

# Apoptotic and antimetastatic activities of betulin isolated from *Quercus incana* against non-small cell lung cancer cells

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**Background:** Globally, the prevalence and mortality rates of lung cancer have been escalated with the increasing trend of tobacco smoking. The toxicity and irresponsive nature of the available drugs for lung cancer treatment demands an alternative approach.

**Methods:** In this study, four known compounds namely, cirsimaritin (4',5, -dihydroxy-6,7-dimethoxyflavone) (1), eupatorin (5,3'-dihydroxy-6,7,4'-trimethoxyflavone) (2), betulin (Lup-20 (29)-ene-3, 28-diol) (3), and  $\beta$ -amyryn acetate (12-Oleanen-3yl acetate) (4) have been isolated from the leaves extract of *Quercus incana*. Preliminary screening of these natural compounds (1–4) was performed against non-small cell lung carcinoma (NCI-H460) and normal mouse fibroblast (NIH-3T3) cell lines.

**Results:** The compounds were found to be antiproliferative against cancer cells with wide therapeutic index in comparison to the normal cells. Effects of betulin (3) on cell migration, invasion, apoptosis, and expression of important apoptosis- and metastasis-related markers were observed at different concentrations. The results showed significant dose-dependent induction of apoptosis after the treatment with betulin (3) followed by increased expression of the caspases family (ie, caspase-3, -6, and -9), proapoptotic genes (*BAX* and *BAK*), and inhibiting anti-apoptotic genes (*BCL-2L1* and *p53*). Furthermore, wound healing and transwell invasion assays suggested that betulin (3) could also regulate metastasis by inhibiting MMP-2/-9. Osteopontin, a central regulator of apoptosis and metastasis was also inhibited in a dose-dependent manner.

**Conclusion:** The present findings suggest that betulin (3) can be an attractive chemotherapeutic target for treating resistant lung cancers.

**Keywords:** *Quercus incana*, betulin, cirsimaritin, eupatorin,  $\beta$ -amyryn acetate, non-small cell lung carcinoma, metastasis, MMP-2, MMP-9, apoptosis, caspases, osteopontin

## Introduction

Lung cancer, being the most prevalent cancer worldwide, has been ranked third among the top 10 malignancies in Pakistan.<sup>1</sup> In 2012, a report confirmed that 9.8% of men were diagnosed with lung cancer per 100,000 cases in Pakistan.<sup>2</sup> Excessive smoking habits in young- and middle-aged men accounted for the high prevalence of lung cancer.<sup>3</sup> Primarily, lung cancer is divided into two types: small cell lung carcinoma (SCLC) and non-small cell lung carcinoma (NSCLC). NSCLC represents 85% of the lung cancer cases occurring worldwide, whereas only 15% accounts for SCLC.<sup>4</sup> Current treatment regimen includes surgery coupled with the use of chemotherapeutic drugs. The commonly used chemotherapy drugs are methotrexate, bevacizumab, carboplatin, cisplatin, docetaxel, gefitinib, vinorelbine, and paclitaxel.<sup>5</sup> Despite the aforementioned

facts, the rate of survival was limited only to 65% because of poor prognosis. High recurrence rate, metastasis potential, and chemoresistance pose a great challenge in treating lung cancer and are also associated with high mortality rates.<sup>3</sup>

Metastasis and inhibition of apoptosis are responsible for cancer advancement, resistance against the drugs, and mortality.<sup>6</sup> Apoptosis is defined as programmed cell death that removes unwanted or mutated cells from the body. An equilibrium between proapoptotic and anti-apoptotic proteins is necessary for programmed cell apoptosis. Cysteine proteases (ie, caspases) and proteins of the BCL-2 family are the key regulators of this process.<sup>7,8</sup> Typically, a cell follows either intrinsic pathway or extrinsic pathway in caspases-mediated apoptosis. The intrinsic pathway is mediated inside the cell in response to DNA damage, hypoxia, and/or oxidative stress. It involves mitochondrial disassembly followed by the release of cytochrome c in the cytoplasm. This cytochrome c with other proapoptotic proteins forms apoptosome thereby activating a family of effector caspases.<sup>9</sup> Extrinsic pathway, however, involves death receptors with adaptor proteins which in turn activate caspases proteins.<sup>10</sup> Once executioner caspases (ie, caspases-3 and caspase-6) are activated, breakdown of cytoskeleton and DNA fragmentation occurs, leading to the formation of apoptotic bodies.<sup>11</sup> The later events are marked as late phase of apoptosis.<sup>12</sup>

Alternatively, metastasis is a complex process of tumor spread from the preliminary site to close or distant body parts. It involves invasion of tumor in the circulatory and lymphatic system followed by metastatic colonization in other organs. The family of matrix metalloproteinases (MMPs) plays important roles in the migration and invasion of in situ carcinoma.<sup>13</sup> MMPs are zinc proteases that belong to the matrixin family.<sup>14</sup> Under this category, gelatinases A and B are of prime importance because of their direct involvement in cancer progression. During tumorigenesis, MMP-2/-9 degrade the basement matrix through which tumor cells enter the blood and spread to other organs.<sup>15</sup> Considerably, lung cancer metastasizes to the adrenal gland, liver, brain, and bones.<sup>16</sup> With the evolution of resistant carcinoma and side effects of the present chemotherapeutic regimens, there is an immense need for new chemical entities with low side effects, high potency, and high selectivity toward molecular targets.

*Quercus incana* belongs to the genus *Quercus* (oak), the largest genus of family Fagaceae. Tropical and subtropical regions of Asia as well as temperate zones of Northern Hemisphere contain various species of genus *Quercus*.<sup>17</sup> The species of this genus are pharmacologically active and possess great medicinal value such as antioxidant, antifungal,

antibacterial, and/or anticancer activities.<sup>18,19</sup> This genus has great medicinal importance and was traditionally used in pharmacology for its astringent, hemostatic, and antiseptic properties.<sup>20</sup> The *Q. incana* has great medicinal uses; it may be used as astringent, diuretic, antidiarrheal and to treat asthma problems.<sup>21</sup> Bark and leaves of this plant are used as antipyretic, antirheumatic, and in wound healing. Acorns bark and leaves are best served for antidiabetic and anti-arthritic purposes.<sup>22</sup> Recently, triterpenes have been reported for their apoptotic and anti-cell migratory activities against breast cancer.<sup>23</sup> Thus, the present study was conducted to isolate and characterize some known flavonoids and triterpenes from the chloroform fraction of *Q. incana* leaves. Flavonoids and triterpenes have already been reported for their potent anticancer activity against various cancers.<sup>24,25</sup> Therefore, this study was designed to evaluate the anticancer potential of isolated compounds against NSCLC (NCI-H460) cell line. Furthermore, the molecular mechanism of the most potent apoptotic and antimetastatic compounds was demonstrated.

## Methods: experimental procedures

### Collection of plant material (leaves of *Q. incana*)

During April to June 2011 plant leaves were collected within Abbottabad region and identified taxonomically as *Q. incana* from the botany department at the Post Graduate College Abbottabad. The sample was deposited at the college herbarium as voucher specimen (#2550).

### Extraction and purification

The leaves of *Q. incana* were shade dried and ground to a coarse powder. The extraction and fractionation of *Q. incana* was described in our previous study.<sup>19</sup> The chloroform fraction was subjected to column chromatography to isolate the bioactive constituents.

### Cell culture

The NSCLC (NCI-H460) and normal mouse fibroblast (NIH-3T3) cell lines were grown and passaged as mentioned earlier by us using RPMI medium.<sup>46</sup> Both cell lines were commercially purchased by cell culture biobank (PCMD, ICCBS) from American Type Culture Collection (ATCC). The biobank provided the cell lines to our research group for experimental purpose.

### Cell viability assay

The efficacy of the isolated compound to inhibit metabolically active cells was determined by MTT assay. NCI-H460 cells at 10,000 cells/well density were seeded in a 96-well

plate for 24 hours followed by treatment at different concentrations (10, 25, 50, 75, and 100  $\mu\text{M}$ ) of the compounds. After 48 hours of treatment the reduction in viability of cells using MTT dye was evaluated as mentioned earlier.<sup>46</sup> Percent inhibition was calculated by using following equation:

$$\text{Cell proliferation inhibited (\%)} \\ = 100 - \left[ \frac{(\text{OD}_{\text{compound}} - \text{OD}_{\text{blank}})}{(\text{OD}_{\text{control}} - \text{OD}_{\text{blank}})} \times 100 \right]$$

By using software (EZ-Fit Enzyme Kinetics; Perrella Scientific Inc., Amherst, NH, USA)  $\text{IC}_{50}$  of the test compounds were calculated. Similarly, these compounds were tested for their cytotoxicity against normal NIH-3T3 cells.

### Cell migration assay

$1 \times 10^6$  NCI-H460 cells were seeded in each well of a 6-well plate and were grown to confluence. The next day a scratch wound was made using 200  $\mu\text{L}$  micropipette tip. After washing with PBS, betulin at varying concentrations was added and incubated for 48 hours. Images of the wounded area, taken immediately after the treatment, were considered as 0 hour, whereas further images were taken at 24 and 48 hours using a phase-contrast microscope at  $10\times$  magnification.<sup>47</sup> Images were processed and percent closure was calculated using image J software.

