



HHS Public Access

Author manuscript

Nat Rev Gastroenterol Hepatol. Author manuscript; available in PMC 2019 September 01.

Published in final edited form as:

Nat Rev Gastroenterol Hepatol. 2018 September ; 15(9): 525–535. doi:10.1038/s41575-018-0022-9.

The role of diet in the etiopathogenesis of inflammatory bowel disease

Hamed Khalili^{1,2}, Simon S. M. Chan^{3,4}, Paul Lochhead¹, Ashwin N. Ananthakrishnan^{1,2}, Andrew R. Hart^{3,4}, and Andrew T. Chan^{*,1,2,5,6}

¹Division of Gastroenterology, Massachusetts General Hospital and Harvard Medical School.

²Clinical and Translational Epidemiology Unit, Massachusetts General Hospital and Harvard Medical School, Boston MA, USA.

³Department of Medicine, Norwich Medical School, University of East Anglia, Norwich, United Kingdom.

⁴Department of Gastroenterology, Norfolk and Norwich University Hospital NHS Trust, Norwich, United Kingdom.

⁵Channing Division of Network Medicine, Harvard Medical School, Boston MA, USA.

⁶Broad Institute, Cambridge MA, USA.

Abstract

Crohn's disease and ulcerative colitis, collectively known as inflammatory bowel disease (IBD), are chronic inflammatory disorders of the gastrointestinal tract. Although the etiopathogenesis of IBD is largely unknown, it is widely thought that diet has a crucial role in the development and progression of IBD. Epidemiological and genetic association studies have identified a few promising dietary and genetic risk factors. These preliminary studies have prompted significant interest in investigating the complex interaction between diet, host genetics, the gut microbiota and immune function in the pathogenesis of IBD. In this Review, we discuss the recent epidemiological, gene–environment interaction, microbiome, and animal studies that have explored the relationship between diet and the risk of IBD. Additionally, we will highlight the limitations of these prior studies, in part explaining their contradictory findings, and review future directions.

Table of Contents Blurb

* achan@mgh.harvard.edu.

Author contributions

Competing interests

The authors declare no competing interests.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

ONLINE ONLY-

Subject Categories

Health sciences / Gastroenterology / Gastrointestinal diseases / Inflammatory bowel disease Health sciences / Gastroenterology / Gastrointestinal system / Microbiota Health sciences / Health care / Public health / Epidemiology

Display items

Emerging data implicate diet as a crucial environmental factor in the development and progression of inflammatory bowel disease (IBD). Here, the authors discuss findings from epidemiological, animal, gene–environment interaction and gut microbiome studies that have investigated the role of diet in the etiopathogenesis of IBD.

Introduction

Crohn’s disease and ulcerative colitis, collectively known as inflammatory bowel disease (IBD), are chronic inflammatory disorders of the gastrointestinal tract that have an increasing incidence¹² and affect over one million individuals in the US and 2.5 million in Europe^{13, 14}, resulting in substantial morbidity¹⁵, health care expenses and loss of productivity^{16, 17}. Although the pathophysiology of IBD is largely unknown, it is thought to be related to an inappropriate immune response to commensal bacteria in genetically susceptible hosts (FIG. 1). A role for genetic risk factors in the development of IBD has been highlighted by the identification of >200 susceptibility loci in genome-wide association studies (GWAS)^{18, 19}. In addition to the genetic contribution to IBD pathophysiology, environmental factors also seem to substantially contribute to both the development and progression of IBD. For instance, twin studies have demonstrated a <50% concordance in the development of Crohn’s disease and ulcerative colitis^{20, 21}. In addition, epidemiological studies have confirmed the rapid increase in the incidence of IBD in the US and developing countries that have witnessed a dramatic ‘westernization’ of lifestyle with the worldwide prevalence of the disease surpassing 0.3% at the turn of 21st century.^{12, 22} Moreover, studies have shown that the incidence of IBD in immigrants to developed countries exceeds that of individuals from their country of origin^{23–26}.

Among environmental factors, diet is widely thought to have a pivotal role in the development of IBD. Although the exact pathophysiological mechanisms remain unknown, a number of plausible explanations have been proposed (FIG. 2) Firstly, diet has a key role in defining the composition of the human gut microbiota and, consequently, that of microbial metabolites²⁷. Secondly, food and nutrients associated with a Western diet, characterized by high intakes of red meat, sugary desserts, high-fat foods, and refined grains have been linked to increased mucosal inflammation as measured by stool calprotectin in human subjects²⁸. Lastly, animal studies have demonstrated that dietary composition regulates mucosal barrier function, a crucial factor in the pathogenesis of IBD^{29, 30}. Despite these data, human observational studies investigating the role of diet in IBD have yielded contradictory and inconclusive results.

In this Review, we will discuss the available data on the role of diet in the pathogenesis of IBD, with a focus on epidemiological,, gene–environment interaction, intervention, gut microbiome, and animal studies. We will also highlight limitation of prior studies, particularly epidemiological and gene-environment studies and review future directions that build on these preliminary studies.

Intervention studies

Recently, there has been a resurgence of interest in the potential of dietary intervention for the treatment of IBD. Although a comprehensive discussion of prior dietary intervention studies is beyond the scope of this Review and has been previously published³¹, the early success of these studies further supports a role for diet in the pathogenesis of IBD. For example, several studies have demonstrated that exclusive enteral nutrition (EEN), which involves the administration of a liquid diet formula for a defined period of time, improves both clinical symptoms and intestinal inflammation in patients with Crohn's disease; one randomized controlled trial demonstrated that a short-term EEN was superior to corticosteroids in promoting mucosal healing in paediatric patients with Crohn's disease^{32, 33}. Nevertheless, similar studies in adults, where compliance to such a restrictive diet might be challenging, have not been as promising³³. Lastly, a number of uncontrolled human intervention studies have also reported promising results for other diets in the treatment of IBD including specific carbohydrate diet (SCD), low-fermentable oligosaccharide, disaccharide, monosaccharide, and polyol (FODMAP) diet and Paleolithic diet³⁴. Although large, well-designed, randomized studies in humans are needed to fully examine the therapeutic effects of these diets in IBD. These early results suggest a role for diet in IBD pathogenesis and support the need for further investigation in this area.

Epidemiological studies

A limited number of prior studies have examined the association between specific dietary factors and risk of IBD^{35–37}. Unfortunately, many of these studies have a number of drawbacks. Specifically, most prior studies were retrospective case–control analyses, and are therefore subject to numerous limitations including recall and selection biases. Recall bias of diet before the onset of IBD symptoms is particularly problematic. For example, if individuals are recruited after an extended period following diagnosis, they may have significantly changed their diet or preferentially recall consumption of specific foods that exacerbate their symptoms. Similarly, recall of diet within a very brief interval post-diagnosis is subject to concerns about reverse causation, whereby symptoms that immediately precede the formal diagnosis might alter dietary intake. In addition, the majority of previous studies collected dietary data at a single time point and were therefore unable to account for variations in dietary intake over time. There have also been very few studies that have rigorously validated dietary assessments within their study populations. Finally, because of the relatively low absolute incidence of IBD in the general population, most prior cohort studies have been limited by sample size. Taken together, these limitations might contribute to the inconsistent associations between dietary factors and the risk of IBD that is generally seen in the scientific literature.

To address these limitations, over the past 10 years, a number of large prospective cohort studies have attempted to better characterize the link between diet and risk of IBD by leveraging validated and updated dietary data collected during long-term follow-up (TABLE 1). The majority of these studies used the semiquantitative food frequency questionnaire (SFFQ) to estimate long-term dietary intake. The SFFQ is a method of assessing food and nutrient intake over a specified period of time by obtaining data on the frequency — and in some cases portion sizes — of food and beverage consumption³⁸. The SFFQ has several

advantages over other forms of dietary assessment (such as 24 hour recall or dietary records) in that it focuses on patterns of dietary intake over longer periods of time and therefore avoids biases that could be introduced through the episodic or occasional consumption of specific foods and beverages. In addition, SFFQs are relatively easy to complete by participants, therefore providing an ideal method to conduct large-scale epidemiological studies of diet in chronic diseases. Lastly, the SFFQ has been extensively validated against diet records (Box 1), allowing for the use of statistical methods to account for measurement errors. By contrast, the SFFQ is limited by its use in specific populations with well-characterized patterns of dietary consumption (i.e. Western societies) and is subject to over-reporting of consumption of healthy foods as a consequence of social desirability bias.

Dietary studies in the Nurses' Health Study.—The Nurses' Health Study (NHS) is a prospective cohort study that began in 1976 when 121,701 female registered nurses aged 30–55 years completed a mailed health questionnaire³⁹. Similar baseline questionnaires were returned in 1989 from a parallel cohort of 116,686 female nurses aged 25–42 years that were enrolled in NHSII. Questionnaires have been mailed to members of both cohorts every two years, starting in 1976 in NHS and 1989 in NHSII, to obtain updated information on diet, lifestyle and medical diagnoses, with a follow-up rate of >90% until 2012 for the NHS and until 2013 for the NHSII. Dietary data is collected every four years in these cohorts through a 161-item SFFQ. This SFFQ has been extensively validated in several studies, including observations that : nutrient intake assessed by the SFFQ strongly correlated with detailed dietary records ($r=0.65$)^{1–3}; specific items between 2 questionnaires administered 1 year apart were strongly correlated ($r=0.67$)^{1–4}; nutrients calculated from the SFFQ correlated with biochemical indicators of dietary intake, including plasma levels of beta-carotene ($r=0.30–0.42$) and vitamin E ($r=0.41–0.53$), as well as red blood cell levels of folate ($r=0.55$)^{5–8}; estimates of fatty acid intake correlated with adipose tissue levels of linoleic acid ($r=0.35–0.37$), trans-fatty acid ($r=0.51$) and n-3 fatty acids ($r=0.48–0.49$)^{9, 10}. Repeated validation studies of the FFQ in the NHS have yielded similar and stable correlations with diet records over time. In a validation study published in 2017, which was conducted in 632 women from the NHS cohort between 2010 and 2012, the FFQ was correlated with two 7-day dietary records that were administered one year apart ($r=0.62$) and automated self-administered 24-hour recalls ($r=0.63$) over a one year period, suggesting that the SFFQ provides reasonably valid estimates of intakes of a wide variety of dietary variables¹¹. These correlations probably underestimate the validity of the SFFQ because they also incorporate the errors related to use of the 6-day dietary record for estimating long-term intake.

