syndrome occurrence in subtropical China. Am J Trop Med Hyg 94: 420–427. https://doi.org/10.4269/ajtmh.15-0486 PMID: 26711521
- 128. Prist PR, Uriarte M, Tambosi LR, Prado A, Pardini R, et al. (2016) Landscape, environmental and social predictors of Hantavirus risk in S\u00e3o Paulo, Brazil. PLoS ONE 11: e0163459. https://doi.org/10.1371/journal.pone.0163459 PMID: 27780250
- 129. Li C, Cui Z, Li S, Magalhaes RJS, Wang B, et al. (2013) Association between hemorrhagic fever with renal syndrome epidemic and climate factors in Heilongjiang Province, China. Am J Trop Med Hyg 89: 1006–1012. https://doi.org/10.4269/ajtmh.12-0473 PMID: 24019443
- 130. Han SS, Kim S, Choi Y, Kim S, Kim YS (2013) Air pollution and hemorrhagic fever with renal syndrome in South Korea: an ecological correlation study. BMC Public Health 13: 347. https://doi.org/10.1186/1471-2458-13-347 PMID: 23587219



- 131. Donalisio MR, Peterson AT (2011) Environmental factors affecting transmission risk for hantaviruses in forested portions of southern Brazil. Acta Tropica 119: 125–130. https://doi.org/10.1016/j.actatropica.2011.04.019 PMID: 21605537
- 132. Nsoesie EO, Mekaru SR, Ramakrishnan N, Marathe MV, Brownstein JS (2014) Modeling to predict cases of hantavirus pulmonary syndrome in Chile. PLoS Negl Trop Dis 8: e2779. https://doi.org/10.1371/journal.pntd.0002779 PMID: 24763320
- 133. Lin H, Zhang Z, Lu L, Li X, Liu Q (2014) Meteorological factors are associated with hemorrhagic fever with renal syndrome in Jiaonan County, China, 2006–2011. Int J Biometeorol 58: 1031–1037. https://doi.org/10.1007/s00484-013-0688-1 PMID: 23793957
- 134. Lin H, Liu Q, Guo J, Zhang J, Wang J, et al. (2007) Analysis of the geographic distribution of HFRS in Liaoning Province between 2000 and 2005. BMC Public Health 7: 207. <a href="https://doi.org/10.1186/1471-2458-7-207">https://doi.org/10.1186/1471-2458-7-207</a> PMID: 17697362
- 135. Bi P, Parton KA, Tong S (2005) El Nino–southern oscillation and vector-borne diseases in Anhui, China. Vector Borne Zoonotic Dis 5: 95–100. https://doi.org/10.1089/vbz.2005.5.95 PMID: 16011424
- 136. Hjelle B, Glass GE (2000) Outbreak of hantavirus infection in the Four Corners region of the United States in the wake of the 1997–1998 El Nino—Southern Oscillation. J Infect Dis 181: 1569–1573. https://doi.org/10.1086/315467 PMID: 10823755
- 137. Fang LQ, Zhao WJ, de Vlas SJ, Zhang WY, Liang S, et al. (2009) Spatiotemporal dynamics of hemorrhagic fever with renal syndrome, Beijing, People's Republic of China. Emerg Infect Dis 15: 2043–2045. https://doi.org/10.3201/eid1512.081078 PMID: 19961697
- 138. Clement J, Maes P, Van Ypersele de Strihou C, van der Groen G, Barrios JM, et al. (2010) Beechnuts and outbreaks of nephropathia epidemica (NE): of mast, mice and men. Nephrol Dial Transplant 25: 1740–1746. https://doi.org/10.1093/ndt/gfq122 PMID: 20237057
- 139. Heyman P, Cochez C, Ducoffre G, Mailles A, Zeller H, et al. (2007) Haemorrhagic fever with renal syndrome: an analysis of the outbreaks in Belgium, France, Germany, the Netherlands and Luxembourg in 2005. Euro Surveill 12: 15–16.
- 140. Hofmann J, Meisel H, Klempa B, Vesenbeckh SM, Beck R, et al. (2008) Hantavirus outbreak, Germany, 2007. Emerg Infect Dis 14: 850–852. https://doi.org/10.3201/eid1405.071533 PMID: 18439382
- 141. Reil D, Imholt C, Eccard JA, Jacob J (2015) Beech fructification and bank vole population dynamics-combined analyses of promoters of human Puumala virus infections in Germany. PLoS One 10: e0134124. https://doi.org/10.1371/journal.pone.0134124 PMID: 26214509
- 142. Yan L, Huang H, Zhang W, Wang J, Ren Y, et al. (2009) The relationship between hemorrhagic fever with renal syndrome cases and time series of NDVI in Dayangshu District. Journal of Remote Sensing 13: 873–886.
- 143. Fang L-Q, Goeijenbier M, Zuo S-Q, Wang L-P, Liang S, et al. (2015) The association between hantavirus infection and selenium deficiency in mainland China. Viruses 7: 333–351. https://doi.org/10.3390/v7010333 PMID: 25609306