### Zymography

$5 \times 10^6$ /well NCI-H460 cells were seeded in a 6-well plate in serum free media with or without test compound. After 48 hours, media from each well was collected and centrifuged at 400 g for 5 minutes. 10  $\mu\text{L}$  of the supernatants were subjected to 10% (w/v) SDS-PAGE in the presence of 0.4% (w/v) gelatin. Following the electrophoresis, gel was thrice washed with 2.5% (v/v) Triton X-100 (Wako Chemicals, Osaka, Japan) for 20 minutes. After washing, the gel was first equilibrated in activation buffer (50 mM Tris HCl pH 7.5, 0.1 mM NaCl, 10 mM  $\text{CaCl}_2$  and 1 mM  $\text{ZnCl}_2$ ) for 20 minutes followed by overnight incubation in the fresh buffer at  $37^\circ\text{C}$ . The next day, gel was stained with 0.2% (w/v) Coomassie Brilliant Blue R-250 (Biobasic, Markham, ON, Canada) in 7.5% glacial acetic acid and 5% methanol, followed by destaining in glacial acetic acid:methanol:water (1:3:6) till gelatinolytic bands appeared. The gel was scanned and gelatinolytic bands were quantified using FluorChem software (Bio-Rad, Hercules, CA, USA).

### Transwell invasion assay

To evaluate the invasive potential of NCI-H460 cells, Transwell inserts (Millipore, Etobicoke, ON, Canada) coated with Geltrex (Life Technologies, Carlsbad, CA, USA) (9 mg/mL) were used. 40,000 cells with or without betulin were added

to the wells and incubated for 48 hours in  $\text{CO}_2$  incubator at  $37^\circ\text{C}$ . 500  $\mu\text{L}$  culture media was added outside the inserts as chemoattractant. Later, the matrix and media were carefully removed from the Transwell inserts using a cotton plug. The inserts were washed with PBS and invaded cells were fixed using 3.7% formaldehyde for 10 minutes at room temperature ( $T_R$ ). Inserts were again washed with PBS and stained with crystal violet for 20 minutes. Excess stain was removed by gentle PBS washing repeatedly. Inserts were then observed and photographed under inverted microscope ( $4\times$  magnification).

### YO-PRO-1™ apoptotic assay

NCI-H460 cells were seeded at a density of  $5 \times 10^5$  in a 6-well plate. After 24 hours, cells were treated with different concentrations of the test compound followed by 48 hours incubation at  $37^\circ\text{C}$  under a humidified 5%  $\text{CO}_2$  incubation. Following the incubation, cells were harvested using 0.05% trypsin and the cell pellet was washed twice with PBS at  $400\times$  g for 5 minutes to remove all the media and trypsin from the cells. After washing, cells were re-suspended in 1 mL PBS and stained using YO-PRO-1™ and PI as supplied by the Vybrant™ Apoptosis Assay Kit #4 (Thermo Fisher Scientific) following the manufacturer protocol. Briefly, cells were incubated with 1  $\mu\text{L}$  YO-PRO-1™ stock solution (component A) and 1  $\mu\text{L}$  PI stock solution (Component B) for 30 minutes on ice. Later, cells were analyzed on FACSCalibur™ (BD, Franklin Lakes, NJ, USA). YO-PRO-1™ and PI were excited at 488 nm, and fluorescence was measured at 530 and 620 nm, respectively. A total of 10,000 events were acquired from each sample. The percentages of live, apoptotic and dead cells were determined using CellquestPro software (BD biosciences, San Jose, CA, USA).

### DAPI staining

$2.2 \times 10^4$  NCI-H460 cells per well were seeded in a 24-well plate and placed overnight at  $37^\circ\text{C}$  in 5% humidified  $\text{CO}_2$  incubator. The next day, following the washing with PBS, cells were treated with betulin. After the incubation of 48 hours, the media was removed, cells were fixed with 3.7% formaldehyde in PBS for 10 minutes followed by PBS washing and DAPI (1:1000) staining for 20 minutes at  $37^\circ\text{C}$ . After staining, cells were again washed with PBS and observed for nuclear changes under Nikon fluorescence microscope at  $40\times$  magnification. Images of random fields were taken and were counted for apoptotic cells.

### JC-1 assay

The change in the mitochondrial membrane potential after the treatment with test compounds was analyzed by JC-1 (5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazolylcarbocyanine iodide) dye using manufacturer (Thermo Fisher Scientific)

protocol. Briefly,  $5 \times 10^5$  NCI-H460 cells were seeded in a 6-well plate. The next day, cells were treated with or without the test compound at different concentrations. After 48 hours of incubation, cells were washed with PBS and harvested using 0.05% trypsin. Later the cell pellet was washed twice with PBS before incubating it with JC-1 dye for 30 minutes at  $37^\circ\text{C}$  in 5%  $\text{CO}_2$  incubator. After incubation, cells were pellet down through centrifugation at  $400 \times g$  for 5 minutes to remove the dye and were resuspended in 500  $\mu\text{L}$  PBS followed by analysis on flow cytometry on FACSCalibur™ (Becton Dickinson). JC-1 was excited at 520 nm, whereas fluorescence was measured at 590 nm. 10,000 events of each sample were recorded and the percentages of cell population in each quadrant were analyzed using CellquestPro software.

## RT-PCR

RNA extraction of NCI-H460 cells after treatment was performed using TRIzol reagent (Thermo Fisher Scientific) following the standard protocol. Briefly,  $1 \times 10^6$  cells/well were seeded in a 6-well plate. The next day, cells were treated with the test compound for 48 hours in a 5%  $\text{CO}_2$  incubator at  $37^\circ\text{C}$ . After the incubation, total RNA of the cells was extracted and absorbance ratio 260/280 was determined using Nanodrop-ND-2000 spectrophotometer (Thermo Fisher Scientific). cDNA from 1  $\mu\text{g}$  of extracted RNA was synthesized using RevertAid First Strand cDNA Synthesis Kit (Thermo Fisher Scientific) according to the manufacturer instructions. Later, real-time PCR was performed in a 25  $\mu\text{L}$  reaction mixture using 1  $\mu\text{L}$  cDNA as a template and primers at given annealing temperatures (Table S1). 1% agarose (w/v) gel electrophoresis was performed to resolve the PCR products. Control of *GAPDH* was used as housekeeping gene.

## Immunocytochemistry

To analyze the effects of betulin (3) on various protein markers, 20,000 NCI-H460 cells were seeded in a 24-well plate with or without betulin. After 48 hours treatment, media was discarded and cells were carefully and thoroughly washed with PBS. Then cells were fixed with 4% paraformaldehyde for 15 minutes at room temperature. Again, wells were washed with PBS and 150  $\mu\text{L}$  Triton X-100 was added to the wells for 10 minutes. Cells were incubated with blocking solution for 30 minutes in a humidified environment followed by addition of primary antibody (1:100 dilution in blocking solution) overnight at  $4^\circ\text{C}$ . The next day, cells were washed with PBS and respective secondary antibody (Thermo Fisher Scientific) was added to the wells for 1

hour. Finally, DAPI staining was done followed by observing expression of markers under fluorescent microscope at  $10 \times$  magnification. The primary antibodies used against the markers include *BCL-2L1* (Santa Cruz Biotechnology Inc., Dallas, TX, USA), Ki-67 (EMD Millipore, Billerica, MA, USA), caspase-3 (EMD Millipore), caspase-6 (EMD Millipore), caspase-8 (EMD Millipore), and osteopontin (Abcam, Cambridge, MA, USA).

## Clonogenic assay

8,000 cells per well in a 6-well plate were seeded and treated with or without betulin the next day. After the treatment of 48 hours, cells were washed with PBS carefully and were allowed to grow in culture media for next 15 days in  $\text{CO}_2$  incubator at  $37^\circ\text{C}$ . The media was changed every third day to ensure the supply of optimal growth conditions to the cells. After incubation, cells were fixed with 3.7% formaldehyde and stained with 0.1% crystal violet. Excess stain was removed by repeated washing with PBS. The colonies of H460 cells were observed and photographed under inverted microscope ( $4 \times$  magnification).