Early work from the NHS and NHSII showed a low risk of Crohn's disease among individuals who consume high amounts of fiber, particularly fiber from vegetables and fruits (Table 1)⁴⁰. By contrast, a high n-3:n-6 fatty acids ratio was associated with a low risk of ulcerative colitis, whereas a high intake of red meat was associated with an increased risk of ulcerative colitis (Table 1)^{41, 42}. Owing to the increasingly recognized role of specific nutrients and electrolytes such as heme, potassium and zinc in regulating the intestinal barrier and immune function^{43–50}, over the past 5 years studies have focused on examining the relationship between quantity of nutrient consumption and risk of IBD. Preliminary data from these cohorts have shown an inverse association between dietary zinc and potassium

intake and risk of Crohn's disease, whereas dietary sodium, heme and iron intake were not associated with IBD risk (Table 1)^{42, 51, 52}.

Dietary studies in the European Prospective Investigation in Cancer study.—

The European Prospective Investigation in Cancer (EPIC) study, which investigated the link between dietary intake and the risk of common malignant diseases in the general population, is comprised of 519,978 men and women recruited from 23 centers in 10 European countries⁵³. The EPIC-IBD study is a subcohort of the EPIC study that involved a total of 401,326 initially healthy men and women aged 20–80 years who did not have Crohn's disease or ulcerative colitis and who were recruited from 12 centers in 8 European countries between 1991 and 1998. EPIC-IBD participants provided information on age, gender and lifestyle factors (such as smoking, physical activity levels and dietary information) through the self-completion of questionnaires at baseline. The follow-up rate until 2010 in EPIC-IBD was >98%. Dietary assessment in EPIC-IBD was conducted at baseline when participants reported dietary intake using country-specific validated questionnaires⁵³. In most centers, a self-administered FFQ was used to assess dietary intake (with 88–266 food-related questions) over the previous 12 months. Interviewer- administered diet questionnaires were used in Greece, Spain and in Ragusa, Italy. In the EPIC study, almost 10% of the cohort (~37,000 individuals) were assessed using a single highly-standardized, 24-hour recall of actual food consumption to account for any between-cohort differences that might exist in systematic exposure measurement error as well as for within-cohort attenuation biases in relative risk estimates. The energy-adjusted intakes were used to estimate correlations between FFQ and 24-hour recall and the mean intake levels estimated from 24-hour recall were used to account for within-cohort and between-cohort variations, respectively⁵⁴.

Prior to the establishment of EPIC-IBD cohort, one study from EPIC that consisted of 260,686 participants from 10 centers across 5 countries, investigated the associations between intake of a total of 18 nutrients, vitamins and minerals and the risk of ulcerative colitis⁵⁵. No associations were detected, with the exception of a weak positive association between increased total polyunsaturated fatty acid (PUFA) intake as a percentage of total energy intake and increased risk of developing ulcerative colitis. A similar broad analysis has yet to be published for Crohn's disease in this particular cohort. However, following these initial results, a subsequent study from the EPIC-IBD study further investigated the associations between intake of PUFAs and ulcerative colitis⁵⁶. On the basis of findings by other groups that n-6 PUFAs are metabolized to pro-inflammatory lipid mediators^{57, 58}, this study hypothesized that individuals with the highest dietary intakes of n-6 PUFAs would have an increased risk of developing ulcerative colitis. Indeed, the highest quintile for dietary intakes of the n-6 PUFA linoleic acid, as measured by the FFQ, was associated with an increased risk of developing ulcerative colitis (OR 2.49, 95% CI 1.23–5.07)⁵⁶ (Table 1). Conversely, the same study found that participants with the highest quintile for dietary intakes of docosahexaenoic acid (DHA), an n-3 PUFA that is metabolized to anti-inflammatory lipid mediators, had a decreased risk of developing ulcerative colitis (OR 0.32, 95% CI 0.06–0.97). Although similar findings for DHA intake and risk of Crohn's disease

have been reported in the EPIC-IBD study⁵⁹, no associations between n-6 PUFA intake and Crohn's disease have been found in EPIC-IBD to date⁵⁹ (Table 1).

In contrast, investigations into the role of monosaccharide, disaccharide and starch intake and the risk of developing ulcerative colitis or Crohn's disease in development of Crohn's disease and ulcerative colitis using EPIC-IBD have not demonstrated any consistent associations⁶⁰ (Table 1). Interestingly, however, EPIC-IBD study participants in the highest quintile for diets rich in sugar and soft drinks had an increased risk of developing ulcerative colitis (IRR 1.68, 95% CI 1.00–2.82)⁶¹ (Table 1). This finding was mainly attributed to those individuals with a dietary pattern of high intake of sugar and soft drinks who had intakes of vegetables, legumes and fruit that were below that of the median population (OR 2.80, 95 % CI 1.54–5.09)⁶¹. High consumption of milk has been shown to be associated with decreased risk of developing Crohn's disease but not ulcerative colitis, independent of dietary calcium intake⁶². However, the association did not show a dose-response relationship and there was no association between total intake of dairy products, yogurt or cheese with either ulcerative colitis or Crohn's disease. Finally, investigations into alcohol intake and the risk of developing ulcerative colitis or Crohn's disease in EPIC-IBD have not reported any statistically significant associations⁶³, despite speculation that isoflavones in alcoholic beverages accumulated during fermentation process⁶⁴ might have antioxidant and immune modulatory effects based on preclinical studies⁶⁵.

Limitations of prospective cohort studies and future directions.—Despite offering unique opportunities to study dietary factors in relation to IBD incidence, both the EPIC and NHS studies are limited by the under-representation of individuals with an early onset of IBD (age < 40 years). This drawback limits the generalizability of the findings of these studies, particularly regarding individuals who were diagnosed with IBD before the age of 40 years. Although dietary data derived from these cohorts has been extensively validated, the correlation between FFQ and dietary records, considered to be the gold standard, is modest for many foods and nutrients, which in turn can be a source of considerable measurement errors. In addition, despite being the largest cohort studies to date that have examined the link between diet and risk of IBD, the number of incident cases of IBD in these studies is limited, precluding the identification of more modest associations.

Thus, there continues to be an unmet need for a comprehensive, prospective examination of association between diet and risk of IBD. Such an effort could perhaps be achieved through the establishment of a large international consortium and could be used to achieve a number of important aims. Firstly, to improve our understanding of the associations between dietary patterns and risk of IBD across cohort studies comprised of different populations with a wide distribution of participant characteristics. Secondly, to examine the presence of dose-response relationships across a wide exposure range. Thirdly, to generate summary estimates that have greater precision than the individual studies due to the larger sample size, therefore allowing examination of the population attributable risk of dietary factors in IBD. Finally, such a collaborative effort could allow for examinations of whether associations differ within specific population subgroups. These analyses are particularly important as they might identify subgroups of the population that might benefit the most from specific lifestyle interventions. In addition, given that IBD is a heterogeneous disease, the evaluation of

whether associations vary by disease subtype might lead to a better understanding of the aetiology of IBD.

Animal studies

Dietary factors are known to induce insulin resistance, influence the balance of fatty acids that elicit or reduce inflammation of the gut epithelium, modify intestinal permeability and contribute to sulfur production which in turn alter the gut microbiota by promoting the growth of sulfate-reducing bacteria^{66, 67}. Animal studies have shown that a Western diet, increases the susceptibility of mice to dextran sodium sulfate (DSS)-induced colitis and increases the infiltration of macrophages compared with a Mediterranean diet, characterized by a high intake of fruits, vegetables, whole-grain foods, poultry and fish⁶⁸. In addition, a Western diet in mice alters host intestinal barrier function by increasing the colonization of adherent-invasive *Escherichia coli*²⁹. Similarly in mice, a high-fat diet was shown to disrupt the gut barrier function by modifying the composition of luminal bile salts³⁰.

Although dietary fiber has been shown to exert its anti-inflammatory effect by modulating the composition of the gut microbiota, increasing colonic fermentation and promoting the production of short-chain fatty acids, studies in mice models of colitis have also shown that fiber might have a direct immunomodulatory effect through inhibition of pro-inflammatory cytokine production in peripheral blood mononuclear cells⁶⁹. Additionally, in a model of *Citrobacter rodentium*-induced acute colitis, a fiber-rich diet reduced inflammation by modifying the host microbiota to increase production of short-chain fatty acids, and by altering host immune mechanisms of bacterial recognition and response to promote host health. In contrast to fiber, dietary red meat exacerbates DSS-induced colitis in mice⁷⁰. Although the exact mechanism underlying this observation is unclear, one potential explanation is that animal proteins increase luminal levels of sphingosine-1-phosphate, a sphingolipid involved in cell signaling, which has been shown to increase inflammation in mice models of colitis^{71–73}.

Over the past 5 years, there has been a growing interest on the role of dietary sodium in regulating the immune response. Several animal studies have shown a role for dietary sodium intake in the development and progression of autoimmune disorders through IL-23–IL-23R-mediated activation of pathogenic pro-inflammatory type 17 T helper (T_H17) cells, which have a critical role in development of IBD^{45–47}. In addition to enhancing the induction of pro-inflammatory T_H17 cells, sodium also appears to suppress immune tolerance through inhibiting the immunosuppressive functions of forkhead box protein P3 positive (FOXP3)⁺ T regulatory (T_{reg}) cells⁷⁴. Sodium might also lead to activation of pro-inflammatory macrophages⁷⁵. In contrast to sodium, a study published in 2016 demonstrated the profound suppressive effect of potassium, released into the extracellular fluid as a result of necrosis in human tumors, on T cell effector function⁴⁴. In turn, the study also demonstrated that augmentation of potassium efflux in tumor-specific T cells enhances their effector function and promotes tumor clearance. Together, these findings demonstrate the critical role of sodium and potassium in regulating the T_H17/T_{reg} balance.

Gene–environment studies

Studies investigating the interaction between environmental components and genetic variants in functionally-annotated genes have been increasingly used to help infer causal associations⁷⁶ and provide insight into potential biological pathways through which an environmental factor such as diet might contribute to the etiopathogenesis of chronic diseases, including IBD. Genetic loci associated with IBD risk can be broadly categorized into those involving abnormalities in the innate and/or adaptive immune response and mucosal barrier function⁷⁷. Experimental data also suggest that many of these pathways are probably influenced by dietary factors⁷⁸. Thus, it is biologically plausible that specific dietary components have differential effects on the incidence of IBD according to the genetic background.

Data on the influence of gene–environment interaction on IBD risk is sparse. This paucity is largely due to the limited availability of well-characterized cohorts with detailed, updated and validated data on dietary and lifestyle factors with simultaneous annotation of genetic variants. In addition, due to the low incidence of IBD in the general population, many cohort studies have a limited number of participants, precluding the possibility of assessing gene–environment interactions. Nevertheless, the investigation of gene–environment interaction is an area of great unmet need as results from such studies will not only shed light on the complex interaction between diet, genetic susceptibility and immune function but could also help to inform dietary recommendations for individuals at a high risk of IBD or those with established disease.