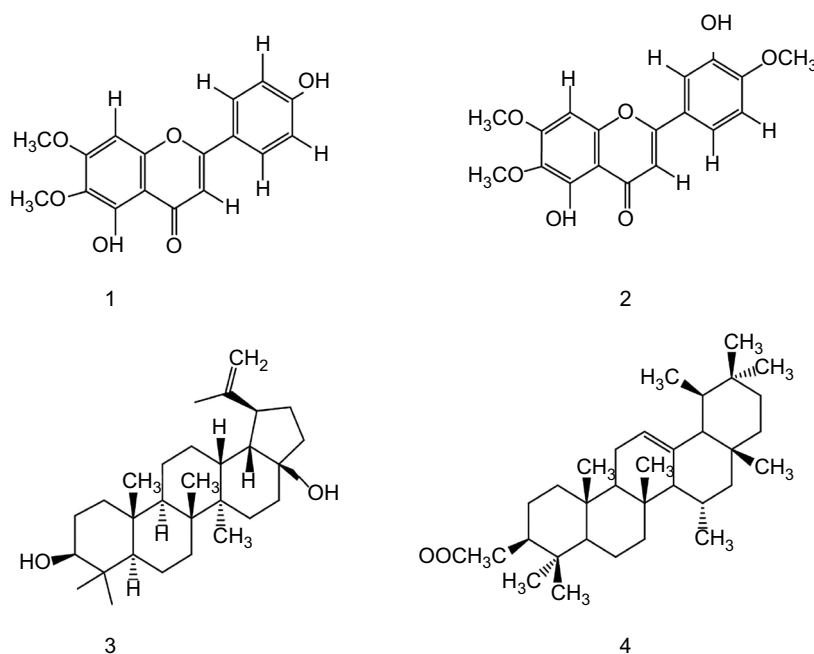
## Statistical analysis

Results of the all presented data are reported as means $\pm$ SD and level of significance were analyzed by Student's *t*-test. The values of  $P < 0.05$  between the control and treated groups were considered as significant.

## Results

### Isolation of compounds from *Q. incana*

The chloroform fraction of leaves of *Q. incana* was fractionated in solvent of increasing polarity (ie, *n*-hexane, *n*-hexane-chloroform, chloroform, chloroform-methanol, and methanol) by column chromatography using silica gel column. On the basis of thin-layer chromatography profile, the eight fractions were combined to get five fractions. Fraction 3 was further fractionated to two subfractions, ie, C-1 and C-2, when eluted with *n*-hexane-chloroform (increasing polarity) and rechromatographed over silica gel. Compounds 1 (34 mg) and 2 (64 mg) were purified from subfraction C-2 when loaded on fresh silica gel with mesh size 230–400  $\mu\text{m}$  and eluted with chloroform: *n*-hexane (9:1). Compound 3 (43 mg) was obtained from fraction 4, when eluted with methanol:chloroform (1:9). However, compound 4 (47 mg) was obtained after elution of fraction 2 in chloroform:*n*-hexane (7:3) (Figure 1). The physical and spectroscopic characterization of the compounds is provided in the Supplementary materials.



**Figure 1** Structure of compounds (1–4).

## Characterization of compounds

### Compound 1

Pale yellow crystal;  $C_{17}H_{14}O_6$ . IR (KBr)  $\nu_{\max}$  1650 (C=O), 1599 (C=C), 1356 (C–O), 1252 (C–O), 650 (unsaturated alkene); EI-MS  $m/z$ : (rel. int.) 315 [M+1]<sup>+</sup> (38), 299 (45), 271 (100), 153 (15), HR-EI-MS:  $m/z$  314.0790 (calculated 314.0896 for  $C_{17}H_{14}O_6$ ); <sup>1</sup>H-NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  3.74 (3H, s, –OCH<sub>3</sub>), 3.94 (3H, s, –OCH<sub>3</sub>), 7.94 (1H, d,  $J=8.8$  Hz, H-2',H-6'), 6.92 (1H, d,  $J=8.5$  Hz, H-3', H-5'), 10.41 (1H, 4'-OH), 12.95 (1H, 5-OH) 6.94, 6.82 (2×1, H, 2× s, H-8, H-3); <sup>13</sup>C-NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  182.0 (C-4), 164.0 (C-2), 161.4 (C -4'), 158.9 (C-7), 152.5 (C-9), 152.0 (C-5), 131.9 (C-6), 128.4 (C -6'), 128.4 (C-2'), 121.0 (C-1'), 115.9 (C-3'), 115.9 (C -5'), 105.1 (C-10), 102.5 (C-3), 91.0 (C-8), 60.0 (C -6-OCH<sub>3</sub>), 56.1 (C -7-OCH<sub>3</sub>). It was characterized as 4',5, -dihydroxy-6,7-dimethoxyflavone or cirsimaritin.<sup>26</sup>

### Compound 2

Yellow crystal;  $C_{18}H_{16}O_7$ . IR (KBr)  $\nu_{\max}$  3470 (OH), 1650 (C=O), 1599 (C=C), 1478 (R), 1365 (C–O). EI-MS  $m/z$ : (rel. int.) 344 [M]<sup>+</sup> (86), 329 (45), 326 (64), 301 (100).HR-EI-MS:  $m/z$  344.0896 (calculated 344.0930 for  $C_{18}H_{16}O_7$ ). <sup>1</sup>H-NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  3.89 (3H, s, 6-OCH<sub>3</sub>), 3.97 (3H, s, OCH<sub>3</sub>-4'), 3.88 (3H, s, 7-OCH<sub>3</sub>), 6.55 (1H, s, H-3), 6.52 (1H, s, H-8), 7.44 (1H,s, H-2'), 10.40 (1H,s, 4'-OH), 12.90 (1H,s, 5-OH), 7.38 (1H, dd,  $J=8.2, 2.1$ , H-6'), 6.91 (1H, d,  $J=8.8$  Hz,

H-5'); <sup>13</sup>C-NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  182.6 (C-4), 163.8 (C-2), 151.2 (C -4'), 158.7 (C-7), 152.1 (C-9), 156.0 (C-5), 132.0 (C-6), 119.1 (C -6'), 113.2 (C-2'), 123.7 (C-1'), 147.1 (C-3'), 111.7 (C -5'), 107.0 (C-10), 104.4 (C-3), 94.8 (C-8), 60.9 (C-6-OCH<sub>3</sub>), 55.9 (C-7-OCH<sub>3</sub>). It was characterized as 5,3'-dihydroxy-6,7,4'-trimethoxyflavone or eupatorin.<sup>27</sup>

### Compound 3

White amorphous powder;  $C_{30}H_{50}O_2$ . IR (KBr)  $\nu_{\max}$  3448 (OH), 3055 (=C–H), 1638 (C=C). EI-MS  $m/z$ : (rel. int.) 442 (35), 412 (26), 344 (12), 288(5), 257 (8), 248 (30), 189 (100). HR-EI-MS:  $m/z$  442.3811 (calculated 442.3843 for  $C_{30}H_{50}O_2$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta$  0.66 (1H, d,  $J=9$  Hz, H-5), 0.88 (3H, s, H-24), 0.87 (3H, s, H-25), 0.95 (3H, s, H-23), 0.74 (3H, s, H-27), 1.02 (3H, s, H-26), 1.59 (1H, s, H-19), 1.68 (3H, br, s, H-28), 4.67, 4.56 (1H, dd,  $J=4.0, 12.3$  Hz, H-30), 3.15 (1H, m, H-3), 1.19 (1H, dd,  $J=3.9, 12.3$  Hz, H-11).; <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 75MHz):  $\delta$  38.7 (C-1), 27.4 (C-2), 79.1 (C-3), 38.8 (C-4), 55.3 (C-5), 18.3 (C-6), 34.3 (C-7), 41.0 (C-8), 50.5 (C-9), 37.3 (C-10), 20.9 (C-11), 25.2 (C-12), 37.2 (C-13), 42.8 (C-14), 27.1 (C-15), 29.2 (C-16), 47.9 (C-17), 48.9 (C-18), 48.8 (C-19), 150.5 (C-20), 29.8 (C-21), 34.0 (C-22), 28.1 (C-23), 15.4 (C-24), 16.1 (C-25), 16.1 (C-26), 14.8 (C-27), 60.6 (C-28), 19.1 (C-29), 109.7 (C-30). It was characterized as Lup-20 (29)-ene-3, 28-diol or betulin.<sup>28</sup>

## Compound 4

White needles;  $C_{32}H_{52}O_2$ . IR (KBr)  $\nu_{max}$  1730 (C=O), 1640 (C=C). EI-MS  $m/z$ : (rel. int.) 468  $[M]^+$  (27), 453 (10), 408 (20), 218 (100), 203 (10). HR-EI-MS:  $m/z$  468.3951 (calculated 468.3968 for  $C_{32}H_{52}O_2$ ).  $^1H$ -NMR ( $CDCl_3$ , 300 MHz):  $\delta$  0.77 (3H, s, H-28), 0.88 (3H, s, H-23), 0.92 (3H, s, H-24), 0.86 (3H, s, H-29), 0.87 (3H, s, H-30), 0.99 (3H, s, H-26), 1.01 (3H, s, H-25), 1.04 (3H, s, H-27), 2.05 (3H, s, OAc), 4.51 (1H, dd,  $J=9.7$  Hz, H-3), 5.13 (1H, t,  $J=3.6$  Hz, H-12);  $^{13}C$ -NMR ( $CDCl_3$ , 75 MHz):  $\delta$  38.5 (C-1), 23.3 (C-2), 80.8 (C-3), 37.5 (C-4), 55.2 (C-5), 18.3 (C-6), 33.0 (C-7), 39.8 (C-8), 47.8 (C-9), 36.9 (C-10), 22.9 (C-11), 124.3 (C-12), 136.5 (C-13), 42.6 (C-14), 27.9 (C-15), 26.7 (C-16), 33.5 (C-17), 59.1 (C-18), 39.4 (C-19), 39.7 (C-20), 31.6 (C-21), 41.5 (C-22), 28.2 (C-23), 15.4 (C-24), 14.9 (C-25), 16.5 (C-26), 17.2 (C-27), 28.5 (C-28), 23.3 (C-29), 21.2 (C-30), 21.4 (C-OMe), 170.4 (C-O). It was characterized as 12-Oleanen-3yl acetate or  $\beta$ -amyrin acetate.<sup>29</sup>

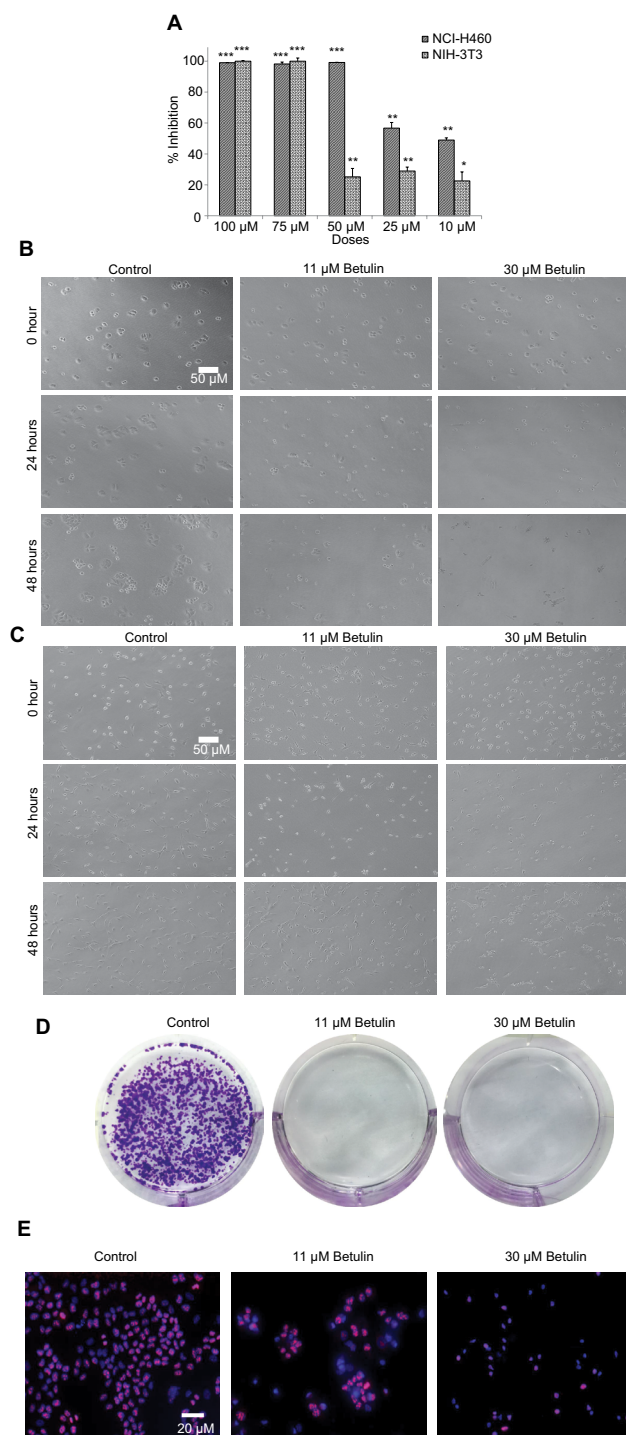
### Selective antiproliferative potential of isolated compounds (1–4) against NSCLC cells

The effect of the compounds isolated from *Q. incana* on the viability of NSCLC cells (NCI-H460) was evaluated by MTT assay. All the four compounds ie, cirsimaritin (4',5,-dihydroxy-6,7-dimethoxyflavone) (1), eupatorin (5, 3'-dihydroxy-6,7,4'-trimethoxyflavone) (2), betulin (Lup-20 (29)-ene-3, 28-diol) (3), and  $\beta$ -amyrin acetate (12-oleanen-3yl acetate) (4) were found to be antiproliferative against the cancer cells with low toxicity against normal fibroblast cells (NIH 3T3 cells; Table 1). Among these, betulin (3) was found to be four times more active against cancer cells compared to normal 3t3 cells and was also found to be more potent ( $IC_{50}$  11.5  $\mu$ M) compared to cisplatin (19  $\mu$ M; Figure 2A). Furthermore, microscopic analysis revealed that after 24 hours of treatment with betulin, H460 cells started to undergo consequential morphological changes subsuming membrane blebbing and formation of

**Table 1**  $IC_{50}$  values of the compounds from chloroform fraction of *Quercus incana* against NSCLC and normal cells

Name/compound no.	NCI-H460 ( $\mu$ M)	NIH-3T3 ( $\mu$ M)
Cirsimaritin (1)	26.23 $\pm$ 0.053	38.84 $\pm$ 0.037
Eupatorin (2)	37.50 $\pm$ 0.070	75.09 $\pm$ 0.086
Betulin (3)	11.55 $\pm$ 0.037	50.54 $\pm$ 0.049
$\beta$ -Amyrin acetate (4)	10.62 $\pm$ 0.045	20.06 $\pm$ 0.073
Cisplatin (standard)	19 $\pm$ 1.24	–

**Abbreviation:** NSCLC, nonsmall cell lung cancer.



**Figure 2** (A) Cytotoxic effects of betulin from *Quercus incana* against NSCLC and normal cells at various concentrations. \* $P < 0.05$  was considered as significant. \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ . Morphological changes in (B) NCI-H460 cells (C) NIH-3T3 cells after 24 and 48 hours of treatment with betulin, observed under phase contrast microscope at 10 $\times$  magnification. (D) Effect of betulin on colony formation of H460 cells after 48 hours treatment of betulin. Cells were supplied with fresh culture media every third day and were incubated at 37 $^{\circ}$ C in humidified 5%  $CO_2$  environment for 15 days after the treatment. (E) Immunocytochemical analysis of expression of proliferative marker Ki-67 after 48 hours of treatment with betulin, observed under fluorescent microscope at 20 $\times$  magnification.

**Abbreviation:** NSCLC, nonsmall cell lung cancer.

apoptotic bodies (Figure 2B). In contrast, 3T3 cells regained their normal morphology and started to propagate after 48 hours of treatment with betulin at 11 and 30  $\mu\text{M}$  concentrations (Figure 2C). The inhibition observed by betulin against NCI-H460 cells was dose and time dependent. In addition, betulin has completely eradicated cancer cells even after 15 days of incubation in culture media during colony formation assay compared to the control (Figure 2D). Immunocytochemical analysis further indicated the significant inhibition of the proliferative marker Ki-67 in the treated cells (Figure 2E).

Based on potent activity against cancer cells in comparison to 3T3 cells (Figure 2), betulin (3) was further evaluated for its apoptotic and antimetastatic potential against lung cancer cells.

### Betulin inhibits migration and invasion of lung carcinoma in vitro

Cell migration and invasion are the crucial steps in the process of metastasis.<sup>30</sup> The antimigratory effect of betulin (3) was established by using bidirectional wound healing assay. Cell migration was significantly inhibited toward the scratch compared with the untreated control at 24 and 48 hours treatment (Figure 3A) in a dose-dependent manner. After 48 hours, betulin treated cells showed 17% migration compared to the control, resulting in 41% wound healing (Figure 3B). Moreover, the invasive ability of NCI-H460 cells was inhibited in a dose-dependent manner as shown by transwell membrane invasion assay (Figure 3C).

It has already been shown that MMP-2 and MMP-9 play major roles in extracellular matrix degradation.<sup>31</sup> The expression of MMP-2/-9 was investigated using RT-PCR and zymography. This result positively correlated with earlier data, with the increasing concentration of betulin (3) significant decrease in the gene expression and enzymatic activity of MMP-2/-9 was observed (Figure 3D, E).

### Betulin induces cell death via mitochondrial-mediated apoptotic pathway

To evaluate whether cell death caused by betulin followed apoptotic dogma, various techniques were employed. Through fluorescence-activated cell sorting (FACS), it was analyzed that betulin treated H460 cells significantly underwent apoptosis in a dose-dependent manner compared to untreated control. It was observed that the majority of the treated cells lied in the late phase of apoptosis which elevated with increasing concentration (Figure 4A and B). As apoptosis is related with the disruption of mitochondrial membrane

potential (MMP,  $\Delta\psi\text{m}$ ), JC-1 staining was performed to investigate the effects of betulin on MMP of treated H460 cells. By principle, high red to green fluorescence ratio is associated with polarized (or functional) mitochondria or vice versa. Therefore, the results revealed collapse of MMP in H460 cells posttreatment of betulin for 48 hours (Figure 4C, D). Furthermore, in agreement with the FACS results, after 48 hours treatment, highly condensed nucleus, apoptotic bodies and DNA fragmentation were clearly observed in H460 cells through DAPI staining. On the contrary, the nucleus of control cells remained intact (Figure 4E).