Preliminary analyses of gene–environment interaction in studies incorporating dietary factors in IBD have been promising^{51, 79–81} (TABLE 2). An analysis from the NHS and NHSII, which included 149 individuals with Crohn’s disease and 172 individuals with ulcerative colitis matched to 650 control participants, examined the interaction between total dietary intake of iron and heme iron and genetic variants associated with risk of IBD⁴². This study demonstrated an interaction between iron and heme iron intake and the ulcerative colitis susceptibility locus rs1801274, a coding variant in the *FCGR2A* (encoding low affinity immunoglobulin γ Fc region receptor II-a) gene. Specifically, among women with the GG genotype, increasing heme iron intake was associated with a substantially lower risk of UC (OR 0.11, 95% CI 0.03–0.37 for each 1 g increase in heme iron intake) (TABLE 2). In contrast, among women with the TT genotype, increasing heme iron intake was associated with an almost 3-fold higher risk of UC (OR 2.76, 95% CI 1.02–7.48). Due to the crucial role of *FCGR2A* in regulating humoral response to infection⁸² and the known importance of the rs1801274 variant in altering the binding capacity of the encoded protein product to C-reactive protein (CRP) and immunoglobulin G2 (IgG2)^{83–85}, this data provides evidence for an intriguing interaction between dietary heme intake and immune function in the development of ulcerative colitis.

Similarly, in a nested case–control study of 202 individuals with ulcerative colitis and 169 individuals with Crohn’s disease in the NHS and NHSII cohorts matched to 740 control participants based on age, menopausal status, month of blood collection and fasting status, an interaction between dietary potassium intake and genetic variants in the IL-23 pathway that have been previously associated with risk of IBD in GWAS was identified⁷⁷.

Specifically, the rs7657746 variant of *IL21* (encoding IL-21) seemed to modify the association between potassium intake and risk of IBD (Table 2). Each 200 mg increase in dietary potassium intake was inversely associated with risk of ulcerative colitis (OR 0.90, 95% CI 0.83–0.98) among participants with the CC genotype, but not among those with the AG or GG genotypes. Similar findings were observed in patients with Crohn's disease in this study. As IL-21 has a key role in the development of T_H17 cells through signal transducer and activator of transcription 3 (STAT3), a transcription factor required for the differentiation of T_H17 cells *in vivo*, the findings from gene–environment interaction studies suggest a potential mechanism for the observed association⁸⁶. IL-21 and IL-23 induce expression of the orphan nuclear receptor ROR γ , which, in synergy with STAT3, promotes IL-17 expression in CD4⁺ T cells, leading to the activation of T_H17 cells⁸⁶. In addition, **IL-21 inhibits the TGF- β -dependent generation of FOXP3⁺ T_{reg} cells, induces Th17 cell activation**⁸⁷. Interestingly, in this study, the gene–environment interaction finding was further supported by *in vitro* studies demonstrating that potassium induces FOXP3 expression in naïve and memory T cells and pro-inflammatory T_H17 cells. This effect was present even in the presence of pro-inflammatory cytokine suggesting that potassium may suppress inflammation in a pro-inflammatory milieu.

Lastly, data from the NHS and NHSII have also demonstrated that two variants in *CYP4F3*, which encodes a cytochrome P450 enzyme (CYP4F3) involved in PUFA metabolism, might modify the association between n-3 and n-6 PUFA intake and risk of ulcerative colitis⁸⁸ (Table 2). Specifically, the association between n-3:n-6 PUFA intake ratio and UC was modified by the rs4646904 SNP in *CYP4F3* ($P_{\text{interaction}}=0.049$). A high (median) n-3:n-6 intake ratio was associated with a lower risk of UC among women with the GG or AG genotypes (OR 0.57, 95% CI 0.32–0.99), but not among those with the AA genotype (OR 0.95, 95% CI 0.47–1.93). Indeed, similar findings were previously reported in a case–control study of children with newly diagnosed Crohn's disease⁸¹, suggesting that the interaction appears to be robust.

Limitations of prior gene–environment studies and future directions.—Despite the early promising results from gene–environment studies, it is important to acknowledge the limitations of these data. First, the majority of these studies had a limited sample size and were therefore not adequately powered to broadly assess (that is, on a genome-wide level) the presence of gene–environment interactions. Secondly, despite the presence of stringent *P* value thresholds, in these preliminary studies, as demonstrated in previous validation studies, even robust preliminary findings may still represent spurious associations further emphasizing the need for validation studies in gene–environment studies^{89, 90}. Thirdly, due to the limited sample size and the adjustment of most of these studies for multiple variables, these preliminary studies might have been prone to type II error. Lastly, although gene–environment interaction analyses might elucidate potential biological mechanisms underpinning the link between diet and IBD, statistical interaction does not imply biological modification; thus, further mechanistic studies are needed to fully characterize the results of these studies. Despite these limitations, preliminary findings highlight the need for larger scale analyses of the interaction between diet and genetics on

risk of IBD, reinforcing the need for large international consortia composed of well-characterized cohort studies with simultaneous assessment of genetic and dietary data.

Gut microbiota studies

Diet is widely linked to the composition of the human gut microbiome and microbial metabolites⁹¹. Although much of the microbial diversity seen in the adult gut might be attained by the age of 4 years, the adult microbiome remains susceptible to the influence of diet⁹². Indeed, dietary patterns have been proposed to explain greater than half of the variation in the adult intestinal microbiome⁹². Accordingly, the gut microbiome has a crucial role in the pathogenesis of IBD, as evidenced by observations that infusion of intestinal luminal contents into excluded ileum triggers post-operative recurrence in patients with Crohn's disease⁹³. Furthermore, in mice models of colitis, microbial colonization is required for the development of active inflammation^{94, 95}. Thus, diet, through its modulatory effect on the gut microbiota, might modify the risk of developing IBD (Figure 2)

Patients with established IBD have dysbiosis of the gut microbiome, characterized by reduced bacterial diversity, enrichment of bacteria of the family Enterobacteriaceae and reduced abundance of bacteria of the phylum Firmicutes and the genus *Bacteroides* (Figure 2)^{92, 96-98}. Interestingly, similar patterns of bacterial enrichment have been reported in observational studies of healthy volunteers fed a high-fat, low-fiber diet, suggesting that prior epidemiological findings demonstrating an inverse association between fiber intake and risk of Crohn's disease might in part be explained by diet-mediated alteration of the gut microbiota in genetically susceptible individuals^{99, 100}.

The majority of prior studies investigating the interaction between diet, the microbiome and intestinal inflammation have been conducted in animal models of colitis. In mice, dietary heme iron was shown to directly injure the colonic surface epithelium through the generation of cytotoxic and oxidative stress^{101, 102}. Moreover, increased dietary heme iron intake has also been associated with marked changes in the composition of the gut microbiota, with an increased ratio of Gram-negative:Gram-positive bacteria in mice¹⁰³. This effect was primarily driven by the increased abundance of Gram-negative species, including those belonging to the genus *Bacteroides* and *Akkermansia*, leading to a marked increase in lipopolysaccharide production¹⁰³. In a 2017 study, *Akkermansia muciniphila* was shown in mice to be a pathobiont, promoting colitis in genetically susceptible hosts¹⁰⁴. Interestingly, alteration in the function of the gut mucosal barrier related to dietary intake of heme iron seems to be dependent on the presence of sulfide-producing and mucin-degrading bacteria (for example, *Akkermansia*)¹⁰¹. By contrast, in a mouse model of spontaneous ileitis, depletion of luminal iron altered gut microbial composition to promote inflammation¹⁰⁵. Finally, a high-fat diet has been linked to changes in phospholipid profile and bacteria taxa in the gut, highlighting the complex interactions between the host and the gut microbiome in response to high fat intake^{106, 107}. More specifically, high intake of saturated fat altered the gut microbiome, characterized by an increase in the sulfite-reducing pathobiont *Bilophila wadsworthia*, and induced colonic inflammation in *III0*-knockout mice¹⁰⁸. This data provided a potential mechanism by which diets high in saturated fat might increase the risk of IBD in a susceptible host.

Human observational studies examining the interaction between diet, the microbiome and IBD are scarce. Among these studies is the ongoing multinational Genetics, Environment and Microbiome study that aims to identify factors that increase the risk of IBD^{109, 110}. This study is examining several environmental exposures, such as being breastfed and the composition of the microbiome, in a cohort of patients at high risk of IBD. Interestingly, preliminary data from this study has not shown a correlation between intestinal permeability¹¹⁰ — a key pathogenic pathway in IBD that is known to be influenced by diet⁷⁸ — and the gut microbiota. Specific patterns of association have been reported in a large Dutch cohort, in which total carbohydrate intake and features of a Western diet, including high caloric intake and consumption of sugar-sweetened beverages, were negatively associated with gut microbiome diversity¹¹¹. By contrast, features of a Mediterranean diet, such as consumption of fruits, vegetables and red wine, were associated with increased diversity of the gut microbiome. Red wine consumption was also associated with the increased abundance of *Faecalibacterium prausnitzii*, which has been proposed to have anti-inflammatory properties in patients with IBD¹¹².

Several studies have examined the correlation between host genetics and the gut microbiome in healthy individuals and patients with IBD^{113–115}. Generally, these studies have established a potential association between several genetic variants, including IBD-specific risk variants, and the gut microbiome. However, perhaps the most intriguing finding has been the observation that gene–diet interaction could in part regulate the composition of human gut microbiome. Using metagenomic data from >1500 healthy individuals, it was found that the abundance of bacteria of the genus *Bifidobacterium* was regulated by a functional variant within the *LCT* gene, which encodes lactase, the enzyme responsible for the breakdown of lactose¹¹⁶. Interestingly, this association was further modified by dairy intake. These studies further illustrate the interaction between diet, host genetics and the gut microbiome in human health and diseases of the gastrointestinal tract and highlights the complex relationship between these factors in studying IBD pathogenesis (FIG. 3).

Conclusions

There is growing body of evidence suggesting a role for diet in the development of IBD, particularly among genetically susceptible individuals. In addition, data from animal studies suggest that dietary modification might alter the risk of IBD. Nevertheless, there continues to be a substantial need for a comprehensive evaluation of the role of diet in the etiopathogenesis of IBD. Such studies will be challenging because of the complex nature of the relationship between diet, genetics, the microbiome and disease activity. A number of large international collaborations are currently in the planning stages and hope to provide an in-depth prospective examination of the role of specific diets in IBD and overcome many of the shortcomings of prior studies. The successful execution of these studies will represent a critical first step in advancing our understanding of the role of diet in the pathogenesis of IBD.