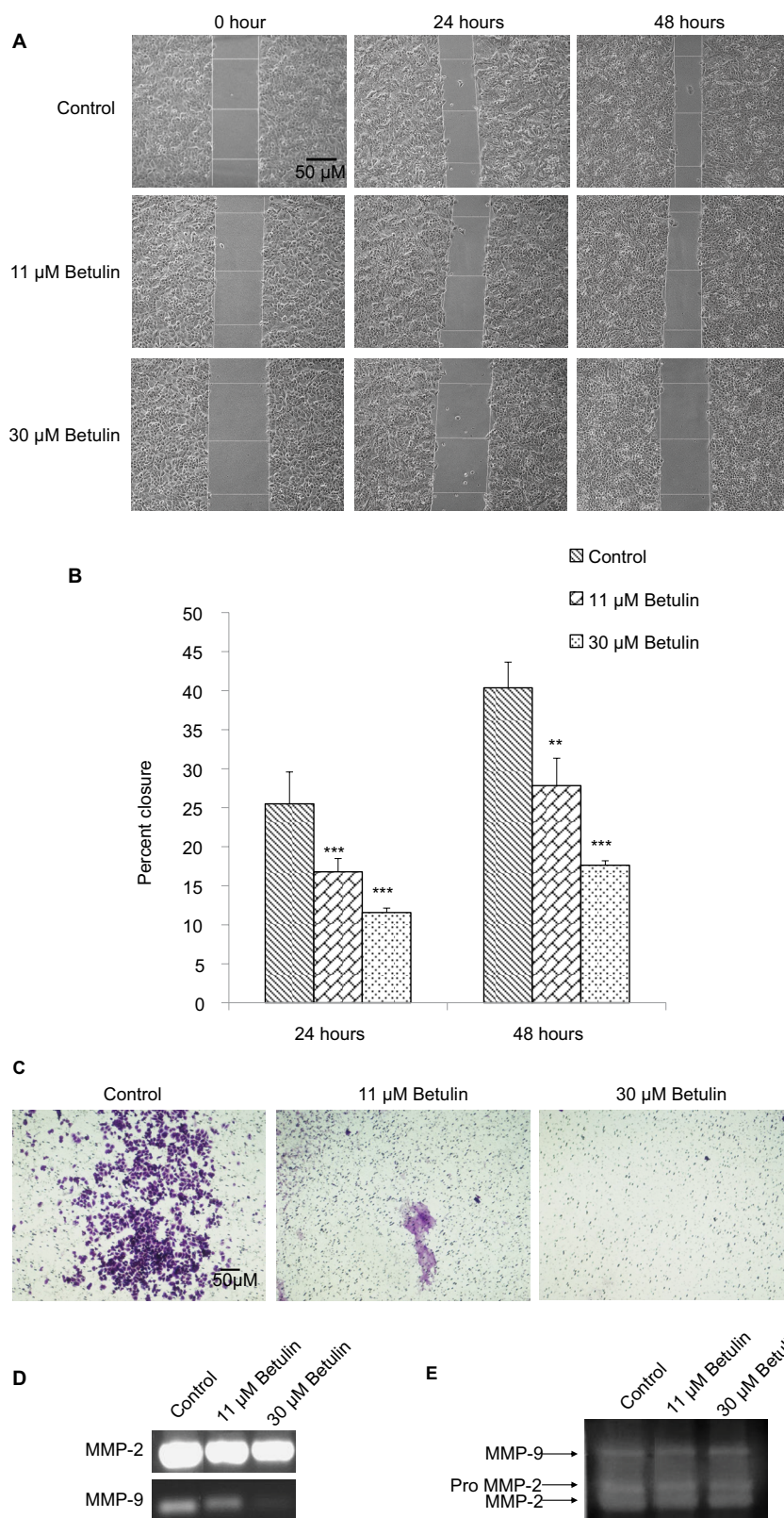
Gene expression analysis of markers responsible for apoptosis was also carried out to elucidate further the molecular mechanism underlying the apoptosis (Figure 5A, B). The results showed dose-dependent upregulation and downregulation of genes involved in apoptosis. The expression of *p53* and *BCL-2L1* genes were significantly downregulated, and are responsible for inhibiting apoptosis. However, the compound upregulated the expression of *BAX* and *BAK* genes responsible for the apoptosis. It also significantly upregulated the expression of caspases-3, -6, and -9; inducers of mitochondrial-mediated apoptosis cascade. However, the expression levels of caspases-6 and -9 were significantly higher in treated cells compared to caspase-3. Furthermore, immunofluorescence results confirmed significant upregulation of caspases-3 and -6 in treated cells compared to control cells (Figure 5C). Interestingly, substantial expression of caspase-8 was observed in control H460 cells, which decreased with the increase in betulin treatment. This supports the findings that betulin mediates mitochondrial-dependent apoptosis in H460 cells. In addition, the gene expressions of *VEGF* and *COX-2*, the most prominent cell proliferative and metastatic markers, were also found to be inhibited after 48 hours of treatment (Figure S1).

### Betulin induces antineoplastic effects via osteopontin regulation

Osteopontin, a phosphorylated glycoprotein, has been widely implicated for cancer progression and metastasis.<sup>32</sup> Therefore, effects of betulin on the expression of osteopontin were determined. The results indicated that betulin positively inhibited osteopontin levels analyzed by RT-PCR and immunocytochemistry (Figure 6). A significant downregulation was observed in a dose-dependent manner (Figure S1).

### Betulin sensitizes cisplatin in inhibiting H460 cells

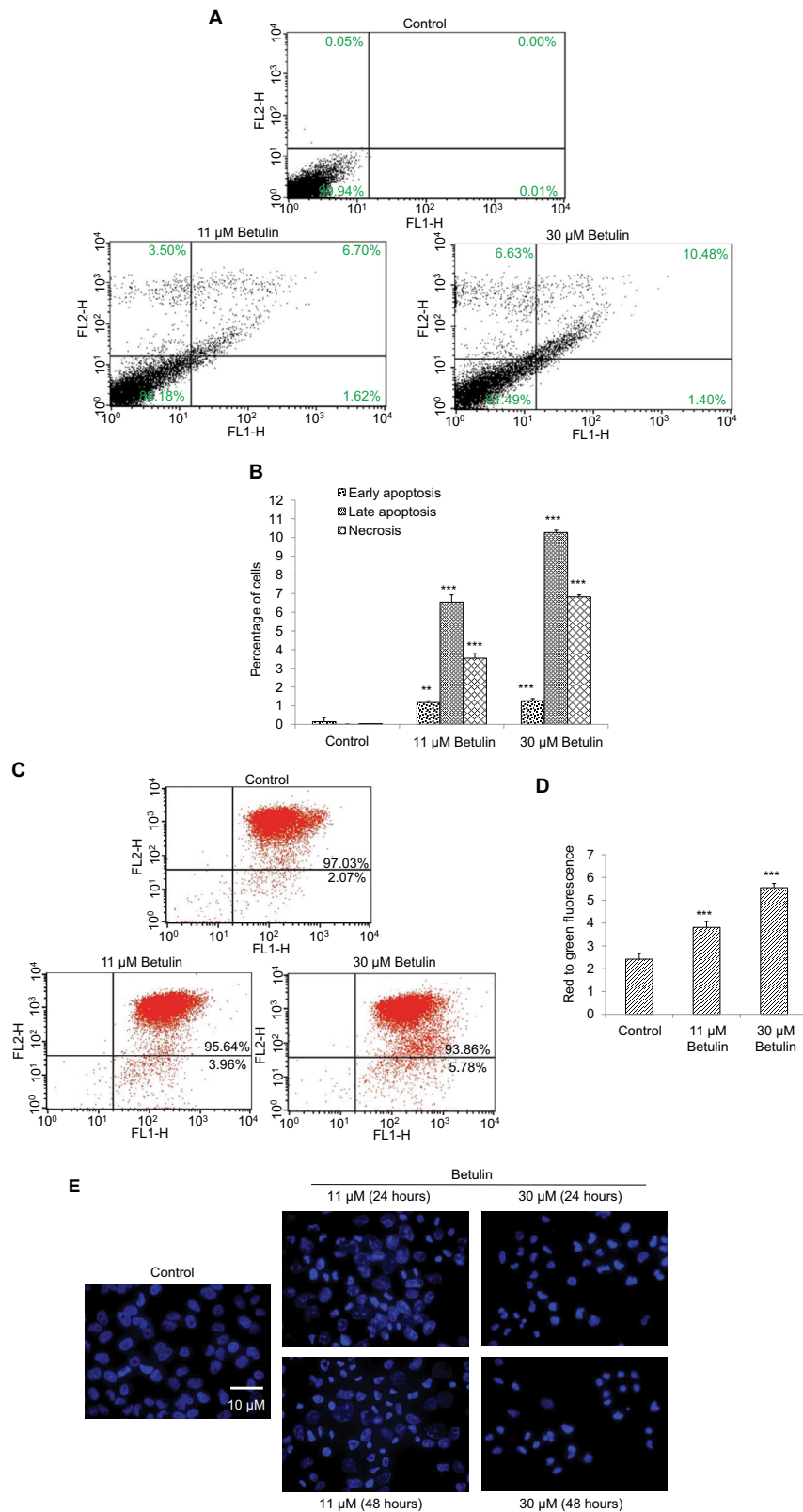
Based on “constant-ratio model”, the combination effects of betulin and cisplatin were studied using MTT assay. After