Acknowledgements

HK is funded by the National Institute of Diabetes and Digestive and Kidney Diseases (K23 DK099681 and R03), ATC is funded by NIH K24 DK098311, a Stuart and Suzanne Steele MGH Research Scholar Award, and a Senior Investigator Award from the Crohn's and Colitis Foundation.

Financial Disclosures: H. K. has received consulting fees from Abbvie and Samsung Bioepis. H. K. also receives funding from Takeda. S. S. M. C. has received consulting fees from Abbvie and Ferring Pharmaceuticals. P. L. is supported by a career development grant by CCF. A. N. A. is a member of the scientific advisory board for Exact Sciences, Abbvie, and Cubist pharmaceuticals. A. T. C. has served as a consultant for Bayer Healthcare, Pfizer Inc., and Takeda.

REFERENCES

1. Feskanich D et al. Reproducibility and validity of food intake measurements from a semiquantitative food frequency questionnaire. *J Am Diet Assoc* 93, 790–6 (1993). [PubMed: 8320406]
2. Rimm E et al. Reproducibility and validity of an expanded self-administered semiquantitative food questionnaire among health professionals. *Am J Epidemiol* 135, 1114–1126 (1992). [PubMed: 1632423]
3. Rockett HR et al. Validation of a youth/adolescent food frequency questionnaire. *Prev Med* 26, 808–16 (1997). [PubMed: 9388792]
4. Rockett HR, Wolf AM & Colditz GA Development and reproducibility of a food frequency questionnaire to assess diets of older children and adolescents. *J Am Diet Assoc* 95, 336–40 (1995). [PubMed: 7860946]
5. Ascherio A et al. Correlations of vitamin A and E intakes with the plasma concentrations of carotenoids and tocopherols among American men and women. *J Nutr* 122, 1792–801 (1992). [PubMed: 1512628]
6. Romieu I et al. Food predictors of plasma beta-carotene and alpha-tocopherol: validation of a food frequency questionnaire. *Am J Epidemiol* 131, 864–76 (1990). [PubMed: 2321629]
7. Willett WC et al. Validation of dietary questionnaire with plasma carotenoid and alpha-tocopherol level. *Am J Clin Nutr* 38, 631–639 (1983). [PubMed: 6624705]
8. Giovannucci E et al. Folate, methionine, and alcohol intake and risk of colorectal adenoma. *J Natl Cancer Inst* 85, 875–84. (1993). [PubMed: 8492316]
9. Hunter DJ et al. Comparison of measures of fatty acid intake by subcutaneous fat aspirate, food frequency questionnaire, and diet records in a free-living population of US men. *Am J Epidemiol* 135, 418–27 (1992). [PubMed: 1550093]
10. London SJ et al. Fatty acid composition of subcutaneous adipose tissue and diet in postmenopausal US women. *Am J Clin Nutr* 54, 340–5 (1991). [PubMed: 1858698]
11. Yuan C et al. Validity of a Dietary Questionnaire Assessed by Comparison With Multiple Weighed Dietary Records or 24-Hour Recalls. *Am J Epidemiol* 185, 570–584 (2017). [PubMed: 28338828]
12. Molodecky NA et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology* 142, 46–54 e42; quiz e30 (2012). [PubMed: 22001864]
13. Burisch J, Jess T, Martinato M, Lakatos PL & EpiCom E The burden of inflammatory bowel disease in Europe. *J Crohns Colitis* 7, 322–37 (2013). [PubMed: 23395397]
14. Kappelman MD et al. The prevalence and geographic distribution of Crohn's disease and ulcerative colitis in the United States. *Clin Gastroenterol Hepatol* 5, 1424–9 (2007). [PubMed: 17904915]
15. Ferguson A, Sedgwick DM & Drummond J Morbidity of juvenile onset inflammatory bowel disease: effects on education and employment in early adult life. *Gut* 35, 665–8 (1994). [PubMed: 8200562]
16. Kappelman MD et al. Direct health care costs of Crohn's disease and ulcerative colitis in US children and adults. *Gastroenterology* 135, 1907–13 (2008). [PubMed: 18854185]
17. Longobardi T, Jacobs P & Bernstein CN Work losses related to inflammatory bowel disease in the United States: results from the National Health Interview Survey. *The American journal of gastroenterology* 98, 1064–72 (2003). [PubMed: 12809829]

18. Franke A et al. Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nature genetics* 42, 1118–25 (2010). [PubMed: 21102463]
19. Kenny EE et al. A genome-wide scan of Ashkenazi Jewish Crohn's disease suggests novel susceptibility loci. *PLoS Genetics* 8, e1002559 (2012).
20. Kugathasan S & Amre D Inflammatory bowel disease–environmental modification and genetic determinants. *Pediatric clinics of North America* 53, 727–49 (2006). [PubMed: 16873002]
21. Tysk C, Lindberg E, Jarnerot G & Floderus-Myrhed B Ulcerative colitis and Crohn's disease in an unselected population of monozygotic and dizygotic twins. A study of heritability and the influence of smoking. *Gut* 29, 990–6 (1988). [PubMed: 3396969]
22. Ng SC et al. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet* 390, 2769–2778 (2018). [PubMed: 29050646]
23. Williams CN Does the incidence of IBD increase when persons move from a low- to a high-risk area? *Inflammatory bowel diseases* 14 Suppl 2, S41–2 (2008). [PubMed: 18816730]
24. Probert CS, Jayanthi V, Pinder D, Wicks AC & Mayberry JF Epidemiological study of ulcerative proctocolitis in Indian migrants and the indigenous population of Leicestershire. *Gut* 33, 687–93 (1992). [PubMed: 1307684]
25. Benchimol EI et al. Inflammatory bowel disease in immigrants to Canada and their children: a population-based cohort study. *Am J Gastroenterol* 110, 553–63 (2015). [PubMed: 25756238]
26. Li X, Sundquist J, Hemminki K & Sundquist K Risk of inflammatory bowel disease in first- and second-generation immigrants in Sweden: a nationwide follow-up study. *Inflamm Bowel Dis* 17, 1784–91 (2011). [PubMed: 21744434]
27. Albenberg LG & Wu GD Diet and the intestinal microbiome: associations, functions, and implications for health and disease. *Gastroenterology* 146, 1564–72 (2014). [PubMed: 24503132]
28. Poullis A, Foster R, Shetty A, Fagerhol MK & Mendall MA Bowel inflammation as measured by fecal calprotectin: a link between lifestyle factors and colorectal cancer risk. *Cancer Epidemiol Biomarkers Prev* 13, 279–84 (2004). [PubMed: 14973103]
29. Martinez-Medina M et al. Western diet induces dysbiosis with increased E coli in CEABAC10 mice, alters host barrier function favouring AIEC colonisation. *Gut* 63, 116–24 (2014). [PubMed: 23598352]
30. Stenman LK, Holma R, Eggert A & Korpela R A novel mechanism for gut barrier dysfunction by dietary fat: epithelial disruption by hydrophobic bile acids. *Am J Physiol Gastrointest Liver Physiol* 304, G227–34 (2013). [PubMed: 23203158]
31. Halmos EP & Gibson PR Dietary management of IBD--insights and advice. *Nat Rev Gastroenterol Hepatol* 12, 133–46 (2015). [PubMed: 25645969]
32. Borrelli O et al. Polymeric diet alone versus corticosteroids in the treatment of active pediatric Crohn's disease: a randomized controlled open-label trial. *Clin Gastroenterol Hepatol* 4, 744–53 (2006). [PubMed: 16682258]
33. Zachos M, Tondeur M & Griffiths AM Enteral nutritional therapy for induction of remission in Crohn's disease. *Cochrane Database Syst Rev*, CD000542 (2007). [PubMed: 17253452]
34. Charlebois A, Rosenfeld G & Bressler B The Impact of Dietary Interventions on the Symptoms of Inflammatory Bowel Disease: A Systematic Review. *Crit Rev Food Sci Nutr* 56, 1370–8 (2016). [PubMed: 25569442]
35. Persson PG, Ahlbom A & Hellers G Diet and inflammatory bowel disease: a case-control study. *Epidemiology* 3, 47–52 (1992). [PubMed: 1313310]
36. Chapman-Kiddell CA, Davies PS, Gillen L & Radford-Smith GL Role of diet in the development of inflammatory bowel disease. *Inflamm Bowel Dis* 16, 137–51 (2010). [PubMed: 19462428]
37. Tragnone A et al. Dietary habits as risk factors for inflammatory bowel disease. *Eur J Gastroenterol Hepatol* 7, 47–51 (1995). [PubMed: 7866810]
38. Willett WC, Reynolds RD, Cottrell-Hoehner S, Sampson L & Browne ML Validation of a semi-quantitative food frequency questionnaire: comparison with a 1-year diet record. *J Am Diet Assoc* 87, 43–7 (1987). [PubMed: 3794132]
39. Colditz GA, Manson JE & Hankinson SE The Nurses' Health Study: 20-year contribution to the understanding of health among women. *J Womens Health* 6, 49–62 (1997). [PubMed: 9065374]

40. Ananthakrishnan AN et al. A prospective study of long-term intake of dietary fiber and risk of Crohn's disease and ulcerative colitis. *Gastroenterology* 145, 970–7 (2013). [PubMed: 23912083]
41. Ananthakrishnan AN et al. Long-term intake of dietary fat and risk of ulcerative colitis and Crohn's disease. *Gut* 63, 776–84 (2014). [PubMed: 23828881]
42. Khalili H et al. Dietary Iron and Heme Iron Consumption, Genetic Susceptibility, and Risk of Crohn's Disease and Ulcerative Colitis. *Inflamm Bowel Dis* 23, 1088–1095 (2017). [PubMed: 28604414]
43. Bettelli E et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* 441, 235–8 (2006). [PubMed: 16648838]
44. Eil R et al. Ionic immune suppression within the tumour microenvironment limits T cell effector function. *Nature* 537, 539–543 (2016). [PubMed: 27626381]
45. Kleiweiefeld M et al. Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. *Nature* 496, 518–22 (2013). [PubMed: 23467095]
46. Wu C et al. Induction of pathogenic TH17 cells by inducible salt-sensing kinase SGK1. *Nature* 496, 513–7 (2013). [PubMed: 23467085]
47. Yosef N et al. Dynamic regulatory network controlling TH17 cell differentiation. *Nature* 496, 461–8 (2013). [PubMed: 23467089]
48. Bao S et al. Zinc modulates the innate immune response in vivo to polymicrobial sepsis through regulation of NF-kappaB. *Am J Physiol Lung Cell Mol Physiol* 298, L744–54 (2010). [PubMed: 20207754]
49. Finamore A, Massimi M, Conti Devirgiliis L & Mengheri E Zinc deficiency induces membrane barrier damage and increases neutrophil transmigration in Caco-2 cells. *J Nutr* 138, 1664–70 (2008). [PubMed: 18716167]
50. Prasad AS Zinc: role in immunity, oxidative stress and chronic inflammation. *Curr Opin Clin Nutr Metab Care* 12, 646–52 (2009). [PubMed: 19710611]
51. Khalili H et al. Identification and Characterization of a Novel Association between Dietary Potassium and Risk of Crohn's Disease and Ulcerative Colitis. *Front Immunol* 7, 554 (2016). [PubMed: 28003811]
52. Ananthakrishnan AN et al. Zinc intake and risk of Crohn's disease and ulcerative colitis: a prospective cohort study. *Int J Epidemiol* 44, 1995–2005 (2015). [PubMed: 26546032]
53. Riboli E et al. European Prospective Investigation into Cancer and Nutrition (EPIC): study populations and data collection. *Public Health Nutr* 5, 1113–24 (2002). [PubMed: 12639222]
54. Ferrari P et al. Within- and between-cohort variation in measured macronutrient intakes, taking account of measurement errors, in the European Prospective Investigation into Cancer and Nutrition study. *Am J Epidemiol* 160, 814–22 (2004). [PubMed: 15466504]
55. Hart AR et al. Diet in the aetiology of ulcerative colitis: a European prospective cohort study. *Digestion* 77, 57–64 (2008). [PubMed: 18349539]
56. Investigators., I.i.E.S. et al. Linoleic acid, a dietary n-6 polyunsaturated fatty acid, and the aetiology of ulcerative colitis: a nested case-control study within a European prospective cohort study. *Gut* 58, 1606–11 (2009). [PubMed: 19628674]
57. Calder PC Fatty acids and inflammation: the cutting edge between food and pharma. *Eur J Pharmacol* 668 Suppl 1, S50–8 (2011). [PubMed: 21816146]
58. Simopoulos AP Evolutionary aspects of diet, the omega-6/omega-3 ratio and genetic variation: nutritional implications for chronic diseases. *Biomed Pharmacother* 60, 502–7 (2006). [PubMed: 17045449]
59. Chan SS et al. Association between high dietary intake of the n-3 polyunsaturated fatty acid docosahexaenoic acid and reduced risk of Crohn's disease. *Aliment Pharmacol Ther* 39, 834–42 (2014). [PubMed: 24611981]
60. Chan SS et al. Carbohydrate intake in the etiology of Crohn's disease and ulcerative colitis. *Inflamm Bowel Dis* 20, 2013–21 (2014). [PubMed: 25265262]
61. Racine A et al. Dietary Patterns and Risk of Inflammatory Bowel Disease in Europe: Results from the EPIC Study. *Inflamm Bowel Dis* 22, 345–54 (2016). [PubMed: 26717318]

62. Opstelten JL et al. Dairy Products, Dietary Calcium, and Risk of Inflammatory Bowel Disease: Results From a European Prospective Cohort Investigation. *Inflamm Bowel Dis* 22, 1403–11 (2016). [PubMed: 27120568]
63. Bergmann MM et al. No association of alcohol use and the risk of ulcerative colitis or Crohn's disease: data from a European Prospective cohort study (EPIC). *Eur J Clin Nutr* 71, 512–518 (2017). [PubMed: 28120853]
64. Gavaler JS, Rosenblum ER, Deal SR & Bowie BT The phytoestrogen congeners of alcoholic beverages: current status. *Proc Soc Exp Biol Med* 208, 98–102 (1995). [PubMed: 7892304]
65. Seifried HE, Anderson DE, Fisher EI & Milner JA A review of the interaction among dietary antioxidants and reactive oxygen species. *J Nutr Biochem* 18, 567–79 (2007). [PubMed: 17360173]
66. Yamamoto T, Nakahigashi M & Saniabadi AR Review article: diet and inflammatory bowel disease--epidemiology and treatment. *Aliment Pharmacol Ther* 30, 99–112 (2009). [PubMed: 19438426]
67. Barton LL, Ritz NL, Fauque GD & Lin HC Sulfur Cycling and the Intestinal Microbiome. *Dig Dis Sci* 62, 2241–2257 (2017). [PubMed: 28766244]
68. Kim IW et al. Western-style diets induce macrophage infiltration and contribute to colitis-associated carcinogenesis. *J Gastroenterol Hepatol* 25, 1785–94.
69. Breton J et al. Intrinsic immunomodulatory effects of low-digestible carbohydrates selectively extend their anti-inflammatory prebiotic potentials. *Biomed Res Int* 2015, 162398 (2015). [PubMed: 25977916]
70. Le Leu RK, Young GP, Hu Y, Winter J & Conlon MA Dietary red meat aggravates dextran sulfate sodium-induced colitis in mice whereas resistant starch attenuates inflammation. *Dig Dis Sci* 58, 3475–82 (2013). [PubMed: 23990000]
71. Degagne E et al. Sphingosine-1-phosphate lyase downregulation promotes colon carcinogenesis through STAT3-activated microRNAs. *J Clin Invest* 124, 5368–84 (2014). [PubMed: 25347472]
72. Jantchou P, Morois S, Clavel-Chapelon F, Boutron-Ruault MC & Carbonnel F Animal protein intake and risk of inflammatory bowel disease: The E3N prospective study. *Am J Gastroenterol* 105, 2195–201 (2010). [PubMed: 20461067]
73. Liang J et al. Sphingosine-1-phosphate links persistent STAT3 activation, chronic intestinal inflammation, and development of colitis-associated cancer. *Cancer Cell* 23, 107–20 (2013). [PubMed: 23273921]
74. Hernandez AL et al. Sodium chloride inhibits the suppressive function of FOXP3+ regulatory T cells. *J Clin Invest* 125, 4212–22 (2015). [PubMed: 26524592]
75. Zhang WC et al. High salt primes a specific activation state of macrophages, M(Na). *Cell Res* 25, 893–910 (2015). [PubMed: 26206316]
76. Hunter DJ Gene-environment interactions in human diseases. *Nat Rev Genet* 6, 287–98 (2005). [PubMed: 15803198]
77. Khor B, Gardet A & Xavier RJ Genetics and pathogenesis of inflammatory bowel disease. *Nature* 474, 307–17 (2011). [PubMed: 21677747]
78. Leone VA, Cham CM & Chang EB Diet, gut microbes, and genetics in immune function: can we leverage our current knowledge to achieve better outcomes in inflammatory bowel diseases? *Curr Opin Immunol* 31, 16–23 (2014). [PubMed: 25214301]
79. Van Der Sloot KW et al. Visceral Adiposity, Genetic Susceptibility, and Risk of Complications Among Individuals with Crohn's Disease. *Inflamm Bowel Dis* 23, 82–88 (2017). [PubMed: 27893544]
80. Ananthkrishnan AN et al. Genetic polymorphisms in fatty acid metabolism modify the association between dietary n3:n6 intake and risk of ulcerative colitis. *Inflamm Bowel Dis* Accepted (5 2017).
81. Costea I et al. Interactions between the dietary polyunsaturated fatty acid ratio and genetic factors determine susceptibility to pediatric Crohn's disease. *Gastroenterology* 146, 929–31 (2014). [PubMed: 24406470]
82. Hargreaves CE et al. Fcγ receptors: genetic variation, function, and disease. *Immunol Rev* 268, 6–24 (2015). [PubMed: 26497510]

83. Stein MP et al. C-reactive protein binding to FcγRIIIa on human monocytes and neutrophils is allele-specific. *J Clin Invest* 105, 369–76 (2000). [PubMed: 10675363]
84. Stein MP, Mold C & Du Clos TW C-reactive protein binding to murine leukocytes requires Fcγ receptors. *J Immunol* 164, 1514–20 (2000). [PubMed: 10640769]
85. Bharadwaj D, Stein MP, Volzer M, Mold C & Du Clos TW The major receptor for C-reactive protein on leukocytes is fcγ receptor II. *J Exp Med* 190, 585–90 (1999). [PubMed: 10449529]
86. Zhou L et al. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. *Nature immunology* 8, 967–74 (2007). [PubMed: 17581537]
87. Fantini MC et al. IL-21 regulates experimental colitis by modulating the balance between Treg and Th17 cells. *Eur J Immunol* 37, 3155–63 (2007). [PubMed: 17918200]
88. Ananthkrishnan AN et al. Genetic Polymorphisms in Fatty Acid Metabolism Modify the Association Between Dietary n3: n6 Intake and Risk of Ulcerative Colitis: A Prospective Cohort Study. *Inflamm Bowel Dis* 23, 1898–1904 (2017). [PubMed: 28991856]
89. Caspi A et al. Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science* 301, 386–9 (2003). [PubMed: 12869766]
90. Risch N et al. Interaction between the serotonin transporter gene (5-HTTLPR), stressful life events, and risk of depression: a meta-analysis. *JAMA* 301, 2462–71 (2009). [PubMed: 19531786]
91. Kau AL, Ahern PP, Griffin NW, Goodman AL & Gordon JI Human nutrition, the gut microbiome and the immune system. *Nature* 474, 327–36 (2011). [PubMed: 21677749]
92. Brown K, DeCoffe D, Molcan E & Gibson DL Diet-induced dysbiosis of the intestinal microbiota and the effects on immunity and disease. *Nutrients* 4, 1095–119 (2012). [PubMed: 23016134]
93. D’Haens GR et al. Early lesions of recurrent Crohn’s disease caused by infusion of intestinal contents in excluded ileum. *Gastroenterology* 114, 262–7 (1998). [PubMed: 9453485]
94. Sellon RK et al. Resident enteric bacteria are necessary for development of spontaneous colitis and immune system activation in interleukin-10-deficient mice. *Infect Immun* 66, 5224–31 (1998). [PubMed: 9784526]
95. Hudcovic T, Stepankova R, Cebra J & Tlaskalova-Hogenova H The role of microflora in the development of intestinal inflammation: acute and chronic colitis induced by dextran sulfate in germ-free and conventionally reared immunocompetent and immunodeficient mice. *Folia Microbiol (Praha)* 46, 565–72 (2001). [PubMed: 11898350]
96. Morgan XC et al. Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment. *Genome Biol* 13, R79 (2012). [PubMed: 23013615]
97. Nagalingam NA & Lynch SV Role of the microbiota in inflammatory bowel diseases. *Inflamm Bowel Dis* 18, 968–84 (2012). [PubMed: 21936031]
98. Spor A, Koren O & Ley R Unravelling the effects of the environment and host genotype on the gut microbiome. *Nat Rev Microbiol* 9, 279–90 (2011). [PubMed: 21407244]
99. Wu GD et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* 334, 105–8 (2011). [PubMed: 21885731]
100. De Filippo C et al. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc Natl Acad Sci U S A* 107, 14691–6 (2010). [PubMed: 20679230]
101. Ijssennagger N et al. Gut microbiota facilitates dietary heme-induced epithelial hyperproliferation by opening the mucus barrier in colon. *Proc Natl Acad Sci U S A* 112, 10038–43 (2015). [PubMed: 26216954]
102. Ijssennagger N, de Wit N, Muller M & van der Meer R Dietary heme-mediated PPARα activation does not affect the heme-induced epithelial hyperproliferation and hyperplasia in mouse colon. *PLoS One* 7, e43260 (2012). [PubMed: 22905243]
103. N IJ et al. Dietary heme alters microbiota and mucosa of mouse colon without functional changes in host-microbe cross-talk. *PLoS One* 7, e49868 (2012). [PubMed: 23239972]
104. Seregin SS et al. NLRP6 Protects Il10(–/–) Mice from Colitis by Limiting Colonization of *Akkermansia muciniphila*. *Cell Rep* 19, 733–745 (2017). [PubMed: 28445725]