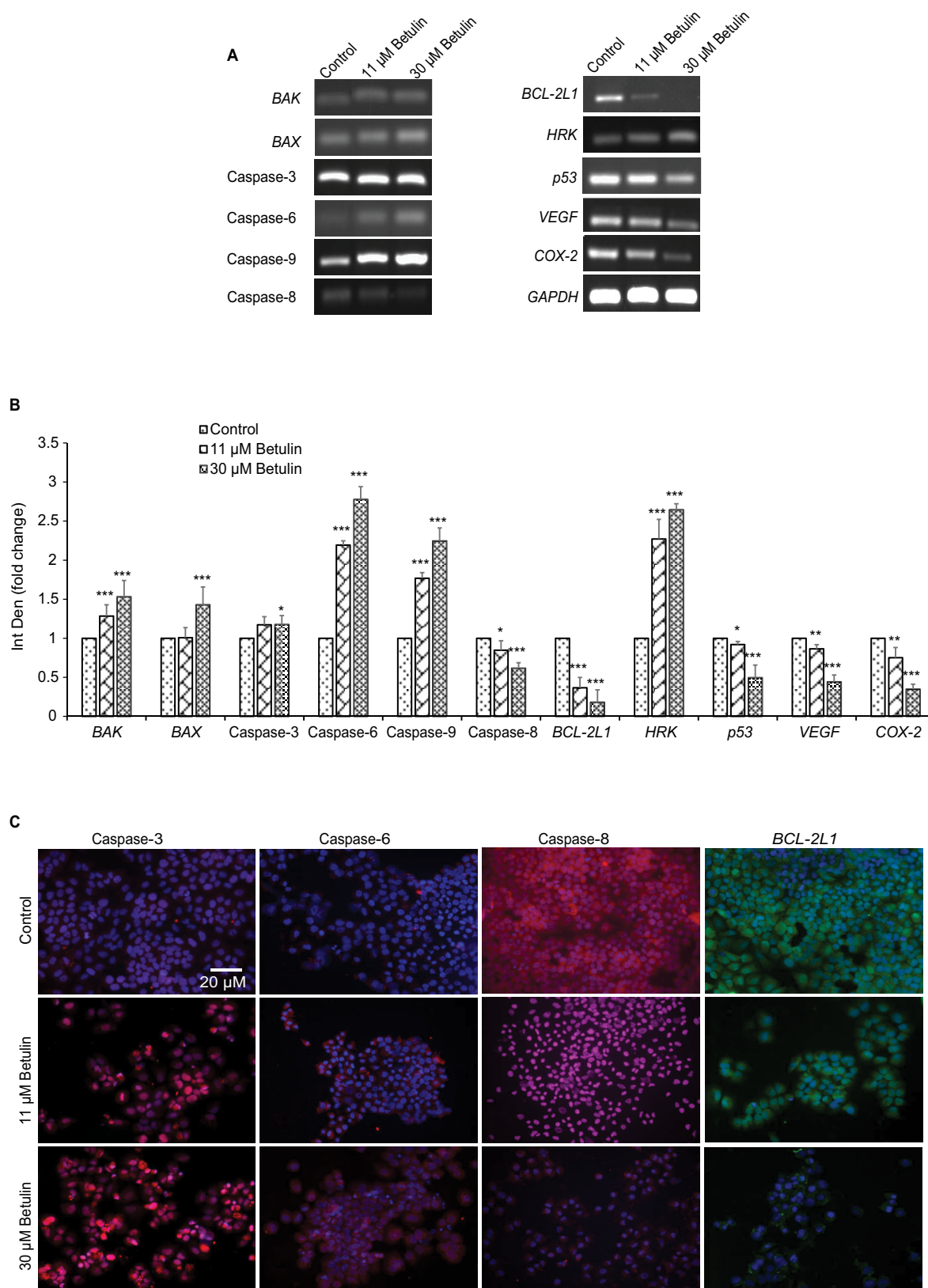
**Figure 3** (A) Effects of betulin on the migration of NCI-H460 cells. Using phase contrast microscope at 10× magnification, scratch areas were photographed and area of the wound was calculated using Image J. (B) Percent closure of the wound after 24 and 48 hours of the treatment. In control well, the rate of migration of the cells toward the wound was increased to 41% after 48 hours in treated groups cells migration was significantly reduced. \*\* $P < 0.01$  vs control. \* $P < 0.05$ ; \*\*\* $P < 0.001$ . (C) Effects of betulin on invasion of NCI-H460 cells using transwell assay. Membrane was photographed under bright field at 4× magnification using inverted microscope. (D) Effects of betulin on expression levels of MMP-2 and -9 in NCI-H460 cells after 48 hours treatment as evaluated by RT-PCR. (E) Gelatinolytic activity of MMP-2/-9 in response to the treatment with different concentrations of betulin as analyzed by zymography.

**Abbreviation:** MMP, matrix metalloproteinases.



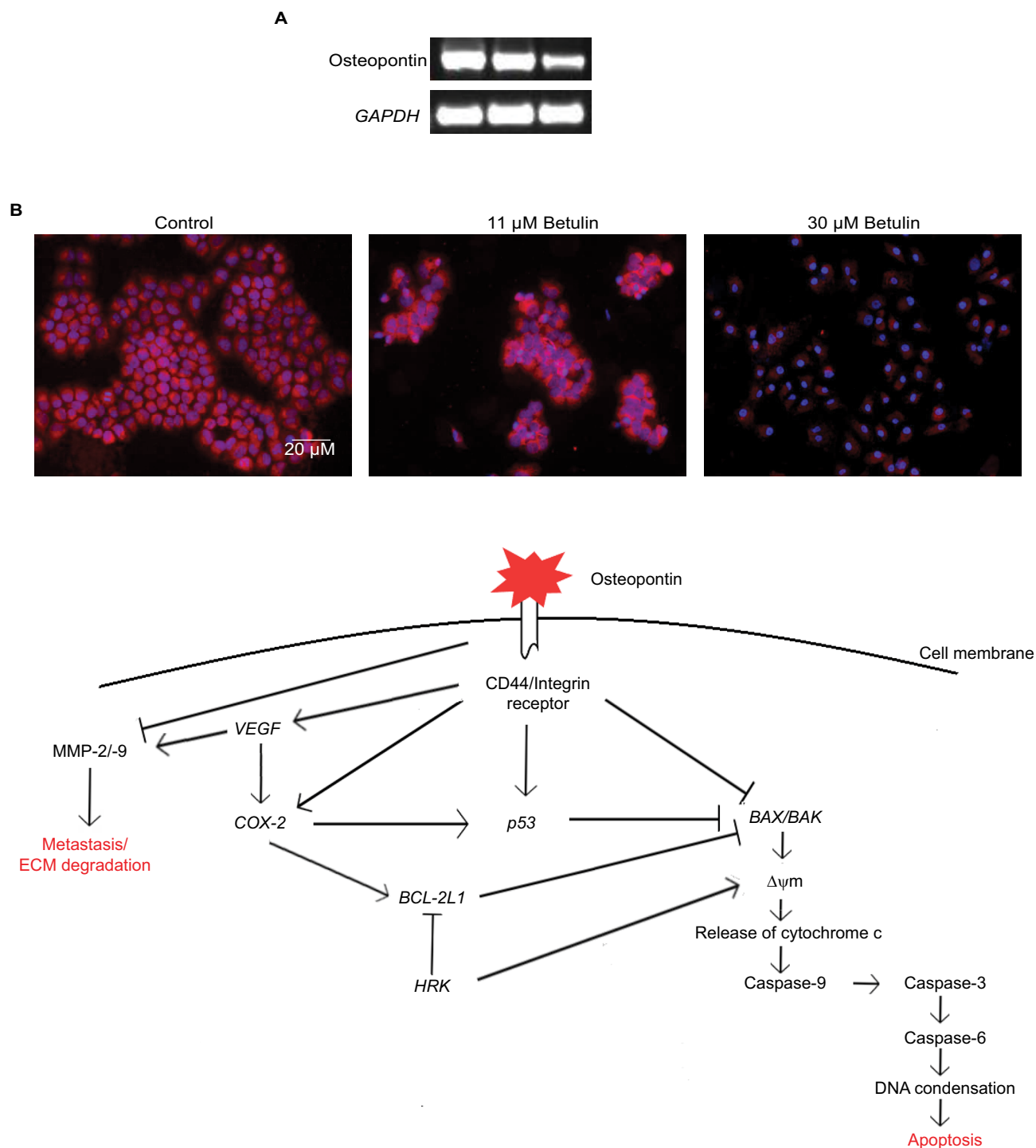


**Figure 4** (A) Betulin (3) induced apoptosis in NSCLC cell line in a dose-dependent manner. Flow cytometric analysis of the apoptosis induced by the indicated concentrations of the compound. An increased number of cells were observed in treated groups compared to control. (B) Graphical representation of the cells in different phases of apoptosis after 48 hours of treatment.  $**P < 0.01$  compared to the untreated control.  $***P < 0.001$ . (C) Effects of betulin (3) on the mitochondrial membrane potential ( $\Delta\psi_m$ ) of the cells after 48 hours. Following the treatment with different concentrations, an increase in positive green fluorescent cells was observed, indicating loss of  $\Delta\psi_m$  compared to the control. (D) Histogram shows the percentages of the cells with disrupted  $\Delta\psi_m$  in treated groups.  $***P < 0.001$  vs the untreated control. (E) Effects of betulin (3) on the nuclear morphology of NCI-H460 cells. Increase in DNA condensation and change in the shape of the nucleus of the treated groups have been observed in a dose and time-dependent manner compared to the control. **Abbreviation:** NSCLC, nonsmall cell lung cancer.