105. Werner T et al. Depletion of luminal iron alters the gut microbiota and prevents Crohn's disease-like ileitis. *Gut* 60, 325–33 (2011). [PubMed: 21076126]
106. Turnbaugh PJ, Backhed F, Fulton L & Gordon JI Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. *Cell Host Microbe* 3, 213–23 (2008). [PubMed: 18407065]
107. David LA et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 505, 559–63 (2014). [PubMed: 24336217]
108. Devkota S et al. Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in *Il10*^{-/-} mice. *Nature* 487, 104–8 (2012). [PubMed: 22722865]
109. Kevans D et al. IBD Genetic Risk Profile in Healthy First-Degree Relatives of Crohn's Disease Patients. *J Crohns Colitis* 10, 209–15 (2016). [PubMed: 26512135]
110. Kevans D et al. Determinants of intestinal permeability in healthy first-degree relatives of individuals with Crohn's disease. *Inflamm Bowel Dis* 21, 879–87 (2015). [PubMed: 25734694]
111. Zhernakova A et al. Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity. *Science* 352, 565–9 (2016). [PubMed: 27126040]
112. Sokol H et al. *Faecalibacterium prausnitzii* is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proc Natl Acad Sci US A* 105, 16731–6 (2008).
113. Imhann F et al. Interplay of host genetics and gut microbiota underlying the onset and clinical presentation of inflammatory bowel disease. *Gut* (2016).
114. Knights D et al. Complex host genetics influence the microbiome in inflammatory bowel disease. *Genome Med* 6, 107 (2014). [PubMed: 25587358]
115. Turpin W et al. Association of host genome with intestinal microbial composition in a large healthy cohort. *Nat Genet* 48, 1413–1417 (2016). [PubMed: 27694960]
116. Bonder MJ et al. The effect of host genetics on the gut microbiome. *Nat Genet* 48, 1407–1412 (2016). [PubMed: 27694959]
117. Ananthakrishnan AN et al. High School Diet and Risk of Crohn's Disease and Ulcerative Colitis. *Inflamm Bowel Dis* 21, 2311–9 (2015). [PubMed: 26236952]
118. Bergmann MM et al. No association of alcohol use and the risk of ulcerative colitis or Crohn's disease: data from a European Prospective cohort study (EPIC). *Eur J Clin Nutr* 71, 566 (2017). [PubMed: 28377581]

Box 1 :**SFFQ validation studies.**

This SFFQ has been extensively validated in several studies, including observations that : nutrient intake assessed by the SFFQ strongly correlated with detailed dietary records ($r=0.65$)¹⁻³; specific items between 2 questionnaires administered 1 year apart were strongly correlated ($r=0.67$)¹⁻⁴; nutrients calculated from the SFFQ correlated with biochemical indicators of dietary intake, including plasma levels of beta-carotene ($r=0.30-0.42$) and vitamin E ($r=0.41-0.53$), as well as red blood cell levels of folate ($r=0.55$)⁵⁻⁸; estimates of fatty acid intake correlated with adipose tissue levels of linoleic acid ($r=0.35-0.37$), trans-fatty acid ($r=0.51$) and n-3 fatty acids ($r=0.48-0.49$)^{9,10}. Repeated validation studies of the FFQ in the NHS have yielded similar and stable correlations with diet records over time. In a validation study published in 2017, which was conducted in 632 women from the NHS cohort between 2010 and 2012, the FFQ was correlated with two 7-day dietary records that were administered one year apart ($r=0.62$) and automated self-administered 24-hour recalls ($r=0.63$) over a one year period, suggesting that the SFFQ provides reasonably valid estimates of intakes of a wide variety of dietary variables¹¹. These correlations probably underestimate the validity of the SFFQ because they also incorporate the errors related to use of the 6-day dietary record for estimating long-term intake

Key points

- 1) Epidemiological studies have identified a few potential dietary risk factors for Crohn's disease and ulcerative colitis.
- 2) Early results from dietary intervention studies in treatment of Crohn's disease and ulcerative colitis have been promising particularly in pediatrics but high quality randomized trials are needed to assess their efficacy
- 3) Preliminary gene-environment and microbiome studies have demonstrated an interaction between diet, host genetics, and the gut microbiota in the etiopathogenesis of Crohn's disease and ulcerative colitis.
- 4) Future large-scale studies to prospectively examine the role of diet in Crohn's disease and ulcerative colitis in the context of host genetics and gut microbiota are needed.

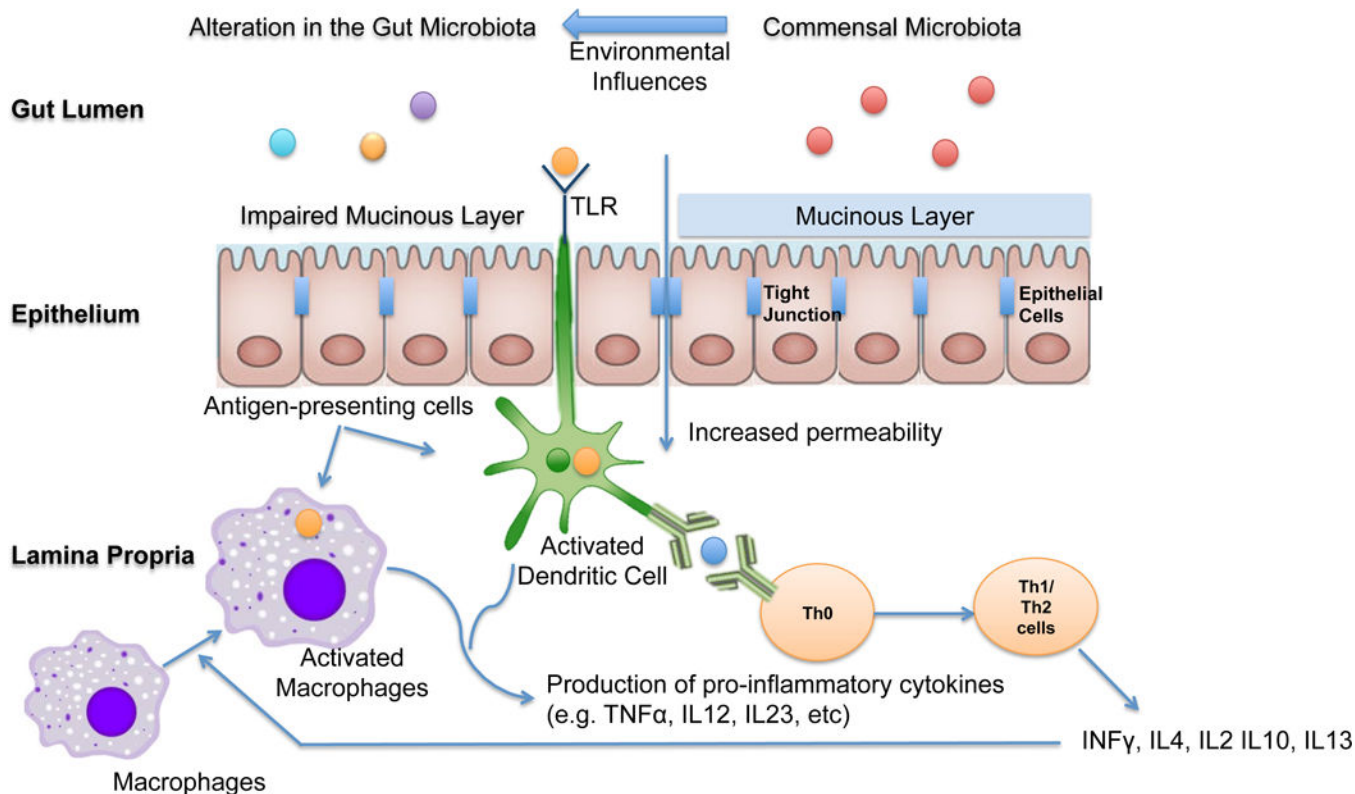


Figure 1 |. The pathophysiology of inflammatory bowel disease.

The pathophysiology of inflammatory bowel disease (IBD) is related to an inappropriate host immune response to commensal bacteria in genetically susceptible individuals. Environmental influences alter the composition of the gut microbiota and change mucosal barrier function. Dendritic cells and macrophages are antigen-presenting cells involved in activation of T cells and production of pro-inflammatory cytokines. Dendritic cells are activated through the recognition of luminal antigens by Toll-like receptors (TLRs) and are in turn necessary for activation of naïve T cells. Macrophages could also serve as antigen-presenting cells once stimulated by INF γ secreted by T cells. Activated macrophages and dendritic cells also produce pro-inflammatory cytokines including TNF-alpha, IL212, and IL23. The result of this pro-inflammatory cytokine cascade is loss of immune tolerance to commensal bacteria and the production of pro-inflammatory cytokines by activated T cells. INF, Inteferon-gamma; TNF, tumour necrosis factor; IL-312, interleukin-12; IL-23, interleukin-23;

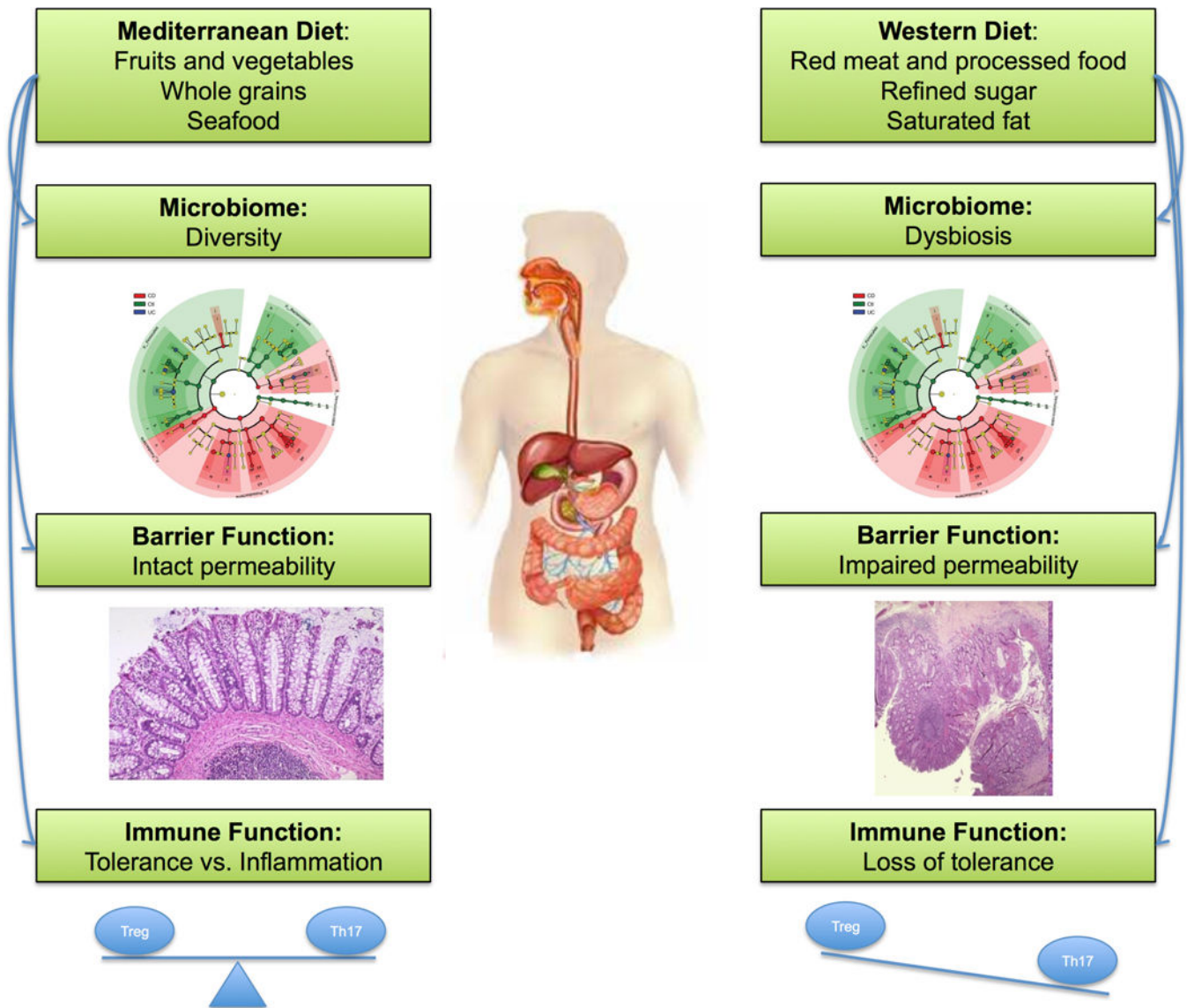


Figure 2 |. Potential mechanisms underpinning the relationship between diet and inflammatory bowel disease.

As compared to Mediterranean diet which is characterized by high intake of fruits and vegetables, whole grains, and sea good, western diet characterized by high intake of red and processed meat, saturated fat, and refined sugar is widely thought to increase risk of IBD. Although the exact mechanism underpinning the association between diet and risk of IBD is unknown, a number of plausible mechanisms have been proposed. Specifically, western diet has been linked to changes in the gut microbiome and epithelial barrier function and appears to have a direct impact on the immune function triggering a pro-inflammatory environment characterized by an imbalance in the T_H17/T_{reg} axis.

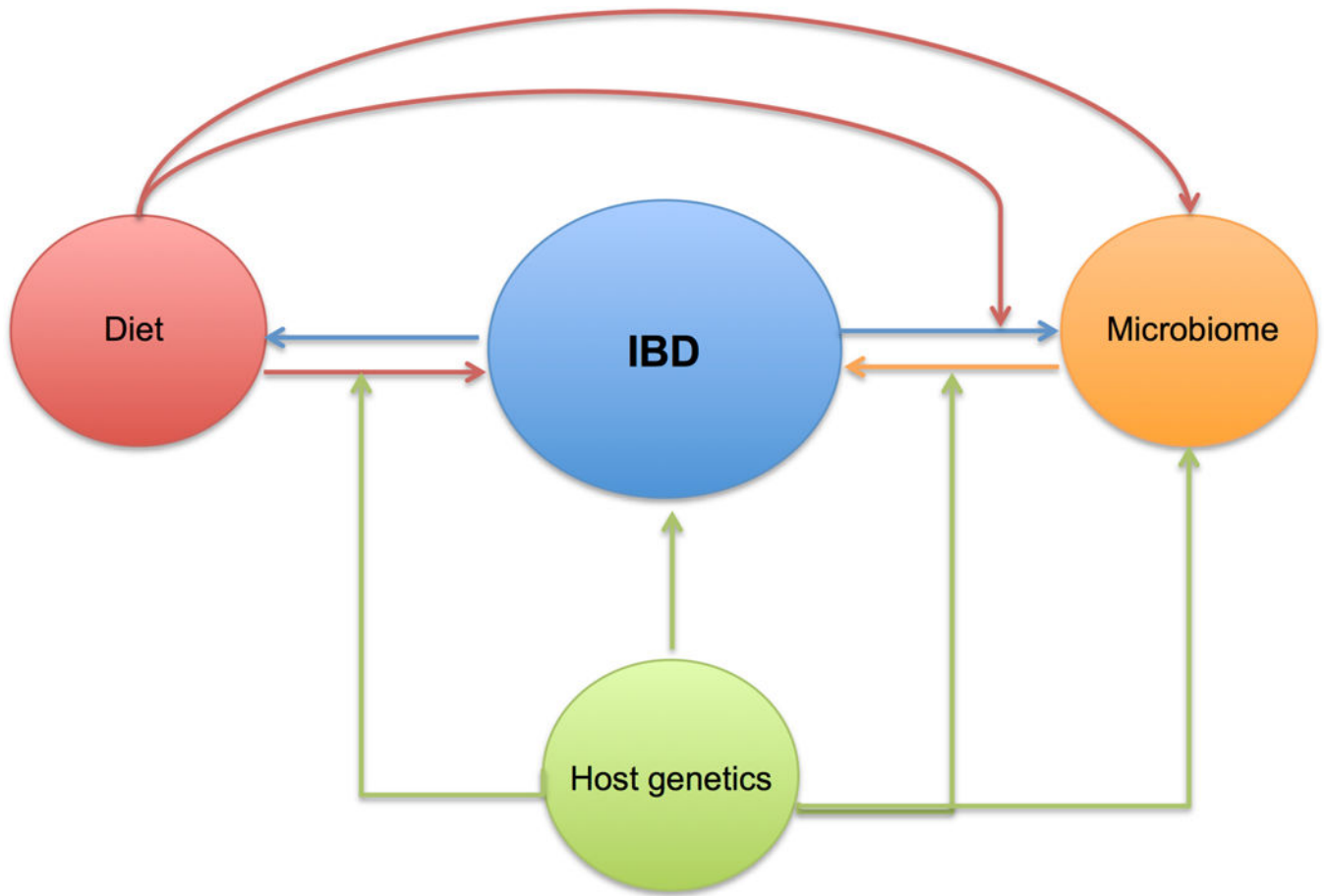


Figure 3 |. The complex causal relationship between diet and inflammatory bowel disease-. The pathophysiology of IBD is thought to be related to inappropriate immune response to the gut microbiome in the genetically susceptible host. The composition of the gut microbiome is primarily dictated by environmental determinants particularly diet and the host genetics. In turn dietary factors may have differential influence on the immune function and risk of IBD according to the host genetics. Studies examining the role of diet, gut microbiome, and host genetics on risk of IBD are particularly complicated by the reciprocal effect of active disease on the gut microbiome and individuals' dietary choices.

Table 1|

Prospective studies of diet and inflammatory bowel disease in the EPIC, NHS and NHSII cohorts