**Figure 5** (A) Analysis of effects of the compounds on the mRNA expression of cell survival and apoptotic genes. An upregulation was observed in genes of caspases family and proapoptotic genes ie, *HRK*, *BAX* and *BAK*, while anti-apoptotic *BCL-2L1* and *p53* genes and cell survival *VEGF* and *COX-2* genes significantly downregulated. Interestingly, initiator caspase-8 of death receptor pathway is also inhibited after the treatment with betulin. *GAPDH* was used as reference gene. (B) Quantitative analysis of expression of apoptotic genes compared to control. \* $P < 0.05$  compared to control. \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ . (C) Immunocytochemical analysis of major apoptotic markers after 48 hours treatment with betulin observed under fluorescent microscope at 20 $\times$  magnification. The caspase-3 and caspase-6 are highly expressed in treated cells while caspase-8 is downregulated, suggesting activation of intrinsic pathway of apoptosis. Furthermore, anti-apoptotic *BCL-2L1* show marked inhibition posttreatment.

**Abbreviation:** Int Den, integrated density.

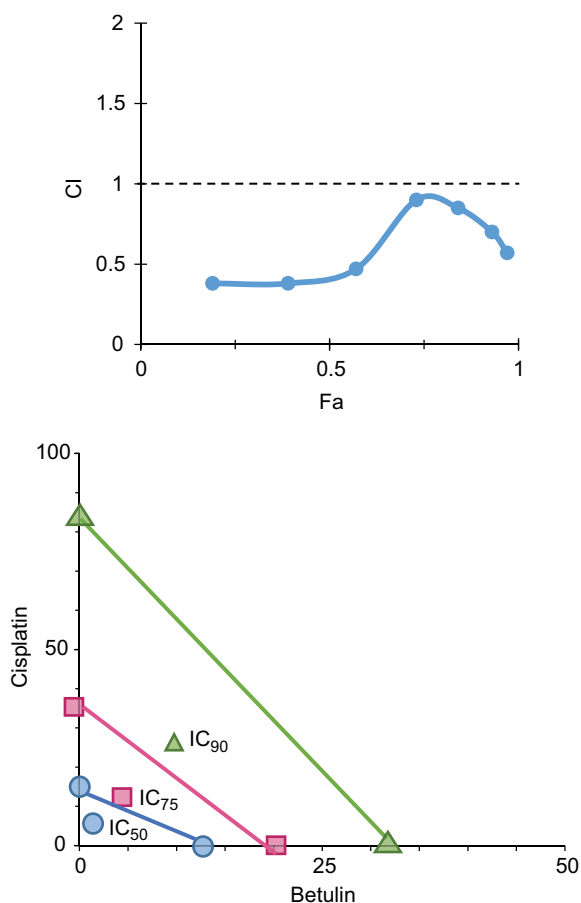


**Figure 6 (A)** The gene expression analysis of osteopontin in response to the treatment with betulin after 48 hours as observed via RT-PCR. The results show pronounced downregulation of expression of the marker in comparison to *GAPDH*, the reference gene. **(B)** Proteomic analysis of osteopontin compared with the untreated control studied through immunofluorescence assay under fluorescent microscope at 20 $\times$  magnification. The expression of osteopontin in the cells after the treatment. **(C)** Proposed mechanism of osteopontin as central regulator of apoptosis and metastasis in H460 cells.

**Abbreviation:** MMP, matrix metalloproteinases.

48 hours of treatment, the data were obtained and analyzed using the Chou–Talalay method. The combination index (CI) of 1/2th, 1/4th, 1/8th, and 1/16th  $\text{IC}_{50}$  showed strong sensitizes with fraction affected (Fa) values in the range 0.4–0.8 (data not shown). In Figure 7, Fa–CI plot indicates CI at a

particular effect produced by the drugs while isobologram represents that the combination doses of betulin and cisplatin were much lower than actual data points indicating  $\text{IC}_{50}$ ,  $\text{IC}_{75}$ , and  $\text{IC}_{90}$  of individual drugs. As indicated, 11.5  $\mu\text{M}$  betulin and 19  $\mu\text{M}$  cisplatin induced 50% inhibition of H460 cells,



**Figure 7** Synergistic effects of betulin with standard drug cisplatin as represented by (A) Fa-CI plot and (B) classic isobologram showing data points of individual drugs vs actual data points of their combination at different inhibitory concentrations observed through MTT assay.

**Abbreviations:** Fa, fraction affected; CI, combination index.

whereas the combination of betulin with cisplatin elicited a similar response at less than 5  $\mu\text{M}$  concentration.

## Discussion

Globally, the mortality and morbidity rate of lung carcinoma is increasing despite of the advancement in the therapeutic strategies. Poor prognosis, resistance against available drugs and inefficacious treatment modalities necessitate the development of new chemotherapy drugs. Triterpenoids, a class of organic compounds, are well known for their multifunctional properties, including antimicrobial and anticancer activities against various cancer types.<sup>33</sup> In the present study, among the four triterpenes and flavonoids, isolated from the leaves of *Q. incana*, betulin (3) possessed the most potent antiproliferative, apoptotic and antimetastatic activities. Earlier studies report lower cytotoxic effects of betulin (3), isolated from other plant sources, against different cancer cell lines.<sup>34-36</sup> In contrast, betulin (3) isolated from *Q. incana* was found highly

and selectively active against the cancer cells at much lower doses (11.55  $\mu\text{M}$ ), even less than standard drug cisplatin (19  $\mu\text{M}$ ). The compound is also noncytotoxic to normal NIH-3T3 cells at higher doses ie 50  $\mu\text{M}$  (Table 1). Therefore, it was subjected to further investigation at IC<sub>50</sub> and IC<sub>80</sub> concentrations (ie, 11  $\mu\text{M}$  and 30  $\mu\text{M}$ ) for in-depth analysis of its apoptotic and antimetastatic potential.

Mutations often result in the uncontrolled cell growth that leads to tumor formation. In order to overcome tumor progression and problems related to chemoresistance, induction of apoptosis and inhibition of metastasis to other organs are important factors.<sup>11</sup> Betulin (3) induced significant apoptosis in H460 cells after 48 hours treatment (Figure 4). Interestingly, apoptosis assay revealed that the majority of the cell population after treatment falls in the late apoptotic phases. These results further complement the findings of DAPI staining, in which the cancer cells, after 48 hours, showed events of late phase apoptosis ie, shrunken nuclei and loss of normal morphology compared to the nuclei of control cells (Figure 4). Among two apoptotic pathways, betulin (3) initiates the mitochondrial pathway as evidenced by altered expression of apoptotic markers, which regulates intrinsic pathway (Figure 5). RT-PCR and immunofluorescence assay revealed, betulin upregulated caspase-9 expression, while caspase-8 was significantly downregulated. Caspase-8 is considered as a caspases initiator in death receptor mediated apoptosis.<sup>9</sup> Therefore, the data suggests that betulin activates intrinsic pathway machinery thereby shutting down proteins of extrinsic pathway in order to induce apoptosis in H460 cells. Furthermore, betulin (3) shows potential antimetastatic activity by inhibiting the wound healing and invasion of cells through transwell inserts compared to the untreated controls. There was an approximate threefold reduction in the wound healing rates of treated cells compared to the untreated cells. Significant inhibition of the expression and enzymatic activity of MMP-2/-9 in a dose-dependent manner was also observed in post betulin treatment (Figure 3).