Study	Dietary exposure(s)	Outcome(s)	Population and design	Key findings*	Refs
Hart <i>et al.</i> 2008	Intake of carbohydrates; fatty acids; protein; fiber; alcohol; and micronutrients, including vitamin C, vitamin D, vitamin E, calcium and iron.	UC	<ul style="list-style-type: none"> Nested case– control study using prospectively-collected dietary data from the EPIC cohort. 139 cases of incident UC matched 1:4 to 556 controls. 	<ul style="list-style-type: none"> No statistically significant associations between dietary factor intakes and UC. A nonsignificant trend towards higher risk of UC associated with PUFA as percentage of total energy intake (OR 1.56, 95% CI 0.87–2.77 for highest versus lowest quartile; $P_{\text{trend}}=0.07$). 	55
Hart <i>et al.</i> 2009	Intake of linoleic acid; α -linoleic acid; oleic acid; DHA; and EPA.	UC	<ul style="list-style-type: none"> Nested case– control study using prospectively-collected dietary data from the EPIC cohort. 126 cases of incident UC over matched 1:4 to 504 controls. 	<ul style="list-style-type: none"> Higher linoleic acid intake was associated with increased risk of UC (OR 2.49, 95% CI 1.23–5.07 for highest versus lowest quartile; $P_{\text{trend}}=0.02$). Higher DHA intake was associated with lower risk of UC (OR 0.23, 95% CI 0.06–0.97 for highest versus lowest quartile; $P_{\text{trend}}=0.03$). 	56
Ananthakrishnan <i>et al.</i> 2013	Energy-adjusted, cumulative average intake of total dietary fiber; and fiber intake from different dietary sources.	CD and UC	<ul style="list-style-type: none"> Prospective analysis of 170,776 women enrolled in the NHS and NHS II. 269 incident cases of CD and 338 incident cases of UC during follow-up 	<ul style="list-style-type: none"> Total fiber intake was associated with lower risk of CD (HR 0.59, 95% CI 0.39–0.90 for highest versus lowest quintile; $P_{\text{trend}}=0.08$). The association was strongest for fiber from fruit sources (HR 0.57, 95% CI 0.38–0.85; $P_{\text{trend}}=0.02$). No statistically significant associations were observed between fiber intake and risk of UC. 	40
Ananthakrishnan <i>et al.</i> 2014	Energy-adjusted, cumulative average intake of total fat; saturated fats; unsaturated fats; and n-6 and n-3 PUFAs.	CD and UC	<ul style="list-style-type: none"> Prospective analysis of 170,805 women enrolled in the NHS and NHS II. 269 incident cases of CD and 338 incident cases of UC during follow-up. 	<ul style="list-style-type: none"> No associations were observed between intakes of total fat, saturated fat, monounsaturated fat or total PUFAs and risk of CD or UC. Higher n-3:n-6 PUFA ratio was associated with a lower risk of UC (HR 0.69, 95% CI 0.49–0.98 for highest versus lowest quintile; $P_{\text{trend}}=0.03$). 	41
Chan <i>et al.</i> 2014	Intake of total fat; linoleic acid; α -linoleic acid; oleic acid; DHA; and EPA.	CD	<ul style="list-style-type: none"> Nested case– control study using prospectively-collected dietary data from the EPIC cohort. 73 cases of incident CD matched 1:4 to 292 controls. 	<ul style="list-style-type: none"> Increasing intake of DHA was associated with a lower risk of CD (OR 0.07, 95% CI 0.02–0.81 for highest versus lowest quintile; $P_{\text{trend}}=0.04$). No statistically significant associations were observed for other fatty acids or total fat intake. 	59
Chan <i>et al.</i> 2014	Intake of total carbohydrates, simple sugars; and starch.	CD and UC	<ul style="list-style-type: none"> Nested case– control study using prospectively-collected dietary data from the EPIC cohort. 110 cases of incident CD and 244 cases of incident UC matched 1:4 to 440 and 976 controls, respectively. 	<ul style="list-style-type: none"> No statistically significant associations were observed between total dietary carbohydrate, sugar or starch intakes and the risk of CD or UC. 	60
Ananthakrishnan <i>et al.</i> 2015	Total zinc intake (from both dietary and supplemental sources).	CD and UC	<ul style="list-style-type: none"> Prospective analysis of 170,809 women enrolled in the NHS and NHS II. 269 incident cases of CD and 338 incident cases of UC during follow-up. 	<ul style="list-style-type: none"> Higher intake of zinc was associated with lower risk of CD (HR 0.57, 95% CI 0.38–0.86 for highest versus lowest quintile; $P_{\text{trend}}=0.003$). 	52

Study	Dietary exposure(s)	Outcome(s)	Population and design	Key findings*	Refs
				<ul style="list-style-type: none"> Reduction in risk of CD was observed with a daily zinc intake of up to 15mg. 	
Ananthakrishnan <i>et al.</i> 2015	'Prudent' and 'Western' high-school dietary patterns (at age 13–17 years).	CD and UC	<ul style="list-style-type: none"> Prospective analysis of 39,511 women enrolled in NHS II. 70 cases of CD and 103 cases of UC during follow-up. 	<ul style="list-style-type: none"> Higher prudent diet score during adolescence was associated with lower risk of CD (HR 0.47, 95% CI 0.23–0.98 for highest versus lowest score quartiles; $P_{\text{trend}}=0.04$) but not UC. 	117
Khalili <i>et al.</i> 2016	Dietary potassium and sodium intake.	CD and UC	<ul style="list-style-type: none"> Prospective analysis of 194,711 women enrolled in the NHS and NHS II. 273 incident cases of CD and 335 incident cases of UC during follow-up. 	<ul style="list-style-type: none"> Increasing dietary potassium intake was associated with lower risk of CD (HR 0.62, 95% CI 0.40–0.95 for highest versus lowest quintile; $P_{\text{trend}}=0.005$). There was a trend toward an inverse association between potassium intake and risk of UC ($P_{\text{trend}}=0.08$). 	51
Racine <i>et al.</i> 2016	aMED score and three <i>a posteriori</i> derived principal component dietary patterns.	CD and UC	<ul style="list-style-type: none"> Nested case–control study using prospectively-collected dietary data from the EPIC cohort. 117 cases of incident CD and 256 cases of incident UC matched 1:4 to 468 and 1022 controls, respectively. 	<ul style="list-style-type: none"> No association was detected between the aMED score and risk of CD or UC. There was an association between the 'sugar and soft drinks' dietary pattern and risk of UC (IRR 1.31, 95% CI 0.85–2.02 for the highest versus the lowest quintile; $P_{\text{trend}}=0.05$). 	61
Opstelten <i>et al.</i> 2016	Intake of milk; cheese; yoghurt; and total dairy products. Dietary calcium intake (without supplements).	CD and UC	<ul style="list-style-type: none"> Nested case – control study using prospectively-collected dietary data from the EPIC cohort. 110 cases of incident CD and 244 cases of incident UC matched 1:4 to 440 controls, respectively. 	<ul style="list-style-type: none"> No statistically significant associations with risk of CD or UC were observed across quartiles of total or individual dairy product intakes or dietary calcium intake. Compared to those who did not drink milk, any milk consumption was associated with a lower risk of CD (OR 0.30, 95% CI 0.13–0.65) but not UC. 	62
Khalili <i>et al.</i> 2017	Dietary total iron and heme iron intakes. Red and processed meat intakes.	CD and UC	<ul style="list-style-type: none"> Prospective analysis of 150,387 women enrolled in the NHS and NHS II. 260 incident cases of CD and 319 incident cases of UC during follow-up. 	<ul style="list-style-type: none"> There was a trend toward higher risk of UC associated with dietary heme iron intake (HR 1.26, 95% CI 0.86–1.83 for highest versus lowest quintile; $P_{\text{trend}}=0.10$). In a lagged analysis, red meat intake was associated with higher risk of UC (HR 1.15, 95% CI 1.01–1.30 for each additional weekly serving of red meat). 	42
Bergman <i>et al.</i> 2017	Habitual alcohol consumption during the 12 months prior to recruitment and at ages 20, 30, 40 and 50 years.	CD and UC	<ul style="list-style-type: none"> Nested case–control study using prospectively-collected dietary data from the EPIC cohort. 84 cases of incident CD and 198 cases of incident UC matched 1:4 to 336 and 792 controls, respectively. 	<ul style="list-style-type: none"> No statistically significant associations between alcohol consumption at recruitment, lifetime average alcohol use or pattern of lifetime alcohol use and risk CD or UC. 	118

* Multivariable estimates are presented with the P value for linear trend across quantiles of the exposure of interest where available. aMED, adapted Mediterranean Diet Score; EPA, eicosapentaenoic acid; EPIC, European Prospective Investigation into Cancer; DHA, docosahexaenoic acid; HR, hazard ratio; IRR, incidence rate ratio; NHS, Nurses' Health Study; OR, odds ratio; PUFA, polyunsaturated fatty acid.

Table 2|

Gene-environment studies of diet in inflammatory bowel disease

Study	Dietary exposure(s)	Genetic variant(s)	Outcome(s)	Population and Design	Key findings*	Refs
Costea et al. 2014	Dietary n-6:n-3 fatty acid intake ratio	Fifteen SNPs in 3 PUFA metabolism genes (<i>FADS1</i> , <i>FADS2</i> and <i>CYP4F3</i>)	CD	• Case-control study • 182 newly diagnosed pediatric patients with CD, matched to 250 controls.	• Statistically significant interactions involving 6 of the candidate SNPs and the n-6:n-3 fatty acid ratio ($P_{\text{interaction}}=0.04$ for all comparisons). • Higher n-6:n-3 ratio was associated with an increased risk of CD among children with the GG genotype at rs1290617 in <i>CYP4F3</i> (OR 2.91, 95% CI 0.90–9.39), but not among those with the TT genotype (OR 0.92, 95% CI 0.44–1.89; $P_{\text{interaction}}=0.04$).	81
Khalili et al. 2016	Dietary potassium and sodium intake	Six SNPs in T_H17 pathway genes (<i>JAK2</i> , <i>STAT3</i> , <i>CCR6</i> , <i>IL21</i> , <i>IL10</i> and <i>IL23R</i>)	CD and UC	• Nested case-control study among women enrolled in the NHS and NHS II. • 169 individuals with CD matched to 740 control participants, and 202 individuals with UC matched to 740 control participants. Yes	• The association between dietary potassium intake and risk of CD was modified by the rs7657746 variant in <i>IL21</i> ($P_{\text{interaction}}=0.004$). • Among women with the GG genotype, increasing potassium intake was associated with a higher risk of CD (OR 1.58, 95% CI 1.15–2.16 per 200 mg increase in potassium intake). • By contrast, increasing potassium intake was associated with a lower risk of CD among those with the AA genotype (OR 0.90, 95% CI 0.82–0.98).	51
Khalili et al. 2017	Total dietary intakes of iron, heme iron and red meat.	180 GWAS-identified CD and UC risk variants	CD and UC	• Nested Case-control study among women enrolled in the NHS and NHS II. • 169 individuals with CD matched to 740 control participants, and 202 individuals with UC matched to 740 control participants.	• Statistically significant interaction between heme iron intake and the UC risk variant rs180121274 in <i>FCGR2A</i> ($P_{\text{interaction}}=7 \times 10^{-5}$). • Among women with the GG genotype, increasing heme iron intake was associated with a substantially lower risk of UC (OR 0.11, 95% CI 0.03–0.37 for each 1 g increase in heme iron intake). • Among women with the TT genotype, increasing heme iron intake was associated with an almost 3-fold higher risk of UC (OR 2.76, 95% CI 1.02–7.48).	42
Ananthkrishnan et al. 2017	Dietary n-3:n-6 fatty acid ratio	Eight SNPs in 3 PUFA metabolism genes (<i>FADS1</i> , <i>FADS2</i> and <i>CYP4F3</i>)	CD and UC	• Nested case-control study among women enrolled in the NHS and NHS II. • 101 individuals with CD and 139 individuals with UC matched to 495 control participants.	• The association between n-3:n-6 PUFA intake ratio and UC was modified by the rs4646904 SNP in <i>CYP4F3</i> ($P_{\text{interaction}}=0.049$). • A high (median) n-3:n-6 intake ratio was associated with a lower risk of UC among women with the GG or AG genotypes (OR 0.57, 95% CI 0.32–0.99), but not among those with the AA genotype (OR 0.95, 95% CI 0.47–1.93).	88

* Estimates are from multivariable-adjusted models. P for interaction is for multiplicative interaction between the dietary exposure and the genetic variant of interest. CD, Crohn's disease; NHS, Nurses' Health Study; OR, odds ratio; PUFA, polyunsaturated fatty acid; SNP, single nucleotide polymorphism; UC ulcerative colitis.