In current era, various treatments and their modalities are being continuously introduced to minimize the cancer-related morbidity and mortality. Early diagnosis is an important factor that could help in early removal of cancer hence increasing the survival capabilities.<sup>37</sup> Biomarkers are the main players of early and easy detection of any abnormality being progressed in the body. Recently, osteopontin has received much attention because of its central functionality in tumor progression in various cancers.<sup>38,39</sup> High levels of serum osteopontin is correlated with tumor advancement.<sup>40,41</sup> This study demonstrates potential inhibition of the gene and

protein expressions of osteopontin after treatment with betulin (3) at various concentrations. As reported, osteopontin has central roles in interlinking many regulatory and physiological pathways, regulating important markers of apoptosis, metastasis, angiogenesis, bone remodeling pathways downstream of it.<sup>42-44</sup> Herein we describe its interplaying role between metastatic and apoptotic pathways of H460 cells (Figure 6). Expression profile indicated that betulin has potential inhibitory effects on the markers of cell survival and proliferation, ie, *VEGF*, *COX-2*, and *p53* proteins and metastatic MMP-2/-9.<sup>43</sup> Osteopontin functions as positive regulator for these proteins, mediating major processes of metastasis and tumor progression leading to cancer advancement. Moreover, betulin (3) further activates the crucial apoptotic markers, ie, *BAK/BAX*, *HRK* and caspases family proteins, while inhibiting *BCL-2L1*.<sup>44</sup> Reports suggest that osteopontin shuts off these proteins and indirectly activates anti-apoptotic *BCL-2L1* via *COX-2* pathway to inhibit the intrinsic pathway of apoptosis.<sup>45</sup>

In conclusion, we report the apoptotic and antimetastatic activities of betulin (3), isolated from new plant source, ie, leaves of *Q. incana*. In addition, the compound followed the cascade of the intrinsic pathway for inducing apoptosis in cancer cells and also inhibited important markers of metastasis. Moreover, strong synergism at lower doses of betulin and cisplatin was also observed after 48 hours (Figure 7). Therefore, the findings suggest that betulin (3) can be an attractive chemotherapeutic target for treating resistant lung cancers.

## Acknowledgments

The authors are grateful to the International Center for Chemical and Biological Sciences, Hussain Ebrahim Jamal Research Institute of Chemistry, University of Karachi, Higher Education Commission (HEC) Pakistan, and COMSATS Institute of Information Technology, Pakistan, for providing financial support for this work.

## Author contributions

BZ and AA are both to be considered as first authors. AK and SAA conceived and designed the study. RS and UF performed isolation. BZ and AA performed anticancer activities. SAA also performed gel experiment. AAH and AK analyzed the data. AK and AA wrote the manuscript with input and comments from all co-authors. All authors contributed toward data analysis, drafting and critically revising the paper, gave final approval of the version to be published, and agreed to be accountable for all aspects of the work.

## Disclosure

The authors report no conflicts of interest in this work.

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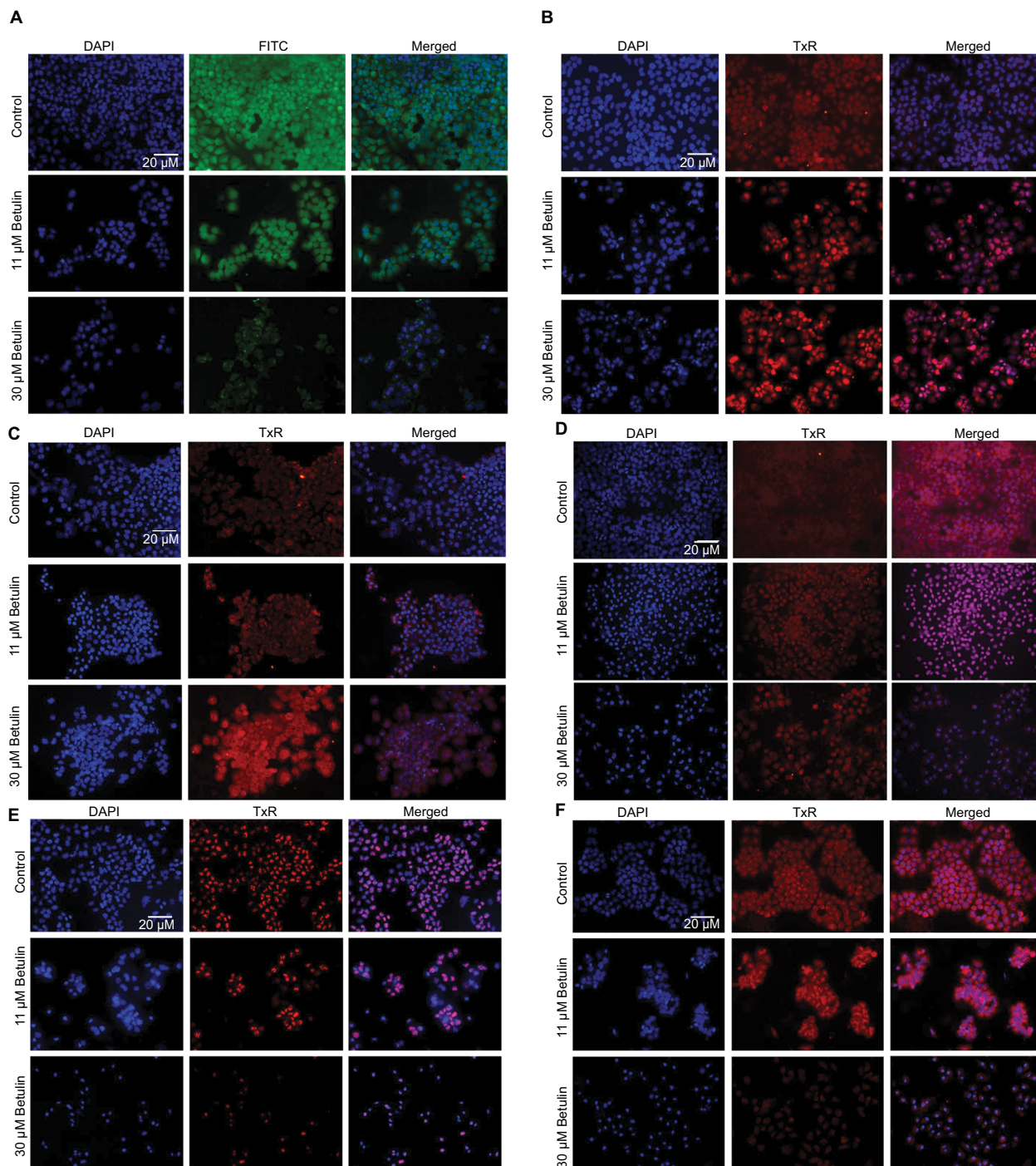
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## Supplementary materials

**Table S1** Primer sequences of the genes used for RT-PCR.

Gene	Sequence	Annealing temperature	References
<b>Caspase-3</b>	R: CAAGCTTGTCGGCATACTGTTTCAG F: TTCAGAGGGGATCGTTGTAGAAGTC	52°C	Hanif et al, <sup>1</sup> 2014
<b>Caspase-6</b>	R: AATTGCACTTGGGTCTTTGC F: ATCCTCACCGGAAACTGTG	54°C	Alharbi, <sup>2</sup> 2015
<b>Caspase-8</b>	R: GGATGAGGCAGACTTTCTGC F: GGCCTCCATCTATGACCTAG	58°C	This study
<b>Caspase-9</b>	R: GCACCACTGGGGTAAGGTTTTCTAG F: ATGGACGAAGCGATCGGCGGCTCC	60°C	Krisnamurti et al, <sup>3</sup> 2016
<b>BAX</b>	R: GGCCCCAGTTGAAGTTGC F: AAGAAGCTGAGCGAGTGTC	54°C	Hanif et al, <sup>1</sup> 2014
<b>BAK</b>	R: CCTGAGAGTCCAACCTGCAA F: GGTCCTGCTCAACTCTACCC	60°C	Guan et al, <sup>4</sup> 2010
<b>BCL-2L1</b>	R: ATGGTCAGTGTCTGGTCATT F: TTGTGGAACCTATGGGAAC	57°C	Choi et al, <sup>5</sup> 2015
<b>p53</b>	R: CTCTCGGAACATCTCGAAGCG F: GCTCTGACTGTACCACCATCC	57°C	Suo et al, <sup>6</sup> 2015
<b>HRK</b>	R: CCCCCRAAAATTAATAAAAAAAAAACT F: GAGTTGAATTTAGGAAAAGGGGAA	58°C	Zaker et al, <sup>7</sup> 2016
<b>Osteopontin</b>	R: ACGGCTGTCCCAATCAGAAG F: AGCAGAATCTCCTAGCCCCA	58°C	This study
<b>VEGF</b>	R: GCTTTCGTTTTTGCCCTTTC F: CAACATCACCATGCAGATTATGC	57°C	Wang et al, <sup>8</sup> 2015
<b>COX-2</b>	R: GCATCTGGCCGAGGCTTTTCTAC F: TGTGGGCAGGAGGTCTTTGGTCT	57°C	Kim et al, <sup>9</sup> 2011
<b>MMP-2</b>	R: GGCATCCAGGTTATCGGGGA F: GGCCCTGCTACTCCTGAGAT	59°C	Benus et al, <sup>10</sup> 2005
<b>MMP-9</b>	R: GCACAGTAGTGCCGTAGAA F: CTATGGTCCTCGCCCTGAAC	54°C	This study
<b>GAPDH</b>	R: GGTCTACATGGCAACTGTGA F: ACGACCACTTTGTCAAGCTC	59°C	Shi et al, <sup>11</sup> 2012

**Abbreviation:** MMP, matrix metalloproteinases.



**Figure S1** Effects on protein expression of (A) *BCL-2L1*, (B) caspase-3, (C) caspase-6, (D) caspase-8, (E) Ki-67, and (F) osteopontin after 48 h treatment with betulin at different concentrations.

**Notes:** DAPI staining indicates nuclei of cells, whereas FITC/TxR image shows expression of the respective marker under fluorescent microscope at 20× magnification.

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